WŨ

# World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 August 14; 28(30): 4053-4060

DOI: 10.3748/wjg.v28.i30.4053

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

# Gut microbiota, inflammatory bowel disease and colorectal cancer

Ana Elisa Valencise Quaglio, Thais Gagno Grillo, Ellen Cristina Souza De Oliveira, Luiz Claudio Di Stasi, Ligia Yukie Sassaki

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Budin C, Romania: Houston KV, United States

Received: January 18, 2022 Peer-review started: January 18, 2022 First decision: March 8, 2022 **Revised:** March 16, 2022 Accepted: July 18, 2022 Article in press: July 18, 2022 Published online: August 14, 2022



Ana Elisa Valencise Quaglio, Luiz Claudio Di Stasi, Departament of Biophysics and Pharmacology, São Paulo State University (Unesp), Institute of Biosciences, Botucatu 18618-689, São Paulo State, Brazil

Thais Gagno Grillo, Ellen Cristina Souza De Oliveira, Ligia Yukie Sassaki, Department of Internal Medicine, São Paulo State University (Unesp), Medical School, Botucatu 18618-686, São Paulo State, Brazil

Corresponding author: Ligia Yukie Sassaki, MD, PhD, Assistant Professor, Ligia Yukie Sassaki, MD, PhD, Assistant Professor, Department of Internal Medicine, São Paulo State University (Unesp), Medical School, s/n. Bairro Rubião Junior, Botucatu 18618-686, São Paulo State, Brazil. ligia.sassaki@unesp.br

### Abstract

The gut microbiota is a complex community of microorganisms that inhabit the digestive tracts of humans, living in symbiosis with the host. Dysbiosis, characterized by an imbalance between the beneficial and opportunistic gut microbiota, is associated with several gastrointestinal disorders, such as irritable bowel syndrome (IBS); inflammatory bowel disease (IBD), represented by ulcerative colitis and Crohn's disease; and colorectal cancer (CRC). Dysbiosis can disrupt the mucosal barrier, resulting in perpetuation of inflammation and carcinogenesis. The increase in some specific groups of harmful bacteria, such as Escherichia coli ( E. coli) and enterotoxigenic Bacteroides fragilis (ETBF), has been associated with chronic tissue inflammation and the release of pro-inflammatory and carcinogenic mediators, increasing the chance of developing CRC, following the inflammationdysplasia-cancer sequence in IBD patients. Therefore, the aim of the present review was to analyze the correlation between changes in the gut microbiota and the development and maintenance of IBD, CRC, and IBD-associated CRC. Patients with IBD and CRC have shown reduced bacterial diversity and abundance compared to healthy individuals, with enrichment of Firmicute sand Bacteroidetes. Specific bacteria are also associated with the onset and progression of CRC, such as Fusobacterium nucleatum, E. coli, Enterococcus faecalis, Streptococcus gallolyticus, and ETBF. Future research can evaluate the advantages of modulating the gut microbiota as preventive measures in CRC high-risk patients, directly affecting the prognosis of the disease and the quality of life of patients.

Key Words: Gut microbiota; Dysbiosis; Ulcerative colitis; Crohn's disease; Inflammatory bowel disease; Colorectal cancer



WJG | https://www.wjgnet.com

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

**Core Tip:** Dysbiosis is present in patients with inflammatory bowel disease (IBD) and colorectal cancer (CRC). Dysbiosis can lead to a disruption of the mucosal barrier of the digestive tract lining, resulting in the perpetuation of inflammation and carcinogenesis, thus increasing the risk of developing CRC in IBD patients. Therefore, the aim of this review was to analyze the correlation between changes in gut microbiota and the development of IBD, CRC, and IBD-associated CRC. Further studies should be carried out to identify bacterial species that cause imbalances in the gut microbiota, enabling the development of prevention strategies or treatment of IBD-associated CRC.

Citation: Quaglio AEV, Grillo TG, De Oliveira ECS, Di Stasi LC, Sassaki LY. Gut microbiota, inflammatory bowel disease and colorectal cancer. *World J Gastroenterol* 2022; 28(30): 4053-4060 URL: https://www.wjgnet.com/1007-9327/full/v28/i30/4053.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i30.4053

#### INTRODUCTION

The human microbiome is composed of several types of microbes that colonize different niches of the human body, such as the skin, lungs, vagina, and gastrointestinal tract[1]. Of these, the gut microbiota is the most studied, due to the presence of greater diversity and number of microbial species compared to other parts of the body[1,2]. The gut microbiota is a complex community of approximately 100 trillion microorganisms, such as fungi, viruses, and protists, that inhabit the digestive tracts of humans, living in symbiosis with the host[3,4].

Alterations in the gut microbiota are associated with several diseases, including inflammatory bowel disease (IBD) and colorectal cancer (CRC)[2]. Thus, studies aimed at elucidating the role of the microbiota in each phase of each disease are necessary. Based on changes in the intestinal microbiota, new diagnostic tools and possible treatments can be developed. Further, studies aiming to correlate alterations in the microbiota or in specific bacterial populations with the diagnosis, prognosis, and/or treatment of diseases are necessary. Therefore, this mini-review was conducted to determine the correlation between changes in the gut microbiota and IBD, CRC, and IBD-associated CRC.

At birth, the intestinal tract is sterile<sup>[2]</sup>, with bacterial colonization beginning as early as during the passage of the baby through the birth canal<sup>[5]</sup>. Colonization continues through feeding and other environmental contacts. Several factors are known to influence colonization, including gestational age, mode of delivery (vaginal birth *vs* cesarean delivery), diet (breast milk *vs* formula) and exposure to antibiotics, and sanitation<sup>[2]</sup>.

During the first year of life, the microbial composition of the mammalian intestine is relatively simple and characterized by low diversity with relative dominance of species from the phyla *Proteobacteria* and *Actinobacteria*[2,5]. Over time, the microbiota becomes more diverse, with the *Firmicutes* and *Bacteroidetes* phyla later emerging as the predominant microbes characterizing the adult microbiota[2,6-8].

In an adult individual, over 90% of intestinal bacteria belong to *Firmicutes, Bacteroidetes, Proteobacteria*, or *Actinobacteria* phyla[4,7]. The gut microbiota can be differentiated into beneficial bacteria and opportunistic bacteria, the latter of which can cause infection[8]. *Lactobacillus, Bifidobacterium, Enterococci*, and *Propionobacteria* are some of the beneficial microbes, while the opportunistic groups include *Bacteriodes, Bacilli, Clostridia, Enterobacteria, Actinobacteria, Peptococci, Staphylococci*, and *Streptococci*[8].

The benefits of the gut microbiota to the host's physiology are vast, including nutrition, immune development, and host defense[4,7]. In the field of nutrition, some bacterial species, such as *Bifidobacterium*, participate in the biosynthesis of several components as vitamin K and vitamin B[7]. In addition, some can provide short-chain fatty acids (SCFAs) by fermenting dietary fibers[7,9]. SCFAs are a group of small fatty acids (2 to 5 carbons) produced by anaerobic microorganisms that, after absorption, have systemic immunomodulatory and anti-inflammatory properties[9].

The most abundant SCFAs in the colon are acetate, propionate, and butyrate[7,9,10], which promote the proliferation of beneficial bacteria and stimulate regulatory T cells to reduce inflammatory mediators, associated with an increase in colonic oxygen consumption by epithelial cells, and enhancing immune regulation, and gut barrier function[9-11].

Imbalance between the beneficial and opportunistic groups causes dysbiosis, which is associated with gastrointestinal disorders such as IBS, IBD, and CRC[4,7,8].

Zaisbideng® WJG | https://www.wjgnet.com

#### **IBD AND GUT MICROBIOTA**

IBD, a heterogeneous chronic and relapsing inflammatory illnesses of the digestive system, is traditionally classified into ulcerative colitis (UC) and Crohn's disease (CD)[4,12,13]. Both UC and CD can cause several symptoms like diarrhea, rectal bleeding, and abdominal pain. However, inflammation in CD is transmural and can involve any part of the gastrointestinal tract. In contrast, inflammation in UC is more superficial and limited to the colon[14].

Although the cause of IBD is not fully known, genetic history is likely involved in the pathophysiology of IBD, thus, several studies have identified a number of susceptible genes[12]. However, environmental factors, such as stress, sleeping patterns, antibiotic use, hygiene, diet, and smoking, are also associated with the development of IBD[4].

While IBD is a disease with the highest rates in industrialized countries, its incidence varies considerably worldwide, and newly industrialized areas such as India and South America presented a rapid increase in case numbers[7,13,14]. This enhancement can be related to dietary habits such as increased consumption of processed foods, sugars, and fats; overutilization of antibiotics; and an overall improvement in sanitary conditions and hygiene[13].

Nevertheless, the exact factors that trigger the first episode of inflammation and subsequent relapse remain unknown, it is now know that inflammatory episodes can result, in genetically predisposed people, from an immune response against intestinal microbial antigens under several environmental conditions[14]. Several IBD-associated susceptible genes are correlated with host responses to gut bacteria, suggesting that the gut microbiota also participate in the pathogenesis of IBD[7]. Dysbiosis is a reduction in microbial diversity, or a combination of the loss of beneficial bacteria and an increase in pathogenic bacteria[15], that may influence the pathogenesis of IBD.

The most prominent changes in the microbiota of IBD patients are the decreased diversity in bacteria species associated with decreased abundance of *Bacteroidetes* and *Firmicutes*[4,7] alongside an increase in the abundance of *Proteobacteria*[4].

In patients with IBD, the mucus layer is compromised, allowing luminal bacteria to penetrate and invade the submucosal layers, leading to the proliferative and inflammatory processes[4,13]. Then, mucosal destruction due to inflammatory injury further exposes the submucosa to more bacteria, leading to a vicious, positive feedback cycle of antigenic exposure and mucosal damage[13].

*Faecalibacterium prausnitzii* is one of the most abundant human gut bacteria and can be used as synonym of the gut health because of its anti-inflammatory and immunoregulatory properties[14]. One of the its products is butyrate, which is associated with an anti-inflammatory effect[7]. Several authors have correlated decreases in *Faecalibacterium prausnitzii* (*F. prausnitzii*) abundance with CD development [7,14,16] and in UC patients, a decrease was also observed during remission, being the recovery of the *F. prausnitzii* population associated with the maintenance of clinical remission[17].

*Roseburia* spp, including *Roseburia faecis, Roseburia intestinalis* (*R. intestinalis*), and *Roseburia hominis* (*R. hominis*), another genus of butyrate-producing bacteria, were significantly lower in healthy individuals with a high genetic risk for IBD compared to healthy individuals with low genetic risk[7]. *R. hominis,* specifically, was significantly reduced in UC patients, and an inverse correlation was observed between *R hominis* and disease severity[18]. In contrast, *R. intestinalis* supernatant suppressed the expression of interleukin (IL)-6 as well as the signal transducer and activator of transcription 3 (STAT3) *via* macrophage regulation in an *in vitro* experiment. Additionally, in dextran sodium sulfate- and 2,4,6-trinitrobenzene sulfonic acid-induced intestinal inflammation models, *R. intestinalis* supernatant reduced macrophages and Th17 cells in the colon, which was associated with the downregulation of IL-6 and STAT3[19].

In the field of pathogenic bacteria, several authors have described a relative increase in *Proteobacteria*, mainly *Escherichia coli* (*E. coli*), in IBD patients[7,20-22]. The exact mechanisms that lead to an increase in *Proteobacteria* during inflammation are not completely understood; however, Rizzatti *et al*[20] proposed two mechanisms: The oxygen hypothesis and the presence of nitrate. In a normal colon, epithelial cells deplete oxygen in the lumen through beta oxidation, creating an anaerobic environment[23]. In contrast, during an inflammatory episode, the beta oxidation capacity of colonic cells is decreased, thus increasing oxygen availability, which promotes dysbiosis and *Proteobacteria* growth[24]. Nitrate generated as a by-product during the inflammatory process conferred a growth advantage to the commensal bacteria *E. coli* in the large intestine, which then became predominant[25].

The increase in pathogenic bacteria in the digestive tract that can adhere to the colonic mucosa could alters the gut permeability, modifying the diversity and composition of intestinal microbiota, and ultimately leading to intestinal inflammation by regulating inflammatory genes expression[7].

Dysbiosis can further alter bacterial metabolites; decreased concentration of SCFAs has been reported as a result of a diminish in butyrate-producing bacteria like *F. prausnitzii*, in patients with IBD and in animal models of intestinal inflammation[7,9,10]. Decreased levels of SCFAs affects the differentiation and expansion of Treg cells and the growth of epithelial cells[26], leading to the loss of intestinal homeostasis.

Zaishidene® WJG | https://www.wjgnet.com

#### **CRC AND GUT MICROBIOTA**

CRC is the third most common cancer, and the second most frequent cause of cancer deaths worldwide being more common in developed than in developing countries [27,28]. Notwithstanding cases in all countries are on the rise, most CRC cases occur in Western countries, where there is an increase in incidence<sup>[28]</sup>. CRC rates in older adults in the United States have declined in recent years probably due to increased screening, but rates in younger adults have been rising, which may be correlated with higher incidence of obesity and other diet and lifestyle trends in the western hemisphere[29].

The exact mechanism for CRC onset and progression has not been fully elucidated, but it is generally believed to be the result of extensive and complex interactions between genetic and environmental factors[30]. According to Olovo et al[28], the progression of adenoma to carcinoma could be linked to the gut microbiota and conversely, a healthy microbiota is correlated with a minor risk of advanced adenoma<sup>[28]</sup>.

Several factors that affect the gut microbiota are thought to be related to colon carcinogenesis, such as obesity, a high-fat diet, smoking, and frequent consumption of alcohol[30].

Patients with CRC have shown reduced bacterial diversity and abundance compared to healthy individuals, with enrichment of Firmicutes and Bacteroidetes[31]. Specific bacteria are also associated with the onset and progression of CRC, such as Fusobacterium nucleatum, E. coli, Enterococcus faecalis, Streptococcus gallolyticus, and enterotoxigenic Bacteroides fragilis (ETBF)[32].

The abundance of Fusobacterium nucleatum, an opportunistic pathogen found normally in the oral cavity, is correlated with age and tumor diameter in CRC patients[31,33,34]. Additionally, an overabundance of F. nucleatum is associated with poor prognosis in metastatic CRC[32]; thus, F. nucleatum could be considered a potential biomarker for predicting the prognosis in patients with proximal colon cancer[34].

There is plenty of evidence of the tumor-promoting effects of *Streptococcus gallolyticus* in colon cells [35-37]. Colonic cells incubated with Streptococcus gallolyticus (S. gallolyticus) presented increased levels of β-catenin, c-Myc, and proliferating cell nuclear antigen (PCNA), all of which are transcription factors associated with cancer development[35]. In addition, in mice, administration of S. gallolyticus leads to more tumors, higher tumor burden, and dysplasia grade, and increased cell proliferation and β-catenin staining in colonic crypts compared to mice incubated with control bacteria[35]. Furthermore, CRC patients present high levels of this bacterium compared to healthy individuals[37]. However, CRCspecific conditions, such as the increased concentration of bile acids, could also promote S. gallolyticus colonization, creating a maintenance cycle for high levels of this bacterium in the gut[37].

ETBF is the most frequent anaerobe isolated from cases of diarrhea, peritonitis, intra-abdominal abscesses, and sepsis, and there is a positive correlation between the presence of ETBF and active IBD and CRC[38]. According to Appunni et al[39], co-colonization of toxigenic E. coli and ETBF in mice resulted in increased production of pro-inflammatory IL-17 and subsequent DNA damage, which could accelerate the development of CRC[39]. In addition, the toxin in ETBF could induce c-myc expression and IL-8 secretion causing oxidative DNA and epithelial barrier damage, and STAT3/Th17 immune responses activation, which are further correlated with an increased risk of CRC[38].

#### IBD-ASSOCIATED CRC AND GUT MICROBIOTA

Intestinal inflammation is one of the most common risk factors for developing CRC. Besides being two to six times more likely to develop CRC than healthy people, IBD patients with cancer are affected younger than sporadic CRC patients[40]. Disease duration, extension of the lesions, inflammation, sclerosing cholangitis, age at onset and onset in childhood are factors linking IBD with an increased incidence of CRC[41,42]

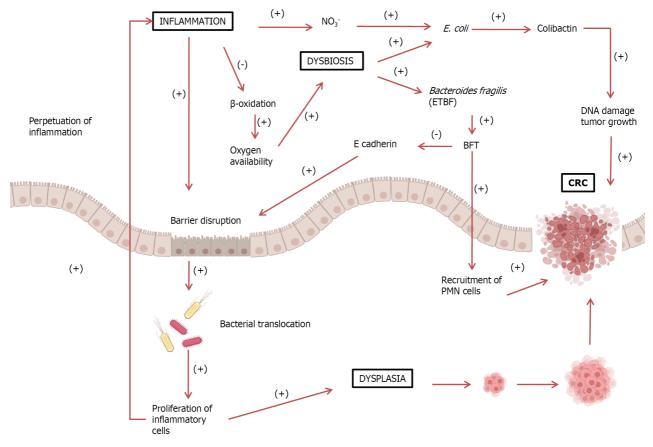
Chronic inflammation initiates and drives tumorigenesis[42], leading to an "inflammation-dysplasiacarcinoma" sequence, not the "adenoma-sequence" classically described in sporadic CRC[40].

Popov et al[43]proposed the three main theories about bacterial involvement in the development of IBD-associated CRC: The alpha-bug hypothesis, driver-passenger hypothesis, and common ground hypothesis. In the alpha-bug hypothesis, a single bacterium (normally ETBF) is thought to cause all modifications and damage that lead to carcinogenesis. The driver-passenger hypothesis is similar, but after the attack by the first bacteria, other opportunistic bacteria start to grow and contribute to cancer development. Finally, in the common ground hypothesis, endogenous and exogenous factors form a "leaky gut," allowing the passage of bacteria into the submucosal tissue, leading to chronic inflammation and the consequent emergence of cancer[43].

It is known that bacteria, through pathogen-associated molecular patterns (PAMPs), are capable of communicating with the tool-like receptors (TLRs), retinoic acid-inducible gene I-like receptors (RLRs), and nucleotide binding oligomerization domain-receptors (NLRs), and trigger an immune response [42]. Nuclear factor kB (NF-kB) can be activated by the TLR and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), inducing the transcription of several tumorigenesis genes such as COX-2, which then leads to the apoptosis of intestinal epithelial cells via tumor suppressor p53 pathways<sup>[40]</sup> and consequent



WJG | https://www.wjgnet.com



DOI: 10.3748/wjg.v28.i30.4053 Copyright ©The Author(s) 2022.

**Figure 1 Increased inflammatory response can lead to barrier disruption, allowing bacterial translocation into the intestinal lumen.** These pathogens trigger an immune and inflammatory response, perpetuating the inflammatory process. Inflammatory cells can also cause dysplasia, leading to the development of colorectal cancer (CRC). Under normal conditions, epithelial cells, during β-oxidation, decrease oxygen availability, thus creating an anaerobic environment. During chronic inflammation, β-oxidation decreases, enhancing oxygen availability. This also leads to an increase in nitrate (NO3-) formation, leading to dysbiosis and growth of proteobacteria such as *Escherichia coli (E. coli)*. *E. coli* produces colibactin, which could damage DNA and stimulate tumor growth. The growth of Bacteroidetes fragilis may also occur, which produces Bacteroides fragilis toxin (BFT). This toxin cleaves E-cadherin, a major constituent of the zonula adherens, which is responsible for cell adhesion, leading to further barrier disruption. Additionally, BFT also stimulates epithelial cells to recruit polymorphonuclear leukocytes. (PMN) cells, promoting the development of CRC. BFT: Bacteroides fragilis toxin; ETBF: Enterotoxigenic *Bacteroides fragilis*; CRC: Colorectal cancer; PMN: Polymorphonuclear leukocytes.

breakdown of the intestinal barrier, allowing microbial translocation.

Increased quantities of *Proteobacteria* are associated with decreased levels of SCFAs and increased colonic inflammation[43]. In addition, intestinal Dysbiosis and barrier defects, associated with alterations in mucin secretion may happen even in the absence of active inflammation[43], perpetuating a favorable environment for the emergence of CRC.

Several studies have demonstrated the participation of bacteria such as *E. coli*, ETBF, and *Fusobacterium nucleatum* in chronic inflammation and cancer development in IBD patients[40,42-45].

A review by Yu *et al*[44] concluded that commensal bacteria, *E. coli* and *E. faecalis*, isolated from healthy subjects are capable of initiating intestinal inflammation in genetically deficient mice, suggesting that opportunistic commensal bacteria may become pathogenic only in genetically predisposed hosts[44]. Nevertheless, some types of *E. coli* can produce colibactin, a genotoxic compound capable of promoting tumor growth, alkylating DNA, and inducing tumorigenic double-stranded breaks[45]. According to Yang *et al*[45], inflammation may upregulate bacterial virulence genes, such as colibactin, and facilitate colonization of the mucosa, leading to increased colibactin-induced DNA damage in colonic epithelial cells, allowing this bacterial strain to exert its carcinogenic activity[42].

The enterotoxin produced by ETBF known as bacteroides fragilis toxin, acted as a metalloprotease cleaving E-cadherin, a major constituent of the zonula adherens, which is responsible for cell adhesion [46]. This toxin can trigger an inflammatory, pro-tumoral signaling caspase in colonic epithelial cells that cause the recruitment of polymorphonuclear immature myeloid cells to promote colon cancers[44].

WJG | https://www.wjgnet.com

#### CONCLUSION

There is a consensus that dysbiosis is present in both IBD and CRC, and that dysbiosis could lead to a disruption of the mucosal barrier with perpetuation of inflammation and carcinogenesis. Dysbiosis with a consequent increase in bacteria, such as E. coli and ETBF, is believed to lead to a breakdown of the intestinal mucosal barrier, allowing the translocation of more bacteria from the lumen to the interior of the tissue. This condition leads to chronic tissue inflammation, with the release of inflammatory and pro-carcinogenic mediators increasing the risk of developing CRC. This positive feedback loop of dysbiosis could be the basis for the inflammation-dysplasia-cancer sequence (Figure 1).

Further studies should be carried out to identify which bacteria or which set of bacteria may be responsible for this feedback cycle. Understanding the mechanism behind dysbiosis cycles may be the basis for preventing or treating IBD-associated CRC and CRC. Future research should evaluate the advantages of modulating the intestinal microbiota as a protective factor for the development of IBDassociated CRC; thus, the use of prebiotics, probiotics, or diet-based treatment can be used as preventive measures in CRC high-risk patients, directly affecting the prognosis of the disease and the quality of life of patients living with IBD.

### FOOTNOTES

Author contributions: Quaglio AEV, Grillo TG, and De Oliveira ECS performed the majority of the writing; Di Stasi LC and Sassaki LY designed the outline and coordinated the writing of the paper; all authors critically revised the manuscript for important intellectual content and approved the final version.

Conflict-of-interest statement: There are no conflicts of interest to report.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is noncommercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

#### Country/Territory of origin: Brazil

ORCID number: Ana Elisa Valencise Quaglio 0000-0002-5998-2382; Thais Gagno Grillo 0000-0002-4351-5034; Ellen Cristina Souza De Oliveira 0000-0001-5357-3468; Luiz Claudio Di Stasi 0000-0002-7864-1073; Ligia Yukie Sassaki 0000-0002-7319-8906.

S-Editor: Chen YL L-Editor: A P-Editor: Chen YL

#### REFERENCES

- 1 Michaudel C. Sokol H. The Gut Microbiota at the Service of Immunometabolism. Cell Metab 2020: 32: 514-523 [PMID: 32946809 DOI: 10.1016/j.cmet.2020.09.004]
- Quigley EM. Gut bacteria in health and disease. Gastroenterol Hepatol (N Y) 2013; 9: 560-569 [PMID: 24729765] 2
- Kc D, Sumner R, Lippmann S. Gut microbiota and health. Postgrad Med 2020; 132: 274 [PMID: 31566046 DOI: 3 10.1080/00325481.2019.1662711]
- Gomaa EZ. Human gut microbiota/microbiome in health and diseases: a review. Antonie Van Leeuwenhoek 2020; 113: 2019-2040 [PMID: 33136284 DOI: 10.1007/s10482-020-01474-7]
- Sekirov I, Russell SL, Antunes LC, Finlay BB. Gut microbiota in health and disease. Physiol Rev 2010; 90: 859-904 [PMID: 20664075 DOI: 10.1152/physrev.00045.2009]
- Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargent M, Gill SR, Nelson KE, Relman DA. Diversity of the human intestinal microbial flora. Science 2005; 308: 1635-1638 [PMID: 15831718 DOI: 10.1126/science.1110591]
- 7 Nishida A, Inoue R, Inatomi O, Bamba S, Naito Y, Andoh A. Gut microbiota in the pathogenesis of inflammatory bowel disease. Clin J Gastroenterol 2018; 11: 1-10 [PMID: 29285689 DOI: 10.1007/s12328-017-0813-5]
- 8 Roy Sarkar S, Banerjee S. Gut microbiota in neurodegenerative disorders. J Neuroimmunol 2019; 328: 98-104 [PMID: 30658292 DOI: 10.1016/j.jneuroim.2019.01.004]
- Almeida-Junior LD, Curimbaba TFS, Chagas AS, Quaglio AEV, Di Stasi LC. Dietary intervention with green dwarf banana flour (Musa sp. AAA) modulates oxidative stress and colonic SCFAs production in the TNBS model of intestinal inflammation. J Funct Foods 2017; 38: 497-504
- Curimbaba TFS, Almeida-Junior LD, Chagas AS, Quaglio AEV, Herculano AM, Di Stasi LC. Prebiotic, antioxidant and 10 anti-inflammatory properties of edible Amazon fruits. Food Biosci 2020; 100599
- Mills S, Stanton C, Lane JA, Smith GJ, Ross RP. Precision Nutrition and the Microbiome, Part I: Current State of the 11



Science. Nutrients 2019; 11 [PMID: 31022973 DOI: 10.3390/nu11040923]

- 12 Matsuoka K, Kanai T. The gut microbiota and inflammatory bowel disease. Semin Immunopathol 2015; 37: 47-55 [PMID: 25420450 DOI: 10.1007/s00281-014-0454-4]
- 13 Ghouri YA, Tahan V, Shen B. Secondary causes of inflammatory bowel diseases. World J Gastroenterol 2020; 26: 3998-4017 [PMID: 32821067 DOI: 10.3748/wjg.v26.i28.3998]
- 14 Ryma T, Samer A, Soufli I, Rafa H, Touil-Boukoffa C. Role of Probiotics and Their Metabolites in Inflammatory Bowel Diseases (IBDs). Gastroenterol Insights 2021; 12: 56-66 [DOI: 10.3390/gastroent12010006]
- Humphreys C. Intestinal Permeability. 5th ed. Pizzorno JE, Murray, MT, editors. Textbook of Natural Medicine, 2020: 15 166-177 [DOI: 10.1016/B978-0-323-43044-9.00019-4]
- Zuo T, Ng SC. The Gut Microbiota in the Pathogenesis and Therapeutics of Inflammatory Bowel Disease. Front Microbiol 16 2018; 9: 2247 [PMID: 30319571 DOI: 10.3389/fmicb.2018.02247]
- Varela E, Manichanh C, Gallart M, Torrejón A, Borruel N, Casellas F, Guarner F, Antolin M. Colonisation by 17 Faecalibacterium prausnitzii and maintenance of clinical remission in patients with ulcerative colitis. Aliment Pharmacol Ther 2013; 38: 151-161 [PMID: 23725320 DOI: 10.1111/apt.12365]
- 18 Machiels K, Joossens M, Sabino J, De Preter V, Arijs I, Eeckhaut V, Ballet V, Claes K, Van Immerseel F, Verbeke K, Ferrante M, Verhaegen J, Rutgeerts P, Vermeire S. A decrease of the butyrate-producing species Roseburia hominis and Faecalibacterium prausnitzii defines dysbiosis in patients with ulcerative colitis. Gut 2014; 63: 1275-1283 [PMID: 24021287 DOI: 10.1136/gutjnl-2013-304833]
- Luo W, Shen Z, Deng M, Li X, Tan B, Xiao M, Wu S, Yang Z, Zhu C, Tian L, Wu X, Meng X, Quan Y, Wang X. 19 Roseburia intestinalis supernatant ameliorates colitis induced in mice by regulating the immune response. Mol Med Rep 2019; 20: 1007-1016 [PMID: 31173202 DOI: 10.3892/mmr.2019.10327]
- Rizzatti G, Lopetuso LR, Gibiino G, Binda C, Gasbarrini A. Proteobacteria: A Common Factor in Human Diseases. 20 Biomed Res Int 2017; 2017: 9351507 [PMID: 29230419 DOI: 10.1155/2017/9351507]
- 21 Nishino K, Nishida A, Inoue R, Kawada Y, Ohno M, Sakai S, Inatomi O, Bamba S, Sugimoto M, Kawahara M, Naito Y, Andoh A. Analysis of endoscopic brush samples identified mucosa-associated dysbiosis in inflammatory bowel disease. J Gastroenterol 2018; 53: 95-106 [PMID: 28852861 DOI: 10.1007/s00535-017-1384-4]
- 22 Vester-Andersen MK, Mirsepasi-Lauridsen HC, Prosberg MV, Mortensen CO, Träger C, Skovsen K, Thorkilgaard T, Nøjgaard C, Vind I, Krogfelt KA, Sørensen N, Bendtsen F, Petersen AM. Increased abundance of proteobacteria in aggressive Crohn's disease seven years after diagnosis. Sci Rep 2019; 9: 13473 [PMID: 31530835 DOI: 10.1038/s41598-019-49833-3
- 23 Rivera-Chávez F, Lopez CA, Bäumler AJ. Oxygen as a driver of gut dysbiosis. Free Radic Biol Med 2017; 105: 93-101 [PMID: 27677568 DOI: 10.1016/j.freeradbiomed.2016.09.022]
- 24 Hughes ER, Winter MG, Duerkop BA, Spiga L, Furtado de Carvalho T, Zhu W, Gillis CC, Büttner L, Smoot MP, Behrendt CL, Cherry S, Santos RL, Hooper LV, Winter SE. Microbial Respiration and Formate Oxidation as Metabolic Signatures of Inflammation-Associated Dysbiosis. Cell Host Microbe 2017; 21: 208-219 [PMID: 28182951 DOI: 10.1016/j.chom.2017.01.005
- Winter SE, Winter MG, Xavier MN, Thiennimitr P, Poon V, Keestra AM, Laughlin RC, Gomez G, Wu J, Lawhon SD, 25 Popova IE, Parikh SJ, Adams LG, Tsolis RM, Stewart VJ, Bäumler AJ. Host-derived nitrate boosts growth of E. coli in the inflamed gut. Science 2013; 339: 708-711 [PMID: 23393266 DOI: 10.1126/science.1232467]
- Atarashi K, Tanoue T, Oshima K, Suda W, Nagano Y, Nishikawa H, Fukuda S, Saito T, Narushima S, Hase K, Kim S, 26 Fritz JV, Wilmes P, Ueha S, Matsushima K, Ohno H, Olle B, Sakaguchi S, Taniguchi T, Morita H, Hattori M, Honda K. Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. Nature 2013; 500: 232-236 [PMID: 23842501 DOI: 10.1038/nature12331]
- 27 Weitz J, Koch M, Debus J, Höhler T, Galle PR, Büchler MW. Colorectal cancer. Lancet 2005; 365: 153-165 [PMID: 15639298 DOI: 10.1016/S0140-6736(05)17706-X]
- Olovo CV, Huang X, Zheng X, Xu M. Faecal microbial biomarkers in early diagnosis of colorectal cancer. J Cell Mol Med 28 2021; 25: 10783-10797 [PMID: 34750964 DOI: 10.1111/jcmm.17010]
- Kehm RD, Lima SM, Swett K, Mueller L, Yang W, Gonsalves L, Terry MB. Age-specific Trends in Colorectal Cancer Incidence for Women and Men, 1935-2017. Gastroenterology 2021; 161: 1060-1062.e3 [PMID: 34058214 DOI: 10.1053/j.gastro.2021.05.050
- Zhang W, An Y, Qin X, Wu X, Wang X, Hou H, Song X, Liu T, Wang B, Huang X, Cao H. Gut Microbiota-Derived 30 Metabolites in Colorectal Cancer: The Bad and the Challenges. Front Oncol 2021; 11: 739648 [PMID: 34733783 DOI: 10.3389/fonc.2021.739648
- Silva M, Brunner V, Tschurtschenthaler M. Microbiota and Colorectal Cancer: From Gut to Bedside. Front Pharmacol 31 2021; 12: 760280 [PMID: 34658896 DOI: 10.3389/fphar.2021.760280]
- 32 Liu K, Yang X, Zeng M, Yuan Y, Sun J, He P, Xie Q, Chang X, Zhang S, Chen X, Cai L, Xie Y, Jiao X. The Role of Fecal Fusobacterium nucleatum and pks+ Escherichia coli as Early Diagnostic Markers of Colorectal Cancer. Dis Markers 2021; 2021: 1171239 [PMID: 34853619 DOI: 10.1155/2021/1171239]
- Brennan CA, Garrett WS. Fusobacterium nucleatum symbiont, opportunist and oncobacterium. Nat Rev Microbiol 2019; 33 17: 156-166 [PMID: 30546113 DOI: 10.1038/s41579-018-0129-6]
- Jin M, Shang F, Wu J, Fan Q, Chen C, Fan J, Liu L, Nie X, Zhang T, Cai K, Ogino S, Liu H. Tumor-Associated 34 Microbiota in Proximal and Distal Colorectal Cancer and Their Relationships With Clinical Outcomes. Front Microbiol 2021; 12: 727937 [PMID: 34650531 DOI: 10.3389/fmicb.2021.727937]
- Kumar R, Herold JL, Schady D, Davis J, Kopetz S, Martinez-Moczygemba M, Murray BE, Han F, Li Y, Callaway E, Chapkin RS, Dashwood WM, Dashwood RH, Berry T, Mackenzie C, Xu Y. Streptococcus gallolyticus subsp. gallolyticus promotes colorectal tumor development. PLoS Pathog 2017; 13: e1006440 [PMID: 28704539 DOI: 10.1371/journal.ppat.1006440]
- Kumar R, Herold JL, Taylor J, Xu J, Xu Y. Variations among Streptococcus gallolyticus subsp. gallolyticus strains in connection with colorectal cancer. Sci Rep 2018; 8: 1514 [PMID: 29367658 DOI: 10.1038/s41598-018-19941-7]



- Oehmcke-Hecht S, Mandl V, Naatz LT, Dühring L, Köhler J, Kreikemeyer B, Maletzki C. Streptococcus gallolyticus 37 abrogates anti-carcinogenic properties of tannic acid on low-passage colorectal carcinomas. Sci Rep 2020; 10: 4714 [PMID: 32170212 DOI: 10.1038/s41598-020-61458-5]
- Haghi F, Goli E, Mirzaei B, Zeighami H. The association between fecal enterotoxigenic B. fragilis with colorectal cancer. 38 BMC Cancer 2019; 19: 879 [PMID: 31488085 DOI: 10.1186/s12885-019-6115-1]
- 39 Appunni S, Rubens M, Ramamoorthy V, Tonse R, Saxena A, McGranaghan P, Kaiser A, Kotecha R. Emerging Evidence on the Effects of Dietary Factors on the Gut Microbiome in Colorectal Cancer. Front Nutr 2021; 8: 718389 [PMID: 34708063 DOI: 10.3389/fnut.2021.718389]
- Keller DS, Windsor A, Cohen R, Chand M. Colorectal cancer in inflammatory bowel disease: review of the evidence. Tech 40 Coloproctol 2019; 23: 3-13 [PMID: 30701345 DOI: 10.1007/s10151-019-1926-2]
- 41 Olén O, Erichsen R, Sachs MC, Pedersen L, Halfvarson J, Askling J, Ekbom A, Sørensen HT, Ludvigsson JF. Colorectal cancer in ulcerative colitis: a Scandinavian population-based cohort study. Lancet 2020; 395: 123-131 [PMID: 31929014 DOI: 10.1016/S0140-6736(19)32545-0]
- 42 Lucafò M, Curci D, Franzin M, Decorti G, Stocco G. Inflammatory Bowel Disease and Risk of Colorectal Cancer: An Overview From Pathophysiology to Pharmacological Prevention. Front Pharmacol 2021; 12: 772101 [PMID: 34744751 DOI: 10.3389/fphar.2021.772101]
- 43 Popov J, Caputi V, Nandeesha N, Rodriguez DA, Pai N. Microbiota-Immune Interactions in Ulcerative Colitis and Colitis Associated Cancer and Emerging Microbiota-Based Therapies. Int J Mol Sci 2021; 22 [PMID: 34768795 DOI: 10.3390/ijms222111365]
- 44 Yu LC. Microbiota dysbiosis and barrier dysfunction in inflammatory bowel disease and colorectal cancers: exploring a common ground hypothesis. J Biomed Sci 2018; 25: 79 [PMID: 30413188 DOI: 10.1186/s12929-018-0483-8]
- 45 Yang Y, Gharaibeh RZ, Newsome RC, Jobin C. Amending microbiota by targeting intestinal inflammation with TNF blockade attenuates development of colorectal cancer. Nat Cancer 2020; 1: 723-734 [PMID: 33768208 DOI: 10.1038/s43018-020-0078-7]
- 46 Zamani S, Taslimi R, Sarabi A, Jasemi S, Sechi LA, Feizabadi MM. Enterotoxigenic Bacteroides fragilis: A Possible Etiological Candidate for Bacterially-Induced Colorectal Precancerous and Cancerous Lesions. Front Cell Infect Microbiol 2019; 9: 449 [PMID: 32010637 DOI: 10.3389/fcimb.2019.00449]





## Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

