

The Role of Alterations in Alpha-Klotho and FGF-23 in Kidney Transplantation and Kidney Donation

Meera Gupta^{1,2*}, Gabriel Orozco^{1,2}, Madhumati Rao³, Roberto Gedaly^{1,2}, Hartmut H. Malluche³ and Javier A. Neyra^{3,4*}

¹ Department of Surgery - Transplant Division, University of Kentucky, College of Medicine, Lexington, KY, United States, ² Department of Surgery, University of Kentucky, Lexington, KY, United States, ³ Department of Internal Medicine -Nephrology, Bone and Mineral Metabolism Division, University of Kentucky, College of Medicine, Lexington, KY, United States, ⁴ Division of Nephrology, Department of Medicine, University of Alabama at Birmingham, Birmingham, AL, United States

Cardiovascular disease and mineral bone disorders are major contributors to morbidity and mortality among patients with chronic kidney disease and often persist after renal transplantation. Ongoing hormonal imbalances after kidney transplant (KT) are associated with loss of graft function and poor outcomes. Fibroblast growth factor 23 (FGF-23) and its co-receptor, α -Klotho, are key factors in the underlying mechanisms that integrate accelerated atherosclerosis, vascular calcification, mineral disorders, and osteodystrophy. On the other hand, kidney donation is also associated with endocrine and metabolic adaptations that include transient increases in circulating FGF-23 and decreases in α -Klotho levels. However, the long-term impact of these alterations and their clinical relevance have not yet been determined. This manuscript aims to review and summarize current data on the role of FGF-23 and α -Klotho in the endocrine response to KT and living kidney donation, and importantly, underscore specific areas of research that may enhance diagnostics and therapeutics in the growing population of KT recipients and kidney donors.

Keywords: alpha-Klotho, FGF-23, kidney donation, kidney transplant, ESKD (end stage kidney disease), bone mineral density (BMD), cardiovascular disease, mineral bone disease

INTRODUCTION

Cardiovascular disease (CVD) and bone mineral disorders are important causes of mortality and morbidity among end-stage kidney disease (ESKD) patients (1). The endocrine adaptation to a decreased glomerular filtration rate (GFR) plays a central role in the cardiovascular and skeletal alterations observed in patients with chronic kidney disease—mineral bone disorder (CKD-MBD). Fibroblast growth factor 23 (FGF-23) and its coreceptor, α -Klotho, are key factors in phosphate homeostasis, and their dysregulation is an essential link between osteodystrophy, left ventricular hypertrophy, atherosclerosis, systemic inflammation, and renal fibrosis observed in patients with ESKD (2).

While the treatment of choice for ESKD is kidney transplant (KT), CVD continues to be a significant cause of morbidity and mortality after transplantation and is a contributing factor to graft failure and loss (3). Post kidney transplant patients (PKTP) have persistent alterations in bone mineral metabolism exacerbated by immunosuppressive therapy, which can lead to progression of bone disease and CVD (4). While the bone-vascular axis has been extensively studied and is significantly affected in patients with ESKD, the endocrine response to kidney transplantation and

OPEN ACCESS

Edited by:

Mostafa Abdelkhalek, Mansoura University, Egypt

Reviewed by:

Alexander Grabner, Duke University, United States Daniela Picciotto, San Martino Hospital (IRCCS), Italy

*Correspondence:

Meera Gupta meera.gupta@uky.edu Javier A. Neyra javier.neyra@uky.edu

Specialty section:

This article was submitted to Nephrology, a section of the journal Frontiers in Medicine

Received: 27 October 2021 Accepted: 03 February 2022 Published: 06 May 2022

Citation:

Gupta M, Orozco G, Rao M, Gedaly R, Malluche HH and Neyra JA (2022) The Role of Alterations in Alpha-Klotho and FGF-23 in Kidney Transplantation and Kidney Donation. Front. Med. 9:803016. doi: 10.3389/fmed.2022.803016

1

kidney donation is not yet fully understood. Among kidney donors, significant derangements of markers of bone metabolism, including serum α -Klotho, have been reported after nephrectomy (5–9). However, the clinical impact, if any, of these alterations in otherwise healthy living donors has not been determined.

In this manuscript, we aim to review and summarize current data on the role of FGF-23 and α -Klotho in the endocrine response to KT and living kidney donation. Importantly, we intend to underpin specific areas of research that may enhance diagnostics and therapeutics in the growing population of KT recipients and kidney donors.

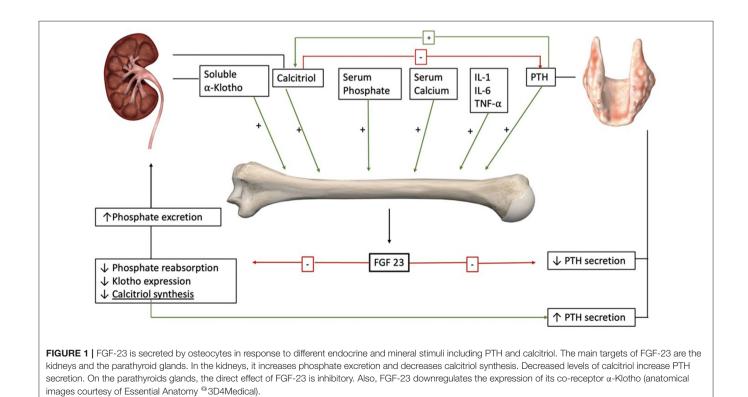
FGF-23 AND α -KLOTHO IN MINERAL METABOLISM

FGF-23 is one of the few fibroblast growth factors that enters the systemic circulation and acts as a hormone (10). FGF-23 is synthesized by osteocytes and secreted in response to rising serum phosphate levels. Its secretion is also stimulated by parathyroid hormone (PTH), vitamin D, soluble α -Klotho, and pro-inflammatory cytokines including IL-1, IL-6, and TNF α (2, 11–13). All fibroblast growth factor receptors (FGF-R) have a low affinity for FGF-23, and the high-affinity interaction depends on the expression of the co-receptor, α -Klotho (14, 15).

The organs most impacted by FGF-23 are the kidneys and parathyroid glands (Figure 1). In the absence of disease, FGF-23 induces the degradation and decline in the synthesis of

sodium-dependent phosphate transport protein 2A (NaPi2A) in the kidney (16). In doing so, FGF-23 directly decreases proximal tubular reabsorption of phosphate (17). Highlighting its physiological relevance, animal models with a deletion on FGF-23 or α-Klotho genes develop phosphate retention (18). FGF-23 also modifies mineral metabolism by decreasing calcitriol levels through two different mechanisms: it decreases calcitriol synthesis by downregulating 25-hydroxyvitamin D₃ 1a hydroxylase in the proximal tubular cells of the kidney, and it increases vitamin D inactivation by up-regulating vitamin D 24-hydroxylase (CYP24A1) expression (19, 20). The decrease in serum calcitriol contributes to PTH secretion despite a direct inhibition of the parathyroid glands by FGF-23 (20, 21). FGF-23 also promotes calcium absorption in the distal convoluted tubules, counterbalancing the inhibition of calcitriol synthesis (22). Notably, FGF-23 auto-regulates its effects by inhibiting the expression of α-Klotho (23). Hence, FGF-23 has a significant direct and indirect effect on phosphate and calcium metabolism.

 α -Klotho, discovered as a gene linked to aging, is predominantly expressed in the kidney, specifically in the distal tubules (24, 25). The α -Klotho gene encodes a 130 kDa single-pass transmembrane protein consisting of two extracellular domains (KL1 and KL2), a transmembrane domain, and a short cytoplasmic tail. Part of the extracellular domain of transmembrane α -Klotho is cleaved by proteases (26–28). A secreted form of α -Klotho has also been postulated from an alternatively spliced transcript (29, 30). Cleaved α -Klotho proteins are released into the circulation and are referred to as soluble α -Klotho. Soluble α -Klotho acts as an endocrine or paracrine factor affecting multiple distant organs such as the



heart, lungs, bone, and brain (31–37). α -Klotho increases FGF-23 expression in the bone (36) and acts as a non-enzymatic scaffold protein that enhances FGF-23 signaling (38).

 α -Klotho is an important regulator of mineral metabolism homeostasis: (1) it decreases renal phosphate reabsorption by acting as a co-receptor for FGF-23 binding to FGFR1 (15); (2) it directly promotes the internalization and degradation of the NaPi2a cotransporter in the renal proximal tubules (39); and (3) it suppresses vitamin D signaling (α -Klotho knockout mice can be rescued from a phenotype of soft tissue calcification by deletion of the CYP27B1 gene) (40–42). In addition, α -Klotho has a plethora of pleiotropic actions as an inhibitor of apoptosis, fibrosis, and cell senescence (43–46).

FGF-23 AND α-KLOTHO IN CHRONIC KIDNEY DISEASE AND ITS COMPLICATIONS

Patients with CKD have significantly reduced functional renal cell mass, leaving the residual renal parenchyma to maintain homeostasis by increasing phosphate excretion. FGF-23through its phosphaturic effects- plays a crucial role in this response (2, 17) and is an early biomarker of CKD (47). As intact FGF-23 is cleared by the kidneys, a decreased glomerular filtration rate (GFR) also independently contributes to its rising levels (48-50). Among patients with CKD, sustained elevated levels of FGF-23 lead to constant inhibition in calcitriol synthesis, with a consequent increase in PTH secretion (51, 52). Because PTH stimulates the release of FGF-23 synthesis, a closed loop of stimulation is created. Supraphysiologic levels of FGF-23 are also perpetuated by downregulation of its co-receptor α -Klotho, resistance to its renal action, and further hyperphosphatemia (53-56). Remarkably, the compensation to impaired kidney function might promote further progression of CKD through different mechanisms: the phosphate hyperfiltration by functional nephrons leads to tubular damage (57), downregulation of α-Klotho promotes renal fibrosis through increasing TGF_β1 (44) and Wnt (58) signaling pathways, and lower α-Klotho levels might decrease its protective effect on the glomerular filtration barrier (59). The resulting endocrine profile that characterizes patients with CKD -high FGF-23 and PTH, with low calcitriol and α-Klotho-contributes to cardiovascular morbidity, systemic inflammation, dysregulation of bone metabolism, and anemia.

Cardiovascular Diseases

Cardiovascular diseases cause more than half of all deaths among patients with ESKD, and they represent the leading cause of mortality in this vulnerable population (1). Among CKD patients older than 65 years, the prevalence of CVD is 65.8%, compared to 31.9% in patients of the same age group but with normal kidney function (60). Epidemiological studies have shown a direct association between FGF-23 levels, mortality, and risk of major cardiovascular events (61–63). Two major contributor mechanisms are left ventricular hypertrophy (LVH) and a proatherosclerotic milieu. LVH is associated with heart failure, changes in heart geometry, and cardiac arrhythmias, while atherosclerosis is associated with coronary, cerebrovascular, and peripheral artery diseases.

High circulating levels of FGF-23 directly induce LVH through the activation of FGFR4 in cardiac myocytes and recruitment of the PLC γ /Calcineurin/NFAT pathway independently of α - Klotho (64–66). FGFR4 is a potential therapeutic target for reducing cardiovascular risk in patients with CKD as pharmacological blockage of this receptor in animal models showed attenuated LVH, while loss of function (FGFR4^{-/-}) protected mice from developing LVH (64).

The pro-atherosclerotic milieu characteristic of patients with CKD is related to alterations in the bone-kidney axis (60, 67). In the initial stages of CKD, increased secretion of FGF-23 helps maintain normal serum phosphate (47). As the GFR decreases, transient or continuous hyperphosphatemia leads to calcium phosphate precipitation and favors the formation of atherosclerotic plaques (68). There is a positive correlation between FGF-23 and vascular calcification in patients with CKD; however, it is still controversial if the association is independent of serum phosphate concentrations or a direct pro-atherosclerotic effect of FGF23 (69).

Aortic stiffness is another biomarker of cardiovascular risk that increases in patients with CKD. While atherosclerosis primarily affects the vascular intima, aortic stiffness represents degenerative changes in the extracellular matrix of the intima and media layers characterized by elastin fracture and collagen deposition (70). Smith et al. demonstrated that increased aortic stiffness among CKD patients is associated with elevated circulating calciprotein particles—which are formed by adsorption of calcium phosphate by Fetuin-A to prevent ectopic calcification (71, 72). Also, *in-vitro* studies have shown that hyperphosphatemia can trigger chronic inflammation, induce endothelial apoptosis and promote an osteoblastic phenotype in vascular smooth muscle cells (2, 73–75). These are other examples of how mineral metabolism affects the vascular system and increases cardiovascular risk in CKD.

Immune System and Inflammation

The systemic inflammation observed in patients with CKD contributes to their elevated morbidity and mortality (76-78). An association between renal function and inflammation is well-established, and eGFR is inversely correlated with circulating levels of pro-inflammatory cytokines (e.g., IL-1β, IL-6, TNF-α, and hs-CRP) (79). Factors that promote inflammation in CKD include increased production of cytokines, hyperphosphatemia, chronic and recurrent infections, acidosis and oxidative stress, altered metabolism of adipose tissue, and gut microbiota dysbiosis (80). FGF-23 directly induces liver secretion of inflammatory cytokines by targeting FGFR4 receptors in hepatocytes and activating PLCy/Calcineurin/NFAT signaling pathway— independently of α -klotho. As an indicator of the therapeutic potential of this finding, blockage of FGFR4 prevented significant elevation in C-Reactive Protein (CRP) secretion by the liver in an animal model of CKD (76). Also, altered immune homeostasis increases the susceptibility to infections. FGF-23 impairs neutrophil recruitment by interfering with chemokine signaling through activation of FGFR2 (81). In an unfavorable feedback loop, FGF-23 increases hepatic secretion of cytokines, inflammatory cytokines stimulate the secretion of FGF-23 by osteocytes, and a predisposition to infections contributes to systemic inflammation.

Bone Metabolism

There is a wide range of bone disorders that affect patients with CKD. The spectrum includes disorders in bone turnover, impaired mineralization, and changes in bone volume (82, 83). These abnormalities cause decreased mechanical strength with increased risk of fractures (84, 85), and altered mineral metabolism with increased risk of ectopic calcification (86). Malluche et al. reported a series of 630 bone biopsies from patients with CKD on dialysis. With 418 cortical bone biopsies available, they found that 82% had altered bone turnover, 52% had high cortical porosity, cortical thickness was low in 44%, cancellous bone volume was altered in 69% of the cohort, and only 3% of patients had impaired mineralization. Notably, bone turnover was low among 58% of patients and high among 24% of patients (87). These data show the high prevalence and variety of bone alterations in patients with CKD. High bone turnover is related to elevated PTH, hyperphosphatemia, and increased cortical porosity. Conversely, low bone turnover is related to low calcium retention and ectopic calcification. Low calcium retention indicates impaired incorporation of calcium into the bone matrix with the consequent hypercalcemia after calcium loads (88).

Animal models have shown that FGF-23 also regulates bone mineralization through a phosphate-independent mechanism. Homozygous FGF-23 knock-out mice develop growth delay, low bone mineral density (BMD), skeletal malformations, increased bone fragility, and impaired osteoid mineralization with concomitant hyperphostemia (89). Furthermore, these skeletal abnormalities persisted in hypophosphatemic animal models (FGF-23 -/- / NaPi2a -/-) (90). On the other hand, in fetal rat calvaria cell cultures overexpressing FGF-23, there was also inhibition of bone mineralization (91). This apparent contradiction-where deficiency and exposure to FGF-23 lead to impaired bone mineralization-favors the possibility that FGF-23 acts as an autocrine/paracrine regulator of mineralization (92) and the inhibition of tissue-non-specific alkaline phosphatase activity (TNAP) by FGF-23 might play a role. TNAP hydrolyzes pyrophosphate and generates inorganic phosphate to enhance mineralization (93). FGF-23 can suppress TNAP activity, cause pyrophosphate accumulation, and inhibit mineralization. The deletion of FGF-23 in Hyp mice (a murine homolog for X-linked hypophosphatemia) rescued the suppressed TNAP activity in osteoblasts (93, 94). However, FGF-23 deficiency ($Fgf^{-/-}$ mice) increases TNAP activity, increases the release of osteopontin, and also leads to inhibition of mineralization. This inhibitory effect of osteopontin in bone mineralization can be attenuated with a low phosphate diet or by deletion of the NaPi2A transporter (92, 95).

Osteocytes and osteoblast synthesize α -Klotho (96). Mice with an osteocyte-specific deletion of the α -Klotho gene have increased osteoblast activity and increased bone volume with normal serum biochemistry. When the same deletion is introduced in mice models of CKD (5/6 nephrectomy and highphosphate diet)—which present with elevated serum phosphate, PTH, and FGF-23— α -Klotho expression in the bone was significantly decreased and the pro-osteoblastic effect was no longer significant. This suggests that α -Klotho might act as a regulator of adequate bone mineralization rate when the systemic mineral homeostasis is preserved (92, 97). As described, the endocrine mechanisms affecting bone metabolism are complex and questions remain as to the role of FGF-23 and α -Klotho in the differential expression of bone disorders observed in patients with CKD.

Anemia

Epidemiological studies have found that FGF-23 is independently associated with anemia (98, 99), and animal studies have reported possible causal mechanisms. The administration of FGF-23 in wild-type mice rapidly decreases erythropoiesis while Knockout mice (FGF-23 $^{-/-}$) have increased circulating erythropoietin and red blood cell number (100). Furthermore, Agoro et al. reported that inhibiting FGF-23 signaling rescues anemia and iron deficiency in a mice model of CKD by normalizing the hemoglobin levels, decreasing erythroid cell apoptosis, and increasing serum erythropoietin, iron, and ferritin. They also found an up-regulation on the liver synthesis of hepcidin that normalized after FGF-23 blocking and provided a possible link between FGF-23, inflammation, and anemia (101).

In summary, elevated serum FGF-23 is an early biomarker of impaired renal function that directly contributes to the morbidity observed in patients with CKD. Clinical and animal studies have shown that potential therapeutic benefits of targeting FGF-23 include attenuating left ventricular hypertrophy, decreasing the synthesis of pro-inflammatory cytokines by the liver, improving neutrophil recruitment, rescuing anemia by stimulating erythropoiesis, and improving iron deficiency by down-regulating the synthesis of hepcidin. Decreased circulating levels of α -klotho is also characteristic of patients with CKD and potential therapeutic benefits of targeting α -klotho-dependent actions include increasing circulating levels of calcitriol, decreasing renal fibrosis, and improving bone mineralization and osteoblast function.

POST KIDNEY TRANSPLANT PATIENTS

Kidney transplantation is associated with a significant reduction in mortality compared to those with ESKD on dialysis. Wolfe et al. reported a 1.7-fold decrease in the annual death rate among patients with ESKD undergoing transplant compared to patients on waiting list (102). Notably, CVD persists as the leading cause of death after transplantation (60). According to the USRDS, the prevalence of CVD is 76.5% in patients on hemodialysis, 65% in patients on peritoneal dialysis, and 53.7% in patients with a functional kidney transplant (1). Life expectancy among kidney transplant recipients remains inferior compared to age-matched controls from the general population. For example, females who undergo kidney transplantation between 40 and 44 years old have a projected lifespan of more than 10 years shorter than similarly aged women from the general population (1). For this reason, it is paramount to identify and characterize the mechanisms underlying CVD in kidney transplant recipients.

Transplant mineral and bone disorder (T-MBD) is a systemic condition characterized by endocrine alterations, vascular or other soft-tissue calcification, and abnormalities in bone metabolism (103, 104). Although there are significant differences with CKD-MBD, there is a continuum in some metabolic pathways conducive to atherosclerosis and bone disorders (104). There are several mechanisms involving the aforementioned biomarkers after kidney transplant with persistent interplay affecting mineral bone metabolism, cardiovascular disease, and load-bearing capacity.

After transplant, as renal function stabilizes, there is a tendency toward normalization of most biochemical indicators of ESKD. A recovered GFR allows excretion of excess serum phosphate and uremic toxins. High basal levels of PTH and FGF-23 make post-transplant hypophosphatemia and hypercalcemia a common finding (105–110). Transient hypophosphatemia is observed in more than 80% of transplant recipients early after surgery, and is related to appropriate allograft function and favorable outcomes (106). On the other hand, transient hypercalcemia is reported in up to 65% of transplants and is associated with poor outcomes (111–113). Hypercalcemia can compromise adequate graft function by acutely inducing vasoconstriction and, in the long-term, by favoring calcinosis and vascular calcification (114, 115).

In the first 3 months following surgery, there is a significant decrease in PTH, FGF-23, and calcium levels and an increase in serum phosphate and calcitriol toward normal levels (107). After this period, these changes normalize at a slower rate. Nonetheless, long-term endocrine alterations are common (106, 107, 109, 116). Among 50 patients undergoing kidney transplantation, Evenepoel et al. reported that 1 year after surgery, 57% had persistently high FGF-23, and 32% had high serum PTH levels. Mild to moderate hypophosphatemia was also present in 14% of patients (117). Elevated FGF-23 is associated with decreased post-transplant GFR (118, 119) and, and increased risk of mortality, graft loss (120, 121), and left ventricular wall strain (119, 121). The effects of FGF-23 in the cardiovascular system were also studied by Yilmaz et al. who reported that endothelium-dependent vasodilation improves after kidney transplantation, and it is independently associated with a decrease in FGF-23 and serum phosphorus (122). As mentioned above, high circulating levels of FGF-23 directly contribute to left ventricular hypertrophy (LVH) and anemia in patients with CKD. Decreases in FGF-23 might contribute to the reduction of LVH and improvements of ventricular contractility after kidney transplantation (123, 124). Baloglu et al. reported that hemoglobin levels are inversely associated with circulating FGF-23, however this association was no longer significant after kidney transplantation (125).

Serum α -Klotho levels increase after transplantation and might contribute to the transient hypophosphatemia by improving the sensitivity to FGF-23 (126, 127). Interestingly, Mizusaki et al. reported that serum α -Klotho was significantly higher in kidney transplant recipients receiving everolimus. They concluded that mTOR inhibition could be related to increased α -Klotho after kidney translantation (127), which underpins a new area of applicable α -Klotho therapeutics in kidney transplantation that needs further study. Higher serum α-Klotho has also been independently associated with improved GFR 1 year after transplant (128). Animal models have shown a renoprotective effect of α -Klotho that could play a role in protecting the renal allograft (129–131). Sugiura et al. reported that α -Klotho induction through gene transferring strategy mitigates apoptosis, histological damage and improves GFR in mice subject to renal ischemia-reperfusion injury. They concluded that the protective mechanism involved increases expression of heat shock protein 70 (129). The antifibrotic effect of α -Klotho through the inhibition of TGFβ1 and Wnt signaling pathways is also an important mechanism in kidney protection against injury (44, 58). Hu et al. studied mice after acute kidney injury and found that early treatment with α-Klotho prevents progression to CKD and significantly reduces uremic cardiomyopathy (37). As an indicator of renal function, α -Klotho activity varies depending on the quality of the transplanted kidney. Among kidneys from deceased donors, lower Kidney Donor Risk Index (KDRI) and age <50 years were associated with higher α -Klotho levels and better GFR after transplant (128, 132). In this context, α -Klotho represents a potentially useful biomarker and therapeutic agent in kidney transplantation.

Bone metabolism and load-bearing capacity are also greatly affected in kidney transplant recipients. The underlying mechanism is multifactorial and includes pre-existing renal osteodystrophy, reduced kidney function, and the effect of immunosuppressive therapy (133, 134). Among 68,814 kidney transplant recipients, Nikkel et al. reported a 22.5% incidence of fractures during the first 5 years after surgery (135). The first 6 months following transplant are associated with a rapid decrease in spine bone mineral density that continues to decline afterward but at a slower rate (136-138). It is well-established that immunosuppressive therapy with glucocorticoids induces osteopenia and is a contributing factor to osteodystrophy. Glucocorticoids are related to a decreased number and impaired function of osteoblasts. They also promote osteoclastic activity by up-regulating RANKL expression and down-regulating osteoprotegerin (139-141). Conversely, the effect of calcineurin inhibitors in the bone is more controversial and difficult to evaluate (141-143). While tacrolimus and cyclosporine have been linked to osteoporosis, population-based studies have failed to find an association with an increased risk of fractures (141, 143-146). Briner et al. conducted a clinical study that combined cyclosporine treatment with no or very low dose of glucocorticoids. They found that glucocorticoid-sparing immunosuppression after transplant prevented cancellous bone loss, and they did not observe any fractures during the 2 years follow-up (147). Also, Westeel et al. reported a series of 52 renal transplanted patients receiving low dose prednisone and cyclosporine. They concluded that cyclosporine, together with the decrease of prednisone dosage, contributes to a transient stimulation of bone remodeling at 6 months after transplant, which counterbalances the bone loss associated with prednisone therapy (142). In addition to immunosuppressive treatment, post-transplant hypophosphatemia is also independently TABLE 1 | Human studies of soluble α -Klotho and FGF-23 among kidney transplant recipients.

Study	Clinical setting	Methods	Observations	FGF-23 and α -Klotho highlights
Evenepoel et al. (117)	50 KTRs and 50 eGFR-matched controls followed for one year after transplant.	FGF-23 (Serum ELISA)	 FGF-23 levels tend to normalize after successful renal transplant. Elevated FGF-23 levels (>50 ng/L) were observed in 66% and 57% at month 3 and month 12, respectively. At month 3, FGF-23 levels were significantly higher than controls. Statistical significance was lost after one year. Hyperphosphatoninism and renal phosphorus wasting also regressed by 1 year after transplantation. 	FGF-23 tends to normalize after renal transplantation.
Trombetti et al. (105)	69 KTRs patients followed evaluated 10-13 days after transplant.	FGF-23 (Serum ELISA)	 Higher serum FGF-23 levels were independently associated with lower serum phosphate and decreased renal tubular reabsorption of phosphate after transplant. FGF-23 was inversely associated with serum calcitriol levels and directly associated with serum calcium levels. Serum levels of PTH and FGF-23 were not correlated. 	Preoperative serum FGF-23 is associated with post-transplant phosphate, calcium, and calcitriol levels.
Kawarazaki et al. (108)	39 KTRs were evaluated one week before and 12 months after transplant.	FGF-23 (Serum ELISA)	 Pre-transplant FGF-23 was the best pre-transplant predictor of persistent hypophosphatemia one year after kidney transplant. 	Preoperative FGF-23 is a predictor of persistent post-transplant hypophosphatemia.
Wolf et al. (120)	984 KTR were followed up for three years.	FGF-23 (Serum ELISA)	 During a median follow-up of 37 months, 87 patients died and 101 patients had allograft loss. High levels of FGF-23 were independently associated with all-cause mortality and graft loss. 	FGF-23 levels are associated with all-cause mortality and graft loss in stable kidney transplant recipients.
Han et al. (107)	20 KTRs followed for 12 weeks.	FGF-23 (Serum ELISA)	 FGF-23 levels decreased by 97% at 4 weeks after transplant but were still above normal. FGF-23 levels, but not PTH levels, were independently associated with post-transplant hypophosphatemia. 	Preoperative FGF-23 levels are associate with post-transplant hypophosphatemia.
Sanchez et al. (118)	Cross-sectional study of 279 maintenance kidney recipients with CKD stages 1-4 and stable allograft function.	FGF-23 (Serum ELISA)	 FGF23, PTH, and phosphorus levels were higher in more advanced stages of CKD, while serum calcitriol levels and phosphate reabsorption rate were lower. A significant inverse correlation was found between eGFR and FGF23. FGF-23 was inversely correlated with serum phosphate levels. High FGF-23 was independently associated with increased time on corticosteroids, increased PTH, increased serum phosphate, and decreased calcitriol. Higher FGF23 values were not correlated with increased phosphate excretion. 	Serum FGF-23 levels remain elevated after renal transplantation and are inversely correlated with eGFR.
Wesseling-Perry et al. (109)	44 pediatric KTRs followed for 6 months after transplant.	FGF-23 (Urinary and serum ELISA)	 Higher pre-transplant FGF-23 values were associated with an increased fractional excretion of phosphorus and a faster decline in circulating phosphate during the first week after transplantation. Circulating 1,25(OH)2 vitamin D levels increased more rapidly and were consistently higher in patients with lower FGF23 values. 25(OH) vitamin D and 24,25(OH)2 vitamin D values were unrelated to FGF23 concentrations. Post-transplantation fractional excretion of FGF-23 was less than 5% at all post-operative times, even in the patients with highest circulating FGF-23 	Higher pre-transplant FGF-23 is related with faster decline in serum phosphate and slower increase in calcitriol levels after renal transplantation.
Baia et al. (121)	593 stable KTRs were evaluated 2.7-11.7 years after transplantation and followed for 6.2 - 7.5 years.	FGF-23 (Serum ELISA)	 FGF23 was independently associated with markers of left ventricular wall strain (pro-A-type natriuretic peptide, N-terminal-pro brain natriuretic peptide, and copeptin). FGF23 serum level was associated with a higher risk of cardiovascular and all-cause mortality. 	Among long-term kidney transplant recipients, higher FGF-23 is associated with markers of left ventricular wall strain, cardiovascular mortality, and all-cause mortality.

(Continued)

Alpha-Klotho and FGF23 in Transplantation

Study	Clinical setting	Methods	Results/Observations	FGF-23 and α -Klotho highlights
Yilmaz et al. (122)	161 adult KTRs were evaluated before and 6 months after transplant.	FGF-23 (Serum ELISA)	Endothelium-dependent vasodilatation improved after kidney transplantation, and it was independently associated with a decrease in FGF23 and serum phosphorus, and an increase in 25-OH-VitaminD levels.	FGF-23 is associated with improved endothelium-dependent vasodilation after kidney transplantation.
Malyszko et al. (119)	84 KTRs with stable graft function and no clinical signs of rejection. Control group: healthy volunteers.	α-Klotho and FGF-23 (Serum ELISA)	 FGF23 was significantly higher and Klotho significantly lower in kidney transplant recipients compared with healthy volunteers. FGF-23 significantly correlated with IL-6, VCAM, and copeptin. α-Klotho significantly correlated with N-Terminal pro brain natriuretic peptide: a marker of left ventricular wall strain. FGF23 was significantly higher and Klotho significantly lower in patients with eGFR >60 mL/min compared with patients with eGFR <60 mL/min. They concluded that disturbances in the FGF23-Klotho system appeared to be related to the endothelial cell injury 	Among kidney transplant recipients, there is an association between markers of endothelial cell injury with high FGF-23 and low α -klotho levels.
Bleskestad et al. (110)	Cross-sectional study of 39 adults KTRs with more 10 years after first transplant. 39 eGFR matched controls. 20 healthy controls.	α-Klotho and FGF-23 (Serum ELISA)	- Among KTR, FGF23 and PTH were significantly higher than in healthy volunteers. - When compared with matched controls for eGFR, PTH was significantly higher and levels of FGF23 had a non-significant trend towards higher levels and α -Klotho towards lower levels.	Among long-term kidney transplant recipients there are persistently high levels of FGF-23 and PTH.
Tartaglione et al. (116)	Cross sectional study of 80 adult KTRs.	30 healthy controls α-Klotho and FGF-23	 Serum level of FGF23 was significantly increased in KTR. Serum α-Klotho was significantly decreased in KTR. FGF-23 had a direct correlation with sclerostin levels (bone-antianabolic protein), α-Klotho had a significant positive association with sclerostin levels on multivariate analysis. 	Sclerostin – a bone antianabolic protein- has a positive association with FGF-23 and α -Klotho levels in kidney transplant recipients.
		(Serum ELISA)		
Tan et al. (126)	29 KTRs were prospectively evaluated before transplant and at 1, 12 and 52 weeks after transplant.	α-Klotho and FGF-23 (Serum ELISA)	 α-Klotho was significantly increased at 52 weeks following an initial decline at 1 week. FGF23 was significantly lower at 52 weeks. 	FGF-23 decreases, and α -Klotho increases after renal transplantation.
Mizusaki et al. (127)	36 KTRs were evaluated before and 1 year after transplantation.	α-Klotho (Serum ELISA)	 α-Klotho levels were higher after transplantation than before transplantation. After transplant, α-Klotho levels were significantly higher in recipients taking everolimus than in those not taking everolimus. mTOR inhibition may augment the increase in Klotho levels in transplant recipients. 	α-Klotho levels were significantly higher in renal transplant recipients receiving immunosuppressive treatment with mTOF inhibitors (everolimus).
Thongprayoon et al. (9)	431 KTRs; meta-analysis with variable follow-up	α-Klotho (Serum ELISA)	 KTRs had a significant increase in serum klotho levels at 4 to 13 months post-KT. KT recipients had a lower serum klotho level compared to healthy unmatched volunteers. 	Serum α -Klotho increases after kidney transplantation, but it is still lower than healthy volunteers.

CKDs, chronic kidney disease patients; ELISA, enzyme-linked immunoassay; FGF, fibroblast growth factor; eGFR, estimated glomerular filtration rate; KTRs, kidney transplant recipients; KT, Kidney transplantation.

Alpha-Klotho and FGF23 in Transplantation

TABLE 2 | Human studies of soluble α -Klotho and FGF-23 among living donors.

Study	Clinical setting	Methods	Results/Observations	FGF-23 and α -Klotho highlights
Westerberg et al. (159)	9 LKDs followed for 6 months post-donation	Serum ELISA	 FGF-23 levels were negatively correlated with serum phosphate and calcitriol prior to nephrectomy FGF-23 levels increased from 31.8 ±12.3 pg/mL to 55.8 ±15.1 pg/mL one week after nephrectomy, and did not correlate with other measures, and return to baseline at 3 months FGF-23 correlated with PTH at 3 and 6 months 	FGF-23 increased after nephrectomy and returned to baseline by 3 months after surgery.
Young et al. (158)	198 LKDs and 98 non-donor controls followed 5.3 years post-donation	Serum Urine ELISA	 Median time after donation was 5.3 years FGF-23 was higher (38.9 vs 29.7 pg/mL) and eGFR was lower in donors compared to controls Donors had higher urine phosphate and iPTH, but lower serum calcitriol and phosphate levels iPTH did not correlate with FGF-23 or calcitriol Unclear if changes impact MBD or fracture rates 	Long-term follow up after nephrectomy showed persistently high FGF-23 and decreased eGFR among kidney donors.
Akimoto et al. (7)	10 LKDs and their LKRs followed for 5 days post-donation/transplant	Serum ELISA	 Baseline serum α-Klotho was 909.8 pg/mL (IQR 754.8–1132.4) among LKDs and 613.0 pg/mL (IQR 445.9-750.8) among recipients prior to surgery Postoperative day 5, serum α-Klotho among donors dropped to 619 pg/mL (IQR 560.3–811.8 pg/mL), while no change in α-Klotho levels was observed in kidney recipients 	After kidney donation, serum α -Klotho decreased by post-operative day five when compared to pre-operative levels.
Ponte et al. (5)	27 LKDs followed for 12 months post-donation	Serum Urine IBL ELISA	 FGF-23 and α-Klotho drop within days post kidney donation 12 months post-donation, LKDs have lower eGFR and vitamin D3, unchanged vitamin D2, higher PTH, lower serum phosphate, unchanged FGF-23, and slightly improved but still lower α-Klotho levels compared to pre-donation 	One year after kidney donation, serum α -Klotho and eGFR were lower and FGF-23 is similar to pre-operative levels.
Kimura et al. (8)	15 LKDs and 6 LKRs followed for 5 days post-donation/transplant	Serum Urine IBL ELISA	 Baseline serum α-Klotho was 827.9 pg/mL (IQR 544.7–993.3) among recipients and 1084 pg/mL (IQR 794.8-1638.2) among LKDs prior to surgery Postoperative days 2 and 5, serum α-Klotho among donors dropped by –23.9 and –25.3 pg/mL, respectively; serum α-Klotho levels among recipients also decreased by –25.7 and –15.5 pg/mL, respectively Baseline urine α-Klotho was 58.6 ng/day (IQR 29.3–142.1) among recipients and 698.8 ng/day (62.3–1619.5) among LKDs Urinary α-Klotho increased well above baseline on postoperative days 2 and 5 among both groups suggesting that the kidney is a major source of urinary α-Klotho 	After kidney donation, serum α-Klotho decreased and urinary α-Klotho increase by postoperative day five when compare to pre-operative levels.
Thorsen et al. (157)	35 LKDs 15 years (median) post-donation, 60 CKDs, and 35 controls	Serum Plasma ELISA	 FGF-23 levels among LKDs [62.6 pg/mL(IQR 6.6–112)] were no different than controls [51.8 pg/mL(25.9–90.0)], but significantly lower than levels among advanced ESKDs α-Klotho levels among LKDs [669.3 pg/mL(IQR 409–1161)] were no different than controls [725.4 pg/mL(IQR 458–1222), but significantly lower among CKDs NGAL was higher among LKDs than controls (2.02±0.10 vs 1.89 ± 0.10 pg/mL; p<0.001), but much lower compared to CKDs FGF-23 and α-Klotho levels are not significantly different between LKDs long-term after donation and controls 	Long-term, FGF-23 and α -Klotho levels were similar between living kidney donors and healthy controls.
Tan et al. (6)	21 LKDs and 20 controls followed for 12 months post-donation	Serum IBL ELISA	 Biochemical and clinical profiles among LKDs and controls were similar pre-donation One month post-donation, serum creatinine increased (from 75 ± 12 to 114 ± 22 μmol/L), FGF23 increased (52 ± 15 to 70 ± 19 pg/mL) and α-Klotho decreased (564 [469-662] to 424 [375-523] pg/mL), all P less than 0.001. Changes persisted 12 months post-donation, while there were no alterations in serum phosphate, PTH, urinary phosphate among LKDs. 	One years after kidney donation, FGF-23 was higher and α -Klotho lower when compared to healthy controls.

Alpha-Klotho and FGF23 in Transplantation

Gupta et al.

TABLE 2 Continued				
Study	Clinical setting	Methods	Results/Observations	FGF-23 and α -Klotho highlights
Thongprayoon et al. (9)	108 LKDs and 431 KTRs; meta-analysis with variable follow-up	Serum	 LKDs had a decrease in α-Klotho (mean -232.24 pg/mL) 3-5 days post-donation, and slight recovery but remained below baseline one year post-donation (mean -110.80 pg/mL); and lower than healthy volunteers (mean -92.41 pg/mL) at one year post-donation There was no difference in FGF-23 (mean +8.19 pg/mL) among LKDs one year post-donation KTRs had increase in α-Klotho within the first year post-transplant (mean +243.11 pg/mL), comparable to eGFR-matched controls 	One year after kidney donation, serum ω -Klotho was lower when compared to pre-operative levels and healthy volunteers. FGF-23 was not similar to pre-operative levels.
Hiemstra et al. (156)	36 LKDs and 36 controls followed for 12 months	Serum IBL ELISA	 α-Klotho among LKDs declined early post-donation compared to controls [677.7pg/mL(536.7 – 833.9)vs 893.4pg/mL(739.8 – 1051.0]], but remained significantly lower than controls at 6-weeks [701.6 pg/mL(548.6 – 874.0]] and 12 months [721.4 pg/mL(562.5 – 956.5]] eGFR among LKDs declined markedly early post-donation compared to controls (52.3±17.5 vs 83.6±25.2), but remained lower at 6-weeks (60±20 vs 87.2±25.9) and 12 months (67.7±22.6 vs 88.6±24.3) There was no difference in FGF-23 between LKDs and controls at all-time points 	Serum <i>a</i> -Klotho and eFGR decreased early post kichey donation and remained low one year after surgery when compared to healthy controls. There was no difference in FGF-23 between LKDs and controls at all-time points
LKDs, living kidney donors; EL kidney transplant recipients; Ck	LKDs, living kidney donors; ELISA, enzyme-linked immunoassay; FGF, kidney transplant recipients; CKDs, chronic kidney disease patients.	libroblast growth f	LKDs, living kidney donors; ELISA, enzyme-linked immunoassay; FGF, fibroblast growth factor; iPTH, intact parathyroid hormone; MBD, mineral bone disease; IQR, interquartile range; eGFR, estimated glomerular filtration rate; KTRs, kidney transplant recipients; CKDs, chronic kidney disease patients.	GFR, estimated glomerular filtration rate; KTRs,

associated with alterations in bone turnover, decreased osteoblast activity, defective mineralization, and osteoblast apoptosis (133, 148). In this regard, high PTH and FGF-23 levels are frequent after transplant and contribute to hypophosphatemia. Persistent tertiary hyperparathyroidism also leads to increased bone turnover, and high FGF-23 has been related to phosphate-independent skeletal abnormalities (89, 90, 149). Human studies of soluble α -Klotho and FGF-23 among kidney transplant recipients are summarized in **Table 1**.

KIDNEY DONORS

In 2020, living kidney donors represented 23% of the total transplanted kidney allografts in the US. The long-term mortality after kidney donation has proven to be similar to the general population or matched controls in several studies (150-152). However, the innocuity of kidney donation is the subject of greater debate. O'Keeffe et al. conducted a meta-analysis that included 118,426 living kidney donors with an average follow-up of one to 24 years. They did not find evidence of increased mortality, cardiovascular diseases, or diabetes among donors compared to controls (153). Nonetheless, they found that donors had an increased risk of ESKD and pre-eclampsia. Muzaale et al. conducted a cohort study involving 96,217 kidney donors and found that 15 years after donation, the risk of ESKD increased from 0.04% in matched non-donors to 0.31% in kidney donors (154). There are scarce data available evaluating the outcomes after donation beyond a follow-up of 20 years. Based on OPTN data as of 2020, 26.5% of kidney donors were under 35 years old. In this regard, an increasing population is expected to live longer than 40 years after donation (155), and the long-term metabolic consequences of unilateral nephrectomy in otherwise healthy individuals are not clearly understood.

As discussed above, α -Klotho is primarily synthesized in the kidneys, and it has a reno-protective effect through different mechanisms. Shortly after kidney donation, α -Klotho levels decrease and remain lower than baseline 1 year after surgery (5–8, 156). However, one study that evaluated serum α -Klotho 5 years after donation did not find a significant difference compared to healthy controls (157). The long-term consequences of changes in α -Klotho levels in response to nephrectomy remain unknown. Further, the scope of the possible therapeutic benefit of increasing α -Klotho levels to prevent the development of ESKD after nephrectomy has not been explored in humans.

Other parameters of the bone-kidney axis are also altered after donation. There is a decrease in GFR, phosphate, and calcitriol synthesis, and an increase in PTH that persists 1 year after surgery (5). However, there is conflicting information about the changes in FGF-23 after donation. Some authors have reported that FGF-23 remains elevated 1 year after donation (6, 158), while others did not find significant long-term changes (5, 157, 159). A meta-analysis conducted by Thongprayoon et al. reported no significant change in FGF-23 1 year after kidney donation. Notably, they also found that serum α -Klotho remained lower than baseline 1 year after nephrectomy among 56 donors (9). As such, there is limited available data considering the clinical relevance of alterations in FGF-23 and α -Klotho after kidney donation (159). Human studies of soluble α -Klotho and FGF-23 among kidney donors are summarized in **Table 2**.

The understanding of long-term adaptations to kidney donation is still the subject of intense investigation. Living kidney donors are essential to close the gap between supply and kidney allograft demand, and sustaining their long-term health is important. Further research and education are necessary to support the kidney health of kidney donors and ensure the maintenance of their endocrine, cardiovascular, and bone mineral health.

CONCLUSION

FGF-23 and α -Klotho are integral endocrine factors that link bone-mineral metabolism, kidney function, and cardiovascular health. Among patients with CKD, this multi-looped axis is significantly impaired and translates into greater cardiovascular morbidity, mortality, and bone disorders. Kidney transplantation partially restores α -Klotho activity and FGF-23 levels. However, persistently high FGF-23 is associated with increased mortality and graft loss, and low circulating α -Klotho is associated with decreased kidney function after transplant. Notably, the

REFERENCES

- System USRD. 2020 Usrds Annual Data Report: Epidemiology of Kidney Disease in the United States. Bethesda, MD: National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases (2020).
- Kuro OM. The klotho proteins in health and disease. *Nat Rev Nephrol.* (2019) 15:27–44. doi: 10.1038/s41581-018-0078-3
- D'Marco L, Bellasi A, Mazzaferro S, Raggi P. Vascular calcification, bone and mineral metabolism after kidney transplantation. *World J Transplant*. (2015) 5:222–30. doi: 10.5500/wjt.v5.i4.222
- Westenfeld R, Schlieper G, Woltje M, Gawlik A, Brandenburg V, Rutkowski P, et al. Impact of sirolimus, tacrolimus and mycophenolate mofetil on osteoclastogenesis-implications for post-transplantation bone disease. *Nephrol Dial Transplant.* (2011) 26:4115–23. doi: 10.1093/ndt/gfr214
- Ponte B, Trombetti A, Hadaya K, Ernandez T, Fumeaux D, Iselin C, et al. Acute and long term mineral metabolism adaptation in living kidney donors: a prospective study. *Bone*. (2014) 62:36–42. doi: 10.1016/j.bone.2014.01.020
- Tan SJ, Hewitson TD, Hughes PD, Holt SG, Toussaint ND. Changes in markers of mineral metabolism after living kidney donation. *Transplant Direct*. (2017) 3:e150. doi: 10.1097/TXD.000000000000660
- Akimoto T, Kimura T, Watanabe Y, Ishikawa N, Iwazu Y, Saito O, et al. The impact of nephrectomy and renal transplantation on serum levels of soluble klotho protein. *Transplant Proc.* (2013) 45:134–6. doi: 10.1016/j.transproceed.2012.07.150
- Kimura T, Akimoto T, Watanabe Y, Kurosawa A, Nanmoku K, Muto S, et al. Impact of renal transplantation and nephrectomy on urinary soluble klotho protein. *Transplant Proc.* (2015) 47:1697–9. doi: 10.1016/j.transproceed.2015. 06.025
- Thongprayoon C, Neyra JA, Hansrivijit P, Medaura J, Leeaphorn N, Davis PW, et al. Serum klotho in living kidney donors and kidney transplant recipients: a meta-analysis. J Clin Med. (2020) 9:61834. doi: 10.3390/jcm9061834
- 10. Goetz R, Beenken A, Ibrahimi OA, Kalinina J, Olsen SK, Eliseenkova AV, et al. Molecular insights into the klotho-dependent, endocrine mode of action

metabolic bone and vascular alterations observed post kidney transplant are not fixed and instead are dynamic, influenced by organ quality, kidney recovery, immunosuppression, recipient comorbidities, and associated treatments. This highlights the concept that patients continue to need personalized medical support after transplant to ensure normalization of endocrine and bone-vascular axes indirectly affected by CKD. On the other hand, while some authors have reported that kidney donors have lower α -Klotho levels and elevated FGF-23 that persist 1 year after surgery, the scarce available data are inconclusive regarding the clinical relevance of these findings.

AUTHOR CONTRIBUTIONS

MG, GO, and JN contributed to conception and design of the study. MG and GO wrote the first draft of the manuscript. JN wrote sections of the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

ACKNOWLEDGMENTS

Figure adapted from anatomical images provided by Essential Anatomy. Many thanks to 3D4Medical.

of fibroblast growth factor 19 subfamily members. *Mol Cell Biol.* (2007) 27:3417–28. doi: 10.1128/MCB.02249-06

- McKnight Q, Jenkins S, Li X, Nelson T, Marlier A, Cantley LG, et al. Il-1beta drives production of fgf-23 at the onset of chronic kidney disease in mice. J Bone Miner Res. (2020) 35:1352–62. doi: 10.1002/jbmr.4003
- Han X, Li L, Yang J, King G, Xiao Z, Quarles LD. Counter-regulatory paracrine actions of Fgf-23 and 1,25(Oh)2 D in macrophages. *FEBS Lett.* (2016) 590:53–67. doi: 10.1002/1873-3468.12040
- Lawrence T. The nuclear factor Nf-kappab pathway in inflammation. *Cold Spring Harb Perspect Biol.* (2009) 1:a001651. doi: 10.1101/cshperspect.a001651
- Kurosu H, Ogawa Y, Miyoshi M, Yamamoto M, Nandi A, Rosenblatt KP, et al. Regulation of fibroblast growth factor-23 signaling by klotho. *J Biol Chem.* (2006) 281:6120–3. doi: 10.1074/jbc.C500457200
- Urakawa I, Yamazaki Y, Shimada T, Iijima K, Hasegawa H, Okawa K, et al. Klotho converts canonical Fgf receptor into a specific receptor for Fgf23. *Nature*. (2006) 444:770–4. doi: 10.1038/nature05315
- Gattineni J, Bates C, Twombley K, Dwarakanath V, Robinson ML, Goetz R, et al. Fgf23 decreases renal Napi-2a and Napi-2c expression and induces hypophosphatemia *in vivo* predominantly *via* Fgf receptor 1. *Am J Physiol Renal Physiol.* (2009) 297:F282–91. doi: 10.1152/ajprenal. 90742.2008
- Juppner H. Phosphate and Fgf-23. *Kidney Int.* (2011) 79121:S24– 7. doi: 10.1038/ki.2011.27
- Kuro OM. Phosphate and Klotho. *Kidney Int.* (2011) 79121:S20– 3. doi: 10.1038/ki.2011.26
- Bai X, Miao D, Xiao S, Qiu D, St-Arnaud R, Petkovich M, et al. Cyp24 inhibition as a therapeutic target in Fgf23-mediated renal phosphate wasting disorders. J Clin Invest. (2016) 126:667–80. doi: 10.1172/JCI81928
- Quarles LD. Skeletal secretion of Fgf-23 regulates phosphate and vitamin D metabolism. Nat Rev Endocrinol. (2012) 8:276–86. doi: 10.1038/nrendo.2011.218
- Ben-Dov IZ, Galitzer H, Lavi-Moshayoff V, Goetz R. Kuro-o M, Mohammadi M, Sirkis R, et al. The parathyroid is a target organ for Fgf23 in rats. J Clin Invest. (2007) 117:4003–8. doi: 10.1172/JCI32409

- 22. Quinn SJ, Thomsen AR, Pang JL, Kantham L, Brauner-Osborne H, Pollak M, et al. Interactions between calcium and phosphorus in the regulation of the production of fibroblast growth factor 23 *in vivo. Am J Physiol Endocrinol Metab.* (2013) 304:E310–20. doi: 10.1152/ajpendo. 00460.2012
- Marsell R, Krajisnik T, Goransson H, Ohlsson C, Ljunggren O, Larsson TE, et al. Gene expression analysis of kidneys from transgenic mice expressing fibroblast growth factor-23. *Nephrol Dial Transplant*. (2008) 23:827–33. doi: 10.1093/ndt/gfm672
- Kuro-o M, Matsumura Y, Aizawa H, Kawaguchi H, Suga T, Utsugi T, et al. Mutation of the mouse Klotho gene leads to a syndrome resembling ageing. *Nature*. (1997) 390:45–51. doi: 10.1038/36285
- Hu MC, Shi M, Zhang J, Addo T, Cho HJ, Barker SL, et al. Renal production, uptake, and handling of circulating alphaklotho. J Am Soc Nephrol. (2016) 27:79–90. doi: 10.1681/ASN.2014101030
- Bloch L, Sineshchekova O, Reichenbach D, Reiss K, Saftig P, Kuro-o M, et al. Klotho is a substrate for alpha-, beta- and gamma-secretase. *FEBS Lett.* (2009) 583:3221–4. doi: 10.1016/j.febslet.2009.0909
- Chen CD, Podvin S, Gillespie E, Leeman SE, Abraham CR. Insulin stimulates the cleavage and release of the extracellular domain of Klotho by Adam10 and Adam17. *Proc Natl Acad Sci USA*. (2007) 104:19796– 801. doi: 10.1073/pnas.0709805104
- Chen CD, Tung TY, Liang J, Zeldich E, Tucker Zhou TB, Turk BE, et al. Identification of cleavage sites leading to the shed form of the anti-aging protein Klotho. *Biochemistry*. (2014) 53:5579–87. doi: 10.1021/bi500409n
- Matsumura Y, Aizawa H, Shiraki-Iida T, Nagai R, Kuro-o M, Nabeshima Y. Identification of the human Klotho gene and its two transcripts encoding membrane and secreted Klotho protein. *Biochem Biophys Res Commun.* (1998) 242:626–30. doi: 10.1006/bbrc.1997.8019
- Shiraki-Iida T, Aizawa H, Matsumura Y, Sekine S, Iida A, Anazawa H, et al. Structure of the mouse Klotho gene and its two transcripts encoding membrane and secreted protein. *FEBS Lett.* (1998) 424:6–10. doi: 10.1016/S0014-5793(98)00127-6
- Panesso MC, Shi M, Cho HJ, Paek J, Ye J, Moe OW, et al. Klotho has dual protective effects on cisplatin-induced acute kidney injury. *Kidney Int.* (2014) 85:855–70. doi: 10.1038/ki.2013.489
- Cheng MF, Chen LJ, Niu HS, Yang TT, Lin KC, Cheng JT. Signals mediating Klotho-induced neuroprotection in hippocampal neuronal cells. *Acta Neurobiol Exp.* (2015) 75:60–71.
- 33. Sun S, Cheng B, Sun PG, Wu XH, Wu QQ, He P. Rtef-1 protects against oxidative damage induced by H2o2 in human umbilical vein endothelial cells through Klotho activation. *Exp Biol Med.* (2015) 240:1606– 13. doi: 10.1177/1535370215587914
- Ravikumar P, Ye J, Zhang J, Pinch SN, Hu MC, Kuro-o M, et al. Alpha-Klotho protects against oxidative damage in pulmonary epithelia. *Am J Physiol Lung Cell Mol Physiol.* (2014) 307:L566–75. doi: 10.1152/ajplung. 00306.2013
- Wang Y, Chen L, Huang G, He D, He J, Xu W, et al. Klotho sensitizes human lung cancer cell line to cisplatin *via* Pi3k/Akt pathway. *PLoS ONE*. (2013) 8:e57391. doi: 10.1371/journal.pone.0057391
- Smith RC, O'Bryan LM, Farrow EG, Summers LJ, Clinkenbeard EL, Roberts JL, et al. Circulating alphaklotho influences phosphate handling by controlling Fgf23 production. J Clin Invest. (2012) 122:4710–5. doi: 10.1172/JCI64986
- Hu MC, Shi M, Gillings N, Flores B, Takahashi M, Kuro OM, et al. Recombinant alpha-Klotho may be prophylactic and therapeutic for acute to chronic kidney disease progression and uremic cardiomyopathy. *Kidney Int.* (2017) 91:1104–14. doi: 10.1016/j.kint.2016.10.034
- Chen G, Liu Y, Goetz R, Fu L, Jayaraman S, Hu MC, et al. Alpha-Klotho is a non-enzymatic molecular scaffold for Fgf23 hormone signalling. *Nature*. (2018) 553:461–6. doi: 10.1038/nature25451
- Hu MC, Shi M, Zhang J, Pastor J, Nakatani T, Lanske B, et al. Klotho: a novel phosphaturic substance acting as an autocrine enzyme in the renal proximal tubule. *FASEB J*. (2010) 24:3438–50. doi: 10.1096/fj.10-154765
- Ohnishi M, Nakatani T, Lanske B, Razzaque MS. Reversal of mineral ion homeostasis and soft-tissue calcification of Klotho knockout mice by deletion of vitamin D 1alpha-hydroxylase. *Kidney Int.* (2009) 75:1166– 72. doi: 10.1038/ki.2009.24

- Razzaque MS. Fgf23, Klotho and Vitamin D Interactions: what have we learned from *in vivo* mouse genetics studies? *Adv Exp Med Biol.* (2012) 728:84–91. doi: 10.1007/978-1-4614-0887-1_5
- Neyra JA, Hu MC. Alphaklotho and chronic kidney disease. Vitam Horm. (2016) 101:257–310. doi: 10.1016/bs.vh.2016.02.007
- Sugiura H, Yoshida T, Shiohira S, Kohei J, Mitobe M, Kurosu H, et al. Reduced Klotho expression level in kidney aggravates renal interstitial fibrosis. *Am J Physiol Renal Physiol.* (2012) 302:F1252– 64. doi: 10.1152/ajprenal.00294.2011
- 44. Doi S, Zou Y, Togao O, Pastor JV, John GB, Wang L, et al. Klotho inhibits transforming growth factor-beta1 (Tgf-Beta1) signaling and suppresses renal fibrosis and cancer metastasis in mice. *J Biol Chem.* (2011) 286:8655– 65. doi: 10.1074/jbc.M110.174037
- Bian A, Neyra JA, Zhan M, Hu MC. Klotho, stem cells, and aging. *Clin Interv* Aging. (2015) 10:1233–43. doi: 10.2147/CIA.S84978
- 46. Shi M, Flores B, Gillings N, Bian A, Cho HJ, Yan S, et al. Alphaklotho mitigates progression of Aki to Ckd through activation of autophagy. J Am Soc Nephrol. (2016) 27:2331–45. doi: 10.1681/ASN.2015060613
- Isakova T, Wahl P, Vargas GS, Gutierrez OM, Scialla J, Xie H, et al. Fibroblast growth factor 23 is elevated before parathyroid hormone and phosphate in chronic kidney disease. *Kidney Int.* (2011) 79:1370–8. doi: 10.1038/ki.2011.47
- Zeng S, Querfeld U, Feger M, Haffner D, Hasan AA, Chu C, et al. Relationship between Gfr, Intact Pth, Oxidized Pth, non-oxidized Pth as well as Fgf23 in patients with Ckd. FASEB J. (2020) 34:15269–81. doi: 10.1096/fj.202000596R
- Mace ML, Gravesen E, Hofman-Bang J, Olgaard K, Lewin E. Key role of the kidney in the regulation of fibroblast growth factor 23. *Kidney Int.* (2015) 88:1304–13. doi: 10.1038/ki.2015.231
- Mace ML, Gravesen E, Nordholm A, Hofman-Bang J, Secher T, Olgaard K, et al. Kidney fibroblast growth factor 23 does not contribute to elevation of its circulating levels in uremia. *Kidney Int.* (2017) 92:165– 78. doi: 10.1016/j.kint.2017.01.015
- Imanishi Y, Inaba M, Nakatsuka K, Nagasue K, Okuno S, Yoshihara A, et al. Fgf-23 in patients with end-stage renal disease on hemodialysis. *Kidney Int.* (2004) 65:1943–6. doi: 10.1111/j.1523-1755.2004.00604.x
- 52. Gutierrez O, Isakova T, Rhee E, Shah A, Holmes J, Collerone G, et al. Fibroblast growth factor-23 mitigates hyperphosphatemia but accentuates calcitriol deficiency in chronic kidney disease. J Am Soc Nephrol. (2005) 16:2205–15. doi: 10.1681/ASN.2005010052
- Olauson H, Vervloet MG, Cozzolino M, Massy ZA, Urena Torres P, Larsson TE. New insights into the Fgf23-Klotho axis. *Semin Nephrol.* (2014) 34:586– 97. doi: 10.1016/j.semnephrol.2014.09.005
- Lim K, Lu TS, Molostvov G, Lee C, Lam FT, Zehnder D, et al. Vascular Klotho deficiency potentiates the development of human artery calcification and mediates resistance to fibroblast growth factor 23. *Circulation*. (2012) 125:2243–55. doi: 10.1161/CIRCULATIONAHA.111.053405
- Christov M, Neyra JA, Gupta S, Leaf DE. Fibroblast growth factor 23 and Klotho in Aki. Semin Nephrol. (2019) 39:57– 75. doi: 10.1016/j.semnephrol.2018.10.005
- Neyra JA, Hu MC, Moe OW. Klotho in clinical nephrology: diagnostic and therapeutic implications. *Clin J Am Soc Nephrol.* (2020) 16:162– 76. doi: 10.2215/CJN.02840320
- Haut LL, Alfrey AC, Guggenheim S, Buddington B, Schrier N. Renal toxicity of phosphate in rats. *Kidney Int.* (1980) 17:722–31. doi: 10.1038/ki. 1980.85
- Liu H, Fergusson MM, Castilho RM, Liu J, Cao L, Chen J, et al. Augmented Wnt signaling in a mammalian model of accelerated aging. *Science*. (2007) 317:803–6. doi: 10.1126/science.1143578
- Kim JH, Xie J, Hwang KH, Wu YL, Oliver N, Eom M, et al. Klotho may ameliorate proteinuria by targeting Trpc6 channels in podocytes. J Am Soc Nephrol. (2017) 28:140–51. doi: 10.1681/ASN.2015080888
- Benjamin EJ, Muntner P, Alonso A, Bittencourt MS, Callaway CW, Carson AP, et al. Heart disease and stroke statistics-2019 update: a report from the American Heart Association. *Circulation*. (2019) 139:e56– e528. doi: 10.1161/CIR.00000000000659
- Gutierrez OM, Mannstadt M, Isakova T, Rauh-Hain JA, Tamez H, Shah A, et al. Fibroblast growth factor 23 and mortality among patients undergoing hemodialysis. N Engl J Med. (2008) 359:584– 92. doi: 10.1056/NEJMoa0706130

- Isakova T, Xie H, Yang W, Xie D, Anderson AH, Scialla J, et al. Fibroblast growth factor 23 and risks of mortality and end-stage renal disease in patients with chronic kidney disease. J Am Med Assoc. (2011) 305:2432– 9. doi: 10.1001/jama.2011.826
- Parker BD, Schurgers LJ, Brandenburg VM, Christenson RH, Vermeer C, Ketteler M, et al. The associations of fibroblast growth factor 23 and uncarboxylated matrix gla protein with mortality in coronary artery disease: the heart and soul study. *Ann Intern Med.* (2010) 152:640– 8. doi: 10.7326/0003-4819-152-10-201005180-00004
- 64. Grabner A, Amaral AP, Schramm K, Singh S, Sloan A, Yanucil C, et al. Activation of cardiac fibroblast growth factor receptor 4 causes left ventricular hypertrophy. *Cell Metab.* (2015) 22:1020–32. doi: 10.1016/j.cmet.2015.09.002
- Faul C, Amaral AP, Oskouei B, Hu MC, Sloan A, Isakova T, et al. Fgf23 induces left ventricular hypertrophy. J Clin Invest. (2011) 121:4393– 408. doi: 10.1172/JCI46122
- Gutierrez OM, Januzzi JL, Isakova T, Laliberte K, Smith K, Collerone G, et al. Fibroblast growth factor 23 and left ventricular hypertrophy in chronic kidney disease. *Circulation.* (2009) 119:2545–52. doi: 10.1161/CIRCULATIONAHA.108.844506
- Seifert ME, Hruska KA. The kidney-vascular-bone axis in the chronic kidney disease-mineral bone disorder. *Transplantation*. (2016) 100:497– 505. doi: 10.1097/TP.000000000000903
- Zhou C, Shi Z, Ouyang N, Ruan X. Hyperphosphatemia and cardiovascular disease. Front Cell Dev Biol. (2021) 9:644363. doi: 10.3389/fcell.2021.644363
- Turan MN, Kircelli F, Yaprak M, Sisman AR, Gungor O, Bayraktaroglu S, et al. Fgf-23 levels are associated with vascular calcification, but not with atherosclerosis, in hemodialysis patients. *Int Urol Nephrol.* (2016) 48:609– 17. doi: 10.1007/s11255-016-1231-1
- Zieman SJ, Melenovsky V, Kass DA. Mechanisms, pathophysiology, and therapy of arterial stiffness. *Arterioscler Thromb Vasc Biol.* (2005) 25:932– 43. doi: 10.1161/01.ATV.0000160548.78317.29
- Jirak P, Stechemesser L, More E, Franzen M, Topf A, Mirna M, et al. Clinical implications of fetuin-A. Adv Clin Chem. (2019) 89:79– 130. doi: 10.1016/bs.acc.2018.12.003
- Smith ER, Ford ML, Tomlinson LA, Rajkumar C, McMahon LP, Holt SG. Phosphorylated fetuin-a-containing calciprotein particles are associated with aortic stiffness and a procalcific milieu in patients with pre-dialysis Ckd. *Nephrol Dial Transplant*. (2012) 27:1957–66. doi: 10.1093/ndt/gfr609
- Valdivielso JM, Rodriguez-Puyol D, Pascual J, Barrios C, Bermudez-Lopez M, Sanchez-Nino MD, et al. Atherosclerosis in chronic kidney disease: more, less, or just different? *Arterioscler Thromb Vasc Biol.* (2019) 39:1938– 66. doi: 10.1161/ATVBAHA.119.312705
- 74. Di Marco GS, Hausberg M, Hillebrand U, Rustemeyer P, Wittkowski W, Lang D, et al. Increased inorganic phosphate induces human endothelial cell apoptosis *in vitro*. *Am J Physiol Renal Physiol*. (2008) 294:F1381– 7. doi: 10.1152/ajprenal.00003.2008
- Guerin AP, London GM, Marchais SJ, Metivier F. Arterial stiffening and vascular calcifications in end-stage renal disease. *Nephrol Dial Transplant*. (2000) 15:1014–21. doi: 10.1093/ndt/15.7.1014
- Singh S, Grabner A, Yanucil C, Schramm K, Czaya B, Krick S, et al. Fibroblast growth factor 23 directly targets hepatocytes to promote inflammation in chronic kidney disease. *Kidney Int.* (2016) 90:985– 96. doi: 10.1016/j.kint.2016.05.019
- Kimmel PL, Phillips TM, Simmens SJ, Peterson RA, Weihs KL, Alleyne S, et al. Immunologic function and survival in hemodialysis patients. *Kidney Int.* (1998) 54:236–44. doi: 10.1046/j.1523-1755.1998.00981.x
- Pereira BJ, Shapiro L, King AJ, Falagas ME, Strom JA, Dinarello CA. Plasma levels of Il-1 beta, Tnf alpha and their specific inhibitors in undialyzed chronic renal failure, capd and hemodialysis patients. *Kidney Int.* (1994) 45:890–6. doi: 10.1038/ki.1994.117
- 79. Gupta J, Mitra N, Kanetsky PA, Devaney J, Wing MR, Reilly M, et al. Association between albuminuria, kidney function, and inflammatory biomarker profile in Ckd in Cric. *Clin J Am Soc Nephrol.* (2012) 7:1938– 46. doi: 10.2215/CJN.03500412
- Mihai S, Codrici E, Popescu ID, Enciu AM, Albulescu L, Necula LG, et al. Inflammation-related mechanisms in chronic kidney

disease prediction, progression, and outcome. J Immunol Res. (2018) 2018:2180373. doi: 10.1155/2018/2180373

- Rossaint J, Oehmichen J, Van Aken H, Reuter S, Pavenstadt HJ, Meersch M, et al. Fgf23 signaling impairs neutrophil recruitment and host defense during Ckd. J Clin Invest. (2016) 126:962–74. doi: 10.1172/JCI83470
- Malluche HH, Porter DS, Pienkowski D. Evaluating bone quality in patients with chronic kidney disease. *Nat Rev Nephrol.* (2013) 9:671– 80. doi: 10.1038/nrneph.2013.198
- Moe S, Drueke T, Cunningham J, Goodman W, Martin K, Olgaard K, et al. Definition, evaluation, and classification of renal osteodystrophy: a position statement from kidney disease: improving global outcomes (Kdigo). *Kidney Int.* (2006) 69:1945–53. doi: 10.1038/sj.ki.5000414
- Iseri K, Carrero JJ, Evans M, Fellander-Tsai L, Berg HE, Runesson B, et al. Incidence of fractures before and after dialysis initiation. J Bone Miner Res. (2020) 35:2372–80. doi: 10.1002/jbmr.4141
- Ensrud KE, Lui LY, Taylor BC, Ishani A, Shlipak MG, Stone KL, et al. Renal function and risk of hip and vertebral fractures in older women. *Arch Intern Med.* (2007) 167:133–9. doi: 10.1001/archinte.167.2.133
- Reiss AB, Miyawaki N, Moon J, Kasselman LJ, Voloshyna I, D'Avino R Jr, et al. Ckd, Arterial calcification, atherosclerosis and bone health: inter-relationships and controversies. *Atherosclerosis.* (2018) 278:49–59. doi: 10.1016/j.atherosclerosis.2018.08.046
- Malluche HH, Mawad HW, Monier-Faugere MC. Renal osteodystrophy in the first decade of the new millennium: analysis of 630 bone biopsies in black and white patients. J Bone Miner Res. (2011) 26:1368– 76. doi: 10.1002/jbmr.309
- Kurz P, Monier-Faugere MC, Bognar B, Werner E, Roth P, Vlachojannis J, et al. Evidence for abnormal calcium homeostasis in patients with adynamic bone disease. *Kidney Int.* (1994) 46:855–61. doi: 10.1038/ki.1994.342
- Sitara D, Razzaque MS, Hesse M, Yoganathan S, Taguchi T, Erben RG, et al. Homozygous ablation of fibroblast growth factor-23 results in hyperphosphatemia and impaired skeletogenesis, and reverses hypophosphatemia in Phex-deficient mice. *Matrix Biol.* (2004) 23:421–32. doi: 10.1016/j.matbio.2004.09.007
- Sitara D, Kim S, Razzaque MS, Bergwitz C, Taguchi T, Schuler C, et al. Genetic evidence of serum phosphate-independent functions of Fgf-23 on bone. *PLoS Genet.* (2008) 4:e1000154. doi: 10.1371/journal.pgen.1000154
- Wang H, Yoshiko Y, Yamamoto R, Minamizaki T, Kozai K, Tanne K, et al. Overexpression of fibroblast growth factor 23 suppresses osteoblast differentiation and matrix mineralization *in vitro*. J Bone Miner Res. (2008) 23:939–48. doi: 10.1359/jbmr.080220
- Moyses RMA, Dusso A. Is osteocyte Klotho bad for bone health? *Kidney Int.* (2017) 92:540–3. doi: 10.1016/j.kint.2017.04.012
- Vimalraj S. Alkaline phosphatase: structure, expression and its function in bone mineralization. *Gene.* (2020) 754:144855. doi: 10.1016/j.gene.2020.144855
- Murali SK, Andrukhova O, Clinkenbeard EL, White KE, Erben RG. Excessive osteocytic Fgf23 secretion contributes to pyrophosphate accumulation and mineralization defect in Hyp mice. *PLoS Biol.* (2016) 14:e1002427. doi: 10.1371/journal.pbio.1002427
- Yuan Q, Jiang Y, Zhao X, Sato T, Densmore M, Schuler C, et al. Increased osteopontin contributes to inhibition of bone mineralization in Fgf23-deficient mice. J Bone Miner Res. (2014) 29:693–704. doi: 10.1002/ jbmr.2079
- Rhee Y, Bivi N, Farrow E, Lezcano V, Plotkin LI, White KE, et al. Parathyroid hormone receptor signaling in osteocytes increases the expression of fibroblast growth factor-23 *in vitro* and *in vivo*. *Bone*. (2011) 49:636– 43. doi: 10.1016/j.bone.2011.06.025
- Komaba H, Kaludjerovic J, Hu DZ, Nagano K, Amano K, Ide N, et al. Klotho expression in osteocytes regulates bone metabolism and controls bone formation. *Kidney Int.* (2017) 92:599–611. doi: 10.1016/j.kint.2017.02.014
- Mehta R, Cai X, Hodakowski A, Lee J, Leonard M, Ricardo A, et al. Fibroblast growth factor 23 and anemia in the chronic renal insufficiency cohort study. *Clin J Am Soc Nephrol.* (2017) 12:1795–803. doi: 10.2215/CJN.039 50417
- 99. Nam KH, Kim H, An SY, Lee M, Cha MU, Park JT, et al. Circulating fibroblast growth factor-23 levels are associated with an increased risk of anemia

development in patients with nondialysis chronic kidney disease. Sci Rep. (2018) 8:7294. doi: 10.1038/s41598-018-25439-z

- Coe LM, Madathil SV, Casu C, Lanske B, Rivella S, Sitara D. Fgf-23 is a negative regulator of prenatal and postnatal erythropoiesis. *J Biol Chem.* (2014) 289:9795–810. doi: 10.1074/jbc.M113.527150
- 101. Agoro R, Montagna A, Goetz R, Aligbe O, Singh G, Coe LM, et al. Inhibition of fibroblast growth factor 23 (Fgf23) signaling rescues renal anemia. *FASEB* J. (2018) 32:3752–64. doi: 10.1096/fj.201700667R
- 102. Wolfe RA, Ashby VB, Milford EL, Ojo AO, Ettenger RE, Agodoa LY, et al. Comparison of mortality in all patients on dialysis, patients on dialysis awaiting transplantation, and recipients of a first cadaveric transplant. *N Engl J Med.* (1999) 341:1725–30. doi: 10.1056/NEJM199912023412303
- KDIGO Guidelines. Chapter 1: introduction and definition of Ckd–Mbd and the development of the guideline statements. *Kidney Int.* (2009) 76:S3– 8. doi: 10.1038/ki.2009.189
- 104. Chapter 5: evaluation and treatment of kidney transplant bone disease. *Kidney Int.* (2009) 76113:S100–10.
- 105. Trombetti A, Richert L, Hadaya K, Graf JD, Herrmann FR, Ferrari SL, et al. Early post-transplantation hypophosphatemia is associated with elevated Fgf-23 levels. *Eur J Endocrinol.* (2011) 164:839–47. doi: 10.1530/EJE-10-1150
- 106. van Londen M, Aarts BM, Deetman PE, van der Weijden J, Eisenga MF, Navis G, et al. Post-transplant hypophosphatemia and the risk of deathcensored graft failure and mortality after kidney transplantation. *Clin J Am Soc Nephrol.* (2017) 12:1301–10. doi: 10.2215/CJN.10270916
- 107. Han SY, Hwang EA, Park SB, Kim HC, Kim HT. Elevated fibroblast growth factor 23 levels as a cause of early postrenal transplantation hypophosphatemia. *Transplant Proc.* (2012) 44:657–60. doi: 10.1016/j.transproceed.2011.11.046
- 108. Kawarazaki H, Shibagaki Y, Fukumoto S, Kido R, Nakajima I, Fuchinoue S, et al. The relative role of fibroblast growth factor 23 and parathyroid hormone in predicting future hypophosphatemia and hypercalcemia after living donor kidney transplantation: a 1-year prospective observational study. *Nephrol Dial Transplant.* (2011) 26:2691–5. doi: 10.1093/ndt/gfq777
- Wesseling-Perry K, Pereira RC, Tsai E, Ettenger R, Juppner H, Salusky IB. Fgf23 and mineral metabolism in the early post-renal transplantation period. *Pediatr Nephrol.* (2013) 28:2207–15. doi: 10.1007/s00467-013-2547-z
- 110. Bleskestad IH, Thorsen IS, Jonsson G, Skadberg O, Bergrem H, Goransson LG. Soluble Klotho and intact fibroblast growth factor 23 in long-term kidney transplant patients. *Eur J Endocrinol.* (2015) 172:343–50. doi: 10.1530/EJE-14-0457
- 111. Vezzoli G, Elli A, Palazzi P, Bertoni T, Scabini M, Quarto di Palo F, et al. High plasma ionized calcium with normal Pth and total calcium levels in normal-function kidney transplant recipients. *Nephron.* (1986) 42:290– 4. doi: 10.1159/000183690
- 112. Stavroulopoulos A, Cassidy MJ, Porter CJ, Hosking DJ, Roe SD. Vitamin D status in renal transplant recipients. Am J Transplant. (2007) 7:2546– 52. doi: 10.1111/j.1600-6143.2007.01978.x
- Boom H, Mallat MJ, de Fijter JW, Paul LC, Bruijn JA, van Es LA. Calcium levels as a risk factor for delayed graft function. *Transplantation*. (2004) 77:868–73. doi: 10.1097/01.TP.0000116417.03114.87
- 114. Messa P, Cafforio C, Alfieri C. Calcium and phosphate changes after renal transplantation. *J Nephrol.* (2010) 23(Suppl.16):S175–81.
- 115. Torregrosa JV, Barros X. Management of hypercalcemia after renal transplantation. *Nefrologia*. (2013) 33:751–7. doi: 10.3265/Nefrologia.pre2013.Aug.11888
- 116. Tartaglione L, Pasquali M, Rotondi S, Muci ML, Leonangeli C, Farcomeni A, et al. Interactions of sclerostin with Fgf23, soluble Klotho and vitamin D in renal transplantation. *PLoS ONE*. (2017) 12:e0178637. doi: 10.1371/journal.pone.0178637
- 117. Evenepoel P, Meijers BK, de Jonge H, Naesens M, Bammens B, Claes K, et al. Recovery of hyperphosphatoninism and renal phosphorus wasting one year after successful renal transplantation. *Clin J Am Soc Nephrol.* (2008) 3:1829–36. doi: 10.2215/CJN.01310308
- 118. Sanchez Fructuoso AI, Maestro ML, Perez-Flores I, Valero R, Rafael S, Veganzones S, et al. Serum level of fibroblast growth factor 23 in

maintenance renal transplant patients. Nephrol Dial Transplant. (2012) 27:4227-35. doi: 10.1093/ndt/gfs409

- 119. Malyszko J, Koc-Zorawska E, Matuszkiewicz-Rowinska J, Malyszko J. Fgf23 and Klotho in relation to markers of endothelial dysfunction in kidney transplant recipients. *Transplant Proc.* (2014) 46:2647–50. doi: 10.1016/j.transproceed.2014.09.015
- 120. Wolf M, Molnar MZ, Amaral AP, Czira ME, Rudas A, Ujszaszi A, et al. Elevated fibroblast growth factor 23 is a risk factor for kidney transplant loss and mortality. J Am Soc Nephrol. (2011) 22:956– 66. doi: 10.1681/ASN.2010080894
- 121. Baia LC, Humalda JK, Vervloet MG, Navis G, Bakker SJ, de Borst MH, et al. Fibroblast growth factor 23 and cardiovascular mortality after kidney transplantation. *Clin J Am Soc Nephrol.* (2013) 8:1968– 78. doi: 10.2215/CJN.01880213
- 122. Yilmaz MI, Sonmez A, Saglam M, Yaman H, Kilic S, Turker T, et al. Longitudinal analysis of vascular function and biomarkers of metabolic bone disorders before and after renal transplantation. *Am J Nephrol.* (2013) 37:126–34. doi: 10.1159/000346711
- Parfrey PS, Harnett JD, Foley RN, Kent GM, Murray DC, Barre PE, et al. Impact of renal transplantation on uremic cardiomyopathy. *Transplantation*. (1995) 60:908–14. doi: 10.1097/00007890-199511000-00005
- 124. Zolty R, Hynes PJ, Vittorio TJ. Severe left ventricular systolic dysfunction may reverse with renal transplantation: uremic cardiomyopathy and cardiorenal syndrome. Am J Transplant. (2008) 8:2219–24. doi: 10.1111/j.1600-6143.2008.02407.x
- 125. Baloglu I, Ozer H, Ozturk Y, Erdur MF, Tonbul HZ, Turkmen K. The relationship between Fgf23 and anemia in Hd and renal transplant patients. *Int Urol Nephrol.* (2021) 21:9. doi: 10.1007/s11255-021-02982-9
- 126. Tan SJ, Crosthwaite A, Langsford D, Obeysekere V, Ierino FL, Roberts MA, et al. Mineral adaptations following kidney transplantation. *Transpl Int.* (2017) 30:463–73. doi: 10.1111/tri.12925
- 127. Mizusaki K, Hasuike Y, Kimura T, Nagasawa Y, Kuragano T, Yamada Y, et al. Inhibition of the mammalian target of rapamycin may augment the increase in soluble Klotho levels in renal transplantation recipients. *Blood Purif.* (2019) 47(Suppl.2):12–8. doi: 10.1159/000496630
- 128. Kim SM, Kim SJ, Ahn S, Min S-I, Min S-K, Ha J. The potential role of klotho as a prognostic biomarker in deceased donor kidney transplantation. *Transplantation*. (2018) 102(Suppl.7S-1):S539– S. doi: 10.1097/01.tp.0000543389.60945.78
- 129. Sugiura H, Yoshida T, Mitobe M, Yoshida S, Shiohira S, Nitta K, et al. Klotho reduces apoptosis in experimental ischaemic acute kidney injury *via* Hsp-70. *Nephrol Dial Transplant.* (2010) 25:60–8. doi: 10.1093/ndt/gfp451
- Sugiura H, Yoshida T, Tsuchiya K, Mitobe M, Nishimura S, Shirota S, et al. Klotho reduces apoptosis in experimental ischaemic acute renal failure. *Nephrol Dial Transplant*. (2005) 20:2636–45. doi: 10.1093/ndt/gfi165
- 131. Hu MC, Shi M, Zhang J, Quinones H, Kuro-o M, Moe OW. Klotho deficiency is an early biomarker of renal ischemia-reperfusion injury and its replacement is protective. *Kidney Int.* (2010) 78:1240–51. doi: 10.1038/ki.2010.328
- 132. Deng G, Yang A, Wu J, Zhou J, Meng S, Zhu C, et al. The value of older donors' Klotho level in predicting recipients' short-term renal function. *Med Sci Monit.* (2018) 24:7936–43. doi: 10.12659/MSM. 913274
- 133. Rojas E, Carlini RG, Clesca P, Arminio A, Suniaga O, De Elguezabal K, et al. The pathogenesis of osteodystrophy after renal transplantation as detected by early alterations in bone remodeling. *Kidney Int.* (2003) 63:1915–23. doi: 10.1046/j.1523-1755.2003.0 0938.x
- Bellorin-Font E, Rojas E, Carlini RG, Suniaga O, Weisinger JR. Bone remodeling after renal transplantation. *Kidney Int Suppl.* (2003) 85:S125– 8. doi: 10.1046/j.1523-1755.63.s85.30.x
- Nikkel LE, Hollenbeak CS, Fox EJ, Uemura T, Ghahramani N. Risk of fractures after renal transplantation in the United States. *Transplantation*. (2009) 87:1846–51. doi: 10.1097/TP.0b013e3181a6bbda
- Mikuls TR, Julian BA, Bartolucci A, Saag KG. Bone mineral density changes within six months of renal transplantation. *Transplantation*. (2003) 75:49– 54. doi: 10.1097/00007890-200301150-00009

- Julian BA, Laskow DA, Dubovsky J, Dubovsky EV, Curtis JJ, Quarles LD. Rapid loss of vertebral mineral density after renal transplantation. N Engl J Med. (1991) 325:544–50. doi: 10.1056/NEJM199108223250804
- 138. Brandenburg VM, Politt D, Ketteler M, Fassbender WJ, Heussen N, Westenfeld R, et al. Early rapid loss followed by long-term consolidation characterizes the development of lumbar bone mineral density after kidney transplantation. *Transplantation.* (2004) 77:1566–71. doi: 10.1097/01.TP.0000131990.13277.28
- Weinstein RS, Jilka RL, Parfitt AM, Manolagas SC. Inhibition of osteoblastogenesis and promotion of apoptosis of osteoblasts and osteocytes by glucocorticoids. Potential mechanisms of their deleterious effects on bone. *J Clin Invest.* (1998) 102:274–82. doi: 10.1172/JCI2799
- 140. Canalis E, Delany AM. Mechanisms of glucocorticoid action in bone. Ann N Y Acad Sci. (2002) 966:73–81. doi: 10.1111/j.1749-6632.2002. tb04204.x
- 141. Malluche HH, Monier-Faugere MC, Herberth J. Bone disease after renal transplantation. Nat Rev Nephrol. (2010) 6:32– 40. doi: 10.1038/nrneph.2009.192
- 142. Westeel FP, Mazouz H, Ezaitouni F, Hottelart C, Ivan C, Fardellone P, et al. Cyclosporine bone remodeling effect prevents steroid osteopenia after kidney transplantation. *Kidney Int.* (2000) 58:1788–96. doi: 10.1046/j.1523-1755.2000.00341.x
- 143. Marcen R, Caballero C, Pascual J, Teruel JL, Tenorio M, Ocana J, et al. Lumbar bone mineral density in renal transplant patients on neoral and tacrolimus: a four-year prospective study. *Transplantation*. (2006) 81:826– 31. doi: 10.1097/01.tp.0000203557.36884.e3
- 144. Ugur A, Guvener N, Isiklar I, Turan M, Erdal R, Haberal M. Osteoporosis after renal transplantation: single center experience. *Transplantation*. (2001) 71:645–9. doi: 10.1097/00007890-200103150-00011
- 145. Vautour LM, Melton LJ 3rd, Clarke BL, Achenbach SJ, Oberg AL, McCarthy JT. Long-term fracture risk following renal transplantation: a population-based study. Osteoporos Int. (2004) 15:160–7. doi: 10.1007/s00198-003-1532-y
- 146. Patel S, Kwan JT, McCloskey E, McGee G, Thomas G, Johnson D, et al. Prevalence and causes of low bone density and fractures in kidney transplant patients. *J Bone Miner Res.* (2001) 16:1863–70. doi: 10.1359/jbmr.2001.16.10.1863
- 147. Briner VA, Thiel G, Monier-Faugere MC, Bognar B, Landmann J, Kamber V, et al. Prevention of cancellous bone loss but persistence of renal bone disease despite normal 1,25 vitamin D levels two years after kidney transplantation. *Transplantation*. (1995) 59:1393– 400. doi: 10.1097/00007890-199505270-00006
- Bouquegneau A, Salam S, Delanaye P, Eastell R, Khwaja A. Bone disease after kidney transplantation. *Clin J Am Soc Nephrol.* (2016) 11:1282– 96. doi: 10.2215/CJN.11371015
- 149. Kanaan N, Claes K, Devogelaer JP, Vanderschueren D, Depresseux G, Goffin E, et al. Fibroblast growth factor-23 and parathyroid hormone are associated with post-transplant bone mineral density loss. *Clin J Am Soc Nephrol.* (2010) 5:1887–92. doi: 10.2215/CJN.00950110
- Lentine KL, Patel A. Risks and outcomes of living donation. Adv Chronic Kidney Dis. (2012) 19:220–8. doi: 10.1053/j.ackd.2011. 09.005

- 151. Segev DL, Muzaale AD, Caffo BS, Mehta SH, Singer AL, Taranto SE, et al. Perioperative mortality and long-term survival following live kidney donation. J Am Med Assoc. (2010) 303:959–66. doi: 10.1001/jama. 2010.237
- 152. Ibrahim HN, Foley R, Tan L, Rogers T, Bailey RF, Guo H, et al. Longterm consequences of kidney donation. N Engl J Med. (2009) 360:459– 69. doi: 10.1056/NEJMoa0804883
- 153. O'Keeffe LM, Ramond A, Oliver-Williams C, Willeit P, Paige E, Trotter P, et al. Mid- and long-term health risks in living kidney donors: a systematic review and meta-analysis. *Ann Intern Med.* (2018) 168:276– 84. doi: 10.7326/M17-1235
- 154. Muzaale AD, Massie AB, Wang MC, Montgomery RA, McBride MA, Wainright JL, et al. Risk of end-stage renal disease following live kidney donation. J Am Med Assoc. (2014) 311:579–86. doi: 10.1001/jama.2013.285141
- 155. Arias E, Tejada-Vera B, Ahmad F. Provisional Life Expectancy Estimates for January through June, 2020. Vital Statistics Rapid Release no 10. Hyattsville, MD: National Center for Health Statistics. (2020). doi: 10.15620/cdc: 100392
- 156. Hiemstra TF, Smith JC, Lim K, Xu D, Kulkarni S, Bradley JA, et al. Effect of kidney donation on bone mineral metabolism. *PLoS ONE*. (2020) 15:e0235082. doi: 10.1371/journal.pone.0235082
- 157. Thorsen IS, Bleskestad IH, Jonsson G, Skadberg O, Goransson LG. Neutrophil gelatinase-associated lipocalin, fibroblast growth factor 23, and soluble klotho in long-term kidney donors. *Nephron Extra.* (2016) 6:31– 9. doi: 10.1159/000450621
- 158. Young A, Hodsman AB, Boudville N, Geddes C, Gill J, Goltzman D, et al. Bone and mineral metabolism and fibroblast growth factor 23 levels after kidney donation. *Am J Kidney Dis.* (2012) 59:761–9. doi: 10.1053/j.ajkd.2011.09.019
- Westerberg PA, Ljunggren O, Larsson TE, Wadstrom J, Linde T. Fibroblast growth factor-23 and mineral metabolism after unilateral nephrectomy. *Nephrol Dial Transplant.* (2010) 25:4068–71. doi: 10.1093/ndt/gfq288

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Gupta, Orozco, Rao, Gedaly, Malluche and Neyra. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.