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Effects of asfotase alfa on fracture healing of adult patient with hypophosphatasia and literature review

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Abstract

Objective Hypophosphatasia (HPP) is a rare inherited disorder caused by *ALPL* gene mutations, with fracture nonunion being a serious complication. This study investigated the effects of teriparatide and asfotase alfa (AA) on femoral fracture healing of an adult patient with HPP, accompanied with a literature review.

Methods A 37-year-old woman wheelchair-bound was diagnosed with HPP due to an extremely low serum alkaline phosphatase (ALP) level (4–10 U/L), who suffered from bilateral femur pain and non-union of femoral shaft fractures on both sides. Compound heterozygous missense mutations (c.382G > A and c.461C > T) were identified in exon5 of *ALPL* gene. The patient received teriparatide sequential AA therapy. Serum levels of ALP, β -isomerized carboxy-telopeptide of type I collagen (β -CTX) and procollagen type 1 amino-terminal peptide (P1NP), bone mineral density (BMD) and skeletal X-ray were measured during the treatment. Literature was searched by keywords of “Hypophosphatasia”, “HPP”, “ALPL”, “TNSALP”, “ALP” combined with “Asfotase alfa”, “AA”, “enzyme replacement therapy”, and “ERT”.

Results After unsuccessful 6-month teriparatide treatment for femoral fracture, AA treatment was initiated, at a dose of 2 mg/kg, 3 times a week. After the first month of AA treatment, serum ALP level increased from 4 to 9206 U/L, and serum calcium and phosphate levels decreased, with increase in PTH, β -CTX, and P1NP levels. After 4 months of AA treatment, her bone pain significantly alleviated, accompanied by significant shortening of the fracture line. After 10 months of AA therapy, the fracture demonstrated complete healing and the patient could walk independently. BMD at lumbar spine and hips was significantly increased. Among 295 adult patients with HPP reported in the literature, 213 (72.2%) exhibited skeletal-related symptoms and 91 (30.8%) presented with bone fractures. In addition to skeletal manifestations, the patients presented with early tooth loss, muscle weakness and ectopic calcification. AA treatment, spanning 9 weeks to 3 years, has been shown to increase ALP levels, promote fracture healing, improve mobility, and alleviate bone pain.

Conclusion Adult HPP patients mainly present with recurrent or poorly healing fractures, bone pain, and early loss of teeth. AA replacement therapy can effectively promote fracture healing, relieve bone pain, and enhance mobility.

Keywords Hypophosphatasia, Adult, Asfotase alfa, Fracture healing

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Introduction

Hypophosphatasia (HPP) is a rare inherited metabolic bone disorder that is caused by pathogenic mutations in the *ALPL* gene, which follows autosomal dominant or recessive inheritance patterns [1]. The *ALPL* gene,



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located on chromosome 1, consists of 12 exons and encodes the tissue-nonspecific isoenzyme of alkaline phosphatase (TNSALP) [2, 3]. TNSALP is predominant in skeleton, liver, kidney, and teeth [4]. Hypophosphatasia is characterized by diminished enzymatic activity of TNSALP, which results in accumulation of its substrates in bone, including pyridoxal 5'-phosphate (PLP), phosphoethanolamine (PEA), and inorganic pyrophosphate (PPi) [5]. PPi impairs bone mineralization and leads to deposition of the pyrophosphate crystals in joints. The estimated prevalence of severe and moderate HPP in European populations is approximately 1 in 300,000 and 1 in 6370, respectively [6, 7].

According to clinical characteristics and the age of onset, HPP is usually categorized into six forms: perinatal, benign prenatal, infantile, childhood, adult, and odonto HPP [8, 9]. Adult patients with HPP typically occur in middle age and commonly present with pain in weight-bearing bones, stress fractures, muscle weakness, early loss of permanent teeth, impaired ambulation [1, 10]. Overall fracture incidence is higher in adult HPP patients than general population over 18 years old, and fractures are often difficult to heal [11, 12], of which nearly three-quarters of the patients require surgery, notably, implant surgeries frequently fail in adult patients with HPP, leading to mobility impairments and psychological problems [13, 14].

HPP used to lack effective treatment and recently enzyme replacement therapy significantly relieves patients' clinical symptoms [15–17]. Asfotase alfa (AA), a recombinant human TNSALP, can improve the symptoms of children with HPP, but there is less research about its application in adult patients with HPP [18, 19]. This study prospectively observed the effects of teriparatide and sequential AA treatment on bone fractures healing in an adult HPP patient and conducted a comprehensive analysis of the therapeutic effect of AA in adult patients with HPP through reviewing all studies on adult patients with HPP receiving AA treatment.

Materials and methods

Object

The patient, a 37-year-old woman, was admitted to the Department of Endocrinology of Peking Union Medical College Hospital (PUMCH) in September 2022 due to "bilateral lower limb pain for four years". She was born at full term with a birth weight of 3.75 kg, and the length was unknown. She was breastfed for one year and she showed no significant differences in growth, development, intelligence or activity compared to her peers. She experienced early loss of her permanent teeth (the exact age was forgotten) and underwent full-mouth dental implant surgery at the age of 24 years. Since 2018, she

has experienced progressively worsening lower limb pain, along with decreased tolerance for activity and a diminished quality of life. By the end of 2022, she was unable to walk independently and relied on assistance for daily activities. She denied that her parents were consanguineously married and reported no family history of similar diseases. Physical examination revealed that her height was 162 cm, weight was 44.0 kg, and BMI was 16.77 kg/m². She entered the clinic room in a wheelchair and had all 28 teeth as dental implants. The thyroid was not enlarged, and there were no obvious abnormalities in the lungs or heart. Both hands displayed ulnar deviation of the middle fingers. No other significant skeletal deformities were noted, although mild atrophy of the lower limb muscles was observed.

The study protocol was approved by the scientific ethics committee of PUMCH. The patient signed informed consent before participating in this study.

Laboratory evaluation

Fasting blood samples were collected in the morning at about 8:00 AM. Serum calcium, phosphate, ALP, alanine aminotransferase (ALT), and creatinine (Cr) were analyzed using an automatic biochemical analyzer (ADVIA 1800, Siemens, Germany). An automated Roche electrochemiluminescence system (Roche Diagnostics, Switzerland) was used to detect serum concentrations of parathyroid hormone (PTH), and 25 hydroxyvitamin D (25-OHD), β -isomerized C-terminal telopeptide of type I collagen (β -CTX), and procollagen type 1 amino-terminal peptide (P1NP). In addition, erythrocyte sedimentation rate (ESR) was measured using an automatic biochemical analyzer (ADVIA 1800, Siemens, Germany). Free triiodothyronine (FT3), free thyroxine (FT4), thyroid-stimulating hormone (TSH), blood cortisol, and carcinoembryonic antigen (CEA) were measured using an automated Roche electrochemiluminescence system (Roche Diagnostics, Switzerland). Serum immunofixation electrophoresis (IFE) was performed using an electrophoresis instrument (ProteomeLab, Beckman Coulter, United States) for differential purposes. All biochemical indicators were measured uniformly in the central laboratory of PUMCH.

Bone mineral density and X-ray examination

Dual-energy X-ray absorptiometry (DXA, Lunar Prodigy, GE Healthcare, Madison, WI, USA) was used to measure areal bone mineral density (BMD) at lumbar spine 1–4 (LS), femoral neck (FN), trochanter, and total hip (TH) at baseline and follow-up. Radiographs of the spine, pelvis, and bilateral lower limbs were obtained at baseline. Based on the patient's clinical presentation and radiological findings, regular follow-up radiographs of both femora

were scheduled every 3–6 months to closely monitor the healing process of the fractures.

Detection of gene mutation

Genomic DNA was extracted from the peripheral leukocytes of the patient using the QIAamp DNA Mini Kit (Qiagen, Germany) according to the manufacturer's instructions. Mutation detection was performed using next-generation targeted exon capture sequencing. All mutations and potential pathogenic variants were validated by Sanger sequencing. The pathogenicity of the variants was assessed according to the American College of Medical Genetics and Genomics and Association for Molecular Pathology (ACMG/AMP) Standards and Guidelines. The pathogenicity of the missense mutation was predicted using MutationTaster software (<http://www.mutationtaster.org/>).

Treatment and follow-up

After being diagnosed with adult-HPP complicated by non-healing bilateral fractures, severe bone pain, and limited mobility, the patient requested further treatment. As AA was not yet available in mainland China, the patient was treated with experimental subcutaneous daily injections of 20 µg teriparatide to promote fracture healing. Simultaneously, the patient received daily supplementation with 600 mg calcium and 0.25 µg calcitriol every other day. However, after 6 months of the above treatment, bone pain persisted, and there was no radiographic evidence of healing in the bilateral femoral fractures.

The patient then initiated AA (Strensiq, Alexion Pharmaceuticals Inc., Boston, MA, USA) therapy and discontinued teriparatide treatment. Based on previous studies, we recommended administration of AA treatment for the patient through subcutaneous injections of 2 mg/kg per dose, three times a week [12, 20]. At 1, 4, 6, and 10 months of AA treatment, bone metabolic markers, BMD, femoral fracture imaging, and drug-related adverse reactions were monitored. The AA dosage was adjusted according to the improvement in the patient's symptoms and fracture healing status. Initially, the patient received 2 mg/kg per dose, three times a week for 4 months, followed by 1 mg/kg per dose, three times a week for 2 months, and finally, 1 mg/kg per dose, once a week for 4 months.

Literature review

We reviewed relevant studies from the following medical databases: PubMed, Web of Science, Embase, and the Cochrane Central Register of Controlled Trials (CENTRAL) up to October 2024 to identify case reports, case series, and research articles written in English on AA

treatment for adult patients with HPP. The keywords used in the literature search included “Hypophosphatasia”, “HPP”, “ALPL”, “TNSALP”, and “ALP” in combination with “Asfotase alfa”, “AA”, “enzyme replacement therapy”, “ERT”. These keywords were combined using the Boolean operators. Relevant studies were identified by screening article titles and abstracts. The inclusion criteria were studies on AA treatment in adult patients with HPP. Exclusion criteria included studies on AA treatment in adolescents, conference abstracts, and articles for which the full text was unavailable.

Statistical analyses of the data from the literature

The Kolmogorov–Smirnov test was performed to assess the normality of continuous variables. Categorical data, such as gender, form of HPP, symptoms, gene classification, and symptom improvement after AA therapy, were expressed as frequencies and percentages. Continuous variables with normal distribution, such as age, serum phosphorus, and serum calcium, were presented as mean ± SD, while those with non-normal distribution, such as serum ALP, were presented as median (interquartile range, IQR). The Mann–Whitney U test was used to compare changes in serum ALP, PLP, phosphorus, calcium, PTH, β-CTX, and P1NP before and after AA treatment. A *P* value of less than 0.05 was considered statistically significant. Statistical analyses were performed using SPSS software (version 25.0; IBM Corp., Armonk, NY, USA).

Results

Baseline characteristics of the patient in this study

The patient was a young woman who experienced early loss of both primary and permanent teeth, along with progressively worsening bilateral thigh pain over time. Laboratory tests revealed extremely low serum ALP levels (4–10 U/L) (reference range: 35–100 U/L), elevated serum phosphorus levels of 1.56 mmol/L (reference range: 0.81–1.45 mmol/L), and low 25-OHD levels of 17.3 ng/mL (reference value: > 30 ng/mL). Other bone metabolic indicators included serum calcium of 2.47 mmol/L (reference range: 2.13–2.70 mmol/L), β-CTX of 0.28 ng/mL (reference range: 0.21–0.44 ng/mL), P1NP of 27.2 ng/mL (reference range: 15.1–58.6 ng/mL), and PTH of 29.5 pg/mL (reference range: 15–65 pg/mL). Serum levels of FT3, FT4, TSH, ALT, Cr, blood cortisol, CEA, IFE were all within normal limits. X-ray examination revealed an incomplete fracture of the bilateral femoral shafts without evidence of healing.

Variant of the ALPL gene

Genetic testing showed compound heterozygous missense mutations (c.382G > A and c.461C > T) in exon 5 of

ALPL gene (Fig. 1). According to the ACMG/AMP Standards and Guidelines, both mutations were pathogenic (the former: PM2+PM3+PP3+PP4+PS3; the latter: PM2+PP3+PP4+PM3). The Mutation Taster software also predicted that both mutations were pathogenic.

Effects of AA treatment on patient with HPP in this study

Changes in clinical symptoms

After 1 month of AA treatment, the patient reported relief from bone pain, allowing her to stand and walk slowly. After 4 months of treatment, bone pain was significantly relieved, and mobility was notably improved. After 6 months of AA treatment, her bone pain completely disappeared and she could walk 2 km independently. By 10 months of AA treatment, the patient resumed normal daily life and work.

Changes in serum biochemical indicators

After 1 month of AA therapy, serum ALP concentration significantly increased to 9206 U/L, up from 4 U/L. Accompanied by an increase in bone metabolism markers, including β -CTX (0.53–0.79 ng/mL), P1NP (83.9–75.9 ng/mL), and PTH (17–49.3 pg/mL), as well as a decrease in serum calcium (2.46–2.21 mmol/L) and phosphorus (1.73–1.45 mmol/L).

After 4 months of AA treatment, the dosage was reduced to 1 mg/kg per dose, three times per week, due to significant improvement in the patient's clinical symptoms and fracture healing. After 2 months, the ALP level had decreased by a half (ALP at 5332 U/L). At the same time, serum level of β -CTX decreased while serum

calcium and phosphate levels increased. Accordingly, PTH levels also decreased. The AA dosage was then further reduced to 1 mg/kg per dose, once a week. After 4 months, the ALP level remained at the upper limit of the normal range (ALP at 97 U/L). β -CTX decreased but remained above the upper limit of the normal range, P1NP normalized, serum calcium and phosphate levels stabilized, and PTH levels continued to decrease. All laboratory data at baseline and during treatment are presented in Table 1 and Fig. 2.

Changes in BMD and fracture imaging

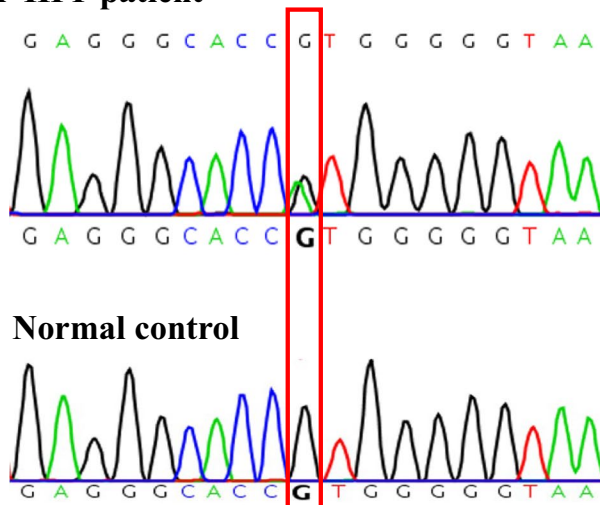
At baseline, BMD at the LS and TH were 1.029 g/cm² and 0.741 g/cm², with Z-scores of -0.2 and -1.4, respectively. After 6 months of teriparatide treatment followed by 6 months of sequential AA therapy, BMD at LS and TH improved to 1.165 g/cm² and 0.800 g/cm², with Z-scores of 1.0 and -1.0 (Table 1).

After 4 months of AA treatment, the fracture line in both femurs were shortened compared to baseline. After 6 months of AA treatment, bilateral femur fractures were nearly healed. Complete fracture healing was achieved with no visible radiolucency after 10 months of AA treatment (Fig. 3).

Safety during AA treatment

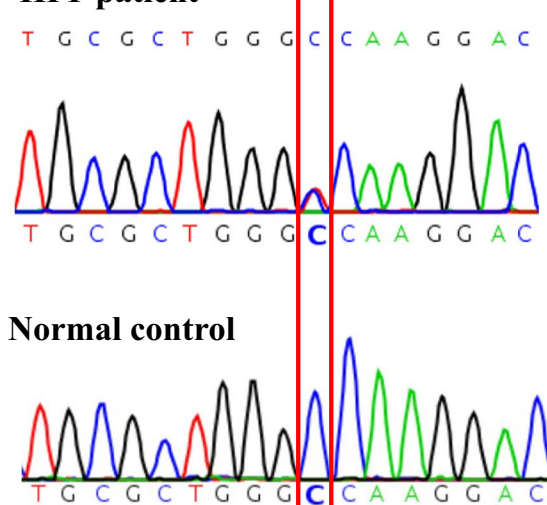
The patient reported a local injection site reaction, including redness and localized itching, within one day of AA injection. Except for that, she did not report symptoms of blurred vision, decreased vision, visual field defects, painful urination, hematuria, or other

A HPP patient



Normal control

B HPP patient



Normal control

Fig. 1 ALPL mutation in this patient revealed by whole-exome sequenc. **A** ALPL exon 5 c.382G>A heterozygous variant. **B** ALPL exon 5 c.461C>T heterozygous variant

Table 1 Changes of bone turnover biomarkers and BMD during teriparatide and sequential AA therapy

	Baseline	Teriparatide for 3 mons	Teriparatide for 6 mons	AA for 1 mon	AA for 4 mons	AA for 6 mons	AA for 10 mons	Reference range
Ca (mmol/L)	2.47	2.38	2.46	2.21	2.29	2.44	2.44	2.13–2.70
P (mmol/L)	1.56	1.68	1.73	1.45	1.51	1.70	1.69	0.81–1.45
PTH (pg/mL)	29.5	14.3	17	49.3	81.2	41.5	32.9	15.0–65.0
25-OHD (ng/mL)	17.3	20.5	19.4	17.4	17.8	21.6	13.2	30–100
ALP (U/L)	4	10	9	9206	11,843	5332	97	35–100
β -CTX (ng/mL)	0.28	0.36	0.53	0.79	1.08	0.87	0.50	0.21–0.44
P1NP (ng/mL)	27.2	91.5	83.9	75.9	52.8	62.1	27.6	15.1–58.6
ALT (U/L)	20	21	18	14	23	18	16	7–40
AST (U/L)	22	26	20	19	25	21	16	13–35
Alb (g/L)	46	43	43	43	42	45	46	35–52
Cr (μ mol/L)	56	52	53	55	57	55	56	45–84
LS aBMD/Z score	1/029/–0.2	–	–	–	–	–	1.165/1.0	–
FN aBMD/Z score	0.788/–0.6	–	–	–	–	–	0.774/–0.8	–
TH aBMD/Z score	0.741/–1.4	–	–	–	–	–	0.800/–1.0	–

AA asfotase alfa, *mon* month, *Ca* total calcium, *P* phosphorus, *PTH* parathyroid hormone, *25-OHD* 25-hydroxyvitamin D, *ALP* alkaline phosphatase, β -*CTX* β -isomerized carboxy-telopeptide of type I collagen, *P1NP* type 1 N-terminal propeptide, *ALT* alanine aminotransferase, *AST* aspartate aminotransferase, *Alb* albumin, *C* creatinine, *LS* lumbar spine, *aBMD* areal bone mineral density, *FN* femur neck, *TH* total hip

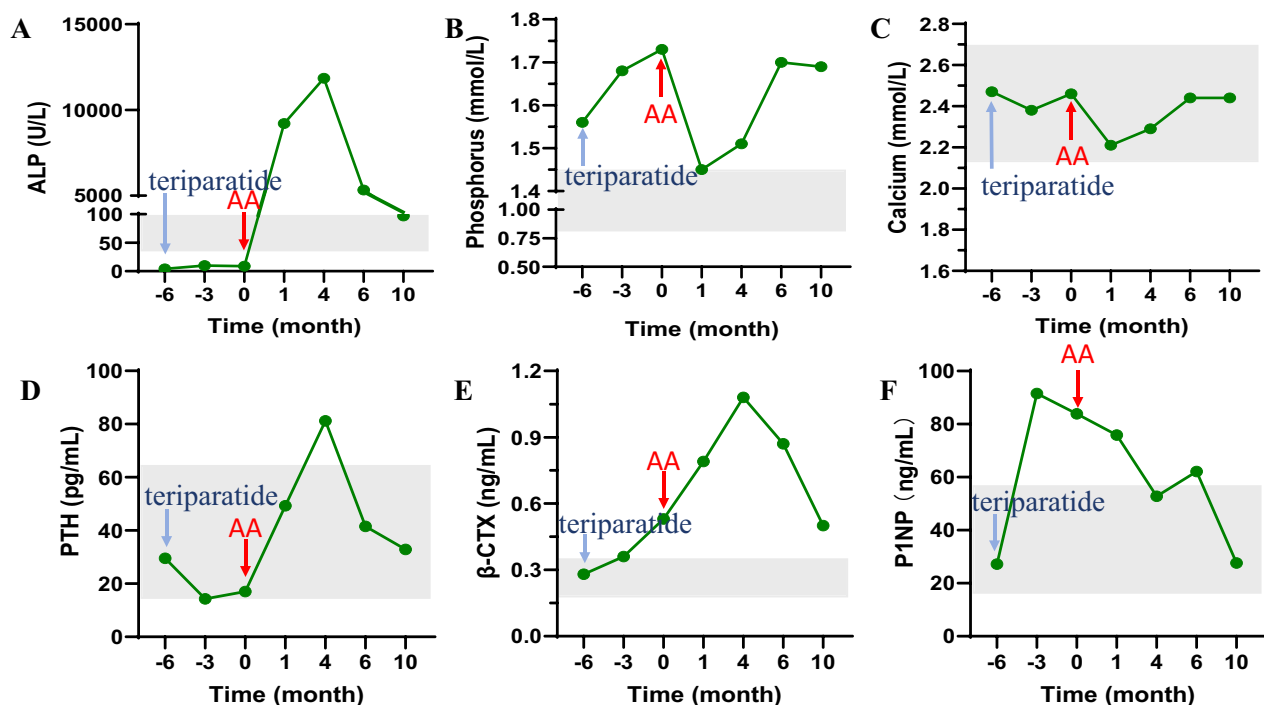


Fig. 2 Changes of bone metabolic markers during teriparatide and sequential AA therapy. **A** Serum levels of ALP during the follow-up. **B** Serum levels of phosphorus during the follow-up. **C** Serum levels of calcium during the follow-up. **D** Serum levels of PTH during the follow-up. **E** Serum levels of β -CTX during the follow-up. **F** Serum levels of P1NP during the follow-up. Annotation: The blue arrow indicates the start of teriparatide treatment. The red arrow indicates the start of AA treatment. The gray area represents the normal reference range. AA asfotase alfa, ALP alkaline phosphatase, PTH parathyroid hormone, β -CTX β -isomerized carboxy-telopeptide of type I collagen, P1NP = procollagen type 1 amino-terminal peptide

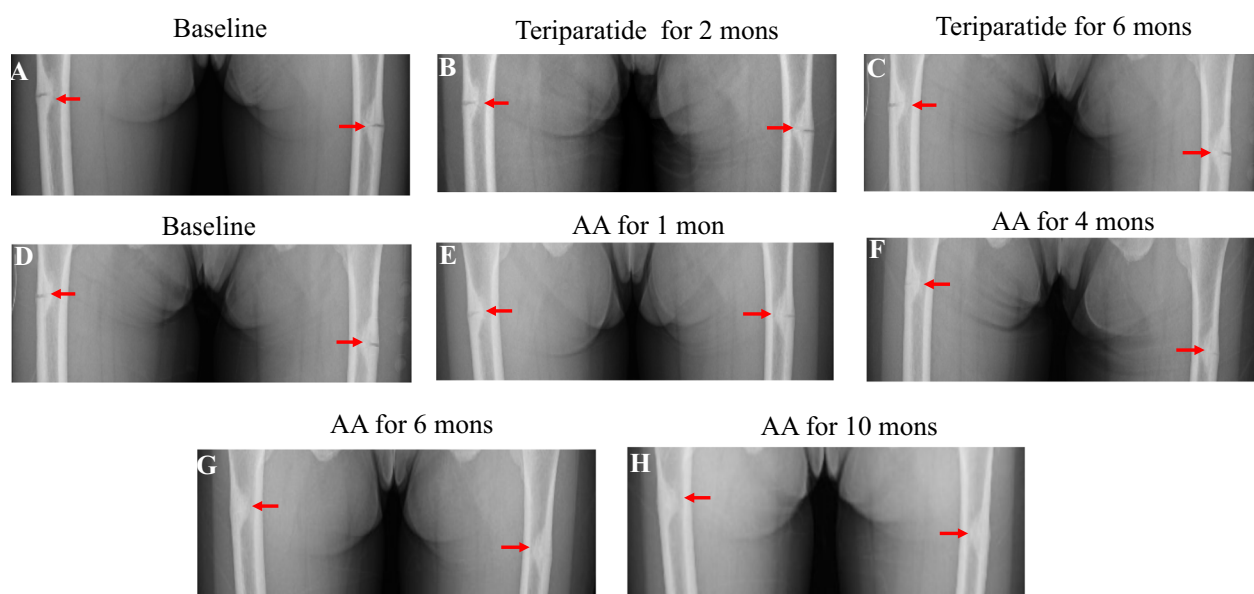


Fig. 3 Changes of bilateral femoral fractures during teriparatide treatment, and sequential AA therapy. Annotation: The arrows in the above figure indicate the bilateral proximal lateral femoral fractures. AA asfotase alfa

related discomforts. Throughout the treatment course, a decrease in the patient's calcium levels was observed following medication, but it remained above the lower limit. Additionally, liver enzyme and Cr levels remained consistently within the normal range.

Results of the literature review

As shown in Fig. 4, a total of 155 references were identified from the databases. Among them, the treatment effects of AA in adult patients with HPP were evaluated in 17 studies, which comprised 12 case reports ($n=21$) and 5 clinical studies ($n=274$) (Tables 2 and 3) [13, 14, 18, 20–33].

Among 295 adult patients with HPP, the mean age was 45.14 ± 16.68 years, with 193 women and 102 men. 91.5% (270/295) had pediatric-onset HPP, and only 8.5% were classified as adult-onset HPP. The majority of reported cases receiving AA treatment were Caucasian, with only four patients being Japanese. No reports or studies have documented Chinese patients receiving AA treatment, as AA has not been approved for marketing in China. Among these patients with HPP, 30.8% (91/295) had a history of bone fractures. 49.4% (146/295) had muscular manifestations of HPP, with some patients experiencing limited daily activities. 71.9% (212/295) had dental manifestations, 79.7% (235/295) had a history of pain, and 17.6% (52/295) had neurological abnormalities.

Among the 17 studies, 76.5% (13/17) used a treatment dosage of 6 mg/kg per week for AA, with some

administering it as three injections per week and others as six. After AA therapy, 83.3% of patients reported relief from bone pain, 98.4% experienced improved mobility, and 93.2% reported enhanced quality of life. However, the literature review identified only a few case reports that explored the effectiveness of AA therapy in fracture healing for adult patients with HPP. Among them, 10 patients showed improvement in fracture healing. The average healing time after AA treatment was 14.25 ± 6.68 months, with the shortest reported healing time being 6 months and the longest 24 months.

After AA treatment, all studies reported a significant increase in serum ALP activity, with a median level of 5873.00 ± 2787.62 U/L and a highest recorded level of 13,336 U/L. Additionally, the levels of PTH, osteocalcin, P1NP, and β -CTX were increased, while serum calcium and phosphate levels decreased.

Regarding safety evaluation, 28.1% (77 of 274) of patients reported injection site reactions (ISRs) during treatment, while 20.8% (57 of 274) developed injection site lipodystrophy resembling lipohypertrophy. Three patients experienced severe allergic reactions, primarily manifesting as difficulty breathing, choking sensation, swelling of the eyes, lips, or tongue, dizziness, vomiting, fever, and chills, which required discontinuation of AA therapy. The patient in this study reported local injection reactions within one day of AA administration, including skin redness and localized itching, with

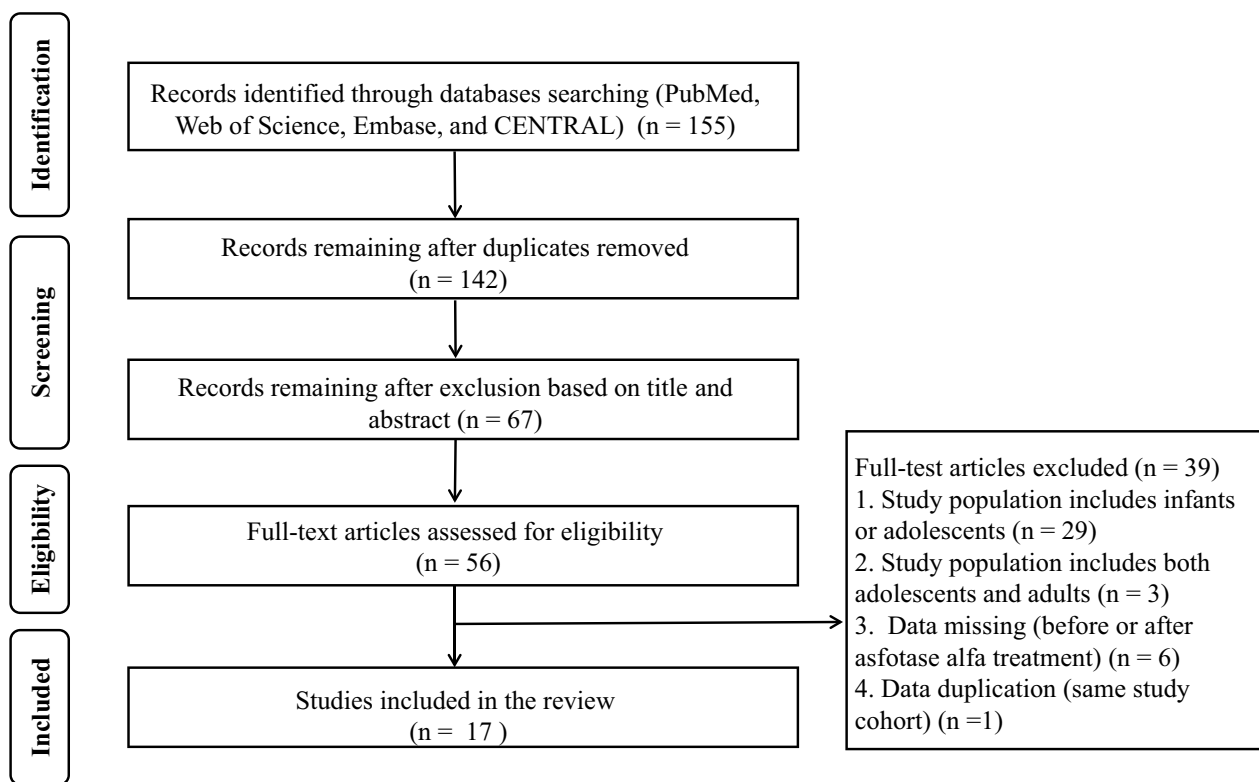


Fig. 4 Flow diagram of study selection process. CENTRAL The Cochrane Central Register of Controlled Trials

no signs of lipodystrophy, hypersensitivity reactions, or abnormalities in liver and kidney function.

Table 2 Summaries of characteristic of adult patients with hypophosphatasia by literature review

	Adult patients with hypophosphatasia (N = 295)
Age (mean ± SD), year	45.14 ± 16.68
Female, % (n)	65.4% (193)
Male, % (n)	34.6% (102)
Form of HPP, % (n)	
Pediatric-onset	91.5% (270)
Adult-onset	8.5% (25)
Symptom, % (n)	
Bony pain	79.7% (235)
Fractures history	30.8% (91)
Muscle weakness	49.4% (146)
Dental	71.9% (212)
Neurologic	17.6% (52)
Gene classification, % (n)	
Heterozygous	92.2% (272)
Homozygous	2.7% (8)
Unknown/missing	5.1% (15)

Discussion

We diagnosed with and treated a female adult patient with HPP who had suffered from bone pain, bilateral femoral fractures, and loss of mobility. Six months of teriparatide treatment failed to relieve her bone pain or promote fracture healing. After 10 months of AA treatment, she achieved fracture healing and regained the ability to walk. As AA has not yet been approved in China, this study presents the first report on the effectiveness of AA treatment in adult patients with HPP in the country, providing valuable clinical experience in AA replacement therapy for this patient group. Through a literature review, we found that adult patients with HPP typically present with bone hypomineralization, severe bone pain, recurrent and non-healing fractures, and poor dental health. Laboratory tests showed extremely low ALP levels and elevated serum calcium and phosphate levels. AA treatment can effectively increase serum ALP levels, reduce TNSALP enzyme substrates, promote fracture healing, relieve bone pain, enhance mobility, and improve the quality of life in adult patients with HPP.

Table 3 Summaries of adult patients with hypophosphatasia before and after receiving AA treatment by literature review

	n	Before AA treatment	After AA treatment	P value
Serum alkaline phosphatase (U/L)	75	11.57 ± 6.35	5873.00 ± 2787.62	0.001
Serum P-pyridoxal-5-phosphate (mmol/L)	26	545.00 ± 87.47	13.87 ± 5.95	0.008
Serum phosphorus (mmol/L)	27	1.68 ± 0.18	1.36 ± 0.07	0.024
Serum calcium (mmol/L)	27	2.39 ± 0.09	2.33 ± 0.09	0.038
parathyroid hormone (pg/ml)	26	30.60 ± 8.98	51.00 ± 14.19	0.005
type 1 N-terminal propeptide	24	47.31 ± 17.45	91.29 ± 15.06	0.009
C-terminal telopeptide of type I collagen (ng/mL)	24	0.37 ± 0.09	0.72 ± 0.24	0.13
Fracture healing	–	–	10/11	–
Bony pain alleviated	–	–	65/78	–
Mobility enhanced	–	–	61/62	–
Quality of life improved	–	–	69/74	–

AA asfotase alfa

HPP is a genetic bone disease caused by the *ALPL* gene mutations, which encodes TNSALP [34, 35]. TNSALP is expressed in multiple tissues and plays a critical role in hydrolyzing PPi, which facilitates bone mineralization through the formation of hydroxyapatite when combined with calcium [36]. When mutations occur in the *ALPL* gene, TNSALP activity is reduced, resulting in the accumulation of its substrates, including PPi, PLP, and PEA [10, 37]. PPi inhibits bone mineralization by osteoblasts and chondrocytes, and its accumulation disrupts the mineralization of hydroxyapatite by impairing calcium and phosphate integration, leading to osteoid accumulation, a hallmark of rickets and osteomalacia [14, 38].

AA is the first pharmacological treatment for HPP by bone-targeted enzyme-replacement therapy [29]. AA is a soluble human recombinant TNSALP fusion protein (726 amino acids, homodimer) comprising the catalytic domain of human TNSALP, the human IgG Fc domain, and a deca-aspartate bone-targeting peptide [39, 40]. It is approved for paediatric-onset HPP and can rapidly improve bone mineralization, pulmonary function, motor function, cognitive development, catch-up height-gain, muscle strength and daily activity ability [41–44]. In adult HPP patients, AA effectively reduces TNSALP substrate levels and markedly improves motor function and health-related quality of life (HRQoL) [45]. AA treatment can also improve fracture healing, and enhance walking ability and reduce bone pain [23].

AA is generally well-tolerated, with most adverse events being mild to moderate in severity. The most frequent adverse reactions are injection site reactions, including erythema, discoloration, pain, pruritus, swelling, induration, macule, bruising, and nodules among others [46]. In addition, most patients exhibit lipodystrophy at the injection site, primarily characterized by faint initial signs of soft tissue distension during the

first 3 months of treatment, including bulging of subcutaneous fat tissue suggesting lipohypertrophy. Upon palpation, no bulky fat masses were identified, instead, there was sagging of the skin, indicating dystrophy of the subcutaneous fat tissue. As long-term adverse effects, patients may experience ectopic calcification in the eyes and kidneys. However, it is currently believed that this ectopic calcification may also result from the HPP disease itself.

A second-generation TNSALP enzyme replacement therapy, efzimfotase alfa, has been developed for the treatment of HPP. It hydrolyzes PPi at approximately twice the rate of AA and supports once-weekly injections. A phase 1 study demonstrated that efzimfotase alfa showed acceptable safety, tolerability, and PK profiles, and resulted in dose-dependent reductions in plasma levels of TNSALP substrates, including PPi and PLP, in adults with HPP [47]. This may be a safe and effective treatment option for HPP patients in the future.

HPP is a heritable disorder caused by pathogenic *ALPL* variants, with over 450 *ALPL* variants and 850 genotypes reported. Severe HPP forms often result from homozygosity or compound heterozygosity, moderate forms from missense variant dominant negative effects, and milder forms from haploinsufficiency [48, 49]. With advances in genome-editing technology, a study using transcription activator-like effector nucleases (TALENs) corrected the c.1559delT mutation, resulting in the recovery of ALP activity in vitro [50]. Another study showed that a single injection of an adeno-associated virus vector serotype 8 harboring TNSALP-D₁₀ effectively improved the long bone phenotype in adult HPP mice [51]. These preclinical studies suggested that gene editing may have important prospects for HPP treatment.

We prospectively observed the effects of AA on bone pain and fracture nonunion for the first time in adult

patients with HPP. AA treatment significantly improved the healing of bilateral fractures, relieved bone pain, and enhanced the patient's mobility. However, there were some limitations to this study. First, we did not measure the concentrations of TNSALP substrates, such as PLP and PPi, during AA therapy. Second, we did not quantify the physical function, pain, or quality of life in the HPP patient. Third, research in the literature on the efficacy of AA in adult HPP patients is extremely limited, which restricts our comprehensive understanding of its effects in this population. Therefore, it is necessary to conduct prospective long-term studies on a large sample of HPP patients receiving AA treatment.

Conclusion

Nonunion of fractures is a serious complication of adult HPP patients. AA treatment can significantly promote fracture healing, alleviate bone pain and improve mobility. The second-generation enzyme replacement therapy shows promising potential. In the future, gene editing therapy is worth further research in HPP patients.

Abbreviations

HPP	Hypophosphatasia
AA	Asfotase alfa
ALP	Alkaline phosphatase
β-CTX	β-isomerized C-terminal telopeptide of type I collagen
P1NP	Procollagen type 1 amino-terminal peptide
BMD	Bone mineral density
TNSALP	The tissue nonspecific isoenzyme of alkaline phosphatase
PLP	Pyridoxal 5'-phosphate
PEA	Phosphoethanolamine
PPi	Pyrophosphate
ALT	Alanine aminotransferase
Cr	Creatinine
PTH	Parathyroid hormone
25-OHD	25 Hydroxyvitamin D
FT3	Free triiodothyronine
FT4	Free thyroxine
TSH	Thyroid-stimulating hormone
CEA	Carcinoembryonic antigen
IFE	Immunofixation electrophoresis
DXA	Dual-energy X-ray absorptiometry
BMD	Areal bone mineral density
LS	Lumbar spine
FN	Femoral neck
TH	Total hip
CENTRAL	The Cochrane Central Register of Controlled Trials
IQR	Interquartile range
ISRs	Injection site reactions
HRQoL	Health-related quality of life

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Author contributions

Mei Li contributed to the conceptualization and design of the research, revising the manuscript. Songqi Wang collected the data, analyzed the data, and drafted the manuscript. Lei Sun, Jing Hu, Qian Zhang, Ou Wang, Yan Jiang, Weibo Xia and Xiaoping Xing contributed to collecting data. All the authors contributed to revising the final version of the manuscript.

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

The study protocol was approved by the scientific ethics committee of PUMCH. The patient signed informed consent before participating in this study.

Consent for publication

This study contains anonymized patient data. No direct patient identifiers have been provided in this manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

- Villa-Suarez JM, Garcia-Fontana C, Andujar-Vera F, et al. Hypophosphatasia: a unique disorder of bone mineralization. *Int J Mol Sci*. 2021. <https://doi.org/10.3390/ijms22094303>.
- Yu Y, Rong K, Yao D, et al. The structural pathology for hypophosphatasia caused by malfunctioned tissue non-specific alkaline phosphatase. *Nat Commun*. 2023;14(1):4048. <https://doi.org/10.1038/s41467-023-39833-3>.
- Whyte MP. Hypophosphatasia: aetiology, nosology, pathogenesis, diagnosis and treatment. *Nat Rev Endocrinol*. 2016;12(4):233–46. <https://doi.org/10.1038/nrendo.2016.14>.
- Mornet E. Hypophosphatasia. *Metabolism*. 2018;82:142–55. <https://doi.org/10.1016/j.metabol.2017.08.013>.
- Dahir KM, Kishnani PS, Martos-Moreno GA, et al. Impact of muscular symptoms and/or pain on disease characteristics, disability, and quality of life in adult patients with hypophosphatasia: a cross-sectional analysis from the Global HPP Registry. *Front Endocrinol (Lausanne)*. 2023;14:1138599. <https://doi.org/10.3389/fendo.2023.1138599>.
- Whyte MP, Madson KL, Phillips D, et al. Asfotase alfa therapy for children with hypophosphatasia. *JCI Insight*. 2016;1(9):e85971. <https://doi.org/10.1172/jci.insight.85971>.
- Inoue D. Diagnosis and treatment of adult hypophosphatasia: Still a big challenge? *Osteoporos Sarcopenia*. 2024;10(1):1–2. <https://doi.org/10.1016/j.afos.2024.03.002>.
- Conti F, Ciullini L, Pugliese G. Hypophosphatasia: clinical manifestation and burden of disease in adult patients. *Clin Cases Miner Bone Metab*. 2017;14(2):230–4. <https://doi.org/10.11138/ccmbm.2017.14.1.230>.
- Khan AA, Brandi ML, Rush ET, et al. Hypophosphatasia diagnosis: current state of the art and proposed diagnostic criteria for children and adults. *Osteoporos Int*. 2024;35(3):431–8. <https://doi.org/10.1007/s00198-023-06844-1>.
- Kishnani PS, Rush ET, Arundel P, et al. Monitoring guidance for patients with hypophosphatasia treated with asfotase alfa. *Mol Genet Metab*. 2017;122(1–2):4–17. <https://doi.org/10.1016/j.ymgme.2017.07.010>.
- Shapiro JR, Lewiecki EM. Hypophosphatasia in adults: clinical assessment and treatment considerations. *J Bone Miner Res*. 2017;32(10):1977–80. <https://doi.org/10.1002/jbmr.3226>.
- Brandi ML, Khan AA, Rush ET, et al. The challenge of hypophosphatasia diagnosis in adults: results from the HPP International Working Group Literature Surveillance. *Osteoporos Int*. 2024;35(3):439–49. <https://doi.org/10.1007/s00198-023-06859-8>.

13. Remde H, Cooper MS, Quinkler M. Successful asfotase alfa treatment in an adult dialysis patient with childhood-onset hypophosphatasia. *J Endocr Soc*. 2017;1(9):1188–93. <https://doi.org/10.1210/js.2017-00307>.
14. Freitas TQ, Franco AS, Pereira RMR. Improvement of bone microarchitecture parameters after 12 months of treatment with asfotase alfa in adult patient with hypophosphatasia: Case report. *Medicine (Baltimore)*. 2018;97(48):e13210. <https://doi.org/10.1097/MD.00000000000013210>.
15. Schindeler A, Ludwig K, Munns CF. Enzyme replacement therapy for hypophosphatasia-The current paradigm. *Clin Endocrinol (Oxf)*. 2024;101(6):593–601. <https://doi.org/10.1111/cen.15063>.
16. Martos-Moreno GA, Rockman-Greenberg C, Ozono K, et al. Clinical profiles of children with hypophosphatasia prior to treatment with enzyme replacement therapy: an observational analysis from the global HPP registry. *Horm Res Paediatr*. 2024;97(3):233–42. <https://doi.org/10.1159/000531865>.
17. Dahir KM, Seefried L, Kishnani PS, et al. Clinical profiles of treated and untreated adults with hypophosphatasia in the Global HPP Registry. *Orphanet J Rare Dis*. 2022;17(1):277. <https://doi.org/10.1186/s13023-022-02393-8>.
18. Klidas P, Severt J, Aggers D, Payne J, Miller PD, Ing SW. Fracture healing in two adult patients with hypophosphatasia after asfotase alfa therapy. *JBMR Plus*. 2018;2(5):304–7. <https://doi.org/10.1002/jbm4.10052>.
19. Dahir KM, Ing SW, Deal C, Messali A, Bates T, Rush ET. Improvement in quality of life after asfotase alfa treatment in adults with pediatric-onset hypophosphatasia: data from 5 patient-reported outcome measures. *JBMR Plus*. 2024;8(8):ziae062. <https://doi.org/10.1093/jbmpr/ziae062>.
20. Seefried L, Rak D, Petryk A, Genest F. Bone turnover and mineral metabolism in adult patients with hypophosphatasia treated with asfotase alfa. *Osteoporos Int*. 2021;32(12):2505–13. <https://doi.org/10.1007/s00198-021-06025-y>.
21. Nishizawa H, Sato Y, Ishikawa M, et al. Marked motor function improvement in a 32-year-old woman with childhood-onset hypophosphatasia by asfotase alfa therapy: evaluation based on standardized testing batteries used in Duchenne muscular dystrophy clinical trials. *Mol Genet Metab Rep*. 2020;25: 100643. <https://doi.org/10.1016/j.ymgmr.2020.100643>.
22. Whyte MP, McAlister WH, Mumm S, Bierhals AJ. No vascular calcification on cardiac computed tomography spanning asfotase alfa treatment for an elderly woman with hypophosphatasia. *Bone*. 2019;122:231–6. <https://doi.org/10.1016/j.bone.2019.02.025>.
23. Kato H, Hidaka N, Koga M, et al. Radiological evaluation of pseudofracture after the administration of asfotase alfa in an adult with benign prenatal hypophosphatasia: a case report. *Bone Rep*. 2022;16: 101163. <https://doi.org/10.1016/j.bonr.2021.101163>.
24. Sturznickel J, Schmidt FN, von Vopelius E, et al. Bone healing and reactivation of remodeling under asfotase alfa therapy in adult patients with pediatric-onset hypophosphatasia. *Bone*. 2021;143: 115794. <https://doi.org/10.1016/j.bone.2020.115794>.
25. Hidaka N, Murata H, Tachikawa K, et al. The effect of asfotase alfa on plasma and urine pyrophosphate levels and pseudofractures in a patient with adult-onset hypophosphatasia. *JBMR Plus*. 2023;7(12): e10842. <https://doi.org/10.1002/jbm4.10842>.
26. Rolvien T, Schmidt T, Schmidt FN, et al. Recovery of bone mineralization and quality during asfotase alfa treatment in an adult patient with infantile-onset hypophosphatasia. *Bone*. 2019;127:67–74. <https://doi.org/10.1016/j.bone.2019.05.036>.
27. Koyama H, Yasuda S, Kakoi S, et al. Effect of asfotase alfa on muscle weakness in a Japanese adult patient of hypophosphatasia with low ALP levels. *Intern Med*. 2020;59(6):811–5. <https://doi.org/10.2169/internalmedicine.3298-19>.
28. Magdaleno AL, Singh S, Venkataraman S, Perilli GA, Lee YY. Adult-onset hypophosphatasia: before and after treatment with asfotase alfa. *AACE Clin Case Rep Nov-Dec*. 2019;5(6):e344–8. <https://doi.org/10.4158/ACCR-2019-0143>.
29. Alsarraf F, Ali DS, Almonaie K, Al-Alwani H, Khan AA, Brandi ML. Hypophosphatasia: presentation and response to asfotase alfa. *Osteoporos Int*. 2024;35(4):717–25. <https://doi.org/10.1007/s00198-023-06943-z>.
30. Kishnani PS, Martos-Moreno GA, Linglart A, et al. Effectiveness of asfotase alfa for treatment of adults with hypophosphatasia: results from a global registry. *Orphanet J Rare Dis*. 2024;19(1):109. <https://doi.org/10.1186/s13023-024-03048-6>.
31. Seefried L, Kishnani PS, Moseley S, et al. Pharmacodynamics of asfotase alfa in adults with pediatric-onset hypophosphatasia. *Bone*. 2021;142: 115664. <https://doi.org/10.1016/j.bone.2020.115664>.
32. Genest F, Rak D, Petryk A, Seefried L. Physical function and health-related quality of life in adults treated with asfotase alfa for pediatric-onset hypophosphatasia. *JBMR Plus*. 2020;4(9): e10395. <https://doi.org/10.1002/jbm4.10395>.
33. Seefried L, Genest F, Petryk A, Veith M. Effects of asfotase alfa in adults with pediatric-onset hypophosphatasia over 24 months of treatment. *Bone*. 2023;175: 116856. <https://doi.org/10.1016/j.bone.2023.116856>.
34. Charoenngam N, Nasr A, Shirvani A, Holick MF. Hereditary metabolic bone diseases: a review of pathogenesis, diagnosis and management. *Genes (Basel)*. 2022. <https://doi.org/10.3390/genes13101880>.
35. Fenn JS, Lorde N, Ward JM, Borovickova I. Hypophosphatasia. *J Clin Pathol*. 2021;74(10):635–40. <https://doi.org/10.1136/jclinpath-2021-207426>.
36. Michigami T, Tachikawa K, Yamazaki M, Kawai M, Kubota T, Ozono K. Hypophosphatasia in Japan: ALPL mutation analysis in 98 unrelated patients. *Calcif Tissue Int*. 2020;106(3):221–31. <https://doi.org/10.1007/s00223-019-00626-w>.
37. Komaru K, Ishida-Okumura Y, Numa-Kinjoh N, Hasegawa T, Oda K. Molecular and cellular basis of hypophosphatasia. *J Oral Biosci*. 2019;61(3):141–8. <https://doi.org/10.1016/j.job.2019.07.003>.
38. Whyte MP. Hypophosphatasia: an overview. *For 2017*. *Bone*. 2017;102:15–25. <https://doi.org/10.1016/j.bone.2017.02.011>.
39. Jaswanthi N, Sindhu R, Nimmy P, et al. Effect of asfotase alfa in the treatment of hypophosphatasia: a systematic review. *J Pharm Bioallied Sci*. 2023;15(Suppl 1):S101–4. https://doi.org/10.4103/jpbs.jpbs_662_22.
40. Whyte MP. Hypophosphatasia: enzyme replacement therapy brings new opportunities and new challenges. *J Bone Miner Res*. 2017;32(4):667–75. <https://doi.org/10.1002/jbm.3075>.
41. Schroth RJ, Long C, Lee VH, Alai-Towfigh H, Rockman-Greenberg C. Dental outcomes for children receiving asfotase alfa for hypophosphatasia. *Bone*. 2021;152: 116089. <https://doi.org/10.1016/j.bone.2021.116089>.
42. Hofmann CE, Harmatz P, Vockley J, et al. Efficacy and safety of asfotase alfa in infants and young children with hypophosphatasia: a phase 2 open-label study. *J Clin Endocrinol Metab*. 2019;104(7):2735–47. <https://doi.org/10.1210/clinem.2018-02335>.
43. Whyte MP, Simmons JH, Moseley S, et al. Asfotase alfa for infants and young children with hypophosphatasia: 7 year outcomes of a single-arm, open-label, phase 2 extension trial. *Lancet Diabetes Endocrinol*. 2019;7(2):93–105. [https://doi.org/10.1016/S2213-8587\(18\)30307-3](https://doi.org/10.1016/S2213-8587(18)30307-3).
44. Padidela R. Asfotase alfa treatment in perinatal and infantile hypophosphatasia: safe and sustained efficacy. *Lancet Diabetes Endocrinol*. 2019;7(2):76–8. [https://doi.org/10.1016/S2213-8587\(18\)30321-8](https://doi.org/10.1016/S2213-8587(18)30321-8).
45. Simon S, Resch H. Treatment of hypophosphatasia. *Wien Med Wochenschr*. 2020;170(5–6):112–5. <https://doi.org/10.1007/s10354-020-00736-3>.
46. Shirinezhad A, Esmaili S, Azarboo A, et al. Efficacy and safety of asfotase alfa in patients with hypophosphatasia: a systematic review. *Bone*. 2024;188: 117219. <https://doi.org/10.1016/j.bone.2024.117219>.
47. Dahir KM, Shannon A, Dunn D, et al. Safety, pharmacokinetics, and pharmacodynamics of efzimfotase alfa, a second-generation enzyme replacement therapy: phase 1, dose-escalation study in adults with hypophosphatasia. *J Bone Miner Res*. 2024;39(10):1412–23. <https://doi.org/10.1093/jbmpr/zjae128>.
48. Allen MR. Recent advances in understanding bisphosphonate effects on bone mechanical properties. *Curr Osteoporos Rep*. 2018;16(2):198–204. <https://doi.org/10.1007/s11914-018-0430-3>.
49. Xiao F, Zhou Z, Song X, et al. Dissecting mutational allosteric effects in alkaline phosphatases associated with different Hypophosphatasia phenotypes: an integrative computational investigation. *PLoS Comput Biol*. 2022;18(3): e1010009. <https://doi.org/10.1371/journal.pcbi.1010009>.
50. Nakano C, Kitabatake Y, Takeyari S, et al. Genetic correction of induced pluripotent stem cells mediated by transcription activator-like effector nucleases targeting ALPL recovers enzyme activity and calcification in vitro. *Mol Genet Metab*. 2019;127(2):158–65. <https://doi.org/10.1016/j.ymgme.2019.05.014>.
51. Amadeu de Oliveira F, Mohamed FF, Kinoshita Y, et al. Gene therapy using recombinant AAV Type 8 vector encoding TNAP-D(10) improves the skeletal phenotypes in murine models of osteomalacia. *JBMR Plus*. 2023;7(1):e10709. <https://doi.org/10.1002/jbm4.10709>.

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