

Review

Role and Treatment of Insulin Resistance in Patients with Chronic Kidney Disease: A Review

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Abstract: Patients with chronic kidney disease (CKD) and dialysis have higher mortality than those without, and cardiovascular disease (CVD) is the main cause of death. As CVD is caused by several mechanisms, insulin resistance plays an important role in CVD. This review summarizes the importance and mechanism of insulin resistance in CKD and discusses the current evidence regarding insulin resistance in patients with CKD and dialysis. Insulin resistance has been reported to influence endothelial dysfunction, plaque formation, hypertension, and dyslipidemia. A recent study also reported an association between insulin resistance and cognitive dysfunction, non-alcoholic fatty liver disease, polycystic ovary syndrome, and malignancy. Insulin resistance increases as renal function decrease in patients with CKD and dialysis. Several mechanisms increase insulin resistance in patients with CKD, such as chronic inflammation, oxidative stress, obesity, and mineral bone disorder. There is the possibility that insulin resistance is the potential future target of treatment in patients with CKD.

Keywords: chronic kidney disease; insulin resistance; vitamin D; cardiovascular disease



Citation: Nakashima, A.; Kato, K.; Ohkido, I.; Yokoo, T. Role and Treatment of Insulin Resistance in Patients with Chronic Kidney Disease: A Review. *Nutrients* **2021**, *13*, 4349. <https://doi.org/10.3390/nu13124349>

Academic Editor: Antonio Brunetti

Received: 5 November 2021

Accepted: 30 November 2021

Published: 2 December 2021

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1. Introduction

Cardiovascular disease (CVD) is the main cause of death among patients with diabetes mellitus and chronic kidney disease (CKD). One of the main causes of CVD is insulin resistance (IR), which is a balance between calorie intake and consumption. Although IR is affected by genetic predisposition, lifestyle factors such as dietary habits and daily exercise affect IR through hypertrophy of fat cells, ultrafiltration of inflammatory cells into adipose tissue, and inappropriate secretion of adipokines [1]. Patients with a higher IR status have higher mortality and morbidity rates than those with a lower IR status [2,3]. Patients with CKD have a higher IR status than patients without because of chronic inflammation, uremic toxins, and vitamin D deficiency [4]. In addition, a recent study reported that CKD-mineral bone disorder (MBD), phosphorus, and fibroblast growth factor-23 (FGF-23) also affect IR [5]. There is the possibility that IR is improved by CKD-MBD, such as phosphate binder and vitamin D. Vitamin D has also received a great deal of attention in recent years in IR. Clinical studies have been conducted to improve IR with activated vitamin D analog and vitamin D supplementation. Although the mechanism of IR has been revealed in recent years, the detailed mechanism is unknown. Therefore, this review summarizes the importance and mechanism of IR in CVD and discusses the current evidence regarding IR and patients with CKD.

1.1. Mechanisms of IR

Insulin mainly targets the liver, skeletal muscle, and fat cells and regulates glucose metabolism via the insulin receptor. Insulin receptors are distributed in various tissues and play important physiological roles. IR is defined as the inability of cells to respond to the action of insulin. As patients with IR have less adipose tissue insulin sensitivity, skeletal muscle, and liver, the blood glucose is elevated due to IR. The main causes of IR

are obesity and overnutrition, which induce imbalance of adipocytokines, lipotoxicity, and chronic inflammation.

1.2. Adipocytokines and IR

Several factors increase IR. Adipocytes not only contain triglycerides but also secrete several adipocytokines. In obese patients, the accumulation of triglycerides induces hypertrophy of adipocytes. This hypertrophy affects the secretion of adipocytokines and causes IR. Tumor necrosis factor- α (TNF- α), free fatty acids (FFAs), and leptin may increase IR. TNF- α is an adipocytokine secreted by adipocytes, and its concentration increased in obese animal models or patients with obesity. The secretion of TNF- α inhibits insulin activity by inducing phosphorylation of insulin receptor substrate-1 (IRS-1) [6,7].

Additionally, the administration of soluble TNF- α receptors attenuated IR in fatty model animals [6]. Although skeletal muscle and the liver are the main organs where IR develops, FFAs are essential sources of lipids, and they play an important role in IR. Adipocyte atrophy induces lipolysis and releases FFA [8]. FFAs are thought to accumulate as skeletal muscle intracellular (intramyocellular lipid) and intrahepatic lipids and induce IR. Leptin is an adipocytokine that regulates appetite and induces the consumption of energy, increasing insulin action. A previous study reported the effect of leptin on obesity and decreased IR in animal models of diabetes. In addition, leptin administration decreased IR in a mouse model. A clinical study also reported a relationship between IR and serum leptin levels. However, the administration of leptin in obese patients was not beneficial.

1.3. Lipotoxicity and IR

Lipotoxicity, defined by Unger et al., is a condition in which free FFAs suppress insulin secretion of glucose responsiveness [9]. FFAs have been shown to inhibit insulin action in insulin-targeted organs, such as the liver, skeletal muscle, and adipose tissue [10]. When a fat emulsion is intravenously administered to healthy patients to increase the blood FFA concentration, insulin's glucose uptake in peripheral tissues is suppressed, resulting in worse IR [11]. FFAs taken up into cells are converted to diacylglycerol (DAG) and activate protein kinase C (PKC) and c-Jun N-terminal kinase (JNK). PKC and JNK induce IR by phosphorylating the serine residue of IRS-1 and suppressing insulin signal transduction.

Excess FFA uptake in the liver induces ectopic fatty accumulation and results in non-alcoholic fatty liver disease (NAFLD) [12]. In patients with NAFLD, the reduction of insulin receptor signaling through DAG and PKC accumulate forkhead transcription factor (FOXO) 1 and FOXO2 and upregulates gluconeogenesis [13].

Chronic inflammation and other mechanisms of IR:

In patients with obesity, inflammatory cytokines, such as interleukin (IL)-1, IL-6, and TNF- α , increase and are reported to induce higher IR conditions [14,15]. Various inflammatory cells, including macrophages, infiltrate the adipose tissue of obese people; many inflammatory cytokines suppress insulin activity, resulting in increased IR [16,17].

A recent clinical study showed that hepatitis C virus clearance by direct-acting antivirals improved IR [18,19]. In addition, another study reported that hepatitis C treatments reduce CVD events in the prediabetic population [20]. These results suggest the importance of hepatitis C virus for glycemic conditions and the possibility as a future therapeutic target for diabetes mellitus.

2. Assessment of IR in Patients with Diabetes Mellitus

Clinicians usually use glycated hemoglobin A1c and glycated albumin to confirm patients' glucose levels and the therapeutic effects of treatment. However, these analyses cannot be used to assess IR. The homeostasis model assessment of insulin resistance (HOMA-IR) and serum insulin levels is usually used to assess the condition of insulin resistance. HOMA-IR is calculated from the fasting blood insulin level in the early morning and the fasting blood glucose level and is often used clinically as a simple index of IR. It correlates well with IR determined by the clamp method when the fasting blood glucose

level is 140 mg/dL or less. The normal value of HOMA-IR is less than 1.6, and patients with a value higher than 2.5 are regarded as having a high IR status. HOMA-IR is a method in which only one blood sample is required, and the cost of the analysis is low. However, HOMA-IR may not be useful in patients receiving basal insulin subcutaneous therapy because the actual status of IR is not reflected. Fasting IR is normal at 2–10 $\mu\text{g}/\text{dL}$, and if it exceeds that, it is considered to be compensated for by insulin hypersecretion. In type 2 diabetes and its reserves, there is often a delay in additional insulin secretion. Even if the insulin index decreases 30 min after loading, insulin secretion is high after 60 or 120 min. This condition often becomes a measure of IR.

The glucose clamp method was devised by DeFronzo et al. and is regarded as the gold standard method for assessing IR [21]. However, the technique is complicated because it requires time for equipment and inspection, making it difficult to use in daily medical treatment and large-scale clinical studies. With the patient in a fasting and resting supine position, insulin is infused from one arm, and the blood insulin concentration is maintained at 100 $\mu\text{U}/\text{mL}$. At the same time, glucose is administered (usually 10% glucose is used), blood is collected from the opposite arm, and the blood glucose level is maintained at 100 mg/dL 90–120 min later. The glucose administration rate (glucose infusion rate, mg/kg/min) is regarded as the glucose absorption rate in response to insulin stimulation. This rate is usually low in patients with IR.

3. IR in Patients with CKD

Previous studies have reported that IR is higher in patients with CKD than in the general population. IR has already been reported to occur during stage 1 of CKD [22], although another large-scale study ($n = 17,157$) reported that a decreased estimated glomerular filtration rate (eGFR) was not related to increased IR [23]. Another study reported a significant association between IR and eGFR, but this association disappeared after adjusting for variables in the multivariate analysis, including the body mass index (BMI) [24]. As the above-mentioned studies included patients with early stage CKD, the association between IR and kidney function was affected by BMI in patients with normal kidney function (Figure 1).

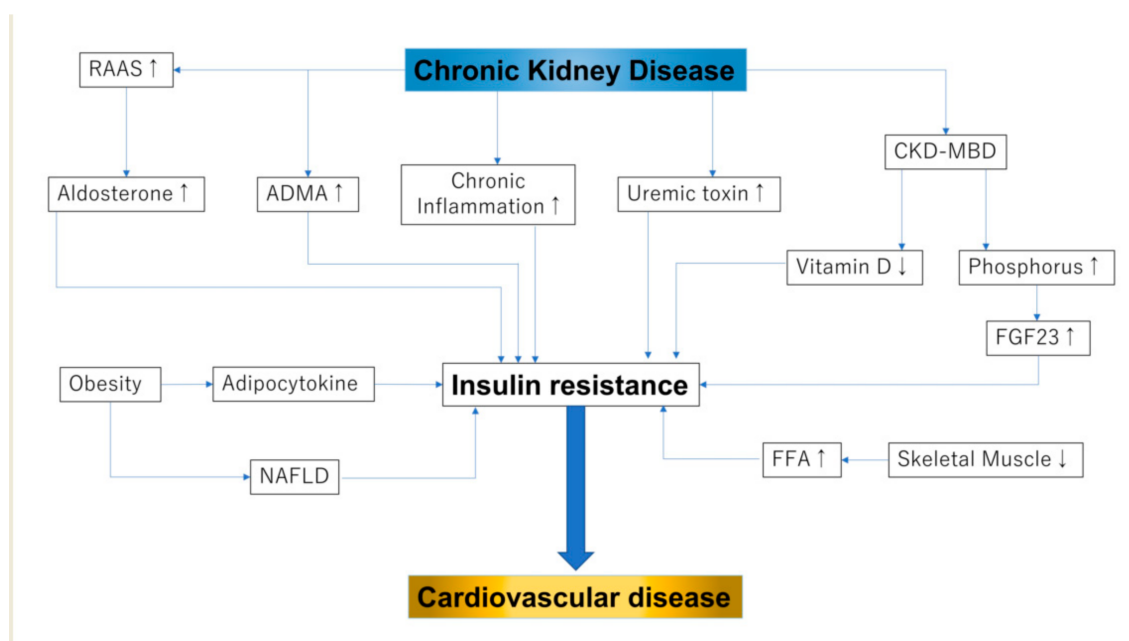


Figure 1. Insulin resistance and chronic kidney disease: RAAS, renin–angiotensin–aldosterone sys; FGF-23, fibroblast growth factor 23; NAFLD, non-alcoholic fatty liver disease; FFA, free fatty acid. In this figure, up-arrow shows enhanced activity and down-arrow shows reduced activity.

In patients with CKD stages 3–4, HOMA-IR was high, and the primary determinant of IR in this population was BMI, specifically fat mass [25]. Among patients undergoing dialysis, HOMA-IR was also high and correlated with inflammatory markers [26]. CKD progression and IR have also been analyzed in several studies. HOMA-IR and metabolic syndrome are risk factors for CKD progression and reduced eGFR [27]. One study reported that a high level of HOMA-IR was associated with the development of albuminuria in relatively healthy individuals without diabetes mellitus [28]. Another study also showed that the urinary albumin-creatinine ratio is associated with HOMA-IR after adjustment for kidney function, BMI, blood pressure, and diabetes mellitus [29]. A recent interventional study showed that multifactorial treatment, not only glycol-metabolic control but also hypertension and dyslipidemia by non-pharmacological and pharmacological approaches, prevent CVD events in diabetic CKD patients. This study suggests the importance of multifactorial approach in treating CKD patients [30].

In a study using renal tissue, IR was reported to correlate with renal tubular interstitial fibrosis and arteriolosclerosis of renal blood vessels among renal tissue disorders [31]. The increase in adiponectin, an inflammatory adipocytokine, such as $\text{TNF}\alpha$, $\text{IL-1}\beta$, and IL-6 , has a large effect on the renal disorders associated with obesity, causing an increase in the mesangial matrix and inflammation of the renal tubular epithelium [32]. The IRS-PI3K/AKT, downstream of insulin binding to its receptor, is believed to cause IR in CKD [33,34].

Uremic toxins accumulate in the body as renal function declines, but their origins vary from endogenous metabolites, gut microbiota metabolites, and exogenous metabolites. Recently, uremic toxins such as blood urea nitrogen (BUN), *p*-cresyl sulfate, indoxyl sulfate, and asymmetric dimethylarginine (ADMA) have attracted attention as causes of renal IR. BUN affects reactive oxygen species (ROS), and tyrosine of the IRS causes ROS production, which has been shown to suppress phosphorylation and PKB/Akt phosphorylation and attenuate glucose transport in CKD model mice [35].

There are several theories for kidney dysfunction and IR upregulation. First, uremic toxins increase IR through oxidative stress and ROS activity [35]. In patients with CKD, the number of insulin receptors, insulin coupling ability, and tyrosine kinase activity are important factors of IR. Second, uremic toxins induce ROS production, cause IR, and increase adipokines.

ADMA also increases renal failure and correlates with the degree of coronary artery calcification, an arteriosclerotic disease [36]. However, when corrected by HOMA-IR, the correlation disappears, so the involvement of ADMA in coronary artery calcification with IR is associated with the upstream of ADMA. ADMA is an antagonist of endogenous NO. It is decomposed by dimethylarginine dimethylaminohydrolase (DDAH), and DDAH has been reported in the ADMA system involved in stimulating insulin secretion in pancreatic β cells [37].

A previous study reported that ADMA treatment led to oxidative stress and steatosis, whereas overexpression of DDAH decreased palmitic acid-induced steatosis, oxidative stress, and inflammation. These results suggest that the ADMA/DDAH pathway is important for IR and NAFLD and that DDAH is a potential therapy for IR [38].

Insulin-induced glucose in adipocytes is associated with decreased DDAH levels and a concomitant increase in ADMA levels. It suppresses metabolism, but these changes are modified by administering the aldosterone antagonist spironolactone [39].

In patients with primary aldosteronism, after treatment with adrenalectomy or a mineralocorticoid receptor antagonist for primary aldosteronism, the insulin response to glucose increases, and insulin clearance decreases. As this effect is independent of kidney function and cortisol levels, a previous study suggested that aldosterone affects insulin secretion and IR [40].

4. CKD-MBD and IR

CKD-MBD is a concept that involves abnormalities in phosphorus, calcium, the parathyroid hormone (PTH), and other bone markers and aims to prevent CVD events

and fractures [41]. The main pathological mechanism of CKD-MBD is the elimination of phosphorus in the renal tubules and the activation disorder of vitamin D [42]. The association between CKD-MBD and IR has also been investigated in recent years (Figure 2).

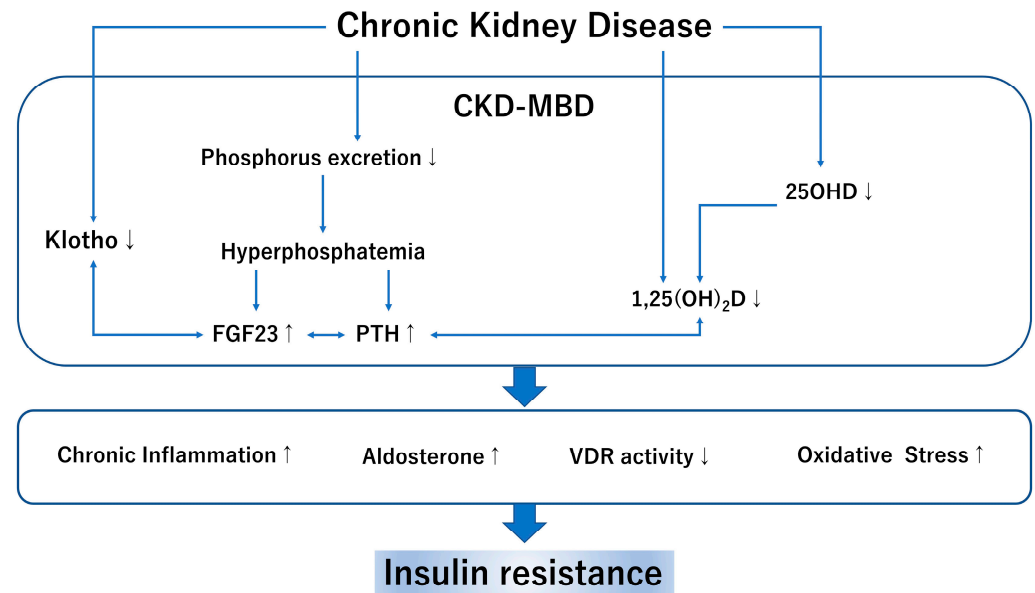


Figure 2. CKD-MBD and insulin resistance: CKD, chronic kidney disease; MBD, mineral bone disorder; FGF23 fibroblast growth factor 23; PTH, parathyroid hormone; 25OHD, 25 hydroxyvitamin D; 1,25(OH)₂D, 1,25 dihydroxy vitamin D; VDR, vitamin D receptor. In this figure, up-arrow shows enhanced activity and down-arrow shows reduced activity.

4.1. Vitamin D and IR

Vitamin D is a fat-soluble steroid hormone that promotes dietary calcium and phosphorus absorption through the vitamin D receptor (VDR) [43]. VDR is expressed in whole-body organs, including the heart, liver, kidney, blood vessels, and central nervous system. Moreover, the beta cells in the pancreas also have the VDR and play an important role [44]. Several mechanisms of the association between vitamin D and IR have been reported. Vitamin D stimulates insulin receptors and is thought to be involved in glucose transport by increasing the reactivity to insulin [45]. As pancreatic beta cells are affected by cytokine-induced apoptosis, a high inflammation status causes worsening glycemic control. Vitamin D can decrease the effects of systemic chronic inflammation and protect against beta-cell cytokine-induced apoptosis by directly modulating the expression and activity of cytokines [46,47]. Another study reported that VDR activation by a vitamin D analog reduced liver inflammation and improved IR [48]. Additionally, vitamin D improves hyperactivity of the aldosterone effect through the renin-angiotensin system, activates the function of beta cells in the pancreas, and improves IR [49,50]. Low vitamin D levels predicted future glycemia and IR in a 10-year follow-up cohort study [51]. Other studies have shown that low vitamin D levels are associated with IR [52–54].

4.2. Phosphorus, FGF-23, and IR

FGF-23, an endocrine hormone secreted by osteocytes, regulates vitamin D metabolism and phosphorus balance through renal tubules. High levels of FGF-23 are associated with all-cause mortality and CVD events in patients with CKD, CKD, and dialysis [55–57]. The association between FGF-23 and IR has been studied recently. Several studies have reported that FGF-23 contributes to IR in patients with CKD and obesity [58–61]. Resistin, expressed in monocytes and macrophages, is the key peptide of IR and is involved in inflammatory processes [62]. We have reported that resistin is associated with FGF-23 in patients with type 2 diabetes mellitus [63]. Because this association is significant after adjustment for

25-hydroxyvitamin D (25OHD), C-peptide, ghrelin, leptin, PTH, FGF-23, and resistin are related to each other. Although the detailed mechanisms of FGF-23 and resistin have not been elucidated, it is considered that chronic inflammation of FGF-23 may induce a higher resistin status. Although phosphorus restriction and phosphate binder decrease serum FGF23 levels, no study investigated whether phosphorus and FGF23 control improved resistin levels in IR status.

5. Treatment of IR

There are two main approaches to the treatment of IR: weight loss and pharmacological therapy. Inflammatory cytokines are produced in obese adipose tissue, and infiltration of immune cells is observed; this causes abnormal glucose metabolism and IR. In addition, the gut microbiota of patients with obesity differs compared to that in the general population. This alteration causes intestinal barrier dysfunction, secretory deficiency of incretin in the intestine, and activation of chronic inflammation. These changes worsen the conditions of IR and cause NAFLD and diabetes mellitus [64–66]. Short-chain fatty acids produced by the breakdown of dietary fiber caused by intestinal bacteria promote energy consumption and increase appetite-suppressing hormones [67].

For severe obesity with a BMI of 35 kg/m² or higher, the goal is to reduce weight by 5–10% with a calorie intake of 20–25 kcal or less per standard daily body weight. As exercise is an important method to reduce body weight, several guidelines recommend regular exercise. To reduce 5–7.5 kg weight, the American College of Sports Medicine recommends 225–420 min of aerobic exercise per week [68].

Thiazolidine improves IR by inhibiting the peroxisome proliferator-activated receptor (PPAR) that regulates fatty cells. It was revealed that thiazolidine increases the blood adiponectin concentration while reducing the expression of inflammatory cytokines, such as TNF- α and monocyte chemoattractant protein-1, and improving IR [69,70]. Pioglitazone was found to reduce CVD events by 10%. Furthermore, the combined incidence of all-cause mortality, non-fatal myocardial infarction, and stroke, which are hard endpoints, was reduced by 16% [71]. When pairing type 2 diabetic patients with and without CVD, the amount of change in the carotid intima-media complex thickening (mean IMT) in thiazolidine was verified. The mean IMT of the glimepiride group increased by 0.012 mm, whereas that of the pioglitazone group decreased by 0.001 mm. It was revealed that pioglitazone suppressed the progression of IMT [72]. However, since there has been concern about using thiazolidine for fracture, bladder cancer, and fluid retention, it should be used carefully [70].

Sodium-glucose transport protein 2 (SGLT2) inhibitors suppress glucose reabsorption in the proximal tubule and improve blood glucose levels by increasing glucose excretion in the urine [73]. SGLT2 decreases weight as a result of catabolic fat burning and improves IR [74]. In an animal diabetic model study, SGLT2 improved IR by stimulating muscle glucose uptake [75].

Is CKD-MBD the Potential Option for Treating IR?

As aforementioned, CKD-MBD, including vitamin and FGF-23, is closely associated with IR. There are two types of research on vitamin D for IR: activated vitamin D analog and vitamin D supplementation. Spoto et al. reported whether paricalcitol improves HOMA-IR in CKD stage 3–4 patients. Although paricalcitol decreases serum parathyroid hormone levels, there are no significant differences in IR between paricalcitol and placebo [76]. Mahmoudi et al. reported that calcitriol, an activated vitamin D analog, improved HOMA-IR in patients with NAFLD [77]. Another study about activated vitamin D analog for IR reported positive effects in NAFLD patients [78]. However, a 2-month crossover randomized controlled trial about paricalcitol or placebo did not show positive effects for IR in the vitamin D group [79]. Although interventional studies about activated vitamin D analogs are relatively fewer than vitamin D supplementation, these studies almost showed that the activated vitamin D analog improved IR.

The studies about vitamin D supplementation, cholecalciferol or ergocalciferol, are much more than activated vitamin D analogs and have shown several results. One study reported that vitamin D supplementation, cholecalciferol, improved HOMA-IR and the insulin sensitivity check index in premenopausal women [78]. On the other hand, there are opposite results for vitamin D regarding IR [48,80,81]. Several meta-analyses of vitamin D supplementation for IR have been reported. He et al. reported that although vitamin D supplementation had no significant effect on IR, insulin resistance was improved for those with 25(OH)D \geq 30 ng/mL by stratified analysis [82]. Other studies also suggested vitamin D supplementation improves IR in prediabetics and polycystic ovary syndrome [83,84]. The evidence of activated vitamin D analogue and vitamin D supplementation for IR was insufficient, and well-designed clinical studies are needed.

Although phosphorus and FGF-23 are associated with IR, these results have been investigated in mainly observational studies [59,61,63,85]. New evidence is needed to determine whether diet phosphate control or phosphate binders improve IR. Phosphate binders are divided into calcium-containing phosphate binders and non-calcium-containing phosphate binders, which have different effects on vascular calcification [86,87]. As earlier studies reported, calcium-containing phosphate and non-calcium-containing phosphate binders decrease FGF23 [88–90]. Future studies that compare phosphate binder and placebo for IR are needed. In addition, researchers should investigate the differences between non-calcium-containing phosphate and calcium-containing phosphate binders for IR.

This review article has some limitations. First, the clinical evidence of IR is lacking, especially randomized controlled trials that explored the possibility of decreasing IR. Therefore, future well-designed studies will be needed. Second, since IR is regarded as a risk factor for CVD events, few data exist on other clinical important events, such as fracture and malignancy.

6. Conclusions

IR is important not only for diabetes mellitus but also for CKD. CKD-MBD is associated with IR and has the potential to be a treatment option. Future well-designed studies are required to investigate this further.

Author Contributions: Conceptualization: A.N., K.K., I.O. and T.Y.; methodology: A.N. and K.K.; writing—original draft preparation: A.N.; writing—review and editing: A.N., K.K., I.O. and T.Y. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: This work was supported by JSPS KAKENHI Grant Number 18K16012 and 20K17259.

Conflicts of Interest: The authors declare that there are no conflict of interest.

Abbreviations

chronic kidney disease	(CKD)
cardiovascular disease	(CVD)
insulin resistance	(IR)
CKD-mineral bone disorder	(MBD)
fibroblast growth factor-23	(FGF-23)
tumor necrosis factor-alpha	(TNF- α),
free fatty acids	(FFAs)
insulin receptor substrate-1	(IRS-1)
diacylglycerol	(DAG)
non-alcoholic fatty liver disease	(NAFLD)

forkhead transcription factor	(FOXO)
interleukin	(IL)-1
homeostasis model assessment of insulin resistance	(HOMA-IR)
estimated glomerular filtration rate	(eGFR)
body mass index	(BMI)
asymmetric dimethylarginine	(ADMA)
blood urea nitrogen	(BUN)
reactive oxygen species	(ROS)
dimethylaminohydrolase	(DDAH)
parathyroid hormone	(PTH)
vitamin D receptor	(VDR)
25 hydroxyvitamin D	(25OHD)
peroxisome proliferator activated receptor	(PPAR)
intima-media complex thickening	(mean IMT)
sodium-glucose transport protein 2	(SGLT2)
activate protein kinase C	(PKC)
c-Jun N-terminal kinase	(JNK)

References

- Czech, M.P. Insulin action and resistance in obesity and type 2 diabetes. *Nat. Med.* **2017**, *23*, 804–814. [[CrossRef](#)]
- Welsh, P.; Preiss, D.; Lloyd, S.M.; De Craen, A.J.; Jukema, J.W.; Westendorp, R.G.; Buckley, B.M.; Kearney, P.; Briggs, A.; Stott, D.J.; et al. Contrasting associations of insulin resistance with diabetes, cardiovascular disease and all-cause mortality in the elderly: PROSPER long-term follow-up. *Diabetologia* **2014**, *57*, 2513–2520. [[CrossRef](#)]
- de Boer, I.H.; Katz, R.; Chonchol, M.B.; Fried, L.F.; Ix, J.H.; Kestenbaum, B.; Mukamal, K.J.; Peralta, C.A.; Siscovick, D.S. Insulin Resistance, Cystatin C, and Mortality Among Older Adults. *Diabetes Care* **2012**, *35*, 1355–1360. [[CrossRef](#)] [[PubMed](#)]
- Xu, H.; Carrero, J.J. Insulin resistance in chronic kidney disease. *Nephrology* **2017**, *22*, 31–34. [[CrossRef](#)]
- Hanks, L.J.; Casazza, K.; Judd, S.E.; Jenny, N.S.; Gutiérrez, O.M. Associations of Fibroblast Growth Factor-23 with Markers of Inflammation, Insulin Resistance and Obesity in Adults. *PLoS ONE* **2015**, *10*, e0122885. [[CrossRef](#)] [[PubMed](#)]
- Hotamisligil, G.S.; Shargill, N.S.; Spiegelman, B.M. Adipose Expression of Tumor Necrosis Factor-Alpha: Direct Role in Obesity-Linked Insulin Resistance. *Science* **1993**, *259*, 87–91. [[CrossRef](#)] [[PubMed](#)]
- Liu, L.S.; Spelleken, M.; Rohrig, K.; Hauner, H.; Eckel, J. Tumor necrosis factor-alpha acutely inhibits insulin signaling in human adipocytes: Implication of the p80 tumor necrosis factor receptor. *Diabetes* **1998**, *47*, 515–522. [[CrossRef](#)] [[PubMed](#)]
- Jenkins, A.B.; Storlien, L.H.; Chisholm, D.J.; Kraegen, E.W. Effects of nonesterified fatty acid availability on tissue-specific glucose utilization in rats in vivo. *J. Clin. Investig.* **1988**, *82*, 293–299. [[CrossRef](#)]
- Unger, R.H. Lipotoxicity in the Pathogenesis of Obesity-Dependent NIDDM: Genetic and Clinical Implications. *Diabetes* **1995**, *44*, 863–870. [[CrossRef](#)]
- Lee, Y.; Ravazzola, M.; Park, B.-H.; Bashmakov, Y.K.; Orci, L.; Unger, R.H. Metabolic Mechanisms of Failure of Intraportally Transplanted Pancreatic -Cells in Rats: Role of Lipotoxicity and Prevention by Leptin. *Diabetes* **2007**, *56*, 2295–2301. [[CrossRef](#)]
- Roden, M.; Price, T.B.; Perseghin, G.; Petersen, K.F.; Rothman, D.L.; Cline, G.W.; Shulman, G. Mechanism of free fatty acid-induced insulin resistance in humans. *J. Clin. Investig.* **1996**, *97*, 2859–2865. [[CrossRef](#)]
- Birkenfeld, A.L.; Shulman, G.I. Nonalcoholic fatty liver disease, hepatic insulin resistance, and type 2 Diabetes. *Hepatology* **2013**, *59*, 713–723. [[CrossRef](#)] [[PubMed](#)]
- Jung, Y.; Lee, M.K.; Puri, P.; Koo, B.K.; Joo, S.K.; Jang, S.Y.; Lee, D.H.; Jung, Y.J.; Kim, B.G.; Lee, K.L.; et al. Circulating lipidomic alterations in obese and non-obese subjects with non-alcoholic fatty liver disease. *Aliment. Pharmacol. Ther.* **2020**, *52*, 1603–1614. [[CrossRef](#)] [[PubMed](#)]
- Tack, C.J.; Stienstra, R.; Joosten, L.A.B.; Netea, M.G. Inflammation links excess fat to insulin resistance: The role of the interleukin-1 family. *Immunol. Rev.* **2012**, *249*, 239–252. [[CrossRef](#)] [[PubMed](#)]
- Wu, H.; Ballantyne, C.M. Skeletal muscle inflammation and insulin resistance in obesity. *J. Clin. Investig.* **2017**, *127*, 43–54. [[CrossRef](#)]
- Lee, Y.S.; Olefsky, J. Chronic tissue inflammation and metabolic disease. *Genes Dev.* **2021**, *35*, 307–328. [[CrossRef](#)]
- Donath, M.Y.; Zeman-Meier, D.; Böni-Schnetzler, M. Inflammation in the Pathophysiology and Therapy of Cardiometabolic Disease. *Endocr. Rev.* **2019**, *40*, 1080–1091. [[CrossRef](#)]
- Adinolfi, L.E.; Petta, S.; Fracanzani, A.L.; Nevola, R.; Coppola, C.; Narciso, V.; Rinaldi, L.; Calvaruso, V.; Pafundi, P.C.; Lombardi, R.; et al. Reduced incidence of type 2 diabetes in patients with chronic hepatitis C virus infection cleared by direct-acting antiviral therapy: A prospective study. *Diabetes Obes. Metab.* **2020**, *22*, 2408–2416. [[CrossRef](#)]
- Adinolfi, L.E.; Petta, S.; Fracanzani, A.L.; Coppola, C.; Narciso, V.; Nevola, R.; Rinaldi, L.; Calvaruso, V.; Staiano, L.; Di Marco, V.; et al. Impact of hepatitis C virus clearance by direct-acting antiviral treatment on the incidence of major cardiovascular events: A prospective multicentre study. *Atherosclerosis* **2020**, *296*, 40–47. [[CrossRef](#)]

20. Sasso, F.C.; Pafundi, P.C.; Caturano, A.; Galiero, R.; Vetrano, E.; Nevola, R.; Petta, S.; Fracanzani, A.L.; Coppola, C.; Di Marco, V.; et al. Impact of direct acting antivirals (DAAs) on cardiovascular events in HCV cohort with pre-diabetes. *Nutr. Metab. Cardiovasc. Dis.* **2021**, *31*, 2345–2353. [[CrossRef](#)]
21. A DeFronzo, R.; Tobin, J.D.; Andres, R. Glucose clamp technique: A method for quantifying insulin secretion and resistance. *Am. J. Physiol. Metab.* **1979**, *237*, E214–E223. [[CrossRef](#)] [[PubMed](#)]
22. Becker, B.; Kronenberg, F.; Kielstein, J.T.; Haller, H.; Morath, C.; Ritz, E.; Fliser, D. Renal Insulin Resistance Syndrome, Adiponectin and Cardiovascular Events in Patients with Kidney Disease: The Mild and Moderate Kidney Disease Study. *J. Am. Soc. Nephrol.* **2005**, *16*, 1091–1098. [[CrossRef](#)]
23. Park, J.H.; Oh, S.W.; Ahn, S.Y.; Kim, S.; Na, K.Y.; Chae, N.-W.; Kim, S.; Chin, H.J. Decreased estimated glomerular filtration rate is not directly related to increased insulin resistance. *Diabetes Res. Clin. Pr.* **2013**, *99*, 366–371. [[CrossRef](#)] [[PubMed](#)]
24. Chen, S.; Chen, Y.; Liu, X.; Li, M.; Wu, B.; Li, Y.; Liang, Y.; Shao, X.; Holthöfer, H.; Zou, H. Association of Insulin Resistance with Chronic Kidney Disease in Non-Diabetic Subjects with Normal Weight. *PLoS ONE* **2013**, *8*, e74058. [[CrossRef](#)]
25. Trirogoff, M.L.; Shintani, A.; Himmelfarb, J.; Ikizler, T.A. Body mass index and fat mass are the primary correlates of insulin resistance in nondiabetic stage 3–4 chronic kidney disease patients. *Am. J. Clin. Nutr.* **2007**, *86*, 1642–1648. [[CrossRef](#)]
26. Gohda, T.; Gotoh, H.; Tanimoto, M.; Sato, M.; Io, H.; Kaneko, K.; Hamada, C.; Tomino, Y. Relationship between Abdominal Fat Accumulation and Insulin Resistance in Hemodialysis Patients. *Hypertens. Res.* **2008**, *31*, 83–88. [[CrossRef](#)]
27. Ma, A.; Liu, F.; Wang, C.; Liang, K.; Yan, F.; Hou, X.; Liu, J.; Chen, L. Both insulin resistance and metabolic syndrome accelerate the progression of chronic kidney disease among Chinese adults: Results from a 3-year follow-up study. *Int. Urol. Nephrol.* **2018**, *50*, 2239–2244. [[CrossRef](#)]
28. Jang, C.M.; Hyun, Y.Y.; Lee, K.B.; Kim, H. Insulin resistance is associated with the development of albuminuria in Korean subjects without diabetes. *Endocrine* **2014**, *48*, 203–210. [[CrossRef](#)] [[PubMed](#)]
29. Gu, S.; Wang, A.; Ning, G.; Zhang, L.; Mu, Y. Insulin resistance is associated with urinary albumin-creatinine ratio in normal weight individuals with hypertension and diabetes: The REACTION study. *J. Diabetes* **2019**, *12*, 406–416. [[CrossRef](#)]
30. Sasso, F.C.; Pafundi, P.C.; Simeon, V.; De Nicola, L.; Chiodini, P.; Galiero, R.; Rinaldi, L.; Nevola, R.; Salvatore, T.; Sardu, C.; et al. Efficacy and durability of multifactorial intervention on mortality and MACEs: A randomized clinical trial in type-2 diabetic kidney disease. *Cardiovasc. Diabetol.* **2021**, *20*, 1–12. [[CrossRef](#)]
31. Ikee, R.; Hamasaki, Y.; Oka, M.; Maesato, K.; Mano, T.; Moriya, H.; Ohtake, T.; Kobayashi, S. Glucose Metabolism, Insulin Resistance, and Renal Pathology in Non-Diabetic Chronic Kidney Disease. *Nephron Clin. Pr.* **2008**, *108*, c163–c168. [[CrossRef](#)]
32. Szeto, H.H.; Liu, S.; Soong, Y.; Alam, N.; Prusky, G.T.; Seshan, S.V. Protection of mitochondria prevents high-fat diet-induced glomerulopathy and proximal tubular injury. *Kidney Int.* **2016**, *90*, 997–1011. [[CrossRef](#)]
33. Koppe, L.; Pelletier, C.C.; Alix, P.M.; Kalbacher, E.; Fouque, D.; Soulage, C.O.; Guebre-Egziabher, F. Insulin resistance in chronic kidney disease: New lessons from experimental models. *Nephrol. Dial. Transplant.* **2013**, *29*, 1666–1674. [[CrossRef](#)] [[PubMed](#)]
34. Thomas, S.S.; Zhang, L.; Mitch, W.E. Molecular mechanisms of insulin resistance in chronic kidney disease. *Kidney Int.* **2015**, *88*, 1233–1239. [[CrossRef](#)]
35. D’Apolito, M.; Du, X.; Zong, H.; Catucci, A.; Maiuri, L.; Trivisano, T.; Pettoello-Mantovani, M.; Campanozzi, A.; Raia, V.; Pessin, J.E.; et al. Urea-induced ROS generation causes insulin resistance in mice with chronic renal failure. *J. Clin. Investig.* **2010**, *120*, 203–213. [[CrossRef](#)] [[PubMed](#)]
36. Alipoor, E.; Hosseinzadeh, F.M.; Hosseinzadeh-Attar, M.J. Adipokines in critical illness: A review of the evidence and knowledge gaps. *Biomed. Pharmacother.* **2018**, *108*, 1739–1750. [[CrossRef](#)]
37. Hasegawa, K.; Wakino, S.; Kimoto, M.; Minakuchi, H.; Fujimura, K.; Hosoya, K.; Komatsu, M.; Kaneko, Y.; Kanda, T.; Tokuyama, H.; et al. The hydrolase DDAH2 enhances pancreatic insulin secretion by transcriptional regulation of secretagogin through a Sirt1-dependent mechanism in mice. *FASEB J.* **2013**, *27*, 2301–2315. [[CrossRef](#)] [[PubMed](#)]
38. Li, T.; Feng, R.; Zhao, C.; Wang, Y.; Wang, J.; Liu, S.; Cao, J.; Wang, H.; Wang, T.; Guo, Y.; et al. Dimethylarginine Dimethylamino-hydrolase 1 Protects Against High-Fat Diet-Induced Hepatic Steatosis and Insulin Resistance in Mice. *Antioxid. Redox Signal.* **2017**, *26*, 598–609. [[CrossRef](#)]
39. Hosoya, K.; Minakuchi, H.; Wakino, S.; Fujimura, K.; Hasegawa, K.; Komatsu, M.; Yoshifuji, A.; Futatsugi, K.; Shinozuka, K.; Washida, N.; et al. Insulin resistance in chronic kidney disease is ameliorated by spironolactone in rats and humans. *Kidney Int.* **2015**, *87*, 749–760. [[CrossRef](#)]
40. Adler, G.K.; Murray, G.R.; Turcu, A.F.; Nian, H.; Yu, C.; Solorzano, C.C.; Manning, R.; Peng, D.; Luther, J.M. Primary Aldosteronism Decreases Insulin Secretion and Increases Insulin Clearance in Humans. *Hypertension* **2020**, *75*, 1251–1259. [[CrossRef](#)]
41. Ketteler, M.; Block, G.A.; Evenepoel, P.; Fukagawa, M.; Herzog, C.A.; McCann, L.; Moe, S.M.; Shroff, R.; Tonelli, M.A.; Toussaint, N.D.; et al. Executive summary of the 2017 KDIGO Chronic Kidney Disease–Mineral and Bone Disorder (CKD-MBD) Guideline Update: What’s changed and why it matters. *Kidney Int.* **2017**, *92*, 26–36. [[CrossRef](#)] [[PubMed](#)]
42. Smith, E.R. The Use of Fibroblast Growth Factor 23 Testing in Patients with Kidney Disease. *Clin. J. Am. Soc. Nephrol.* **2014**, *9*, 1283–1303. [[CrossRef](#)]
43. Holick, M.F. Vitamin D Deficiency. *N. Engl. J. Med.* **2007**, *357*, 266–281. [[CrossRef](#)] [[PubMed](#)]
44. Gal-Moscovici, A.; Sprague, S.M. Use of vitamin D in chronic kidney disease patients. *Kidney Int.* **2010**, *78*, 146–151. [[CrossRef](#)]
45. Pittas, A.G.; Dawson-Hughes, B. Vitamin D and diabetes. *J. Steroid Biochem. Mol. Biol.* **2010**, *121*, 425–429. [[CrossRef](#)] [[PubMed](#)]

46. Pradhan, A. Obesity, Metabolic Syndrome, and Type 2 Diabetes: Inflammatory Basis of Glucose Metabolic Disorders. *Nutr. Rev.* **2008**, *65*, S152–S156. [[CrossRef](#)]
47. Oh, J.; Riek, A.E.; Darwech, I.; Funai, K.; Shao, J.; Chin, K.; Sierra, O.L.; Carmeliet, G.; Ostlund, R.E., Jr.; Bernal-Mizrachi, C. Deletion of Macrophage Vitamin D Receptor Promotes Insulin Resistance and Monocyte Cholesterol Transport to Accelerate Atherosclerosis in Mice. *Cell Rep.* **2015**, *10*, 1872–1886. [[CrossRef](#)]
48. Dong, B.; Zhou, Y.; Wang, W.; Scott, J.; Kim, K.H.; Sun, Z.; Guo, Q.; Lu, Y.; Gonzales, N.M.; Wu, H.; et al. Vitamin D Receptor Activation in Liver Macrophages Ameliorates Hepatic Inflammation, Steatosis, and Insulin Resistance in Mice. *Hepatology* **2019**, *71*, 1559–1574. [[CrossRef](#)]
49. Leung, P.S. The Potential Protective Action of Vitamin D in Hepatic Insulin Resistance and Pancreatic Islet Dysfunction in Type 2 Diabetes Mellitus. *Nutrients* **2016**, *8*, 147. [[CrossRef](#)]
50. Vaidya, A.; Williams, J. The relationship between vitamin D and the renin-angiotensin system in the pathophysiology of hypertension, kidney disease, and diabetes. *Metabolism* **2012**, *61*, 450–458. [[CrossRef](#)] [[PubMed](#)]
51. Forouhi, N.G.; Luan, J.; Cooper, A.; Boucher, B.J.; Wareham, N.J. Baseline Serum 25-Hydroxy Vitamin D Is Predictive of Future Glycemic Status and Insulin Resistance: The Medical Research Council Ely Prospective Study 1990–2000. *Diabetes* **2008**, *57*, 2619–2625. [[CrossRef](#)]
52. A Alvarez, J.; Ashraf, A.P.; Hunter, G.R.; A Gower, B. Serum 25-hydroxyvitamin D and parathyroid hormone are independent determinants of whole-body insulin sensitivity in women and may contribute to lower insulin sensitivity in African Americans. *Am. J. Clin. Nutr.* **2010**, *92*, 1344–1349. [[CrossRef](#)]
53. Weiler, H.A.; Lowe, J.; Krahn, J.; Leslie, W.D. Osteocalcin and vitamin D status are inversely associated with homeostatic model assessment of insulin resistance in Canadian Aboriginal and white women: The First Nations Bone Health Study. *J. Nutr. Biochem.* **2013**, *24*, 412–418. [[CrossRef](#)]
54. Heaney, R.P.; French, C.B.; Nguyen, S.; Ferreira, M.; Baggerly, L.L.; Brunel, L.; Veugelers, P. A Novel Approach Localizes the Association of Vitamin D Status with Insulin Resistance to One Region of the 25-Hydroxyvitamin D Continuum. *Adv. Nutr.* **2013**, *4*, 303–310. [[CrossRef](#)] [[PubMed](#)]
55. Scialla, J.J.; Kao, W.L.; Crainiceanu, C.; Sozio, S.M.; Oberai, P.C.; Shafi, T.; Coresh, J.; Powe, N.R.; Plantinga, L.C.; Jaar, B.G.; et al. Biomarkers of Vascular Calcification and Mortality in Patients with ESRD. *Clin. J. Am. Soc. Nephrol.* **2014**, *9*, 745–755. [[CrossRef](#)] [[PubMed](#)]
56. Gutiérrez, O.M.; Mannstadt, M.; Isakova, T.; Rauh-Hain, J.A.; Tamez, H.; Shah, A.; Smith, K.; Lee, H.; Thadhani, R.; Jüppner, H.; et al. Fibroblast Growth Factor 23 and Mortality among Patients Undergoing Hemodialysis. *N. Engl. J. Med.* **2008**, *359*, 584–592. [[CrossRef](#)]
57. Kendrick, J.; Cheung, A.K.; Kaufman, J.S.; Greene, T.; Roberts, W.L.; Smits, G.; Chonchol, M.; The HOST Investigators. FGF-23 Associates with Death, Cardiovascular Events, and Initiation of Chronic Dialysis. *J. Am. Soc. Nephrol.* **2011**, *22*, 1913–1922. [[CrossRef](#)]
58. Mirza, M.A.; Alsiö, J.; Hammarstedt, A.; Erben, R.G.; Michaëlsson, K.; Tivesten, Å.; Marsell, R.; Orwoll, E.; Karlsson, M.K.; Ljunggren, Ö.; et al. Circulating Fibroblast Growth Factor-23 Is Associated With Fat Mass and Dyslipidemia in Two Independent Cohorts of Elderly Individuals. *Arter. Thromb. Vasc. Biol.* **2011**, *31*, 219–227. [[CrossRef](#)]
59. Chen, P.C.; Chang, Y.D.; Lee, M.C.; Hsu, B.G. High Serum Fibroblast Growth Factor 23 Level Is Associated with Metabolic Syndrome in Kidney Transplantation Patients. *Transplant Proc.* **2020**, *520*, 3168–3172. [[CrossRef](#)] [[PubMed](#)]
60. Holecki, M.; Chudek, J.; Owczarek, A.; Olszanecka-Glinianowicz, M.; Bożentowicz-Wikarek, M.; Dulawa, J.; Mossakowska, M.; Zdrojewski, T.; Skalska, A.; Więcek, A. Inflammation but not obesity or insulin resistance is associated with increased plasma fibroblast growth factor 23 concentration in the elderly. *Clin. Endocrinol.* **2015**, *82*, 900–909. [[CrossRef](#)] [[PubMed](#)]
61. Fayed, A.; El Nokeety, M.M.; Heikal, A.A.; Abdulazim, D.O.; Naguib, M.M.; El Din, U.A.A.S. Fibroblast growth factor-23 is a strong predictor of insulin resistance among chronic kidney disease patients. *Ren. Fail.* **2018**, *40*, 226–230. [[CrossRef](#)] [[PubMed](#)]
62. Steppan, C.M.; Brown, E.J.; Wright, C.M.; Bhat, S.; Banerjee, R.R.; Dai, C.Y.; Enders, G.H.; Silberg, D.G.; Wen, X.; Wu, G.D.; et al. A family of tissue-specific resistin-like molecules. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 502–506. [[CrossRef](#)]
63. Nakashima, A.; Yokoyama, K.; Kawanami, D.; Ohkido, I.; Urashima, M.; Utsunomiya, K.; Yokoo, T. Association between resistin and fibroblast growth factor 23 in patients with type 2 diabetes mellitus. *Sci. Rep.* **2018**, *8*, 13999. [[CrossRef](#)] [[PubMed](#)]
64. Dabke, K.; Hendrick, G.; Devkota, S. The gut microbiome and metabolic syndrome. *J. Clin. Investig.* **2019**, *129*, 4050–4057. [[CrossRef](#)] [[PubMed](#)]
65. Aron-Wisniewsky, J.; Vigliotti, C.; Witjes, J.; Le, P.; Holleboom, A.G.; Verheij, J.; Nieuwdorp, M.; Clément, K. Gut microbiota and human NAFLD: Disentangling microbial signatures from metabolic disorders. *Nat. Rev. Gastroenterol. Hepatol.* **2020**, *17*, 279–297. [[CrossRef](#)]
66. Khalyfa, A.; Ericsson, A.; Qiao, Z.; Almendros, I.; Farré, R.; Gozal, D. Circulating exosomes and gut microbiome induced insulin resistance in mice exposed to intermittent hypoxia: Effects of physical activity. *EBioMedicine* **2021**, *64*, 103208. [[CrossRef](#)] [[PubMed](#)]
67. Zhao, L.; Zhang, F.; Ding, X.; Wu, G.; Lam, Y.Y.; Wang, X.; Fu, H.; Xue, X.; Lu, C.; Ma, J.; et al. Gut bacteria selectively promoted by dietary fibers alleviate type 2 diabetes. *Science* **2018**, *359*, 1151–1156. [[CrossRef](#)] [[PubMed](#)]
68. Vetter, M.L.; Faulconbridge, L.F.; Webb, V.L.; Wadden, T.A. Behavioral and pharmacologic therapies for obesity. *Nat. Rev. Endocrinol.* **2010**, *6*, 578–588. [[CrossRef](#)]

69. Pitta, E.; Liaras, K. Thiazoles and Thiazolidinones as Antioxidants. *Curr. Med. Chem.* **2013**, *20*, 4460–4480. [[CrossRef](#)]
70. Manjal, S.K.; Kaur, R.; Bhatia, R.; Kumar, K.; Singh, V.; Shankar, R.; Kaur, R.; Rawal, R.K. Synthetic and medicinal perspective of thiazolidinones: A review. *Bioorganic Chem.* **2017**, *75*, 406–423. [[CrossRef](#)]
71. Dormandy, J.A.; Charbonnel, B.; Eckland, D.J.A.; Erdmann, E.; Massi-Benedetti, M.; Moules, I.K.; Skene, A.M.; Tan, M.H.; Lefèbvre, P.J.; Murray, G.D.; et al. Secondary prevention of macrovascular events in patients with type 2 diabetes in the PROactive Study (PROspective pioglitAzone Clinical Trial In macroVascular Events): A randomised controlled trial. *Lancet* **2005**, *366*, 1279–1289. [[CrossRef](#)]
72. Mazzone, T.; Meyer, P.M.; Feinstein, S.B.; Davidson, M.H.; Kondos, G.T.; D'Agostino, R.B.; Perez, A.; Provost, J.-C.; Haffner, S.M. Effect of Pioglitazone Compared with Glimpiride on Carotid Intima-Media Thickness in Type 2 Diabetes. *JAMA* **2006**, *296*, 2572–2581. [[CrossRef](#)] [[PubMed](#)]
73. Cowie, M.R.; Fisher, M. SGLT2 inhibitors: Mechanisms of cardiovascular benefit beyond glycaemic control. *Nat. Rev. Cardiol.* **2020**, *17*, 761–772. [[CrossRef](#)]
74. Khan, R.S.; Bril, F.; Cusi, K.; Newsome, P.N. Modulation of Insulin Resistance in Nonalcoholic Fatty Liver Disease. *Hepatology* **2018**, *70*, 711–724. [[CrossRef](#)] [[PubMed](#)]
75. O'Brien, T.P.; Jenkins, E.C.; Estes, S.K.; Castaneda, A.V.; Ueta, K.; Farmer, T.D.; Puglisi, A.E.; Swift, L.L.; Printz, R.L.; Shiota, M. Correcting Postprandial Hyperglycemia in Zucker Diabetic Fatty Rats with an SGLT2 Inhibitor Restores Glucose Effectiveness in the Liver and Reduces Insulin Resistance in Skeletal Muscle. *Diabetes* **2017**, *66*, 1172–1184. [[CrossRef](#)]
76. Spoto, B.; Pizzini, P.; Cutrupi, S.; Tripepi, G.; Curatola, G.; Mallamaci, F.; Zoccali, C. Vitamin D receptor activation by paricalcitol and insulin resistance in CKD. *Nutr. Metab. Cardiovasc. Dis.* **2018**, *28*, 291–297. [[CrossRef](#)] [[PubMed](#)]
77. Mahmoudi, L.; Asadi, S.; Al-Mousavi, Z.; Niknam, R. A randomized controlled clinical trial comparing calcitriol versus cholecalciferol supplementation to reduce insulin resistance in patients with non-alcoholic fatty liver disease. *Clin. Nutr.* **2020**, *40*, 2999–3005. [[CrossRef](#)]
78. Trummer, C.; Theiler-Schwetz, V.; Kollmann, M.; Wölfler, M.; Münzker, J.; Pilz, S.; Pieber, T.R.; Heijboer, A.C.; Obermayer-Pietsch, B.; Lerchbaum, E. Effects of vitamin D supplementation on metabolic and endocrine parameters in healthy premenopausal women: A randomized controlled trial. *Clin. Nutr.* **2019**, *39*, 718–726. [[CrossRef](#)]
79. de Boer, I.H.; Sachs, M.; Hoofnagle, A.N.; Utzschneider, K.M.; Kahn, S.E.; Kestenbaum, B.; Himmelfarb, J. Paricalcitol does not improve glucose metabolism in patients with stage 3–4 chronic kidney disease. *Kidney Int.* **2013**, *83*, 323–330. [[CrossRef](#)]
80. Wallace, H.J.; Holmes, L.; Ennis, C.N.; Cardwell, C.R.; Woodside, J.V.; Young, I.; Bell, P.M.; Hunter, S.J.; McKinley, M.C. Effect of vitamin D3 supplementation on insulin resistance and β -cell function in prediabetes: A double-blind, randomized, placebo-controlled trial. *Am. J. Clin. Nutr.* **2019**, *110*, 1138–1147. [[CrossRef](#)]
81. Wood, A.D.; Secombes, K.R.; Thies, F.; Aucott, L.; Black, A.J.; Mavroei, A.; Simpson, W.G.; Fraser, W.D.; Reid, D.M.; Macdonald, H.M. Vitamin D3 Supplementation Has No Effect on Conventional Cardiovascular Risk Factors: A Parallel-Group, Double-Blind, Placebo-Controlled RCT. *J. Clin. Endocrinol. Metab.* **2012**, *97*, 3557–3568. [[CrossRef](#)]
82. He, S.; Yu, S.; Zhou, Z.; Wang, C.; Wu, Y.; Li, W. Effect of vitamin D supplementation on fasting plasma glucose, insulin resistance and prevention of type 2 diabetes mellitus in non-diabetics: A systematic review and meta-analysis. *Biomed. Rep.* **2018**, *8*, 475–484. [[CrossRef](#)] [[PubMed](#)]
83. Mirhosseini, N.; Vatanparast, H.; Mazidi, M.; Kimball, S.M. Vitamin D Supplementation, Glycemic Control, and Insulin Resistance in Prediabetics: A Meta-Analysis. *J. Endocr. Soc.* **2018**, *2*, 687–709. [[CrossRef](#)] [[PubMed](#)]
84. Łagowska, K.; Bajerska, J.; Jamka, M. The Role of Vitamin Oral Supplementation in Insulin Resistance in Women with Polycystic Ovary Syndrome: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Nutrients* **2018**, *10*, 1637. [[CrossRef](#)]
85. Wojcik, M.; Dolezal-Oltarzewska, K.; Janus, D.; Drozd, D.; Sztéfko, K.; Starzyk, J.B. FGF23 contributes to insulin sensitivity in obese adolescents—Preliminary results. *Clin. Endocrinol.* **2011**, *77*, 537–540. [[CrossRef](#)]
86. Block, G.A.; Spiegel, D.M.; Ehrlich, J.; Mehta, R.; Lindbergh, J.; Dreisbach, A.; Raggi, P. Effects of sevelamer and calcium on coronary artery calcification in patients new to hemodialysis. *Kidney Int.* **2005**, *68*, 1815–1824. [[CrossRef](#)]
87. Kakuta, T.; Tanaka, R.; Hyodo, T.; Suzuki, H.; Kanai, G.; Nagaoka, M.; Takahashi, H.; Hirawa, N.; Oogushi, Y.; Miyata, T.; et al. Effect of Sevelamer and Calcium-Based Phosphate Binders on Coronary Artery Calcification and Accumulation of Circulating Advanced Glycation End Products in Hemodialysis Patients. *Am. J. Kidney Dis.* **2011**, *57*, 422–431. [[CrossRef](#)]
88. Toida, T.; Fukudome, K.; Fujimoto, S.; Yamada, K.; Sato, Y.; Chiyotanda, S.; Kitamura, K. Effect of lanthanum carbonate vs. calcium carbonate on serum calcium in hemodialysis patients: A crossover study. *Clin. Nephrol.* **2012**, *78*, 216–223. [[CrossRef](#)]
89. Spatz, C.; Roe, K.; Lehman, E.; Verma, N. Effect of a Non-Calcium-Based Phosphate Binder on Fibroblast Growth Factor 23 in Chronic Kidney Disease. *Nephron Clin. Pr.* **2013**, *123*, 61–66. [[CrossRef](#)]
90. Covic, A.; Passlick-Deetjen, J.; Krocak, M.; Büschges-Seraphin, B.; Ghenu, A.; Ponce, P.; Marzell, B.; De Francisco, A.L. A comparison of calcium acetate/magnesium carbonate and sevelamer-hydrochloride effects on fibroblast growth factor-23 and bone markers: Post hoc evaluation from a controlled, randomized study. *Nephrol. Dial. Transplant.* **2013**, *28*, 2383–2392. [[CrossRef](#)]