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RESEARCH

Sertility

L-carnitine and L-acetylcarnitine supplementation for idiopathic male infertility

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Abstract

Fifteen percent of couples are globally estimated to be infertile, with up to half of these cases attributed to male infertility. Reactive oxidative species (ROS) are known to damage sperm leading to impaired quantity and quality. Although not routinely assessed, oxidative stress is a common underlying pathology in infertile men. Antioxidants have been shown to improve semen analysis parameters by reducing ROS and facilitating repair of damage caused by oxidative stress, but it remains unclear whether they improve fertility. Carnitines are naturally occurring antioxidants in mammals and are normally abundant in the epididymal luminal fluid of men. We conducted a systematic review and meta-analysis to evaluate the safety and efficacy of carnitine supplementation for idiopathic male infertility. We searched ClinicalKey, ClinicalTrials.gov, Cochrane Central Register of Controlled Trials (CENTRAL), EMBASE, MEDLINE, PubMed and ScienceDirect for relevant studies published from 1 January 2000 to 30 April 2020. Of the articles retrieved, only eight randomised controlled trials were identified and included. Analysis showed that carnitines significantly improve total sperm motility, progressive sperm motility and sperm morphology, but without effect on sperm concentration. There was no demonstrable effect on clinical pregnancy rate in the five studies that included that outcome, although patient numbers were limited. Therefore, the use of carnitines in male infertility appears to improve some sperm parameters but without evidence of an increase in the chance of natural conception.

Lay summary

Although male infertility affects 1:15 men, there is no obvious reason in the vast majority of cases. Reactive oxidative species (ROS) are highly active molecules containing oxygen and are natural byproducts of normal metabolism. However, high concentrations of ROS have been shown to damage sperm, which negatively impacts a couple's ability to conceive. Carnitines are natural antioxidants found in the body that counterbalance the damaging effects of ROS. We conducted a comprehensive review of published studies to assess whether carnitine supplements are safe and effective in improving sperm quality and pregnancy rates. Our analysis shows that carnitines improve sperm swimming and production of normal-shaped sperm cells but do not affect sperm count or pregnancy rates, although there are only a few studies and scientific evidence is limited. Whilst it is possible that carnitines may benefit male infertility, more evidence is required regarding chances of pregnancy after carnitine therapy.

Key Words:
 antioxidants

▶ carnitine male infertility

reactive oxidative species ▶ sperm

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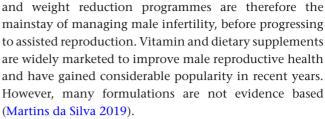


Introduction

Infertility is the inability to conceive naturally within 1 year for a sexually active couple not using contraception (Rowe et al. 2000, World Health Organization 2020). Worldwide, 15% of couples are estimated to be infertile and approximately 50% of these cases are due to male factor, either as the sole underlying cause or a contributory factor (Agarwal et al. 2015). Diagnosis of male infertility usually follows semen analysis. The results may show abnormal semen parameters such as oligozoospermia, asthenozoospermia teratozoospermia and or combination of these, or a complete absence of sperm in the ejaculate (azoospermia), which is identified in 10-15% of infertile men (Rowe et al. 2000, Gudeloglu & Parekattil 2013, Colpi et al. 2018). Notably, up to 75% of male infertility is thought to be idiopathic (i.e. with no cause identified) (Punab et al. 2017).

Oxidative stress (OS) occurs when there is an overproduction of oxidative free radicals and ROS, which damage spermatozoa and cause male infertility by impairing both the structure and function of sperm (Aitken et al. 2003, Aitken & Baker 2006, Valko et al. 2007, Venkatesh et al. 2011, Agarwal et al. 2014). Although the exact mechanism(s) of OS in reducing sperm quality is unknown, it is widely acknowledged that depleted intracellular ATP levels, insufficient axoneme phosphorylation and lipid peroxidation of the cell membrane manifests as poor motility and sperm dysfunction, including reduced ability of sperm to fertilise the oocyte (Storey 1997, Gomez et al. 1998, Valko et al. 2007). Sperm are vulnerable to OS as they have minimal cytoplasm and endogenous antioxidant protection (Martins da Silva 2019). This leads to the production of malondialdehyde (MDA) and 4-hydroxynonenal (4HNE), which oxidises the lipid membrane and causes fragmentation of both nuclear and mitochondrial DNA in sperm (de Lamirande & Gagnon 1992, Kodama et al. 1997, Gomez et al. 1998, Aitken et al. 2012, Iommiello et al. 2015).

There are currently no clinically established treatments available for unexplained male infertility (Martins da Silva *et al.* 2017). Empirical medical treatments such as human menopausal gonadotrophin (hMG)/ human chorionic gonadotrophin (hCG), androgen, antioestrogens (clomiphene and tamoxifen), prolactin inhibitors (bromocriptine), and steroids have been used. However, beneficial effects on semen parameters are not proven (Isidori *et al.* 2006, Jungwirth *et al.* 2012). Lifestyle modification advice such as smoking, alcohol cessation



Carnitines are naturally occurring compounds in mammals (Bremer 1983, Reuter & Evans 2012). Primary sources of carnitines are through dietary intake, de novo biosynthesis, and renal tubular reabsorption (Reuter & Evans 2012). Foods rich in carnitines include red meats, fish, poultry and dairy products (Steiber et al. 2004). Aside from dietary consumption, approximately 25% of total body carnitine is synthesised by the body from the essential amino acids lysine and methionine (Vaz & Wanders 2002, Shekhawat et al. 2013). Endogenous plasma and tissue concentrations of carnitines are preserved at relatively precise limits to facilitate mitochondrial and peroxisomal fatty acid oxidation (Bremer 1983, Reuter & Evans 2012). L-carnitine facilitates the β-oxidation of long-chain fatty acids, and in its active form of L-acetylcarnitine, is a vital antioxidant that protects the sperm mitochondria from oxidative stress (Kerner & Hoppel 1998, Russo et al. 2000, Abdelrazik et al. 2009). Carnitines participate in the metabolism of branch-chain amino acids and stabilise cellular membranes (Shalev et al. 1986, Adeva-Andany et al. 2017) and can also act as free radicle scavengers, thereby increasing antioxidative capabilities in spermatozoa resulting in reduction of OS (Balercia et al. 2005, Dokmeci 2005, Adewovin et al. 2017). In vitro, addition of carnitine to culture media increases sperm motility and vitality (Tanphaichitr 1977, Banihani et al. 2014). Notably, men with abnormal semen parameters have been reported to have significantly lower carnitine serum levels (Zopfgen et al. 2000, Mongioi et al. 2016). In this review we have aggregated and analysed currently available data from clinical trials of L-carnitine and/or L-acetylcarnitine in idiopathic male infertility to determine whether carnitine supplements indeed improve sperm quality, and therefore male reproductive potential, in couples with male factor infertility (Haje & Naoom 2015, Moolenaar et al. 2015).

Materials and methods

Our study is based on the Preferred Reporting Items for Systematic Reviews and Meta-Analyses Protocols (PRISMA-P) and the systematic review is reported according to PRISMA guidelines (Moher *et al.* 2015,



Shamseer *et al.* 2015). The study protocol is registered in PROSPERO (CRD42020181104).

Eligibility criteria

Analysis specifically included only Randomised Controlled Trials (RCTs). The RCTs had to be human studies with male patients between the ages of 18 and 65, with abnormal semen characteristics according to WHO normative ranges (2010) and treated with L-carnitine and/or L-acetyl-carnitine (World Health Organization 2010). Studies required at least one control group treated with placebo or without treatment.

Reviews, commentaries, observational studies, retrospective studies, quasi-randomised trials, case series and case reports were excluded. We also excluded literature with animal studies, laboratory and *in vitro* studies, female factor infertility, undiagnosed patients, infertility <1 year, couples with no regular sexual intercourse and other causes of male infertility not related to abnormal semen analysis.

Information sources

Literature search strategies were developed using medical subject heading (MeSH) terms and text relating to the impact of L-carnitine and L-acetylcarnitine on male reproductive potential. We searched ClinicalKey, ClinicalTrials.gov, Cochrane Central Register of Controlled Trials (CENTRAL), EMBASE, MEDLINE, PubMed and ScienceDirect thoroughly according to PRISMA guidelines (Moher et al. 2015, Shamseer et al. 2015). The literature search was limited to the English language and published between 1 January 2000 and 30 April 2020. Articles were also sourced by screening through the references of included studies or relevant reviews during the selection process.

Search strategy

Our MeSH terms were 'male infertility' or 'male reproductive potential' or 'male subfertility' or 'spermatozoa' or 'asthenozoospermia' or 'oligospermia' or 'oligoasthenozoospermia' or 'teratozoospermia' or 'DNA damage' or 'oxidative stress and 'Carnitine' or 'Levocarnitine' or 'L-carnitine' or 'L-acetylcarnitine' or 'L-acetyl Carnitine' or 'L-acetyl-carnitine' or 'L-acetyl Carnitine' or 'Levoacetylcarnitine' or 'Levo-acetylcarnitine' or 'Levoacetyl Carnitine' or 'Levo-acetyl Carnitine' or 'Acetyl-L-carnitine' or 'Acetyl L-carnitine' or 'Acetyl-L Carnitine' or 'Acetyl-Levocarnitine' or 'Acetyl Carnitine'.

Data management and collection

Covidence was used to filter duplicates and conduct the systematic review data collection process (Veritas Health Innovation). The selection was performed according to PRISMA guidelines (Moher et al. 2015, Shamseer et al. 2015). All available pieces of literature were thoroughly screened using the inclusion and exclusion criteria through Covidence (Veritas Health Innovation). Literature was first screened by title and abstract. Full-text articles that fulfilled the inclusion criteria were then reviewed. If two or more reports had repeated data, the study with the largest sample size, most extended follow-up, and most specific intervention and outcomes were selected. The screening and selection process was carried out by two independent review authors simultaneously (Khaw and Wong) using a standardised form to include study characteristics such as methodology, number of participants, demographics of participants, detailed test and control interventions, primary and secondary outcomes of the studies, the effect of treatment and risk of bias. Missing data were requested from study authors. Any discrepancies were resolved through consensus.

Risk of bias in individual studies

The Cochrane Collaboration's tool for assessing the risk of bias in randomised trials was used to determine the six domains of bias (Higgins *et al.* 2011). Two independent review authors conducted this assessment (Khaw and Wong) and any discrepancies were resolved through consensus. Articles were assessed for bias based on the following aspects: random sequence generation, allocation concealment, blinding of participants and personnel, blinding of outcome assessment, incomplete outcome data, selective reporting and other bias (Higgins *et al.* 2011). Each aspect was then classified as high, low, or unclear risk of material bias (Higgins *et al.* 2011). The risk of bias assessment chart was then generated using Review Manager (RevMan) 5.4 software (The Nordic Cochrane Centre 2014).

Data synthesis

We then carried out a descriptive analysis of included studies focusing on the methodology of the study,



type and details of the intervention, target population demographics, primary outcomes, secondary outcomes, adverse outcomes and intervention effects. A meta-analysis was conducted for studies with the same intervention and comparator with equal outcome measures. For the metaanalysis, we conducted a random-effects meta-analysis using risk ratios for dichotomous outcomes and mean differences with s.D. The raw mean differences were used instead of standardised mean differences, as all studies used the same continuous outcomes and units of measure. We then used the statistical significance of 95% CIs and *P*-values for each outcome. Where there were results from multiple durations of therapy, the results after the most prolonged period of treatment was used. Higgins's I² test statistic (>50% indicative of substantial heterogeneity) was utilised to assess heterogeneity among the studies. Cochran's Q test was not used to analyse heterogeneity as there were only small numbers of available studies. We then proceed with a stratified meta-analysis for study quality, trial size, concealment of allocation, blind adjudication of events, analysis according to the intention-to-treat principle, and intervention method. Assessment evidence of publication bias was carried out for the included studies and plots were generated to visually inspect the data through a funnel plot generated by RevMan 5.4 software (The Nordic Cochrane Centre 2014). In study outcomes that had substantial heterogeneity, the data were also synthesised through a narrative and qualitative approach.

The s.D. from Sigman *et al.* (2006) was calculated according to the Cochrane Handbook for

Systematic Reviews of Interventions section 6.5.2.3(3) (Higgins *et al.* 2019).

Overall quality of evidence

The quality and consistency of each comparison was assessed using the Grading of Recommendations Assessment, Development and Evaluation (GRADE) guidelines through GRADEpro (Balshem *et al.* 2011, Schünemann *et al.* 2013, Evidence Prime 2015). The strength of evidence for critical and essential outcomes was rated based on study design, risk of bias, consistency, limitations, directness, reporting precision and publication bias (Balshem *et al.* 2011, Schünemann *et al.* 2013, Evidence Prime 2015). Table 1 shows a summary of the included studies and their GRADE assessments. Effect (risk) of carnitine is expressed as mean difference (MD).

Results

Study characteristics

The search strategy identified 1176 citations dated 1 January 2000 to 30 April 2020. After 440 duplicates were removed, 736 abstracts were assessed. Six hundred and ninety-eight records were excluded as they did not meet inclusion criteria. Thirty-eight full-text articles were searched for eligibility according to inclusion and exclusion criterion. All included studies were of randomised controlled trials without cross over.

Table 1 Summary of findings of carnitine compared to placebo or no treatment for idiopathic male infertility.

Anticipated Risk with placebo or no treatment, range	absolute effects Risk with			Part	ticipants		ertainty
	Risk with	cornitino			•		
no treatment range		i carnitine		in	studies	of	evidence
no a cauneira, range	Value	Range	RR (95% CI)	n	Studies	Evidence	Grade
0.8–33.73 million/mL	MD 2.7 million/mL higher	2.04 lower to 7.44 higher	-	438	6 RCTs	000	VERY LOW ^{a,b,c}
3.3-43.4%	MD 10.72% higher	3.94 higher to 17.5 higher	-	459	7 RCTs	$\oplus \oplus \Theta \Theta$	LOW ^{a,c}
4–24.41%	MD 9.82% higher	2.01 higher to 17.62 higher	-	231	3 RCTs	$\oplus \oplus \ominus \ominus$	LOW ^{a,c}
1.39–32.73%	MD 2.41% higher	0.79 higher to 4.03 higher	-	438	6 RCTs	$\oplus \oplus \Theta \Theta$	LOW ^{a,c}
113 per 1000	116 per 1000	61–221	1.03 (0.54–1.96)	301	5 RCTs	$\oplus \oplus \Theta \Theta$	LOW ^{a,b}
2 2 4	0.8–33.73 million/mL 8.3–43.4% I–24.41% .39–32.73%	0.8-33.73 MD 2.7 million/mL million/mL higher MD 10.72% 8.3-43.4% MD 9.82% H-24.41% MD 9.82% higher MD 2.41% MD 2.41% higher	D.8-33.73 MD 2.7 2.04 lower to million/mL million/mL 7.44 higher higher 3.94 higher to 17.5 higher 8.3-43.4% MD 9.82% 2.01 higher to 1-24.41% MD 9.82% 2.01 higher to .39-32.73% MD 2.41% 0.79 higher to	Notice Name Name 0.8–33.73 MD 2.7 2.04 lower to - million/mL million/mL 7.44 higher higher 3.94 higher to - bigher 17.5 higher - H-24.41% MD 9.82% 2.01 higher to - higher 17.62 higher - .39–32.73% MD 2.41% 0.79 higher to -	Notes Notes <th< td=""><td>Noted and the product of the second secon</td><td>Notedet end (ange)MargeAngeNStearesEndertee$0.8-33.73$MD 2.72.04 lower to-4386 RCTs$\oplus \odot \odot \odot$million/mLmillion/mL7.44 higher-4597 RCTs$\oplus \odot \odot \odot$$0.8-33.4\%$MD 10.72%3.94 higher to-4597 RCTs$\oplus \odot \odot \odot$$0.8-24.41\%$MD 9.82%2.01 higher to-2313 RCTs$\oplus \odot \odot$$0.9-24.41\%$MD 2.41%0.79 higher to-4386 RCTs$\oplus \odot \odot$$0.39-32.73\%$MD 2.41%0.79 higher to-4386 RCTs$\oplus \odot \odot$</td></th<>	Noted and the product of the second secon	Notedet end (ange)MargeAngeNStearesEndertee $0.8-33.73$ MD 2.72.04 lower to-4386 RCTs $\oplus \odot \odot \odot$ million/mLmillion/mL7.44 higher-4597 RCTs $\oplus \odot \odot \odot$ $0.8-33.4\%$ MD 10.72%3.94 higher to-4597 RCTs $\oplus \odot \odot \odot$ $0.8-24.41\%$ MD 9.82%2.01 higher to-2313 RCTs $\oplus \odot \odot$ $0.9-24.41\%$ MD 2.41%0.79 higher to-4386 RCTs $\oplus \odot \odot$ $0.39-32.73\%$ MD 2.41%0.79 higher to-4386 RCTs $\oplus \odot \odot$

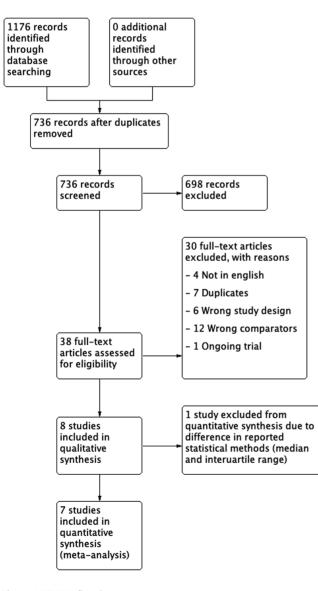
The population was men with abnormal semen characteristics. The intervention was L-carnitine and/or L-acetylcarnitine. The table compares placebo or no treatment. The outcomes measured were semen analysis parameters; clinical pregnancy; adverse events in a clinic or hospital. aLack of blinding; bCrosses the line of no effect; cHiggins's l² test >50%.

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We excluded Lenzi 2003 as it had a cross-over design (Lenzi *et al.* 2003). A total of eight studies were included in the review and the findings from seven studies were pooled into a meta-analysis (Cavallini *et al.* 2004, Lenzi *et al.* 2004, Balercia *et al.* 2005, Sigman *et al.* 2006, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2018). The data by Cavellini *et al.* was excluded from the meta-analysis as the authors reported their results as medians and interquartile ranges rather than means and s.d. (Cavallini *et al.* 2004). Our search process is summarised in the PRISMA flowchart (Fig. 1).

The included articles were assessed based on seven aspects: random sequence generation, allocation concealment, blinding of participants and personnel, blinding of outcome assessment, incomplete outcome

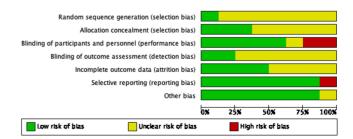


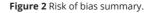
data, selective reporting and other bias (Figs 2 and 3). Where additional information was required, the study authors were contacted but this was unsuccessful. Overall, none of the included studies explicitly mentioned the method of randomisation (Cavallini *et al.* 2004, Lenzi *et al.* 2004, Balercia *et al.* 2005, Sigman *et al.* 2006, Mehni *et al.* 2014, Dimitriadis *et al.* 2010, Haje & Naoom 2015, Tsounapi *et al.* 2018). Hence, all literature had an unclear risk of selection bias in this aspect. The majority of articles also had unclear risks of detection bias as the process of assessment was not reported in detail (Lenzi *et al.* 2004, Sigman *et al.* 2006, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2006, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2016, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2016, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2016, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2015, Tsounapi *et al.* 2016, Dimitriadis *et al.* 2016).

One study (Tsounapi *et al.* 2018) was five-armed, four studies (Balercia *et al.* 2005, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015) were four-armed, one study (Cavallini *et al.* 2004) was three-armed, and two studies (Lenzi *et al.* 2004, Sigman *et al.* 2006) were twoarmed. The total duration of treatment ranged from 3 months (Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2018) to 6 months (Cavallini *et al.* 2004, Lenzi *et al.* 2004, Balercia *et al.* 2005, Haje & Naoom 2015) while follow-up time varied between from 3 months (Mehni *et al.* 2014) to9 months (Cavallini *et al.* 2004, Balercia *et al.* 2005). Table 2 shows an overview of the included studies.

Population

Our meta-analysis only included studies with idiopathic male infertility. The participants were treated with carnitine supplementation, placebo or did not receive any treatment. Six studies (438 men) recorded sperm concentration and morphology, seven studies (459 men) reported total sperm motility and three studies (231 men) evaluated the progressive sperm motility after carnitine therapy. Only four studies (252 men) reported pregnancy outcomes following carnitine supplementation. All participants were between the ages of 18 and 65 with infertility of more than 1 year.







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Figure 1 PRISMA flowchart.

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Blinding of participants and personnel (performance bias) Blinding of outcome assessment (detection bias) Random sequence generation (selection bias) Incomplete outcome data (attrition bias) Allocation concealment (selection bias) Selective reporting (reporting bias) Other bias Balercia 2005 ? Ŧ + + + + Cavallini 2004 + + + + Dimitriadis 2010 ? ? ? ? Haje 2015 ? ? ? ? ? + + Lenzi 2004 ? ? + + + + + Mehni 2014 ? ? ? ? + ? Sigman 2006 ? ? ? + + + + Tsounapl 2018 ? ? ?

Figure 3 Risk of bias graph.

A study by Cavallini *et al.* (2004) was also included in our systematic review, but not in the meta-analysis, as their results were recorded in medians and interquartile ranges. Their study also enrolled men with varicocele but only the results from men with idiopathic male infertility were included in our review (Cavallini *et al.* 2004).

Interventions

Data included in our analysis compared L-carnitine (LC) and/or L-acetylcarnitine (LAC) to placebo or no treatment. Four studies (Cavallini *et al.* 2004, Lenzi *et al.* 2004, Balercia

et al. 2005, Sigman *et al.* 2006) compared LC and LAC to placebos, five studies compared LC to placebo (Balercia *et al.* 2005, Mehni *et al.* 2014, Haje & Naoom 2015) or no treatment (Dimitriadis *et al.* 2010, Tsounapi *et al.* 2018) while only one study (Balercia *et al.* 2005) compared LAC to placebo.

Outcomes

The primary outcomes for our review are sperm concentration, total sperm motility, progressive sperm motility, sperm morphology, pregnancy rate and live birth rate. We also included sperm DNA damage and adverse events such as side effects and miscarriage as secondary outcomes. Seven studies (Lenzi et al. 2004, Balercia et al. 2005, Sigman et al. 2006, Dimitriadis et al. 2010, Mehni et al. 2014, Haje & Naoom 2015, Tsounapi et al. 2018) recorded the total sperm motility while only four studies (Cavallini et al. 2004, Lenzi et al. 2004, Balercia et al. 2005, Tsounapi et al. 2018) reported progressive sperm motility. Sperm concentration and morphology were recorded by seven studies (Cavallini et al. 2004, Lenzi et al. 2004, Balercia et al. 2005, Dimitriadis et al. 2010, Mehni et al. 2014, Haje & Naoom 2015, Tsounapi et al. 2018). None of the studies included DNA damage assessment. Although five studies (Cavallini et al. 2004, Balercia et al. 2005, Sigman et al. 2006, Haje & Naoom 2015, Tsounapi et al. 2018) reported on pregnancy rate, none included live birth or miscarriage data. We contacted the authors where details were unclear or if a different statistical approach was used in their study; however, we received no response.

Sperm concentration

Seven studies reported the effects of carnitines on sperm concentration (Fig. 4) (Cavallini *et al.* 2004, Lenzi *et al.* 2004, Balercia *et al.* 2005, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2018). Although the study by Cavallini *et al.* reported higher sperm concentrations after LC+LAC therapy (20.6%, IQR 24.9–15.1%) when compared to a placebo (10.9%, IQR 15.1–9.0%), findings were reported as median and interquartile range rather than mean and s.D. and no statistical analysis was reported (Cavallini *et al.* 2004). This study was therefore excluded from the meta-analysis.

Overall, our findings showed that carnitines did not significantly improve sperm concentration (P > 0.05). However, the six studies showed a high heterogeneity (MD 2.70, 95% CI -2.04 to 7.44; n = 438, RCT=6,



Tabl	Table 2 Study characteristics.	teristics.								
		Study					Duration of	Sample size	size	
	Study	design	Age (years)	Treatment/day	Control /	Arms	treatment (Weeks)	Therapy	Control	Therapy Control Total follow-up time
2004	Cavallini <i>et al.</i> (2004)	RCT	27-40	2 g LC + 1 g LAC	Placebo	m	24	39	47	9 months
2004	_	RCT	20-40	2 g LC + 1 g LAC	Placebo	2	24	30	26	8 months
2005	Balercia <i>et al.</i> (2005)	RCT	24-38	2 g LC + 1 g LAC (<i>n</i> = 14) vs 3 g LC (<i>n</i> = 15) vs 3 g LAC (<i>n</i> = 15)	Placebo	4	24	44	15	9 months
2006	Sigman <i>et al.</i> (2006)	RCT	36.2 ± 1.7	2 g LC + 1 g LAC	Placebo	7	16	12	б	6 months
2010		RCT	NR	1 g LC	No TT	4	12	26	22	13 weeks (6 days after the experimental period)
2014	2	RCT	25-40	1 g LC	Placebo	4	12	51	59	3 months
2015	±	RCT	37.54 ± 2.46 1 g LC	1 g LC	Placebo	4	12-24	20	29	4–7 months (as two samples were taken after treatment – 1 month apart)
2018	2018 Tsounapi <i>et al.</i> (2018)	RCT	NR	1 g LC	TT oN	ъ	12.8	44	42	Experimental period of 90 days; up to 180 days for pregnancy rate

P > 0.05, $I^2 = 97\%$) (Lenzi *et al.* 2004, Balercia *et al.* 2005, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2018) primarily due to two studies with very small s.d. compared to the others (Mehni *et al.* 2014, Haje & Naoom 2015). A sensitivity analysis after removal of these two studies showed homogeneity between the remaining studies but the meta-analysis still showed no significant effects of carnitines on sperm concentration (MD 0.79, 95% CI –0.39 to 1.96; n = 279, RCT = 4, P > 0.05, $I^2 = 0\%$) (Lenzi *et al.* 2004, Balercia *et al.* 2005, Dimitriadis *et al.* 2010, Tsounapi *et al.* 2018).

Total sperm motility

Seven studies compared the efficacy of carnitines to placebo or no treatment (Lenzi *et al.* 2004, Balercia *et al.* 2005, Sigman *et al.* 2006, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2018). Analysis of the mean difference in total sperm motility showed that carnitines improved total sperm motility by 10.72% (95% CI 3.94–17.50; n = 459, RCT=7, P < 0.05, $I^2 = 97\%$) (Fig. 5).

The studies showed high heterogeneity. In studies comparing LC and LAC to placebo, Balercia et al. showed a significant increase in total sperm motility in all three study arms when compared to a placebo (MD 18.75, 95% CI 14.78–22.73; *n* = 30, *P* < 0.05) (Balercia *et al.* 2005). However two other studies did not show significant differences between the treatment and control groups (Lenzi et al. 2004, Sigman et al. 2006). Lenzi et al. showed a mean difference of 1.56 (95% CI -4.48 to 7.60; n = 56, P > 0.05), while Sigman *et al.* showed a mean difference of -7.70 (95% CI -20.68 to 5.28; n = 21, P > 0.05). In studies that compared LC to placebo or no treatment, four studies (Balercia et al. 2005, Dimitriadis et al. 2010, Mehni et al. 2014, Haje & Naoom 2015) showed significant improvements after receiving LC while one study (Tsounapi et al. 2018) reported no significant differences. Balercia et al. (2005) was the only study that assessed LAC treatment alone.

A detailed statistical analysis is shown in Fig. 5.

Progressive sperm motility

Four studies showed an increase in progressive sperm motility after carnitine when compared to control groups (Cavallini *et al.* 2004, Lenzi *et al.* 2004, Balercia *et al.* 2005, Tsounapi *et al.* 2018). Overall, carnitines significantly improved progressive sperm motility in idiopathic male

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Not reported; TT, treatment.

NR,



infertility (MD 9.82, 95% CI 2.01, 17.62; *n* = 231, *P* < 0.05) (Fig. 6).

This outcome also showed high heterogeneity (I² = 94%). Balercia *et al.* (2005) showed significant improvement in progressive sperm motility in all LC, LAC and LC+LAC therapy groups (MD 16.02, 95% CI 11.98–20.06; n = 30, P < 0.05). Tousnapi *et al.* (2018) reported a significant increase in progressive sperm motility after LC therapy (MD 2.00, 95% CI 0.93–3.07; n = 86, P < 0.05). Lenzi *et al.* (2004) did not show a significant increase in progressive sperm motility after LC+LAC therapy (MD 0.59, 95% CI –5.30 to 6.48; n = 56, P > 0.05). Cavallini *et al.* (2004) reported an increase in progressive sperm motility after LC+LAC therapy (MD 0.59, 95% CI –5.30 to 6.48; n = 56, P > 0.05). Cavallini *et al.* (2004) reported an increase in progressive sperm motility after LC+LAC therapy (3.6%, IQR 28.9–16.0%) when compared to controls (13.2%, 18.6–9.0%) but no raw data or *P*-values were provided to draw a statistically significant conclusion.

Sperm morphology

The results of sperm morphology were recorded by seven studies (Cavallini *et al.* 2004, Lenzi *et al.* 2004, Balercia *et al.* 2005, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2018). Overall, our results showed a significant improvement in sperm morphology (P < 0.05) as seen in Fig. 7.

This outcome also had a high heterogeneity (I² = 91%). Balercia *et al.* (2005) documented a significant improvement of 9.29% when compared to a placebo (MD 9.29, 95% CI 6.51–12.06; n = 30, P < 0.05).

Lenzi *et al.* (2004) showed no significant differences between the treatment and control groups after LC+LAC therapy. Among the remaining studies of treatment after LC, only the study by Haje and Naoom (2015) showed a significant improvement (MD 2.14, 95% CI 1.99–2.29; n = 49, P < 0.05) in sperm morphology; while three other studies (Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Tsounapi *et al.* 2018) recorded no significant changes. Cavallini *et al.* (2004) recorded an improvement of sperm morphology in their study after LC+LAC therapy (27.3%, IQR 32.0–22.6% vs 15.3%, IQR 22.0–12.1% in the placebo group) but the data provided is not sufficient for a test of statistical significance.

Clinical pregnancy rate

A total of five RCTs were analysed for their reported pregnancies and four studies were consolidated into a meta-analysis. There was low heterogeneity ($I^2 = 0\%$) and some concern with the risk of bias. The meta-analysis (Fig. 8) showed that there was no significant improvement in pregnancy rates when compared to control groups (RR 1.17, 95% CI 0.55–2.46; n = 252, RCT = 4, P > 0.05).

In patients treated with a combination of LC and LAC, Cavallini *et al.* (2004) showed a significant improvement in pregnancy rate ($X^2 = 20.795$, P < 0.01) when compared to controls. In contrast, the three other studies (Lenzi *et al.* 2004, Balercia *et al.* 2005, Sigman *et al.* 2006) did not show significant differences between the two groups (RR 1.89, 95% CI 0.67–5.36; n = 106, RCT=3, P > 0.05, $I^2 = 0\%$).

		rnitine			ntrols			Mean Difference		Mean Difference	Risk of Bias
Study or Subgroup	Mean [million/mL]	SD [million/mL]	Total	Mean [million/mL]	SD [million/mL]	Total	Weight	IV, Random, 95% CI	Year	IV, Random, 95% CI	ABCDEFG
2.3.1 LC + LAC											
Lenzi 2004	22.09	9.05	30		16.95	26	12.0%				? • • ? • • •
Balercia 2005 Subtotal (95% CI)	37.4	16.42	1 4 44		14.36	15 41	8. 6% 20.5%		2005	•	? • • • • • • •
	= 0.00; Chl ² = 0.30, d : Z = 0.33 (P = 0.74)	f = 1 (P = 0.58); ŕ	° = 0%								
2.3.2 LC											
Balercia 2005	45.53	21.42	15	33.73	14.36	15	7.4%	11.80 [-1.25, 24.85]	2005		?99999
Dimitriadis 2010	15.4	6.7	26	16.3	7	22	15.0%	-0.90 [-4.80, 3.00]	2010		22022
Mehni 2014	9.3	1.7	51	0.8	1.8	59	16.6%	8.50 [7.85, 9.15]	2014	•	? ? ? ? ? ? ?
Haje 2015	6.17	2.22	20	8.2	2.69	29	16.4%	-2.03 [-3.41, -0.65]	2015	+	2 2 2 3 3
T sounapi 2018 Subtotal (95% CI)	6.6	3.1	44 156		2.9	42 167	16.4% 71.7%		2018	-	?? 🗣 ? ? 🗣 ?
	= 36.24; Chl ² = 258.9 : Z = 0.94 (P = 0.35)	3, df = 4 (P < 0.0)	0001);	r² = 98%							
2.3.3 LAC											
Balercia 2005 Subtotal (95% CI)	39.57	19.99	15 15	33.73	14.36	15 15	7.7% 7.7%		2005		? • • • • • • •
Heterogeneity: Not ap Test for overall effect	plicable : Z = 0.92 (P = 0.36)										
Total (95% CI)			215			223	100.0%	2.70 [-2.04, 7.44]		•	
	- 35.20; Chi ² = 261.2	2, df = 7 (P < 0.0	0001);	r ² = 97%						-20 -10 0 10 20	
	Z = 1.12 (P = 0.26)									Favours Controls Favours Carnitine	
	ferences: Chl ² = 0.50,	df = 2 (P = 0.78),	۳ = 0	×							
Risk of bias legend											
	e generation (selection	bias)									
(B) Allocation conceal											
	pants and personnel (p										
	ne assessment (detecti	on bias)									
(E) Incomplete outcom	ne data (attrition bias)										

(F) Selective reporting (reporting bias)

Figure 4 Forest plot of comparison for sperm concentration.



⁽G) Other bias

Carnitine for idiopathic ma infertility **1**:1

		rnitine			ntrols			Mean Difference		Mean Difference	Risk of Bias
Study or Subgroup	Mean [%]	SD [%]	Total	Mean [%]	SD [%]	Total	Weight	IV, Random, 95% CI	Year	IV, Random, 95% CI	ABCDEFO
2.1.1 LC + LAC											
Lenzi 2004		13.46	30		9.5	26	11.3%	1.56 [-4.48, 7.60]		_ 	?~~?~~~~~~~~~~~~~
Balercia 2005		9.07	14		9.85	15		17.67 [10.78, 24.56]			?..........
Sigman 2006 Subtotal (95% CI)	32.3	19.39	12 56		10.61	9 50	8.5X 30.8%	-7.70 [-20.68, 5.28] 4.59 [-9.05, 18.22]	2006		??
Heterogeneity: Tau ² -	124.87; C	'hi² = 17	.25, di	F = 2 (P = 0	.0002);	r ² = 66	×				
lest for overall effect	: Z = 0.66 (P = 0.51	0								
2.1.2 LC											
Balercia 2005	64.53	8.41	15	43.4	9.85	15	11.1%	21.13 [14.58, 27.68]	2005		?99999
Dimitriadis 2010	35.6	15.5	26	24.7	10.8	22	10.8%	10.90 [3.43, 18.37]	2010	│ _ •	??
Mehni 2014	24.6	1.5	51		2.7	59	12.4%	21.30 [20.50, 22.10]	2014	•	??????
Haje 2015	23.33	3.63	20		4.96	29	12.2%	8.40 [5.99, 10.81]	2015	-	??????
Tsounapi 2018	19	9	44		6	42	12.0%	2.00 [-1.60, 5.60]	2018		22022
Subtotal (95% CI)			156			167	58.4%	12.70 [3.81, 21.59]			
Heterogeneity: Tau ² = Test for overall effect 2.1.3 LAC					.00001)						
	~~ ~~								2005		
Balercia 2005 Subtotal (95% CI)	60.43	10.46	15 15		9.85	15 15	10.6% 10.8%	17.03 [9.76, 24.30] 17.03 [9.76, 24.30]	2005	•	?99999
Heterogeneity: Not ap Test for overall effect		P < 0.00)001)								
Total (95% CI)			227			232	100.0%	10.72 [3.94, 17.50]			
Heterogeneity: Tau ² -	• 96.54: Ch	r ² = 245	.30. df	F = 8 (P < 0	.00001)	: l ² = 9	7%				
Test for overall effect	Z = 3.10 (P = 0.00)2)			•	-			-20 -10 0 10 20 Favours Controls Favours Carnitine	
Test for subgroup dif	ferences: Ch	1 ² = 2.5	7, df =	2 (P = 0.2	6), i ² = 1	22.2%				Favours Controis Favours Carnitine	
Risk of bias legend											
A) Random sequence	e generation	(selectio	on bias))							
B) Allocation conceal	ment (select	ion bias)									
(C) Blinding of particip	pants and p	ersonnel	(perfo	rmance bias	;)						
(D) Blinding of outcom	ne assessme	ent (dete	ction bi	ias)							
(E) Incomplete outcom			s)								
(F) Selective reporting	(reporting l	bias)									
(G) Other bias											

(G) Other bias

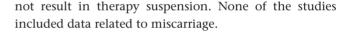
Figure 5 Forest plot of comparison for total sperm motility.

Patients treated with either LC (RR 0.73, 95% CI 0.28– 1.87; n = 165, RCT=3, P > 0.05, $I^2 = 0\%$) or LAC (RR 0.67, 95% CI 0.13–3.44; n = 30, RCT=1, P > 0.05) did not show any significant changes in pregnancy rates in comparison to their control groups (Balercia *et al.* 2005, Haje & Naoom 2015, Tsounapi *et al.* 2018). Haje and Naoom (2015) studied the effect of carnitine supplementation in patients undergoing ICSI and reported no significant increase in pregnancy rates.

Overall, the results were imprecise as there were very few events and thus confidence intervals were wide. Moreover, the quality of evidence is also low.

Adverse events

Six studies did not report adverse events (Lenzi *et al.* 2004, Balercia *et al.* 2005, Dimitriadis *et al.* 2010, Mehni *et al.* 2014, Haje & Naoom 2015, Tsounapi *et al.* 2018). Sigman *et al.* (2006) confirmed that there were no adverse events in their study. Cavallini *et al.* (2004) reported four cases of mild euphoria (two in the LC+LAC group and two in control groups). Their study also recorded two cases of gastrointestinal side effects (mild epigastria and nausea) from both the treatment and control groups. However, these side effects were reported as negligible as they did



Carnitine versus other arms in the included studies

Further studies were identified that compared carnitine to other compounds, rather than placebo or no treatment. They were therefore not included in the meta-analysis but are summarised in Table 3.

Discussion

Several published studies have reported that carnitines have beneficial effects on improving sperm quality in men with idiopathic male infertility (Steiber *et al.* 2004, Isidori *et al.* 2005, Isidori *et al.* 2006, Mongioi *et al.* 2016, Smits *et al.* 2019). Notably, concentrations of carnitine have also been documented to be higher in the sperm and seminal plasma of fertile men, compared to men with abnormal semen parameters (Zopfgen *et al.* 2000, Banihani *et al.* 2014, Mongioi *et al.* 2016, Smits *et al.* 2019). The scientific rationale behind this is a vital role played by carnitines during spermatogenesis (Jeulin & Lewin 1996, Agarwal & Sekhon 2011, Aliabadi *et al.* 2012). Carnitines



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lean [%]				ntrols	_		Mean Difference		Mean Difference	Risk of Bias
icun [/o]	SD [%]	Total	Mean [%]	SD [%]	Total	Weight	IV, Random, 95% CI	Year	IV, Random, 95% CI	ABCDEFG
25	12.00	20	24.41	0.21	20	10.00	A 5A 7 5 3A 6 481	2004		2442444
										2444444
30.13	0.23	44	24	0.5	41	39.0%	7.34 [-5.93, 20.61]	2005		
			1 (P = 0.00))2);	90%					
43.8	7.12	15	24	8.5	15	19.8%	19.80 [14.19, 25.41]	2005		?@@@@@@
6	3	44 59	4	2	42 57	22.0% 41.7%		2018	+	?? @ ?? @ @
			= 1 (P < 0	.00001)	; ² = 9	7%				
37.5	9.2	15 15	24	8.5	15 15	19.2% 19.2%	13.50 [7.16, 19.84] 13.50 [7.16, 19.84]	2005		? @@@@@@
cable = 4.17 (i	P < 0.00	01)								
		118			113	100.0%	9.82 [2.01, 17.62]			
1.92; Chi	² = 61.6	i9. df -	4 (P < 0.0	00001);	r ² = 94	×				
= 2.47 (P = 0.01)								
nces: Ch	r ² = 0.7	1, df =	2 (P = 0.7)	0), f ² = (0%					
		n bias)								
)						
			as)							
)								
porting b	110.5)									
	25 38.13 2.33; Chi = 1.08 (i 43.8 6 54.17; Ci = 1.20 (i 37.5 cable = 4.17 (i 1.92; Chi = 2.47 (i ences: Ch encration tt (selecti ts and pe issessme lata (attri	25 13.06 38.13 8.23 2.33; Ch ² = 9.82 = 1.08 (P = 0.28 43.8 7.12 6 3 54.17; Ch ² = 37 = 1.20 (P = 0.23 37.5 9.2 cable = 4.17 (P < 0.00 1.92; Ch ² = 61.6 = 2.47 (P = 0.01 sinces: Ch ² = 0.72 eneration (selection bias) ts and personnel issessment (deter	25 13.06 30 38.13 8.23 14 44 2.33; Chi ² = 9.82, df = = 1.08 (P = 0.28) 43.8 7.12 15 6 3 44 59 54.17; Chi ² = 37.29, df = 1.20 (P = 0.23) 37.5 9.2 15 15 cable = 4.17 (P < 0.0001) 118 1.92; Chi ² = 61.69, df = = 2.47 (P = 0.01) mces: Chi ² = 0.71, df = eneration (selection bias) ts and personnel (perfor issessment (detection bi lata (attrition bias)	25 13.06 30 24.41 36.13 8.23 14 24 44 2.33; Ch ² = 9.82, df = 1 ($P = 0.0$ ($= 1.08$ ($P = 0.28$) 43.8 7.12 15 24 6 3 44 4 59 54.17; Ch ² = 37.29, df = 1 ($P < 0$ = 1.20 ($P = 0.23$) 37.5 9.2 15 24 15 cable = 4.17 ($P < 0.0001$) 118 1.92; Ch ² = 61.69, df = 4 ($P < 0.6$ = 2.47 ($P = 0.01$) mores: Ch ² = 0.71, df = 2 ($P = 0.76$) eneration (selection bias) tt (selection bias) ts and personnel (performance bias) ts and personnel (performance bias) tata (attrition bias)	25 13.06 30 24.41 9.31 36.13 8.23 14 24 8.5 44 2.33; Ch ² = 9.82, df = 1 ($P = 0.002$); $I^2 =$ = 1.08 ($P = 0.28$) 43.8 7.12 15 24 8.5 6 3 44 4 2 59 54.17; Ch ² = 37.29, df = 1 ($P < 0.00001$) = 1.20 ($P = 0.23$) 37.5 9.2 15 24 8.5 15 cable = 4.17 ($P < 0.0001$) 118 1.92; Ch ² = 61.69, df = 4 ($P < 0.00001$); = 2.47 ($P = 0.01$) mores: Ch ² = 0.71, df = 2 ($P = 0.70$), $I^2 = 0$ eneration (selection bias) tt (selection bias) ts and personnel (performance bias) issessment (detection bias) lata (attrition bias)	25 13.06 30 24.41 9.31 26 38.13 8.23 14 24 8.5 15 44 4 4 2.33; Ch ² = 9.82, df = 1 (P = 0.002); $r^2 = 90\%$ = 1.08 (P = 0.28) 43.8 7.12 15 24 8.5 15 6 3 44 4 2 42 59 59 54.17; Ch ² = 37.29, df = 1 (P < 0.00001); $r^2 = 9$ = 1.20 (P = 0.23) 37.5 9.2 15 24 8.5 15 15 cable = 4.17 (P < 0.0001) 118 113 1.92; Ch ² = 61.69, df = 4 (P < 0.00001); $r^2 = 94$ = 2.47 (P = 0.01) inces: Ch ² = 0.71, df = 2 (P = 0.70), $r^2 = 0\%$ eneration (selection bias) it (selection bias) its and personnel (performance bias) issessment (detection bias) lata (attrition bias)	25 13.06 30 24.41 9.31 26 19.6% 36.13 8.23 14 24 8.5 15 19.4% 44 45 15 19.4% 44 24 8.5 15 19.4% 42.33; Ch ² = 9.82, df = 1 (P = 0.002); l^2 = 90% = 1.08 (P = 0.28) 43.8 7.12 15 24 8.5 15 19.8% 6 3 44 4 2 42 22.0% 59 57 41.7% 54.17; Ch ² = 37.29, df = 1 (P < 0.00001); l^2 = 97% = 1.20 (P = 0.23) 37.5 9.2 15 24 8.5 15 19.2% able = 4.17 (P < 0.0001) 118 113 100.0% 1.92; Ch ² = 61.69, df = 4 (P < 0.00001); l^2 = 94% = 2.47 (P = 0.01) mores: Ch ² = 0.71, df = 2 (P = 0.70), l^2 = 0% eneration (selection bias) tt (selection bias) tt (selection bias) ts and personnel (performance bias) issessment (detection bias) lata (attrition bias)	25 13.06 30 24.41 9.31 26 19.6% 0.59 [-5.30, 6.46] 38.13 8.23 14 24 8.5 15 19.4% 14.13 [8.04, 20.22] 44 41 39.0% 7.34 [-5.93, 20.61] 2.33; Ch ² = 9.82, df = 1 (P = 0.002); $r^2 = 90\%$ = 1.08 (P = 0.26) 43.8 7.12 15 24 8.5 15 19.8% 19.80 [14.19, 25.41] 6 3 44 4 2 42 22.0% 2.00 [0.93, 3.07] 59 57 41.7% 10.68 [-6.76, 28.12] 54.17; Ch ² = 37.29, df = 1 (P < 0.00001); $r^2 = 97\%$ = 1.20 (P = 0.23) 37.5 9.2 15 24 8.5 15 19.2% 13.50 [7.16, 19.84] 15 15 19.2% 13.50 [7.16, 19.84] cable = 4.17 (P < 0.0001) 118 113 100.0% 9.82 [2.01, 17.62] 1.92; Ch ² = 61.69, df = 4 (P < 0.00001); $r^2 = 94\%$ = 2.47 (P = 0.01) inces: Ch ² = 0.71, df = 2 (P = 0.70), $r^2 = 0\%$ eneration (selection bias) it (selection bias) its and personnel (performance bias) issessment (detection bias) lata (attrition bias)	25 13.06 30 24.41 9.31 26 19.6% 0.59 [-5.30, 6.48] 2004 38.13 8.23 14 24 8.5 15 19.4% 14.13 [8.04, 20.22] 2005 44 41 39.0% 7.34 [-5.93, 20.61] 2.33; Ch ² = 9.82, df = 1 (P = 0.002); $r^2 = 90\%$ = 1.08 (P = 0.28) 43.8 7.12 15 24 8.5 15 19.8% 19.80 [14.19, 25.41] 2005 6 3 44 4 2 42 22.0% 2.00 [0.93, 3.07] 2018 59 57 41.7% 10.68 [-6.76, 28.12] 54.17; Ch ² = 37.29, df = 1 (P < 0.00001); $r^2 = 97\%$ = 1.20 (P = 0.23) 37.5 9.2 15 24 8.5 15 19.2% 13.50 [7.16, 19.84] 2005 15 19.2% 13.50 [7.16, 19.84] 2005 16 4.17 (P < 0.0001) 118 113 100.0% 9.82 [2.01, 17.62] 1.92; Ch ² = 61.69, df = 4 (P < 0.00001); $r^2 = 94\%$ = 2.47 (P = 0.01) inces: Ch ² = 0.71, df = 2 (P = 0.70), $r^2 = 0\%$ int (selection bias) it (selection bias) it (selection bias) its and personnel (performance bias) issessment (detection bias) itata (attrition bias)	25 13.06 30 24.41 9.31 26 19.6% 0.59 [-5.30, 6.48] 2004 38.13 8.23 14 24 8.5 15 19.4% 14.13 [8.04, 20.22] 2005 44 24 8.5 15 19.4% 14.13 [8.04, 20.22] 2005 2.33; Ch ² = 9.82; df = 1 ($P = 0.002$); $P = 90\%$ = 1.08 ($P = 0.28$) 43.8 7.12 15 24 8.5 15 19.8% 19.80 [14.19, 25.41] 2005 6 3 44 4 2 42 22.0% 2.00 [0.93, 3.07] 2018 59 57 41.7% 10.68 [-6.76, 28.12] 54.17; Ch ² = 37.29, df = 1 ($P < 0.00001$); $P = 97\%$ = 1.20 ($P = 0.23$) 37.5 9.2 15 24 8.5 15 19.2% 13.50 [7.16, 19.84] 2005 15 15 19.2% 13.50 [7.16, 19.84] 2005 cable = 4.17 ($P < 0.0001$) 118 113 100.0% 9.82 [2.01, 17.62] 1.92; Ch ² = 61.69, df = 4 ($P < 0.00001$); $P = 94\%$ = 2.47 ($P = 0.01$) sinces: Ch ² = 0.71, df = 2 ($P = 0.70$), $P = 0\%$ int (selection bias) it (selection bias) it (selection bias) its and personnel (performance bias) issessment (detection bias) its and personnel (performance bias) issessment (detection bias)

Figure 6 Forest plot of comparison for progressive sperm motility.

are concentrated in the epididymal luminal fluid (Jeulin & Lewin 1996), and likely to be associated with sperm maturation. Carnitines also scavange free oxygen radicles and ROS, thus protecting against OS, as well as aiding

cellular repair in mitochondria during β -oxidation of long-chain fatty acids (Fritz 1963, Steiber *et al.* 2004, Reuter & Evans 2012, Smits *et al.* 2019). However, whilst improved semen characterisitics have been reported,

Study or Subgroup		nitine	Total		ntrols	Total	Weight	Mean Difference IV, Random, 95% CI	Year	Mean Difference IV, Random, 95% CI	Risk of Bias A B C D E F G
2.4.1 LC + LAC	incuit [/v]	55 [74]	Total	incuit [/o]	55 [74]	Iotai	neight	11, 14, 14, 14, 15, 16, 17, 17, 17, 17, 17, 17, 17, 17, 17, 17	. cui		
Lenzi 2004	24.41	7.79	30	28.89	9.1	26	8.2%	-4.48 [-8.95, -0.01]	2004		?
Balencia 2005 Subtotal (95% CI)	40.4	5.82	1 4 44	32.73	6.42	15 41	6.2% 16.4%	7.67 [3.21, 12.13] 1.60 [-10.31, 13.50]	2005		? • • • • • •
Heterogeneity: Tau ² = Test for overall effect:				• 1 (P = 0.0	0002); P	93%	i				
2.4.2 LC											
Balercia 2005	45.13	7.27	15	32.73	6.42	15	7.3%	12.40 [7.49, 17.31]	2005		?
Dimitriadis 2010	25.8	9.8	26	23.7	8.1	22	7.0%	2.10 [-2.96, 7.16]	2010	- -	?? 🗣 ?? ? 🗣 🥊
Mehni 2014	2.2	2.2	51	1.7	0.8	59	20.9%	0.50 [-0.14, 1.14]	2014	+	???????????????????????????????????????
Haje 2015	3.53	0.25	20	1.39	0.28	29	21.6%	2.14 [1.99, 2.29]	2015		??????
T sounapi 2018 Subtotal (95% CI)	7	3	44 156	8	4	42 167	18.3× 75.0%	-1.00 [-2.50, 0.50] 1.77 [0.08, 3.45]	2018	-+	2 ? 🗣 ? ? 🗣 🤅
Heterogeneity: Tau ² = Fest for overall effect:				4 (P < 0.0)	0001); ř	93%	i				
2.4.3 LAC											
Balencia 2005 Subtotal (95% CI)	41.07	5.62	15 15	32.73	6.42	15 15	8.6X 8.6%	8.34 [4.02, 12.66] 8.34 [4.02, 12.66]	2005		?@@@@
Heterogeneity: Not ap Test for overall effect:		P = 0.00)02)								
Total (95% CI)			215			223	100.0%	2.41 [0.79, 4.03]		•	
Heterogeneity: Tau ² = Test for overall effect: Test for subgroup diff	Z = 2.91 (P = 0.00)4)				i			-20 -10 0 10 20 Favours Controls Favours Carnitine	
Risk of bias legend											
A) Random sequence	generation	(selectio	on bias)								
(B) Allocation conceal											
C) Blinding of particip					5)						
D) Blinding of outcom				as)							
E) Incomplete outcom	e data (attr	ition bia:	5)								
(F) Selective reporting	(reporting b	oias)									
G) Other bias											

Figure 7 Forest plot of comparison for normal sperm morphology.



very few studies have recorded pregnancy outcomes after treatment of infertile men with carnitines, and none have considered live birth as a primary outcome (Zini *et al.* 1993, Shekarriz *et al.* 1995, Hakonsen *et al.* 2011, Poljsak 2011, Moolenaar *et al.* 2015).

This meta-analysis presents evidence supporting the improvement of sperm parameters with carnitine supplementation. Carnitines significantly improve total sperm motility (+10.72%), progressive sperm motility (+9.82%) and sperm morphology (+2.41%) (Cavallini et al. 2004, Lenzi et al. 2004, Balercia et al. 2005, Sigman et al. 2006, Dimitriadis et al. 2010, Mehni et al. 2014, Haje & Naoom 2015, Tsounapi et al. 2018). There does not appear to be a positive effect of carnitine supplementation on sperm concentration (Cavallini et al. 2004, Lenzi et al. 2004, Balercia et al. 2005, Dimitriadis et al. 2010, Mehni et al. 2014, Haje & Naoom 2015, Tsounapi et al. 2018). However, it is notable that the studies are characterised by high heterogeneity, and the quality of the evidence was low (total sperm motility, progressive sperm motility and sperm morphology) or very low (sperm concentration) when assessed through GRADEpro (Table 1).

The data indicate that carnitine does not significantly improve pregnancy rates in infertile couples with male infertility, despite improvements in sperm motility and morphology. However, natural conception was not a primary outcome in most studies, indeed most did not follow-up until pregnancy. Therefore, more evidence is required to study the effects of carnitines on pregnancy outcomes. Although multiple attempts have been made to encourage RCTs to report on fertility outcomes, very few RCTs achieve this in studies relating to male infertility (Tournaye, 2006). Nonetheless, two recently reported large RCTs showed that folic acid and zinc supplements (FAZST) or combination antioxidant treatment including Vitamin C, Vitamin E, folic acid, selenium, zinc, and L-carnitine (MOXI trial) did not improve clinical pregnancy or live birth rates when compared to placebo (Schisterman et al. 2020, Steiner et al. 2020).

Our findings are consistent with previously published systematic reviews researching the efficacy and safety of antioxidants in idiopathic male infertility. Two systematic reviews of empirical dietary and/or supplementary intervention recorded improved total sperm motility,

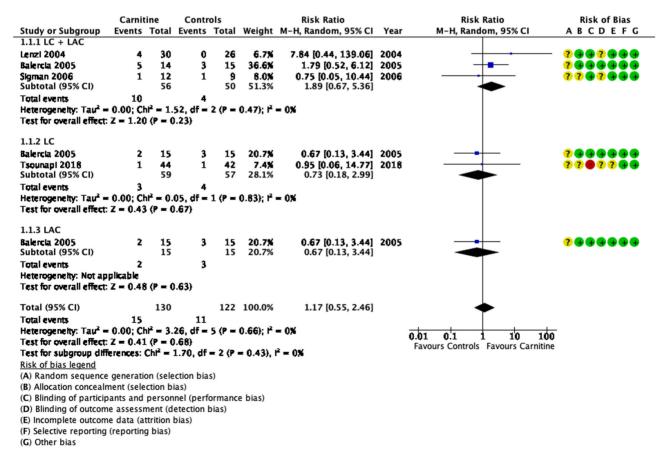


Figure 8 Forest plot of comparison for clinical pregnancy.



Table 3	Table 3 Carnitine versus other arms in the included	arms in the inc	luded studies.					
						Improved Outcome	e	
Published year	l Study	Age (years)	Treatment/day	Sperm concentration, Total sperm (×10°/mL) motility (%)	Total sperm motility (%)	Progressive sperm Sperm motility (%) morph	Sperm morphology (%)	Clinical pregnancy rate (compared to carnitines or controls)
2004	Cavallini <i>et al.</i> (2004) 27–40	27-40	LC + LAC and cinnoxicam	Improved*	Unchanged*	Improved*	Improved*	$x^2 = +5.743; P < 0.05$
2010	Dimitriadis <i>et al.</i> (2010)	NR	Vardenafil Sildenafil	+12.0, <i>P</i> < 0.05 +14.8. <i>P</i> < 0.05	+19.9, <i>P</i> < 0.05 +21.4. <i>P</i> < 0.05	NR NR	+16.3, <i>P</i> < 0.05 +17.7, <i>P</i> < 0.05	NR
2014 2015	Mehni <i>et al.</i> (2014)	25-40 27 EA ± 2 A6	LC and Pentoxifylline	$P = 0.001^{\circ}$	$P = 0.045^{\circ}$	NR ND	$P = 0.052^{\circ}$	NR 18 006 B > 0.05
CI 07	наје & Naoom (2015)	31.54 ± 2.40	i amoxiren Tamoxifen and carnitine	+3.23, P = 0.016 +0.6, P = 0.01	دں، مرحر +5.75, P = 0.045	NR NR	+1.11, P = 0.026	48.3%, P > 0.05
2018	Tsounapi <i>et al.</i> (2018)	NR	Profertil Avanafil Combination of Profertil and Avanafil	+2.1, <i>P</i> > 0.05 +3.5, <i>P</i> > 0.05 +2.9, <i>P</i> > 0.05	+16, <i>P</i> < 0.05 +30, <i>P</i> < 0.05 +24, <i>P</i> < 0.05	+9, <i>P</i> < 0.05 +12, <i>P</i> < 0.05 +7, <i>P</i> < 0.05	+2, <i>P</i> > 0.05 <i>P</i> > 0.05 +2, <i>P</i> > 0.05	R
*Detailed	statistical data was not repc	orted by the author	*Detailed statistical data was not reported by the authors, ^{\$} The raw data was not provided by the authors.	vided by the authors.				

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Not reported

NR,

progressive sperm motility and sperm morphology (Salas-Huetos et al. 2018, Omar et al. 2019). However, it is notable that our findings of effects on total sperm motility and morphology differ from a recent meta-analysis of carnitines in men with idiopathic oligoasthenoteratozoospermia conducted by Zhang et al. (2020). Their systematic review included studies of carnitine plus other antioxidants/ compounds and one study that used active controls (Zhang et al. 2020). In contrast, we selected studies of carnitine-only treatment vs non-active controls so these studies were excluded during our full-text screening. We also included additional studies from other database searches (Zhang et al. 2020). However, the other authors similarly commented on inconsistent data and high heterogeneity amongst the published trials.

A major limitation of this systematic review, and others, is the inability to assess robustly the effect of carnitines on natural conception and pregnancy outcomes as this has not been comprehensively studied to date. Critically, our findings in regards to pregnancy rates did not support carnitine supplementation as an intervention for male infertility, which disagrees with Zhang et al. and is reflective of the different study data included (Zhang et al. 2020).

Conclusion

Overall, our systematic review shows that carnitine supplementation can improve sperm motility and morphology. However, there were only eight randomised controlled trials that specifically compared carnitine(s) to placebo or no treatment and study outcomes had high heterogeneity and were derived from low-quality evidence (Table 1). The majority of studies included found that carnitines were most effective in men with severe idiopathic infertility (Cavallini et al. 2004, Balercia et al. 2005, Sigman et al. 2006, Mehni et al. 2014), supporting their use as a potential treatment. However, whilst it is accepted that gains in male fertility are likely to be seen with improvement in total motile count, particularly when at the lower end of the range (Hamilton et al. 2015), studies included in this metaanalysis have not demonstrated increase in chance of conception, pregnancy and live birth with carnitine supplementation. It therefore remains unclear whether carnitines are a suitable intervention for idiopathic male infertility and randomised placebo-controlled trials reporting on pregnancy and live births are required to clarify this.



Declaration of interest

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Author contribution statement

S C K conceived and designed the study. S C K and Z Z W performed the acquision of data and quality assessment of included studies. S C K conducted the meta-analysis. S C K, R A A and S M d S analysed and interpreted the data. S C K wrote the manuscript with support and input from R A A and S M d S. R A A and S M d S revised the manuscript critically for important intellectual content. All authors approved the final version of the manuscript to be published.

References

- Abdelrazik H, Sharma R, Mahfouz R & Agarwal A 2009
 L-carnitine decreases DNA damage and improves the in vitro blastocyst development rate in mouse embryos. *Fertility and Sterility* 91 589–596. (https://doi.org/10.1016/j.fertnstert.2007.11.067)
- Adeva-Andany MM, López-Maside L, Donapetry-García C, Fernández-Fernández C & Sixto-Leal C 2017 Enzymes involved in branched-chain amino acid metabolism in humans. *Amino Acids* 49 1005–1028. (https://doi.org/10.1007/s00726-017-2412-7)
- Adewoyin M, Ibrahim M, Roszaman R, Isa MLM, Alewi NAM, Rafa AAA & Anuar MNN 2017 Male infertility: the effect of natural antioxidants and phytocompounds on seminal oxidative stress. *Diseases* 5 9. (https://doi.org/10.3390/diseases5010009)
- Agarwal A & Sekhon LH 2011 Oxidative stress and antioxidants for idiopathic oligoasthenoteratospermia: is it justified? *Indian Journal of Urology* 27 74–85. (https://doi.org/10.4103/0970-1591.78437)
- Agarwal A, Virk G, Ong C & Du Plessis SS 2014 Effect of oxidative stress on male reproduction. *World Journal of Men's Health* **32** 1–17. (https://doi.org/10.5534/wjmh.2014.32.1.1)
- Agarwal A, Mulgund A, Hamada A & Chyatte MR 2015 A unique view on male infertility around the globe. *Reproductive Biology and Endocrinology* 13 37. (https://doi.org/10.1186/s12958-015-0032-1)
- Aitken RJ & Baker MA 2006 Oxidative stress, sperm survival and fertility control. *Molecular and Cellular Endocrinology* 250 66–69. (https://doi.org/10.1016/j.mce.2005.12.026)
- Aitken RJ, Baker MA & Sawyer D 2003 Oxidative stress in the male germ line and its role in the aetiology of male infertility and genetic disease. *Reproductive Biomedicine Online* 7 65–70. (https://doi. org/10.1016/s1472-6483(10)61730-0)
- Aitken RJ, Gibb Z, Mitchell LA, Lambourne SR,
 Connaughton HS & De Iuliis GN 2012 Sperm motility is lost in vitro as a consequence of mitochondrial free radical production and the generation of electrophilic aldehydes but can be significantly rescued by the presence of nucleophilic thiols. *Biology of Reproduction* 87 110. (https://doi.org/10.1095/biolreprod.112.102020)
- Aliabadi E, Soleimani Mehranjani M, Borzoei Z, Talaei-Khozani T, Mirkhani H & Tabesh H 2012 Effects of L-carnitine

and L-acetyl-carnitine on testicular sperm motility and chromatin quality. *Iranian Journal of Reproductive Medicine* **10** 77–82.

- Balercia G, Regoli F, Armeni T, Koverech A, Mantero F &
 Boscaro M 2005 Placebo-controlled double-blind randomized trial on the use of L-carnitine, L-acetylcarnitine, or combined L-carnitine and L-acetylcarnitine in men with idiopathic asthenozoospermia. *Fertility and Sterility* 84 662–671. (https://doi.org/10.1016/j.fertnstert.2005.03.064)
- Balshem H, Helfand M, Schunemann HJ, Oxman AD, Kunz R, Brozek J, Vist GE, Falck-Ytter Y, Meerpohl J, Norris S et al. 2011 GRADE guidelines: 3. Rating the quality of evidence. *Journal* of Clinical Epidemiology 64 401–406. (https://doi.org/10.1016/j. jclinepi.2010.07.015)
- Banihani S, Agarwal A, Sharma R & Bayachou M 2014 Cryoprotective effect of L-carnitine on motility, vitality and DNA oxidation of human spermatozoa. *Andrologia* **46** 637–641. (https:// doi.org/10.1111/and.12130)
- Bremer J 1983 Carnitine metabolism and functions. *Physiological Reviews* 63 1420–1480. (https://doi.org/10.1152/ physrev.1983.63.4.1420)
- Cavallini G, Ferraretti AP, Gianaroli L, Biagiotti G & Vitali G 2004 Cinnoxicam and L-carnitine/acetyl-L-carnitine treatment for idiopathic and varicocele-associated oligoasthenospermia. *Journal of Andrology* 25 761–770; discussion 771–772. (https://doi. org/10.1002/j.1939-4640.2004.tb02853.x)
- Colpi GM, Francavilla S, Haidl G, Link K, Behre HM, Goulis DG, Krausz C & Giwercman A 2018 European Academy of Andrology Guideline Management of oligo-astheno-teratozoospermia. *Andrology* 6 513–524. (https://doi.org/10.1111/andr.12502)
- **de Lamirande E & Gagnon C** 1992 Reactive oxygen species and human spermatozoa. I. Effects on the motility of intact spermatozoa and on sperm axonemes. *Journal of Andrology* **13** 368–378.
- Dimitriadis F, Tsambalas S, Tsounapi P, Kawamura H, Vlachopoulou E, Haliasos N, Gratsias S, Watanabe T, Saito M, Miyagawa I et al. 2010 Effects of phosphodiesterase-5 inhibitors on Leydig cell secretory function in oligoasthenospermic infertile men: a randomized trial. *BJU International* **106** 1181–1185. (https://doi.org/10.1111/j.1464-410X.2010.09243.x)
- **Dokmeci D** 2005 Oxidative stress, male infertility and the role of carnitines. *Folia Medica* **47** 26–30.
- **Evidence Prime Inc.** 2015 *GRADEpro GDT: GRADEpro Guideline Development Tool [Software].* McMaster University.
- Fritz IB 1963 Carnitine and its role in fatty acid metabolism. Advances in Lipid Research 1 285–334. (https://doi.org/10.1016/B978-1-4831-9937-5.50014-4)
- Gomez E, Irvine DS & Aitken RJ 1998 Evaluation of a spectrophotometric assay for the measurement of malondialdehyde and 4-hydroxyalkenals in human spermatozoa: relationships with semen quality and sperm function. *International Journal of Andrology* 21 81–94. (https://doi.org/10.1046/j.1365-2605.1998.00106.x)
- **Gudeloglu A & Parekattil SJ** 2013 Update in the evaluation of the azoospermic male. *Clinics* **68** (Supplement 1) 27–34. (https://doi. org/10.6061/clinics/2013(sup01)04)
- Haje M & Naoom K 2015 Combined tamoxifen and L-carnitine therapies for the treatment of idiopathic male infertility attending intracytoplasmic sperm injection: a randomized controlled trial. *International Journal of Infertility and Fetal Medicine* 6 20–24. (https:// doi.org/10.5005/jp-journals-10016-1096)
- Hakonsen LB, Thulstrup AM, Aggerholm AS, Olsen J,
 Bonde JP, Andersen CY, Bungum M, Ernst EH, Hansen ML,
 Ernst EH et al. 2011 Does weight loss improve semen quality and reproductive hormones? Results from a cohort of severely obese men.
 Reproductive Health 8 24. (https://doi.org/10.1186/1742-4755-8-24)
- Hamilton JA, Cissen M, Brandes M, Smeenk JM, De Bruin JP, Kremer JA, Nelen WL & Hamilton CJ 2015 Total motile sperm count: a better indicator for the severity of male factor infertility



than the WHO sperm classification system. *Human Reproduction* **30** 1110–1121. (https://doi.org/10.1093/humrep/dev058)

- Higgins JP, Altman DG, Gotzsche PC, Juni P, Moher D, Oxman AD, Savovic J, Schulz KF, Weeks L, Sterne JA et al. 2011 The Cochrane Collaboration's tool for assessing risk of bias in randomised trials. *BMJ* 343 d5928. (https://doi.org/10.1136/bmj. d5928)
- Higgins J, Thomas J, Chandler J, Cumpston M, Li T, Page M & Welch VE 2019 Cochrane Handbook for Systematic Reviews of Interventions, Version 6.0 (Updated July 2019). Cochrane. (available at: https://training.cochrane.org/handbook/archive/v6)
- Iommiello VM, Albani E, Di Rosa A, Marras A, Menduni F, Morreale G, Levi SL, Pisano B & Levi-Setti PE 2015 Ejaculate oxidative stress is related with sperm DNA fragmentation and round cells. *International Journal of Endocrinology* 2015 321901. (https://doi. org/10.1155/2015/321901)
- Isidori A, Latini M & Romanelli F 2005 Treatment of male infertility. *Contraception* **72** 314–318. (https://doi.org/10.1016/j. contraception.2005.05.007)
- Isidori AM, Pozza C, Gianfrilli D & Isidori A 2006 Medical treatment to improve sperm quality. *Reproductive Biomedicine Online* 12 704–714. (https://doi.org/10.1016/s1472-6483(10)61082-6)
- Jeulin C & Lewin LM 1996 Role of free L-carnitine and acetyl-Lcarnitine in post-gonadal maturation of mammalian spermatozoa. *Human Reproduction Update* **2** 87–102. (https://doi.org/10.1093/ humupd/2.2.87)
- Jungwirth A, Giwercman A, Tournaye H, Diemer T, Kopa Z, Dohle G, Krausz C & European Association of Urology Working Group on Male Infertility 2012 European Association of Urology guidelines on male infertility: the 2012 update. *European Urology* 62 324–332. (https://doi.org/10.1016/j.eururo.2012.04.048)
- Kerner J & Hoppel C 1998 Genetic disorders of carnitine metabolism and their nutritional management. *Annual Review of Nutrition* 18 179–206. (https://doi.org/10.1146/annurev.nutr.18.1.179)
- Kodama H, Yamaguchi R, Fukuda J, Kasai H & Tanaka T 1997 Increased oxidative deoxyribonucleic acid damage in the spermatozoa of infertile male patients. *Fertility and Sterility* **68** 519–524. (https://doi.org/10.1016/s0015-0282(97)00236-7)
- Lenzi A, Lombardo F, Sgro P, Salacone P, Caponecchia L, Dondero F & Gandini L 2003 Use of carnitine therapy in selected cases of male factor infertility: a double-blind crossover trial. *Fertility and Sterility* **79** 292–300. (https://doi.org/10.1016/s0015-0282(02)04679-4)
- Lenzi A, Sgro P, Salacone P, Paoli D, Gilio B, Lombardo F, Santulli M, Agarwal A & Gandini L 2004 A placebo-controlled double-blind randomized trial of the use of combined l-carnitine and l-acetyl-carnitine treatment in men with asthenozoospermia. *Fertility and Sterility* 81 1578–1584. (https://doi.org/10.1016/j. fertnstert.2003.10.034)
- Martins da Silva SJ 2019 Male infertility and antioxidants: one small step for man, no giant leap for andrology? *Reproductive Biomedicine Online* 39 879–883. (https://doi.org/10.1016/j.rbmo.2019.08.008)
- Martins da Silva SJ, Brown SG, Sutton K, King LV, Ruso H, Gray DW, Wyatt PG, Kelly MC, Barratt CLR & Hope AG 2017 Drug discovery for male subfertility using high-throughput screening: a new approach to an unsolved problem. *Human Reproduction* **32** 974–984. (https://doi.org/10.1093/humrep/dex055)
- Mehni N, Ketabchi AA & Hosseini E 2014 Combination effect of pentoxifylline and L-carnitine on idiopathic oligoasthenoteratozoospermia. *Iranian Journal of Reproductive Medicine* 12 817–824.
- Moher D, Shamseer L, Clarke M, Ghersi D, Liberati A, Petticrew M, Shekelle P & Stewart LA & PRISMA-P Group 2015 Preferred reporting items for systematic review and metaanalysis protocols (PRISMA-P) 2015 statement. *Systematic Reviews* **4** [epub]. (https://doi.org/10.1186/2046-4053-4-1)

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- Mongioi L, Calogero AE, Vicari E, Condorelli RA, Russo GI, Privitera S, Morgia G & La Vignera S 2016 The role of carnitine in male infertility. *Andrology* 4 800–807. (https://doi.org/10.1111/ andr.12191)
- Moolenaar LM, Cissen M, De Bruin JP, Hompes PG, Repping S, Van Der Veen F & Mol BW 2015 Cost-effectiveness of assisted conception for male subfertility. *Reproductive Biomedicine Online* **30** 659–666. (https://doi.org/10.1016/j.rbmo.2015.02.006)
- Omar MI, Pal RP, Kelly BD, Bruins HM, Yuan Y, Diemer T, Krausz C, Tournaye H, Kopa Z, Jungwirth A et al. 2019 Benefits of empiric nutritional and medical therapy for semen parameters and pregnancy and live birth rates in couples with idiopathic infertility: a systematic review and meta-analysis. *European* Urology 75 615–625. (https://doi.org/10.1016/j.eururo.2018.12.022)
- Poljsak B 2011 Strategies for reducing or preventing the generation of oxidative stress. Oxidative Medicine and Cellular Longevity 2011 194586. (https://doi.org/10.1155/2011/194586)
- Punab M, Poolamets O, Paju P, Vihljajev V, Pomm K, Ladva R, Korrovits P & Laan M 2017 Causes of male infertility: a 9-year prospective monocentre study on 1737 patients with reduced total sperm counts. *Human Reproduction* **32** 18–31. (https://doi. org/10.1093/humrep/dew284)
- Reuter SE & Evans AM 2012 Carnitine and acylcarnitines: pharmacokinetic, pharmacological and clinical aspects. *Clinical Pharmacokinetics* 51 553–572. (https://doi.org/10.1007/BF03261931)
- Rowe PJ, Comhaire FH, Hargreave TB & Mahmoud AMA 2000 WHO Manual for the Standardized Investigation and Diagnosis of the Infertile Male. Cambridge, UK: Cambridge University Press.
- Russo A, Acquaviva R, Campisi A, Sorrenti V, Di Giacomo C, Virgata G, Barcellona ML & Vanella A 2000 Bioflavonoids as antiradicals, antioxidants and DNA cleavage protectors. *Cell Biology and Toxicology* 16 91–98. (https://doi. org/10.1023/a:1007685909018)
- Salas-Huetos A, Rosique-Esteban N, Becerra-Tomas N, Vizmanos B, Bullo M & Salas-Salvado J 2018 The effect of nutrients and dietary supplements on sperm quality parameters: a systematic review and meta-analysis of randomized clinical trials. Advances in Nutrition 9 833–848. (https://doi.org/10.1093/advances/ nmy057)
- Schisterman EF, Sjaarda LA, Clemons T, Carrell DT, Perkins NJ, Johnstone E, Lamb D, Chaney K, Van Voorhis BJ, Ryan G *et al.* 2020 Effect of folic acid and zinc supplementation in men on semen quality and live birth among couples undergoing infertility treatment: a randomized clinical trial. *JAMA* 323 35–48. (https://doi. org/10.1001/jama.2019.18714)
- Schünemann H, Brożek J, Guyatt G & Oxman A 2013 GRADE Handbook for Grading Quality of Evidence and Strength of Recommendations. The Grade Working Group.
- Shalev DP, Soffer Y & Lewin LM 1986 Investigations on the motility of human spermatozoa in a defined medium in the presence of metabolic inhibitors and of carnitine. *Andrologia* 18 368–375. (https://doi.org/10.1111/j.1439-0272.1986.tb01792.x)
- Shamseer L, Moher D, Clarke M, Ghersi D, Liberati A, Petticrew M, Shekelle P, Stewart LA & GROUP P-P 2015 Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation. *BMJ* 350 g7647. (https://doi.org/10.1136/bmj.g7647)
- Shekarriz M, Thomas Jr AJ & Agarwal A 1995 Effects of time and sperm concentration on reactive oxygen species formation in human semen. *Archives of Andrology* **34** 69–75. (https://doi. org/10.3109/01485019508987833)
- Shekhawat PS, Sonne S, Carter AL, Matern D & Ganapathy V 2013 Enzymes involved in L-carnitine biosynthesis are expressed by small intestinal enterocytes in mice: implications for gut health. *Journal of Crohn's and Colitis* **7** e197–e205. (https://doi.org/10.1016/j. crohns.2012.08.011)



- Sigman M, Glass S, Campagnone J & Pryor JL 2006 Carnitine for the treatment of idiopathic asthenospermia: a randomized, doubleblind, placebo-controlled trial. *Fertility and Sterility* 85 1409–1414. (https://doi.org/10.1016/j.fertnstert.2005.10.055)
- Smits RM, Mackenzie-Proctor R, Yazdani A, Stankiewicz MT, Jordan V & Showell MG 2019 Antioxidants for male subfertility. Cochrane Database of Systematic Reviews 3 CD007411. (https://doi. org/10.1002/14651858.CD007411.pub4)
- Steiber A, Kerner J & Hoppel CL 2004 Carnitine: a nutritional, biosynthetic, and functional perspective. *Molecular Aspects of Medicine* 25 455–473. (https://doi.org/10.1016/j.mam.2004.06.006)
- Steiner AZ, Hansen KR, Barnhart KT, Cedars MI, Legro RS, Diamond MP, Krawetz SA, Usadi R, Baker VL, Coward RM et al. 2020 The effect of antioxidants on male factor infertility: the Males, Antioxidants, and Infertility (MOXI) randomized clinical trial. *Fertility and Sterility* **113** 552.e3–560.e3. (https://doi.org/10.1016/j. fertnstert.2019.11.008)
- Storey BT 1997 Biochemistry of the induction and prevention of lipoperoxidative damage in human spermatozoa. *Molecular Human Reproduction* **3** 203–213. (https://doi.org/10.1093/molehr/3.3.203)
- **Tanphaichitr N** 1977 In vitro stimulation of human sperm motility by acetylcarnitine. *International Journal of Fertility* **22** 85–91.
- **The Nordic Cochrane Centre** 2014 *Review Manager (RevMan).* Version 5.3 ed. Copenhagen: The Cochrane Collaboration.
- Tournaye H 2006 Evidence-based management of male subfertility. *Current Opinion in Obstetrics and Gynecology* 18 253–259. (https://doi.org/10.1097/01.gco.0000192994.37965.c6)
- Tsounapi P, Honda M, Dimitriadis F, Koukos S, Hikita K, Zachariou A, Sofikitis N & Takenaka A 2018 Effects of a micronutrient supplementation combined with a phosphodiesterase type 5 inhibitor on sperm quantitative and qualitative parameters, percentage of mature spermatozoa and sperm capacity to undergo hyperactivation: a randomised controlled trial. *Andrologia* **50** e13071. (https://doi.org/10.1111/and.13071)
- Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M & Telser J 2007 Free radicals and antioxidants in normal physiological

functions and human disease. *International Journal of Biochemistry and Cell Biology* **39** 44–84. (. (https://doi.org/10.1016/j. biocel.2006.07.001)

- Vaz FM & Wanders RJ 2002 Carnitine biosynthesis in mammals. Biochemical Journal 361 417–429. (https://doi.org/10.1042/0264-6021:3610417)
- Venkatesh S, Shamsi MB, Deka D, Saxena V, Kumar R & Dada R 2011 Clinical implications of oxidative stress & sperm DNA damage in normozoospermic infertile men. *Indian Journal of Medical Research* 134 396–398.
- **Veritas Health Innovation**. *Covidence Systematic Review Software* [Online]. Melbourne, Australia. (available at: www.covidence.org). Accessed.
- **World Health Organization** 2010 Reference values and semen nomenclature. In *WHO Laboratory Manual for the Examination and Processing of Human Semen.* Geneva: World Health Organization.
- World Health Organization 2020 Infertility definitions and terminology [Online]. World Health Organization. (available at: https://www.who.int/reproductivehealth/topics/infertility/ definitions/en/). Accessed 16 July 2020.
- Zhang X, Cui Y, Dong L, Sun M & Zhang Y 2020 The efficacy of combined l-carnitine and l-acetyl carnitine in men with idiopathic oligoasthenoteratozoospermia: a systematic review and meta-analysis. *Andrologia* 52 e13470. (https://doi.org/10.1111/ and.13470)
- Zini A, De Lamirande E & Gagnon C 1993 Reactive oxygen species in semen of infertile patients: levels of superoxide dismutaseand catalase-like activities in seminal plasma and spermatozoa. *International Journal of Andrology* **16** 183–188. (https://doi. org/10.1111/j.1365-2605.1993.tb01177.x)
- Zopfgen A, Priem F, Sudhoff F, Jung K, Lenk S, Loening SA & Sinha P 2000 Relationship between semen quality and the seminal plasma components carnitine, alpha-glucosidase, fructose, citrate and granulocyte elastase in infertile men compared with a normal population. *Human Reproduction* **15** 840–845. (https://doi. org/10.1093/humrep/15.4.840)

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