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Short communication

Pleurotus spp. cultivation on *Brachiaria* sp. straw treatment with alkaline water Oyster mushroom and substrate treatment



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ABSTRACT

The aim of this research was to evaluate the efficiency of aqueous alkali-treated *Brachiaria* straw for the cultivation of appropriate species of oyster mushroom. The substrate used in the cultivation of various *Pleurotus* spp. was soaked for 20 min by using two different procedures: (i) 0.5–2.0% Ca(OH)₂ in 100 L water, and (ii) 50–250 L water. As a result, 1% Ca(OH)₂ dissolved in 100 L water and 3.5 kg of *Brachiaria* straw presented the best production. The most suitable species for the application of the present method were *P. pulmonarius* and *P. sapidus*. The success of this technique is directly related to the concentration of Ca(OH)₂ and water, the species, and the origin and quality of raw material used as the substrate in the production of oyster mushroom.

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Worldwide, diverse treatments have been used to prepare substrates for mushroom cultivation, with the objective to eliminate fungal contamination and microorganisms.^{1–3} The principal methods are composting and steam pasteurization, but these treatments require a high energy outlay.^{4,5} The alkaline method is a simple procedure that is mainly used in rural communities, where small-scale production will suffice. Compared to other strategies, this method presents many

advantages, such as low-cost, the highest biological efficiency, no fungal contamination, a shorter colonization time,^{6,7} and energy absence.

The principal substrates studied in the cultivation of mushroom using the alkaline method include banana leaves, “palmareca” leaves, corn cob, and corn straw.^{7,8} Conversely, the food industry is always producing high quantities of agricultural wastes, like *Brachiaria* straw.⁹ Growers of mushroom

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are continually searching for various substrates to improve the production with low cost.¹⁰ The ability of *Pleurotus* genera to grow on a variety of organic materials is a valuable differential to increase the world production and consumption of this mushroom, represented by various species, such as *P. ostreatus* var. Florida, *P. sapidus*, *P. pulmonarius*, *P. djamor*, and *P. cornucopiae* var. *citrinopileatus*.

Hence, this research aimed to evaluate the efficiency of aqueous alkali-treated *Brachiaria* straw for the cultivation of appropriate species of culinary-medicinal oyster mushroom. Particular focus was given to the $\text{Ca}(\text{OH})_2$ and water contents, to determine the optimal formulation to achieve the highest yield parameters.

The following three fungi strains were used: *P. sapidus*, *P. ostreatus* var. Florida, and *P. pulmonarius*. Stocks of all strains are deposited at the fungal collection of the São Paulo State University, Câmpus de Dracena (São Paulo, Brazil) and accessible to other researchers who are interested in continuing the present research.

The grain spawn was produced using sorghum seeds, as outlined in the method presented by Zied et al.¹¹ Briefly, the seeds were boiled at 100 °C for 30 h and then placed (0.5 kg wet weight) in polyethylene bags and mixed with CaSO_4 (1%) and limestone (0.5%). Afterward, the bags were inoculated with the *Pleurotus* species and incubated in a dark room at 25 °C for 15 days.

Edible fungi were produced on *Brachiaria* straw, broken into 4–8 cm pieces using a hammer mill. The chemical composition of the straw was 6.6% protein, 29.5% fiber, 57.4% carbon, 0.3% phosphorus, 1.06% nitrogen, and a C/N ratio of 54/1. Two experimentation conditions of substrate preparation were performed. In both experiments, water and $\text{Ca}(\text{OH})_2$ were mixed for 2 min to ensure a perfect homogenization before soaking of the *Brachiaria* straw. In the first, the substrate was soaked (3.5 kg wet weight) in a plastic container with 100 L water and 0.0 (control), 0.5, 1.0, and 2.0% $\text{Ca}(\text{OH})_2$, respectively, for 20 min (Supporting File). In the second, the *Brachiaria* straw

was soaked (3.5 kg wet weight) in a plastic container containing 1.0% $\text{Ca}(\text{OH})_2$ (best level reported in first procedure) and 50, 100, 150, 200, and 250 L water, respectively, for 20 min. In both experiment after soaking the *Brachiaria* straw was drained in plastic box for 60 min (Supporting File). Afterward, in plastic bags the substrate (2.5 kg wet weight) were inoculated with 2% (w/w) spawn (sorghum seeds), and incubated in a dark room at 25 °C and 70% relative humidity.

The yield parameters of the fruit bodies were evaluated based on the yield (Y), biological efficiency (BE), number of mushrooms (NM), and weight of mushrooms (AFB).^{12–15} For Y, the basidiocarp fresh weight was divided by the compost fresh weight, multiplied by 100, and expressed as a percentage. BE was calculated as the basidiocarp fresh weight divided by the compost dry weight, multiplied by 100, and expressed as a percentage. The count of the harvested basidiocarps represented the NM. For calculation of the AFB, the basidiocarp fresh weight was divided by the number of basidiocarps and expressed in grams.

Both experimental procedures adopted a double factorial design. In the first, eight treatments (two species \times four $\text{Ca}(\text{OH})_2$ contents) were considered, with seven replicates, resulting in a degree of residual freedom with a value of 18. The second consisted of ten treatments (two species \times five water contents), with seven replicates, resulting in a degree of residual freedom with a value of 24. ANOVA was used to analyze the data, and Tukey's test was employed to establish significant differences between means ($p \leq 0.05$). All calculations were performed using the SAS JMP software.

According to the results in Table 1, compared to cultivation of *P. sapidus* in the presence of $\text{Ca}(\text{OH})_2$, the treatment control showed the lowest yield parameters for this species while *P. ostreatus* var. Florida was not produced under this condition (no $\text{Ca}(\text{OH})_2$). Otherwise, 1% $\text{Ca}(\text{OH})_2$ presented the highest yield for both strains, with *P. sapidus* recording the highest values of Y (19.07%), BE (198.38%), NM (57), and AFB (1.60 g). In comparison, values of Y (5.98%), BE (74.75%), NM (16), and AFB

Table 1 – Yield parameters of *Pleurotus* spp. using different amounts of $\text{Ca}(\text{OH})_2$.

Species	$\text{Ca}(\text{OH})_2$ (%)			
	0.0	0.5	1.0	2.0
	Yield, %			
<i>P. sapidus</i>	0.83 b A	5.57 b A	19.07 a A	15.78 a A
<i>P. ostreatus</i> var. Florida	N.F. ^a	0.49 a A	5.98 a B	3.01 a B
	Biological efficiency, %			
<i>P. sapidus</i>	10.38 b A	69.30 b A	198.38 a A	159.75 a A
<i>P. ostreatus</i> var. Florida	N.F.	6.13 a A	74.75 a B	37.63 a B
	Number of mushrooms, n			
<i>P. sapidus</i>	3 c A	23 bc A	57 a A	47 ab A
<i>P. ostreatus</i> var. Florida	N.F.	11 a A	16 a B	6 a B
	Weight of mushrooms, g			
<i>P. sapidus</i>	0.29 a A	0.98 a A	1.60 a A	1.73 a A
<i>P. ostreatus</i> var. Florida	N.F.	0.10 b A	1.42 ab A	2.46 a A

^a N.F.: not fructified.

Values followed by different lowercase letters within a line and uppercase letters within a column are significantly different among the yield parameters at $p < 0.05$, according to Tukey's test, $n = 7$.

Table 2 – Production of *Pleurotus* spp. using 1% Ca(OH)₂ and different water contents.

Species	Water (L)				
	50	100	150	200	250
	Yield, %				
<i>P. sapidus</i>	5.50 abc A	7.68 a A	1.68 bc A	1.51 c B	7.58 ab A
<i>P. pulmonarius</i>	8.18 a A	10.84 a A	5.59 a A	8.46 a A	7.26 a A
	Biological efficiency, %				
<i>P. sapidus</i>	68.75 abc A	96.00 a A	21.00 bc A	18.80 c B	94.75 ab A
<i>P. pulmonarius</i>	102.25 a A	135.50 a A	69.87 a A	105.70 a A	90.75 a A
	Number of mushrooms, n				
<i>P. sapidus</i>	11 ab A	18 ab A	5 b A	4 b B	24 a A
<i>P. pulmonarius</i>	18 a A	15 a A	13 a A	16 a A	12 a B
	Weight of mushrooms, g				
<i>P. sapidus</i>	0.41 a A	0.64 a A	0.25 a A	0.19 a A	0.44 a A
<i>P. pulmonarius</i>	0.50 a A	0.76 a A	0.32 a A	0.46 a A	0.64 a A

Values followed by different lowercase letters within a line and uppercase letters within a column are significantly different among the yield parameters at $p < 0.05$, according to Tukey's test, $n = 7$.

(1.42 g) were recorded for *P. ostreatus* var. Florida. Significantly superior, *P. sapidus* species can be cultivated in small and low technological mushroom growers using 1–2% Ca(OH)₂.

Depending on the concentrations and alkaline chemical, literature results vary among the studies. Villa-Cruz et al.¹⁶ obtained a BE of *P. ostreatus* between 70.6% and 72.0% when a mixture of corn-cobs and coffee pulp was soaked in a solution of water with lime (2%). Bernabé-González et al.¹⁷ cultivated *P. pulmonarius* on dry banana leaves using two treatments. In the first treatment, the substrate was immersed in a solution of water with lime (2%) for 24 h, reaching a BE of 120.1%. By contrast, a lower BE (41.4–81.2%) was obtained when the substrate was immersed in hot water at 80 °C for 1 h. In the present study, the best results of BE with the species *P. ostreatus* was 74.75% and *P. pulmonarius* was 135.50%.

According to the positive results obtained by *P. sapidus* species for the 1% Ca(OH)₂ diluted in 100L water, a second procedure was done, to verify the viability of the production of the *P. pulmonarius* species. In this trial, we also established if the volume of 3.5 kg *Brachiaria* would be influenced by the amount of water used to soak the straw.

Again, the 1% Ca(OH)₂ in 100L water showed the highest yield for both species. In this instance, *P. pulmonarius* exhibited the highest values of Y (10.84%), BE (135.50%), NM (15), and AFB (0.76 g). Conversely, *P. sapidus* displayed values of Y (7.68%), BE (96%), NM (18), and AFB (0.64 g). The water content for soaking of the *Brachiaria* does not influence the average weight of the mushrooms. The treatments that obtained a superior Y and BE provided a high NM harvested (Table 2).

The differences in the Y and BE of *P. sapidus* between the first and second procedures was due to the quality of the *Brachiaria* straw used. In the first study, the presence of pests did not occur, whereas, in the second study, the presence of larvae and adult sciarid flies was verified (Supporting File). Thus, the presented method does not have efficient insect control unlike that over contaminating fungi. Stölzer and Grabbe¹⁸ reported that the alkaline method is favored owing to its ability to reduce the microorganisms and also for its low cost and efficiency.

Therefore, batches of old straw and those stored under poor conditions should be avoided when applying this approach. Another alternative would be the use of a biological or chemical treatment together with the soaking of the straw. Rodríguez Estrada and Pecchia⁵ reported that sciarids are the main fly that affects *Pleurotus* crops in North America. These pests heavily impact the crop if they enter the cultivation rooms during the incubation stage when the substrate temperature is around 24 °C.

Thus, the success of this technique is directly related to the concentration of Ca(OH)₂ and water, the species, and the origin and quality of raw material used as the substrate in the production of oyster mushroom.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.bjm.2018.06.003](https://doi.org/10.1016/j.bjm.2018.06.003).

REFERENCES

- Hernández D, Sánchez JE, Yamasaki K. A simple procedure for preparing substrate for *Pleurotus ostreatus* cultivation. *Bioresour Technol.* 2003;90:145–150.
- Mejía SG, Albertó E. Heat treatment of wheat straw by immersion in hot water decreases mushroom yield in *Pleurotus ostreatus*. *Rev Iberoam Micol.* 2013;30:125–129.
- Colavolpe MB, Mejía SJ, Albertó E. Efficiency of treatments for controlling *Trichoderma* spp. during spawning in cultivation of lignicolous mushrooms. *Braz J Microbiol.* 2014;4:1263–1270.
- Ali MA, Mehmood MI, Nawaz R, Hanif MA, Wasim R. Influence of substrate pasteurization methods on the yield

- of oyster mushroom (*Pleurotus* species). *Pak J Agric Sci*. 2007;44:300–303.
5. Rodríguez Estrada AE, Pecchia J. Cultivation of *Pleurotus ostreatus*. In: Zied DC, Pardo-Giménez A, eds. *Edible and medicinal mushrooms: technology and applications*. Chichester, UK: John Wiley & Sons; 2017:339–360.
 6. Philippoussis A, Zervakis G, Diamantopoulou P. Bioconversion of agricultural lignocellulosic wastes through the cultivation of the edible mushroom *Agrocybe aegerita*, *Volvariella volvacea* and *Pleurotus* spp. *World J Microbiol Biotechnol*. 2001;17:191–200.
 7. Contreras EP, Sokolov M, Mejía G, Sánchez JE. Soaking of substrate in alkaline water as a pretreatment for the cultivation of *Pleurotus ostreatus*. *J Hort Sci Biotechnol*. 2004;79:234–240.
 8. Bernabé-González T, Cayetano-Catarino M, Adán-Díaz A, Torres-Pastrana MA. Cultivo de *Pleurotus pulmonarius* sobre diversos subproductos agrícolas de Guerrero México. *Rev Mex Mic*. 2004;18:77–80.
 9. Nascente AS, Guimarães CM, Cobucci T, Crusciol CAC. *Brachiaria ruziziensis* and herbicide on yield of upland rice. *Planta Daninha*. 2012;30:729–736.
 10. Royse DJ, Rhodes TW, Ohga S, Sanchez JE. Yield, mushroom size and time to production of *Pleurotus cornucopiae* (oyster mushroom) grown on switch grass substrate spawned and supplemented at various rates. *Bioresour Technol*. 2004;91:85–91.
 11. Zied DC, Pardo JE, Tomaz RS, Miasaki CT, Pardo-Giménez A. Mycochemical characterization of *Agaricus subrufescens* considering their morphological and physiological stage of maturity on the traceability process. *Biomed Res Int*. 2017;7422713:1–10.
 12. Pardo-Giménez A, Catalán L, Carrasco J, Álvarez-Ortí M, Zied DC, Pardo JE. Effect of supplementing crop substrate with defatted pistachio meal on *Agaricus bisporus* and *Pleurotus ostreatus* production. *J Sci Food Agric*. 2016;96:3838–3845.
 13. Salmones D, Gaitán-Hernández R, Pérez R, Guzmán G. Estudios sobre el género *Pleurotus* VIII Interacción entre crecimiento micelial y productividad. *Rev Iberoam Mico*. 1997;14:173–176.
 14. Dias ES, Zied DC, Rinker DL. Physiologic response of *Agaricus subrufescens* using different casing materials and practices applied in the cultivation of *Agaricus bisporus*. *Fungal Biol*. 2013;117:569–575.
 15. Zied DC, Dourado FA, Dias ES, Pardo-Giménez A. First study of hormesis effect on mushroom cultivation. *World J Microbiol Biotechnol*. 2017;33:195.
 16. Villa-Cruz V, Huerta-Palacios G, Sánchez-Vázquez JE. Solid fermentation of a corn cob-coffee pulp mixture for the cultivation of *Pleurotus ostreatus*. *Micol Neotrop Apl*. 1999;12:67–74.
 17. Bernabé-González T, Cayetano-Catarino M. Cultivation of *Pleurotus pulmonarius* on substrates treated by immersion in alkaline water in Guerrero Mexico. *Micol Apl Int*. 2009;21:19–23.
 18. Stölzer S, Grabbe K. Mechanisms of substrate selectivity in the cultivation of edible fungi. *Mush Sci*. 1991;13:141–146.