

Occult HCV Infection (OCI) Diagnosis in Cirrhotic and Non-cirrhotic Naïve Patients by Intra-PBMC Nested Viral RNA PCR

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Abstract

Background and Aims: Occult HCV infections (OCIs) include IgG antibody seronegative cryptogenic (COCIs), as well as seropositive secondary naïve (SNOCIs) and experienced (SEOCIs) cases. We used peripheral-blood-mononuclear-cell (PBMC)-PCR to evaluate COCIs and SNOCIs prevalence, serum HCV spontaneous disappearance (SCSD) in naïve cirrhotics and non-cirrhotics, intra-PBMC HCV-RNA strands in relation to cirrhosis density in naïve non-viremia cases, and HCV-RNA seroconversion after 1 year of solitary naïve intra-PBMC infection. *Methods:* The anti-HCV IgG antibodypositive naïve-patients (n = 785) were classified into viremic (n = 673) and non-viremic [n = 112, including non-cirrhotics (n = 55) and cirrhotics (n = 57)], and 62 controls without evidence of HCV-infection. Controls and post-HCV non-viremia cases (n = 62+112 = 174) were submitted to hepatic Fibroscan-Elastography evaluation. All subjects (n = 847) were screened for intra-PBMC HCV-RNA sense and antisense strands by nested-PCR. Results: Naïve-OCI cases (4.84%) that were diagnosed by PBMC-PCR significantly raised the total numbers of HCV-infection to 714 (p = 0.01). The percent positivity of SNOCIs (34.82%) was significantly higher than for asymptomatic-COCIs (3.125%, p = 0.0001). Comparing PBMC-PCR with single-step-reverse-transcription (SRT)-PCR for identification of SCSD in naïve IgG antibody-positive nonviremia patients (n = 112) revealed a decline in SCSD prevalence by PBMC-PCR (from 14.27% to 9.3%), regardless of presence of hepatic cirrhosis (p = 0.03). SCSD was found to be higher by PBMC-PCR in non-cirrhotics compared to cirrhotics (p = 0.0001), with an insignificant difference when using SRT-PCR (p = 0.45). Intra-PBMC HCV-RNA infection was significantly more frequent in cirrhotics compared to both non-cirrhotics and controls (p < 0.0005). An increased

hepatic fibrosis density was recognized in intra-PBMC HCV-RNA infection with sense (p = 0.0001) or antisense strand (p = 0.003). HCV-RNA seroconversion was associated with intra-PBMC infection when both sense and antisense strands were detected (p = 0.047). **Conclusions:** Intracellular HCV-RNA evaluation is crucial for diagnosing OCIs and addressing relapse probability.

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Introduction

In 2004, Pham *et al.*¹ reported occult HCV infections (OCIs) in anti-HCV IgG antibody-positive naïve patients who had spontaneous recovery after self-limited HCV-infections and in experienced cases with sustained virologic response (SVR) after interferon treatment. During the same year, Castillo *et al.*² described HCV-RNA existence in anti-HCV IgG antibodynegative naïve patients who presented with active hepatocellular damage. Further refinement of the definitions of OCIs included: a) recognition of intracellular RNA strands in postinfection non-viremia naïve cases, which is termed spontaneous HCV disappearance (SCSD) and manifests as secondary naïve OCIs (SNOCIs); b) identification of post-treatment non-viremia cases, which have intracellular HCV-RNA strand infections, and manifest as secondary experienced OCIs (SEOCIs).^{3,4}

Recently, baseline diagnosis and post-treatment follow-up of both SNOCIs and SEOCIs by peripheral blood mononuclear cell (PBMC) HCV-PCR in Egyptian populations addressed both validation of PBMC-PCR as a diagnostic test that can diagnose intracellular HCV when single-step reverse transcription (SRT)-PCR is negative and the association of intra-PBMC infections with liver cirrhosis in naïve non-viremia patients. The authors recommended eradication of intra-PBMC RNAstrands in SEOCIs to avoid HCV-RNA seroconversion and subsequent post-treatment relapse.⁵ A third category of OCIs is known as cryptogenic occult HCV-infection (COCIs), which is diagnosed in the IgG-seronegative population by detecting the intracellular RNA strands. Its prevalence is around 3.5% in asymptomatic populations,³ which might

Keywords: OCIs; PBMCs; Naïve; Cirrhosis.

Abbreviations: COCIs, cryptogenic occult HCV infection; DAA, direct-acting antiviral; HCC, hepatocellular carcinoma; HCV, hepatitis C virus; OCIs, occult HCV infections; PBMCs, peripheral blood mononuclear cells; RAVs, resistance-associated variants; SCSD, serum HCV spontaneous disappearance; SEOCIs, secondary experienced occult HCV infections; SNOCIs, secondary naïve occult HCV infection; SRT, single-step reverse transcription; SVR, sustained virologic response.

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have active replication of HCV inside PBMCs, as identified by detection of either the viral NS3-protein or antisense-strand.⁶

In 1999, El-Awady *et al.*⁷ predicted that assay for viral replication in PBMCs enhances sensitivity of diagnosing and monitoring HCV associated hepatitis. Accordingly, intra-PBMC detection of HCV-RNA by PCR is currently used to diagnose OCIs in non-viremia patients.⁵ It is obvious that the immune system is not only a part of an OCI patient's reaction to the infection but that it also serves as a host to the viral RNA, thereby facilitating its replication, as indicated by antisense HCV-RNA strands found in the PBMCs of these patients.⁸ Keeping in mind that HCV-infection is an important and treatable cause of liver disease and the fact that patients with cryptogenic liver diseases can potentially develop cirrhosis and hepatocellular carcinoma, investigating the prevalence of HCV and developing more accurate techniques to diagnose OCIs in these patients are critically important.⁹

Some researchers¹⁰ have expressed doubts about using SRT-PCR in post-treatment follow-up. They have concluded that HCV cure in individuals achieving a state of undetectable viremia after an 8 to 12-week course of direct-acting antiviral (DAA) therapy may not be entirely valid. The same researchers recommended careful longitudinal follow-up utilizing highly sensitive assays and unique approaches to viral detection. Because PBMCs present a convenient extrahepatic home for HCV adoption, translation, transcription, assembly and finally release into the serum and/or other adjacent cells,⁶ cellular PCR can be used to further refine the definitions of clinical presentations of spontaneous HCV clearance. Recognition of intra-PBMC HCV strands in SNOCIs⁵ should refer to regression of using the 'spontaneous HCV-clearance' term. In populations of HCV endemic areas, the SCSD is estimated to be around 25%.¹¹ However, cellular HCV spontaneous disappearance has not been evaluated before the current report.

On the other hand, the post-treatment RNA genomic seroconversion has been attributed to either treatment failure or new infection, and is known as HCV-relapse, ¹² which has not been described in naïve subjects who present with spontaneous viral clearance because no temporal follow-up is recommended. Till now, naïve cirrhotic patients who present with nonviremia, but have evidence of hepatocellular damage and positive intra-PBMC HCV-RNA strand infection, have not been considered as candidates for DAA therapy.² However, eradication of intra-PBMC HCV-infection is recommended in naïve cirrhotics and experienced post-treatment patients, respectively, because of association with cirrhotic changes and probability of relapse in more than 18%.⁵

Considering the high cure rate of DAA therapy, physicians are looking for effective treatments for COCIs and SNOCIs. Patients with OCIs should be afforded the option of getting rid of the intracellular HCV genomic materials, to prevent the subsequent hepatic fibrosis.⁹ For this reason, the current study was designed to shed more light on the OCI problem among a population of cases from one of the most endemic areas in the world for HCV-infection, in the hopes of promoting attention to this unmet challenge to satisfy clinical practice demands.

The specific aims of the current study included the use of PBMC-PCR in the evaluation of a) prevalence of asymptomatic COCIs and SNOCIs, b) SCSD in naïve cirrhotics and noncirrhotics, c) intra-PBMC HCV-RNA sense and antisense strand infections in relation to cirrhosis density in naïve non-viremia cases, and d) rate of HCV-RNA seroconversion after 1 year of solitary naïve intra-PBMC infection.

Methods

Study subjects

All study subjects (n = 847) were treatment-naïve and outpatient-visitors to the Infectious Disease Clinic. All subjects were screened by SRT-PCR between January 2015 and February 2017. Inclusion criteria were age between 18 and 70 years-old and positivity for serum anti-HCV IgG antibodies; controls were recruited from the population that was clinically and serologically free of HCV. Exclusion criteria included hepatocellular carcinoma (HCC) and Child C classification. Ethical committee approval for the study was obtained before patient enrollment (Registration No. 10231, National Research Center).

Sample size in each group depended upon availability of subjects that fulfill the inclusion criteria during the study period. Study subjects (n = 847) included the following subgroups: anti-HCV IgG antibody-positive naïve patients (n = 785) who were classified into chronic HCV-viremic (n = 673) and post-HCV non-viremic cases (n = 112), with the last group further divided into non-cirrhotic (n = 55) and cirrhotic (n = 57) subgroups. Controls included 62 participants without evidence of HCV-infection. Controls and post-HCV non-viremia cases (n = 174) submitted to hepatic elastography evaluation by Fibroscan. All subjects (n = 847) were screened for presence of intra-PBMC HCV-RNA sense and antisense strands by nested-PCR.

ELISA for HCV IgG antibody detection

The procedure was performed as described.¹³ The thirdgeneration ELISA contained reconfigured NS3 and core antigens and in addition a newly incorporated antigen from the NS5 region. This assay was used as a preliminary screening test to enroll cases in the current study.

Real-time PCR for quantification of HCV RNA

The collection and transportation of specimens, RNA isolation, SRT-PCR procedure, internal control of the isolated RNA and/ or contamination, and quantification of the HCV PCR were all performed as described by Abd Alla and El-Awady⁵ in 2017.

Amplification of intracellular HCV RNA genomes by strand-specific RT-PCR⁵

Extraction of RNA from PBMCs

Peripheral blood (200 μ L) was diluted with 10 mL freshly prepared red blood cell samples in alkaline buffer (38.8 mmol/L NH₄Cl, 2.5 mmol/L K₂HCO₃, 1 mmol/L EDTA; pH 8.0). After 10 min incubation at room temperature, nucleated cells were washed with the same buffer, the cells were dissolved in 500 μ L anti nuclease solution (4 mol/L guanidinium isothiocyanate containing 25 mmol/L sodium citrates, 0.5 sarcosyl and 0.1 mol/L β-mercaptoethanol). A single-step method described by¹⁴ and modified by^{15,16} followed to carry out the RNA extraction.

Retrotranscription PCR of sense and antisense strands of HCV RNA

Detection of HCV RNA strands in PBMCs was performed as described by Lohr *et al.*¹⁷ The sequences of primers used in

this study are: 1CH, 5'ggt gca cgg tct acg aga cct c3'; 2CH, 5' aac tca tgt ctt cac gca gaa3'; P2, 5'tgc tca tgg tgc acg gtc ta3'; P3, 5'ctt tcg cga ccc aac act ac3'; and P4, 5'aga gcc ata gtg gtc tgc gg3'. The other steps for the retrotranscription PCR procedure for sense and antisense strands of HCV RNA were done as previously reported by Abd Alla and El Awady.⁵

Elastography Fibroscan

The procedure was carried out as previously described.¹⁸ Briefly, patients were placed in supine position, with their right arm in maximal abduction. All measurements were performed in the right lobe of the liver through the costal space. The tip of the probe transducer was covered with coupling gel and placed on the skin between the ribs at the right liver lobe level. The operator, assisted by ultrasound time motion and a mode images provided by the system, located a portion of the liver that was at least 6 cm thick and free of any large vascular structures. Once the area of measurement was located, the operator pressed the probe button to begin image acquisition. The measurement depth ranged from 25 mm to 45 mm, and 10 valid measurements were obtained for each patient. The results were expressed in units of kilopascal (kPa). All the non-viremic subjects underwent the examination with a normal prob. The success rate¹⁹ of the examination was calculated as the ratio between the number of measurements validated by the machine and the total number of attempted measurements. The liver stiffness corresponds to the median value of the validated measurements. The interguartile range (IQR) was used as the interval around the median that contained 50% of the valid measurements. To be considered interpretable and valid, the examination must have included at least 10 measurements, with a SR of at least 66%; and, the IQR must not have exceeded 33% of the results of the examination.

Statistical analysis

The diagnostic procedure (PBMC-PCR) used was compared to the SRT-PCR (gold standard test) in a cross-sectional study that contained two groups of selected subjects. All cases that fit the inclusion criteria during 25 consecutive month period were included in the study. All tests performed were two-sided and statistical significance was considered at a *p*-value of 0.05. SSPS version 9.0 for Windows (Chicago, Illinois, USA) was used in the data analysis. We compared the mean results by the Student's *t*-test (for variables with normal distribution) or by the Mann-Whitney U test (for variables with non-normal distribution). Categorical variables were compared using the chi-square or Fisher's exact tests. To study the existence of correlation between the variables, Pearson's (for variables with Gaussian distribution) or Spearman's (for non-normally distributed variables) correlation coefficients were determined.

Results

Advantages of combined screening with both SRT-PCR and PBMC-PCR

As illustrated in Figure 1, both SRT-PCR and PBMC-PCR were used in screening of study populations with (Fig. 1A and 1B) and without (Fig. 1C) controls for HCV-infection. All cases which tested positive for HCV-infection by SRT-PCR (n = 673/847, 79.46%; Fig. 1A&B and 673/785, 85.733%; Fig. 1C) were also positive for intra-PBMC HCV RNA strands. Patients who presented with non-viremia, as identified by SRT-PCR, consisted of anti-HCV IgG antibody negative controls (n = 62) and IgG antibody-positive naïve patients (n = 112).

Testing all RNA-seronegative subjects (controls n = 62 + IgG-seropositive n = 112) by cellular-PCR for intra-PBMC HCV-infection revealed 60/847 (7.1%) of negative controls, 73/847 (8.62%) of negative naïve post-HCV infection patients, and 41 (4.84%) patients positive for intra-PBMC RNA strands, as demonstrated in Figure 1B. Adding the extra positive patients (n = 41) who presented with solitary intra-PBMC HCV-RNA strand infection to the baseline RNA-seropositive patients (n = 673/847, 79.46%) significantly raised the overall percent positive patients for HCV-infection

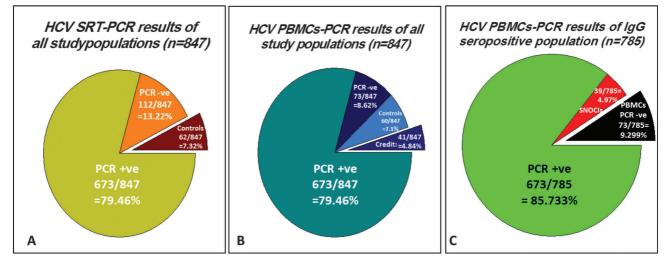


Fig. 1. Advantages of combined serum and PBMCs (B and C) screening over solitary serum screening for HCV infection (A) by PCR. PBMC testing of nonviremic subjects detected an extra 41 (4.84%) and 39 (4.97%) infections upon adding (B) or subtracting (C) controls, respectively. The higher numbers of HCV diagnosis obtained by detecting intracellular RNA strands significantly raised the overall frequencies of HCV diagnosis by PBMCs compared to SRT-PCR (p = 0.011 for panel B and p = 0.0029 for panel C). As noted in panel C, the frequency of SNOCIs (39/785, 4.968%) is significantly lower than the frequency among patients with negative PCR (73/785, 9.3%) (p = 0.001).

Table 1. HCV RNA genomic detection within PBMCs compared to serum in naïve post-HCV infection

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Type of HCV genomic PCR (<i>n</i>)	HCV RNA- negative, <i>n</i> (%)	HCV RNA- positive, <i>n</i> (%)
SRT-PCR (785)	112 (14.27)	673 (85.73)
PBMC-PCR (785)	73 (9.30)	712 (90.70)
Fisher's exact 2-tailed <i>p</i>	0.00285545	

HCV diagnosis by intra-PBMCs RNA genomic detection was significantly higher compared to SRT-PCR (p = 0.0029). PBMC-PCR-based frequency of negative HCV infection among naïve patients (9.31%) was significantly lower than the SRT-PCR-based frequency (14.16%; p < 0.00001).

Table 2. COCIs and SNOCIs in anti-HCV IgG antibody-positive and RNA seronegative naïve patients compared to negative controls

Studied group (n)	COCIs, n (%)	SNOCIs, n (%)	Non-OCIs, <i>n</i> (%)
Negative controls (62)	2 (3.125)	0.0 (0.0)	60 (96.875)
IgG-positive/ SRT-PCR- negative (112)	0.0 (0.0)	39 (34.82)	73 (65.18)
Fisher's exact 2-tailed <i>p</i>	0.00000025		

As diagnosed by PBMC-PCR, the prevalence of SNOCIs in anti-HCV IgG antibodypositive patients (34.82%) is highly significant compared to the prevalence of COCIs (3.1%) in negative controls (p < 0.00001).

(n = 714/847, 84.3%; p = 0.012). Figure 1C shows that prevalence of SNOCIs in the HCV IgG antibody-seropositive population (39/785, 4.968%) was significantly lower than the frequency of RNA-seronegative PBMC-PCR (73/785, 9.3%; p = 0.001).

Table 1 demonstrates results of both SRT-PCR and PBMC-PCR regarding evaluation of naïve post-HCV RNA-seronegative and naïve RNA-seropositive infections without negative controls. Diagnosis of intra-PBMC RNA strand infection (n = 712/785, 90.7%) was significantly higher than that of SRT-PCR (673/785, 85.73%; p = 0.0029). The number of HCV infection-free cases diagnosed by SRT-PCR (112/785, 14.27%) was significantly higher than for those diagnosed by PBMC-PCR (73/785, 9.3%; p < 0.00001).

Table 2 demonstrates results of PBMC-PCR in diagnosing SNOCIs and COCIs. Prevalence of SNOCIs who presented with solitary positive anti-HCV IgG antibody in non-viremia patients (34.82%) was highly significant (higher) compared to prevalence of COCIs (3.125%) in negative controls for both IgG antibodies and viremia (p < 0.00001). Thus, SNOCIs are highly recommended for diagnostic testing and meticulous evaluation before proposal of HCV eradication.

SCSD of RNA-genomic materials

Tables 1 and 3 illustrate evaluation of SCSD as detected by SRT-PCR and PBMC-PCR in anti-HCV IgG antibody-positive naïve patients. The overall prevalence of SCSD dropped from 14.26% by SRT-PCR to 9.3% by PBMC-PCR (p =0.003), as shown in Table 1. Correlation of SCSD with hepatic cirrhosis in naïve cirrhotic cases, non-cirrhotic patients and controls is demonstrated in Table 3. SCSD of RNA genomic materials was significantly lower in post-HCV non-viremia patients regardless of cirrhosis compared to controls, as diagnosed by both SRT-PCR and PBMC-PCR (p <0.00001). Upon comparing the diagnostic yields of SRT-PCR with PBMC-PCR regarding evaluation of SCSD, the latter procedure was associated with significantly lower SCSD of RNA (p = 0.03) because of higher sensitivity. Correlation of results of SRT-PCR and PBMC-PCR with hepatic cirrhosis is illustrated in Figure 2. SCSD of HCV RNA genomic materials, as diagnosed by SRT-PCR, had insignificant difference when comparing cirrhotic patients with non-cirrhotic naïve patients (p = 0.446). On the other hand, PBMC-PCR had a highly significant difference in SCSD of HCV strands when comparing non-cirrhotic patients with cirrhotic naïve patients (p = 0.0001).

Distribution of sense and antisense strands in cirrhotic versus non-cirrhotic naïve HCV infection

As demonstrated in Table 4, naïve cirrhotic patients showed significantly increased prevalence of sense and antisense strands compared to naïve non-cirrhotic and negative controls (p < 0.001). The naïve non-cirrhotic patients, who presented with solitary positive serum anti-HCV IgG antibodies, had a significantly increased frequency of both sense and antisense strands compared to controls (p < 0.0001). On the other hand, Figure 3 shows the relationship between density of hepatic cirrhosis and the intracellular HCV RNA strands' distribution in naïve patients. Association of intra-PBMC HCV genomic materials with hepatic fibrosis was

Table 3. Comparison of PBMC-PCR with SRT-PCR in evaluation of spontaneous HCV RNA disappearance in cirrhotic and non-cirrhotic naïve patients

HCV IgG Abs and PCR based grouping (n)	Cirrhotic naïve n (%)	Non-cirrhotic Naïve n (%)	
IgG abs-negative/PCR negative Controls (62)	0.0(0.0%)	62(100%)	
IgG abs-positive/SRT-PCR negative Naïve (112)	57(50.89%)	55(49.11%)	
IgG abs-positive/PBMC-PCR negative Naïve (73)	25(34.25%)	48(65.75%)	
Fisher's exact 2-tailed <i>p</i> : A vs. B : A vs. C : B vs. C	0.00000000 0.00000001 0.03384352		

Spontaneous disappearance of HCV RNA genomic materials from PBMCs is significantly lower than in serum from post-HCV naïve patients regardless of cirrhotic changes (p = 0.03). All controls are non-cirrhotic and showed significant absence of both serum and intra-PBMC HCV RNA genomic infection than all naïve post-HCV infection (p > 0.00001).

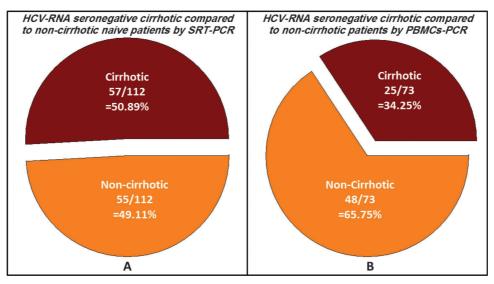


Fig. 2. SCSD of HCV RNA genomic materials from IgG antibody-positive and PCR negative populations in relation to hepatic cirrhosis. SCSD of HCV RNA as diagnosed by A) SRT-PCR and B) PBMC-PCR. A) Insignificant difference was found when cirrhotic and non-cirrhotic naïve patients were compared (Fisher's exact 1-tailed, p = 0.446). B) Significant difference was found when non-cirrhotic naïve patients were compared (Fisher's exact 1-tailed, p = 0.446). B) Significant difference was found when non-cirrhotic naïve patients were compared (Fisher's exact 1-tailed, p = 0.0001).

	PBMC-PCR			
Study subjects (n)	Antisense strand <i>n</i> (% positive)	Sense strand <i>n</i> (% positive)	Negative <i>n</i> (% positive)	
Controls (62)	0.0 (0.00)	2 (3.2300)	60 (96.77)	
Cirrhotic naïve (57)	11 (19.30)	21 (36.84)	25 (43.86)	
Non-cirrhotic naïve (55)	3 (5.45)	4 (7.270)	48 (87.28)	
Fisher's exact 2-tailed <i>p</i> : A vs. B : A vs. C : B vs. C	0.00000000 0.00000000 0.00000010	0.000000028 0.00000000 0.000371340	0.0034618535 0.000000000 0.0026551955	

Naïve cirrhotic patients have significantly higher frequencies of both sense and antisense strands compared to naïve non-cirrhotic and negative controls (p < 0.001). Naïve non-cirrhotic patients have a significantly high frequency of sense and antisense strands compared to controls (p < 0.0001).

found to be highly significant in those who have intracellular RNA strand infection in comparison to the post-hepatitis PCR negative patients (p < 0.00001 and 0.003 for intra-PBMC sense and antisense strands, respectively).

Hepatic fibrosis showed insignificant changes in distribution among the naïve HCV patients who had RNA-antisense or sense strand infections (p = 0.68). Quantitatively, the difference in hepatic fibrosis as kPa was very highly significant when HCV PCR negative, antisense and sense strand-infected patients were compared with negative controls (p <0.00001). Statistical analysis of absolute kPa values was done by using the mean values of hepatic fibrosis ± standard deviation (SD) of the illustrated groups in Figure 3. The increased means of hepatic fibrosis were significantly associated with intra-cellular infections with sense (19.17 ± 14.28) and antisense (20.8 \pm 16.29) strands compared to controls (3.34 ± 0.34) and post-HCV PCR negative patients $(7.51 \pm$ 8.28; T-value > 1.0). The mean values of hepatic fibrosis were insignificantly different for those who had intra-PBMC infection with HCV-RNA antisense strand (20.8 \pm 16.29) upon comparison with those who had infection with sense strands (19.17 \pm 14.28; T = 0.74), and on comparison of post-HCV PCR negative groups (7.51 ± 8.28) with negative controls $(3.34 \pm 0.34; T = 0.0001)$. These results confirm the statistically analyzed data that were retrieved from Figure 3.

HCV RNA seroconversion compared to spontaneous disappearance of intracellular infection

The fate of intra-PBMC HCV-RNA infection in 7 patients determined at 1-year follow-up is described in Table 5. At baseline, all 7 naïve patients were cirrhotic and had positive serum anti-HCV IgG antibodies, with 4 having both HCV strands inside their PBMCs and the other 3 having the intracellular RNA sense strand but lacking the antisense one. Follow-up of the 7 patients for 1 year by SRT-PCR and PBMC-PCR revealed the following: a) 3 out of 4 F4 patients (75%) who had positive intra-PBMC sense and antisense strands at baseline showed RNA seroconversion (p = 0.047), with the fourth patient was still positive for intracellular sense and antisense RNA strands without RNA seroconversion; b) absence of antisense strand in the other 3 naïve patients who started with positive intra-PBMC HCV-RNA sense

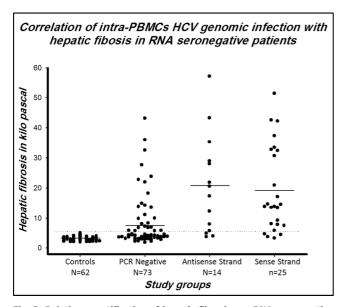


Fig. 3. Relative quantification of hepatic fibrosis per RNA seronegative subject in relation to intra-PBMC HCV-infection. Intracellular RNA was significantly associated with liver fibrosis upon comparison of currently infected groups with the post-HCV PCR negative group (Fisher's exact 2-tailed, p = 0.00001 and p = 0.0028, respectively, for intra-PBMC sense and antisense strands). Hepatic fibrosis was almost equally distributed among patients who presented with intra-PBMC HCV RNA antisense or sense strand infection (Fisher's exact 1-tailed, p = 0.68). Fisher's exact 1-tailed p < 0.00001 was found upon comparing controls with each of the other three groups. Dotted line, cutoff point; solid line, mean kPa values per group. Data cutoff point = mean+3SD of controls KPa values.

strand were associated with clearance of the intracellular infection after 1 year.

Discussion

The current study presents the advantage of using PBMC-PCR in diagnosing OCIs. PBMC-PCR added 3.125% of asymptomatic COCIs and 34.82% of SNOCIs, significantly raising the

overall number of infected cases from 673 to 712. Infections that are usually recognized in clinical practice as cases with history of HCV infection (i.e. having no viremia despite positive serum IgG-antibodies) were designated as patients with SCSD in the current study. The number of cases presented with SCSD was found to be significantly lower because of the increased sensitivity of PBMC-PCR in diagnosing SNOCIs. The association of hepatic fibrosis with SNOCIs was also significant in the current data set. Furthermore, the increased density of hepatic fibrosis is accompanied by intra-PBMC infection with either sense or antisense strands. The HCV RNA seroconversion in this cohort is attributed to intra-PBMC infection with both sense and antisense strands in cirrhotic patients.

The increased sensitivity of diagnosing dormant HCVinfection addressed by a previous study⁵ adds major advantages to the management of both SNOCIs and SEOCIs in medical practice. The appreciated high predictive values of HCV-PCRs provide a trustworthy diagnostic tool to trace extra-and intra-cellular RNA replication. The current study looked for RNA strands in PBMCs because of their considerable degrees of HCV tropism and because their nuclei are essential to viral RNA genomic replication.⁶ The solitary intracellular recognition of HCV RNA, without viremia, has been known to occur during the incubation periods or as a post-infection presentation after successful elimination of genomic materials from sera.²⁰

Spontaneous and drug-induced HCV RNA disappearance from sera have been collectively termed as sustained virologic disappearance; unfortunately, that naming convention doesn't correspond to elimination of viral genomic materials from the rest of body tissues.²¹ RNA seroconversion in both post-treatment or naïve patients is usually related to persistent intracellular infection with HCV genomic strands.^{9,11,22–25} We have recently evaluated PBMC-PCR of HCV-RNA as one of the most sensitive and specific tools to assess extrahepatic HCV infections.⁵ In current cohort, we utilized PBMC-PCR to identify HCV infections in naïve cases that tested negative by SRT-PCR (COCIs and SNOCIs). The significant increases in the numbers of the infected cases

Table 5. RNA seroconversion versus spontaneous intracellular disappearance of naïve HCV infection as recognized by concomitant SRT-PCR and PBMC-PCR

	Baseline PCR data at study entry			Follow-up by PCR after one year			
		PBMC-PCR			SRT-PCR	PBMC-PCR	
Serial number	SRT-PCR Serum RNA	Sense strand	Antisense strand	Fibroscan results at baseline in kPa	Serum RNA (IU/ml)	Sense strand	Antisense strand
1	-ve	+ve	+ve	F4 = 51.4	21700	+ve	+ve
2	-ve	+ve	+ve	F4 = 57.3	242000	+ve	+ve
3	-ve	+ve	+ve	F4 = 28.0	299000	+ve	+ve
4	-ve	+ve	+ve	F4 = 35.3	-ve	+ve	+ve
5	-ve	+ve	-ve	F4 = 17.1	-ve	-ve	-ve
6	-ve	+ve	-ve	F2 = 9.50	-ve	-ve	-ve
7	-ve	+ve	-ve	F1 = 5.80	-ve	-ve	-ve

Positive cases for both intra-PBMC HCV RNA strands at baseline are associated with RNA seroconversion in 3 out of 4 (uncorrected 2-tailed, p = 0.047) F4 naïve patients; the remaining patient failed to clear the intracellular sense and antisense RNA strands. All naïve patients who had positivity for intra-PBMC HCV-RNA sense but negativity for antisense strand cleared the intracellular infection within 1 year. kPa, Kilo Pascal; +ve, positive; -ve, negative. F0 <5.5 kPa, F1 = 5.5-7.5 kPa, F2 = 7.5-10 kPa, F3 = 10-15 kPa, F4 > 15.0 kPa.

after adding the newly diagnosed cases by PBMC-PCR recommends cellular screening for HCV-RNA infection to diagnose SNOCIs. On the other hand, the present results showed that COCIs prevalence is almost the same as described in other reports. $^{2-4}$

It is important to remember that the studied population has a higher incidence of HCV genotype 4 and a lower incidence of genotype 1, a profile that is spreading widely in western countries. However, the current study did not aim to investigate OCIs for any particular genotype, but instead aimed to provide insight into a new use of an established diagnostic tool that would facilitate the diagnostic workup and furnish more accurate information regardless to viral genotype. The clinical application of the data analysis method that was adopted in the current study would be valid for studying of other populations and a spectrum of genotypes. However, calculation of the real prevalence of HCV infection in any population worldwide is dependent upon the combination of multiple diagnostic procedures (i.e. enzyme-linked immunosorbent assay, traditional PCR, PBMC-PCR, tissue PCR, ultracentrifugation-based PCR approach, etc.),^{5,20} a countless number of tissues (i.e. serum, blood cells, whole blood, bone marrow, liver tissues, etc.),^{20,24,26} and application in various clinical case situations (i.e. treatment-naïve, treatment-experienced, in contact with HCV patients, and with chronic active hepatocellular damage).^{5,23-28}

Our study concluded that SCSD is inversely proportional to the degree of hepatic cirrhosis; in addition, the increased hepatic cirrhosis is associated with diminished frequency of SCSD. These results confirm other studies regarding the association of liver cirrhosis with active HCV infection in viremic patients,¹⁰ but is considered a novel finding in SNOCIs as we reported in a previous study.⁵ It is convenient to consider the presence of HCV IgG antibody in naïve nonviremia (SCSD) patients as a remark for history of HCV infection, despite the belonging of dormant HCV infection in SCSD patients to the SNOCIs, as documented in the current study. Considering the clinical importance of PBMC-PCR, the significantly higher number of SCSD cases (73/112) compared to SNOCIs (39/112) supports the importance of anti-HCV IgG antibody testing as a preliminary screening approach, although it is not sufficient as a baseline diagnostic test for treatment. The numbers of SCSD cases are expected to drop down upon searching nucleated cells from other tissues (e.g., bone marrow, lymph nodes, spleen, liver, etc.) or using combined serologic diagnostic tests.²⁰ The rising titers of anti-HCV IgG antibodies in naïve and post-treatment cases might be a good predictor for current chronic HCV-infection.

Diagnosis of hepatic fibrosis in non-viremia patients is a confusing clinical situation that might push both physician and patients to face unexpected morbidity and mortality outcomes over time. Results of intra-PBMC HCV-RNA detection in the current study concern the above-mentioned clinical situation. Fortunately, the current report addresses HCV as an inducing factor for liver cirrhosis in SNOCIs, which will cause impaired liver function. The impact of hepatic fibrosis on SCSD of infection is most probably dependent upon interactions between the host immune system and the underlying pathogenic etiology. The current data set linked density of hepatic fibrosis with the presence of intra-PBMC HCV-RNA sense and antisense strands. Cumulative data during the next few years might help in creation of effective guidelines for HCV therapy to address treatment feasibility of solitary intracellular chronic HCV infection problems in non-viremia patients. Drawbacks of neglecting progressive hepatic cirrhosis are shown by evidence of active liver cell damage, so that lack of appropriate and timely medical management is extremely hazardous. Narrowing the spectrum of the etiologic factors in anti-HCV IgG antibody-positive non-viremia patients by management of SNOCIs in cirrhotic patients should minimize the over-time morbidity and mortality rates among chronic HCV infections.

On the other hand, differential recognition of intra-PBMC sense and antisense strands by PCR in our study provided elective information regarding their relationship to HCV RNA seroconversion in naïve patients. The present results indicate that solitary identification of intra-PBMC HCV-RNA sense strand would not be enough to induce viral genomic material seroconversion in naïve patients. The co-existence of sense with antisense strands seems to be crucial for HCV-RNA genomic materials to reappear in patient's sera. The authors believe that this is the first-report of HCV-RNA spontaneous seroconversion in naïve non-viremia cirrhotic patients occurring during the follow-up period without intervention, despite the small number (7 patients) that limits clinical significance. Contrary to these clues, the presence of intracellular antisense strand in naïve patients was reported as a good prognostic marker of response to antiviral therapy, and the post-treatment persistence of HCV-RNA antisense strands is an ominous sign because of the high probability of relapse.

Findings of the current study include: a) diagnosis of both cryptogenic and SNOCIs by PBMC-PCR having significant impact upon the total prevalence of naïve HCV-infection; b) recognition of SNOCIs in all IgG antibody seropositive non-viremic patients being highly recommended because of their frequent detectability compared to controls; c) use of PBMC-PCR being strongly recommended to diagnose SCSD, in addition to SRT-PCR; d) cases of SCSD, as diagnosed by HCV PBMC-PCR, being more prevalent in non-cirrhotic patients than in cirrhotic patients; e) both intra-PBMC HCV-RNA sense and antisense strands being associated with increased density of hepatic fibrosis in kPa units; f) and primary seroconversion of HCV-RNA in naïve patients being associated with combined intra-PBMC existence of sense and antisense strands.

In conclusion, evaluation of intra-PBMC HCV RNA infection is crucial for diagnosing OCIs, evaluating post-infection sequelae in RNA seronegative naïve patients, and addressing the probability of viral RNA serologic relapse.

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Conflict of interest

The authors have no conflict of interests related to this publication.

Author contributions

Responsible for study design, patient management, data analysis, and writing of the manuscript (MDAA), recruited patients, revised and formatted the manuscript, and performed the virological investigation (SAE, GYW, MKEA).

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