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# **REVIEW ARTICLE**

# Sigma receptors [ $\sigma$ Rs]: biology in normal and diseased states

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#### Abstract

This review compares the biological and physiological function of Sigma receptors [\sigmaRs] and their potential therapeutic roles. Sigma receptors are widespread in the central nervous system and across multiple peripheral tissues.  $\sigma Rs$  consist of sigma receptor one ( $\sigma_1 R$ ) and sigma receptor two ( $\sigma_2 R$ ) and are expressed in numerous regions of the brain. The sigma receptor was originally proposed as a subtype of opioid receptors and was suggested to contribute to the delusions and psychoses induced by benzomorphans such as SKF-10047 and pentazocine. Later studies confirmed that  $\sigma Rs$  are non-opioid receptors (not an  $\mu$  opioid receptor) and play a more diverse role in intracellular signaling, apoptosis and metabolic regulation.  $\sigma_1$ Rs are intracellular receptors acting as chaperone proteins that modulate Ca<sup>2+</sup> signaling through the IP<sub>3</sub> receptor. They dynamically translocate inside cells, hence are transmembrane proteins. The  $\sigma_1 R$  receptor, at the mitochondrial-associated endoplasmic reticulum membrane, is responsible for mitochondrial metabolic regulation and promotes mitochondrial energy depletion and apoptosis. Studies have demonstrated that they play a role as a modulator of ion channels (K<sup>+</sup> channels; N-methyl-Daspartate receptors [NMDAR]; inositol 1,3,5 triphosphate receptors) and regulate lipid transport and metabolism, neuritogenesis, cellular differentiation and myelination in the brain.  $\sigma_1 R$ modulation of Ca<sup>2+</sup> release, modulation of cardiac myocyte contractility and may have links to Gproteins. It has been proposed that  $\sigma_1$ Rs are intracellular signal transduction amplifiers. This review of the literature examines the mechanism of action of the  $\sigma$ Rs, their interaction with neurotransmitters, pharmacology, location and adverse effects mediated through them.

## Introduction

Sigma receptors  $[\sigma Rs]$  are a relatively novel group of receptors originally discovered in the central nervous system [CNS] of mammals in 1976 (1). They represent a ubiquitously expressed unique binding site in the CNS and other peripheral tissues (2–6).  $\sigma$ Rs are a member of the orphan receptor class for which no endogenous ligand was known until recently dimethyltryptamine [DMT] (7-9). They also bind with high affinity to several classes of chemically unrelated ligands such as neurosteroids (10), neuroleptics, dextrobenzomorphans [DEX] and several psychostimulants such as cocaine (11), methamphetamine [METH] (12,13) methylenedioxymethamphetamine [MDMA] (14) and methacathinone (15,16). Consequently, it is thought that the  $\sigma R$  may mediate the immunosuppressant, antipsychotic and neuroprotective effects of many drugs (17).

#### **Keywords**

Apoptosis, cannabinoids, central nervous system, glutamate, neoplasia, non-opioid receptors

## History

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Historically, the  $\sigma R$  was identified as one of the subtypes of opiate receptors, differentiated using a chronic spinal pain model in the dog, the unique psychomimetic effects induced by N-allylnormetazocine [SKF-10,047] (18) ( $\sigma$ -syndrome), from the effects induced by morphine ( $\mu$ -syndrome) and ketocyclazocine ( $\kappa$ -syndrome) (1). However, subsequent studies established that  $\sigma R$  sites possess negligible affinity for naloxone or naltrexone (19,20); thus, establishing a complete distinction between the non-opiate  $\sigma$  binding sites and the classical  $\mu$ -,  $\delta$ - and  $\mu$ -opiate receptors (21,22). It has recently been suggested that  $\sigma_1 R$  antagonism be used with opioids to increase pain control without increasing the adverse effects of the opioids (23).

Two subtypes of  $\sigma Rs$  were found originally: sigma-1 [ $\sigma_1 R$ ] and sigma-2 [ $\sigma_2 R$ ] (24–27). Although another subtype, sigma-3 [ $\sigma_3 R$ ], has been suggested, it has not been defined adequately (28,29).  $\sigma_1 Rs$  have been cloned (2), assayed (30) and their biological and physiological roles have been examined more intensively than  $\sigma_2 Rs$ , as until now  $\sigma_2 Rs$  have not been cloned (31).

 $\sigma_1 Rs$  regulate a number of neurotransmitter systems, including the glutamatergic [Glu], dopaminergic [DA], serotonergic [5HT], noradrenergic [NE] and cholinergic [Ch] systems. As these transmitters, which interact with the  $\sigma_1 Rs$ , are involved in many neuropsychiatric disorders their role has been evaluated in a number of these disorders (32). In fact,

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several lines of evidence have demonstrated that  $\sigma_1 R$  play a role in the pathophysiology of neuropsychiatric disorders such as mood (33), anxiety disorders (34,35) and schizophrenia (9).

Hence,  $\sigma R$  ligands are potential therapeutic agents for several neuropsychiatric disorders (36,37).  $\sigma_1 R$  has also been suggested as a target for the treatment of neuropathic pain (38,39) and a treatment for dementia, such as seen associated with Alzheimers disease [AD] (40). In addition,  $\sigma_1 R$  mutations have been implicated in frontotemporal lobar degeneration and motor neuron disease [MND] (41), diseases in which they have been shown to have a low density (42). It appears that there is an association between a variant of the  $\sigma_1 R$  gene and AD (43) where genetic polymorphisms in  $\sigma_1 R$  and apolipoprotein E interact to influence the severity of AD (44).

Many psychostimulant drugs, including cocaine (45) and METH (46,47), interact with  $\sigma Rs$  in the brain and heart, offering a logical target for medication development efforts (48).  $\sigma R$  antagonists and antisense oligonucleotides ameliorate cocaine-induced convulsions, lethality and locomotor activity (49,50), as well as sensitization, and conditioned place-preference in rodents (51). They also reduce alcohol consumption in alcohol-drinking rats (52,53) and Swiss mice (54). Interestingly, the interaction of fluvoxamine [Luvox], a selective serotonin reuptake inhibitor [SSRI], and the  $\sigma Rs$ may account for its potential amelioration of psychotic depression (55,40), where increased glutamate [Glu] release occurs through activation of serotonin [5-HT<sub>3</sub>] mediated by  $\sigma_1 Rs$  (56), and in patients with schizophrenia (57,40). These findings are supported by research on a depressive phenotype in  $\sigma_1 R$  knockout mice (53). In contrast, the SSRI sertraline worsens the symptoms (58). Not all SSRIs induce their antidepressant activity via the  $\sigma_1 R$ , e.g. paroxetine (59). This detailed review explores the  $\sigma Rs$  in normal homeostatic and diseased states. First, the structure and function of these receptors are described. Next, sites of  $\sigma Rs$ , disease states and their relationship to  $\sigma Rs$  are discussed.

#### Molecular biology of $\sigma Rs$

Due to their CNS pharmacological action, most work has been focused on evaluation of  $\sigma$ Rs in the CNS; however, considerable current research has also been directed toward neoplasia, its treatment and imaging ( $\sigma_2$ R) (60).  $\sigma$ Rs are highly expressed in all parts of the brain (25,61,62), where they are predominantly localized in the cell plasma membrane and at the endoplasmic reticulum [ER] of *both* neurons and oligodendrocytes (63). They are dynamically translocated upon ligand binding into cells from the cell membrane (64–66).  $\sigma_1$ Rs agonists provide protection of the ER from oxidative stress (67).

More recently, a  $\sigma_1 R$  receptor knockout mouse has been developed that displays a depressive-like phenotype, supporting the receptors importance in this psychiatric disorder (53). The database concerning the molecular biology of  $\sigma Rs$  is large.

# Sigma-1 receptors [ $\sigma$ 1Rs]

The two subclasses of  $\sigma R$  sites ( $\sigma_1 R$  and  $\sigma_2 R$ ), distinguished based on their different drug selectivity patterns and

molecular weights (21) have no homology to any other mammalian protein (2,68). However, several biochemical features have been observed for  $\sigma_1$ Rs, such as an allosteric modulation by phenytoin (69) and sensitivity to pertussis toxin or G-protein modulators (70–73), probably though potentiation of opioid transduction independent from receptor binding (74). The  $\sigma_1$ R site also shows a stereo selectivity with high affinity for the dextro isomers of benzomorphans [BZM], whereas  $\sigma_2$ R sites show the reverse stereo selectivity with a lower affinity range. 1,3,Di-*O*- tolylguanidin [DTG], 3-(3-Hydroxyphenyl)-N-n-propyliperidin (+) 3-PPP [preclamol] and haloperidol [Haladol<sup>®</sup>] are non-discriminating ligands with high affinity for both  $\sigma_1$ R and  $\sigma_2$ R subtypes (75).

The  $\sigma_1 R$  is a 29 kDa single polypeptide that has been cloned in mice, rats and humans (2,3,6,76,77), the ligand binding profile of which is similar to those described in brain homogenates studies (78,79). The  $\sigma_1 R$  gene, located on chromosome 9, band p13, in human and chromosome 2 in rodents, is approximately 7 kbp long and contains four exons, interrupted by three introns, where exon 3 is the shortest (93 bp) and exon 4 is the longest (1132 bp) (68). Exon 2 encodes 25 kDa membrane proteins for the single transmembrane domain, identified at present, but two other hydrophobic regions exist and one of them may putatively constitute a second transmembrane domain (80).

The  $\sigma_1 R$  sequence contains a 22 amino acid [AA] retention signal for the ER at its N-terminal region and two short Cterminal hydrophobic AA sequences that are probably involved in sterol binding (2). The 223 amino acid sequence of the purified protein is highly preserved, with 87–92% identity and 90–93% homology among tissues and animal species (81). This protein is identical in peripheral tissues and brain, and probably is similar in other tissues as well. It shares a similarity, 33% identity and 66% homology, with a sterol  $C_8-C_7$  isomerase (82), but nevertheless is different from any other mammalian protein identified (2,68), outlining the uniqueness of the  $\sigma_1 R$  as compared with any other known receptor.

Hydropathic analysis of the  $\sigma_1 R$  indicates three hydrophobic regions, with some evidence for two transmembrane segments. A crystal structure of the  $\sigma_1 R$  was unavailable at the time of writing, but a 3D model has recently been validated showing agreement of the *in vitro* and the *in* silico model (83).

The  $\sigma_1 R$  gene also has been isolated from human, guinea pig, mouse and rat (2,6,76). AA substitutions in transmembrane domains do not alter the expression levels of the protein but suppresses ligand binding activity (80), suggesting that these AAs belong to the binding site pharmacophore located within the transmembrane domain. In addition, anionic AA residues have been identified that also appear critical for ligand binding (68,77).

Exon-2 codes for a single transmembrane domain present in the  $\sigma R$  (68). The fact that the gene for the  $\sigma_1 R$  is located on chromosome 9p13, a region associated with psychiatric disorders (68), helps explain the psychiatric effects of  $\sigma_1 R$ agonists and antagonists.

A splice variant of the  $\sigma_1 R$  has been found in Jurkat cells, an immortalized line of T-lymphocyte cells (84) and in The  $\sigma_1 R$  has been cloned from guinea pig and mouse liver, human placental cell line, and human, mouse and rat brain (2–6). The protein cloned is a 223 AA, 1 transmembrane protein with potent (+)-pentazocine [PTZ], haloperidol, ditolylguanidine (1,3,di-*O*-tolylguanidin) [DTG] and (+)-3-PPP binding, but does not couple with G-proteins (5,76).

At this point, it is not completely clear whether the cloned  $\sigma_1 R$  is the ligand binding subunit of a multi-subunit complex or represents one subtype of the  $\sigma_1 R$ . A study investigating putative transmembrane segments based on homology identified two putative transmembrane segments for the  $\sigma_1 R$  (88). Thus, as research investigates the  $\sigma_R R$  further, subtypes of the  $\sigma_1 R$ ,  $\sigma_2 R$  and possibly the  $\sigma_3 R$  might be found.

Regardless, cloning has led to an important focus on the molecular biology and signal transduction mechanisms of  $\sigma_1 R$ , e.g. inhibition of Ca<sup>2+</sup> entry into epithelial cells (89). This is discussed in more detail in Sections " $\sigma_1 R$  ligands" and "Neoplasia". However, given the one-transmembrane segment cloned, it is most likely that it does not represent the complete functional receptor. More experiments using techniques such as the use of selective  $\sigma_1 R$  gene antisense will elucidate the exact structure of the functional  $\sigma R$  in the future (63).

## Sigma-2 receptors [ $\sigma_2 Rs$ ]

The  $\sigma_2 R$  site has not been cloned as of yet, but a comprehensive ligand based mapping of the receptor binding pocket has been done (90). The  $\sigma_2 R$  site was first characterized in pheochromocytoma PC12 cells (91), and has a low affinity for (+)-BZM and has an apparent molecular weight of 18 to 21 kDa (92). Some selective and high affinity  $\sigma_2 R$  site ligands are now available such as 1'-(4-(1-(4-fluorophenyl))-1H-indol-3-yl)-1-butyl)spiro (isobenzofuran-1(3H),4'piperidine [Lu 28-179] (93), N-[2-(3,4-dichlorophenyl)ethyl]-N-methyl-2-(1-pyrrolidinyl) ethylamine [BD1008] (92), and ibogaine (94). The site also appears to be important in the modulation of cellular Ca<sup>2+</sup> concentrations (Figure 1) (95).

Several attributes have been proposed for  $\sigma_2 R$  sites: stem cell differentiation (96); regulation of motor functions (97–99), induction of dystonia after *in situ* administration in the red nucleus (97), regulation of ileal function (100). The sites are also important in the blockade of tonic K<sup>+</sup> channels (101), potentiation of the neuronal response to *N*-methyl-D-aspartate [NMDA] in the CA<sub>3</sub> region of the rat dorsal hippocampus (102), or activation of a novel p53- and caspaseindependent apoptotic pathway. The mechanism of the induction of apoptosis is distinct from other apoptotic stimuli (103).

The  $\sigma_2 R$  is an  $\sigma R$  that preferentially binds to siramesine<sup>®</sup> (26), selective  $\sigma_2 R$  agonist and also PB28 (104). Activation of the  $\sigma_2 R$  causes apoptosis (104) via triggering of cancer selective cell death signaling (105) by multiple pathways (106). This finding is an important observation for potential antineoplastic drug development. The mechanism by which  $\sigma_2 R$  stimulation induces apoptosis may result from its

modulation of intracellular  $Ca^{2+}$  stores in some tumors (95). This is of particular importance in those tumors that induce hypercalcemia, e.g. some lymphomas.

The molecular nature of the  $\sigma_2 R$  is still to be fully characterized; however, a structure-affinity and comparative molecular field analysis of  $\sigma_2 R$  receptor ligands has been reported (107). A photo affinity labeling study, using DTG, revealed the existence of two protein bands of MW 25 000 and 21 500 (92). Because the  $\sigma_1 R$  has been cloned (6,77) and shown to be a protein of MW 25 300, it has been presumed that the  $\sigma_2 R$  gene encodes a protein of MW 21 500.

Despite efforts to define the gene for the  $\sigma_2 R$ , it remains unidentified. It has been suggested that the  $\sigma_2 R$  characteristics are, in fact, a consequence of  $\sigma_1$  gene alternative splicing (108). However, in the  $\sigma_1 R$  knockout mouse, although  $\sigma_1 R$ specific drug binding is significantly reduced, binding of nonspecific  $\sigma R$  drugs, such as DTG, is not affected, suggesting that the  $\sigma_2 R$  is unaffected (63).

Recently, a novel iodinated  $\sigma_2 R$  ligand (a conformationally-flexible benzamide derivative, 5-bromo-2,3-dimethoxy-N-[2-(6,7-dimethoxy-3,4-dihydro-1H-isoquinolin-2-yl)-butyl]benzamide, which has 1000-fold selectivity for  $\sigma_2 R$ ) has been evaluated as a cell proliferation marker (109).

 $\sigma_2 Rs$  have been implicated in a number of neoplasms, e.g. pancreatic carcinoma (110), urinary bladder tumors (111,112) and breast tumor cell lines (103); therefore, they have been primarily investigated for possible use as cancer chemotherapy targets (113). A more detailed discussion regarding the  $\sigma_2 Rs$  and neoplasia can be found later in the Section "Neoplasia".

Sigma<sub>3</sub> receptors [ $\sigma_3$ Rs] have been proposed (28,29) and were suggested to be linked to the conversion of tyrosine to dopamine [DA] and the activation of protein kinase C [PKC] (114). Here, the proposed  $\sigma_3$ R agonists may increase the rate of DA synthesis. In addition, putative  $\sigma_3$ Rs have been imaged in the mammalian brain, and appear to have histamine receptor [H<sub>1</sub>R] properties (115,116). Regardless of these findings, the molecular basis for this diversity is not clear, and the limited amount of literature regarding the subject questions whether the  $\sigma_3$ Rs really exist, or whether they are a subtype of  $\sigma_1$ Rs or  $\sigma_2$ Rs.

#### Mechanism of action

 $\sigma_1 Rs$  are intracellular receptors acting as chaperone proteins (46,117). Chaperone proteins assist in the correct folding of other proteins, either during their synthesis or function (118). More specifically,  $\sigma_1 Rs$  modulate Ca<sup>2+</sup> signaling through the inositol triphosphate [IP<sub>3</sub>] receptor. They dynamically translocate inside cells, hence are transmembrane proteins (118). In fact, it has been suggested that the  $\sigma_1 R$  receptor at the mitochondrial-associated endoplasmic reticulum membrane is responsible for mitochondrial metabolic regulation (119).  $\sigma_1 R$  also promotes mitochondrial energy depletion, Ca<sup>2+</sup> influx and apoptosis (120). The  $\sigma_1 R$  chaperone protein can be activated or deactivated by specific ligands (121).

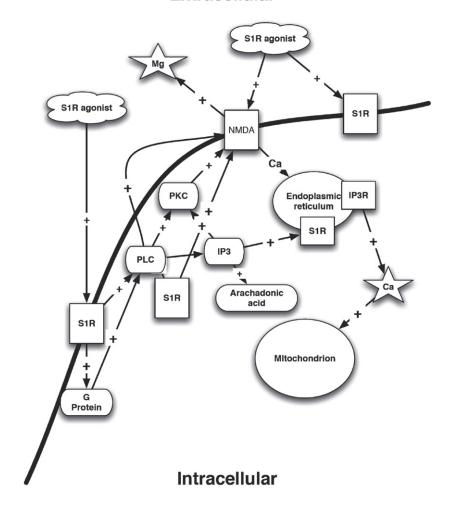
These  $\sigma_1 R$  chaperones act at the functional inositol triphosphate receptor [IP<sub>3</sub>R] to the ER and mitochondrion interface to ensure proper Ca<sup>2+</sup> signaling from ER into mitochondrion. However, under pathological conditions

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Figure 1. σRs and their effect on intracellular calcium concentrations.

PLC – phospholipases C; PKC – protein kinase C; S1R – sigma1; IP3 – inositol triphosphate; IP3R – inositol triphosphate receptor; NMDA – N-methyl-D-aspartate receptor; Mg – magnesium; Ca – calcium.

# Extracellular



where cells encounter excess stress that results in the ER losing its global Ca<sup>2+</sup> homeostasis, the  $\sigma_1 R$  translocates and counteracts the potential apoptosis. Thus, the  $\sigma_1 R$  is a receptor chaperone essential for the metabotropic receptor signaling and for the survival against cellular stress (46).  $\sigma_2 R$  is now thought to be a histone binding protein (111).

Although the precise mechanism of the biological response of  $\sigma Rs$  is still uncertain, it is accepted that  $\sigma R$  can modulate a number of neurotransmitter systems, including neurosteroids (49), glutamatergic [Glu] (56), noradrenergic [NA] (122) and dopaminergic [DAergic] ones (26,98) thought to be especially important functional modulators of Glu activity at this site (123–128).

Neurochemical and electrophysiological studies have been crucial in revealing that the  $\sigma$ Rs regulate the NMDA receptormediated glutamatergic, cholinergic and catecholaminergic neuronal responses (26,129,130).  $\sigma_1$ Rs, at least in part, are intracellular amplifiers creating a super sensitized state for signal transduction (82,131).

#### Signal transduction by σRs

The cloning of a one transmembrane domain  $\sigma_1 R$ , which does not correspond to a G-protein-coupled receptor, reactivated the debate over whether or not  $\sigma Rs$  act through

G-protein-dependent signaling cascades (132). Manipulation of G-proteins alters  $\sigma$ R-mediated effects on K<sup>+</sup> currents (133), acid sensing ion channels (134) and NMDA-evokes release of [<sup>3</sup>H]norepinephrine [NE] (135–137). Yet this manipulation has no effect on K<sup>+</sup> currents in other models, or on the NMDA response with other  $\sigma$ R ligands (138,139). Contrasting evidence exists for the effects of G-proteins on  $\sigma_1$ R ligand binding (140–143). Therefore, the data concerning the mechanism by which  $\sigma$ Rs act at the cell membrane level is often conflicting, if not controversial. Given the presumed heterogeneity of the  $\sigma_1$ R subgroup, it is likely that one subtype of the  $\sigma$ R interacts with G-proteins, while another subtype relies on G-protein-independent signal transduction mechanisms, probably via NMDAR.

## G-proteins

Studies on the modulation of ion channels by  $\sigma_1 Rs$  have made advances in deducing the nature of the signal transduction mechanism (144). It has been suggested, despite the lack of homology between the  $\sigma_1 R$  and classic G-protein-coupled receptors, that  $\sigma_1 Rs$  use G-proteins (74,133,145,146). Accordingly, the  $\sigma_1 R$  could interact functionally with G-proteins through a mechanism that differs from that of classical G-protein-coupled receptors (147). However, many physiological experiments suggest that  $\sigma_2 R$  signal transduction does not involve any G-protein. Experiments on rat neurohypophysis also produced negative results for secondary messenger or G-protein mediation of  $\sigma_1 R$  signaling (138). This finding may be a result of the dose response curve previously described.

In support of  $\sigma Rs'$  association with G-proteins, manipulating GTP and 5 guanylylimidodiphosphate [Gpp(NH)p] alters the binding of  $\sigma R$  some ligands (70,71,148,149). Contrasting results have also been found for the effects of G-proteins on  $\sigma_1 R$  ligand binding (142,143). Chronic treatments with haloperidol [Haladol<sup>®</sup>] in rats cause decrease responsiveness to guanine nucleotides following repeated exposure (72). Some selective  $\sigma R$  agonists stimulate GTPase activity (132).

The mechanisms of these  $\sigma R$  effects are not well understood, even though  $\sigma_1 Rs$  have been linked circumstantially to a wide variety of signal transduction pathways (150). Links between  $\sigma_1 Rs$  and G-proteins have been suggested, but there is also some evidence against this hypothesis (142). Regardless of their involvement of G-proteins, it is more likely that  $\sigma_1 Rs$  act through the NMDAR rather than through these G-proteins (138,139,151).

## Ion channels and cations

In support of the majority of effects of  $\sigma_1 R$  stimulation being mediated by the ionotropic glutamate receptors [iGluRs], such as the NMDAR, the  $\sigma_1 R$  has been shown to appear in a complex with voltage-gated K<sup>+</sup> channels, leading to the suggestion that these receptors are auxiliary subunits of the voltage-gated channels (88,138). For example, K<sup>+</sup> conductance is the prominent target of  $\sigma_1 R$  in rat cortical synaptosomes, C6 glioma cells (101), NCB-20 cells (152), rat neurohypophysis (139) and frog melanotropic cells (133,145).

*Calcium.* An interaction between  $\sigma Rs$  and  $Ca^{2+}$  channels is probable, as (+)-PTZ inhibits the rise in  $Ca^{2+}$  levels induced by depolarization of cell membranes and  $\sigma R$  ligands decrease basal intracellular  $Ca^{2+}$  concentration ( $[Ca^{2+}]_i$ ). This finding supports the hypothesis that the  $\sigma R$  activation alone affects  $[Ca^{2+}]_i$  (2,153,154) and that the  $\sigma_1 R$  is likely coupled to the nicotine-receptor-associated  $Ca^{2+}$  ionophore (155).

 $\sigma$ R-induced increases in Ca<sup>2+</sup> currents, which develop progressively following relatively long lasting applications of  $\sigma$ R ligands, suggest a direct intracellular coupling of  $\sigma$ R to Ca<sup>2+</sup> channels, through which  $\sigma$ R ligands can stimulate voltage-activated Ca<sup>2+</sup> conductance, independent of the K<sup>+</sup> channel pathway (156). It is possible that an atypical  $\sigma_1$ R subtype might also interfere with [Ca<sup>2+</sup>]<sub>i</sub> homoeostasis (153,154,157).

In rat sympathetic and parasympathetic neurons,  $\sigma Rs$  have been shown to modulate high-voltage-activated Ca<sup>2+</sup> channels including N-, L-, P/Q- and R-type Ca<sup>2+</sup> channels (158). Although  $\sigma_2 R$ -selective  $\sigma R$  ligands were not used, the rank order potency observed, which was haloperidol > ibogaine (an indole alkaloid (159)>(+)-PTZ>DTG, would suggest that this effect may be mediated by  $\sigma_2 Rs$ . In addition to reducing the peak amplitude of the Ca<sup>2+</sup> current,  $\sigma Rs$  altered the kinetic properties of these channels. Several lines of evidence have added further arguments for the involvement of  $\sigma_1 R$  in Ca<sup>2+</sup> signaling (160). Specifically, the  $\sigma_1 R$  ligands (+)-PTZ and PRE-084 modulate Ca<sup>2+</sup> signaling in NG108 cells via  $\sigma_1 Rs$  by two different modes of action. Firstly, intracellularly, perhaps on the ER,  $\sigma_1 R$ ligands potentiate bradykinin-induced increase in cytosolic free Ca<sup>2+</sup> in a biphasic manner, which can be blocked by  $\sigma_1 R$ antisense oligodeoxynucleotide (161), and a second mode of action at the plasma membrane (153,161).

However, the NMDA receptor is probably involved, as such an interaction explains the potentiating action of  $\sigma_1 R$ drugs on NMDA receptor-mediated responses (137,162,163). Further support for this notion is provided by the parallel between their effect on  $[Ca^{2+}]_i$  mobilization and on the neuronal response to NMDA (135,163,164). It is possible that the major physiological function of the  $\sigma_1 R$  in the CNS is to regulate both types of intracellular Ca<sup>2+</sup> equilibrium (165).

The changes reported above may cause the reported amplification of Glu, acetylcholine [ACh] and DA responses via the  $\sigma_2 R$  (82,157,164). For example, DTG decreases, whereas reduced haloperidol increases,  $[Ca^{2+}]_i$  mobilization in colon and mammary adenocarcinoma cells independently of any effect on  $Ca^{2+}$  entry through the plasma membrane (153,166). These observations suggest that the biological effect of  $\sigma_1 R$  drugs may be more complex in the regulation of the  $[Ca^{2+}]_i$  equilibrium; regardless, these results give support to the suggestion that  $\sigma_2 R$  also impacts  $[Ca^{2+}]_i$  homoeostasis (95,135,153,167).

It has been proposed that the modulation of  $Ca^{2+}$  signaling mediated by  $\sigma_1 Rs$  involves the formation of a multiprotein complex, or  $\sigma_1 Rs$  that form multiunit complexes responsible for the modulation of these ion channels (163,165). Specifically,  $\sigma_1 Rs$  have recently been found to anchor ankyrin, a cytoskeletal adaptor protein, to the ER membrane and modulate the function of ankyrin and IP<sub>3</sub> on the ER (82,164). In this model, the presence of the  $\sigma R$  agonist (+)-PTZ leads to the  $\sigma_1 R$ -ankyrin complex dissociating from the IP<sub>3</sub> (168). This dissociation leads to an increased binding of IP<sub>3</sub>, which in turn increases  $Ca^{2+}$  efflux. On the other hand, in the presence of the  $\sigma_1 R$  antagonist NE-100 (156), the  $\sigma_1 R$  dissociates from ankyrin, which remains coupled to IP<sub>3</sub> on the ER (164).

According to the heterogeneity of the  $\sigma R$  subtypes, it has been proposed that in the guinea-pig brain, which expresses mainly the  $\sigma_2 R$  protein, bivalent cations zinc  $[Zn^{2+}]$ , nickel  $[Ni^{2+}]$ , sodium  $[Na^+]$ , strontium  $[Sr^{2+}]$ , magnesium  $[Mg^{2+}]$ and  $Ca^{2+}$  inhibit  $[^{3}H]DTG$  binding in a monophasic manner within a micromolar concentration range (169). However,  $[^{3}H](+)$ -PTZ binds in a biphasic manner within an mM concentration range, thereby supporting a hypothesis of preferential involvement of the  $\sigma_2 R$  subtype as modulator of  $Ca^{2+}$  entry (170). Subsequent dissociation experiments performed with  $[^{3}H]DTG$  show that verapamil and amidirone, but not nifedipine, BAY-K8644 or amiloride, enhanced the dissociation of  $[^{3}H]DTG$  from  $\sigma R$ -binding sites further supporting the involvement of  $\sigma_2 R$  in the modulation of  $Ca^{2+}$  channels.

*Potassium.* K<sup>+</sup> conductance is the prominent target of  $\sigma_1 R$  in rat cortical synaptosomes, C6 glioma cells (101), NCB-20 cells (152) rat neurohypophysis (139), or frog melanotropic

cells (101,133,145,146). An observation has been made that there is interaction between  $\sigma Rs$  and  $K^+$  channels. Here the  $\sigma R$  ligands DTG and (+)-PTZ inhibit  $K^+$  currents (133,138,139).

The inhibition of K<sup>+</sup> channels by  $\sigma R$  agonists and antagonists in NCB-20 cells is not affected by pretreatment with A23187, forskolin, phorbol-12,13-dibutyrate, cholera toxin, or pertussis toxin has been shown (152). These results are consistent with the well-known intracellular secondary messenger systems not being essential for the modulation of voltage-gated K<sup>+</sup> channels by  $\sigma_1 R$ .

Further investigations of this modulation suggest that a protein-protein interaction is the likely mechanism of signal transduction by  $\sigma$ Rs, as  $\sigma$ R ligands do not interact directly with K<sup>+</sup> channels (88,138), although this effect is enhanced in the presence of  $\sigma$ R ligands (138). Therefore,  $\sigma$ Rs may serve as auxiliary subunits to voltage-gated K<sup>+</sup> channels in the plasma membrane (88), which also may involve other proteins such as ankyrin and IP<sub>3</sub>R.

Studies on  $\sigma_1 R$  modulation of K<sup>+</sup> channels, to date, have led to the conclusion that the signal transduction mechanism of  $\sigma_1 Rs$  is membrane independent of G-protein coupling and protein phosphorylation (158) reconstructable in a heterologous system, not requiring cytoplasmic factors, and necessitating the  $\sigma_1 R$  and the K<sup>+</sup> channel to be in close proximity (138), probably to form a stable macro-molecular complex (88).

Additional studies are required to determine whether the  $\sigma_1 R$  modulation of  $K^+$  channels is through a direct proteinprotein interaction or through intermediate signaling molecules. Given the wide variety of functions that the  $\sigma_1 Rs$  are reported to serve, the most likely explanation is a  $\sigma_1 R$ signaling mechanism involving one or more intermediate signaling molecules, which are localized at or in the plasma membrane, rather than a direct interaction.

## $\sigma_I R$ as an intracellular amplifier

Acute activation of the  $\sigma_1 R$  results in a direct modulation of  $([Ca^{2+}]_i)$  mobilization (161,163), and prevents intracellular  $Ca^{2+}$  dysregulation in neurons follow an ischemic event. After depletion of intracellular  $Ca^{2+}$  from ER stores, the depolarization-induced increase in  $[Ca^{2+}]_i$  in the cells is modulated by  $\sigma_1 R$  agonists. Both effects are blocked by an antisense oligodeoxynucleotide targeting the  $\sigma_1 R$  (161). Therefore, activation of the  $\sigma_1 R$  results in a complex, bipolar modulation of  $Ca^{2+}$  homeostasis.

At the ER level, the  $\sigma_1 R$  activation facilitates the mobilization of IP<sub>3</sub>R-gated intracellular Ca<sup>2+</sup> pools. This change also occurs at the plasma membrane level. A co-immunoprecipitation study further revealed that the  $\sigma_1 R$  could regulate the coupling of the IP<sub>3</sub>R with the cytoskeleton via an ankyrin B anchor protein, a cytoskeletal protein originally attached to ER membranes (164).

As stated previously, activation of the  $\sigma_1 R$  dissociates ankyrin B from IP<sub>3</sub>R in NG-108 cells, and this dissociation correlate with the efficacy of each ligand in potentiating the Ca<sup>2+</sup> efflux induced by bradykinin. These results, in conjunction with the  $\sigma_1 R$  subcellular localization (171,165), show that the  $\sigma_1 R$  might act as a sensor or modulator for the neuronal intracellular  $Ca^{2+}$  mobilizations and consecutively for extracellular  $Ca^{2+}$  influx.

Stimulation of the  $\sigma_1 R$  results in its translocation from the ER (64,163,164), via lipid droplets, to plasma membranes when stimulated by agonists (65,172,173). Thus the translocation of  $\sigma_1 Rs$  at the plasma membrane, associated with the ankyrin B protein consequently affects Ca<sup>2+</sup> mobilization at the ER (174).

Lipid droplets are formed by coalescence of neutral lipids within the ER membrane bilayer when the coalesced lipids reach a critical size they bud off to form cytosolic lipid droplets, serving as a new transport pathway of lipids between the ER and Golgi apparatus or plasma membrane (65,172,173). Therefore,  $\sigma_1 R$  on the ER may play a role in the compartmentalization of lipids into the ER lipid storage sites and in the export of lipids to peripheries of cells (64).

Lipid rafts play a role in a variety of cellular functions including vesicle transport, receptor clustering and internalization, and coupling of receptors with proteins involved signal transduction (175). Over-expression of functional  $\sigma_1 R$ increases cholesterol contents and alters glycosphingolipid components in lipid rafts of NG108 or PC-12 cells (65,176,177), suggesting that up-regulation of  $\sigma_1 Rs$  potentiates lipid raft formation. Since glycosylated moieties of gangliosides have been proposed to play a role in regulating the localization of growth factor receptors in lipid rafts (175), chronic activation of  $\sigma_1 R$  may present substantial consequences in cell viability and differentiation.

#### Potential endogenous ligand

It has been demonstrated that alterations in endogenous hormonal levels, via adrenalectomy [ADX], castration [CX] (178), ovariectomy [OVX], or pregnancy, affect  $\sigma R$  ligands activity when these have been evaluated in the electrophysiological model of the modulation of the NMDA response in the hippocampus (179,180). Similar findings have been seen when investigating the "antidepressant-like" effects of  $\sigma R$  ligands in behavioral models of depression (181). Moreover, radioligand binding studies show a 30–40% decrease in [<sup>3</sup>H]SKF-10,047 binding during pregnancy, while ADX/CX enhances [<sup>3</sup>H]SKF-10,047 binding. Subsequent treatment with finasteride, which increases progesterone [PROG] levels, produces decreased [<sup>3</sup>H]SKF-10,047 binding (178,182–184).

Steroid hormones had been original proposed as endogenous ligands of  $\sigma_1 Rs$ , and more recently DMT, a natural tryptamine alkaloid, has been defined as the  $\sigma_1 Rs$  endogenous ligand (7,8). DMT is a hallucinogen found endogenously in human brain. It is commonly recognized to target the 5-hydroxytryptamine 2A receptor [5HT<sub>2A</sub>R] or the trace amine-associated receptor to exert its psychedelic effect. DMT has been recently shown to bind the  $\sigma_1 R$  molecular chaperones, whose function includes inhibiting various voltage-sensitive ion channels (9). Thus, it is possible that the psychedelic action of DMT might be mediated in part through  $\sigma_1 Rs$ .

#### Cell development and plasticity

 $\sigma R$  drugs and neurosteroids, acting at the level of the  $\sigma_1 R$  protein, may act in cell development and cell trophic actions

(82,185). For example, they have been shown to suppress multiple aspects of microglial activation (186), probably increasing intracellular Ca<sup>2+</sup>. These morphological changes have been previously ascribed to the prominent role of Ca<sup>2+</sup> in cellular plasticity. This plasticity, which is associated both with the same prerequisite enhancement of NMDA-mediated glutamatergic neurotransmission and protein dephosphorylation that occur downstream from the massive entry of Ca<sup>2+</sup> into the cell cytoplasm, as well as  $[Ca^{2+}]_i$  mobilization from the ER and the mitochondria. These events often occur synergistically (187,188).

The amplitude and reliability of both induction and maintenance of long-term potentiation [LTP] in neurons represent an effective model for memory acquisition and consolidation (189). The blockade of LTP and of several learning processes in mice, including spatial learning or passive avoidance, by  $Ca^{2+}$  depletion further supports the notion that  $Ca^{2+}$  influx and  $Ca^{2+}$  compartments are mandatory for memory (187,188). Additional evidence is provided by the poor capacity for acquisition and storage of spatial memory, combined with the lack of hippocampal LTP in transgenic strains of mice lacking subtypes of ryanodine receptors and IP<sub>3</sub> kinase. This receptor-mediated postsynaptic  $Ca^{2+}$  accumulation ( $Ca^{2+}$  influx plus massive  $Ca^{2+}$  release from internal stores) is reinforced by subsequent activation of kinases such as  $Ca^{2+}/calmodulin-dependent protein kinase II$ 

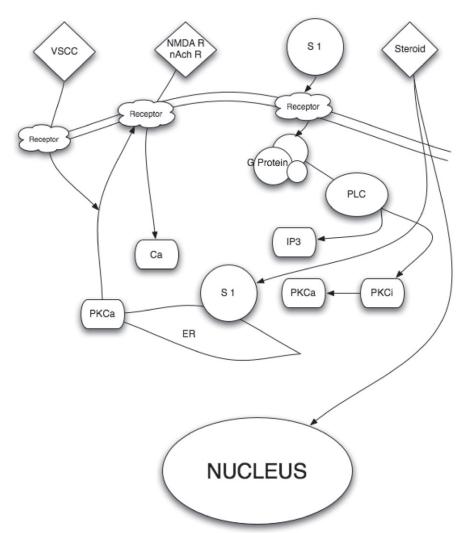
[CaMKII] and PKC (17). Thus,  $\sigma_1 R$  are probably involved in LTP via altering Ca^{2+} influx.

The initial statement that drugs acting via  $\sigma$ Rs may affect the regulation of  $[Ca^{2+}]_i$  equilibrium and likely the  $Ca^{2+}$  entry through the plasma membrane emerged from *in vitro* binding studies (163,190). The binding studies showed that inorganic Ca<sup>2+</sup> channel blockers, such as cadmium  $[Cd^{2+}]$ , nickel  $[Ni^{2+}]$  and Ca<sup>2+</sup>, and the nonselective Na<sup>+</sup> and Ca<sup>2+</sup> channel blockers phenylamine, cinnarizine, amidirone and amiloride, reduced the labeling of  $[^{3}H]$ dextromethorphan (191) and  $[^{3}H]$ DTG to  $\sigma$ R sites (192,193) (Figure 2).

An interesting feature of  $\sigma$ Rs is that they do not follow the classical pharmacology of a more or less linear dose-response curve followed by a plateau effect. A biphasic bell-shaped dose response curve has been observed for  $\sigma$ R ligands in various behavioral, biochemical and electrophysiological paradigms (135,161,182,194). For example, because of the bell-shaped dose response curves, in the electrophysiological paradigm of the modulation of the NMDA response, low doses of  $\sigma$ R agonists induce a potentiation of the NMDA response (162,195). At higher doses, the effects of  $\sigma$ R agonists such as DTG and JO-1784 progressively decrease and disappear and these molecules act as antagonists by preventing the potentiation induced by low doses of other  $\sigma$ R agonists (194).

Figure 2. Putative biological action of the  $\sigma_1 R$  on neuronal function.

PLC - phospholipases C; PKCa - protein kinase C alpha; PKCi - protein kinase C inhibitor; S 1 - sigma1 receptor; IP3 - inositol triphosphate; nAch - nicotinic acetylcholine; nAchR - nicotinic acetylcholine receptor; NMDAR - N-methyl-d-aspartate receptors; Ca - calcium: VSCC - voltage-sensitive calcium channels. Once a neuron has been activated, e.g. via Glu or acetylcholine, a concomitant influx of  $Ca^{2+}$  and  $[Ca^{2+}]_i$ mobilization occur, facilitated by the activation of the endoplasmic-reticulum-bound  $\sigma_1 R$ , which is also triggered by numerous xenobiotics and steroids. The subsequent activation of PLC and the recruitment of the PKCs from its inactive form  $[PKC_i]$  to its active form [PKCa], which is translocated to the plasma membrane, result in the activation of various enzymatic processes, as well as the phosphorylation of membrane-bound neurotransmitter receptors. In turn, the  $\sigma_1 R$ translocates to the plasma membrane where it decreases the excitatory neurotransmitterinduced Ca<sup>2+</sup> influx.



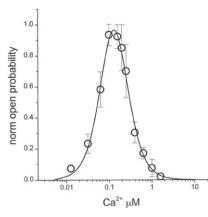


Figure 3. Bell curve dose response. Bell-shaped Ca<sup>2+</sup> dependence of recombinant IP<sub>3</sub>R. Recombinant IP<sub>3</sub>R activity was measured in bilayers in the presence of  $2 \,\mu$ M InsP<sub>3</sub> and 1 mM Na<sub>2</sub>ATP at cis (cytosolic) Ca<sup>2+</sup> concentrations in the range between 10 nM and 5  $\mu$ M Ca<sup>2+</sup>. Ca<sup>2+</sup> concentration in the cis chamber was adjusted by using calibrated 20 mM CaCl<sub>2</sub> stock solution and 1 mM mixture of HEDTA and EGTA. Po in each experiment was normalized to maximum Po observed in the same experiment, and then data from three independent experiments were averaged together at each Ca<sup>2+</sup> concentration ( $\circ$ ) (477).

A similar shaped dose response curve has also been described with  $\sigma R$  ligands in other models such as in release experiments (135) and in behavioral models (182,183). The exact reason for such dose response curves obtained in so many models have not been well established. It has been proposed that they may be due to the fact that low doses of  $\sigma R$ ligands activate one subtype of  $\sigma Rs$  for which they have high affinity, whereas higher doses may activate another subtype(s) of the  $\sigma R$  for which they have a lower affinity. Such activity would counteract the effects observed at lower doses (194,196,197). Nonetheless, it is important to note that the different, and sometimes opposite, results obtained with low and high doses of  $\sigma R$  ligands could constitute a very important factor to explain much of the controversy seen in the literature regarding  $\sigma Rs$  (Figure 3). The importance of the curves seen in these and other experiments will be discussed further on.

Throughout adulthood, differences in the motor changes elicited by drugs affecting  $\sigma Rs$  are correlated with the number of receptors in the P<sub>2</sub>, and not the P<sub>3</sub>, cellular fraction (198), which supports the hypothesis that translocation of the  $\sigma_1 R$  from the ER to the cell membrane occurs (190). This change decreases with age in motor structures as has been observed in the aged monkey brain where an increase of  $\sigma_1 Rs$ has been found (199).

 $\sigma R$  agonists enhance memory performance in young rodents and in rodent models of cognitive impairment (200–205). For this reason, it has been suggested that agerelated memory deficits may be responsive to up regulation of the  $\sigma Rs$ , implying that  $\sigma_1 R$  agonists may have therapeutic potential in dementia (204). In fact, such ability to alleviate memory deficits during aging has also been confirmed in humans for the selective  $\sigma_1 R$  agonist Igmesine<sup>®</sup> [(+)-*N*cyclopropylmethyl-*N*-methyl-1,4-diphenyl-1-ethyl-but-3-en-1-ylamine hydrochloride], which appears more efficient among the elderly (206).

Conversely, the  $\sigma_2 R$  subtype exhibits no stereo selectivity and only low affinities for the (+)-BZM (91). It does not appear to be modulated by pertussis toxin-sensitive  $G_{i/o}$  proteins (207), and is predominantly located in the motor system and periphery (21). Clinically, the  $\sigma_2 R$  subtype may be preferentially involved in the motor and anxiolytic effects of  $\sigma R$  ligands, as well as in diseases affecting motor and postural control (208). Interestingly, brainstem motor function, which is profoundly sensitive to  $\sigma R$  drugs, decreases with age, during which the accuracy and consistency of fine and complex motor performance decrease (208).

The modulatory role of neurosteroids on neuronal function is typified by dihydroepiandrosterone (sulfate) [DHEA(S)] and its effect on  $\sigma$ Rs (209). NE release induced by NMDA via the stimulation of the  $\sigma$ R is significantly enhanced by the addition of DHEAS (210). These findings have been replicated (123,183,210) and the overall data are consistent with the activity of DHEAS as a  $\sigma_1$ R agonist; hence, neurosteroids potentiate NMDA-induced neuronal excitability (180).

It now appears as though DHEA(S) has an ability to modulate neurotransmitter receptors in the CNS that are primarily involved in learning and memory (209).  $\sigma R$ agonists (205) enhance memory performance in young rodents and in rodent models of cognitive impairment (183,200,203,211,212), probably via the NMDAR which is involved in the development of LTP (213–216), an essential element of neural plasticity.

#### Activity through neurotransmitters

Neurotransmitters rarely act alone. The delicate balance of the major neurotransmitters, receptors and other methods of transmission control are central to normal homeostasis. These interactions make a reductionist approach to determining the effect of one specific neurotransmitter difficult (217). In fact, experiments that address only one major neurotransmitter may be misleading due to the lack of evaluation of other neurotransmitters and associated receptors.

As  $\sigma Rs$  are central to a number of CNS and other actions, it is not surprising that they interact with many other concurrent events within and outside the cell membranes on cells of many types. A functional interaction between  $\sigma R$  ligands and neurosteroids, such as PROG (218), GluR and opioids, DA and 5-HT exists (98,126–128,183,194,195,219–222).

## Neuroactive steroids (neurosteroids)

Neurosteroids (223–225), such as PROG, pregnenolone [PREG], dihydroepiandrosterone [DHEA(S)] and their respective sulfate esters PREGS or DHEAS, are involved in regulating the imbalance between excitation and inhibition in the CNS (226); hence, they have been suggested as a treatment for anxiety (227).

The initial proposition that steroids behave like endogenous  $\sigma_1 R$  ligands emerged from binding studies (222) and pharmacological experiments (210) leading to the hypothesis that neurosteroids may constitute endogenous ligands for the  $\sigma_1 R$  (2). A functional interaction between  $\sigma R$  ligands and neurosteroids, such as PROG (218), GluRs and neurotransmitters exists (Figure 4) (98,194,195,220). Early studies found that neurosteroids bind to  $\sigma_1 R$  (183,228–230), but not to  $\sigma_2 R$  (231). For example, the neurosteroids PROG and DHEA(S) dose-dependently inhibit the *in vivo* binding of [<sup>3</sup>H]-SKF-10,047, an  $\sigma$ R agonist, PROG being the most potent (228, 230). These binding data led to the hypothesis that PROG might be the endogenous ligand for  $\sigma_1$ Rs, which is controversial, as the affinity of PROG for  $\sigma_1$ R does not appear very high for an endogenous ligand (232). DMT, a natural tryptamine alkaloid, is now recognized as the  $\sigma_1$ Rs endogenous ligand (8).

The non-neuronal physiological actions of the neurosteroids, demonstrated from embryogenesis through adult life, are mediated secondarily by steroid receptors translocating into the nucleus, and non-genomic neuromodulatory actions affecting directly several ion channels, neurotransmitter receptors and second messenger systems (32). Neurosteroids activate transcription factors; hence, they regulate gene expression and stimulate protein synthesis (233–238). Only the human  $\sigma_1 R$  gene contains a steroid-binding component (239). These neurosteroids are found in the cortex, hippocampus and brainstem, areas of the brain containing high densities of  $\sigma_1 R$  (98).

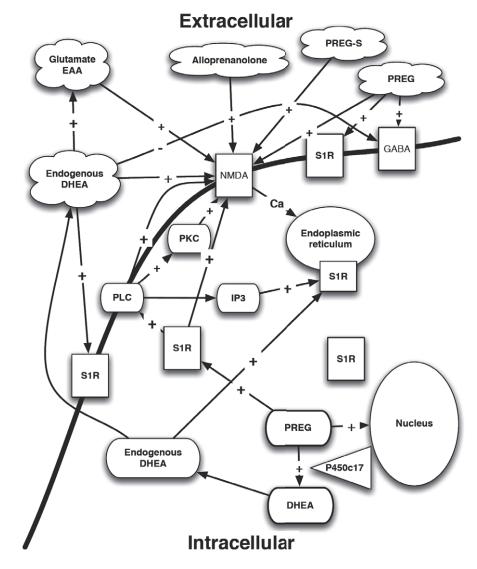
The neurosteroids  $3\alpha$ -hydroxy- $5\alpha$ -pregnan-20-one (allopregnanolone) [ALLO], allotetrahydrodeoxy-corticosterone, PREGS and DHEAS possess anti-stress, anxiolytic and

Figure 4. Neurosteroids and their interactions with  $\sigma Rs$ .

PLC – phospholipases C; PKC – protein kinase C; PKCi – protein kinase C inhibitor; S1R – sigma1 receptor; IP3 – inositol triphosphate; EAA – excitotoxic amino acid; GABA –  $\gamma$ -aminobutyric acid; NMDA – N-methyl-D-aspartate receptor; Ca – calcium; DHEA – dihydroepiandrosterone; PREG – pregnenolone; PREG-S – pregnenolone sulfate ester; P450c17 – cytochrome P450 C17. antiamnesic properties in experimental animal models (212,240–246), and have a possible neuroprotective effect in AD (247). In AD, decreased levels of PREG(S), DHEA(S) and PROG have been identified in the hippocampus (248), cortex and cerebellum, compared to the control animals (249–251). Their actions are mediated via the  $\sigma_1 R$  (Figure 4).

DHEAS and PREGS may also play an important role in depression (252), as decreased levels of DHEA, DHEAS and PREGS have been associated with clinical depression (253), cognitive dysfunction (254,255), dementia (253,256,257) and other neurological conditions (190,258–260). Although there is still controversy as to whether and how the steroidogenic enzymes are involved in the physiology of nervous system (261) and the pathophysiology of neuro-psychiatric disorders (262),  $\sigma$ Rs are critical to their cellular effects.

Clinical investigations in humans have produced evidence for an involvement of neurosteroids in conditions such as fatigue during pregnancy, premenstrual syndrome, postpartum depression, catamenial epilepsy, depressive disorders (252). They possibly alter the expression of conditioned fear stress response in mice (263). However, the exact mechanism underlying the beneficial effects of neurosteroids is not yet fully elucidated (82,263–267).



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Modulation of GABA<sub>A</sub>, NMDA, nicotinic, muscarinic, serotonin [5-HT<sub>3</sub>], kainate [Ka], glycine [Gly] and  $\sigma$ Rs, plus neuroprotection and induction of neurite outgrowth, dendritic spine development, and synaptogenesis are properties of specific neurosteroids (268,269). However, only the human  $\sigma_1$ R gene contains a steroid-binding component (239), which exerts effects on the genome via individual intracellular steroid receptors (270). Neurosteroids rapidly alter neuronal excitability through interaction with neurotransmitter-gated ion channels, e.g. NMDA.

The  $3\alpha$ -hydroxy ring A-reduced pregnane steroids, ALLO and tetrahydrodeoxycorticosterone, enhance  $\gamma$ -aminobutyric acid [GABA]-mediated chloride [Cl<sup>-</sup>] currents, whereas PREG sulfate and DHEAS display functional antagonistic properties at GABA<sub>A</sub>Rs (271–275). At physiologically relevant concentrations, that is, below 100 nM, these steroids directly activated the GABA<sub>A</sub>R–channel complex (276,277) and exerted a GABAmimetic effect sufficient to suppress excitatory neurotransmission (277).

Certain steroids, including PREG, DHEA, PROG, ALLO and their S (sulfate) esters, rapidly affect neuronal excitability through the modulation of voltage-gated ion channels (278), e.g. voltage-sensitive Ca<sup>2+</sup> channels [VSCC]s (226,279–283), and neurotransmitter-gated ion channels, such as at the NMDAR level (210,226,284–288). These steroids act at specific extracellular sites that are distinct from one another and from the spermine, redox, Gly Mg<sup>2+</sup>, phencyclidine [PCP] and arachidonic acid sites (289,290). In addition, DHEA(S), but not PREG(S), potentiates the NMDA-evoked catecholaminergic release (210) and firing activity of CA<sub>3</sub> hippocampal neurons (123). Moreover, the NMDA-stimulates NE release is inhibited by PREGS (210).

It remains unclear whether  $\sigma_1 R$  and neurosteroids exert a common action via the regulation of Ca<sup>2+</sup> influx and  $[Ca^{2+}]_i$  regarding amnesic and age-dependent cognitive abilities (163,291). In humans, plasma levels of DHEAS decline with age (258,259), PREG and PREG(S), DHEA and DHEA(S), or PROG decrease in aged mice (292–295) and PREGS decreases in aged Sprague Dawley rats correlating

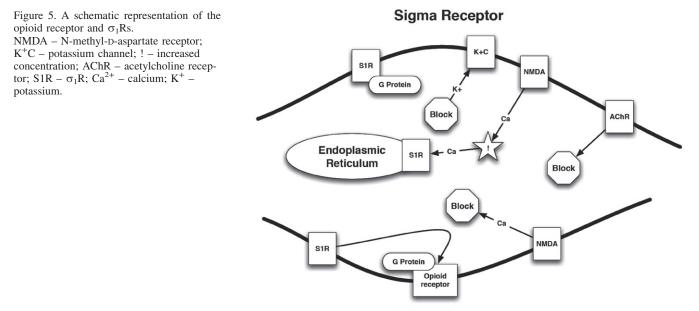
with impaired memory functions (260). However, attenuating effects of DHEAS and PREGS on the conditioned fear stress response are mediated via  $\sigma_1 Rs$  and that PROG has a  $\sigma_1 R$  antagonistic property (263).

It should be noted that  $\sigma R$  sites are different from high affinity PCP binding sites, located within the ion channel associated with NMDAR (21). PCP receptor is dependent on the presence of L-Glu; this has led to the suggestion that there may exist an NMDA/PCP receptor complex (296). The lack of selectivity between the  $\sigma_1 Rs$  and PCP binding sites seen following exposure to several compounds, including BZM or PCP derivatives, has led to confusion that was eventually clarified by the availability of new highly selective drugs (212). These compounds are now reference compounds in terms of selectivity between  $\sigma R$  and PCPRs.

## Opioids

Opioids have subtle differences in binding to the  $\mu$ ,  $\kappa$  and  $\sigma$ Rs; however,  $\sigma$ Rs are a receptor class in their own right (297,298). Although  $\sigma$ Rs now have been classified as a separate group of receptors from the opioid receptors, the outcome of  $\sigma$ R binding is not necessarily different from that when these receptors are bound to opiates (Figure 5) (299), especially since interactions between  $\kappa$ Rs and  $\sigma$ Rs have been reported (278,300).

 $\sigma_1$ Rs have been implicated in the modulation of opioid analgesia. It has been shown that coadministration of a  $\sigma_1$ R agonist decrease the analgesic power of morphine, whereas the use of an antagonist, DEX, increase analgesia (301), thus illustrating the pharmacological importance of  $\sigma_1$ R in the brainstem modulation of opioid analgesia (302). Interestingly, the dysphoric and psychomimetic side effects of  $\sigma$ Rs reside in the levorotatory (*L*) or (–) and not in the dextrorotatory (*D*) or (+)-isomer (303) as exemplified by nalorphine, levallorphan, (–)-PTZ, (–)-3-hydroxy-N-propargylmorphinan and MR 2034. All *L* opiates, produce dysphoria and psychomimetic effects, whereas the *D* isomers of PTZ and MR 2034 do not. Despite this selective response, both (+) and (–) PTZ



**Opioid Receptor** 

improve memory via the  $\sigma Rs$  rather than via the  $\mu$  and  $\kappa$  opioid receptors *per se* (304–306).

#### Serotonin [5-HT]

There is controversial evidence regarding possible interactions between  $\sigma R$  and the 5-HT system (Figure 6). The distribution of 5-HT binding sites in the CNS has been well described (307). These sites include  $\sigma_1 Rs$ . 5-HT and tryptophan (308) play a key role in depression and the mechanism of action of many antidepressants (88,309), probably via a decrease in the firing activity of 5-HT neurons (310–313).

Peripheral 5-HT- $\sigma$ R interactions have been proposed, as DTG, haloperidol and BMY-14802 inhibit the 5-HT evoked contractions of the guinea pig ileum longitudinal muscle and myenteric plexus preparations, showing high correlation with their potency to compete with DTG binding (314). However, the  $\sigma_1$ R agonist ligand EMD 57445 does not affect 5-HT-related parameters such as 8-OH-DPAT induced behavioral syndrome, m-chlorophenylpiperazine-induced hypothermia or L-5-hydroxytryptophaninduced head twitches (130). In addition, EMD 57445 and the  $\sigma_1$ R ligand PD144418 do not induce any change in 5-HT or 5-hydroxyindoleacetic acid [5-HI<sub>AA</sub>] levels in various brain regions, suggesting that these ligands exert no effect on 5-HTR populations or 5-HT metabolism (130,264). Interestingly, EMD 57445 and PhmD 144415 have been suggested to be  $\sigma_1$ R antagonists.

Whether or not  $\sigma R$  ligands can modulate 5-HT neuronal activity *in vivo*, the effects of short- and long-term administration of various  $\sigma R$  ligands on 5-HT basal neuronal activity in the dorsal raphe nucleus [DRN], have shown that acute treatments with SSRIs and MAOIs induce a decrease in the firing activity of DRN 5-HT neurons (310,312,313,315). There is an eventual restoration of the firing activity of these neurons (312,313,316,317) due to the desensitization of the 5-HT<sub>1A</sub> autoreceptors in the CNS (311,318–321).

In contrast to what has been observed in the dorsal hippocampus, acute iv administration of (+)-PTZ has no effect in the DRN. Interestingly, however, the  $\sigma R$  ligands,

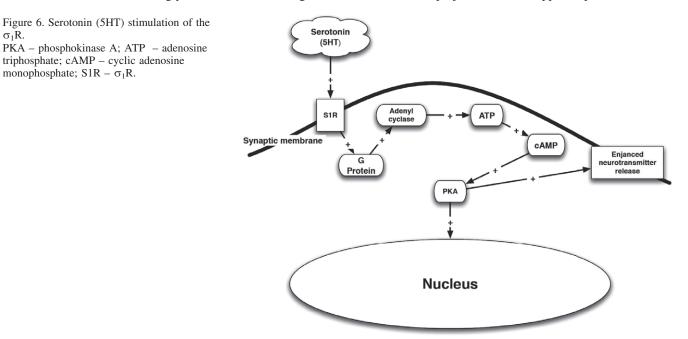
4-IBP, (+)-PTZ and DTG, after either two or 21 days of treatment induce a 50% increase in the firing activity of 5-HT neurons of the DRN (322). These findings suggest modulation of 5-HT neurotransmission by  $\sigma R$  ligands *in vivo*, a novel finding with respect to  $\sigma R$  research, again supporting a role for  $\sigma R$  in depression, probably mediated by  $\sigma_1 Rs$  via the 5-HT<sub>1A</sub> receptor (323–325).

Interestingly, OPC-14523, a  $\sigma_1 R$  agonist, decreases the responsiveness of the 5-HT<sub>1A</sub>R after two days of treatment (326,327). This is particularly significant given that classical antidepressant medications require chronic treatment for this decreased receptor response to occur (313,319,320). If this effect is shown to be a general effect, present with other  $\sigma_1 R$  agonists, the rapid desensitization of the 5-HT<sub>1A</sub> autoreceptor, in addition to the observed rapid increase in the firing activity of 5-HT neurons after only two days of treatment with  $\sigma_1 R$  agonists, would constitute another argument to suggest that  $\sigma R$  agonists have potential to produce a fast onset of antidepressant effect.

The neurosteroid  $\sigma_1 R$  agonist PROG does not have any effect by itself on 5-HT neuronal activity in the DRN, but several of its metabolites, such as ALLO or DHEA, increase the firing activity of DRN 5-HT neurons. Interestingly, at least part of the effects of neurosteroids is mediated through an activation of  $\sigma Rs$  as they are reversed by NE-100 (328).

The precise mechanism by which  $\sigma R$  ligands increase the firing activity of DRN 5-HT neurons has not been established. One possibility is that the effect is mediated locally, in the DRN, as a consequence of the modulation of the Glu neurotransmission, since AMPA and NMDA GluRs have been shown to mediate glutamatergic excitatory input in the DRN (329).

The  $\sigma_1$ R-mediated effect on firing could also be an indirect one, as  $\sigma R$  ligands rapidly modulate NMDAR-mediated transmission in the hippocampus, which leads to a modulation of 5-HT neurotransmission in the DRN via feedback loops to DRN 5-HT neurons. In fact, an afferent connection has been identified that projects from the hippocampus to the DRN via



the lateral habenula (330–336), and the long feedback loop that projects from the DRN to the prefrontal cortex [PFC] and back to the DRN (329,333,335,337–340).

Therefore, the activity of  $\sigma Rs$  on the DRN neurons is dependent on the balance between the excitatory input (the Glu system) from various brain regions (e.g. lateral habenula and mPFC) and inhibitory input from GABAergic interneurons in distal areas (e.g. periaqueductal gray area) and local GABAergic interneurons situated in the DRN (341,336).

Another factor likely contributing to the requirement of a sustained treatment of  $\sigma R$  agonists to observe an antidepressant effect is based on the density of  $\sigma Rs$  at the plasma membrane, which is progressively altered by the presence of  $\sigma R$  ligands.  $\sigma R$  agonists induce an increase in the  $\sigma R$  density at the plasma membrane following a minimum of two days of treatment (185) exerting effects on NMDAR-mediated signaling.

## Dopamine

The  $\sigma_1 R$  subtype is involved in the facilitation of cortical Dopamine [DA] transmission in the rat brain (342).  $\sigma_1 Rs$  are located in limbic areas, including nucleus accumbens [NAC] (343) and PFC, both of which are thought to be involved in schizophrenia (344). Many antipsychotics, including haloperidol (345), bind with high affinity to  $\sigma_1 Rs$ , where the DAergic hyperactivity in the NAC is thought to underlie positive symptoms of schizophrenia (including delusions, disordered thoughts and speech, and tactile, auditory, visual, olfactory and gustatory hallucinations, typically regarded as manifestations of psychosis), while DAergic hypoactivity in PFC the negative symptoms (including deficits of normal emotional responses or of other thought processes).  $\sigma_1 R$ ligand agonists increase extracellular DA levels in rats (346) whereas their antagonism inhibits DA-induced abnormal involuntary movements (347).

 $\sigma$ Rs regulate NMDA-[<sup>3</sup>H]DA release in caudate-putamen [CP], the neuroanatomical substrate for extrapyramidal side effects resulting from chronic 2-amino-7-phosphonoheptanoic acid [AP-7] treatment (348). In that study, in the NAC, regulation of DA release by the prototypical  $\sigma$ R agonist (+)PTZ mediated predominantly by the  $\sigma_1$ R, whereas in the PFC a portion of the (+)PTZ effect is likely mediated by the  $\sigma_2$ R.

In both the NAC and PFC, regulation of DA release by the  $\sigma R$  agonist BD737 is mediated primarily by the  $\sigma_1 R$ , not via the opioid receptors, the NMDAR-operated cation channel, or by  $\sigma R$  effects upon [<sup>3</sup>H]DA accumulated by noradrenergic terminals in PFC (349). In fact, the action of NMDA in primary cortical neurons is regulated differently by ligands with differential affinities at DA D<sub>2</sub> and  $\sigma Rs$  (291).

The effects of different selective  $\sigma R$  ligands on DA and Glu-NMDA neurotransmissions, both in origin (A10 and A9 areas) and terminal NAC and CP regions of the rat mesolimbic and nigrostriatal DA-ergic systems, have been evaluated. The selective  $\sigma_1 R$  ligands 2-[4-(4-methoxy-ben-zyl)piperazin-1-yl-methyl]4-oxo[4H]-benzo-thiazolin-2-one [S-21377] and 2[(4-benzyl piperazin-1-yl) methyl] naphthalene, dichlorydrate [S-21378] slightly increase the spontaneous firing rate and potentiate the NMDA-induced neuronal

activation of DA-ergic neurons in the A9 and A10 regions. (+)N-cyclopropylmethyl-N-methyl-1,4-diphenyl-1-ethylbutyl-2-N [JO-1784], another selective  $\sigma_1 R$  ligand, has produced no or little effect in these areas (350).

A selective  $\sigma_2 R$  ligand 1,4-bis-spiro[isobenzofuran-1(3H), 4'-piperidin-1'yl]butane [Lu 29–252] significantly potentiates the NMDA-induced increase in firing activity of A<sub>10</sub> DA neurons. Functional interaction between  $\sigma_2 R$  and NMDARs in the A<sub>10</sub> region has been reported (350); thus, DA release in the striatum may be modulated by multiple  $\sigma R$  subtypes. In such a situation, NMDARs may mediate the stimulatory effect of  $\sigma R$  ligands on DA release in the striatum (351).

In addition,  $\sigma R$  may regulate the release of DA along with an action at the NMDAR, e.g. the pharmacological effects of amantadine on DAergic transmission are proposed to result from an uncompetitive antagonism at this receptor (352). These data demonstrate that aminoadamantanes behave as  $\sigma_1 R$  agonists, and confirm an involvement of this receptor in modulating DA receptors exerted by therapeutically relevant concentrations of amantadine (352,353).

The regulation of DA release is much more complicated than has been alluded above. Regardless, work has showed that activation of  $\sigma_2 R$  results in the regulation of dopamine transporter [DAT] activity via a Ca<sup>2+</sup>- and PKC-dependent signaling mechanism (354).

#### Nicotine and acetyl choline [ACh]

 $\sigma_1 R$  ligands noncompetitively inhibit nicotine-stimulated catecholamine release from bovine adrenal chromaffin cells in a concentration-dependent and reversible manner (355). The rank order of potency of ligands to inhibit nicotine stimulated catecholamine release is correlated with that observed in radioligand binding assays selective for the  $\sigma_1 R$ subtype. This naltrexone-insensitive effect is paralleled by an inhibition of nicotine-stimulated increases in  $[Ca^{2+}]_i$ .  $\sigma R$ ligands are without effect on catecholamine release or  $[Ca^{2+}]_i$ in the absence of nicotine (155), although the inhibitory effect of  $\sigma R$  ligands on the nicotine-evoked  $Ca^{2+}$  uptake is not directly coupled with either the  $\sigma_1 R$  or  $\sigma_2 R$  sites (356).

Nicotine accelerates the association of the receptor selective radioligand,  $[{}^{3}H](+)PTZ$ , to adrenal medullary homogenates while having no effect on the rate of ligand dissociation, consistent with a  $\sigma R$  ligand binding site closely associated with and allosterically modulated by the nicotinic acetylcholine receptor [AChR] (155). Thus, the actions of agonists at the nicotinic AChR are modulated by  $\sigma_1 R$  selective ligands (160). In addition, the increased ACh level seen in rat frontal cortex induced by (+)N-allylnormetazocine supports the activity of  $\sigma Rs$  in ACh regulation (357–359).

# Nitric oxide

It has been shown *in vitro* that  $\sigma R$  ligands prevent Gluinduced activation of nitric oxide synthetase [NOS] (360). Nitric oxide [NO] is an important mediator in ischemic brain injury (361–363), and in many other disease states. Specifically, NO derived from constitutively expressed NOS in neurons [nNOS] and the inducible isoform expressed by many cells [iNOS] are important in excitotoxic injury cascades (363,364), such as can be seen following exposure to EAAs. Pharmacologically selective inhibitors of nNOS and iNOS, such as the  $\sigma_1 R$  (365), attenuate infarction volume after focal cerebral ischemia (362,366,367).

A potent  $\sigma_1 R$  infusion into normal striatum by microdialysis attenuates basal, and NMDA-evoked, striatal NO production *in situ* (368); therefore, it is not surprising that systemic  $\sigma R$  ligand treatment reduces stroke damage by preventing ischemia-induced NO production (369) with reduced infarct volume (370). These findings have been reproduced more recently (187). For this reason it has been suggested that  $\sigma_1 R$  agonists should be considered as neuroprotective drugs, where some of the protection offered occurs through inhibition of inducible NOS (365).

## Glutamate [Glu]

Although many AAs play a role in neurotransmission, Glu, Gly and GABA are among the more common and betterunderstood neurotransmitters (371–375). Glu mediates an estimated 50% of all the synaptic transmissions in the CNS. Glu, glycine and GABA are metabolic intermediates and neurotransmitters, where Glu is the major excitatory neurotransmitter, and Gly and GABA are the major inhibitory neurotransmitters (326,350,376–380). Glu is involved in nearly all aspects of normal brain function including learning, memory, movement, cognition and development (381–390).

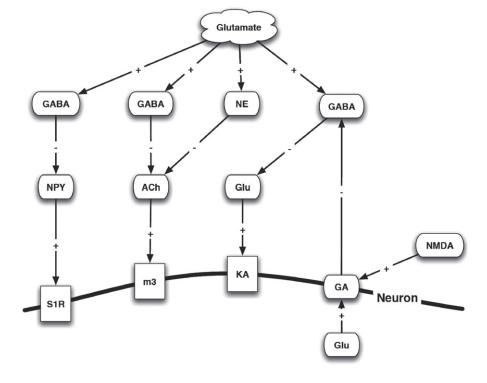
Glu is synthesized, stored and released from the presynaptic terminal, has specific neurotransmitter receptors are localized on the postsynaptic cells, and is eliminated from the synaptic cleft by specific transporters. In addition to Glu, aspartate [Asp] also acts as a major excitatory neurotransmitter (382,391–395) by stimulating or exciting postsynaptic neurons.

From Glu labeling studies, the average concentration of Glu in ganglion cells is 5 mM (396). Physiological studies using isolated cells indicate that only  $\mu M$  levels of

Figure 7. Interaction of glutamate, neurotransmitters and the  $\sigma R.$ NMDA – N-methyl-D-aspartate receptor; NE – norepinephrine; NPY – neuropeptide Y; ACh – acetylcholine; M3 – rat muscarinic acetyl choline receptor; GABA –  $\gamma$ -aminobutyric acid; GA – Ga-binding protein  $\alpha$ -chain; Ka – kainate; Glu – glutamate; S1R –  $\sigma_1 R.$  Glu are required to activate GluRs (397–399). Thus, the amount of Glu released into the synaptic cleft is several orders of magnitude higher than the concentration required to activate most postsynaptic receptors. As  $\sigma$ Rs seem to mediate a number of processes through the Glu system, a more detailed discussion of the Glu system is provided (Figure 7).

N-Acetyl-aspartyl-glutamate [NAAG] is abundant in the mammalian CNS, which has led to the hypothesis that this dipeptide is the storage form of Glu (400,401). Brain tissue has a remarkable ability to accumulate Glu, an ability resulting from Glu transporter [GluT] proteins present in the plasma membranes of both glial cells and neurons (402). Glu is at the center of other metabolic events, e.g. Glu serves as substrate for the synthesis of N-acetyl Glu, an essential allosteric activator of carbamyl phosphate synthetase I, a key regulatory enzyme in the urea cycle (403). It has a welldescribed transdeamination system involving aminotransferases and Glu dehydrogenase, where Glu plays a key catalytic role in the removal of  $\alpha$ -amino nitrogen from AAs. Finally, the "Glu family" of AAs (arginine, ornithine, proline, histidine and glutamine) requires the conversion of these AAs to Glu for their metabolic disposal. The Glu system is probably the mediator of excitatory effects seen following  $\sigma Rs$  stimulation (404) by  $\sigma R$  agonists such as phencyclidine [PCP] (201).

At toxic concentrations, Glu acts as a neurotoxin (excitatory amino acid [EAA]) capable of inducing severe neuronal damage and necrosis by causing over excitation of neurons through receptor-mediated depolarization and Ca<sup>2+</sup> influx (373,405–411). However, Glu is not the only EAA that can cause excitotoxicity and cell death in the CNS (382,391,392,394,412,413). The  $\sigma_1 R$  ligand PRE-084 protects against excitotoxic perinatal brain injury in newborn mice (414), indicating a central role for the  $\sigma Rs$  in modulating the excitatory effects of Glu.



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Other EAAs access the brain tissue of the circumventricular organs located outside the blood brain barrier [BBB] (415–420). An array of GluRs are known to be present on preand postsynaptic membranes that are used to transduce integrated signals using an increased ion flux and second messenger pathways (382,389–392,421,422,). It is the excessive activation of these receptors that leads to neurotoxicity, often referred to as "excitotoxicity".

There are five main factors necessary for the transition of Glu and Asp from neurotransmitters to excitotoxins, including inadequate neuronal ATP levels, inadequate neuronal levels of  $Mg^{2+}$ ; high concentrations of inflammatory prostaglandins; excessive free radical formation (423,424) and inadequate removal of synaptic Glu (296,373,408,425,426). It has been postulated that excitotoxicity is involved in the pathogenesis of many types of acute and chronic insults to the CNS (416) and peripheral tissues (418), and interestingly, excitotoxicity has also been suggested as a central mechanism in fluoride neurotoxicity (427).

Glu and its structural analogues may enter the food supply during preparation or processing as contaminants or additives in its free form or bound to peptides and proteins (428-437). These analogues include monosodium Glu [MSG], L-aspartate, L-cysteine, related sulfur AAs, B-N -oxalyamino-Lalanine [BOAA or ODAP], B-N-methyl-amino-L-alanine [BMAA] and the seafood toxin domoic acid [DomA] (429,432,435,438–440). Structurally similar environmental dietary excitotoxins (441), such as DomA, one of the most potent neurotoxins in seafood can enter our food supply (439). Contamination of mussels by sea diatoms producing DomA (429-431,442), results in neuronal excitation resulting in severe seizures (429-431,433,434,439,442). Survivors of severe cases suffered permanent loss of short-term memory, a phenomenon that lead to the term amnesic shellfish poisoning (415,418,431,434,437,439).

It now is clear that the  $\sigma Rs$  are important in modulating Glu-mediated seizures (443), and protects neurons against Glu toxicity *in vitro* (444), although direct interaction with NMDARs should not be forgotten as a crucial element in the neuroprotective effects of  $\sigma Rs$  ligands with affinity for NMDARs (445,446).

Although excitotoxic effects can be pronounced during acute events such as ischemic stroke and trauma, they can occur in prolonged chronic neurodegenerative diseases such as AD (425), Parkinsons disease [PD] (447), Huntingtons disease [HD] (448) and Amyotrophic Lateral Sclerosis (373,449) schizophrenia, [ALS] anxiety, depression (425,450,451). These are likely associated with  $\sigma R$  stimulation. Recently, a mutation in the  $\sigma_1 R$  has been associated with juvenile ALS (452); therefore, it is not surprising that  $\sigma_1 R$  agonists improve motor function and motor neuron survival in ALS mice (453). In fact, loss of  $\sigma_1 R$  has been associated with defective autophagy and lipid raft disturbances (454).

In contrast to the effects of  $\sigma R$  stimulation, antagonism of the  $\sigma Rs$  blocks compulsive-like eating behavior (455), enhances brain plasticity (456) and exacerbates other addictions (457). In addition, glutamatergic dysfunction has been postulated as being part of the development of disorders associated with long-term plastic changes in the CNS such as chronic pain (458), drug tolerance, dependence, addiction, partial complex seizures and tardive dyskinesia (373,459).

L-Glu acts through both ligand-gated ion channels at the iGluR and at G-protein-coupled metabotropic glutamate receptors [mGluR] (Figure 8). Activation of these receptors is responsible for basal excitatory synaptic transmission and many forms of synaptic plasticity such as LTP and long-term depression [LTD], which are thought to underlie learning and memory (216,371,460–473).

Transporter proteins (Glutamate transporter [GluT]) represent the only significant mechanism for removal of Glu from the extracellular fluid and are important for the long-term maintenance of low and non-toxic concentrations of Glu and appear to have more sophisticated functions in the modulation of neurotransmission (402). A number of soluble compounds, including Glu, cytokines and growth factors, influence the GluT expression and activities (474). It is not known as to whether the  $\sigma$ Rs are involved in regulation of this transport.

The genes encoding GluT proteins have been cloned both from rats and humans (475–480). They are found in astroglia and microglia widely distributed throughout the CNS (481,482) and provide Glu for synthesis of GABA, glutathione and protein (402,483). They rapidly remove Glu from the synaptic cleft to prevent cell death (484).

Many tissues demonstrate Glu, GluR and GluTs, (396,418, 432,434,485–526). mGluRs and L-Glu, L-aspartate and D-aspartate are substrates for the transporters (217,495,521, 527), whereas GluR agonists (528) and antagonists (495,529) are not.

GluTs incorporate Glu into cells along with the cotransport of three Na<sup>+</sup> ions (527,530) and the antiport of one K<sup>+</sup> ion (529,531) and either one OH<sup>-</sup> or one HCO<sub>3</sub><sup>-</sup> ion. The excess Na<sup>+</sup> ions generate a net positive inward current, which drives the GluT (527). In addition, a Glu-elicited Cl<sup>-</sup> current is also associated with some GluT (475,532). In contrast, the vesicular transporter selectively concentrates Glu into synaptic vesicles in a Na<sup>+</sup>-independent, ATP-dependent manner (533–535) that requires Cl<sup>-</sup> (374,375,533,536–542). Given the complexity of the Glu system and the limited information regarding the interaction of the multiple components with  $\sigma$ Rs, further research is necessary to fully elucidate interaction of  $\sigma$ R and the system components.

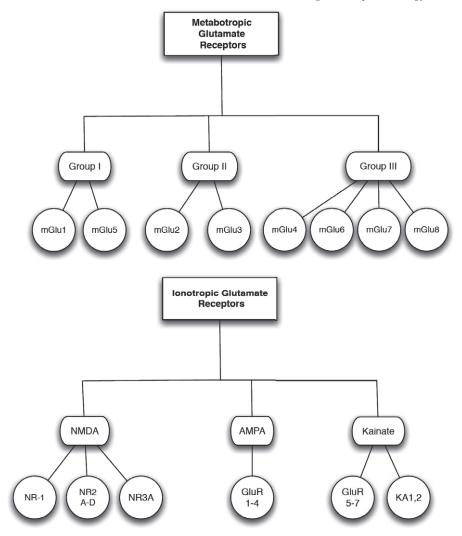
*Glu receptors [GluR].* Two classes of GluRs have been characterized based on studies in the CNS: iGluRs and mGluRs (382,388,389,391,392,421,422,518,543,544). The iGluRs are ligand-gated ion channels that mediate the vast majority of excitatory neurotransmission in the brain. They are classified into three major subtypes: NMDARs, AMPARs and KaRs (296,373,381,382,392–394,449,543,545–554). These receptors exhibit varied pharmacological and electrophysiological properties, including ionic channel selectivity to Na<sup>+</sup>, K<sup>+</sup> and Ca<sup>2+</sup> (389,543).

#### NMDA receptors [NMDAR]

The NMDAR is perhaps the best characterized of the iGlu, in part due to the existence of selective agonists and antagonists that can be used to study its physiology. These receptors are

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Figure 8. Glutamate receptors types. NMDA – N-methyl-D-aspartate receptor; AMPA –  $\alpha$ -amino-3-hydroxy-5-methyl-4isoxazolepropionic acid; KA – kainate; GluR – glutamate receptor; NR – NMDA receptor subtype.



modulated by  $\sigma_1 R$ . NMDAR are ubiquitous (555). NMDARs are composed of assemblies of NR<sub>1</sub> subunits and NR<sub>2</sub> subunits, which can be one of four separate gene products [NR<sub>2A-D</sub>]. Expression of both subunits is required to form functional channels (556).

NMDARs are structurally complex, with separate binding sites for Glu, Gly,  $Mg^{2+}$  (557,558),  $Zn^{2+}$  and a polyamine recognition site, where  $Mg^{2+}$  ions provide a voltage-dependent block of NMDA-gated channels (394).

NMDAR are highly permeable for  $Ca^{2+}$ . They show slower gating kinetics with the channel blocked in a voltage- and usedependent manner by physiological concentrations of Mg<sup>2+</sup> ions (371,372,458,459,462,559). It is this property of the NMDAR that enables  $\sigma$ Rs to trigger cell death via Ca<sup>2+</sup> overload.

## AMPA receptors [AMPAR]

AMPAR are involved in mediating most forms of fast glutamatergic neurotransmission, which corresponds to a  $Ca^{2+}$  influx, a function controlled by the GluR<sub>2</sub> subunit (560). There are four known subunits GluR<sub>1</sub> to GluR<sub>4</sub>, sometimes referred to as GluR<sub>A</sub> to GluR<sub>D</sub>, are widely, but differentially, distributed throughout the CNS (392). AMPARs play an important role in memory function. They are localized in the

hippocampus and striatum and also may play a role in the generation of seizures (560–562).

## Kainate receptors [KaR]

Kainate receptors [KaR] constitute a separate group from the NMDAR and AMPAR, although they share many of the same structural characteristics. KaRs and AMPARs are localized in the hippocampus and striatum and also may play a role in the generation of seizures (563–566). Also they are involved post-synaptically in neurotransmission in some pathways (566–569).

*Metabotropic Glu receptors [mGluR].* mGluRs form a family of currently eight subtypes (mGluR<sub>1-8</sub>), subdivided into three groups (I–III) (570–572). Activation of group-II (mGluR<sub>2,3</sub>) or group-III mGluRs (mGluR<sub>4,6-8</sub>) has been established to be neuroprotective *in vitro* and *in vivo* (572), and for the NMDA iGluR (573). In contrast, group-I mGluRs (mGluR<sub>1,5</sub>) need to be antagonized in order to evoke protection (448) antagonists, and drugs acting on 5-HT<sub>2A</sub>,  $\alpha_2$ -adrenergic, adenosine (A2A) and cannabinoid [CB<sub>1</sub>] receptors may be helpful (574).

Members of this family of mGluR exert their effects either on the second messengers or ion channels via the activation of the GTP-binding proteins and regulate the synthesis of different intracellular second messengers such as IP<sub>3</sub>, cAMP or cGMP, as do  $\sigma$ Rs (382,422). They function to modulate the presynaptic release of Glu and the post-synaptic sensitivity of the cell to Glu excitation (382,389,390,392,422).

mGluRs have both chemical and electrical signaling properties (575). Glu binding onto an mGluR opens non-selective cation channels more permeable to Na<sup>+</sup> and K<sup>+</sup> ions than Ca<sup>2+</sup> (548,576). mGlu binding elicits a rapidly activating inward and outward current and Ka, quisqualate and AMPA are the specific agonists at these receptors (399,577–583).

As with iGluRs, the mGluRs are classified into 4 groups (Group I–IV) based on AA sequence similarities, agonist pharmacology and the signal transduction pathways to which they are coupled (584). Each receptor is formed from the co-assembly of several subunits (584–587). To date, eight subunits (named GluR<sub>1</sub> through GluR<sub>8</sub>) have been cloned (393,576,586,588–591).

 $\sigma Rs$  and Glu neurotransmission. Numerous studies have shown interactions between  $\sigma Rs$  and NMDAR-mediated responses. For example,  $\sigma R$  ligands, including haloperidol, (+)-PTZ, 4-IBP (592), (+)-3-PPP, (+)-SKF-10,047 (593) and DTG (594), antagonize NMDAR currents in Xenopus oocytes (595). The effects of  $\sigma R$  ligands on NMDARs in are thought to be indirect; however, high doses ( $\mu$ M) and nonselective  $\sigma$ R ligands have been used in past studies. Furthermore, there was no correlation between the potency of NMDAR inhibition and the affinity or stereo selectivity for  $\sigma R$  sites (595–597). Thus, it is difficult to assess whether these observations have been based on  $\sigma R$  mediated actions rather than on non-specific effects. In vitro radioligand binding studies have shown that haloperidol, (+)-PTZ, DTG, (+)-SKF-10,047 and (+)-3-PPP inhibited [3H]TCP binding to NMDARs in neuronal cells, with a potency correlated with the affinity for DTG binding sites (64,598).

In a model of modulation of the NMDA response in dorsal hippocampal pyramidal neurons of the CA<sub>3</sub> region, it was found that low doses of the  $\sigma R$  ligands DTG, JO-1784, (+)-PTZ and L-687,384 selectively potentiated the response of these neurons to microiontophoretic applications of NMDA (137,194,195,197). Other  $\sigma R$  ligands such as BD-737, 4-IBP and OPC-14523 were less selective (196,376).

Interestingly, it was also found that depending on the initial level of excitatory response to QUIS and NMDA,  $\sigma R$  agonists could increase or decrease NMDA-induced responses, thus suggesting a real modulatory role of  $\sigma R$  ligands on the Glu response (377). Antagonists including SA4503 (593), BMY-14802, (+)-3-PPP and NE-100, suppress the potentiation induced by  $\sigma R$  agonists (162,195,197).

The effects of all  $\sigma_1 R$  agonists on the NMDA response produce a biphasic dose response curve, which will be discussed later (194,376,377). As stated above, this particular pharmacological profile could explain the discrepancies observed for the effects of  $\sigma R$  ligands with respect to inhibition versus potentiation on NMDAR-mediated responses, as most *in vitro* studies may have used high doses, at which the  $\sigma R$  ligands were acting as antagonists.

In contrast, the antidepressants paroxetine and tranylcypromine, which have a low affinity for  $\sigma Rs$ , have no effect on the NMDA response despite their similar monoaminergic profiles to sertraline and clorgyline. Moreover, the effects of sertraline and clorgyline are suppressed by the  $\sigma R$  antagonist haloperidol but not by spiperone, suggesting that their effects are likely mediated by  $\sigma Rs$  (197). The  $\sigma_2 R$  ligands Lu 28-179 (599) and BD-1008 (600) have also been shown to modulate NMDA mediated responses.

Despite their high affinity for  $\sigma_2 Rs$ , the doses required for antidepressant activity are 5–10 times higher than  $\sigma_1 R$ ligands (102). The effects of the specific  $\sigma_2 R$  ligand Lu 28–179, are not blocked by the  $\sigma_1 R$  antagonists NE-100, PROG, or haloperidol, suggesting that these effects are mediated through  $\sigma_2 R$  (102).

In vitro models have also suggested a modulatory role for  $\sigma R$  agonists on NMDA-mediated responses. For example, JO-1784, BD-737, (+)-PTZ and (+)-3-PPP potentiates in a concentration-dependent manner NMDA-induced [<sup>3</sup>H]NE release from preloaded rat hippocampal slices (135, 162,210), whereas DTG and BD-737 act as inverse agonists, by concentration dependently inhibiting the overflow of [<sup>3</sup>H]NE evoked by NMDA. Haloperidol and BD-1063 (208) alone do not modify [<sup>3</sup>H]NE release, but completely prevent the effects of JO-1784, BD-737, (+)-PTZ, DTG and (+)-3-PPP (162), whereas DuP734 inhibits that of BD-737 (122).

Neurosteroids, acting as  $\sigma R$  agonists, have also been shown to modulate NMDAR-mediated effects (601), as DHEA at low doses potentiates the NMDA response in extracellular recordings from the dorsal hippocampus. The effect of DHEA is blocked by NE-100 and haloperidol (123,179). In this model, neither PREG nor PREGS modifies the NMDA response or act as antagonists (602), which may be due to their lower affinity for the  $\sigma_1 R$  (82,283).

Endogenous hormone levels also affect the  $\sigma Rs$  modulatory effect on NMDA-mediated responses. For example, two weeks following OVX, the potentiation of the NMDA response induced by DTG was significantly greater than in control female rats, suggesting that  $\sigma Rs$  may be tonically inhibited by endogenous PROG (123,180). In agreement, 10 times higher doses of (+)-PTZ and DHEA are required in pregnant females to potentiate the NMDA response. This reduction of effect of  $\sigma R$  agonists in late pregnancy may be due to occupation of  $\sigma R$  by high concentrations of PROG and the apparent super sensitivity of  $\sigma R$  observed during the postpartum period that might be due to the rapid drop of PROG levels after parturition (603–605). Overall, many  $\sigma_1 R$  ligands have demonstrated the ability to modulate NMDA-mediated Glu neurotransmission.

## $\gamma$ -Aminobutyric acid (GABA)

Glutamic acid decarboxylase [GAD] in mouse brain is capable of decarboxylating Glu to GABA but requires pyridoxal 5-phosphate as a cofactor (606–611). The role of GABA as a neurotransmitter is that of inhibitory neurotransmission, although this property has been questioned recently (612). Following the purification of GAD and the generation of GAD antisera, immunohistochemical studies reveal that many GABAergic neurons in brain are interneurons and are, therefore, uniquely able to alter the excitability of local circuits within a given brain region (611,613). From these and other studies it has been confirmed that 30–40% of all CNS neurons utilize GABA as their primary neurotransmitter.

GABA is formed *in vivo* via a metabolic pathway called the "GABA shunt." The initial step in this pathway utilizes  $\alpha$ -ketoglutarate formed from glucose metabolism via the Krebs cycle.  $\alpha$ -Ketoglutarate is then transaminated by  $\alpha$ -oxoglutarate transaminase (GABA-T) to form Glu, the immediate precursor of GABA. Finally, Glu is decarboxylated to form GABA by the GAD (607,608,614). GAD is expressed only in GABAergic neurons and in certain peripheral tissues, which are also known to synthesize GABA (615).

The principal neuronal GABA transporter is a 70–80 kDa glycoprotein that contains 12 hydrophobic membrane-spanning domains (616,617). Specific inhibitors of GABA uptake that directly bind to the transporter have anticonvulsant and antinociceptive properties in laboratory animals (500). The role of  $\sigma R$  interaction with GABA and GABA transporters has yet to be elucidated, but given their role in NMDARs, a role for them could be postulated.

Conformationally-restricted analogues of GABA have been used to help identify three major GABARs, termed GABA<sub>A</sub>(618–620), GABA<sub>B</sub> and GABA<sub>C</sub> receptors (621,622) GABA<sub>A</sub> and GABA<sub>C</sub> receptors are members of a superfamily of transmitter-gated ion channels that include nACh (623), strychnine-sensitive Gly and 5-HT<sub>3</sub> receptors (618,619). On the other hand, GABA<sub>B</sub>Rs are seven transmembrane receptors that are coupled to G-proteins and activate second messenger systems and Ca<sup>2+</sup> and K<sup>+</sup> ion channels, resembling the activity of mGluRs (624).

The large numbers of drug recognition sites associated with GABA<sub>A</sub>Rs, suggested that there may be an endogenous receptor ligand including two natural reduced steroid metabolites of PROG and deoxycorticosterone: ALLO and allotetrahydro-DOC (619,625,626). However there is little compelling evidence at present that any interact with GABA<sub>A</sub>Rs *in vivo* (627). More recently, N,N-dimethyltryp-tamine [DMT] has been shown to be the endogenous ligand of  $\sigma$ Rs (7,8), not the neurosteroids as previously thought.

To date, five distinct classes of polypeptide subunits ( $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$  and  $\rho$ ) have been cloned (618,620) and multiple isoforms of each have been shown to (628). There is approximately 70% sequence identity between the polypeptide subunits within a given class, but only approximately 30% between classes (629,630–632).

### Glycine [Gly]

Gly is the major inhibitory neurotransmitter in the brainstem and spinal cord and functions as a co-agonist at the NMDA subtype of GluR, finely modulated by local expression of specific Gly transporters such as  $GLYT_1$  (633) in the forebrain, where it promotes the actions of Glu, the major excitatory neurotransmitter (449). Thus, Gly serves both inhibitory and excitatory functions within the CNS.

The actions of Gly are terminated primarily by reuptake via  $Na^+$ -Cl<sup>-</sup>-dependent, high-affinity Gly transporters [GlyT]. Like GABA, this increase in Cl<sup>-</sup> ion conductance results in a hyperpolarization of the neuronal membrane and an antagonism of other depolarizing stimuli (634). Given their

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impact on NMDARs,  $\sigma_1 Rs$  and their activation are probably potentiating factors for glycine transmission.

# Cannabinoids

Cannabis and cannabinoids exert most of their biological functions through receptor-mediated mechanisms. Two types of cannabinoid receptors [CB] have been identified – namely CB<sub>1</sub> and CB<sub>2</sub> – both coupled to a G protein (635). CB<sub>1</sub> receptors have been detected and quantified in the CNS (636). They are responsible for the characteristic effects of cannabis, including catalepsy, depression of motor activity, analgesia and feelings of relaxation/well being. Cannabis also affects peripheral neurons; activation of CBs produces suppression in neurotransmitter release in the heart, bladder, intestine and *vas deferens* (637,638).

CB<sub>1</sub> cannabinoid receptors appear to mediate most, if not all of the psychoactive effects of  $\delta$ -9-tetrahydrocannabinol [ $\delta$ THC] and related cannabinoid compounds. This G proteincoupled receptor has a characteristic distribution in the nervous system: It is particularly enriched in cortex, hippocampus, amygdala, basal ganglia outflow tracts and cerebellum, a distribution that corresponds to the most prominent behavioral effects of cannabis (637).

Cannabinoid  $CB_2$  receptors have only been detected outside the central nervous system, mostly in cells of the immune system, presumably mediating cannabinoid-induced immunosuppression and anti-inflammatory effects (639). With the discovery of cannabinoid receptors for exogenous cannabinoids, endogenous cannabinoids (anandamide, 2arachidonylglycerol [2-AG]) have been described subsequently (638,640).

Endocannabinoids not only act at cannabinoid receptors, but potentially also at vanilloid and 5-HT<sub>3</sub> receptors, both of which are expressed in the gastrointestinal tract. The interactions between endocannabinoids and these other important receptor systems have not been extensively investigated (641). Additionally, experimental evidence also suggests that endocannabinoids mediate neuron-astrocyte communication (642).

The relationship of cannabinoids to the  $\sigma Rs$  has received little attention, although the interaction of  $CB_1$  with the classical opiate receptors has been investigated (635). Decades ago, it was been shown that the morphine-induced dopamine release in the nucleus accumbens requires the CB<sub>1</sub> activation (643). Although studies seldom include investigation of the  $\sigma Rs$ , the effects on other neurotransmitter systems suggest a possibility of interaction of the CB and  $\sigma$ Rs. For example, both the serotonergic (644) and endocannabinoid systems modulate frontocortical Glu release (645). Cannabinoid CB1 receptor antagonists rimonabant (SR141716) and AM251 directly potentiate  $\mbox{GABA}_{\rm A}$ receptors (646), inferring that CB<sub>1</sub> receptor agonists may do the reverse; thus damping the excitatory effects of Glu. In fact, endocannabinoids control GABA effects (647), mediates inhibition Glu transmission in the hippocampus (648) leading to the neuroprotective role on the cannabinoids (649) via negative signaling through the G-protein-coupled cannabinoid receptors (650). Although specific, direct data are absent for the role that  $\sigma Rs$  play in the cannabinoid

modulation, the role that  $\sigma Rs$  play in Glu modulation suggests that they are probably involved in the modulation of Glu produced by the endocannabinoids. To support this hypothesis, the endocannabinoid system plays a central role in the phenomenon of addiction (651), as do the  $\sigma Rs$ . Hence, some of the changes in  $\sigma R$  signaling seen in addictions probably occur in concert with the endocannabinoid system. In fact, it has recently been suggested that  $\sigma_1 R$ dysfunction might increase vulnerability to cannabis-induced psychosis (652).

#### Summary

Summarizing the interactions of  $\sigma R$  with neurotransmitters is difficult. Data are scarce and incomplete. In addition, the dose-response of stimulation of the  $\sigma R$  to an agonist can show stimulatory effects at a low dose and inhibitory effects at high doses, when used experimentally using greater concentrations than physiological levels. As most work is done in *in vitro*, doses are often excessive and may reflect an overexposure that would not be seen in the *in vivo* situation. Nonetheless, it seems clear that the  $\sigma Rs$  have a core and only partly defined role in regulation of neurotransmission.

# Pharmacology

A diverse class of psychotropic drugs bind to  $\sigma_1 Rs$  (653), including antipsychotics, e.g. haloperidol [Haldol<sup>®</sup>], which have the highest affinity for  $\sigma_1 R$  (176,177,654), SSRIs, which have medium to high affinities for  $\sigma_1 Rs$ , and tricyclic antidepressants [TCAs], which have less (176,655). Other compounds that bind to the receptor include morphinans (e.g. DEX) (69), guanidines (e.g. DTG) (193), phenothiazines (e.g. chlorpromazine) (656), butyrophenones (e.g. haloperidol) (657), TCAs (e.g. imipramine) (658), monoamine oxidase inhibitors [MAOI] (e.g. clorgyline) (659), SSRIs (e.g. sertraline) (660), cytochrome P<sub>450</sub> inhibitors (e.g. proadifen) (192), anticonvulsants (e.g. phenytoin) (141), addictive drugs (e.g. cocaine, METH) (661), polyamines (e.g. ifenprodil) (662) and certain steroids (e.g. progesterone [PROG] and testosterone) (98). The effects of cocaine occur through direct involvement of  $\sigma_1 R$  and the DA<sub>1</sub> receptor (663). In addition, the anticonvulsant drug phenytoin allosterically modulates  $\sigma_1 Rs$ (141). These receptors also exhibit a high affinity for (+)-isomers and are proposed to be associated with both pertussis toxin-sensitive  $G_{i/o}$  and cholera toxin-sensitive  $G_s$ proteins, PLC and PKC (165).

The  $\sigma_1$ Rs have an affinity for a number of specific stereoisomers of these drugs (e.g. (+) PTZ and (+) cyclazocine) (65,79,179,252,653,664). The lack of selectivity between the  $\sigma$  and PCP binding sites seen following exposure to several compounds, including BZM or PCP derivatives, led to a confusion resolved by the availability of new highly selective drugs. Among them, the reference PCP non-competitive antagonist (+)MK-801 maleate [dizocilpine] failed to displace radioligands labeling the  $\sigma$ R sites. Selective  $\sigma$ R agonists like 1,3-di-O-tolylguanidine [DTG], (+)N-cyclopropylmethyl-N-methyl-1,4-diphenyl-1-ethyl-but-3-en-1-ylamine hydrochloride [JO-1784, igmesine], 2-(4-morpholino)ethyl-1-phenylcyclohexane-1-carboxylate

hydrochloride [PRE-084] and 1-(3,4-dimethoxyphenethyl)-4-(3-phenylpropyl)piperazine dihydrochloride [SA4503], do not bind to the NMDAR-associated PCP site (212). These compounds are now reference compounds in terms of selectivity between  $\sigma$  and PCP receptors.

Neurochemical and electrophysiological studies have then been crucial in revealing the function of the  $\sigma R$ (26,129,130). These studies have demonstrated that  $\sigma Rs$ play a role as modulators of Ca<sup>2+</sup> release (164,665) and inhibitors of voltage-gated potassium K<sup>+</sup> channels (138,139), NMDARs, tyrosine kinase [TK]-related processes (666), *IP*<sub>3</sub>R activation (139,161,164,165), other iGluR and mGluR functions (548), neurosteroids (667) and other neurotransmitter activities (211).

 $\sigma_1$ Rs also regulate compartmentalization of lipids in the ER (64,65,164,668,669), and have antitumor activity *in vitro* and *in vivo* (670). Studies also have suggested that  $\sigma_1$ Rs regulate lipid transport and metabolism, neurogenesis (671), cellular differentiation and myelination in the brain (672); the latter has implications for diseases such as multiple sclerosis [MS].

The actions mediated by  $\sigma_1 Rs$  at the cellular level can be considered either as acute or chronic. The acute actions include the modulation of ion channels (e.g. K<sup>+</sup> channel), NMDARs, IP<sub>3</sub>R and  $\sigma_1 R$  translocation. Chronic actions of  $\sigma_1 Rs$  are basically considered to be the result of an up- or down regulation of the  $\sigma_1 R$  itself. For example, the up regulation of  $\sigma_1 R$  *per se*, even without exogenous ligands, promotes cellular differentiation and reconstitution of lipid "micro domains" in cultured cells (65,673). Recent *in vitro* and *in vivo* studies strongly point to the possibility that  $\sigma_1 Rs$ participate in membrane remodeling and cellular differentiation in the nervous system (65).

Metabolic studies support the view that  $\sigma Rs$  have functional significance in brain glucose metabolism as glucose utilization is affected by ligands in areas of brain that show high densities of  $\sigma Rs$  (78). The findings of up and down regulation, suggest that  $\sigma_1 Rs$  might possess a constitutive biological activity, and that  $\sigma_1 R$  ligands might merely work as modulators of the innate activity of this protein.

 $\sigma_1 Rs$  are present throughout vertebrate evolution, with conserved pharmacologic properties (674), in sea anemones, planaria, earthworm, crayfish, cricket, hadfish, shark, goldfish, frog, turtle, chicken, guinea pig and monkey. There does not appear to be a family-related trend in quantity, e.g. monkey has only 20% of the  $\sigma$ Rs seen in the guinea pig (675). Also,  $\sigma Rs$  differ from classical neurotransmitter receptors in that they show no postnatal ontogeny in the rat and no agedependent change in the receptor density. The lack of postnatal development of receptors in the CNS, as compared with postnatal changes in other classical neurotransmitter receptors, and the fact that  $\sigma R$  sites are much denser in peripheral organs, such as the liver (418), immune and endocrine tissues (676, 677), suggest a universal role for  $\sigma Rs$ in cellular function. Because of their widespread modulatory role,  $\sigma_1 R$  ligands have been proposed to be useful in several therapeutic fields such as amnesic and cognitive deficits, depression and anxiety, schizophrenia, analgesia and against some effects of drugs of abuse (such as cocaine and METH) (32,678).

#### $\sigma_1 R$ ligands

 $\sigma R$  agonists and antagonists are common in easily available drugs, e.g. DEX in cough medications. These antagonists

Table	1.	Some	$\sigma R$	ligands	in	order	of	their	potency.

$\sigma R$ ligands in rough order of potency					
Sigma <sub>1</sub> ligands	Sigma <sub>2</sub> ligands				
(+)-pentazocine [PTZ] Haloperidol [Haldol <sup>®</sup> ] 1,3di-o-tolyl-guanidine [DTG] (+)-3-PPP Dextromethorphan [DEX] (+)-SKF 10,047 (+)-cyclazocine (-)-pentazocine [PTZ] Phencyclidine (-)-SKF 10,047	1,3 di-o-tolyl-guanidine [DTG] Haloperidol [Haldol <sup>®</sup> ] (+)-3-PPP [preclamol] (-)-pentazocine [PTZ] Phencyclidine (+)-pentazocine [PTZ] (-)-SKF 10,047 BD1047 BD1063				

Sigma receptor biology show a GTP-sensitive high affinity binding to the  $\sigma_1 R$  (679).

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A complete list of  $\sigma R$  ligands is difficult to obtain, as many compounds only have been used in research (680) and are not available, due to the proprietary nature of drug development. A short list of  $\sigma_1 R$  and  $\sigma_2 R$  ligands can be seen in Table 1.

Each of the above acts as either an agonist or antagonists often depending on the dose. This biphasic dose-response makes evaluation of studies difficult, but does help to explain conflicting findings (681). Bearing in mind the biphasic doseresponse of agonists and antagonists, a summary of some ligands and their agonistic or antagonistic activities can be seen in Table 2.

## Dose response

As previously mentioned,  $\sigma Rs$  do not show a linear doseresponse curve, but show a biphasic dose response curve in various behavioral, biochemical and electrophysiological

Table 2.  $\sigma R$  agonists and antagonists.

$\sigma_1 R$ and $\sigma_2 R$ ligands as	s agonists or antagonists
$\sigma R$ ligands – agonists	$\sigma R$ ligands – antagonists
(+)-N-allylnormetazocine [(+)-SKF 10,047]	(1-[2-(3,4-dichlorophenyl)ethyl]-methylpiperazine [BD1063]
2-(4-morpholino)ethyl-1-phenylcyclohexane-1-carboxylate [PRE-084]	(N-(3,4-dichloropheny)ethyl]—4-methylpiperazine [BD 1008]
1-(3,4-dimethoxyphenethyl)-4-(3-phenylpropyl)piperazine [SA 4503]	[1-cyclopropylmethyl)-4-(2'(4"-flurophenyl)-2'-oxoethyl)piperidine [DuP 734]
1'-[4-[1-(4-fluorophenyl)-1H-indol-3-yl]-1-butyl]- spiro[isobenzofuran-1(3H), 4'piperidine] [Lu 28-179]	2-amino-7phosphonoheptanoic acid [AP-7]
Fluoxamine	NE-100
Pregenolone-S	E-5842
DHEA-S	BD1139
Donepezil	BIMU-8
PPBP	BMY 14802
Amitriptyline	Cabetapentane
BD 737	Dextromethorphan [DEX]
Ibogane	Eliprodil [SL 82.0715]
Haloperidol ( $\sigma_2 R$ ) [Haldol <sup>®</sup> ]	Fenpropimorph
BD 737	Haloperidol ( $\sigma_1 R$ ) [Haldol <sup>®</sup> ]
4-(N-benzylpiperidin-4-yl)-4-iodobenzamide [4-IBP]	Ifenprodil tartrate
3,4-methylenedioxymethamphetamine [MDMA]	N,N-dipropyl-2-[4-mrthoxy-3-(2-phenylethoxy)phenyl]ethylamine monohydrochloride [NE-100])
Dehyroepiandrosterone sulfate [DHEA-S] [suggested as	N-2-[2-(3,4-dichlorophenyl)ethyl]-N-methyl-2-(dimethylamino)
the endogenous $\sigma R$ agonist]	ethylamine [BD 1047]
(+)Cyclozocine	N-methyl-D-aspartate [NMDA]
Siramesine	N-phenthylpiperidine
Igmesine	Opipramole
Fluvoxamine	Panamasine
Dextromethorphan [DEX]	Pregnalone
OPC-14523	<b>PROG</b> (suggested as the endogenous $\sigma R$ antagonist)
CB-64D	Rimcazole
Ditolylguanidine [1,3-di-O-tolyguanidin] [DTG]	Sabeluzole
Memantine	Testosterone
Certain steroids (agonist plus steroidal effect)	Tiopirone
Phencyclidine [PCP]	Verapamil
Donepezil	WAY 100635
Igmesine [J01783]	Sertraline
Interleukin 10 [I L-10]	
(+)3H-3-3(3-Hydroxyphenyl)-N-(1-propyl)-piperidine [SA4503]	
Methamphetamine	
Phenothazines	
(+)-pentazocine [PTZ]	
Heroin	
Cocaine	
2-(4-morholinethyl)1-phenycyclohexanecarboxylate	
Amantadine	
CB-184	
Dimemorfan	

paradigms (135,137,161,182,194,195). For example, the  $\sigma R$  agonist, SA4503, both attenuates and enhances the effects of methamphetamine depending on the dose (682).

A similar dose response curve has also been described with  $\sigma R$  ligands in other models such as in release experiments (135) and in behavioral models (182,184). It has been proposed that the different dose response may be due to low doses of  $\sigma R$  ligands activating one subtype of  $\sigma Rs$  for which they have high affinity, whereas higher doses may activate another subtype(s) of the  $\sigma R$  for which they have a lower affinity). Such activity would counteract the effects observed at lower doses (194,196,197). Nonetheless, it is important to note that the different, and sometimes opposite, results obtained with low and high doses of  $\sigma R$  ligands may explain much of the controversy seen in the literature on  $\sigma Rs$ .

The fact that low dose effects do not follow the classical dose-response curve has been known for many years and has only been recently revived under the title of hormesis (683–709). The *Arndt-Schulz rule* or *Schulz' ''law''* is a basically a hypothesis concerning the observed effects of many chemicals in low concentrations (710–713). According to the Arndt-Schulz rule, highly diluted chemicals enhance life processes, while strong concentrations of the same chemical may inhibit these processes and even terminate these processes (714).

Depending on the process affected, this interplay results in either a J-shaped or inverted J-shaped dose response, which are sometimes called "bell-shaped", "U-shaped," "inverted U-shaped," "biphasic" or " $\beta$ -curve" (685,686, 710,715–732). The point at which the hormetic curve crosses the reference level of response (i.e. the threshold) is the zero equivalent point [ZEP]; in other words, the point at which there is no toxic or stimulatory effect.

At low doses of an  $\sigma R$  agonist induction and potentiation of the NMDA response is seen (162,195). In contrast, at higher doses the effects of  $\sigma R$  agonists such as DEX, and Igmesine (733) progressively decrease and disappear (194). A similar dose response curve has also been described with  $\sigma R$  ligands in other models such as in neurotransmitter release experiments (135) and in behavioral models (182).

It has been proposed that the biphasic dose response curves may be explained by low doses of an  $\sigma R$  ligand activating one subtype of  $\sigma R$  for which they have high affinity, whereas higher doses activate another or other subtype(s) of the  $\sigma Rs$  for which they have a lower affinity.

Such a mechanism would counteract the effects observed at lower doses (194,196,197). Nonetheless, it is important to note that the different, and sometimes opposite, results obtained with low and high doses of  $\sigma R$  ligands.

## Models

 $\sigma R$  ligands have been proposed for tumor imaging studies (734), particularly in the detection of pulmonary and abdominal tumors (735), despite irreversible binding in some cases, e.g. (<sup>11</sup>C)-SA5845 (736). In fact, selective  $\sigma_2 R$  ligands preferentially bind to pancreatic adenocarcinomas; thus, expanding the possibility of  $\sigma_2 R$ -based applications in diagnostic imaging, in addition to therapy (110) or drug development (239). A haloperidol challenge has shown that

[<sup>123</sup>I]TPCNE is a novel is a single photon emission-coupled tomography [SPECT] tracer for the  $\sigma_1 R$  (737).

A  $\sigma_1 R$  knockout mouse has been developed. The mice demonstrated no overt abnormal phenotype when compared to the wild type. The activity of  $\sigma_2 Rs$  seems to be unaffected in  $\sigma_1 R$ -mutant mice. (63). As expected, however, they do lack the locomotor response to the  $\sigma R$  ligand (+)-SKF100047 and display reduced response to pain via the  $\sigma_1 R$  (39).

#### Cell development and plasticity

 $\sigma R$  drugs and neurosteroids, acting at the level of the  $\sigma_1 R$  protein, are important for plasticity, cell development and trophic actions. These are probably mediated by Ca<sup>2+</sup> (82, 185,190,210). This observed plasticity, which is both associated with the same prerequisite enhancement of NMDA-mediated glutamatergic neurotransmission and protein dephosphorylation that occur downstream from the massive entry of Ca<sup>2+</sup>, and [Ca<sup>2+</sup>]<sub>*i*</sub> mobilization from the endoplasmic reticulum and the mitochondria, often occur synergistically (188,738).

Intracellular chaperones, reside specifically at the endoplasmic reticulum (ER)-mitochondrial interface, referred to as the mitochondrial-associated ER membrane [MAM]. Here,  $\sigma_1 Rs$  is an inter-organelle signaling moderator (665) and regulates ER-mitochondrion Ca<sup>2+</sup> signaling (739).

As previously mentioned (88,133,139,145,146,740), K<sup>+</sup> channels, which control the fine tuning of Ca<sup>2+</sup> entry through both VSCCs and SOCs (store-operated channels), are also prominent targets of the  $\sigma_1 R$  agonist and antagonist drugs.

## Age changes in $\sigma \text{Rs}$

Throughout adulthood, differences in the motor changes elicited by drugs affecting  $\sigma R$  are correlated with the number of receptors in the P<sub>2</sub>, and not the P<sub>3</sub>, cellular fraction (198). Thus, translocation of the  $\sigma_1 R$  from the ER to the cell membrane (190) decreases with age in motor neuron regions. An increase in density of  $\sigma_1 Rs$  found in the aged monkey brain supports this hypothesis (199), as they are not as readily translocated and, therefore, increase in density.

For this reason it has been suggested that age-related memory deficits associated with advancing age may be responsive to up regulation of the  $\sigma$ Rs. In fact, such ability to alleviate memory deficits during aging has been confirmed in humans for the selective  $\sigma_1$ R agonist (+)-*N*-cyclopropyl-methyl-*N*-methyl-1,4-diphenyl-1-ethyl-but-3-en-1-ylamine hydrochloride [Igmesine<sup>®</sup>], which appears more efficient among the elderly (206).

Conversely, the  $\sigma_2 R$  subtype exhibits no stereo selectivity and only low affinities for the (+)-BZM. It does not appear to be modulated by pertussis toxin-sensitive  $G_{i/o}$  proteins (207), and is predominantly located in the motor system and periphery (21). Interestingly, brainstem motor function, which is profoundly sensitive to  $\sigma R$  drugs, decreases with age, resulting in the reduced accuracy and consistency of fine and complex motor performance (208).

In addition to other neurosteroids, which change with age, the discussion above reflects the effects of DHEA(S) on  $\sigma_1$ Rs. Therefore, the age-related changes in neurosteroids probably affect the  $\sigma$ Rs. Alteration of age-related changes in memory

Table 3. Some  $\sigma R$  ligands in order of their potency.

$\sigma_1 R$ ligands	$\sigma_2 R$ ligands
(+)-pentazocine [PTZ] Haloperidol 1,3 di-o-tolyl-guanidine [DTG] (+)-3-PPP Dextromethorphan [DEX] (+)-SKF 10,047 (+)-cyclazocine (-)-pentazocine Phencyclidine (-)-SKF 10,047	1,3 di-o-tolyl-guanidine [DTG] Haloperidol (+)-3-PPP [preclamol) (-)-pentazocine Phencyclidine (+)-pentazocine (-)-SKF 10,047 BD1047 BD1063

probably relates to the balance of excitatory and inhibitory effects on the CNS, in which  $\sigma$ Rs play a role. Several studies in rodents show that GABA<sub>A</sub> agonists impair learning and memory while GABA<sub>A</sub> antagonists enhance memory (213,214).

In humans benzodiazepines [BZD] ( $\sigma$ R agonist) may impair cognition (741–743). On the other hand,  $\sigma$ R agonists enhance memory performance in young rodents and in rodent models of cognitive impairment (182,200,201,203,744). In addition, the NMDAR is involved in the development of longterm potentiation [LTP] (215,216), an essential element of neural plasticity. In addition, it now appears as though DHEA(S) has an ability to modulate neurotransmitter receptors in the CNS that are primarily involved in learning and memory (209). Thus,  $\sigma$ Rs appear to be essential for maintaining neural health and protecting against age-related mammary defects.

## Sigma ligands

Numerous  $\sigma R$  agonists and antagonists have been previously described. Some are common and easily available drugs, e.g. DEX in cough medications. A short list of  $\sigma_1 R$  and  $\sigma_2 R$  ligands can be seen in Table 3.

Each of the above acts as either an agonist or antagonists. A list of agonists and antagonists can be seen in Table 4.

## Known σRs – location and effects

Steroids binding to  $\sigma Rs$  has suggested that  $\sigma Rs$  serve as a link among endocrine, nervous, heart, lung, kidney, liver, intestines, and sexual (745) and immune systems (221); hence, evaluation of  $\sigma Rs$  in one organ in isolation can miss a significant lesion that is only apparent when viewed in conjunction with other  $\sigma R$ -containing organs showing similar, but subtler, lesions.

The tissue density of  $\sigma Rs$  is not uniform and is different for each subtype (376). The highest concentration of receptors is seen in the CNS, followed by the periphery (liver, spleen, endocrine, GIT lung) (171,676,746–749).

Links between  $\sigma_1 Rs$  and G-proteins and mGlu (133,138,139,142,750) implies that the  $\sigma_1 R$  mediates a large number of its effects via the Glu system and as such Glu-related diseases probably have an  $\sigma R$  component to them.

The  $\sigma_1 R$  has been implicated in myriad of disease phenomena, including cardiovascular arrhythmias (751,167), schizophrenia (15), clinical depression [DEP] (752), Parkinsons disease [PD] (574), Alzheimers disease [AD] (753), the effects of cocaine abuse (754) and cancer (95,377,755,756).  $\sigma_1$ Rs are distributed throughout the brain in normal subjects, but decreased in the frontal, temporal and occipital lobes, cerebellum and thalamus in patients with early AD and in the putamen in patients with PD (757). Compromising  $\sigma_1$ Rs at the endoplasmic reticulum results in cytotoxicity in a dose response manner at physiologically relevant concentrations of dopamine (758). In fact, the cytotoxicity of  $\sigma$ R agonists is associated with major changes in cellular metabolism when there is occupancy of the  $\sigma_2$ R (759). More recently, the pharmacological stimulation of the  $\sigma_1$ R has shown some neuro-restorative effects in experimental PD (760).

## Central nervous system

As previously outlined, the CNS appears to be the primary site of  $\sigma R$  activity and effects. Specific regions that have been shown to have concentrations of  $\sigma_1 R$  include, but are not limited to, corpus striatum, nucleus accumbens (61), *substantia nigra, pars compacta* (656), hippocampal pyramidal cell layer (761), hypothalamus, central grey and red nucleus, pontine and cranial nerve nuclei, pontine nuclei, pons – medulla (761), amygdala (762) and cerebellum (763).  $\sigma_1 Rs$ have also been seen in the spinal cord, particularly the ventral and dorsal route ganglia (761), a site that is important for  $\sigma R$ agonist induction of neck dystonia in rats (764). More specifically, the regional distribution of  $\sigma R$  binding within the brain has shown densities at sites as follows: medullapons > midbrain > cerebellum > thalamus > striatum > cortex > hippocampus (78,79).

Studies comparing  $\sigma_1 R$  versus  $\sigma_2 R$  distributions have established that  $\sigma_1 R$  sites are most abundant in the dentate gyrus of the hippocampus, facial nucleus, thalamic and hypothalamic nuclei, with moderate densities found in the striatum, cerebellum, dorsal raphe nucleus and locus coeruleus (171,302,765,766). In agreement, studies of  $\sigma_1 R$ mRNA levels have found high levels of expression in all layers of the cerebral cortex, striatum, hippocampus and cerebellum (767). In comparison,  $\sigma_2 R$  sites are prominent in the *substantia nigra*, central gray matter, oculomotor nuclei, cerebellum, *nucleus accumbens*, amygdala, olfactory bulb, hippocampus and motor cortex (766,768).

The  $\sigma Rs$  are probably essential for Glu regulation. Glu neurons, also containing GluRs, make up an extensive network throughout the cortex, hippocampus, striatum, thalamus, hypothalamus, cerebellum, and visual and auditory centers in the CNS (386,417,769,770).

At the cellular level, in the CNS, the  $\sigma_1 R$  is expressed in neurons, ependymocytes, oligodendrocytes and in the peripheral nervous system [PNS], Schwann cells (171,672,771– 773). GluRs are similarly expressed in these regions (485), which may mediate the excitatory influence of the  $\sigma Rs$ .

The highest levels of  $\sigma_1 R$  immunostaining can be observed in the neurons of the granular layer of the olfactory bulb, hypothalamic nuclei and pyramidal layers of the hippocampus (171). Among other areas that exhibit intense to moderate  $\sigma_1 R$  immunostaining are the superficial cortical layers, striatal areas including the CP and nucleus accumbens (core

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Table 4.  $\sigma R$  agonists and antagonists.

$\sigma_{1}R$ and	$\sigma_{2}R$	ligands	28	agonists	or	antagonists
OIN and	021	nganus	as	agomsts	UI.	antagomsts

$\sigma R$ ligands – agonists	$\sigma R$ ligands antagonists
(+)-N-allylnormetazocine [(+)-SKF 10,047]	(1-[2-(3,4-dichlorophenyl)ethyl]-methylpiperazine [BD1063]
2-(4-morpholino)ethyl-1-phenylcyclohexane-1-carboxylate [PRE-084]	(N-(3,4-dichloropheny)ethyl]-4-methylpiperazine [BD 1008]
1-(3,4-dimethoxyphenethyl)-4-(3-phenylpropyl)piperazine [SA 4503]	[1-cyclopropylmethyl)-4-(2'(4"-flurophenyl)-2'-oxoethyl)piperidine [DuP 734]
1'-[4-[1-(4-fluorophenyl)-1H-indol-3-yl]-1-butyl]-spiro[isobenzofuran-1(3H), 4'piperidine] [Lu 28-179]	2-amino-7phosphonoheptanoic acid [AP-7]
Amitriptyline	E-5842
BD 737	BD1139
Ibogane	BIMU-8
Haloperidol $(\sigma_2 R)$	BMY 14802
BD 737	Cabetapentane
4-(N-benzylpiperidin-4-yl)-4-iodobenzamide [4-IBP]	Dextromethorphan [DEX]
3,4-methylenedioxymethamphetamine [MDMA]	Eliprodil [SL 82.0715]
Dehyroepiandrosterone sulfate [DHEA-S] [suggested as the endogenous	Fenpropimorph
$\sigma R$ agonist]	
(+)Cyclozocine	Haloperidol ( $\sigma_1 R$ )
Siramesine	Ifenprodil tartrate
Igmesine	N,N-dipropyl-2-[4-mrthoxy-3-(2-phenylethoxy)phenyl] ethylamine monohydrochloride [NE-100])
Fluvoxamine	N-2-[2-(3,4-dichlorophenyl)ethyl]-N-methyl-2- (dimethylamino)ethylamine [BD 1047]
Dextromethorphan [DEX]	N-methyl-D-aspartate [NMDA]
OPC-14523	N-phenthylpiperidine
CB-64D	Opipramole
Ditolylguanidine [1,3-di-O-tolyguanidin] [DTG]	Panamasine
Memantine	Pregnalone
Certain steroids (agonist plus steroidal effect)	PROG (suggested as the endogenous $\sigma R$ antagonist)
Phencyclidine [PCP]	Rimcazole
Donepezil	Sabeluzole
Igmesine [J01783]	Testosterone
Interleukin 10 [I L-10]	Tiopirone
(+)3H-3-3(3-Hydroxyphenyl)-N-(1-propyl)-piperidine [SA4503]	Verapamil
Methamphetamine	WAY 100635
Phenothazines	
(+)-pentazocine	
Heroin	
Cocaine	
2-(4-morholinethyl)1-phenycyclohexanecarboxylate	
Amantadine	
CB-184	
3-(4-(4-cyclohexylpiperazin-1-yl)butyl)benzo[d]thiazole-2(3H)-thione (CM156)	
Dimemorfan	

and shell), the midbrain, the motor nuclei of the hindbrain, cerebellar Purkinje cells in the cerebellum and the dorsal horn of the spinal cord. At the subcellular level, the  $\sigma_1 R$  is mostly present within neuronal perikarya and dendrites, where it is associated with microsomal, plasmic, nuclear, or ER membranes (171).

There is adequate direct and circumstantial evidence for abnormal Glu and Glu analogue neurotransmission, suggesting altered  $\sigma R$  activity, in the etiology and pathophysiology of many neurological and psychiatric disorders such as epilepsy, schizophrenia, addiction, DEP, anxiety, AD, HD, PD and ALS. In fact, it has been suggested that the lack, or dysfunction, of  $\sigma_1 R$  exacerbates ALS (774) and AD (42).

 $\sigma$ Rs probably dampen the excitotoxic effect Glu. Excessive Glu effects can be pronounced during acute events such as ischemic stroke and trauma, or milder but prolonged in chronic neurodegenerative diseases such as AD, PD, HD and ALS (425,447,450,451,775,). In addition there appears to be a role for Glu system, and hence  $\sigma$ Rs, in regulation of

manganese  $[Mn^{2+}]$ , mercury  $[Hg^{2+}]$  and lead  $[Pb^{2+}]$  neurotoxicity (379) (Table 5).

#### Memory loss

At the behavioral level,  $\sigma_1 R$  agonists are antiamnesic (602,776–779) and improve the cognitive abilities in experimental animals via the cholinergic system (777,780). In studies using amnesic rodents, the animals' amnesia seemed to be alleviated by  $\sigma_1 R$  agonist ligands (781). Examples include PCP-induced cognitive dysfunctions, and amnesias induced by scopolamine (782), the Ca<sup>2+</sup> channel blocker nimodipine or carbon monoxide (212). In addition,  $\sigma_1 R$  agonists show an enhanced efficacy in animal models of AD-related learning impairments or DEP responses (783,784).

The cognition-improving action of neurosteroids has been shown to be mediated via  $\sigma_1 R$  (181,212). Indeed,  $\sigma_1 R$  ligands and related neurosteroids interfere with the cocaine-induced state of memory loss (785) mediated through inhibition of iNOS (365).  $\sigma_1 R$  agonists also have a similar effect (376).

Table 5. Summary of  $\sigma_1 R$  and associated psychiatric diseases.

Disorder	Substances tested	Study type	Events (121)
Schizophrenia	Haloperidol Eliprodil	Nonclinical	$\sigma_1 R$ ligands modulate NMDA receptors effecting dopamine regulation
	Fluoxamine	Clinical	Reduction of $\sigma_1 R$ receptors in the postmortem schizophrenic brain
			Adjunctive medication of $\sigma_1 R$ ligands effective for cognitive deficits of schizophrenia
Major depressive disorder	Fluoxamine SA4503 Igmesine Neurosteroids	Nonclinical	$\sigma_1 R$ ligands show antidepressive effects in the forced swimming test Neurosteroids, considered as endogenous $\sigma_1 R$ ligands, show antidepressive effects
Obsessive-compulsive disorder	Fluoxamine	Clinical Nonclinical Clinical	Psychotic major depression is improved by fluvoxamine monotherapy Fluvoxamine improves marble-burying behavior in mice through $\sigma_1 R$ activity Fluvoxamine effective for obsessive-compulsive disorder
Alzheimers disease	Donepezil	Nonclinical Clinical	Fluvoxamine enhances the effect of cognitive behavioral therapy Donepezil show neuroprotective properties against $A\beta_{25-35}$ peptide-induced toxicity Donepezil show anti-amnesic effects which are antagonized by $\sigma_1 R$ antagonists Decrease of $\sigma_1 R$ receptors in the Alzheimers disease brain

## Neuroprotection

The detailed mechanism by which  $\sigma Rs$  protect the nervous system is not clear (786). The basic mechanism of neuroprotection can be seen in (Figure 9). At least two subtypes of  $\sigma_1 R$  may affect differentially the Glu-medicated NMDA neurotransmission in the terminal and origin regions of the mesolimbic and nigrostriatal DA-ergic systems. There also probably exists a functional interaction between  $\sigma_2 R$  and NMDARs in the hippocampus (360). However, there is some question as to whether the interaction is direct or indirect. Even so, administration of a  $\sigma_1 R$  agonist delays middle cerebral artery occlusion induced neurodegeneration and white matter injury (787), thus confirming the neuroprotective effect of the  $\sigma_1 R$ . Similarly,  $\sigma R$  agonists have been show to attenuate brain injury after experimental focal cerebral ischemia in several species (788). The current hypothesis is that  $\sigma_1 R$  agonists protect neurons by a mechanism involving the anti-apoptotic protein bcl-2 (27).

## $\sigma Rs$ and neurogenesis

Recent evidence has shown hippocampal atrophy can persist long after CNS damage is resolved resulting in major depression (789-792). This atrophy could be due to a regression of dendritic processes, an inhibition of neurogeneration or the loss of hippocampal neurons (793). It has also been shown that hippocampal atrophy can be reversed by successful antidepressant treatments and that in vitro, classical antidepressants promote neurogenesis (794).  $\sigma_1 R$  has a role in cell morphological changes, specifically in the initiation of neurite outgrowth and sprouting (164,185). In fact, in addition to noted neuronal regeneration, functional recovery has been described following SA-4503 administration (32). Additional support for the neuroprotective effects of  $\sigma_1 R$  is the finding that mutations of the receptor are associated with frontotemporal lobe degeneration and MND (41).

 $\sigma_1 R$  and ankyrins are highly concentrated in the growth cone of NG-108 cells, a region related to neurite sprouting, extension and guidance (164). The  $\sigma_1 R$  agonist (+)- PTZ has

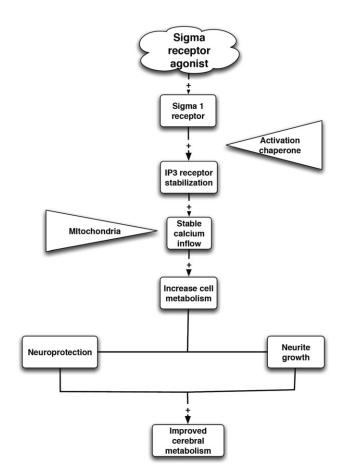


Figure 9. The basic mechanism of neuroprotection by  $\sigma_1 R$  agonists.

no effect by itself on neurite sprouting, but potentiates the neurite-sprouting, induced by nerve growth factor [NGF] (185). In contrast, neurite sprouting, induced by cAMP in PC12 cells, is not affected by (+)-PTZ. The  $\sigma_1 R$  antagonist NE-100, regardless of the presence of NGF, does not affect neurite sprouting, but antagonizes the potentiation induced by (+)-PTZ, thus clearly indicating mediation via  $\sigma_1 R$  (185).

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Interestingly, similar to  $\sigma R$  agonists, the antidepressants imipramine and fluvoxamine potentiate the effects of NGF induce neurite sprouting in PC12 cells (185). These effects of imipramine and fluvoxamine were antagonized by NE-100, while no concentration of 5-HT tested affected neurite sprouting induced by NGF (185), suggesting that the effect on NGF-induced neurite outgrowth of both  $\sigma_1 R$  agonists and classical antidepressants are mediated by  $\sigma_1 R$ . Moreover, cell treatments with NGF, even in the absence of  $\sigma_1 R$  agonists, increased the level of  $\sigma_1 Rs$  in a dose-dependent manner, and the effects of (+)-PTZ and NGF were additive (185).

Interestingly, treatment with imipramine and fluvoxamine also increased  $\sigma_1 R$ . In another *in vitro* model, MT40 cells expressing high levels of  $\sigma_1 R$ , NGF was found to be more potent in inducing neurite sprouting, whereas treatments with  $\sigma_1 R$  antisense DNA significantly reduced the degree of neurite sprouting (176,177,185). Together, these data suggest a primary role for  $\sigma_1 R$  ligands in enhancing NGF-induced neurite growth.

One member of the neurotrophin family, brain derived neurotrophic factor [BDNF], has been heavily implicated in the actions of antidepressants, and perhaps  $\sigma R$  agonists, since chronic treatments with a variety of antidepressant therapies induce an increase in BDNF expression (795–798). Moreover, BDNF administration itself has been shown to produce antidepressant effects in behavioral models of depression (799–801).

The effects of  $\sigma_1 R$  ligands on BDNF expression need to be defined further. Thus far, this has only been studied with the  $\sigma R$  ligand E-5842, which showed no effects on BDNF or NGF levels following chronic treatments (802). However, E-5842 presents an  $\sigma R$  antagonist profile, so its lack of efficacy cannot be considered as an indication of what might be the effects generated by  $\sigma_1 R$  agonists. It is still conceivable that  $\sigma_1 R$  agonists would potentiate the effects of BDNF, similarly to that observed previously with NGF.

Another growth factor of importance is epithelial growth factor [EGF] (803). EGF is present in the CNS and known to stimulate cell proliferation in PC12 cells. A recent report indicated that in PC12 cells, the overexpression of  $\sigma_1 R$ induces a three-fold increase in neurite sprouting. This effect is suppressed by the  $\sigma_1 R$  antagonist NE-100 (176,177). The overexpression of the  $\sigma_1 R$  in squamous cell carcinomas has shown a strong positive correlation with tumor node metastases [TNM], indicating a potential prognostic tool based on pathology assessment and  $\sigma_1 R$  expression (804). In the context of this review, these data are even more interesting if one considers that EGF has been shown to enhance NMDA-induced modulation of intracellular Ca<sup>2+</sup>.

More research will be required to elucidate the exact basis for the observed potentiation of neurotrophic effects by  $\sigma R$ agonists and whether  $\sigma R$  ligands always affect neuronal survival and neurogenesis (805).

#### Depression, antidepressants and stress

There are a number of possible mechanisms of action for  $\sigma R$  ligands to act as antidepressants, including  $\sigma R$ , Glu, 5-HT neurotransmission and Ca<sup>2+</sup> regulation (26,806).  $\sigma_1 R$  ligands may present a novel mechanism of antidepressant action with

potential for a faster onset of action than classical antidepressants (805,807) and SSRI drugs (55,808).

Depression often coexists with cardiovascular diseases, such as hypertension and heart failure, in which sympathetic hyperactivation is critically involved. Reduced  $\sigma_1 R$  brain function in depression decreases heart rate via neuronal activity modulation (809). Reduced brain  $\sigma_1 R$  exacerbates heart failure, especially when combined with pressure overload via sympathetic hyperactivation and worsening depression (810).

The first interest in  $\sigma R$  ligands as antidepressants originated from the observation that the antidepressants fluvoxamine (811,812), fluoxetine (813), citalopram, sertraline, clorgyline and imipramine all possess moderate to high affinity (Ki 36–343 nM) for  $\sigma_1 R$  sites (655,659,660,814). Antidepressant treatments, or other modifications of the 5-HT system, induce changes in  $\sigma R$  binding properties. For example, repeated treatments with the TCA imipramine (14 days) causes a decrease in the total number of  $\sigma_1 R$  binding sites without affecting the affinity of [<sup>3</sup>H]DTG binding to  $\sigma R$ sites in the striatum, hippocampus and cortex of the rat (800,815). Therefore, certain differences in the clinical effects of various antidepressants may, in part, be explained by their distinct influence on cerebral  $\sigma Rs$  (800,815).

More direct evidence of the potential antidepressant properties of  $\sigma R$  ligands was obtained from behavioral experiments. SA 4503 (359), (+)- PTZ, DTG, JO-1784 and SKF-10,047 agonists decrease in a dose dependent fashion the immobility in the FST, whereas the  $\sigma R$  antagonists NE-100 and BD1047 blocked these effects (181,208,816). In addition, SA 4503 and (+)-PTZ also decreased immobility time in the Tail Suspension Test [TST], an effect also antagonized by NE-100 (817). Interestingly the antidepressant-like effect of SA 4503 in the FST, a test of a rodents behavioral response to the threat of drowning, was potentiated by the non-competitive NMDA antagonist amantadine (129).

OPC-14523, a combined  $\sigma_1 R$  and 5-HT<sub>1A</sub> receptor ligand (323), decreases immobility time where the effect of OPC-14523 can be enhanced by its daily administration for 7 days using the FST as a behavioral biomarker (324). Both the  $\sigma_1 R$  antagonist NE-100 and the selective 5-HT<sub>1A</sub> antagonist, WAY 100635 (818) antagonized the behavioral effects of a single dose of OPC-14523 in the FST (324).

Moreover, a one-week pretreatment with *para*(4)chloroamphetamine [p-CPA] depletion of brain 5-HT, failed to diminish the antidepressant effects of OPC-14523 in the FST (819), suggesting that  $\sigma$ Rs alone can mediate the antidepressant effects produced by OPC-14523 and that the combination of the  $\sigma$ R and 5-HT<sub>1A</sub>-receptor activity could induce a more potent or rapid "antidepressant-like" effect.

In keeping with this hypothesis, a potentiation of the "antidepressant-like" effects in the rodent FST has been observed following the combined administration of  $\sigma R$  and 5-HT<sub>1A</sub>-receptor agonists compared with their separate administration (820). In the chronic mild stress behavioral [CMS] model (chronic stress is believed to be involved in the etiology of affective psychiatric disorders), the  $\sigma R$  ligands SKF-10,047 (821) and DEX reversed the motor suppression induced by stress (767,805).

Most of the data regarding  $\sigma R$  and depression have focused on the  $\sigma_1 R$ ; however, the  $\sigma_2 R$  ligand Lu 28-179 also has shown "antidepressant like" activity in the CMS model of depression. Specifically, three-week treatments with antidepressants led to a normalized sucrose intake in rats, which reversed the decreased intake caused by the stress. Lu 28-179 did not affect sucrose intake in non-stressed controls, but produced a significant increase in sucrose intake in rats exposed to CMS (822,823). However, even if Lu 28-179 has a higher affinity for  $\sigma_2 R$ , it also has affinity for  $\sigma_1 R$  (822); therefore, a role of the  $\sigma_1 R$  in these "antidepressant-like" effects of Lu 28-179 cannot be excluded.

In animal models, neurosteroids with affinity for  $\sigma$ Rs have also been shown to exert "antidepressant-like" effects that are dependent on the endogenous neurosteroidal systems. For example, the effect of JO-1784  $\sigma$ R agonist on the FST was enhanced in ADX/CX mice compared to control animals, whereas another  $\sigma$ R agonist, PRE-084 (127), demonstrated a significant antidepressant effect only in ADX/CX mice (181); however, this effect has been reported more recently in C57BL/6J and to a lesser degree in Albino Swiss mice (824).

The  $\sigma_1$ R-antagonist BD 10047 (208) blocked all these effects (181). Furthermore, treatments with finasteride, which lead to the accumulation of PROG, also blocked  $\sigma_1$ Rmediated antidepressant effects. Thus, as discussed previously, circulating steroids appear to exert a tonic modulatory effect on the  $\sigma_1$ R and therefore on  $\sigma_1$ R-mediated "antidepressant-like" effects (181). It follows that the potency of  $\sigma_1$ R agonists as antidepressants is highly dependent on the endogenous PROG levels. Depressed patients such as the elderly with decreased levels of neurosteroids, which would be tonically inhibiting  $\sigma$ R to a lesser degree, might be particularly sensitive to such treatments (181).

Only a few controlled clinical trial data are available regarding the effect of  $\sigma R$  ligands in depressed patients (825). The results from a double-blind placebo controlled study, obtained from an interim analysis, showed that a dose of 20 mg/day of JO-1784 was superior to placebo and to 20 mg/day of fluoxetine. However, at 100 mg/day, JO-1784 was not different from the placebo (206), which is in keeping with the dose response curves mentioned above (206). A phase II study of SA4503 (cutamesine) in patients with major depression is currently underway (121).

Therefore, even if very limited, the clinical data support the hypothesis that  $\sigma R$  agonists could be effective antidepressant medications. However, the mechanisms of action through which  $\sigma R$  ligands could exert their antidepressant effects have not been clearly identified. Recent work points to the involvement of the executive function of the PFC (826). As attention deficit disorder [ADD] responds to the stimulant methylphenidate operating via the  $\sigma_1 R$  (827), investigation of these stimulant effects might help elucidate the mechanism of action of  $\sigma R$  ligands in depression.

Attempts to identify the mechanisms by which  $\sigma R$  ligands exert their effects have brought to light the role of  $\sigma Rs$  in the regulation of Ca<sup>2+</sup> (153,161,164,165,376) or K<sup>+</sup> signaling (88,133,138,139). The effects of JO-1784 in the FST were demonstrated to be Ca<sup>2+</sup>-dependent, since the extracellular Ca<sup>2+</sup> chelator ethylenediamine tetraacetic acid [EDTA] prevented the effect of JO-1784 in a dose-dependent manner. In addition, a lower dose of JO-1784 had no effect by itself, but co-administered with the L-type voltagedependent  $Ca^{2+}$  channel [VDCC] positive modulator (–)-Bay K8644, it significantly reduced immobility time in the TST.

In agreement with the hypothesis that  $\sigma Rs$  affect Ca<sup>2+</sup> regulation, the L-type VDCC antagonist, verapamil and the *N*-type VDCC antagonist,  $\alpha$ -conotoxin, blocks the effects of JO-1784 (181,249–251). Therefore,  $\sigma_1 R$  may be interacting with pre- or postsynaptic VDCCs to exert antidepressant-like effects in the FST (181).

Bradykinin, which increases IP<sub>3</sub> levels, enhances the effect of JO-1784 (249), whereas the IP<sub>3</sub>R antagonist, xestospongin C, blocks the effect of JO-1784. Thus the mobilization of intracellular Ca<sup>2+</sup> from IP<sub>3</sub>R-sensitive pools appears to participate initially in the behavioral effects mediated by  $\sigma_1$ Rs located on the ER membranes (249). The  $\sigma_1$ R then putatively moves to the plasma membrane and interacts with the VDCCs (154,161).

It is likely that  $\sigma R$  ligands' ability to modulate both Glu and 5-HT transmissions also contribute to the antidepressantlike effect observed in behavioral models. The molecular mechanism underlying  $\sigma Rs'$  ability to modulate 5-HT and Glu-ergic transmissions may involve  $\sigma Rs'$  ability to modulate Ca<sup>2+</sup>. This modulation could represent a secondary target involved in the effects of  $\sigma R$  ligands on both the Glu and the 5-HT systems, thus leading to one primary target (377,828). Recently, practical treatment efforts have shown that  $\sigma 1Rs$  are also one of the major pharmacological therapeutic targets of selective serotonin reuptake inhibitors [SSRIs] (829).

#### Schizophrenia and psychosis

Schizophrenia is one of the most devastating diseases for both the affected patient and those close to him or her. In the search for medications the antipsychotic effect of stimulation  $\sigma_1 Rs$ has been investigated (830). Experimentation with a number of typical and atypical antipsychotics has been investigated, but the SIGMAR1 gene ( $\sigma_1 R$  gene) does not confer susceptibility to schizophrenia (831). Interestingly,  $\sigma_1 R$  polymorphism is associated with an increased risk of schizophrenia and differential activation of the PFC (344) and the severity of AD (44).

The effect of chronic administration of the atypical antipsychotic E-5842, a preferential  $\sigma_1 R$  ligand, on iGluR subunit levels of mRNA and protein reveals differentially regulated levels of the NMDA<sub>2A</sub> and of GluR<sub>2</sub> subunits in a regionally specific manner. Concentrations of immunoreactivity for the NMDA<sub>2A</sub> subunit are unregulated in the medial PFC, the frontoparietal cortex, the cingulate cortex and in the dorsal striatum, while they are down regulated in the nucleus accumbens. Concentrations of the GluR<sub>2</sub> subunit of the AMPAR are up regulated in the medial PFC and the *nucleus accumbens* and down-regulation is observed in the dorsolateral striatum, indicating that E-5842 is able to modify levels of several GluR subunits (15,380,802).

#### Psychosis

Psychosis occurs in 10% to 37.1% of patients with mood disorders (832,833). Psychotic depression is a clinical subtype

of major depressive disorder and is characterized by psychosis accompanied by greater severity of depressive symptoms that include psychomotor impairment (retardation or agitation), morbid cognition (involving guilt and a sense of deserving punishment), suicidal ideation and neuropsychological impairment (834,835). Psychotic depression has been shown to have poor prognosis when compared to nonpsychotic depression (i.e. higher rates of recurrence, greater incapacitation, more frequent hospitalization, longer episodes and greater mortality) (836–839). Although several reports suggest abnormalities of endocrine, DA-ergic and serotonergic systems in psychotic depression (840,841), pathophysiology of psychotic depression is still unclear.

Psychotic depression has traditionally been treated with electroconvulsive therapy and classical antipsychotics, such as respiridone (842), in conjunction with tricyclic antidepressants, such as desipramine (843), although tardive dyskinesia may occur following protracted exposure (844). More recent studies have demonstrated the efficacy of atypical antipsychotics and SSRIs in treating psychotic depression (845,846).

Interestingly, SSRI monotherapy, especially fluvoxamine (Luvox), has been shown effective against both the psychotic and depressive symptoms of this disorder (845,847–851). Based on these findings, it has been recently proposed that SSRIs might have multiple action sites in the brain, in addition to serotonin transporters: perhaps  $\sigma_1$ Rs might play a role in the therapeutic action of SSRIs (752).

Some studies have demonstrated the possible link of psychotic depression to dysregulation of neurotransmitters, such as DA and 5HT; abnormality of brain lipid ganglioside; or hyperactivation of the neuroendocrine and the DA system (840,852). In addition, some studies have suggested that the abnormality of cortisol responses to the dexamethasone suppression test is more prevalent in psychotic depression than in nonpsychotic depression (837,838,841); hence it is possible that psychotic symptoms in depression could be due to increased DA activity secondary to hypothalamic-pituitaryadrenal [HPA] axis over activity (853,854). The observation that psychotic depression frequently appears in patients with neuroendocrine diseases such as Cushings syndrome supports the involvement of the abnormalities of the endocrine system in psychotic depression (855–858).

As previously stated  $\sigma_1 R$  antagonists show antipsychotic effects *in vivo*. Although some  $\sigma_1 R$  antagonists have been shown to inhibit apomorphine- or amphetamine-induced behavioral alterations (859), other studies clearly show that selective  $\sigma_1 R$  antagonists more specifically inhibit the PCP-induced behaviors (156,771).

 $\sigma_1 R$  antagonists rimcazole and BMY-14802 have been tested in clinical trials of acute psychotic symptoms of schizophrenia, but the antipsychotic actions of these compounds have not been confirmed (860,861). The synthesized  $\sigma_1 R$  ligands SL82.0715 and EMD 57445 (panamesine) have been shown to improve negative symptoms in open clinical trials (862–864).

Fluvoxamine (Luvox), showing the utmost potent effectiveness in the treatment of psychotic depression, has the highest affinity for  $\sigma_1 R$  ( $K_i = 36 \text{ nM}$ ) among SSRIs (655). Indeed, the efficacy of SSRIs in psychotic depression appears to correlate better with their affinities for  $\sigma_1 R$  than with those for 5HT transporters (655,849,865). Chronic fluvoxamine exposure *in vitro* causes an up regulation of  $\sigma_1 R$  and potentiates the neuritogenesis in a  $\sigma_1 R$ -dependent, but 5HTindependent, manner (185). Studies have also demonstrated that chronic fluvoxamine increases, in a 5HT-independent manner, ALLO in rat brains and in the cerebrospinal fluid [CSF] of patients with depression (866,867). One study demonstrated a statistically significant correlation between symptomatology improvement and the increase in ALLO following fluoxetine (Prozac) or fluvoxamine treatment (867).

Selective  $\sigma_1 R$  ligands potently stimulate adrenocorticotropic hormone release after central or peripheral administrations in rats (868,869). Therefore, it is possible that one of the action sites of fluvoxamine may involve  $\sigma_1 R$  that regulate the neuroendocrine system in the brain (870).

## Seizures

Seizures associated with cocaine intoxication are a serious clinical problem requiring immediate and adequate treatment. The seizures appear to arise from the interaction of cocaine with GABAergic and Glu systems (871). Accordingly, pharmacological studies have demonstrated that GABA<sub>A</sub>R agonists and NMDAR antagonists can efficiently inhibit cocaine-induced seizures, whereas Na<sup>+</sup> and Ca<sup>2+</sup> channel blockers were ineffective (457). The likely interactions are extremely complex; hence, looking at one component in isolation could be misleading.

An involvement of 5-HT<sub>2</sub>, DA and  $\sigma$ Rs in cocaine-induced seizures has also been proposed (872). Some of these changes, such as expression of immediate early genes and increase in neuropeptide biosynthesis may play a compensatory anticonvulsive role; however, other alterations e.g. up-regulation of NMDARs may increase susceptibility to seizures (538). Stimulation of  $\sigma$ Rs down-regulate electro-acupuncture induced seizures (873). In fact, sigma receptor-mediated events may play some role in seizure processes in the central nervous system and can modulate the protective activity of some conventional antiepileptic drugs (874).

#### Pain

Although no specific  $\sigma R$  ligand has reached the market for the treatment of pain, different pharmacological approaches to the alleviation and treatment of pain have been investigated using  $\sigma_1 R$  agonists and antagonists (15,38,875–879), particularly regarding potential interaction with opioid analgesics and the effect on analgesia (302,880). Activation of  $\sigma_1 Rs$ antagonizes opioid analgesia (881,882), where antagonists potentiate opioid analgesia (681,883).  $\sigma_1 Rs$  differentially modulate acute vs. chronic pain (884-887) and possibly migraine headaches (888): they are also involved in visceral pain (889). In fact, the  $\sigma Rs$  have been proposed as a modulatory system influencing the analgesic activity of opioid drugs (39). The most promising effects of the  $\sigma Rs$ lie in the potentiation or modification of the action of other analgesics such as acetaminophen (890) and morphine (891). Such observations may provide a starting point for the development of novel analgesics.

Work has been carried out to identify the mechanism of action of  $\sigma$ Rs in pain (892). The findings that EAAs have actions on  $\sigma$ Rs indicated that the EAAs might act via the Glu system in the transmission of nociceptive information (483). In fact, NMDARs receptors play an important role in the potentiation of morphine antinociception (893), and it has been shown that activation of spinal  $\sigma_1$ R enhances NMDA-induced pain via PKC- and PKA-dependent phosphorylation of the NMDA receptor NR<sub>1</sub> subunit (458,894).

## Addiction

Many drugs of abuse, including cocaine and METH, produce effects that can be mitigated through  $\sigma Rs$ , particularly the  $\sigma_1 R$  subtype (872) including neurotoxicity (12); hence, it has been suggested that  $\sigma_1 Rs$  should be considered as potential compound for substance abuse (895). More specifically, agonists at  $\sigma_1 R$  and  $\sigma_2 R$  inhibit NMDA-stimulated DA release from motor and limbic areas of rat brain (896). Both cocaine and METH exhibit significant affinities for  $\sigma Rs$ , and about a 10- to 20-fold preference for the  $\sigma_1 R$  subtype (897,898). Because of these effects, it has been suggested that  $\sigma R$  antagonists are an obvious potential medications for the treatment of drug abuse (872), and CM156 has been shown to attenuate the neurotoxic effects of METH (13).

These interactions appear physiologically relevant because treatment of animals with selective  $\sigma R$  antagonists significantly attenuates cocaine-induced locomotor activity, conditioned place preference, behavioral sensitization, convulsions, lethality and changes in gene expression (897,899–902). The importance of the  $\sigma_1 R$  subtype is supported by the ability of antisense oligonucleotides against them to prevent a number of cocaine-induced behaviors including locomotor hyperactivity, conditioned place preference and convulsions (897,902).

Under normal conditions the brain maintains a delicate balance between inputs of reward seeking controlled by neurons having the D<sub>1</sub>-like family of dopamine receptors and inputs of aversion coming from neurons having the D<sub>2</sub>-like ones (663). Cocaine is able to subvert these balanced inputs by altering the cell signaling of these two pathways such that D<sub>1</sub> reward seeking pathway dominates. D<sub>2</sub> receptors (the long isoforms of the D<sub>2</sub> receptor) can complex with  $\sigma_1$ Rs, a result that is specific to D<sub>2</sub> receptors; thus, signaling via D<sub>2</sub> receptor containing neurons, destabilizes the delicate signaling balance influencing drug seeking that emanates from the D<sub>1</sub> and D<sub>2</sub> receptor containing neurons in the brain (663).

Antagonism of  $\sigma Rs$ , using either putative antagonists or antisense oligonucleotides, also reduces METH-induced locomotor activity and behavioral sensitization (754,898). In addition,  $\sigma R$  proteins levels become up regulated in the brains of rodents who self-administer or are repeatedly injected with METH (71,903). Despite the known interactions between  $\sigma Rs$ and psycho stimulants such as cocaine and METH, other than an early abstract reporting the binding of 3,4-methylenedioxymethamphetamine [MDMA] to  $\sigma Rs$ , no other studies to investigate this interaction have been conducted (14). METH and MDMA are structurally similar so the question of whether the interaction between  $\sigma R$  and MDMA is similar. Experimental evidence has now shown that indeed the interaction is similar (14).

 $\sigma_1$ Rs are critically involved in the rewarding effect of cocaine (900,904). Cocaines mechanism of action involves initial inhibition of neuronal monoamine transporters primarily in the DA reuptake systems located on mesolimbic neurons. Cocaine rapidly increases the DA neurotransmission and triggers adaptive changes in numerous neuronal circuits underlying reinforcement, reward, sensitization and the high addictive potential of cocaine (122,784).

There appears to be regional differences as to the upregulation of the  $\sigma_1 R$ . At present the major up-regulation has been recorded in the regions involved in addiction and reward (899). The observation that repeated administration of cocaine rapidly provokes over expression of the  $\sigma_1 R$ outlines its major role in these first psychological steps of addictive processes (905). Indeed, there is little question that the behavioral effects of cocaine can be related to the  $\sigma_1 R$  (897).

In utero cocaine [IUC] exposure results in offspring rats with complex neurochemical and behavioral alterations, particularly affecting learning and memory processes (871). However an investigation into the impact of IUC exposure on memory functions in male and female offspring rats revealed that the activation of the  $\sigma_1 R$  neuromodulatory receptor *in utero* allows a complete behavioral recovery of the memory functions in prenatally cocaine-exposed rats (779).

# Neurodegeneration

Seizure activity, by overstimulation of the  $\sigma R$  is mediated via iGlu, particularly NMDAR; this has been associated with neurodegenerative processes such as status epilepticus (906, 907), cerebral ischemia (908), perinatal asphyxia and traumatic brain injury (407).

Because the iGluRs are ion-gated channels selective to Na<sup>+</sup>, K<sup>+</sup> and Ca<sup>2+</sup>, any sustained stimulation of the GluRs results in osmotic damage due to the entry of excessive ions, in particular Ca<sup>2+</sup> and water. The entry of this material results in apoptosis and necrosis in neurons (382,384–386,394,412, 438,511).

This increase in the intracellular Ca<sup>2+</sup> concentration in neurons is crucial to the determinant of injury that occurs following activation of several enzyme pathways and signaling cascades including as phospholipases, PKC, proteases, protein phosphatases, nitric acid synthases, oxidative stress (423) and the generation of oxygen-based free radicals [ROS] (382,384,386,387,394,412,415,428,486,489,490,491,543,909, 910,911). Activation of the  $\sigma_1$ R inhibits glutamate-induced death of neuron by reducing ROS (912,913).

Neurons are not the only cell type in the nervous system to be damaged by high concentrations of Glu (914). Functional NMDAR recently have been reported in brain glia (915), astrocytes (247,916) and oligodendroglia (409,559,917,). Glial and neuronal NMDARs are functionally and structurally different from the neuronal NMDR; however, the structure of  $\sigma$ Rs in these cell types is not known at present, and it can only be speculated that the alteration of  $\sigma$ Rs in these cell types would ameliorate damage caused by overstimulation of NMDARs. Activation upon ischemia triggers Ca<sup>2+</sup>-dependent damage of oligodendrocytes and myelin (559,918), a finding that has implications for many central nervous disorders such as MS. Because of the association between  $\sigma R$  and Glu, it is likely that  $\sigma Rs$  are involved in exogenous and endogenous Glu toxicity in oligodendrocytes.

## Heart and vessels

 $\sigma$ Rs have been implicated in the regulation of the cardiovascular system [CV], and  $\sigma_1$ R transcripts have been found in parasympathetic intracardiac neurons (756) and the human ether-a-go-go-related gene [hERG] channel (919). These structures are central to cardiac excitation and rhythmic control (920–922).

Both  $\sigma_1 R$  and  $\sigma_2 R$  interact with the human ether-à-gogorelated gene (hERG). hERG encodes a cardiac channel that is also abnormally expressed in many primary human cancers, potentiating tumor progression through the modulation of extracellular matrix adhesive interactions.  $\sigma_1 R$  potentiates hERG current by stimulating channel subunit biosynthesis and  $\sigma_1 R$  silencing does not modify hERG mRNA contents but reduces hERG mature form densities. A physical association has been shown in HEK cells expressing hERG and  $\sigma_1 R$ : both proteins co-immunoprecipitate.  $\sigma_1 R$  expression enhances both channel protein maturation and stability (919).

In rabbits, all  $\sigma_2 R$  agonists have been shown to reduce phenylephrine-induced cardiac arrhythmias. They prolonged action potential duration in rabbit Purkinje fibers and reduced human ether-a-go-go-related gene (HERG) K(+) currents. It has been suggested that  $\sigma_2 R$ -receptor ligands block I(Kr) and that this effect could explain part of the antiarrhythmic properties of this ligands family. Nevertheless, an interaction with HERG channels not involving  $\sigma_2 R$  seems to share this pharmacological property. The repolarization prolongation and the early-after depolarization can be responsible for "torsades de pointe" and sudden cardiac death. It is for this reason that particular caution has to be taken using ligands with affinity for  $\sigma_2 R$  with respect to abnormal cardiac function (923,924).

The relationship between depression and heart failure is known, but the mechanism has not been fully elucidated. Depression is associated with a substantial increase in the risk of developing heart failure and is independently associated with increased cardiovascular morbidity and mortality. Reduced  $\sigma_1$ Rs density in depression decreases heart rate via the sympathetic stimulation in the autonomic nervous system [ANS] (809) and exacerbates heart failure, especially when combined with pressure overload and worsening depression (810). Conversely, cardiovascular disease can lead to severe depression. Thus, therapy with SSRIs used for treatment of depression, has been recommended to reduce cardiovascular disease morbidity and mortality (829,925).

Similarly, GluR have been found in cardiac intramural nerve fibers and ganglia cells as the main structures expressing GluRs in the conducting system (926) and similar findings have been seen in human hearts (500).

These effects of the  $\sigma_1 Rs$  are probably mediated via PKCand PKA-dependent phosphorylation of the NMDA receptor (458); however, the exact cellular function of  $\sigma_1 R$  in these cells remains to be determined. Regardless, a reduction of brain  $\sigma_1 Rs$  also contribute to sympathetic hyperactivation of the heart (810,927), probably via altered Na<sup>+</sup> channels (150,928,929). In the reverse, stimulation of brain  $\sigma_1 Rs$  ameliorates hypertrophy in mice (925) and cardiac function following myocardial infarction (927).

 $\sigma R$  ligands have been shown to modulate contractility,  $Ca^{2+}$  influx and cardiac rate *in vitro* (930,931), where  $\sigma R$  stimulation causes changes in beating frequencies, followed by irregular contractions. In this case, changes in Ca<sup>2+</sup> are not mediated by sarcoplasmic reticulum Ca<sup>2+</sup> transport systems (923,930).

Experimentally, pre-treatment with an  $\sigma R$  agonist improves the reperfusion recovery of cardiac pump function in rat hearts (932) and is cardioprotective (925,933). Activation of the cardiac  $\sigma R$  prompts an augmentation of tolerance to the reperfusion damage; however, this effect decreases with time, indicating a possible desensitization of the receptor (934).

In any case,  $\sigma R$  activation prevents reperfusion contracture, increases pressure in the left ventricle, and improves survival of cardiac myocytes after ischemia and reperfusion. Conversely, pre-treatment with an  $\sigma R$  antagonist augments the reperfusion systolic dysfunction of the myocardium and prevents post-ischemic contractures and cardiac cell lesions (932). Interestingly, the electrical stability in the rat model of post-infarction cardiac sclerosis and stress, activation of either  $\mu$ - or  $\kappa_1$ -opioid receptors or blockade of  $\sigma_1 R$  reverses the decrease in ventricular fibrillation threshold (935) increasing the probability of sudden death.

By contrast, L-Glu increases the frequency of Ca<sup>2+</sup> oscillations in cardiac excitation and rhythmic control (436), which has been positively correlated with increased contraction frequency in myocardial cells. Such an increase may reduce cardiac filling, hypoxia and angina-like chest pains (936,937). It would appear that in the case of the heart, GluR and  $\sigma$ Rs might have opposing actions.

Activation of  $\sigma R$  reversibly blocks the delay in outwardly rectifying K<sup>+</sup> channels, large conductance Ca<sup>2+</sup> sensitive K<sup>+</sup> channels and the M-current. This blockade is dose-dependent suggesting the effect is mediated by  $\sigma_1 R$  activation (930,931). Thus, activation of  $\sigma_1 R$  depresses the excitability of intracardiac neurons and is likely to block parasympathetic input to the heart.  $\sigma R$  stimulation has been shown to cause changes in beating frequencies, which are followed by irregular contractions (923), probably mediated through NMDARs. It also has been suggested that in the heart the signal transduction pathway does not involve a diffusible cytosolic second messenger or a G protein (158,756), a finding that is supported (207) and refuted by others (174). Therefore, the activation of  $\sigma_1 R$  is most likely mediated via iGluRs rather than mGluRs.

In addition to myocardial contraction modulation,  $\sigma Rs$  also are involved in the regulation of coronary and peripheral arterial vascular tension (923). Experiments have shown that the changes in Ca<sup>2+</sup> induced by  $\sigma_1 R$  stimulation are not mediated by sarcoplasmic reticulum Ca<sup>2+</sup> transport systems and do not affect the apparent sensitivity of the myofilaments to Ca<sup>2+</sup> (930). In fact,  $\sigma R$  agonists increase the intracellular

 $Ca^{2+}$  levels by stimulating IP3 production and, thus, modulate contractility (167).

# Muscle and bones

Many neuroleptic drugs reported to play a role in the control of movement bind with high affinity to  $\sigma_2 R$ . The high affinity of some neuroleptics for these sites suggests their possible involvement in some  $\sigma_2 R$ -mediated side effects, such as druginduced dystonia (938). A correlation between the clinical incidence of neuroleptic-induced acute dystonia and binding affinity of drugs at the  $\sigma R$ , indicate that the  $\sigma R$  might be involved in neuroleptic-induced acute dystonia, which has been confirmed by  $\sigma R$  agonist induced neck dystonia of rats (764).

As bone has been shown to express many of the molecules associated with Glu- mediated signaling (939–941), it is probably that  $\sigma$ Rs are involved in normal bone function as well as in disease states, although there is some debate regarding the role of Glu in controlling bone growth (942–944). Nonetheless, all osteoblasts (943,944), osteocytes and osteoclasts express one or more of the GluR subunits, including NMDARs (492,497,945–948). The Glu Asp transporter [GLAST] has also been identified in bone (492,949,950). As activation of  $\sigma$ Rs is an integral part of Glu system, it is likely that they act in conjunction with GluRs to affect cellular changes.

## Lung

Considerable data are available for the presence of  $\sigma$ Rs in lung tissue and a role for them with respect to cancer and chemotherapy (951). In fact,  $\sigma$ Rs are expressed in a wide variety of tumour cell lines (755,952,953), including nonsmall-cell lung carcinoma, large-cell-carcinoma (NCI-H1299 and NCI-H838), lung cancer cell line (NCI-H727) (755,953,954) and small-cell lung cancer (NCI-H209/N417) (955). More recently, in material obtained from patients with lung tumors elevated PROG receptor membrane component was seen associated with increased  $\sigma_2$ Rs levels in the tumor mass and blood plasma (956).

The anatomical sites of  $\sigma Rs$  in normal lung are likely to be associated with pulmonary nerves (951). However, expression of the  $\sigma R$  has been used to visualize cancerous cells in the lung (86,108,676).

The presence  $\sigma Rs$  in the airway structures such as the larynx, esophagus and mast cells also implicate the GluRs (and probably the  $\sigma Rs$ ) in the mediation of asthmatic episodes (516,957,958). Thus the excitation of GluRs in the air passages may be important in airway inflammation (959) and hyper reactivity observed in bronchial asthma (440,960). Their presence also could explain the enhancement of acute asthmatic attacks by Glu-containing foods (957).

Current antitussive medications have limited efficacy and often contain the opiate-like agent DEX, which is an  $\sigma R$  agonist, or antagonist, depending on the dose administered (194,733). The mechanism whereby DEX inhibits cough is ill defined; however, DEX displays affinity at both NMDARs and  $\sigma Rs$ , suggesting that the antitussive activity may involve central or peripheral activity at either of these

receptors. Experimental findings in guinea pigs support the argument that antitussive effects of DEX may be mediated via  $\sigma R$ , since both systemic and aerosol administration of  $\sigma_1 R$  agonists experimentally inhibit citric-acid-induced cough (961).

## **Endocrine system**

Visualization using autoradiography with  $\sigma R$  radioligands has revealed these receptors in the rat pituitary, adrenal, testis and ovary (676). The highest density of  $\sigma Rs$  is present in the ovary, with progressively lower densities present in the testis, pituitary, adrenal and cerebellum, respectively (676). This distribution is not surprising given that studies have found that PROG and DHEAS bind to  $\sigma_1 Rs$  (183,228–230).

 $\sigma$ Rs are believed to be responsible for important regulatory functions in the endocrine system (222,962,963). However, the role of  $\sigma$ Rs in endocrine cells remains unclear, particularly given the plethora of possible neurotransmitter interactions in the HPA (964). It has been suggested that endogenous  $\sigma$ R ligand(s) would contribute, together with other endocrine factors such as DA, neuropeptide Y, or GABA, to the control of pituitary functions (868).

Because steroids have been shown to interact with  $\sigma_1 R$  (123,210,222) and because they exhibit a significant physiological relevance in the modulation of the electrical activity of frog melanotrope cells (965,966), it can be hypothesized that they represent a very interesting class of endogenous  $\sigma R$  modulators in endocrine cells.

Because of the many effects of the endocrine system are related to homeostasis, it is not surprising to note that manipulation of the  $\sigma$ Rs has numerous potential effects (967). For example, long-term administration of neuroleptic agents, such as haloperidol, has been associated with the development of a drug induced syndrome of inappropriate antidiuretic hormone release, which occurs in the absence of other abnormalities in endocrine function (968). Furthermore, it has now been hypothesized that interaction with some neuroleptic agents and the posterior pituitary  $\sigma$ R ligands can inhibit K<sup>+</sup>-channel function (138,139).

There are minimal data concerning role of  $\sigma$ Rs and the involvement of GluRs in diabetes mellitus and associated dysfunctional islet cells (418,435,486–488,512,523,524,669, 969–973), and abnormalities of HPA function (507,974,975). It is likely that  $\sigma$ Rs also play a role in these related disorders.

#### Reproduction

 $\sigma_1$ Rs are expressed in the placenta (976), in spermatozoa (977) and other parts of the reproductive system. As stated previously, the highest density of  $\sigma$ Rs is present in the ovary, with lower densities present in the testis (676). In the ovary,  $\sigma$ Rs are seen in highest density in the maturing follicles, and lower densities in resting follicles. In the testis, they are present in highest concentrations in the ductuli efferentes and ductus epididymis. Lower densities of binding sites are present in the seminiferous tubules, but none in the interstitial tissue (676). This pattern is mirrored by the distribution of GluRs (978). In addition,  $\sigma$ Rs (977) and GluRs (418) are abundant in spermatozoa and may affect they signaling

pathways (977) in conjunction with PROG or prostaglandin  $E_1$  [PGE<sub>1</sub>].

The developing fetus may be indirectly affected by PROG levels, which have been shown to decrease brain  $\sigma R$  function (179). Here PROG acts as an antagonist ligand for the  $\sigma R$  during pregnancy. At parturition, Glu output from the fetal liver reduces, leading to a fall in fetal arterial Glu concentrations, which correlate with a marked decrease in PROG output from the pregnant uterus (979), with probable up regulation of  $\sigma Rs$  (179).

Modulation of ion channels in *Xenopus* oocytes was observed in the presence or absence of  $\sigma_1 R$  ligands, suggesting that the  $\sigma_1 R$  may form a functional complex with the expressed ion channels (88). In fact, these authors went on to show that the  $\sigma_2 R$  forms an immunoprecipitating complex with ion channels both in rat neurohypophysis and when co expressed in *Xenopus* oocytes (88).

## Liver and kidney

Liver contains high densities of  $\sigma_1 R$  and  $\sigma_2 R$  (92,980), and these receptors are specifically localized to lipid rafts in rat liver phospholipid membranes (981,982), particularly mitochondria (113,983).

iGluRs and mGluRs also have been demonstrated in the liver (502,520) and mGluRs are involved in the hydrolysis of IP3 and reduction of viable hepatocytes (984). In fact, it has been suggested that GluR is activated by the Glu present in the portal blood and may contribute to toxic liver damage. At present the relationship of  $\sigma$ Rs and liver disease is still to be elucidated. As a number of  $\sigma_1$ R agonists are being developed for cancer treatment, especially those with EGFR activity, and as the liver is endodermal in origin, it is likely that a lot more findings will result from further hepatic cancer research (981).

Similarly, the kidney contains high densities of  $\sigma_1 R$  and  $\sigma_2 R$  as determined by using selective  $\sigma R$  probes and photo affinity labeling (92). Interestingly, this work, using kidney tissue *in vitro* shows that the 25 and 21.5 kDa proteins represent  $\sigma_1 R$  and  $\sigma_2 Rs$ , respectively. The role of these  $\sigma Rs$  in renal disease has yet to be determined.

#### Eye

Loss of retinal ganglion cells [RGC] is a hallmark of many ophthalmic diseases including glaucoma, diabetes retinopathy, retinal ischemia due to central artery occlusion, anterior ischemic optic neuropathy and may be significant in optic neuritis, optic nerve trauma and AIDS (985). The expression of  $\sigma_1$ R mRNA in the mammalian retina is greatest in ganglion cells (986), as determined via mRNA expression, cells of the inner nuclear layer, inner segments of the photoreceptor cells and retinal pigment epithelial cells (987). As Glu toxicity is seen mainly in the ganglion cells, a possibility of neuroprotection by  $\sigma$ R ligands against ganglion cell Glu toxicity has been suggested (987). In fact, the  $\sigma$ R ligand (+)-PTZ prevents Glu-induced apoptosis in retinal ganglion cells (988).

Expression, subcellular localization and regulation of  $\sigma R$  experiments have been undertaken in retinal Mueller cells (989). Mueller cells express  $\sigma_1 R$  and demonstrate robust  $\sigma_1 R$  binding activity, which is inhibited by  $\sigma_1 R$  ligands and is stimulated during oxidative stress (913,990). A similar

response is seen for  $\sigma_2 Rs$  (991). Additionally, late-onset inner retinal dysfunction in mice lacking  $\sigma_1 R$  has been reported, confirming the importance of  $\sigma_1 R$  in retinal health (992,993). In adult Mueller cells,  $\sigma_1 Rs$  are bound and stimulated under the conditions of oxidative stress, an effect that is amplified when cells were incubated with NO and reactive oxygen species [ROS] (989).

Exposure of lens cells to  $\sigma R$  antagonists has been shown to lead to growth inhibition and pigment granule production (994,995), implying that  $\sigma Rs$  are important during lens development.

## Gastrointestinal system

 $\sigma R$  binding sites have been shown to be present in the myenteric plexus of the guinea pig ileum (747) and are important in the regulation of ileal contractions (314), as are the GluRs (64,390,513,518,996,971,997). As Glu and Asp are both involved in regulating acid secretion in the stomach (998,999), it is likely that  $\sigma Rs$  are also involved.

DTG and its  $\sigma$ R-active congeners inhibit electrically or 5-HT-evoked contractions of the longitudinal muscle and myenteric plexus [LMMP] preparation by a neuronal mechanism (314), and as such  $\sigma$ R agonists might be possible novel targets for antisecretory therapy in diarrhea (1000). In fact, the importance of  $\sigma$ R manipulation in a number of diseases is highlighted by the recent patent applications (1001), where a method of stimulation of salivary secretion using oral administration of certain  $\sigma$ R ligands which may be generally described as N,N-disubstituted phenylalkylamine (US Patent 5387614).

 $\sigma$ Rs induce emesis in a number of species, probably mediated centrally via the vagus as has been shown for Glu, GluR and GLUTs (1002–1004). It is not surprising to note that emesis and nausea are often associated with the use of  $\sigma$ R agonists and antagonists (1005).

Unfortunately, nausea is a difficult endpoint to measure in animal studies; hence, most endpoints used with respect to the gastrointestinal system have been limited to vomiting. Nonetheless, symptoms reported by human subjects include nausea following chemical manipulation of  $\sigma R$  *in vivo*, but their quality of life scores improved (1006). Nausea and vomiting is probably a common endpoint for EAA poisoning mediated via the Glu, and most likely,  $\sigma R$  systems in such poisonings, such as is seen in DomA toxicity (415,418,434, 437,439,442).

 $\sigma$ Rs stimulate physiological motility and inhibit experimentally induced colonic hypermotility. They stimulate the postprandial colonic motility in dogs by acting selectively on sigma receptors located peripherally and probably by affecting the release of cholecystokinin octapeptide through a central adrenergic mechanism (1007). Other findings indicate that  $\sigma$ R ligand igmesine, blocks the corticotropin releasing factor and emotional stress-induced colonic hypermotility also via an interaction with central cholecystokinin octapaptide mechanisms (1008,1009).

## Immune system

Pharmacological studies initially identified high-affinity  $\sigma Rs$  on human peripheral blood mononuclear cells using DTG and

haloperidol (677). Subsequent studies employing the  $\sigma R$  selective radioligand, [<sup>3</sup>H] (+)-PTZ (600) then identified a lower affinity-binding site on murine B- and T-enriched lymphocytes (1010), human and rat lymphocytes (1011). Here, high concentrations of PCP (gM) compete for binding to  $\sigma Rs$  on the lymphocytes (98).  $\sigma_2 Rs$  also inhibit T lymphocyte activation (1012).

Evaluation of the effects of  $\sigma R$  and the immune system has helped solidify the understanding of the link between the endocrine, nervous and immune systems, although there is still an enormous amount of work required to sort out this relationship (222).  $\sigma R$  ligands have potent immunoregulatory properties, including the induction of I L-10 (1013) and the suppression of IFN- $\gamma$  and granulocyte colony stimulating factor [GM-CSF] (1014).

In murine studies, treatment with  $\sigma R$  ligands prevents both graft versus host reactions and delayed-type hypersensitivity granuloma formation (1014). These studies indicate that  $\sigma R$ dependent signaling plays a role in immune-mediated responses. Cocaine, a  $\sigma_1 R$  ligand, is also known to modulate immune function *in vivo* and *in vitro* (1015,1016).  $\sigma_1 R$  has been shown to regulate early steps of viral RNA replication at the onset of hepatitis C virus infection (1017) and a reovirus nonstructural protein  $\sigma_1 R$  is required for establishment of viremia and systemic dissemination (1018).

Because immunocompetent animal models of tumorigenicity and tumor progression can serve as sensitive indicators of immune dysfunction, it has been found that  $\sigma R$  ligands do impact host antitumor immunity, probably through a  $\sigma R$ dependent cytokine modulation (1013). Sigma ligands, especially  $\sigma_2 R$  agonists, can inhibit proliferation and induce apoptosis by a mechanism involving changes in cytosolic Ca<sup>2+</sup>, ceramide and sphingolipid concentrations (1019).

Specific cell types that assist in immunoregulation have been investigated  $\sigma R$  activity. Studies initially identified high-affinity  $\sigma Rs$  on human peripheral blood mononuclear cells using DTG and haloperidol (677). Subsequent studies employing the  $\sigma R$  selective radioligand, [<sup>3</sup>H] (+)-PTZ (600) have identified a lower affinity-binding site on murine B- and T-enriched lymphocytes (1010).

Previous work has shown high that concentrations (pM) of PCP suppressed lymphocyte proliferation, mitogen-induced IgG and IgM production, and LPS-induced IL-1 production (1020). Another report indicates pM concentrations of PCP and PCP analogues inhibit IL-2 production by concanavalin A [Con A]-stimulated murine splenocytes (1021). Similarly, DTG, haloperidol, (+)-PTZ and (–)-PTZ have been shown to enhance LPS-stimulated murine splenocyte proliferation while PCP was without effect (1010). Lymphocytes do not possess PCP-selective receptors as determined in radio receptor assays using the PCP-selective ligand, [<sup>3</sup>H]N-[1-(2-thienyl)cyclohexyl]-piperidine (677), but a high concentration of PCP (gM) competes for binding to  $\sigma$ Rs on splenocytes (98).

Regardless of the literature available, it is possible that current research into the effects of manipulation of  $\sigma Rs$  is not widely known to the public due to proprietary efforts to develop new treatment regimens using stimulation or antagonism of  $\sigma Rs$  on the immune system and the cells thereof. Due to the many body systems, cell types and substances involved in immunoregulation, tissues that also contain  $\sigma Rs$ , the manipulation of  $\sigma Rs$  holds promise for increasing our understanding of immune mediated diseases, cancer and "difficult" infections in which immune dysregulation is an essential part of the pathogenesis of the disease, e.g. HIV and AIDS.

# Neoplasia

Pharmaceutical agents acting at the  $\sigma R$  have been used in the treatment of cancer and are receiving considerable attention (1022). A large number of drugs are known to bind with high affinity to  $\sigma_2 Rs$  and these receptors are overexpressed in many cancer tissues, suggesting potential applications for  $\sigma R$  ligands in cancer diagnosis and therapy (1023). The potential and specific signal transduction pathways and mechanisms involved in the actions of  $\sigma R$  ligands in cancer biology include modulations of the plasma membrane and lipid raft components, intracellular Ca<sup>2+</sup> levels, cytoskeletal protein functions and ER stress (1022).

# Expression of $\sigma Rs$ in neoplastic cell lines and tissues

Both  $\sigma R$  subtypes,  $\sigma_1 R$  and  $\sigma_1 R$ , are highly expressed in tumor cell lines from various human cancer tissues, including, but not limited to, small- and non-small-cell lung carcinoma (755,953–955), large-cell carcinoma (954), renal carcinoma (952), colon carcinoma (952), sarcoma (952), brain tumors (1024), breast cancer (103,755,953), melanoma (755,953), glioblastoma (755,953), neuroblastoma (755,953) and prostate cancer (755,953).

Comparable findings available from rat cancer cell lines, such as C6 glioma (755), N1E-115 neuroblastoma (94,953) and NG108–15 neuroblastoma X glioma hybrid (755), which generally agree with the human data. Many of these observations are based on the binding of labeled  $\sigma R$  ligands that are  $\sigma_1 R$ - or  $\sigma_2 R$ - non-specific. In some cases,  $\sigma_1 R$  sites are masked with DEX so as to determine the relative amounts of  $\sigma_1 R$  and  $\sigma_2 R$  sites in the cell preparations. However, these results await confirmation by Western blotting and reverse transcription PCR [RT-PCR] studies (Table 6).

A comparative study on mouse mammary adenocarcinoma revealed that proliferative cells possessed 10 times more  $\sigma_2 R$ than did quiescent cells (1027); hence, the development of pharmaceuticals to block these receptors is a field of endeavor. The density of  $\sigma R_2$  sites have been evaluated after the stimulation of mitosis and progression through the cell cycle in the human mammary tumor cell lines T47D and MCF-7 as well as in the prostate tumor cell line DU-145 (1025). The results suggest that there is a direct correlation between the binding of the  $\sigma R$  drug [*N*-[1 $\alpha$ (2-piperidinyl)ethyl]-4-[I<sup>125</sup>]iodobenzamide [<sup>125</sup>I-PAB], moderately selective for  $\sigma_1 Rs$  and proliferative status; and an up-regulation of  $\sigma R$  binding sites occurred before mitosis.

Using *N*-[2-(1'-piperidinyl)- ethyl]-3-123I-iodo-4 methoxybenzamide, also moderately selective for  $\sigma_1 Rs$ , another study also found that  $\sigma_1 Rs$  and  $\sigma_2 Rs$  were present at high density on human breast tumor biopsies but virtually absent in normal tissues (980). Expression of the  $\sigma_1 R$ , monitored immunocytochemically, has been suggested as a possible marker for predicting the aggressiveness of breast tumors, in particular, where there was a significant correlation between

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Table 6.  $\sigma R$  drug binding in tumor tissues and cell lines.

Cell line or tumor tissue	$\sigma R$ ligands tested	Reference
Non-small-cell lung carcinoma	IPAB, haloperidol, DTG	(954)
Large-cell-carcinoma (NCI-H1299 and NCI-H838)	IPAB, haloperidol, DTG	(954)
Lung cancer cell line (NCI-H727)	IPAB, haloperidol, PTZ, DTG, (+/-) dextrallorphan	(755,953)
Breast ductal carcinoma (T47D)	PTZ, DTG, $(+/-)$ dextrallorphan	(755,953)
Renal carcinoma	DTG	(952)
Colon carcinoma	DTG	(952)
Sarcoma	DTG	(952)
Brain tumor tissue	DTG	(1024)
(Nude mouse) neuroblastoma and glioma	DTG	(1024)
Rat neuroblastoma(NIE-115), rat glioma (c6)	PTZ, DTG, $(+/-)$ dextrallorphan	(755,953)
U-138MG glioblastomas	PTZ, DTG, $(+/-)$ dextrallorphan	(755,953)
Breast cancer cell line (MCF-7; T47D; SKBr3)	Haloperidol, CB-64D, CB-184, IPAB	(103,1025)
Small-cell lung cancer (NCI-H209/N417)	IBP, haloperidol	(955)
Neuroblastoma [BE(2)]; SK-N-SH)	PTZ, DTG, $(+/-)$ dextrallorphan	(755,953,1026)
Prostate tumor cells (DU-145) (LnCap)	IPAB, PTZ, DTG, (+/-) dextrallorphan	(755,953,1025)
Mammary adenocarcinoma (line 66)	DTG	(1027)
Melanoma (A375)	PTZ, DTG, $(+/-)$ dextrallorphan	(955)
C6 glioma cells	<sup>11</sup> C-SA4503	(1028)

 $\sigma_1 R$  expression and PROG receptor status (1028,1029). The age-related decrease in PROG may be important in the binding of  $\sigma_1 R$  agonists in tumor cells (759).

#### $\sigma R$ ligands as tumor imaging agents

The high densities of  $\sigma_1 R$  and  $\sigma_2 R$  binding sites in tumor cell lines and tissues are indicative of their involvement in the cellular pathophysiology of cancer, and as such could have diagnostic potential in tumor imaging. In fact, previous work has developed probes for imaging  $\sigma_2 R$  both *in vitro* and *in vivo* (1030). Most of what is known about  $\sigma_2 R$  has been obtained using either radiolabeled or fluorescent probes, or biochemical analysis of the effect of  $\sigma_2 R$  selective ligands on cells growing under tissue culture conditions. Now it has been shown that the PGRMC1 protein complex is the putative  $\sigma_2 R$ binding site (31).

Numerous nonclinical studies have evaluated the usefulness of radiolabeled  $\sigma R$  ligands (1031,1032), as tumor imaging agents in melanoma (734,1033-1036,), breast cancer (954,980,1030,1032,1037-1039), prostate cancer (954,1038,1040) and non-small-cell lung cancer in mouse tumor models (1035). These observations suggested that  $\sigma_2 R$ ligands could be effective for tumor imaging, including radiotracers (1041), coupled with techniques such as positron emission tomography [PET] (757) or single-photon emission computerized tomography [SPECT] (737,1025,1027,1031, 1040) and two-photon confocal microscopy (1042). Recently, development of these tracers ( $\sigma_2 R$  ligands) has allowed differentiation of tumors from inflammation (1043), especially T cell lymphocytes (1012), or mast cells (1044). Most of these  $\sigma R$  ligands are nonselective for the  $\sigma_1 Rs$  and  $\sigma_2 Rs$ , but, more recently,  $\sigma_2 R$ -selective agents have shown the most promise in this regard (1037,1039).

#### *Physiology and pathophysiology of* $\sigma Rs$ *in neoplasia*

*Effects of*  $\sigma R$  *ligands on cancer cell proliferation and death.* Several studies have tested the potential effectiveness of  $\sigma R$  ligands on proliferation of tumor cells *in vitro*. The effects of various  $\sigma R$  ligands (e.g. haloperidol, DTG,

SKF10047, PTZ and Rimcazole) on the *in vitro* growth of human mammary adenocarcinoma, colon carcinomas and melanomas show promise (1045).

Cellular proliferation is inhibited, and cell detachment and rounding subsequent to cell death are observed by light microscopy. Of the  $\sigma R$  ligands tested, the  $\sigma_1 R$ - and  $\sigma_2 R$ nonspecific rimcazole, and reduced haloperidol, which is the main metabolite of haloperidol in humans (657), were the most potent inhibitors of cell proliferation (1045). Similar inhibitory effects of  $\sigma R$  ligands [e.g. *N*-[2-(piperidino) ethyl]-2-iodobenzamide [2-IBP], haloperidol and N-(2-piperidinoethyl)4-iodobenzamide [IPAB] were observed on smallcell lung cancer (NCI-H209 and NCI-N417) cells (955). IPAB or 2-IBP also inhibited the *in vivo* xenograft proliferation of NCI-N417 cells (955).

The question of the mechanism(s) underlying the inhibitory effect of  $\sigma R$  ligands on tumor cell proliferation is an important one. The morphological effect of treating C6 glioma cells with various  $\sigma R$  ligands (generally  $\sigma_2 R$ - and  $\sigma_2 R$ -nonspecific) has been examined (755,953). These compounds cause loss of cellular processes, assumption of spherical shape and cessation of cell division, and the time course and magnitude of these effects are dependent on the concentration of the various  $\sigma R$ ligands used. Continued exposure to  $\sigma R$  ligands for 3-24 h results in cell death, although the morphological effects are reversible if the drug is removed shortly after rounding (755,953). Reduced haloperidol also potently inhibited proliferation of WIDr colon and MCF-7 breast adenocarcinoma cell lines, where in these cells, the intracellular  $Ca^{2+}$  levels were raised, and apoptosis was observed (166), although a direct link between them has not been shown.

The ability of  $\sigma_2 R$  ligands to induce cell death in the human breast tumor cell lines MCF-7, MCF-7/Adr<sup>-</sup>, T47D and SKBr3 also has been demonstrated (103). Both  $\sigma_2 R$  subtype-specific and  $\sigma_2 R$  non-selective  $\sigma R$  ligands result in cell death by a mechanism that involves apoptosis. This has been suggested to be a novel p53- and caspase-independent apoptotic pathway (103).

The effects of  $\sigma R$  ligands on cell growth and apoptosis are thought to occur via the sphingolipid pathway. Therefore,

it is not surprising to note that  $\sigma_2 R$  ligands applied to MCF-7/ Adr<sup>-</sup> and T47D breast tumor cells induce a dose-dependent increase in ceramide and concomitant decreases in sphingomyelin (103).

Progress is being made in the development of potential treatment modalities. Nanoparticles have been extensively used as carriers to deliver molecules into tumors through the enhanced permeation and retention effect, and to regulate the release of a chemical or biological effector in response to environmental stimuli such as temperature or pH change. In these cases, cell uptake of nanoparticles has been studied to maximize their delivery into the target cells (1046). Recently, the surface of gold nanocages was functionalized with SV119, a synthetic small molecule specific to  $\sigma_2 R$ , and then was shown to be effective in for targeting cancer cells (1047).

Possible mechanisms of  $\sigma R$  signal transduction and relevance to cancer cell biology. Although there is considerable evidence for the involvement of  $\sigma Rs$  in cancer cell biology, the mechanism(s) through which these effects occur has not fully been discerned. As has been discussed previously,  $\sigma Rs$  have been implicated in a wide range of functions, and formulating a unifying hypothesis for the molecular physiology of  $\sigma Rs$  to account for all of the varied functions will be a great challenge. Few reports exist that deal directly with the mode of action of  $\sigma Rs$ .

The homology between the  $\sigma_2 R$  and the sterol isomerase, *ERG2*, of yeast is interesting, given that both the  $\sigma_1 R$  and the sterol isomerase have high affinity for  $\sigma_1 R$  ligands (1048). However, the  $\sigma_2 R$  has never been demonstrated to possess sterol isomerase activity. On the other hand, emopamilbinding protein, which also binds  $\sigma R$  ligands, was found to complement a yeast strain containing a deletion of the *ERG2* gene and is a sterol isomerase like ERG2 (147).

Modulation of ion channels. Ion channels are expressed in cell lines derived from several different cancer types and can play an important role in metastasis, an integral aspect of which is the control of cell growth and proliferation (1049–1052). The dual observation that  $\sigma R$  expression is increased in tumor cell lines or tissues, and that  $\sigma_1 Rs$  act as secondary subunits for some ion channels including Cl<sup>-</sup> channels (1053) might be of importance, given the accumulating evidence for the involvement of different types of ion channels in proliferation (1049,1050,1052) and metastatic activities of cancer cells (1050,1054-1056). Because down-regulation of K<sup>+</sup> channel amplitude has been associated with the metastatic phenotype in human prostate and breast cancer (1049,1057), such an effect could underlie the proposed association between cancer progression and  $\sigma R$  ligands.

In addition to roles such as proliferation (1057–1059), there are a number of ways in which ion channel activity may contribute to the cancer cell behavior, including migration (1060), apoptosis (1061), adhesion and cytoskeletal organization (1013,1062,1063) and secretion (1064). It remains to be determined whether ion channels, such as the voltage-gated Na<sup>+</sup> channel (1050,1064–1067) are also modulated by  $\sigma R$  ligands in the cancer process.

*Modulation of Ankyrin.*  $\sigma_1 R$  may play a role in controlling the functioning of cytoskeletal proteins (46,66,164). Using immunocytochemical techniques,  $\sigma_1 R$ , ankyrin B and IP3R have been co-localized in perinuclear areas and areas of cellto-cell communication. It has been proposed that this trimeric complex may regulate Ca<sup>2+</sup> signaling (164). Although the exact underlying molecular mechanism has not yet been described, it is well known that adhesion and cytoskeletal organization are important factors in cancer cell biology (1068,1069).

Modulation of Intracellular Ca<sup>2+</sup>. Evidence suggest that  $\sigma R$ in neuroblastoma cells may use Ca<sup>2+</sup> signals to produce cellular effects (95). By using  $\sigma R$ -inactive (but structurally similar) ligands,  $\sigma_2 R$ -selective agents such as CB-64D, and  $\sigma_1 R$ -selective agents have shown that a fast and transient release of Ca<sup>2+</sup> from the ER is induced specifically by the action of the  $\sigma_1 R$  and  $\sigma_2 Rs$  (1070). In turn, intracellular Ca<sup>2+</sup> modulation can affect PKC activity. Indeed, in rat brain synaptosomes, DA transporter activity is modulated by  $\sigma_2 R$ ligands via activation of PKC (354). Because intracellular Ca<sup>2+</sup> signaling is broadly important for many cellular processes, this may be an important mechanism through which  $\sigma_2 R$  ligands produce their documented effects on cancer cells.

Modulation of sphingolipid levels. Sphingolipid levels in MCF-7/Adr<sup>-</sup> and T47D breast tumor cell lines have been investigated following application of  $\sigma_2 R$  specific agonists in order to understand further the molecular mechanism by which  $\sigma_2 R$  ligands could cause their observed morphological and apoptotic effects in various cancer cell lines (103). CB-184 causes a dose-dependent increase in ceramide levels and concomitant decrease in sphingomyelin within the MCF-7/Adr<sup>-</sup> and T47D breast tumor cell lines.

These effects can be attenuated by *N*-phenethylpiperidine, a nonspecific  $\sigma R$  antagonist. These results suggest that  $\sigma_2 Rs$ may use sphingolipid products to affect Ca<sup>2+</sup> signaling, cell proliferation and survival (86,103). In fact, imaging of  $\sigma_1 R$  in the human brain using SPECT radioligands has started to investigate whether  $\sigma Rs$  can be used as prognostic indicators (808), even though it is already known that  $\sigma_2 R$  are potentially useful tumor imaging ligands.

Immunological changes. The mechanism by which  $\sigma$ Rs affect tumor cells has been more recently investigated with respect to immunological alterations (1071).  $\sigma$ R agonists in mice promote the *in vivo* growth of a syngeneic lung cancer cell lines, which was accompanied by an increase in IL-10 and a decrease in interferon production in spleen cells and at the tumor site. The tumor-promoting effects produced were abrogated by administration of specific antibodies to IL-10, or by administration of a  $\sigma_1$ R antagonist, indicating that  $\sigma_1$ R agonist ligands augment tumor growth via a cytokine-dependent, receptor-mediated mechanism that involves regulation of T helper<sub>1</sub>/T helper<sub>2</sub> cytokine balance (1071). Most likely, the alteration of immune cells and function will impact the process of carcinogenesis.

#### Vascular effects

As tumors progress to increased malignancy, cells within them develop the ability to invade into surrounding normal tissues and through tissue boundaries to form metastases at sites distinct from the primary tumor. The molecular mechanisms involved in this process are incompletely understood but those associated with cell-cell and cell-matrix adhesion, with the degradation of extracellular matrix, and with the initiation and maintenance of early growth at the new site are generally accepted to be critical (1068).  $\sigma$ R ligands have also been shown to inhibit stem cell differentiation (96), and modulate endothelial cell proliferation and can control angiogenesis, which makes them a promising target for oncology applications (239).

#### Apoptosis

Apoptosis is a key process in cancer development and progression. The ability of cancer cells to avoid apoptosis and continue to proliferate is one of the fundamental hallmarks of cancer and is a major target of cancer therapy development (110). Apoptosis is the most common mechanism by which the body eliminates damaged or unneeded cells without local inflammation from leakage of cell contents. As the  $\sigma$ Rs have known apoptotic effects on tumors (86), a more detailed review on their anticancer effects follows.

 $\sigma_1 R$ .  $\sigma_1 R$  ligands cause a cell cycle arrest underlined by p27 accumulation. Studies indicate  $\sigma_1 Rs$  modulate cell regulating volume processes in physiological conditions, indicating that  $\sigma_1 Rs$  protect cancer cells from apoptosis. It appears that the  $\sigma_1 Rs$  modulate differentiation (1053). However, other findings suggest that the  $\sigma_2 Rs$  play a very significant role in  $\sigma R$  associated toxicity (1072).

4-(N-benzylpiperidin-4-yl)-4-iodobenzamide [4-IBP], a selective  $\sigma_1 R$  agonist, has been used to investigate whether this compound modifies the migration and proliferation of human cancer cells. 4-IBP has weak antiproliferative effects on human U373-MG glioblastoma and C32 melanoma cells but induces marked concentration-dependent decreases in the growth of human A549 NSCLC and PC3 prostate cancer cells by eliciting apoptosis. The compound was also significantly antimigratory in all four cancer cell lines (1073). These results indicate that up regulation of the  $\sigma_1 R$  decrease growth and migration of malignant human cells in vitro, a finding that has been supported by investigations using cells from other species and non-malignant cell types (1074). These authors investigated the expression of  $\sigma_1 R$  in various human cancer cell lines in comparison to non-cancerous cell lines, using real time RT-PCR and by western blotting with a  $\sigma_1 R$  specific antibody. Also investigated were the effect of  $\sigma_1 R$  and  $\sigma_2 R$ drugs and a  $\sigma_1 R$  silencing construct. The results suggest  $\sigma_1 R$ plays a role in proliferation and adhesion of breast cancer cells (1074).

 $\sigma_2 R$ . Over expression of  $\sigma_2 R$  induces apoptosis (740).  $\sigma_2 R$  proteins are over expressed in several tumor cell lines, but the bimolecular mechanism of this over expression still needs further clarification, although two-photon confocal has shown  $\sigma_2 Rs$  are present in mitochondria, lysozomes, endoplasmic

reticulum and plasma membranes (1075). There is a possibility that this over expression can be used with a radioligand to visualize in human bladder cancer specimens, then if a possible correlation could be established between  $\sigma_2 R$  over expression and tumor tissue stage and grade. In studies done so far, results demonstrate that  $\sigma_2 R$  protein is normally expressed in human bladder and over expressed in the case of high-grade transitional cell carcinomas (112), indicating this technique shows promise for staging of some cancers (111).

 $σ_2R$  agonists induce apoptosis in drug-resistant cancer cells (1038), enhance the potency of DNA damaging agents, and down-regulates expression of p-glycoprotein mRNA (1076).  $σ_2R$  agonists increase lysosomal membrane permeability in the early stages of  $σ_2R$ -induced cell death (1077). Thus,  $σ_2R$  agonists may be useful in treatment of drugresistant cancers and the  $σ_2R$  may serve as a novel signaling pathway to apoptosis (15,981). Further work has demonstrated that the  $σ_2Rs$  are located in lipid rafts in the cell membrane, and these lipid rafts may play an important role in the mechanism of  $σ_2R$ -induced apoptosis (982).

Several  $\sigma_2 Rs$  ligands have been shown to trigger apoptosis in pancreatic cancer cells. More importantly,  $\sigma_2 Rs$  ligands are internalized rapidly by the cancer cells and are capable of delivering other small-molecule therapeutics (1077).

A summary of references related to the expression of  $\sigma Rs$  are outlined in Table 7. A summary of the references that describe the molecular action of  $\sigma Rs$  are outlined in Table 8. A summary of the references describing  $\sigma R$  binding are outlined in Table 9. A summary of the references describing the role of  $\sigma Rs$  in pathophysiology are outlined in Table 10.

## Conclusions

The neuropharmacological properties of  $\sigma_1 R$  ligands relate to the neuron modulatory role of  $\sigma_1 R$ .  $\sigma_1 Rs$  act as intracellular amplifiers for signal transductions involving IP3R and modulate neurotransmitter systems (mainly through NMDA receptors).  $\sigma_1 Rs$  and ion channels may play an important role in neuroplasticity processes.  $\sigma_1 R$  ligands are highly active when a pharmacological or pathological imbalanced state arises.

The combined administration of  $\sigma_1 R$  receptor ligands and medications with a known therapeutic effect has been shown to improve these effects due to the modulatory role of  $\sigma_1 R$ receptors resulting in the need for lower doses to reach therapeutic concentrations. Of particular interest is the nonlinear dose response curve of  $\sigma_1 R$  agonists in *in vitro* experiments, in which  $\sigma_1 R$  agonists are active, e.g. learning and memory processes, depression (1078) and anxiety. These findings imply that researchers should take hormesis into account in order to design informative experiments or clinical trials with  $\sigma_1 R$  agonists. For example, the  $\sigma_1 R$  SA4503 agonist attenuates or enhances the effects of methamphetamine depending on the dose (682).

The most promising therapeutic targets for  $\sigma_1 R$  antagonism are nociception and some deleterious effects of certain drugs of abuse such as cocaine, methamphetamine and ethanol (1079). Many drugs used routinely show affinity for  $\sigma_1 R$  receptors and exert the same effects as other more selective  $\sigma_1 R$  ligands in many behavioral tests and *in vitro* assays. Therefore, the therapeutic properties of these drugs

Location of $\sigma R$ in tissues	Function(s)	Reference(s)
CNS: corpus striatum, nucleus accumbens		(61)
Brain: substantia nigra, pars compacta		(656)
CNS: dentate gyrus of hippocampus, facial		(171,765,766,302)
nucleus, thalamic, hypothalamic nucei, strai-		
tum, cerebellum dorsal raphe nucleus and locus coeruleus		
Hippocampal pyramidal cell layer, hypothala-		(761)
mus, central grey and red nucleus, pontine,		(701)
cranial nerve nuclei, pontine nuclei, Pons –		
medulla, spinal cord – ventral and dorsal route		
ganglia		
Brain: cortex limbic area amygdala		(762)
Brain: cerebellum		(763)
Brain: Medulla – pons, midbrain, cerebellum,		(78,79)
thalamus, straitum, cortex, hippocampus		
Brain: cerebral cortex, straitum, hippocampus, cerebellum		(767)
Brain: substantia nigra, central grey matter,		(766,768)
oculomotor nuclei, cerebellum, nucleus		(700,708)
accumbens, amygdale, olfactory bulb, hippo-		
campus, motor cortex		
CNS	Glu regulation, regulates excitotoxic effect of	(386,417,425,447,450,451,
	Glu	769,770,775,)
CNS	Regulation of $Mn_{2+}$ , $Hg_{2+}$ and $Pb_{2+}$	(379)
	neurotoxicity.	
Neurons: ependymocytes, oligodendrocytes and		(171,672,771–773)
peripheral nervous system Schwann cells	- Description of the section of the	(27.181.010.265.276.600.752
CNS	$\sigma_1 R$ agonist are antiamnesic, improve cognitive	(27,181,212,365,376,602,753,
CNS	abilities Neuroprotection – two subtypes of $\sigma_1 R$ may	776–781,783,785) (360,786–788, )
ens	affect differentially the Glu-mediated NMDA	(500,780-788, )
	neurotransmission in the terminal and origin	
	regions of the mesolimbic and nigrostriatal	
	DA-ergic systems. Also, functional interaction	
	between $\sigma_2 R$ and NMDARs in the hippo-	
	campus. $\sigma_1 R$ agonist may protect neurons by	
	mechanism involving anti-apototic protein	
CNG	bcl-2.	(22 41 164 176 177 105 002)
CNS	$\sigma_1 R$ initiates neurite outgrowth and sprouting.	(32,41,164,176,177,185,803)
	$\sigma_1 R$ agonist potentiates neurite-sprouting by nerve growth factor. $\sigma_1 R$ agonist may	
	potentiate effects of BDNF and EGF	
CNS	Mechanism of action of $\sigma R$ ligands in depression	(88,133,139,153,161,164–166,181,249–251,
	– regulation of Ca <sup>2+</sup> or K <sup>+</sup> signaling. Interacts	376,377,664,828, 829)
	with VDCC's. Modulate Glu and 5-HT	
	transmissions. May be a target for serotonin	
	reuptake inhibitors.	
Brain: frontoparietal cortex, cingulated cortex,	$\sigma_1 R$ ligands affect iGluR subunit levels of	(15,380,802)
dorsal striatum, nucleus accumbens	mRNA and protein, differentially regulating	
	levels of NMDA <sub>2A</sub> and $GluR_2$ in a regionally	
CNS	specific manner. Activation of $\sigma_1 Rs$ antagonize opioid analgesia	(150 102 601 001 002 001)
CINS	whereas antagonists potentiate opioid anal-	(458,483,681,881–883,894)
	gesia. Excitatory amino acids have actions on	
	$\sigma$ Rs indicating action via the Glu system.	
	Activation of spinal $\sigma_1 R$ enhances NMDA-	
	induced pain via PKC- and PKA-dependent	
	phosphorylation of the NMDA receptor NR <sub>1</sub>	
	subunit	
CNS	Agonist of $\sigma_1 R$ and $\sigma_2 R$ inhibit NMDA-	(896)
	stimulated DA release from motor and limbic	
	areas of rat brain.	(75( 000 000)
ANS: Parasympathetic intracardiac neurons	Cardiac excitation and rhythmic control	(756,920–922)
Ion Channel: hERG channel	Cardiac excitation and rhythmic control	(919–922) (150,458,028,020)
Heart and vessels	Effects of $\sigma_1 R$ mediated via PKC- and PKA	(150,458,928,929)
	dependent phosphorylation of the NMDA receptor, altered Na <sup>+</sup> channels	
Heart	$\sigma R$ ligands modulate contractility, Ca <sup>2+</sup> influx	(923,930–932)
	games in an and a sind	

Table 7.	Continued
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Location of $\sigma R$ in tissues	Function(s)	Reference(s)
	reperfusion contracture, increases pressure in left ventricle and improves survival of cardiac myocytes after ischemia and reperfusion. Activation of $\sigma R$ reversibly blocks the delay	
	in outwardly rectifying $K^+$ channels, con- ductance Ca <sup>2+</sup> sensitive $K^+$ channels and the M-current.	
Peripheral arteries	$\sigma R$ agonist increase intracellular Ca <sup>2+</sup> levels by stimulating IP3 production, modulating contractility.	(167)
Muscle	Dystonia	(764,938)
Osteoblasts	Act in conjunction with GluRs to affect cellular changes	(943,944)
Osteocytes, osteoclasts	Act in conjunction with GluRs to affect cellular changes	(492,497,765,945–947)
Lung: pulmonary nerves		(951)
Larynx, esophagus, mast cells		(516,957,958)
Airway passage	Excitation of GluRs may be important in airway inflammation and hyper reactivity observed in bronchial asthma	(440,959,960)
Airways	Antitussive	(194,733,961)
Pituitary, adrenal, testis and ovary		(676)
Endocrine system	Regulatory functions	(127,222,962)
Pituitary	Control of pituitary functions	(868)
Endocrine system	Antidiuretic hormone release	(968)
Posterior pituitary	Inhibit K <sup>+</sup> channel function	(138,139)
Placenta		(976)
Spermatozoa	May affect signaling pathways in conjunction with PROG or prostaglandin $E_{1.}$	(976,977)
Ovary – follicles Testis: ductuli efferentes, ductus epididymis, seminiferous tubules		(676) (676)
Xenopus oocytes, neurohypophysis	Modulation of ion channels. Forms a immuno- precipitating complex with ion channels	(88)
Liver, localized to lipid rafts in rat liver phospholipid membranes, mitochondria		(92,113,980–983)
Kidney	New sector time and instance line Charterisity	(92)
Eye: retinal ganglion cells, inner nuclear mem- brane, inner segments of the photoreceptors, retinal pigment epithelial cells, retinal Mueller cells,	Neuroprotection against ganglion Glu toxicity, apoptosis, $\sigma_1 R$ and $\sigma_2 R$ binding activity stimulated during oxidative stress, important during lens development	(74,783,913,987,989–991,994,995)
Myenteric plexus of the guinea pig ileum	Regulation of ileal contractions, may be involved in regulating acid secretion in stomach	(314,747,998,999)
Gastrointestinal longitudinal muscle and myen- teric plexus	Inhibit electrically or 5-HT-evoked contractions, stimulation of salivary secretion	(314,1001)
Vagus	Induce emesis	(1002–1005)
Human peripheral blood mononuclear cells, lymphocytes	σ2Rs inhibit lymphocyte activation. Potent immunoregulatory properties including induction of IL-10, suppression of IFN-γ and suppression of granulocyte colony stimulating factor.	(98,677,1010–1014)
Splenocytes	Lymphocyte proliferation, mitogen-induced IgG and IgM production, LPS-induced IL-I production	(98,1020)
Viral RNA	Regulate early steps in viral RNA replication	(1017,1018)
Host antitumor immunity	$\sigma$ R-dependent cytokine modulation. Ligand can induce apoptosis by changes in cytosolic Ca <sup>2+</sup> , ceramide and sphingolipid concentrations.	(1019,1013)
Neoplasia	Receptors overexpressed in many cancer tissues	(1022,1023)
reoptasta	Receptors overexpressed in many cancer tissues	(1022,1023)

Table 8. Reference Summary: Molecular Action of  $\sigma Rs$ .

Molecular action of $\sigma R$	Tissue	Reference(s)
Glu regulation: regulates excitotoxic effect of Glu. two subtypes of $\sigma 1R$ may affect differentially the Glu- mediated NMDA neurotransmission in the terminal and origin regions of the mesolimbic and nigrostriatal DA- ergic systems. Functional interaction between $\sigma 2R$ and NMDARs in the hippocampus. $\sigma 1R$ agonist may protect neurons by mechanism involving anti-apototic protein bcl-2	CNS	(27,360,386,417,425,447,450,451, 769,770,775,786–788)
Regulation of $Mn_{2+}$ , $Hg_{2+}$ and $Pb_{2+}$ neurotoxicity	CNS	(379)
$\sigma_1 R$ initiates neurite outgrowth and sprouting. $\sigma_1 R$ agonist potentiates neite-sprouting by nerve growth factor. $\sigma_1 R$ agonist may potentiate effects of BDNF and EGF	CNS	(32,41,164,176,177,185,803)
Regulation of $Ca^{2+}$ or $K^+$ signaling. Interacts with VDCC's. Modulate Glu and 5-HT transmissions.	CNS	(88,133,138,139,153,154,161,164–166, 181,249–251,377,821,829)
$\sigma_1 R$ ligands affect iGluR subunit levels of mRNA and protein, differentially regulating levels of NMDA <sub>2A</sub> and GluR <sub>2</sub> in a regionally specific manner.	Frontoparietal cortex, cingu- lated cortex, dorsal striatum, nucleus accumbens	(15,380,802)
Activation of spinal σ <sub>1</sub> R enhances NMDA-induced pain via PKC- and PKA-dependent phosphorylation of the NMDA receptor NR <sub>1</sub> subunit	CNS	(458,483,681,881–883,894)
Agonist of $\sigma_1 \hat{R}$ and $\sigma_2 \hat{R}$ inhibit NMDA-stimulated DA release from motor and limbic areas of rat brain.	CNS	(896)
Cardiac excitation and rhythmic control	Parasympathetic intracardiac neurons, hERG channel	(756,919–922)
Effects of $\sigma_1 R$ mediated via PKC- and PKA dependent phosphorylation of the NMDA receptor, altered Na <sup>+</sup> channels	Heart and vessels	(150,458,928,929)
$\sigma R$ ligands modulate contractility, Ca <sup>2+</sup> influx and cardiac rate. $\sigma R$ activation prevent reperfusion contracture, increases pressure in left ventricle and improves survival of cardiac myocytes after ischemia and reperfusion. Activation of $\sigma R$ reversibly blocks the delay in out- wardly rectifying K <sup>+</sup> channels, conductance Ca <sup>2+</sup> sen- sitive K <sup>+</sup> channels and the M-current.	Heart	(923,930–932)
$\sigma R$ agonist increase intracellular Ca <sup>2+</sup> levels by stimulating IP3 production, modulating contractility	Peripheral arteries	(167)
Ca <sup>2+</sup> influx	Muscle	(764,938)
Act in conjunction with GluRs to affect cellular changes	Osteoblasts, osteocytes, osteoclasts	(492,497,943–948)
Modulate ion channels	Lungs	(194,440,733,959–961)
Regulatory functions. Control of pituitary functions. Antidiuretic hormone release. Inhibit K <sup>+</sup> channel function.	Endocrine system, pituitary	(127,222,868,962,968)
May affect their signaling pathways in conjunction with PROG or prostaglandin $E_{1}$ .	Spermatozoa	(976,977)
Modulation of ion channels. Forms a immunoprecipitating complex with ion channels	Xenopus oocytes, neurohypophysis	(88)
Neuroprotection against ganglion Glu toxicity, apoptosis, $\sigma_1 R$ and $\sigma_2 R$ binding activity stimulated during oxida- tive stress, important during lens development	Eye – retinal ganglion cells, inner nuclear membrane, inner segments of the photoreceptors, retinal pig- ment epithelial cells, retinal Mueller cells	(74,783,913,987,989–991,994,995)
Regulation of ileal contractions, may be involved in	Myenteric plexus of the guinea	(314,747,998,999)
regulating acid secretion in stomach Inhibit electrically or 5-HT-evoked contractions, stimula- tion of salivary secretion	pig ileum Gastrointestinal longitudinal muscle and myenteric plexus	(314,1001)
5-HT transmissions. σ2Rs inhibit lymphocyte activation. Potent immunoregu- latory properties including induction of IL-10, suppres- sion if IFN-γ and suppression of granulocyte colony stimulating factor.	Vagus Human peripheral blood mononuclear cells, lymphocytes	(1002–1005) (98,677,1010–1014)
Lymphocyte proliferation, miogen-induced IgG and IgM production, LPS-induced IL-I production	Splenocytes	(98,1020)

Table 9. Reference Summary: oR Binding (679, 680).

Drug	Target tissue	
(+)-pentazocine [PTZ]	Sigma <sub>1</sub> Sigma <sub>2</sub> ligands	
Haloperidol [Haldol <sup>®</sup> ]	Sigma <sub>1</sub> Sigma <sub>2</sub> ligands	
1,3 di-o-tolyl-guanidine [DTG]	Sigma <sub>1</sub> Sigma <sub>2</sub> ligands	
(+)-3-PPP [preclamol]	Sigma <sub>1</sub> Sigma <sub>2</sub> ligands	
(+)-SKF 10,047	Sigma <sub>1</sub> Sigma <sub>2</sub> ligands	
(+)-pentazocine [PTZ]	Sigma <sub>1</sub> Sigma <sub>2</sub> ligands	
Phencyclidine	Sigma <sub>1</sub> Sigma <sub>2</sub> ligands	
Dextromethorphan [DEX]	Sigma <sub>1</sub> ligands	
(+)-cyclazocine	Sigma <sub>1</sub> ligands	
BD1047	Sigma <sub>2</sub> ligands	
BD1063	Sigma <sub>2</sub> ligands	

Table 10. Reference Summary: Role of  $\sigma Rs$  in Pathophysiology.

might be due, at least in part, to their interaction with  $\sigma_1 R$  receptors.

The involvement of  $\sigma Rs$  in the cellular pathophysiology of cancer is apparent from the high density of  $\sigma_1 R$  and  $\sigma_2 R$ -binding sites found in various tumor cell lines and tissues. Consequently,  $\sigma R$  drugs have been suggested to be potentially useful tumor imaging agents.

The ability of  $\sigma_2 R$  drugs to inhibit tumor cell proliferation through mechanisms that may involve apoptosis, intracellular Ca<sup>2+</sup> and sphingolipids have been investigated, and such findings may lead to the development of  $\sigma$  drugs as cancer therapeutic agents. It is possible that an increase in  $\sigma_2 R$ 

Tissue type	– disorder	Function or role in pathology	Reference(s)
CNS	Memory loss	$\sigma R$ ligands may be antiamnesic, improve cognitive abilities	(181,212,365,376,602,753, 776–783,785)
CNS	Neurodegeneration	Delays cerebral artery occlusion-induced neurodegeneration and white matter injury. $\sigma_1 R$ agonist protect neurons by a mechanism involving the anti-apoptotic protein bcl-2. Initiation of neurite outgrowth and sprouting. Overstimulation of $\sigma R$ is mediated via Glu, particularly NMDAR leading to osmotic damage, apoptosis and necrosis.	(27,32,41,164,176,177,185, 382,384–386,394,412, 438,511,787,788)
CNS	Schizophrenia	$\sigma_1 R$ polymorphism is associated with increased rick of schizophrenia and differential activation of PFC and the severity of AD.	(44,344)
CNS	Depression, stress	$\sigma$ R, Glu, 5-HT neurotransmission and Ca <sup>2+</sup> regulation. Reduced brain $\sigma_1$ R exacerbates heart failure. Regulation of Ca <sup>2+</sup> or K <sup>+</sup> in neurotransmission.	(26,57,88,133,138,139,153, 154,161,164,165,181, 249–251,326,376, 805–810,828,829)
CNS	Psychosis	Selective $\sigma_1 R$ ligans potentially stimulate adrenocorticotropic hormone release, regulation of neuroendocrine system in brain.	(868–870)
CNS	Seizures	Complex involvement of 5-HT <sub>2</sub> , DA and $\sigma$ Rs. Increase in neuropeptide biosynthesis may play a compensatory antic- onvulsive role. Seizure activity by overstimulation of $\sigma$ R is mediated via Glu, particularly NMDAR.	(407,538,872,873,906–908)
CNS	Pain	Activation of $\sigma_1 Rs$ antagonize opioid analgesia whereas antagonists potentiate opioid analgesia. Excitatory amino acids have actions on $\sigma Rs$ indicating action via the Glu system. Activation of spinal $\sigma_1 R$ enhances NMDA-induced pain via PKC- and PKA-dependent phosphorylation of the NMDA receptor NR <sub>1</sub> subunit.	(483,458,681,881–883,894)
CNS	Addiction	Both cocaine and METH exhibit a significant affinities for $\sigma Rs$ . Agonist of $\sigma_1 R$ and $\sigma_2 R$ inhibit NMDA-stimulated DA release from motor and limbic areas of rat brain.	(872,896–898)
Heart and blood vessels	Heart failure	Reduced $\sigma_1 Rs$ density in depression decreases heart rate via the sympathetic stimulation in the autonomic nervous system. Reduction of brain $\sigma_1 Rs$ also contribute to sympa- thetic hyperactivation of the heart via altered Na <sup>+</sup> channels. Activation of $\sigma_1 R$ depresses the excitability of intracardiac neurons causing changes in beating frequencies, which are followed by irregular contractions. $\sigma Rs$ are involved in the regulation of coronary and peripheral arterial vascular tension.	(150,751,809,810,923,927– 933)
Muscle	drug-induced dystonia	High affinity of some neuroleptics for these sites suggests their possible involvement in some $\sigma_2$ R-mediated side effects.	(764,938)
Bone	Bone	All osteoblasts, osteocytes and osteoclasts express one or more of the GluR subunits, including NMDARs. Possible that $\sigma$ Rs are involved in normal bone function as well as in disease states.	(946,943,944,947,492,945, 497,948)
Lung	Asthma	The presence $\sigma Rs$ in the airway structures such as the larynx, esophagus and mast cells also implicate the GluRs (and	(440,516,957–960)
Endocrine	Drug induced syn- drome of	probably the $\sigma Rs$ ) in the mediation of asthmatic episodes. Interaction with some neuroleptic agents and the posterior pituitary $\sigma R$ ligands can inhibit K <sup>+</sup> -channel function.	(138,139,968)

Tissue t	ype – disorder	Function or role in pathology	Reference(s)
	inappropriate antidiuretic hor- mone release.		
Endocrine	Diabetes mellitus	Dysfunctional islet cells	(435,486–488,499,523,524, 669,969,970,971,512, 972,973)
Eye	Glaucoma, diabetes retinopathy, ret- inal ischemia due to central artery occlusion, anter- ior ischemic optic neuropathy, optic neuritis, optic nerve trauma	Possible neuroprotection by $\sigma R$ ligands against ganglion cell Glu toxicity. Late-onset inner retinal dysfunction in mice lacking $\sigma_1 R$ . Exposure of lens cells to $\sigma R$ antagonists has been shown to lead to growth inhibition and pigment granule production implying importance during lens devel- opment. Gene silencing of the $\sigma_1 R$ induces cell death.	(987,988,992–995)
Gastrointestinal	Emesis	$\sigma Rs$ induce emesis in a number of species mediated centrally via the vagus.	(1002–1004)
Immune system	Graft versus host reactions and delayed-type hypersensitivity granuloma formation. Im- mune dysregulation	$ σ_2 Rs $ inhibit T lymphocyte activation. σR ligands have potent immunoregulatory properties, including the induc- tion of IL-10 and the suppression of IFN-γ and granulocyte colony stimulating factor [GM-CSF].	(98,1012–1014)

expression is a significant event in transition from normal to malignant cells. Further research would be interesting to determine whether  $\sigma R$  are involved in other metastatic cell behaviors such as adhesion, secretion, motility and invasion.

The interaction of  $\sigma Rs$  and other neurotransmitters is complex. As has been discussed in this review,  $\sigma Rs$  are intimately involved with the glutamate system, and are probably an essential part of the expression of excitotoxicity (1080). Other interactions with opiates, neurosteroids, serotonin, dopamine and cannabinoids have been difficult to fully elucidate due to the biphasic nature of dose response curves and the large combination of potential effects. In addition, as most work is done in *in vitro*, doses are often excessive and may reflect an overexposure that would not be seen in the *in vivo* situation. Even though the interaction of  $\sigma Rs$  with various tissues is complex, it is apparent that  $\sigma Rs$  play a central role in neurotransmission and apoptosis. The development of new knockout mice and transgenic initiatives will be important to further research.

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## **Declaration of interest**

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# References

 Martin WR, Eades CG, Thompson JA, et al. The effects of morphine- and nalorphine- like drugs in the nondependent and morphine-dependent chronic spinal dog. J Pharm Exp Therap 1976; 197:517–32.

- Hanner M, Moebius F, Flandorfer A, et al. Purification, molecular cloning, and expression of the mammalian sigma1-binding site. Proc Natl Acad Sci USA 1996;93:8072–7.
- Pan YX, Mei J, Xu J, et al. Cloning and characterization of a mouse sigma1 receptor. J Neurochem 1998;70:2279–85.
- Prasad PD, Srinivas SR, Wang H, et al. Electrogenic nature of rat sodium-dependent multivitamin transport. Biochem Biophys Res Commun 2000;270:836–40.
- Seth P, Fei YJ, Li HW, et al. Cloning and functional characterization of a sigma receptor from rat brain. J Neurochem 1998;70: 922–31.
- Seth P, Leibach FH, Ganapathy V. Cloning and structural analysis of the cDNA and the gene encoding the murine type 1 sigma receptor. Biochem Biophys Res Commun 1997;241: 535–40.
- Cozzi N, Gopalakrishnan A, Anderson L, et al. Dimethyltryptamine and other hallucinogenic tryptamines exhibit substrate behavior at the serotonin uptake transporter and the vesicle monoamine transporter. J Neural transmission (Vienna, Austria: 1996) 2009; 116:1591–9.
- Fontanilla D, Johannessen M, Hajipour A, et al. The hallucinogen N,N-dimethyltryptamine (DMT) is an endogenous sigma-1 receptor regulator. Science (NY) 2009;323:934–7.
- 9. Su TP, Hayashi T, Vaupel DB. When the endogenous hallucinogenic trace amine N,N-dimethyltryptamine meets the sigma-1 receptor. Sci Signal 2009;2:pe12.
- Bastianetto S, Ramassamy C, Poirier J, Quirion R. Dehydroepiandrosterone (DHEA) protects hippocampal cells from oxidative stress-induced damage. Brain Res Mol Brain Res 1999; 66:35–41.
- 11. Menkel M, Terry P, Pontecorvo M, et al. Selective sigma ligands block stimulant effects of cocaine. Eur J Pharmacol 1991;201: 251–2.
- 12. Kaushal N, Matsumoto RR. Role of sigma receptors in methamphetamine-induced neurotoxicity. Curr Neuropharmacol 2011;9: 54–7.
- Kaushal N, Seminerio MJ, Shaikh J, et al. CM156, a high affinity sigma ligand, attenuates the stimulant and neurotoxic effects of methamphetamine in mice. Neuropharmacology 2011;61: 992–1000.
- Brammer M, Gilmore D, Matsumoto R. Interactions between 3,4methylenedioxymethamphetamine and sigma1 receptors. Eur J Pharmacol 2006;553:141–5.

- Guitart X, Codony X, Monroy X. Sigma receptors: biology and therapeutic potential. Psychopharmacology 2004;174:301–19.
- Meririnne E, Kankaanpaa A, Lillsunde P, Seppala T. The effects of diazepam and zolpidem on cocaine- and amphetamine-induced place preference. Pharmacol Biochem Behav 1999;62:159–64.
- Cormaci G, Mori T, Hayashi T, Su TP. Protein kinase A activation down-regulates, whereas extracellular signal-regulated kinase activation up-regulates sigma-1 receptors in B-104 cells: implication for neuroplasticity. J Pharm Exp Therap 2007;320:202–10.
- Freeman A, Bunney B. The effects of phencyclidine and Nallylnormetazocine on midbrain dopamine neuronal activity. Eur J Pharmacol 1984;104:287–93.
- Ohno M, Watanabe S. Intrahippocampal administration of (+)-SKF 10,047, a sigma ligand, reverses MK-801-induced impairment of working memory in rats. Brain Res 1995;684:237–42.
- Vaupel DB. Naltrexone fails to antagonize the sigma effects of PCP and SKF 10,047 in the dog. Eur J Pharmacol 1983;92:269–74.
- Quirion R, Bowen WD, Itzhak Y, et al. A proposal for the classification of sigma binding sites. Trends Pharmacol Sci 1992; 12:85–6.
- 22. Su TP. Evidence for sigma opioid receptor: binding of [3H]SKF-10047 to etorphine-inaccessible sites in guinea-pig brain. J Pharm Exp Therap 1982;223:284–90.
- Vidal-Torres A, de la Puente B, Rocasalbas M, et al. Sigma-1 receptor antagonism as opioid adjuvant strategy: enhancement of opioid antinociception without increasing adverse effects. Eur J Pharmacol 2013;711:63–72.
- Bowen W, Hellewell S, McGarry K. Evidence for a multi-site model of the rat brain sigma receptor. Eur J Pharmacol 1989;163: 309–18.
- 25. Kitaichi K, Chabot JG, Moebius FF, et al. Expression of the purported sigma(1) (sigma(1)) receptor in the mammalian brain and its possible relevance in deficits induced by antagonism of the NMDA receptor complex as revealed using an antisense strategy. J Chem Neuroanat 2000;20:375–87.
- 26. Skuza G, Rogoz Z. The synergistic effect of selective sigma receptor agonists and uncompetitive NMDA receptor antagonists in the forced swim test in rats. J Physiol Pharmacol 2006;57: 217–29.
- Yang S, Bhardwaj A, Cheng J, et al. Sigma receptor agonists provide neuroprotection in vitro by preserving bcl-2. Anesth Analg 2007;104:1179–84.
- Meyer JH, Lee S, Wittenberg GF, et al. Neurosteroid regulation of inhibitory synaptic transmission in the rat hippocampus in vitro. Neuroscience 1999;90:1177–83.
- Myers AM, Charifson PS, Owens CE, et al. Conformational analysis, pharmacophore identification, and comparative molecular field analysis of ligands for the neuromodulatory sigma 3 receptor. J Med Chem 1994;37:4109–17.
- Lee IT, Chen S, Schetz JA. An unambiguous assay for the cloned human sigmal receptor reveals high affinity interactions with dopamine D4 receptor selective compounds and a distinct structureaffinity relationship for butyrophenones. Eur J Pharmacol 2008; 578:123–36.
- Xu J, Zeng C, Chu W, Pan F, et al. Identification of the PGRMC1 protein complex as the putative sigma-2 receptor binding site. Nature Commun 2011;2:380–94.
- 32. Cobos E, Entrena J, Nieto F, Cendan C, Del Pozo E. Pharmacology and therapeutic potential of sigma(1) receptor ligands. Current neuropharmacology 2008;6:344–66.
- Paschos KA, Veletza S, Chatzaki E. Neuropeptide and sigma receptors as novel therapeutic targets for the pharmacotherapy of depression. CNS Drugs 2009;23:755–72.
- Kulkarni SK, Dhir A. sigma-1 receptors in major depression and anxiety. Expert review of neurotherapeutics 2009;9:1021–34.
- Navarro JF, Beltran D, Cavas M. Effects of (+) SKF 10,047, a sigma-1 receptor agonist, on anxiety,tested in two laboratory models in mice. Psicothema 2012;24:427–30.
- 36. Hashimoto K. Comments on "An innovative design to establish proof of concept of the antidepressant effects of the NR2B subunit selective N-methyl-D-aspartate antagonist, CP-101,606 in patients with treatment-refractory major depressive disorder". J Clin Psychopharmacol 2009;29:411–12.
- 37. Hashimoto T, Nguyen Q, Rotaru D, et al. Protracted developmental trajectories of GABAA receptor alpha1 and alpha2 subunit

expression in primate prefrontal cortex. Biol Psychiatry 2009;65: 1015–23.

- Diaz J, Zamanillo D, Corbera J, et al. Selective sigma-1 (sigma1) receptor antagonists: emerging target for the treatment of neuropathic pain. Central Nerv Syst Agents Med Chem 2009;9: 172–83.
- Zamanillo D, Romero L, Merlos M, Vela JM. Sigma 1 receptor: a new therapeutic target for pain. Eur J Pharmacol 2013;716: 78–93.
- Furuse T, Hashimoto K. Sigma-1 receptor agonist fluvoxamine for delirium in patients with Alzheimer's disease. Ann Gen Psychiatry 2010;9:6.
- Luty AA, Kwok JB, Dobson-Stone C, Loy CT, et al. Sigma nonopioid intracellular receptor 1 mutations cause frontotemporal lobar degeneration-motor neuron disease. Ann Neurol 2010;68: 639–49.
- 42. Mishina M, Ohyama M, Ishii K, et al. Low density of sigma1 receptors in early Alzheimer's disease. Ann Nuclear Med 2008;22: 151–6.
- Feher A, Juhasz A, Laszlo A, et al. Association between a variant of the sigma-1 receptor gene and Alzheimer's disease. Neurosci Lett 2012;517:136–9.
- Huang Y, Zheng L, Halliday G, et al. Genetic polymorphisms in sigma-1 receptor and apolipoprotein E interact to influence the severity of Alzheimer's disease. Current Alzheimer research 2011; 8:765–70.
- Mesangeau C, Narayanan S, Green AM, et al. Conversion of a highly selective sigma-1 receptor-ligand to sigma-2 receptor preferring ligands with anticocaine activity. J Med Chem 2008; 51:1482–6.
- Hayashi T, Justinova Z, Hayashi E, et al. Regulation of sigma-1 receptors and endoplasmic reticulum chaperones in the brain of methamphetamine self-administering rats. J Pharm Exp Therap 2010;332:1054–63.
- Smith KJ, Butler TR, Prendergast MA. Inhibition of sigma-1 receptor reduces N-methyl-D-aspartate induced neuronal injury in methamphetamine-exposed and -naive hippocampi. Neurosci Lett 2010;481:144–8.
- Hayashi T, Su T. An update on the development of drugs for neuropsychiatric disorders: focusing on the sigma 1 receptor ligand. Expert Opin Therap Targets 2008;12:45–58.
- Dhir A, Kulkarni S. Possible involvement of sigma-1 receptors in the anti-immobility action of bupropion, a dopamine reuptake inhibitor. Fundam Clin Pharmacol 2008;22:387–94.
- Lever JR, Miller DK, Fergason-Cantrell EA, et al. Relationship between cerebral sigma-1 receptor occupancy and attenuation of cocaine's motor stimulatory effects in mice by PD144418. J Pharm Exp Therap 2014;351:153–63.
- 51. Matsumoto RR, Shaikh J, Wilson LL, et al. Attenuation of methamphetamine-induced effects through the antagonism of sigma (sigma) receptors: evidence from in vivo and in vitro studies. Eur Neuropsychopharmacol 2008;18:871–81.
- Sabino V, Cottone P, Blasio A, et al. Activation of sigma-receptors induces binge-like drinking in Sardinian alcohol-preferring rats. Neuropsychopharmacology 2011;36:1207–18.
- Sabino V, Cottone P, Parylak SL, et al. Sigma-1 receptor knockout mice display a depressive-like phenotype. Behav Brain Res 2009; 198:472–6.
- Bhutada P, Mundhada Y, Ghodki Y, et al. Influence of sigma-1 receptor modulators on ethanol-induced conditioned place preference in the extinction-reinstatement model. Behav Pharmacol 2012; 23:25–33.
- Furuse T, Hashimoto K. Fluvoxamine monotherapy for psychotic depression: the potential role of sigma-1 receptors. Ann Gen Psychiatry 2009;8:26–9.
- Fu Y, Yu S, Guo X, et al. Fluvoxamine increased glutamate release by activating both 5-HT(3) and sigma-1 receptors in prelimbic cortex of chronic restraint stress C57BL/6 mice. Biochim Biophys Acta 2012;1823:826–37.
- 57. Furuse T, Hashimoto K. Fluvoxamine for aripiprazole-associated akathisia in patients with schizophrenia: a potential role of sigma-1 receptors. Ann Gen Psychiatry 2010;9:11–14.
- Kishimoto Y, Terada S, Sato S, et al. Repetitive questioning behavior in Alzheimer's disease: relationship to regional cerebral blood flow. Psychiatry Res 2010;184:151–6.

- 59. Sugimoto Y, Tagawa N, Kobayashi Y, et al. Involvement of the sigmal receptor in the antidepressant-like effects of fluvoxamine in the forced swimming test in comparison with the effects elicited by paroxetine. Eur J Pharmacol 2012;696:96–100.
- Ogawa K, Shiba K, Akhter N, et al. Evaluation of radioiodinated vesamicol analogs for sigma receptor imaging in tumor and radionuclide receptor therapy. Cancer Sci 2009;100:2188–92.
- 61. Angulo J, McEwen B. Molecular aspects of neuropeptide regulation and function in the corpus striatum and nucleus accumbens. Brain Res Brain Res Rev 1994;19:1–28.
- Heroux J, Tam S, De Souza E. Autoradiographic identification and characterization of sigma receptors in guinea pig brain using [3H]1(cyclopropylmethyl)-4-(2'-(4"-fluorophenyl)-2'-oxoethyl) piperidine ([3H]DuP 734): a novel sigma receptor ligand. Brain Res 1992;598:76–86.
- Langa F, Codony X, Tovar V, et al. Generation and phenotypic analysis of sigma receptor type I (sigma 1) knockout mice. Eur J Neurosci 2003;18:2188–96.
- Hayashi T, Su T. Intracellular dynamics of sigma-1 receptors (sigma(1) binding sites) in NG108-15 cells. J Pharm Exp Therap 2003;306:726–33.
- 65. Hayashi T, Su T. The potential role of sigma-1 receptors in lipid transport and lipid raft reconstitution in the brain: implication for drug abuse. Life Sci 2005;77:1612–24.
- 66. Hayashi T, Su T. Sigma-1 receptors (sigma(1) binding sites) form raft-like microdomains and target lipid droplets on the endoplasmic reticulum: roles in endoplasmic reticulum lipid compartmentalization and export. J Pharm Exp Therap 2003;306:718–25.
- 67. Mitsuda T, Omi T, Tanimukai H, et al. Sigma-1Rs are upregulated via PERK/eIF2alpha/ATF4 pathway and execute protective function in ER stress. Biochem Biophys Res Commun 2011;415: 519–25.
- Prasad PD, Li HW, Fei YJ, et al. Exon-intron structure, analysis of promoter region, and chromosomal localization of the human type 1 sigma receptor gene. J Neurochem 1998;70:443–51.
- Musacchio JM, Klein M, Santiago LJ. High affinity dextromethorphan binding sites in guinea pig brain: further characterization and allosteric interactions. J Pharm Exp Therap 1988;247:424–31.
- Itzhak Y. Different modulation of the binding to two phencyclidine (PCP) receptor subtypes: effects of N-methyl-D-aspartate agonists and antagonists. Neurosci Lett 1989;104:314–19.
- Itzhak Y. Multiple affinity binding states of the sigma receptor: effect of GTP-binding protein-modifying agents. Mol Pharmacol 1989;36:512–17.
- 72. Itzhak Y, Stein I. Regulation of sigma receptors and responsiveness to guanine nucleotides following repeated exposure of rats to haloperidol: further evidence for multiple sigma binding sites. Brain Res 1991;666:166–72.
- Itzhak Y, Stein I. Sigma binding sites in the brain; an emerging concept for multiple sites and their relevance for psychiatric disorders. Life Sci 1990;47:1073–81.
- Kim FJ, Kovalyshyn I, Burgman M, et al. Sigma 1 receptor modulation of G-protein-coupled receptor signaling: potentiation of opioid transduction independent from receptor binding. Mol Pharmacol 2010;77:695–703.
- Witkin JM, Steele TD, Sharpe LG. Effects of strychnineinsensitive glycine receptor ligands in rats discriminating dizocilpine or phencyclidine from saline. J Pharm Exp Therap 1997;280: 46–52.
- Kekuda R, Prasad PD, Fei YJ, et al. Cloning and functional expression of the human type 1 sigma receptor (hSigmaR1). Biochem Biophys Res Commun 1996;229:553–8.
- 77. Seth P, Ganapathy ME, Conway SJ, et al. Expression pattern of the type 1 sigma receptor in the brain and identity of critical anionic amino acid residues in the ligand-binding domain of the receptor. Biochim Biophys Acta 2001;1540:59–67.
- Weissman AD, Dam M, London ED. Alterations in local cerebral glucose utilization induced by phencyclidine. Brain Res 1987;435: 29–40.
- Weissman AD, Su TP, Hedreen JC, London ED. Sigma receptors in post-mortem human brains. J Pharm Exp Therap 1988;247: 29–33.
- Yamamoto H, Miura R, Yamamoto T, et al. Amino acid residues in the transmembrane domain of the type 1 sigma receptor critical for ligand binding. FEBS Lett 1999;445:19–22.

- Kavanaugh MP, Tester BC, Scherz MW, et al. Identification of the binding subunit of the sigma-type opiate receptor by photoaffinity labeling with 1-(4-azido-2-methyl[6-3H]phenyl)-3-(2-methyl[4,6-3H]phenyl)guanidine. Proc Natl Acad Sci USA 1988;85:2844–8.
- Su TP, Hayashi T. Understanding the molecular mechanism of sigma-1 receptors: towards a hypothesis that sigma-1 receptors are intracellular amplifiers for signal transduction. Curr Med Chem 2003;10:2073–80.
- Laurini E, Marson D, Dal Col V, et al. Another brick in the wall. Validation of the sigma1 receptor 3D model by computer-assisted design, synthesis, and activity of new sigma1 ligands. Mol Pharm 2012;9:3107–26.
- Ganapathy M, Prasad P, Huang W, et al. Molecular and ligandbinding characterization of the sigma-receptor in the Jurkat human T lymphocyte cell line. J Pharm Exp Therap 1999;289: 251–60.
- Wang LM, Shelness GS, Childers SR, et al. Cloning and expression of an alternative splice variant of the mouse sigma-1 receptor. Proc Am Assoc Cancer Res 2000;41:205–6.
- Aydar E, Palmer C, Djamgoz M. Sigma receptors and cancer: possible involvement of ion channels. Cancer Res 2004;64: 5029–35.
- Wang K, Hackett JT, Cox ME, et al. Regulation of the neuronal nicotinic acetylcholine receptor by SRC family tyrosine kinases. J Biol Chem 2004;279:8779–86.
- Aydar E, Palmer C, Klyachko V, Jackson M. The sigma receptor as a ligand-regulated auxiliary potassium channel subunit. Neuron 2002;34:399–410.
- Amer M, McKeown L, Tumova S, et al. Inhibition of endothelial cell Ca(2)(+) entry and transient receptor potential channels by Sigma-1 receptor ligands. Br J Pharmacol 2013;68:1445–55.
- Rhoades DJ, Kinder DH, Mahfouz TM. A comprehensive ligand based mapping of the sigma2 receptor binding pocket. Med Chem (Shariqah (UAE)) 2014;10:98–121.
- Hellewell S, Bowen W. A sigma-like binding site in rat pheochromocytoma (PC12) cells: decreased affinity for (+)-benzomorphans and lower molecular weight suggest a different sigma receptor form from that of guinea pig brain. Brain Res 1990;527: 244–53.
- 92. Hellewell S, Bruce A, Feinstein G, et al. Rat liver and kidney contain high densities of sigma 1 and sigma 2 receptors: characterization by ligand binding and photoaffinity labeling. Eur J Pharmacol 1994;268:9–18.
- Moltzen EK, Perregaard J, Meier E. Sigma ligands with subnanomolar affinity and preference for the sigma 2 binding site. 2: spiro-joined benzofuran, isobenzofuran, and benzopyran piperidines. J Med Chem 1995;38:2009–17.
- Bowen W, Vilner B, Williams W, et al. Ibogaine and its congeners are sigma 2 receptor-selective ligands with moderate affinity. Eur J Pharmacol 1995;279:R1–3.
- Vilner BJ, Bowen WD. Modulation of cellular calcium by sigma-2 receptors: release from intracellular stores in human SK-N-SH neuroblastoma cells. J Pharm Exp Therap 2000;292:900–11.
- Haller J, Panyutin I, Chaudhry A, et al. Sigma-2 receptor as potential indicator of stem cell differentiation. Molecular imaging and biology: MIB 2012;14:325–35.
- 97. Walker JM, Bowen WD, Patrick SL, et al. A comparison of (-)-deoxybenzomorphans devoid of opiate activity with their dextrorotatory phenolic counterparts suggests role of sigma 2 receptors in motor function. Eur J Pharmacol 1993;231:61–8.
- Walker JM, Bowen WD, Walker FO, et al. Sigma receptors: biology and function. Pharmacol Rev 1990;42:355–402.
- Walker R. The significance of excursions above the ADI. Case study: monosodium glutamate. Regul Toxicol Pharmacol 1999;30: S119–21.
- Kinney GG, Harris EW, Ray R, Hudzik TJ. sigma2 Site-mediated inhibition of electrically evoked guinea pig ileum longitudinal muscle/myenteric plexus contractions. Eur J Pharmacol 1995;294: 547–53.
- 101. Jeanjean AP, Mestre M, Maloteaux JM, Laduron PM. Is the sigma 2 receptor in rat brain related to the K+ channel of class III antiarrhythmic drugs? Eur J Pharmacol 1993;241:111–16
- Couture S, Debonnel G. Modulation of the neuronal response to N-methyl-D-aspartate by selective sigma2 ligands. Synapse (NY) 1998;29:62–71.

- 368 C. G. Rousseaux & S. F. Greene
- Crawford K, Bowen W. Sigma-2 receptor agonists activate a novel apoptotic pathway and potentiate antineoplastic drugs in breast tumor cell lines. Cancer Res 2002;62:313–22.
- Cassano G, Gasparre G, Contino M, et al. The sigma-2 receptor agonist PB28 inhibits calcium release from the endoplasmic reticulum of SK-N-SH neuroblastoma cells. Cell Calcium 2006; 40:23–8.
- 105. Spitzer D, Simon Jr PO, Kashiwagi H, et al. Use of multifunctional sigma-2 receptor ligand conjugates to trigger cancerselective cell death signaling. Cancer Res 2012;72:201–9.
- Zeng C, Rothfuss J, Zhang J, et al. Sigma-2 ligands induce tumour cell death by multiple signalling pathways. Br J Cancer 2012;106: 693–701.
- Abate C, Mosier P, Berardi F, Glennon R. A structure-affinity and comparative molecular field analysis of sigma-2 (sigma2) receptor ligands. Central Nerv Syst Agents Med Chem 2009;9:246–57.
- Wang B, Rouzier R, Albarracin CT, et al. Expression of sigma 1 receptor in human breast cancer. Breast Cancer Res Treat 2004; 87:205–14.
- 109. Hou X, Hui YN, Han QH, et al. Effects of magnetic field on MAPK signaling pathways of human retinal pigment epithelial cells bound with beads in vitro. [Zhonghua yan ke za zhi] Chin J Opthamol 2006;42:1103–8.
- 110. Kashiwagi H, McDunn JE, Simon Jr PO, et al. Selective sigma-2 ligands preferentially bind to pancreatic adenocarcinomas: applications in diagnostic imaging and therapy. Mol Cancer 2007;6:48–60.
- 111. Colabufo N, Berardi F, Abate C, et al. Is the sigma2 receptor a histone binding protein? J Med Chem 2006;49:4153–8
- 112. Schepmann D, Lehmkuhl K, Brune S, Wunsch B. Expression of sigma receptors of human urinary bladder tumor cells (RT-4 cells) and development of a competitive receptor binding assay for the determination of ligand affinity to human sigma(2) receptors. J Pharm Biomed Anal 2011;55:1136–41.
- Mukherjee A, Prasad TK, Rao NM, Banerjee R. Haloperidolassociated stealth liposomes: a potent carrier for delivering genes to human breast cancer cells. J Biol Chem 2005;280:15619–27.
- 114. Fu Y, Zhao Y, Luan W, et al. Sigma-1 receptors amplify dopamine D1 receptor signaling at presynaptic sites in the prelimbic cortex. Biochim Biophys Acta 2010;1803:1396–408.
- 115. Booth R, Owens C, Brown R, et al. Putative sigma(3) sites in mammalian brain have histamine H(1) receptor properties: evidence from ligand binding and distribution studies with the novel H(1) radioligand [(3)H]-(-)-trans-1-phenyl-3-aminotetralin. Brain Res 1999;837:95–105.
- 116. Lathe R, Seckl JR. Neurosteroids and brain steroids. Boca Raton: CRX Press; 2002.
- Tsai SY, Rothman RK, Su TP. Insights into the Sigma-1 receptor chaperone's cellular functions: a microarray report. Synapse (NY) 2012;66:42–51.
- Tsai SY, Hayashi T, Mori T, Su TP. Sigma-1 receptor chaperones and diseases. Central Nerv Syst Agents Med Chem 2009;9:184–9.
- Marriott KS, Prasad M, Thapliyal V, Bose HS. sigma-1 receptor at the mitochondrial-associated endoplasmic reticulum membrane is responsible for mitochondrial metabolic regulation. J Pharm Exp Therap 2012;343:578–86.
- 120. Shioda N, Ishikawa K, Tagashira H, et al. Expression of a truncated form of the endoplasmic reticulum chaperone protein, sigmal receptor, promotes mitochondrial energy depletion and apoptosis. J Biol Chem 2012;287:23318–31.
- 121. Ishikawa M, Hashimoto K. The role of sigma-1 receptors in the pathophysiology of neuropsychiatric diseases. J Receptor Ligand Channel Res 2009;33:25–36.
- Gonzalez-Alvear G, Werling L. Sigma receptor regulation of norepinephrine release from rat hippocampal slices. Brain Res 1995;673:61–9.
- 123. Bergeron R, de Montigny C, Debonnel G. Potentiation of neuronal NMDA response induced by dehydroepiandrosterone and its suppression by progesterone: effects mediated via sigma receptors. J Neurosci 1996;16:1193–202.
- 124. Debonnel G. Current hypotheses on sigma receptors and their physiological role: possible implications in psychiatry. J Psychiatry Neurosci 1993;18:157–72.
- 125. Debonnel G, de Montigny C. Modulation of NMDA and dopaminergic neurotransmissions by sigma ligands: possible

implications for the treatment of psychiatric disorders. Life Sci 1996;58:721–34.

- Su TP. Delineating biochemical and functional properties of sigma receptors: emerging concepts. Crit Rev Neurobiol 1993;7: 187–203.
- 127. Su TP. Sigma receptors: putative links between nervous, endocrine and immune systems. Eur J Biochem 1991;200:633–42.
- Su TP, Shukla K, Gund T. Steroid binding at sigma receptors: CNS and immunological implications. Ciba Found Symp 1990; 153:107–13; discussion 113–106.
- Skuza G, Rogoz Z. A potential antidepressant activity of SA4503, a selective sigma 1 receptor agonist. Behav Pharmacol 2002;13: 537–43.
- Skuza G, Rogoz Z, Golembiowska K. EMD 57445, the selective sigma receptor ligand, has no effect on the 5-hydroxytryptamine system. Pol J Pharmacol 1997;49:489–93.
- Maurice T, Privat A. SA4503, a novel cognitive enhancer with sigmal receptor agonist properties, facilitates NMDA receptordependent learning in mice. Eur J Pharmacol 1997;328:9–18.
- 132. Tokuyama S, Hirata K, Ide A, Ueda H. Sigma ligands stimulate GTPase activity in mouse prefrontal membranes: evidence for the existence of metabotropic sigma receptor. Neurosci Lett 1997;233: 141–4.
- 133. Soriani O, Vaudry H, Mei YA, et al. Sigma ligands stimulate the electrical activity of frog pituitary melanotrope cells through a G-protein-dependent inhibition of potassium conductances. J Pharm Exp Therap 1998;286:163–71.
- Carnally S, Johannessen M, Henderson R, et al. Demonstration of a direct interaction between sigma-1 receptors and acid-sensing ion channels. Biophys J 2010;98:1182–91.
- Monnet FP, de Costa BR, Bowen WD. Differentiation of sigma ligand-activated receptor subtypes that modulate NMDA-evoked [3H]-noradrenaline release in rat hippocampal slices. Br J Pharmacol 1996;119:65–72.
- 136. Monnet FP, Debonnel G, Bergeron R, et al. The effects of sigma ligands and of neuropeptide Y on N-methyl-D-aspartate-induced neuronal activation of CA3 dorsal hippocampus neurones are differentially affected by pertussin toxin. Br J Pharmacol 1994; 112:709–15.
- 137. Monnet FP, Debonnel G, de Montigny C. In vivo electrophysiological evidence for a selective modulation of N-methyl-Daspartate-induced neuronal activation in rat CA3 dorsal hippocampus by sigma ligands. J Pharm Exp Therap 1992;261:123–30.
- Lupardus PJ, Wilke RA, Aydar E, et al. Membrane-delimited coupling between sigma receptors and K+ channels in rat neurohypophysial terminals requires neither G-protein nor ATP. J Physiol 2000;526:527–39.
- Wilke RA, Lupardus PJ, Grandy DK, et al. K+ channel modulation in rodent neurohypophysial nerve terminals by sigma receptors and not by dopamine receptors. J Physiol 1999; 517:391–406.
- DeHaven-Hudkins D, Fleissner L, Ford-Rice F. Characterization of the binding of [3H](+)-pentazocine to sigma recognition sites in guinea pig brain. Eur J Pharmacol 1992;227:371–8.
- DeHaven-Hudkins D, Ford-Rice F, Allen J, Hudkins R. Allosteric modulation of ligand binding to [3H](+)pentazocine-defined sigma recognition sites by phenytoin. Life Sci 1993;53:41–8.
- Hong W, Werling LL. Evidence that the sigma(1) receptor is not directly coupled to G proteins. Eur J Pharmacol 2000;408:117–25.
- McCann DJ, Weissman AD, Su TP. Sigma-1 and sigma-2 sites in rat brain: comparison of regional, ontogenetic, and subcellular patterns. Synapse (NY) 1994;17:182–9.
- 144. Palmer CP, Aydar E, Jackson MB. Sigma receptor modulation of ion channels. New York: Kluwer Academic Publishers; 2004.
- 145. Soriani O, Foll FL, Roman F, et al. A current down-modulated by sigma receptor in frog pituitary melanotrope cells through a G protein-dependent pathway. J Pharm Exp Therap 1999;289:321–8.
- Soriani O, Le Foll F, Galas L, et al. The sigma-ligand (+)-pentazocine depresses M current and enhances calcium conductances in frog melanotrophs. Am J Physiol 1999;277:E73–80.
- 147. Jbilo O, Vidal H, Paul R, et al. Purification and characterization of the human SR 31747A-binding protein. A nuclear membrane protein related to yeast sterol isomerase. J Biol Chem 1997;272: 27107–15.

- 148. Beart P, O'Shea R, Manallack D. Regulation of sigma-receptors: high- and low-affinity agonist states, GTP shifts, and up-regulation by rimcazole and 1,3-Di(2-tolyl)guanidine. J Neurochem 1989;53:779–88.
- Connick J, Hanlon G, Roberts J, et al. Multiple sigma binding sites in guinea-pig and rat brain membranes: G-protein interactions. Br J Pharmacol 1992;107:726–31.
- Johannessen M, Ramachandran S, Riemer L, et al. Voltage-gated sodium channel modulation by sigma-receptors in cardiac myocytes and heterologous systems. Am J Physiol Cell Physiol 2009;296:C1049–57.
- Kemp JA, Foster AC, Wong EH, Middlemiss DN. A comment on the classification and nomenclature of phencyclidine and sigma receptor sites. Trends Neurosci 1998;11:388–9.
- Morio Y, Tanimoto H, Yakushiji T, Morimoto Y. Characterization of the currents induced by sigma ligands in NCB20 neuroblastoma cells. Brain Res 1994;637:190–6.
- 153. Brent P, Herd L, Saunders H, et al. Protein phosphorylation and calcium uptake into rat forebrain synaptosomes: modulation by the sigma ligand, 1,3-ditolylguanidine. J Neurochem 1997;68: 2201–11.
- 154. Brent P, Saunders H, Dunkley P. Intrasynaptosomal free calcium levels in rat forebrain synaptosomes: modulation by sigma (sigma) receptor ligands. Neurosci Lett 1996;211:138–42.
- Paul IA, Basile AS, Rojas E, et al. Sigma receptors modulate nicotinic receptor function in adrenal chromaffin cells. FASEB J 1993;7:1171–8.
- 156. Okuyama S, Imagawa Y, Ogawa S, et al. NE-100, a novel sigma receptor ligand: in vivo tests. Life Sci 1993;53:Pl285–90.
- 157. Church J, Fletcher E. Blockade by sigma site ligands of high voltage-activated Ca2+ channels in rat and mouse cultured hippocampal pyramidal neurones. Br J Pharmacol 1995;116: 2801–10.
- Zhang H, Cuevas J. Sigma receptors inhibit high-voltage-activated calcium channels in rat sympathetic and parasympathetic neurons. J Neurophysiol 2002;87:2867–79.
- 159. Bowen W. Sigma receptors and iboga alkaloids. Alkaloids Chem Biol 2001;56:173–91.
- Paul IA, Layer RT, Skolnick P, Nowak G. Adaptation of the NMDA receptor in rat cortex following chronic electroconvulsive shock or imipramine. Eur J Pharmacol 1993;247:305–11.
- 161. Hayashi T, Maurice T, Su T. Ca(2+) signaling via sigma(1)receptors: novel regulatory mechanism affecting intracellular Ca(2+) concentration. J Pharm Exp Therap 2000;293:788–98.
- 162. Monnet FP, Blier P, Debonnel G, de Montigny C. Modulation by sigma ligands of N-methyl-D-aspartate-induced [3H]noradrenaline release in the rat hippocampus: G-protein dependency. Naunyn– Schmiedeberg's Arch Pharmacol 1992;346:32–9.
- Monnet FP, Morin-Surun MP, Leger J, Combettes L. Protein kinase C-dependent potentiation of intracellular calcium influx by sigmal receptor agonists in rat hippocampal neurons. J Pharm Exp Therap 2003;307:705–12.
- Hayashi T, Su T. Regulating ankyrin dynamics: roles of sigma-1 receptors. Proc Natl Acad Sci USA 2001;98:491–6.
- 165. Morin-Surun MP, Collin T, Denavit-Saubie M, et al. Intracellular sigmal receptor modulates phospholipase C and protein kinase C activities in the brainstem. Proc Natl Acad Sci USA 1999;96: 8196–9.
- 166. Brent P, Pang G, Little G, et al. The sigma receptor ligand, reduced haloperidol, induces apoptosis and increases intracellularfree calcium levels [Ca2+]i in colon and mammary adenocarcinoma cells. Biochem Biophys Res Commun 1996;219:219–26.
- 167. Novakova M, Ela C, Bowen WD, et al. Highly selective sigma receptor ligands elevate inositol 1,4,5-trisphosphate production in rat cardiac myocytes. Eur J Pharmacol 1998;353:315–27.
- Kostyuk P, Akaike N, Osipchuk Y, et al. Gating and permeation of different types of Ca channels. Ann NY Acad Sci 1989;560: 63–79.
- 169. Morin D, Sapena R, Elimadi A, et al. [3H]-Trimetazidine mitochondrial binding sites: regulation by cations, effect of trimetazidine derivatives and other agents and interaction with an endogenous substance. Br J Pharmacol 2000;130:655–63.
- Basile A, Paul I, Mirchevich A, et al. Modulation of (+)-[3H]pentazocine binding to guinea pig cerebellum by divalent cations. Mol Pharmacol 1992;42:882–9.

- 171. Alonso G, Phan V, Guillemain I, et al. Immunocytochemical localization of the sigma(1) receptor in the adult rat central nervous system. Neuroscience 2000;97:155–70.
- 172. Murphy DJ, Vance J. Mechanisms of lipid-body formation. Trends Biochem Sci 1999;24:109–15.
- Ohashi M, Mizushima N, Kabeya Y, Yoshimori T. Localization of mammalian NAD(P)H steroid dehydrogenase-like protein on lipid droplets. J Biol Chem 2003;278:36819–29.
- Monnet FP. Sigma-1 receptor as regulator of neuronal intracellular Ca2+: clinical and therapeutic relevance. Biol Cell 2005;97: 873–83.
- Simons K, Ikonen E. Functional rafts in cell membranes. Nature 1997;387:569–72.
- Takebayashi M, Hayashi T, Su TP. A perspective on the new mechanism of antidepressants: neuritogenesis through sigma-1 receptors. Pharmacopsychiatry 2004;37:S208–13.
- 177. Takebayashi M, Hayashi T, Su TP. Sigma-1 receptors potentiate epidermal growth factor signaling towards neuritogenesis in PC12 cells: potential relation to lipid raft reconstitution. Synapse (NY) 2004;53:90–103.
- 178. Maurice T, Su TP, Privat A. Sigmal (sigma 1) receptor agonists and neurosteroids attenuate B25-35-amyloid peptide-induced amnesia in mice through a common mechanism. Neuroscience 1998;83:413–28.
- 179. Bergeron R, de Montigny C, Debonnel G. Pregnancy reduces brain sigma receptor function. Br J Pharmacol 1999;127:1769–76.
- Debonnel G, Bergeron R, de Montigny C. Potentiation by dehydroepiandrosterone of the neuronal response to N-methyl-Daspartate in the CA3 region of the rat dorsal hippocampus: an effect mediated via sigma receptors. J Endocrinol 1996;150: S33–42.
- 181. Urani A, Roman FJ, Phan VL, et al. The antidepressant-like effect induced by sigma(1)-receptor agonists and neuroactive steroids in mice submitted to the forced swimming test. J Pharm Exp Therap 2001;298:1269–79.
- 182. Maurice T, Hiramatsu M, Itoh J, et al. Behavioral evidence for a modulating role of sigma ligands in memory processes. I: attenuation of dizocilpine (MK-801)-induced amnesia. Brain Res 2994;647:44–56.
- Maurice T, Roman FJ, Privat A. Modulation by neurosteroids of the in vivo (+)-[3H]SKF-10,047 binding to sigma 1 receptors in the mouse forebrain. J Neurosci Res 1996;46:734–43.
- Maurice T, Su TP, Parish DW, et al. PRE-084, a sigma selective PCP derivative, attenuates MK-801-induced impairment of learning in mice. Pharmacol Biochem Behav 1994;49:859–69.
- 185. Takebayashi M, Hayashi T, Su TP. Nerve growth factor-induced neurite sprouting in PC12 cells involves sigma-1 receptors: implications for antidepressants. J Pharm Exp Therap 2002;303: 1227–37.
- Hall A, Herrera Y, Ajmo Jr C, et al. Sigma receptors suppress multiple aspects of microglial activation. Glia 2009;57:744–54.
- 187. Ajmo Jr C, Vernon D, Collier L, et al. Sigma receptor activation reduces infarct size at 24 hours after permanent middle cerebral artery occlusion in rats. Curr Neurovasc Res 2006;3:89–98.
- Luscher C, Nicoll RA, Malenka RC, Muller D. Synaptic plasticity and dynamic modulation of the postsynaptic membrane. Nature Neurosci 2000;3:545–50.
- 189. Moschovos C, Kostopoulos G, Papatheodoropoulos C. Long-term potentiation of high-frequency oscillation and synaptic transmission characterize in vitro NMDA receptor-dependent epileptogenesis in the hippocampus. Neurobiol Dis 2008;29:368–80.
- Monnet FP, Maurice T. The sigmal protein as a target for the nongenomic effects of neuro(active)steroids: molecular, physiological, and behavioral aspects. J Pharmacol Sci 2006;100: 93–118.
- Carpenter C, Marks S, Watson D, Greenberg D. Dextromethorphan and dextrorphan as calcium channel antagonists. Brain Res 1988;439:372–5.
- 192. Klein M, Canoll PD, Musacchio JM. SKF 525-A and cytochrome P-450 ligands inhibit with high affinity the binding of [3H]dextromethorphan and sigma ligands to guinea pig brain. Life Sci 1991;48:543–50.
- 193. Rothman RB, Reid A, Mahboubi A, et al. Labeling by [3H]1,3di(2-tolyl)guanidine of two high affinity binding sites in guinea pig brain: evidence for allosteric regulation by calcium channel

antagonists and pseudoallosteric modulation by sigma ligands. Mol Pharmacol 1991;39:222-32.

- 194. Bergeron R, de Montigny C, Debonnel G. Biphasic effects of sigma ligands on the neuronal response to N-methyl-D-aspartate. Naunyn–Schmiedeberg's Arch Pharmacol 1995;351:252–60.
- Monnet FP, Debonnel G, Junien JL, De Montigny C. N-methyl-Daspartate-induced neuronal activation is selectively modulated by sigma receptors. Eur J Pharmacol 1990;179:441–5.
- 196. Bergeron R, Debonnel G. Effects of low and high doses of selective sigma ligands: further evidence suggesting the existence of different subtypes of sigma receptors. Psychopharmacology 1997;129:215–24.
- 197. Bergeron R, Debonnel G, De Montigny C. Modification of the N-methyl-D-aspartate response by antidepressant sigma receptor ligands. Eur J Pharmacol 1993;240:319–23.
- Hemstreet M, Matsumoto R, Bowen W, Walker J. Sigma binding parameters in developing rats predict behavioral efficacy of a sigma ligand. Brain Res 1993;627:291–8.
- 199. Kawamura K, Kimura Y, Tsukada H, et al. An increase of sigma receptors in the aged monkey brain. Neurobiol Aging 2003;24: 745–52.
- 200. Matsuno K, Senda T, Matsunaga K, Mita S. Ameliorating effects of sigma receptor ligands on the impairment of passive avoidance tasks in mice: involvement in the central acetylcholinergic system. Eur J Pharmacol 1994;261:43–51.
- Maurice T, Lockhart BP. Neuroprotective and anti-amnesic potentials of sigma (sigma) receptor ligands. Progr Neuropsychopharmacology Biol Psychiatry 1997;21:69–102.
- 202. Maurice T, Phan VL, Noda Y, et al. The attenuation of learning impairments induced after exposure to CO or trimethyltin in mice by sigma (sigma) receptor ligands involves both sigma1 and sigma2 sites. Br J Pharmacol 1999;127:335–42.
- Mayo W, Dellu F, Robel P, et al. Infusion of neurosteroids into the nucleus basalis magnocellularis affects cognitive processes in the rat. Brain Res 1993;607:324–8.
- Niitsu T, Iyo M, Hashimoto K. Sigma-1 receptor agonists as therapeutic drugs for cognitive impairment in neuropsychiatric diseases. Current Pharm Design 2012;18:875–83.
- Senda T, Matsuno K, Okamoto K, et al. Ameliorating effect of SA4503, a novel sigma 1 receptor agonist, on memory impairments induced by cholinergic dysfunction in rats. Eur J Pharmacol 1996;315:1–10.
- Pande AC, Geneve J, Scherrer B. Igmesine, a novel sigma ligand, has antidepressant properties. Int J Neuropsychopharmacol 1998; 1:S8–S9.
- 207. Odagaki Y, Toyoshima R, Yamauchi T. Lack of G protein-coupled sigma receptors in rat brain membranes: receptor-mediated highaffinity GTPase activity and [35S]GTPgammaS binding studies. J Neural Transmission (Vienna, Austria: 1996) 2005;112:873–83.
- Matsumoto RR, Bowen WD, Tom MA, et al. Characterization of two novel sigma receptor ligands: antidystonic effects in rats suggest sigma receptor antagonism. Eur J Pharmacol 1995;280: 301–10.
- Racchi M, Balduzzi C, Corsini E. Dehydroepiandrosterone (DHEA) and the aging brain: flipping a coin in the "fountain of youth". CNS Drug Rev 2003;9:21–40.
- Monnet FP, Mahe V, Robel P, Baulieu EE. Neurosteroids, via sigma receptors, modulate the [3H]norepinephrine release evoked by N-methyl-D-aspartate in the rat hippocampus. Proc Natl Acad Sci USA 1995;92:3774–8.
- 211. Matsuno K, Senda T, Kobayashi T, Mita S. Involvement of sigma 1 receptor in (+)-N-allylnormetazocine-stimulated hippocampal cholinergic functions in rats. Brain Res 1995;690: 200–6.
- Maurice T, Junien JL, Privat A. Dehydroepiandrosterone sulfate attenuates dizocilpine-induced learning impairment in mice via sigma 1-receptors. Behav Brain Res 1997;83:159–64.
- Castellano C, McGaugh J. Effects of post-training bicuculline and muscimol on retention: lack of state dependency. Behav Neural Biol 1990;54:156–64.
- Introini-Collison IB, Castellano C, McGaugh JL. Interaction of GABAergic and beta-noradrenergic drugs in the regulation of memory storage. Behav Neural Biol 1994;61:150–5.
- 215. Izquierdo I. Pharmacological evidence for a role of long-term potentiation in memory. FASEB J 1994;8:1139–45.

- 216. Rison RA, Stanton PK. Long-term potentiation and N-methyl-Daspartate receptors: foundations of memory and neurologic disease? Neurosci Biobehav Rev 1995;19:533–52
- 217. Sabbatini R. Neurons and synapses. Brain Mind 2003;17:1-6.
- Falkenstein E, Schmieding K, Lange A, et al. Localization of a putative progesterone membrane binding protein in porcine hepatocytes. Cell Mol Biol (Noisy-le-Grand, France) 1998;44: 571–8.
- 219. McCann DJ, Su TP. Haloperidol-sensitive (+)[3H]SKF-10,047 binding sites (sigma sites) exhibit a unique distribution in rat brain subcellular fractions. Eur J Pharmacol 1990;188:211–18.
- Nabeshima T, Okuyama S. [Physiological function of sigma receptors: central pharmacological effects of sigma ligands]. Nihon Shinkei Seishin Yakurigaku Zasshi 1994;14:51–76.
- 221. Su TP, London ED, Jaffe JH. Response: steroid binding at sgr-"opioid" receptors. Science (NY) 1989;246:1637–8.
- 222. Su TP, London ED, Jaffe JH. Steroid binding at sigma receptors suggests a link between endocrine, nervous, and immune systems. Science (NY) 1988;240:219–21.
- 223. Baulieu E, Robel P. Neurosteroids: a new brain function? J Steroid Biochem Mol Biol 1990;37:395–403
- 224. Corpechot C, Robel P, Axelson M, et al. Characterization and measurement of dehydroepiandrosterone sulfate in rat brain. Proc Natl Acad Sci USA 1981;78:4704–7.
- 225. Corpechot C, Synguelakis M, Talha S, et al. Pregnenolone and its sulfate ester in the rat brain. Brain Res 1983;270:119–25.
- 226. Wu FS, Gibbs TT, Farb DH. Pregnenolone sulfate: a positive allosteric modulator at the N-methyl-D-aspartate receptor. Mol Pharmacol 1991;40:333–6.
- 227. Longone P, di Michele F, D'Agati E, et al. Neurosteroids as neuromodulators in the treatment of anxiety disorders. Front Endocrinol 2011;2:55–64.
- Ramamoorthy JD, Ramamoorthy S, Mahesh VB, et al. Cocainesensitive sigma-receptor and its interaction with steroid hormones in the human placental syncytiotrophoblast and in choriocarcinoma cells. Endocrinology 1995;136:924–32.
- 229. Yamada K, Nabeshima T. Stress-induced behavioral responses and multiple opioid systems in the brain. Behav Brain Res 1995;67: 133–45.
- Yamada M, Nishigami T, Nakasho K, et al. Relationship between sigma-like site and progesterone-binding site of adult male rat liver microsomes. Hepatology (Baltimore, MD) 1994;20:1271–80.
- 231. Elfverson M, Johansson T, Zhou Q, et al. Chronic administration of the anabolic androgenic steroid nandrolone alters neurosteroid action at the sigma-1 receptor but not at the sigma-2 or NMDA receptors. Neuropharmacology 2011;61:1172–81.
- Schwarz S, Pohl P, Zhou GZ. Steroid binding at sigma-"opioid" receptors. Science (NY) 1989;246:1635–8.
- Cheney D, Uzunov D, Guidotti A. Pregnenolone sulfate antagonizes dizocilpine amnesia: role for allopregnanolone. Neuroreport 1995;6:1697–700.
- 234. Rupprecht R, Hauser CA, Trapp T, Holsboer F. Neurosteroids: molecular mechanisms of action and psychopharmacological significance. J Steroid Biochem Mol Biol 1996;56:163–8.
- Rupprecht R, Reul JM, Trapp T, et al. Progesterone receptormediated effects of neuroactive steroids. Neuron 1993;11:523–30.
- Schumacher M, Akwa Y, Guennoun R, et al. Steroid synthesis and metabolism in the nervous system: trophic and protective effects. J Neurocytol 2000;29:307–26.
- 237. Schumacher M, Weill-Engerer S, Liere P, et al. Steroid hormones and neurosteroids in normal and pathological aging of the nervous system. Progr Neurobiol 2003;71:3–29.
- McEwen BS, Coirini H, Westlind-Danielsson A, et al. Steroid hormones as mediators of neural plasticity. J Steroid Biochem Mol Biol 1991;39:223–32.
- Collier T, Waterhouse R, Kassiou M. Imaging sigma receptors: applications in drug development. Current Pharm Design 2007;13: 51–72.
- Bitran D, Hilvers R, Kellogg C. Anxiolytic effects of 3 alphahydroxy-5 alpha[beta]-pregnan-20-one: endogenous metabolites of progesterone that are active at the GABAA receptor. Brain Res 1991;561:157–61.
- 241. Brot M, Akwa Y, Purdy R, et al. The anxiolytic-like effects of the neurosteroid allopregnanolone: interactions with GABA(A) receptors. Eur J Pharmacol 1997;325:1–7.

- 242. Maurice T, Phan VL, Urani A, et al. Neuroactive neurosteroids as endogenous effectors for the sigmal (sigma1) receptor: pharmacological evidence and therapeutic opportunities. Jpn J Pharmacol 1999;81:125–55.
- Reddy DS, Kaur G, Kulkarni SK. Sigma (sigma1) receptor mediated anti-depressant-like effects of neurosteroids in the Porsolt forced swim test. Neuroreport 1998;9:3069–73.
- 244. Reddy DS, Kulkarni SK. The effects of neurosteroids on acquisition and retention of a modified passive-avoidance learning task in mice. Brain Res 1998;791:108–16.
- Urani A, Privat A, Maurice T. The modulation by neurosteroids of the scopolamine-induced learning impairment in mice involves an interaction with sigma1 (sigma1) receptors. Brain Res 1998;799: 64–77.
- Wieland S, Lan NC, Mirasedeghi S, Gee KW. Anxiolytic activity of the progesterone metabolite 5 alpha-pregnan-3 alpha-o1-20one. Brain Res 1991;565:263–8.
- Brown D. Neurons depend on astrocytes in a coculture system for protection from glutamate toxicity. Mol Cell Neurosci 1999;13: 379–89.
- 248. Weill-Engerer S, David JP, Sazdovitch V, et al. Neurosteroid quantification in human brain regions: comparison between Alzheimer's and nondemented patients. J Clin Endocrinol Metab 2002;87:5138–43.
- 249. Urani A, Romieu P, Portales-Casamar E, et al. The antidepressantlike effect induced by the sigma(1) (sigma(1)) receptor agonist igmesine involves modulation of intracellular calcium mobilization. Psychopharmacology 2002;163:26–35.
- Urani A, Romieu P, Roman FJ, Maurice T. Enhanced antidepressant effect of sigma(1) (sigma(1)) receptor agonists in beta(25–35)-amyloid peptide-treated mice. Behav Brain Res 2002;134: 239–47.
- 251. Urani A, Romieu P, Roman FJ, et al. Enhanced antidepressant efficacy of sigmal receptor agonists in rats after chronic intracerebroventricular infusion of beta-amyloid-(1–40) protein. Eur J Pharmacol 2004;486:151–61.
- Stoffel-Wagner B. Neurosteroid biosynthesis in the human brain and its clinical implications. Ann NY Acad Sci 2003;1007: 64–78.
- Wolkowitz OM, Reus VI, Roberts E, et al. Dehydroepiandrosterone (DHEA) treatment of depression. Biol Psychiatry 1997;41:311–18.
- 254. Flood J, Morley J, Roberts E. Memory-enhancing effects in male mice of pregnenolone and steroids metabolically derived from it. Proc Natl Acad Sci USA 1992;89:1567–71.
- 255. Flood J, Morley J, Roberts E. Pregnenolone sulfate enhances posttraining memory processes when injected in very low doses into limbic system structures: the amygdala is by far the most sensitive. Proc Natl Acad Sci USA 1995;92:10806–10.
- 256. Bonnet K, Brown R. Cognitive effects of DHEA replacement therapy. New York: Walter de Gruyter; 1990.
- Flood J, Roberts E. Dehydroepiandrosterone sulfate improves memory in aging mice. Brain Res 1988;448:178–81.
- 258. Orentreich N, Brind JL, Rizer RL, Vogelman JH. Age changes and sex differences in serum dehydroepiandrosterone sulfate concentrations throughout adulthood. J clin Endocrinol Metab 1984;59: 551–5.
- 259. Orentreich N, Brind JL, Vogelman JH, et al. Long-term longitudinal measurements of plasma dehydroepiandrosterone sulfate in normal men. J clin Endocrinol Metab 1992;75:1002–4.
- Vallee M, Mayo W, Darnaudery M, et al. Neurosteroids: deficient cognitive performance in aged rats depends on low pregnenolone sulfate levels in the hippocampus. Proc Natl Acad Sci USA 1997; 94:14865–70.
- 261. Kurata K, Takebayashi M, Morinobu S, Yamawaki S. betaestradiol, dehydroepiandrosterone, and dehydroepiandrosterone sulfate protect against N-methyl-D-aspartate-induced neurotoxicity in rat hippocampal neurons by different mechanisms. J Pharm Exp Therap 2004;311:237–45.
- 262. Hayashi T, Tsai SY, Mori T, et al. Targeting ligand-operated chaperone sigma-1 receptors in the treatment of neuropsychiatric disorders. Expert Opin Therap Targets 2011;15:557–77.
- 263. Noda Y, Kamei H, Kamei Y, et al. Neurosteroids ameliorate conditioned fear stress: an association with sigma receptors. Neuropsychopharmacology 2000;23:276–84.

- Akunne H, Whetzel S, Wiley J, et al. The pharmacology of the novel and selective sigma ligand, PD 144418. Neuropharmacology 1997;36:51–62.
- Farb D, Gibbs T, Wu F, Gyenes M, et al. Steroid modulation of amino acid neurotransmitter receptors. Adv Biochem Psychopharmacol 1992;47:119–31.
- Gasior M, Carter R, Witkin J. Neuroactive steroids: potential therapeutic use in neurological and psychiatric disorders. Trends Pharmacol Sci 1999;20:107–12.
- 267. Takebayashi M, Kagaya A, Uchitomi Y, et al. Differential regulation by pregnenolone sulfate of intracellular Ca2+ increase by amino acids in primary cultured rat cortical neurons. Neurochem Int 1998;32:205–11.
- Martins RA, Linden R, Dyer MA. Glutamate regulates retinal progenitors cells proliferation during development. Eur J Neurosci 2006;24:969–80.
- Martins RA, Pearson RA. Control of cell proliferation by neurotransmitters in the developing vertebrate retina. Brain Res 2008;1192:37–60.
- Smith SS. Progesterone administration attenuates excitatory amino acid responses of cerebellar Purkinje cells. Neuroscience 1991;42: 309–20.
- Belelli D, Lambert J. Neurosteroids: endogenous regulators of the GABA(A) receptor. Nat Rev Neurosci 2005;6:565–75.
- 272. Harrison N, Simmonds M. Modulation of the GABA receptor complex by a steroid anaesthetic. Brain Res 1984;323:287–92.
- Lambert JJ, Belelli D, Hill-Venning C, et al. Neurosteroid modulation of native and recombinant GABAA receptors. Cell Mol Neurobiol 1996;16:155–74.
- Mathis C, Paul SM, Crawley JN. The neurosteroid pregnenolone sulfate blocks NMDA antagonist-induced deficits in a passive avoidance memory task. Psychopharmacology 1994;116:201–6.
- 275. Mathis C, Vogel E, Cagniard B, et al. The neurosteroid pregnenolone sulfate blocks deficits induced by a competitive NMDA antagonist in active avoidance and lever-press learning tasks in mice. Neuropharmacology 1996;35:1057–64.
- 276. Callachan H, Cottrell G, Hather N, et al. Modulation of the GABAA receptor by progesterone metabolites. Proc R Soc Lond ser B 1987;231:359–69.
- Shu HJ, Eisenman LN, Jinadasa D, et al. Slow actions of neuroactive steroids at GABAA receptors. J Neurosci 2004;24: 6667–75.
- Hong W, Nuwayhid SJ, Werling LL. Modulation of bradykinininduced calcium changes in SH-SY5Y cells by neurosteroids and sigma receptor ligands via a shared mechanism. Synapse 2004;54: 102–10.
- Bukusoglu C, Sarlak F. Pregnenolone sulfate increases intracellular Ca2+ levels in a pituitary cell line. Eur J Pharmacol 1996; 298:79–85.
- Fahey J, Lindquist D, Pritchard G, Miller L. Pregnenolone sulfate potentiation of NMDA-mediated increases in intracellular calcium in cultured chick cortical neurons. Brain Res 1995;669:183–8.
- French-Mullen J, Danks P, Spence K. Neurosteroids modulate calcium currents in hippocampal CA1 neurons via a pertussis toxin-sensitive G-protein-coupled mechanism. J Neurosci 1994; 14:1963–77.
- French-Mullen J, Spence K. Neurosteroids block Ca2+ channel current in freshly isolated hippocampal CA1 neurons. Eur J Pharmacol 1991;202:269–72.
- 283. Maurice T. Neurosteroids and sigma1 receptors, biochemical and behavioral relevance. Pharmacopsychiatry 2004;37:S171–82.
- Compagnone N, Mellon S. Dehydroepiandrosterone: a potential signalling molecule for neocortical organization during development. Proc Natl Acad Sci USA 1998;95:4678–83.
- Dubrovsky B. Neurosteroids, neuroactive steroids, and symptoms of affective disorders. Pharmacol Biochem Behav 2006;84: 644–55.
- Irwin RP, Lin SZ, Rogawski MA, et al. Steroid potentiation and inhibition of N-methyl-D-aspartate receptor-mediated intracellular Ca++ responses: structure-activity studies. J Pharm Exp Therap 1994;271:677–82.
- Irwin RP, Maragakis NJ, Rogawski MA, et al. Pregnenolone sulfate augments NMDA receptor mediated increases in intracellular Ca2+ in cultured rat hippocampal neurons. Neurosci Lett 1992;141:30–4.

- 372 C. G. Rousseaux & S. F. Greene
- Wakerley JB, Richardson CM. The neurosteroid pregnenolone sulphate enhances NMDA-induced phasic firing of vasopressin neurones in the rat supraoptic nucleus. Neurosci Lett 1997;226: 123–6.
- Park-Chung M, Wu FS, Farb DH. 3 alpha-Hydroxy-5 betapregnan-20-one sulfate: a negative modulator of the NMDAinduced current in cultured neurons. Mol Pharmacol 1994;46: 146–50.
- Park-Chung M, Wu FS, Purdy RH, et al. Distinct sites for inverse modulation of N-methyl-D-aspartate receptors by sulfated steroids. Mol Pharmacol 1997;52:1113–23.
- 291. Hayashi T, Kagaya A, Takebayashi M, et al. Modulation by sigma ligands of intracellular free Ca++ mobilization by N-methyl-D-aspartate in primary culture of rat frontal cortical neurons. J Pharm Exp Therap 1995;275:207–14.
- 292. Phan VL, Miyamoto Y, Nabeshima T, Maurice T. Age-related expression of sigmal receptors and antidepressant efficacy of a selective agonist in the senescence-accelerated (SAM) mouse. J Neurosci Res 2005;79:561–72.
- 293. Phan VL, Su TP, Privat A, Maurice T. Modulation of steroidal levels by adrenalectomy/castration and inhibition of neurosteroid synthesis enzymes affect sigma1 receptor-mediated behaviour in mice. Eur J Neurosci 1999;11:2385–96.
- 294. Phan VL, Urani A, Romieu P, Maurice T. Strain differences in sigma(1) receptor-mediated behaviours are related to neurosteroid levels. Eur J Neurosci 2002;15:1523–34.
- 295. Phan VL, Urani A, Sandillon F, et al. Preserved sigma1 (sigma1) receptor expression and behavioral efficacy in the aged C57BL/6 mouse. Neurobiol Aging 2003;24:865–81.
- 296. Danysz W, Wroblewski J, Brooker G, Costa E. Modulation of glutamate receptors by phencyclidine and glycine in the rat cerebellum: cGMP increase in vivo. Brain Res 1989;479: 270–6.
- Swegle JM, Logemann C. Management of common opioidinduced adverse effects. Am Fam Phys 2006;74:1347–54.
- 298. Tam SW. Naloxone-inaccessible sigma receptor in rat central nervous system. Proc Natl Acad Sci USA 1983;80:6703–7.
- 299. May EL, Aceto MD, Bowman ER, et al. Antipodal alpha-N-(methyl through decyl)-N-normetazocines (5,9 alpha-dimethyl-2'hydroxy-6,7-benzomorphans): in vitro and in vivo properties. J Med Chem 1994;37:3408–18.
- Young GA, Hudson GM, Stamidis H, Steinfels GF. Interactions between U-50,488H and sigma receptor antagonists: EEG, EEG power spectral and behavioral correlates. Eur J Pharmacol 1993; 231:473–6.
- 301. Grisel J, Allen S, Nemmani K, et al. The influence of dextromethorphan on morphine analgesia in Swiss Webster mice is sex-specific. Pharmacol Biochem Behav 2005;81:131–8.
- Mei J, Pasternak GW. Modulation of brainstem opiate analgesia in the rat by sigma 1 receptors: a microinjection study. J Pharm Exp Therap 2007;322:1278–85.
- Musacchio JM. The psychotomimetic effects of opiates and the sigma receptor. Neuropsychopharmacology 1990;3:191–200.
- Hiramatsu M, Hoshino T. Improvement of memory impairment by (+)- and (-)-pentazocine via sigma, but not kappa opioid receptors. Brain Res 2005;1057:72–80.
- 305. Hiramatsu M, Hoshino T, Kanematsu K. Pharmacological characterization of the ameliorating effect on short-term memory impairment and antinociceptive effect of KT-90 in mice. Behav Brain Res 2005;160:374–81.
- 306. Hiramatsu M, Mizuno N, Kanematsu K. Pharmacological characterization of the ameliorating effect on learning and memory impairment and antinociceptive effect of KT-95 in mice. Behav Brain Res 2006;167:219–25.
- 307. Kohler C. The distribution of serotonin binding sites in the hippocampal region of the rat brain: an autoradiographic study. Neuroscience 1984;13:667–80.
- Heninger G, Delgado P, Charney D, et al. Tryptophan-deficient diet and amino acid drink deplete plasma tryptophan and induce a relapse of depression in susceptible patients. J Chem Neuroanat 1992;5:347–8.
- Andrade R, Nicoll R. Pharmacologically distinct actions of serotonin on single pyramidal neurones of the rat hippocampus recorded in vitro. J Physiol 1987;394:99–124.

- Blier P, De Montigny C. Electrophysiological investigations on the effect of repeated zimelidine administration on serotonergic neurotransmission in the rat. J Neurosci 1983;3:1270–8.
- 311. Blier P, de Montigny C. Serotoninergic but not noradrenergic neurons in rat central nervous system adapt to long-term treatment with monoamine oxidase inhibitors. Neuroscience 1985;16: 949–55.
- 312. Blier P, de Montigny C, Tardif D. Effects of the two antidepressant drugs mianserin and indalpine on the serotonergic system: single-cell studies in the rat. Psychopharmacology 1984;84:242–9.
- 313. Chaput Y, de Montigny C, Blier P. Effects of a selective 5-HT reuptake blocker, citalopram, on the sensitivity of 5-HT autoreceptors: electrophysiological studies in the rat brain. Naunyn–Schmiedeberg's Arch Pharmacol 1986;333:342–8.
- 314. Campbell B, Scherz M, Keana J, Weber E. Sigma receptors regulate contractions of the guinea pig ileum longitudinal muscle/ myenteric plexus preparation elicited by both electrical stimulation and exogenous serotonin. J Neurosci 1989;9:3380–91.
- 315. Bermack J, Haddjeri N, Debonnel G. Effects of the potential antidepressant OPC-14523 [1-[3-[4-(3-chlorophenyl)-1piperazinyl]propyl]-5-methoxy-3,4-dihydro-2-quinolino ne monomethanesulfonate] a combined sigma and 5-HT1A ligand: modulation of neuronal activity in the dorsal raphe nucleus. J Pharm Exp Therap 2004;310:578–83.
- 316. Blier P, Chaput Y, de Montigny C. Long-term 5-HT reuptake blockade, but not monoamine oxidase inhibition, decreases the function of terminal 5-HT autoreceptors: an electrophysiological study in the rat brain. Naunyn–Schmiedeberg's Arch Pharmacol 1998;337:246–54.
- 317. Chaput Y, de Montigny C, Blier P. Presynaptic and postsynaptic modifications of the serotonin system by long-term administration of antidepressant treatments. An in vivo electrophysiologic study in the rat. Neuropsychopharmacology 1991;5:219–29.
- Kia HK, Miquel MC, Brisorgueil MJ, et al. Immunocytochemical localization of serotonin1A receptors in the rat central nervous system. J Compar Neurobiol 1996;365:289–305.
- Kreiss DS, Lucki I. Effects of acute and repeated administration of antidepressant drugs on extracellular levels of 5-hydroxytryptamine measured in vivo. J Pharm Exp Therap 1995;274:866–76.
- Le Poul E, Boni C, Hanoun N, et al. Differential adaptation of brain 5-HT1A and 5-HT1B receptors and 5-HT transporter in rats treated chronically with fluoxetine. Neuropharmacology 2000;39: 110–22.
- 321. Le Poul E, Laaris N, Doucet E, et al. Early desensitization of somato-dendritic 5-HT1A autoreceptors in rats treated with fluoxetine or paroxetine. Naunyn–Schmiedeberg's Arch Pharmacol 1995;352:141–8.
- 322. Finn D, Phillips T, Okorn D, et al. Rewarding effect of the neuroactive steroid 3 alpha-hydroxy-5 alpha-pregnan-20-one in mice. Pharmacol Biochem Behav 1997;56:261–4.
- 323. Oshiro Y, Sakurai Y, Sato S, et al. 3,4-dihydro-2(1H)-quinolinone as a novel antidepressant drug: synthesis and pharmacology of 1-[3-[4-(3-chlorophenyl)-1-piperazinyl]propyl]-3,4dihydro-5methoxy-2(1H)-quinolinone and its derivatives. J Med Chem 2000;43:177–89.
- 324. Tottori K, Miwa T, Uwahodo Y, et al. Antidepressant-like responses to the combined sigma and 5-HT1A receptor agonist OPC-14523. Neuropharmacology 2001;41:976–88.
- 325. Tottori K, Nakai M, Uwahodo Y, et al. Attenuation of scopolamine-induced and age-associated memory impairments by the sigma and 5-hydroxytryptamine(1A) receptor agonist OPC-14523 (1-[3-[4-(3-chlorophenyl)-1-piperazinyl]propyl]-5-methoxy-3,4-dihydro-2[1H]-quino linone monomethanesulfonate). J Pharm Exp Therap 2002;301:249–57.
- Bermack J, Debonnel G. Modulation of serotonergic neurotransmission by short- and long-term treatments with sigma ligands. Br J Pharmacol 2001;134:691–9.
- 327. Marcinkiewicz M, Verge D, Gozlan H, et al. Autoradiographic evidence for the heterogeneity of 5-HT1 sites in the rat brain. Brain Res 1984;291:159–63.
- Robichaud M, Beauchemin V, Lavoie N, et al. Effects of bilateral olfactory bulbectomy on N-methyl-D-aspartate receptor function: autoradiographic and behavioral studies in the rat. Synapse (NY) 2001;42:95–103.

- 329. Celada P, Puig M, Casanovas J, et al. Control of dorsal raphe serotonergic neurons by the medial prefrontal cortex: Involvement of serotonin-1A, GABA(A), and glutamate receptors. J Neurosci 2001;21:9917–29.
- 330. Aghajanian G, Wang R. Habenular and other midbrain raphe afferents demonstrated by a modified retrograde tracing technique. Brain Res 1997;122:229–42.
- Aghajanian G, Wang R, Baraban J. Serotonergic and nonserotonergic neurons of the dorsal raphe: reciprocal changes in firing induced by peripheral nerve stimulation. Brain Res 1978; 153:169–75.
- Ferraro G, Montalbano M, Sardo P, La Grutta V. Lateral habenular influence on dorsal raphe neurons. Brain Res Bull 1996;41:47–52.
- 333. Hajos M, Gartside S, Varga V, Sharp T. In vivo inhibition of neuronal activity in the rat ventromedial prefrontal cortex by midbrain-raphe nuclei: role of 5-HT1A receptors. Neuropharmacology 2003;45:72–81.
- 334. Kalen P, Karlson M, Wiklund L. Possible excitatory amino acid afferents to nucleus raphe dorsalis of the rat investigated with retrograde wheat germ agglutinin and D-[3H]aspartate tracing. Brain Res 1985;360:285–97.
- 335. Peyron C, Petit JM, Rampon C, et al. Forebrain afferents to the rat dorsal raphe nucleus demonstrated by retrograde and anterograde tracing methods. Neuroscience 1998;82:443–68.
- 336. Varga V, Kocsis B, Sharp T. Electrophysiological evidence for convergence of inputs from the medial prefrontal cortex and lateral habenula on single neurons in the dorsal raphe nucleus. Eur J Neurosci 2003;17:280–6.
- 337. Ceci A, Baschirotto A, Borsini F. The inhibitory effect of 8-OH-DPAT on the firing activity of dorsal raphe serotoninergic neurons in rats is attenuated by lesion of the frontal cortex. Neuropharmacology 1994;33:709–13.
- 338. Hajos M, Richards C, Szekely A, Sharp T. An electrophysiological and neuroanatomical study of the medial prefrontal cortical projection to the midbrain raphe nuclei in the rat. Neuroscience 1998;87:95–108.
- O'Hearn E, Molliver ME. Organization of raphe-cortical projections in rat: a quantitative retrograde study. Brain Res Bull 1984; 13:709–26.
- Varga V, Szekely AD, Csillag A, et al. Evidence for a role of GABA interneurones in the cortical modulation of midbrain 5-hydroxytryptamine neurones. Neuroscience 2001;106:783–92.
- Varga EV, Rubenzik MK, Stropova D, et al. Converging protein kinase pathways mediate adenylyl cyclase superactivation upon chronic delta-opioid agonist treatment. J Pharm Exp Therap 2003; 306:109–15.
- 342. Kobayashi T, Matsuno K, Murai M, Mita S. Sigma 1 receptor subtype is involved in the facilitation of cortical dopaminergic transmission in the rat brain. Neurochem Res 1997;22:1105–9.
- 343. Barrot M, Vallee M, Gingras M, et al. The neurosteroid pregnenolone sulphate increases dopamine release and the dopaminergic response to morphine in the rat nucleus accumbens. Eur J Neurosci 1999;11:3757–60.
- 344. Ohi K, Hashimoto R, Yasuda Y, et al. The SIGMAR1 gene is associated with a risk of schizophrenia and activation of the prefrontal cortex. Progr Neuro-psychopharmacol Biol Psychiatry 2011;35:1309–15.
- Bonomo V, Fogliani A. Citalopram and haloperidol for psychotic depression. Am J Psychiatry 2000;157:1706–7.
- 346. Patrick SL, Walker JM, Perkel JM, et al. Increases in rat striatal extracellular dopamine and vacuous chewing produced by two sigma receptor ligands. Eur J Pharmacol 1993; 231:243–9.
- 347. Paquette MA, Foley K, Brudney EG, et al. The sigma-1 antagonist BMY-14802 inhibits L-DOPA-induced abnormal involuntary movements by a WAY-100635-sensitive mechanism. Psychopharmacology 2009;204:743–54.
- Steinfels GF, Tam SW. Selective sigma receptor agonist and antagonist affect dopamine neuronal activity. Eur J Pharmacol 1989;163:167–70.
- Weatherspoon JK, Gonzalez-Alvear GM, Frank AR, Werling LL. Regulation of [3H] dopamine release from mesolimbic and mesocortical areas of guinea pig brain by sigma receptors. Schizophr Res 1996;21:51–62.

- 350. Gronier B, Debonnel G. Involvement of sigma receptors in the modulation of the glutamatergic/NMDA neurotransmission in the dopaminergic systems. Eur J Pharmacol 1999;368:183–96.
- Gudelsky G. Biphasic effect of sigma receptor ligands on the extracellular concentration of dopamine in the striatum of the rat. J Neural Transmission (Vienna, Austria: 1996) 1999;106:849–56.
- 352. Peeters M, Romieu P, Maurice T, et al. Involvement of the sigma 1 receptor in the modulation of dopaminergic transmission by amantadine. Eur J Neurosci 2004;19:2212–20.
- Sershen H, Hashim A, Lajtha A. The effect of ibogaine on sigmaand NMDA-receptor-mediated release of [3H]dopamine. Brain Res Bull 1996;40:63–7.
- Derbez A, Mody R, Werling L. Sigma(2)-receptor regulation of dopamine transporter via activation of protein kinase C. J Pharm Exp Therap 2002;301:306–14.
- 355. von Ruden L, Neher E. A Ca-dependent early step in the release of catecholamines from adrenal chromaffin cells. Science (NY) 1993;262:1061–5.
- Sagi N, Yamamoto H, Yamamoto T, et al. Possible expression of a sigma 1 site in rat pheochromocytoma (PC12) cells. Eur J Pharmacol 1996;304:185–90.
- 357. Matsuno K, Matsunaga K, Mita S. Increase of extracellular acetylcholine level in rat frontal cortex induced by (+)N-allylnormetazocine as measured by brain microdialysis. Brain Res 1992;575:315–19.
- 358. Matsuno K, Matsunaga K, Senda T, Mita S. Increase in extracellular acetylcholine level by sigma ligands in rat frontal cortex. J Pharm Exp Therap 1993;265:851–9.
- 359. Matsuno K, Senda T, Kobayashi T, et al. SA4503, a novel cognitive enhancer, with sigma 1 receptor agonistic properties. Behav Brain Res 1997;83:221–4.
- Lesage AS, De Loore KL, Peeters L, Leysen JE. Neuroprotective sigma ligands interfere with the glutamate-activated NOS pathway in hippocampal cell culture. Synapse (NY) 1995;20:156–64.
- Dirnagl U, Iadecola C, Moskowitz M. Pathobiology of ischaemic stroke: an integrated view. Trends Neurosci 1999;22:391–7.
- Iadecola C, Zhang F, Casey R, et al. Delayed reduction of ischemic brain injury and neurological deficits in mice lacking the inducible nitric oxide synthase gene. J Neurosci 1997;17:9157–64.
- Samdani AF, Dawson TM, Dawson VL. Nitric oxide synthase in models of focal ischemia. Stroke 1997;28:1283–8.
- 364. Iadecola C. Bright and dark sides of nitric oxide in ischemic brain injury. Trends Neurosci 1997;20:132–9.
- Vagnerova K, Hurn PD, Bhardwaj A, Kirsch JR. Sigma 1 receptor agonists act as neuroprotective drugs through inhibition of inducible nitric oxide synthase. Anesth Analg 2006;103:430–4.
- 366. Jiang MH, Kaku T, Hada J, Hayashi Y. 7-Nitroindazole reduces nitric oxide concentration in rat hippocampus after transient forebrain ischemia. Eur J Pharmacol 1999;380:117–21.
- 367. Nanri K, Montecot C, Springhetti V, et al. The selective inhibitor of neuronal nitric oxide synthase, 7-nitroindazole, reduces the delayed neuronal damage due to forebrain ischemia in rats. Stroke 1998;29:1248–53; discussion 1253–1244.
- 368. Bhardwaj A, Sawada M, London E, et al. Potent sigma1-receptor ligand 4-phenyl-1-(4-phenylbutyl) piperidine modulates basal and N-methyl-D-aspartate-evoked nitric oxide production in vivo. Stroke 1998;29:2404–10; discussion 2411.
- 369. Goyagi T, Goto S, Bhardwaj A, et al. Neuroprotective effect of sigma(1)-receptor ligand 4-phenyl-1-(4-phenylbutyl) piperidine (PPBP) is linked to reduced neuronal nitric oxide production. Stroke 2001;32:1613–20.
- 370. Allahtavakoli M, Jarrott B. Sigma-1 receptor ligand PRE-084 reduced infarct volume, neurological deficits, pro-inflammatory cytokines and enhanced anti-inflammatory cytokines after embolic stroke in rats. Brain Res Bull 2011;85:219–24.
- Collingridge G, Bliss T. Memories of NMDA receptors and LTP. Trends Neurosci 1995;18:54–6.
- 372. Collingridge G, Singer W. Excitatory amino acid receptors and synaptic plasticity. Trends Pharmacol Sci 1990;11:290–6.
- Parsons CG, Danysz W, Quack G. Glutamate in CNS disorders as a target for drug development: an update. Drug News Perspect 1998;11:523–69.
- Rousseaux CG. A review of Glutamate receptors I: current understanding of their biology. Japan J Toxicol Pathol 2008;21: 21–51.

- 375. Rousseaux CG. A review of Glutamate receptors II: Pathophysiology and pathology. Japan J Toxicol Pathol 2008;21: 133–73.
- 376. Bermack J, Debonnel G. Distinct modulatory roles of sigma receptor subtypes on glutamatergic responses in the dorsal hippocampus. Synapse (NY) 2005;55:37–44.
- 377. Bermack J, Debonnel G. The role of sigma receptors in depression. J Pharmacol Sci 2005;97:317–36.
- 378. Dong H, Hayar A, Ennis M. Activation of group I metabotropic glutamate receptors on main olfactory bulb granule cells and periglomerular cells enhances synaptic inhibition of mitral cells. J Neurosci 2007;27:5654–63.
- 379. Fitsanakis V, Aschner M. The importance of glutamate, glycine, and gamma-aminobutyric acid transport and regulation in manganese, mercury and lead neurotoxicity. Toxicol Appl Pharmacol 2005;204:343–54.
- 380. Guitart X, Mendez R, Ovalle S, et al. Regulation of ionotropic glutamate receptor subunits in different rat brain areas by a preferential sigma(1) receptor ligand and potential atypical antipsychotic. Neuropsychopharmacology 2000;23:539–46.
- Asztely F, Erdemli G, Kullmann D. Extrasynaptic glutamate spillover in the hippocampus: dependence on temperature and the role of active glutamate uptake. Neuron 1997;18:281–93.
- Dingledine R, Borges K, Bowie D, Traynelis S. The glutamate receptor ion channels. Pharmacol Rev 1999;51:7–61.
- Frye C, Sturgis J. Neurosteroids affect spatial/reference, working, and long-term memory of female rats. Neurobiol Learn Memory 1995;64:83–96.
- Lipton SA. Prospects for clinically tolerated NMDA antagonists: open-channel blockers and alternative redox states of nitric oxide. Trends Neurosci 1993;16:527–32.
- Lipton SA, Gendelman HE. Seminars in medicine of the Beth Israel Hospital, Boston. Dementia associated with the acquired immunodeficiency syndrome. N Eng J Med 1995;332:934–40.
- Lipton SA, Rosenberg PA. Excitatory amino acids as a final common pathway for neurologic disorders. N Eng J Med 1994; 330:613–22.
- 387. Meldrum BS. The role of glutamate in epilepsy and other CNS disorders. Neurology 1994;44:S14–23.
- Miller S, Kesslak JP, Romano C, Cotman CW. Roles of metabotropic glutamate receptors in brain plasticity and pathology. Ann NY Acad Sci 1995;757:460–74.
- Ozawa S, Kamiya H, Tsuzuki K. Glutamate receptors in the mammalian central nervous system. Progr Neurobiol 1998;54: 581–618.
- Schoepp DD. Novel functions for subtypes of metabotropic glutamate receptors. Neurochem Int 1994;24:439–49.
- Gasic G, Hollmann M. Mol Neurobiol of glutamate receptors. Annu Rev Physiol 1992;54:507–36.
- 392. Hollmann M, Heinemann S. Cloned glutamate receptors. Annu Rev Neurosci 1994;17:31–108.
- 393. Hollmann M, O'Shea-Greenfield A, Rogers S, Heinemann S. Cloning by functional expression of a member of the glutamate receptor family. Nature 1989;342:643–8.
- 394. McBain CJ, Mayer ML. N-methyl-D-aspartic acid receptor structure and function. Physiol Rev 1994;74:723–60.
- Young VR, Ajami AM. Glutamate: an amino acid of particular distinction. J Nutr 2000;130:892s–900s.
- Marc RE, Liu WL, Kalloniatis M, et al. Patterns of glutamate immunoreactivity in the goldfish retina. J Neurosci 1990;10: 4006–34.
- 397. Aizenman E, Frosch M, Lipton S. Responses mediated by excitatory amino acid receptors in solitary retinal ganglion cells from rat. J Physiol 1988;396:75–91.
- Sasaki T, Kaneko A. L-Glutamate-induced responses in OFF-type bipolar cells of the cat retina. Vis Res 1996;36:787–95.
- 399. Zhou ZJ, Fain GL, Dowling JE. The excitatory and inhibitory amino acid receptors on horizontal cells isolated from the white perch retina. J Neurophysiol 1993;70:8–19.
- Jackson PF, Slusher BS. Design of NAALADase inhibitors: a novel neuroprotective strategy. Curr Med Chem 2001;8:949–57.
- 401. Neale JH, Bzdega T, Wroblewska B. N-Acetylaspartylglutamate: the most abundant peptide neurotransmitter in the mammalian central nervous system. J Neurochem 2000;75:443–52.
- 402. Danbolt N. Glutamate uptake. Progr Neurobiol 2001;65:1-105.

- 403. Brosnan J. Glutamate, at the interface between amino acid and carbohydrate metabolism. J Nutr 2000;130:988s–90s.
- Skuza G. Pharmacology of sigma (sigma) receptor ligands from a behavioral perspective. Current Pharm Design 2012;18:863–74.
- 405. Ankarcrona M, Dypbukt J, Bonfoco E, et al. Glutamate-induced neuronal death: a succession of necrosis or apoptosis depending on mitochondrial function. Neuron 1995;15:961–73.
- 406. Lipton SA, Nicotera P. Calcium, free radicals and excitotoxins in neuronal apoptosis. Cell Calcium 1998;23:165–71.
- 407. Nicotera P, Lipton SA. Excitotoxins in neuronal apoptosis and necrosis. J Cerebral Blood Flow Metab 1999;19:583–91.
- Obrenovitch TP, Urenjak J. Altered glutamatergic transmission in neurological disorders: from high extracellular glutamate to excessive synaptic efficacy. Progr Neurobiol 1997;51:39–87.
- 409. Rosin C, Bates TE, Skaper SD. Excitatory amino acid induced oligodendrocyte cell death in vitro: receptor-dependent and -independent mechanisms. J Neurochem 2004;90:1173–85.
- Truong DD, Bhidayasiri R, Wolters E. Management of non-motor symptoms in advanced Parkinson disease. J Neurol Sci 2008;266: 216–28.
- 411. Underhill SM, Goldberg MP. Hypoxic injury of isolated axons is independent of ionotropic glutamate receptors. Neurobiol Dis 2007;25:284–90.
- 412. Choi D. Excitotoxic cell death. J Neurobiol 1992;23:1261-76.
- Ozono R, O'Connell DP, Vaughan C, et al. Expression of the subtype 1A dopamine receptor in the rat heart. Hypertension 1996;27:693–703.
- 414. Griesmaier E, Posod A, Gross M, et al. Neuroprotective effects of the sigma-1 receptor ligand PRE-084 against excitotoxic perinatal brain injury in newborn mice. Exp Neurol 2012;237:388–95.
- 415. Bruni J, Bose R, Pinsky C, Glavin G. Circumventricular organ origin of domoic acid-induced neuropathology and toxicology. Brain Res Bull 1991;26:419–24.
- Choi D. Calcium: still center-stage in hypoxic-ischemic neuronal death. Trends Neurosci 1995;18:58–60.
- 417. Choi D. Glutamate neurotoxicity and diseases of the nervous system. Neuron 1988;1:623–34.
- 418. Gill S, Pulido O. Glutamate receptors in peripheral tissues: current knowledge, future research, and implications for toxicology. Toxicol Pathol 2001;29:208–23.
- 419. Meldrum BS. Glutamate as a neurotransmitter in the brain: review of physiology and pathology. J Nutr 2000;130:1007s–15s.
- 420. Price MT, Olney JW, Cicero TJ. Acute elevations of serum luteinizing hormone induced by kainic acid, N-methyl aspartic acid or homocysteic acid. Neuroendocrinology 1978;26: 352–8.
- Conn P, Pin J. Pharmacology and functions of metabotropic glutamate receptors. Annu Rev Pharmacol Toxicol 1997;37: 205–7.
- 422. Cunningham M, Ferkany J, Enna S. Excitatory amino acid receptors: a gallery of new targets for pharmacological intervention. Life Sci 1994;54:135–48.
- Boldyrev A, Bulygina E, Makhro A. Glutamate receptors modulate oxidative stress in neuronal cells: a mini-review. Neurotoxic Res 2004;6:581–7.
- 424. Singh P, Mann KA, Mangat HK, Kaur G. Prolonged glutamate excitotoxicity: effects on mitochondrial antioxidants and antioxidant enzymes. Mol Cell Biochem 2003;243:139–45.
- 425. Beal M. Aging, energy, and oxidative stress in neurodegenerative diseases. Ann Neurol 1995;38:357–66.
- Li S, Stys PK. Mechanisms of ionotropic glutamate receptormediated excitotoxicity in isolated spinal cord white matter. J Neurosci 2000;20:1190–8.
- 427. Blaylock R. Excitotoxicity: a possible central mechanism in fluoride neurotoxicity. Fluoride 2004;37:301–14.
- 428. Butcher S, Sandberg M, Hagberg H, Hamberger A. Cellular origins of endogenous amino acids released into the extracellular fluid of the rat striatum during severe insulin-induced hypoglycemia. J Neurochem 1987;48:722–8.
- 429. Krogsgaard-Larsen P, Hansen JJ. Naturally-occurring excitatory amino acids as neurotoxins and leads in drug design. Toxicol Lett 1992;64:409–16.
- Peng YG, Taylor TB, Finch RE, et al. Neuroexcitatory and neurotoxic actions of the amnesic shellfish poison, domoic acid. Neuroreport 1994;5:981–5.

- 431. Perl TM, Bedard L, Kosatsky T, et al. Amnesic shellfish poisoning: a new clinical syndrome due to domoic acid. Can Dis Week Rep 1990;16:7–8.
- 432. Rockhold RW, Acuff CG, Clower BR. Excitotoxin-induced myocardial necrosis. Eur J Pharmacol 1989;166:571–6.
- 433. Teitelbaum JS, Zatorre RJ, Carpenter S, et al. Neurologic sequelae of domoic acid intoxication due to the ingestion of contaminated mussels. N Eng J Med 1990;322:1781–7.
- 434. Tryphonas L, Truelove J, Iverson F, et al. Neuropathology of experimental domoic acid poisoning in non-human primates and rats. Can Dis Week Rep 1990;16:75–81.
- 435. Watters MR. Organic neurotoxins in seafoods. Clin Neurol Neurosurg 1995;97:119–24.
- 436. Winter CR, Baker RC. L-glutamate-induced changes in intracellular calcium oscillation frequency through non-classical glutamate receptor binding in cultured rat myocardial cells. Life Sci 1995;57:1925–34.
- 437. Zautcke JL, Schwartz JA, Mueller EJ. Chinese restaurant syndrome: a review. Ann Emerg Med 1986;15:1210–13.
- 438. Erdo S. Excitatory amino acid receptors in the mammalian periphery. Trends Pharmacol Sci 1991;12:426–9.
- Iverson F, Truelove J, Tryphonas L, Nera EA. The toxicology of domoic acid administered systemically to rodents and primates. Can Dis Week Rep 1990;16:15–8; discussion 18–19.
- 440. Olney JW. Glutamate: a neurotoxic transmitter. J Child Neurol 1989;4:218–26.
- Meldrum B. Amino acids as dietary excitotoxins: a contribution to understanding neurodegenerative disorders. Brain Res Brain Res Rev 1993;18:293–314.
- 442. Truelove J, Mueller R, Pulido O, Iverson F. Subchronic toxicity study of domoic acid in the rat. Food Chem Toxicol 1996;34: 525–9.
- 443. Kim HC, Jhoo WK, Kim WK, et al. Carbetapentane attenuates kainate-induced seizures via sigma-1 receptor modulation. Life Sci 2001;69:915–22.
- DeCoster M, Klette K, Knight E, Tortella F. Sigma receptormediated neuroprotection against glutamate toxicity in primary rat neuronal cultures. Brain Res 1995;671:45–53.
- 445. Farber N, Jiang X, Heinkel C, Nemmers B. Antiepileptic drugs and agents that inhibit voltage-gated sodium channels prevent NMDA antagonist neurotoxicity. Mol Psychiatry 2002;7:726–33.
- 446. Nishikawa J, Saito K, Sasaki M, et al. Molecular cloning and functional characterization of a novel nuclear receptor similar to an embryonic benzoate receptor BXR. Biochem Biophys Res Commun 2000;277:209–15.
- 447. Beal M. Excitotoxicity and nitric oxide in Parkinson's disease pathogenesis. Ann Neurol 1998;44:S110–14.
- 448. Flor P, Battaglia G, Nicoletti F, et al. Neuroprotective activity of metabotropic glutamate receptor ligands. Adv Exp Med Biol 2002;513:197–23.
- Danysz W, Parsons C. Glycine and N-methyl-D-aspartate receptors: physiological significance and possible therapeutic applications. Pharmacol Rev 1998;50:597–664.
- 450. Shaw PJ, Ince PG. Glutamate, excitotoxicity and amyotrophic lateral sclerosis. J Neurol 1997;244:S3–14.
- 451. Starr MS. Antiparkinsonian actions of glutamate antagonistsalone and with L-DOPA: a review of evidence and suggestions for possible mechanisms. J Neural Transm Parkinson's Dis Demen Sect 1995;10:141–85.
- 452. Al-Saif A, Al-Mohanna F, Bohlega S. A mutation in sigma-1 receptor causes juvenile amyotrophic lateral sclerosis. Ann Neurol 2011;70:913–19.
- 453. Mancuso R, Olivan S, Rando A, et al. Sigma-1R agonist improves motor function and motoneuron survival in ALS mice. Neurotherapeutics 2012;9:814–26.
- 454. Vollrath JT, Sechi A, Dreser A, et al. Loss of function of the ALS protein SigR1 leads to ER pathology associated with defective autophagy and lipid raft disturbances. Cell Death Dis 2014;5: e1290.
- Cottone P, Wang X, Park J, et al. Antagonism of sigma-1 receptors blocks compulsive-like eating. Neuropsychopharmacology 2012; 37:2593–604.
- 456. Ruscher K, Shamloo M, Rickhag M, et al. The sigma-1 receptor enhances brain plasticity and functional recovery after experimental stroke. Brain 2011;134:732–46.

- 457. Katz JL, Su TP, Hiranita T, et al. A role for sigma receptors in stimulant self administration and addiction. Pharmaceuticals (Basel, Switzerland) 2011;4:880–914.
- 458. Kim HW, Roh DH, Yoon SY, et al. Activation of the spinal sigma-1 receptor enhances NMDA-induced pain via PKC- and PKAdependent phosphorylation of the NR1 subunit in mice. Br J Pharmacol 2008;154:1125–34.
- 459. Trujillo KA, Akil H. Excitatory amino acids and drugs of abuse: a role for N-methyl-D-aspartate receptors in drug tolerance, sensitization and physical dependence. Drug Alcohol Depend 1995;38: 139–54.
- Artola A, Hensch T, Singer W. Calcium-induced long-term depression in the visual cortex of the rat in vitro. J Neurophysiol 1996;76:984–94.
- Artola A, Singer W. Long-term depression of excitatory synaptic transmission and its relationship to long-term potentiation. Trends Neurosci 1993;16:480–7.
- 462. Bear M, Malenka R. Synaptic plasticity: LTP and LTD. Curr Opin Neurobiol 1994;4:389–99.
- Christie B, Kerr D, Abraham W. Flip side of synaptic plasticity: long-term depression mechanisms in the hippocampus. Hippocampus 1994;4:127–35.
- Christie B, Magee J, Johnston D. Dendritic calcium channels and hippocampal long-term depression. Hippocampus 1996;6:17–23.
- 465. Cummings J, Mulkey R, Nicoll R, Malenka R. Ca<sup>2+</sup> signaling requirements for long-term depression in the hippocampus. Neuron 1996;16:825–33.
- 466. Derrick B, Martinez Jr J, Associative, bidirectional modifications at the hippocampal mossy fibre-CA3 synapse. Nature 1996;381: 429–34.
- Edwards F. LTP a structural model to explain the inconsistencies. Trends Neurosci 1995;18:250–5.
- 468. Hansel C, Artola A, Singer W. Different threshold levels of postsynaptic [Ca2+]i have to be reached to induce LTP and LTD in neocortical pyramidal cells. J Physiol 1996;90:317–19.
- 469. Jeffery KJ. LTP and spatial learning where to next? Hippocampus 1997;7:95–110
- Kirkwood A, Rioult MC, Bear MF. Experience-dependent modification of synaptic plasticity in visual cortex. Nature 1996;381: 526–8.
- 471. Malenka RC. Synaptic plasticity in the hippocampus: LTP and LTD. Cell 1994;78:535–8.
- 472. Maren S, Tocco G, Standley S, et al. Postsynaptic factors in the expression of long-term potentiation (LTP): increased glutamate receptor binding following LTP induction in vivo. Proc Natl Acad Sci USA 1993;90:9654–8.
- 473. Tsumoto T, Yasuda H, Fukuda M, Akaneya Y. Postsynaptic calcium and calcium-dependent processes in synaptic plasticity in the developing visual cortex. J Physiol (Paris) 1996;90:151–6.
- 474. Bode B. Recent molecular advances in mammalian glutamine transport. J Nutr 2001;131:2475S–85S; discussion 2486S–7S.
- Arriza J, Fairman W, Wadiche J, et al. Functional comparisons of three glutamate transporter subtypes cloned from human motor cortex. J Neurosci 1994;14:5559–69.
- 476. Fairman W, Vandenberg R, Arriza J, et al. An excitatory aminoacid transporter with properties of a ligand-gated chloride channel. Nature 1995;375:599–603.
- 477. Kanai Y, Hediger MA. Primary structure and functional characterization of a high-affinity glutamate transporter. Nature 1992; 360:467–71.
- 478. Malandro MS, Kilberg MS. Molecular biology of mammalian amino acid transporters. Annu Rev Biochem 1996;65:305–36.
- 479. Pines G, Danbolt NC, Bjoras M, et al. Cloning and expression of a rat brain L-glutamate transporter. Nature 1992;360:464–7.
- Schultz K, Stell WK. Immunocytochemical localization of the high-affinity glutamate transporter, EAAC1, in the retina of representative vertebrate species. Neurosci Lett 1996;211:191–4.
- 481. Bai L, Xu H, Collins J, Ghishan F. Molecular and functional analysis of a novel neuronal vesicular glutamate transporter. J Biol Chem 2001;276:36764–9.
- 482. Moriyama Y, Yamamoto A. Glutamatergic chemical transmission: look! Here, there, and anywhere. J Biochem 2004;135:155–63.
- Aanonsen L, Wilcox G. Nociceptive action of excitatory amino acids in the mouse: effects of spinally administered opioids,

phencyclidine and sigma agonists. J Pharm Exp Therap 1987;243: 9–19.

- 484. Kanai Y, Bhide PG, DiFiglia M, Hediger MA. Neuronal highaffinity glutamate transport in the rat central nervous system. Neuroreport 1995;6:2357–62.
- 485. Aas P, Tanso R, Fonnum F. Stimulation of peripheral cholinergic nerves by glutamate indicates a new peripheral glutamate receptor. Eur J Pharmacol 1989;164:93–102.
- 486. Barb C, Campbell R, Armstrong J, Cox N. Aspartate and glutamate modulation of growth hormone secretion in the pig: possible site of action. Domestic Anim Endocrinol 1996;13: 81–90.
- 487. Bertrand G, Gross R, Puech R, et al. Evidence for a glutamate receptor of the AMPA subtype which mediates insulin release from rat perfused pancreas. Br J Pharmacol 1992;106:354–9.
- 488. Bertrand G, Gross R, Puech R, et al. Glutamate stimulates glucagon secretion via an excitatory amino acid receptor of the AMPA subtype in rat pancreas. Eur J Pharmacol 1993;237: 45–50.
- 489. Burns G, Stephens K, Benson J. Expression of mRNA for the Nmethyl-D-aspartate (NMDAR1) receptor by the enteric neurons of the rat. Neurosci Lett 1994;170:87–90.
- Carlton S, Hargett G, Coggeshall R. Localization and activation of glutamate receptors in unmyelinated axons of rat glabrous skin. Neurosci Lett 1995;197:25–8.
- Chaudhari N, Yang H, Lamp C, et al. The taste of monosodium glutamate: membrane receptors in taste buds. J Neurosci 1996;16: 3817–26.
- 492. Chenu C, Serre C, Raynal C, et al. Glutamate receptors are expressed by bone cells and are involved in bone resorption. Bone 1998;22:295–9.
- 493. Coggeshall R, Carlton S. Ultrastructural analysis of NMDA, AMPA, and kainate receptors on unmyelinated and myelinated axons in the periphery. J Compar Neurobiol 1998;391:78–86.
- 494. Dememes D, Lleixa A, Dechesne C. Cellular and subcellular localization of AMPA-selective glutamate receptors in the mammalian peripheral vestibular system. Brain Res 1995;671:83–94.
- 495. Eliasof S, Werblin F. Characterization of the glutamate transporter in retinal cones of the tiger salamander. J Neurosci 1993;13: 402–11.
- 496. Genever P, Maxfield S, Kennovin G, et al. Evidence for a novel glutamate-mediated signaling pathway in keratinocytes. J Invest Dermatol 1999;112:337–42.
- 497. Genever P, Skerry T. Regulation of spontaneous glutamate release activity in osteoblastic cells and its role in differentiation and survival: evidence for intrinsic glutamatergic signaling in bone. FASEB J 2001;15:1586–8.
- 498. Genever P, Wilkinson D, Patton A, et al. Expression of a functional N-methyl-D-aspartate-type glutamate receptor by bone marrow megakaryocytes. Blood 1999;93:2876–83.
- 499. Gill S, Barker M, Pulido O. Neuroexcitatory targets in the female reproductive system of the nonhuman primate (Macaca fascicularis). Toxicol Pathol 2008;36:478–84.
- Gill S, Veinot J, Kavanagh M, Pulido O. Human heart glutamate receptors – implications for toxicology, food safety, and drug discovery. Toxicol Pathol 2007;35:411–17.
- Gill S, Pulido O, Mueller R, McGuire P. Immunochemical localization of the metabotropic glutamate receptors in the rat heart. Brain Res Bull 1999;48:143–6.
- Gill S, Pulido O, Mueller R, McGuire P. Molecular and immunochemical characterization of the ionotropic glutamate receptors in the rat heart. Brain Res Bull 1998;46:429–34.
- 503. Gonoi T, Mizuno N, Inagaki N, et al. Functional neuronal ionotropic glutamate receptors are expressed in the non-neuronal cell line MIN6. J Biol Chem 1994;269:16989–92.
- Hardy M, Younkin D, Tang C, et al. Expression of non-NMDA glutamate receptor channel genes by clonal human neurons. J Neurochem 1994;63:482–9.
- Haxhiu M, Erokwu B, Dreshaj I. The role of excitatory amino acids in airway reflex responses in anesthetized dogs. J automat Nerv Syst 1997;67:192–9.
- 506. Hayashi Y, Zviman M, Brand J, et al. Measurement of membrane potential and [Ca2+]i in cell ensembles: application to the study of glutamate taste in mice. Biophys J 1996;71:1057–70.

- 507. Kiyama H, Sato K, Tohyama M. Characteristic localization of non-NMDA type glutamate receptor subunits in the rat pituitary gland. Brain Res 1993;19:262–8.
- Lindstrom P, Ohlsson L. Effect of N-methyl-D, L-aspartate on isolated rat somatotrophs. Endocrinology 1992;131:1903–7.
- 509. Liu HP, Tay SS, Leong SK. Localization of glutamate receptor subunits of the alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionate (AMPA) type in the pancreas of newborn guinea pigs. Pancreas 1997;14:360–8.
- Marc RE, Lam DM. Uptake of aspartic and glutamic acid by photoreceptors in goldfish retina. Proc Natl Acad Sci USA 1981; 78:7185–9.
- 511. Mick G. Non-N-methyl-D-aspartate glutamate receptors in glial cells and neurons of the pineal gland in a higher primate. Neuroendocrinology 1995;61:256–64.
- 512. Molnar E, Varadi A, McIlhinney RA, Ashcroft SJ. Identification of functional ionotropic glutamate receptor proteins in pancreatic beta-cells and in islets of Langerhans. FEBS Lett 1995;371: 253–7.
- Moroni F, Luzzi S, Franchi-Micheli S, Zilletti L. The presence of N-methyl-D-aspartate-type receptors for glutamic acid in the guinea pig myenteric plexus. Neurosci Lett 1986;68:57–62.
- 514. O'Connell DP, Aherne AM, Lane E, et al. Detection of dopamine receptor D1A subtype-specific mRNA in rat kidney by in situ amplification. Am J Physiol 1998;274:F232–41.
- Pulido O, Veinot J, Mueller R, et al. Toxicol pathol of glutamate receptors (GluRs) – an opportunity for pharmaceutical development. Part I – human heart. Washington (DC): STP Annual General Meeting, 2005.
- 516. Said SI. Glutamate receptors and asthmatic airway disease. Trends Pharmacol Sci 1999;20:132–4.
- 517. Sasa M, Takeshita S, Amano T, Kurisu K. Primary neurotransmitters and regulatory substances onto vestibular nucleus neurons. Uchu Seibutsu Kagaku 2001;15:371–4.
- Shannon HE, Sawyer BD. Glutamate receptors of the N-methyl-Daspartate subtype in the myenteric plexus of the guinea pig ileum. J Pharm Exp Therap 1989;251:518–23.
- 519. Stuhmer T, Amar M, Harvey RJ, et al. Structure and pharmacological properties of a molluscan glutamate-gated cation channel and its likely role in feeding behavior. J Neurosci 1996;16: 2869–80.
- 520. Sureda F, Copani A, Bruno V, et al. Metabotropic glutamate receptor agonists stimulate polyphosphoinositide hydrolysis in primary cultures of rat hepatocytes. Eur J Pharmacol 1997;338: R1–2.
- 521. Tachibana M, Kaneko A. L-glutamate-induced depolarization in solitary photoreceptors: a process that may contribute to the interaction between photoreceptors in situ. Proc Natl Acad Sci USA 1988;85:5315–19.
- 522. Watanabe M, Mishina M, Inoue Y. Distinct gene expression of the N-methyl-D-aspartate receptor channel subunit in peripheral neurons of the mouse sensory ganglia and adrenal gland. Neurosci Lett 1994;165:183–6.
- 523. Weaver CD, Gundersen V, Verdoorn TA. A high affinity glutamate/aspartate transport system in pancreatic islets of Langerhans modulates glucose-stimulated insulin secretion. J Biol Chem 1998;273:1647–53.
- 524. Weaver CD, Yao TL, Powers AC, Verdoorn TA. Differential expression of glutamate receptor subtypes in rat pancreatic islets. J Biol Chem 1996;271:12977–84.
- 525. Yamaguchi I, Jose PA, Mouradian MM, et al. Expression of dopamine D1A receptor gene in proximal tubule of rat kidneys. Am J Physiol 1993;264:F280–5.
- Yang JH, Wu SM. Characterization of glutamate transporter function in the tiger salamander retina. Vis Res 1997;37:827–38.
- 527. Brew H, Attwell D. Electrogenic glutamate uptake is a major current carrier in the membrane of axolotl retinal glial cells. Nature 1987;327:707–9.
- Schwartz EA, Tachibana M. Electrophysiology of glutamate and sodium co-transport in a glial cell of the salamander retina. J Physiol 1990;426:43–80.
- 529. Barbour B, Brew H, Attwell D. Electrogenic uptake of glutamate and aspartate into glial cells isolated from the salamander (Ambystoma) retina. J Physiol 1991;436:169–93.

- 531. Bouvier M, Szatkowski M, Amato A, Attwell D. The glial cell glutamate uptake carrier countertransports pH-changing anions. Nature 1992;360:471–4.
- Eliasof S, Jahr C. Retinal glial cell glutamate transporter is coupled to an anionic conductance. Proc Natl Acad Sci USA 1996; 93:4153–8.
- Fykse E, Fonnum F. Amino acid neurotransmission: dynamics of vesicular uptake. Neurochem Res 1996;21:1053–60.
- 534. Naito S, Ueda T. Adenosine triphosphate-dependent uptake of glutamate into protein I-associated synaptic vesicles. J Biol Chem 1983;258:696–9.
- 535. Tabb JS, Ueda T. Phylogenetic studies on the synaptic vesicle glutamate transport system. J Neurosci 1991;11:1822–8.
- Grant G, Dowling J. On bipolar cell responses in the teleost retina are generated by two distinct mechanisms. J Neurophysiol 1996; 76:3842–9.
- 537. Grant G, Werblin F. A glutamate-elicited chloride current with transporter-like properties in rod photoreceptors of the tiger salamander. Vis Neurosci 1996;13:135–44.
- Harrington E, Moddel G, Najm I, Baraban S. Altered glutamate receptor – transporter expression and spontaneous seizures in rats exposed to methylazoxymethanol in utero. Epilepsia 2007;48: 158–68.
- Kew JN, Kemp JA. Ionotropic and metabotropic glutamate receptor structure and pharmacology. Psychopharmacology 2005;179:4–29.
- 540. Picaud S, Larsson HP, Wellis DP, et al. Cone photoreceptors respond to their own glutamate release in the tiger salamander. Proc Natl Acad Sci USA 1995;92:9417–21.
- 541. Picaud SA, Larsson HP, Grant GB, et al. Glutamate-gated chloride channel with glutamate-transporter-like properties in cone photoreceptors of the tiger salamander. J Neurophysiol 1995;74: 1760–71.
- 542. Takamori S, Rhee JS, Rosenmund C, Jahn R. Identification of a vesicular glutamate transporter that defines a glutamatergic phenotype in neurons. Nature 2000;407:189–94.
- Asztely F, Gustafsson B. Ionotropic glutamate receptors: their possible role in the expression of hippocampal synaptic plasticity. Mol Neurobiol 1996;12:1–11.
- Seeburg PH. The TINS/TiPS lecture: the molecular biology of mammalian glutamate receptor channels. Trends Neurosci 1993; 16:359–65.
- Cancela J, Churchill G, Galione A. Coordination of agonistinduced Ca2+-signalling patterns by NAADP in pancreatic acinar cells. Nature 1999;398:74–6.
- 546. Laube B, Hirai H, Sturgess M, et al. Molecular determinants of agonist discrimination by NMDA receptor subunits: analysis of the glutamate binding site on the NR2B subunit. Neuron 1997;18: 493–503.
- 547. Laube B, Kuhse J, Betz H. Evidence for a tetrameric structure of recombinant NMDA receptors. J Neurosci 1998;18:2954–61.
- 548. Mayer ML, Armstrong N. Structure and function of glutamate receptor ion channels. Annu Rev Physiol 2004;66:161–81.
- McFeeters RL, Oswald RE. Emerging structural explanations of ionotropic glutamate receptor function. FASEB J 2004;18:428–38.
- Rosenmund C, Stern-Bach Y, Stevens CF. The tetrameric structure of a glutamate receptor channel. Science (New York, NY) 1998; 280:1596–9.
- 551. Wisden W, Seeburg PH. Mammalian ionotropic glutamate receptors. Curr Opin Neurobiol 1993;3:291–8.
- 552. Yuzaki M. New insights into the structure and function of glutamate receptors: the orphan receptor delta2 reveals its family's secrets. Keio J Med 2003;52:92–9.
- 553. Mansour M, Nagarajan N, Nehring RB, et al. Heteromeric AMPA receptors assemble with a preferred subunit stoichiometry and spatial arrangement. Neuron 2001;32:841–53.
- 554. Yoneda Y, Kuramoto N, Kitayama T, Hinoi E. Consolidation of transient ionotropic glutamate signals through nuclear transcription factors in the brain. Progr Neurobiol 2001;63:697–719.
- 555. Sharp CD, Hines I, Houghton J, et al. Glutamate causes a loss in human cerebral endothelial barrier integrity through activation of

NMDA receptor. Am J Physiol Heart Circulatory Physiol 2003; 285:H2592–8.

- 556. Mayer ML, Westbrook GL. Permeation and block of N-methyl-Daspartic acid receptor channels by divalent cations in mouse cultured central neurones. J Physiol 1987;394:501–27.
- 557. Johnson JW, Ascher P. Glycine potentiates the NMDA response in cultured mouse brain neurons. Nature 1987;325:529–31.
- Kleckner NW, Dingledine R. Requirement for glycine in activation of NMDA-receptors expressed in *Xenopus* oocytes. Science (NY) 1988;241:835–7.
- 559. Verkhratsky A, Kirchhoff F. NMDA Receptors in glia. Neuroscientist 2007;13:28–37.
- Chavez A, Singer J, Diamond J. Fast neurotransmitter release triggered by Ca influx through AMPA-type glutamate receptors. Nature 2006;443:705–8.
- Monyer H, Seeburg PH, Wisden W. Glutamate-operated channels: developmentally early and mature forms arise by alternative splicing. Neuron 1991;6:799–810.
- Sommer B, Keinanen K, Verdoorn TA, et al. Flip and flop: a cellspecific functional switch in glutamate-operated channels of the CNS. Science (NY) 1990;249:1580–5.
- Chittajallu R, Braithwaite S, Clarke V, Henley J. Kainate receptors: subunits, synaptic localization and function. Trends Pharmacol Sci 1999;20:26–35.
- 564. Contractor A, Swanson G, Heinemann S. Kainate receptors are involved in short- and long-term plasticity at mossy fiber synapses in the hippocampus. Neuron 2001;29:209–16.
- Schmitz D, Mellor J, Nicoll RA. Presynaptic kainate receptor mediation of frequency facilitation at hippocampal mossy fiber synapses. Science (NY) 2001;291:1972–6.
- 566. Vignes M, Collingridge GL. The synaptic activation of kainate receptors. Nature 1997;388:179–82.
- Abram S, Olson E. Systemic opioids do not suppress spinal sensitization after subcutaneous formalin in rats. Anesthesiology 1994;80:1114–19.
- Lerma J, Morales M, Vicente MA, Herreras O. Glutamate receptors of the kainate type and synaptic transmission. Trends Neurosci 1997;20:9–12.
- Wilding TJ, Huettner JE. Activation and desensitization of hippocampal kainate receptors. J Neurosci 1997;17:2713–21.
- 570. Kunishima N, Shimada Y, Tsuji Y, et al. Structural basis of glutamate recognition by a dimeric metabotropic glutamate receptor. Nature 2000;407:971–7.
- 571. Marin YE, Chen S. Involvement of metabotropic glutamate receptor 1, a G protein coupled receptor, in melanoma development. J Mol Med (Berlin, Germany) 2004;82:735–49.
- 572. Senkowska A, Ossowska K. Role of metabotropic glutamate receptors in animal models of Parkinson's disease. Pol J Pharmacol 2003;55:935–50.
- 573. Hallett P, Standaert D. Rationale for and use of NMDA receptor antagonists in Parkinson's disease. Pharmacol Therap 2004;102: 155–74.
- 574. Konitsiotis S. Novel pharmacological strategies for motor complications in Parkinson's disease. Expert Opinion Invest Drugs 2005;14:377–92.
- Coutinho V, Knopfel T. Metabotropic glutamate receptors: electrical and chemical signaling properties. Neuroscientist 2002;8:551–61.
- Cartmell J, Schoepp D. Regulation of neurotransmitter release by metabotropic glutamate receptors. J Neurochem 2000;75: 889–907.
- 577. Boos R, Schneider H, Wassle H. Voltage- and transmitter-gated currents of all-amacrine cells in a slice preparation of the rat retina. J Neurosci 1993;13:2874–88.
- 578. Cohen E, Miller R. The role of NMDA and non-NMDA excitatory amino acid receptors in the functional organization of primate retinal ganglion cells. Vis Neurosci 1994;11:317–32.
- Gilbertson T, Scobey R, Wilson M. Permeation of calcium ions through non-NMDA glutamate channels in retinal bipolar cells. Science (NY) 1991;251:1613–15.
- Kessler M, Arai A, Quan A, Lynch G. Effect of cyclothiazide on binding properties of AMPA-type glutamate receptors: lack of competition between cyclothiazide and GYKI 52466. Mol Pharmacol 1996;49:123–31.

- 581. Kessler M, Arai A, Vanderklish P, Lynch G. Failure to detect changes in AMPA receptor binding after long-term potentiation. Brain Res 1991;560:337–41.
- Yamada KA, Tang CM. Benzothiadiazides inhibit rapid glutamate receptor desensitization and enhance glutamatergic synaptic currents. J Neurosci 1993;13:3904–15.
- Yu W, Miller RF. NBQX, an improved non-NMDA antagonist studied in retinal ganglion cells. Brain Res 1995;692:190–4.
- Nakanishi S. Molecular diversity of glutamate receptors and implications for brain function. Science (NY) 1992;258:597–603.
- Boulter J, Hollmann M, O'Shea-Greenfield A, et al. Molecular cloning and functional expression of glutamate receptor subunit genes. Science (NY) 1990;249:1033–7.
- 586. Nakanishi N, Shneider NA, Axel R. A family of glutamate receptor genes: evidence for the formation of heteromultimeric receptors with distinct channel properties. Neuron 1990;5:569–81.
- Nakanishi S. Metabotropic glutamate receptors: synaptic transmission, modulation, and plasticity. Neuron 1994;13:1031–7.
- Bettler B, Boulter J, Hermans-Borgmeyer I, et al. Cloning of a novel glutamate receptor subunit, GluR5: expression in the nervous system during development. Neuron 1990;5:583–95.
- 589. Bettler B, Egebjerg J, Sharma G, et al. Cloning of a putative glutamate receptor: a low affinity kainate-binding subunit. Neuron 1992;8:257–65.
- 590. Egebjerg J, Bettler B, Hermans-Borgmeyer I, Heinemann S. Cloning of a cDNA for a glutamate receptor subunit activated by kainate but not AMPA. Nature 1991;351:745–8.
- Keinanen K, Wisden W, Sommer B, et al. A family of AMPAselective glutamate receptors. Science (NY) 1990;249:556–60.
- 592. John CS, Vilner BJ, Bowen WD. Synthesis and characterization of [1251]-N-(N-benzylpiperidin-4-yl)-4-iodobenzamide: a new sigma receptor radiopharmaceutical: high-affinity binding to MCF-7 breast tumor cells. J Med Chem 1994;37:1737–9.
- 593. Zou LB, Yamada K, Nabeshima T. Sigma receptor ligands (+)-SKF10,047 and SA4503 improve dizocilpine-induced spatial memory deficits in rats. Eur J Pharmacol 1998;355:1–10.
- 594. Wallace DR, Mactutus CF, Booze RM. Sigma binding sites identified by [(3)H] DTG are elevated in aged Fischer-344 × Brown Norway (F1) rats. Synapse (NY) 2000;35:311–13.
- 595. Whittemore ER, Ilyin VI, Woodward RM. Antagonism of Nmethyl-D-aspartate receptors by sigma site ligands: potency, subtype-selectivity and mechanisms of inhibition. J Pharm Exp Therap 1997;282:326–8.
- 596. Fletcher E, Church J, Abdel-Hamid K, MacDonald J. Blockade by sigma site ligands of N-methyl-D-aspartate-evoked responses in rat and mouse cultured hippocampal pyramidal neurones. Br J Pharmacol 1995;116:2791–800.
- 597. Pontecorvo MJ, Karbon EW, Goode S, et al. Possible cerebroprotective and in vivo NMDA antagonist activities of sigma agents. Brain Res Bull 1991;26:461–5.
- Yamamoto H, Yamamoto T, Sagi N, et al. Sigma ligands indirectly modulate the NMDA receptor-ion channel complex on intact neuronal cells via sigma 1 site. J Neurosci 1995;15:731–6.
- Perregaard J, Moltzen EK, Meier E, Sanchez C. Sigma ligands with subnanomolar affinity and preference for the sigma 2 binding site. 1: 3-(omega-aminoalkyl)-1H-indoles. J Med Chem 1995;38: 1998–2008.
- 600. de Costa B, Bowen W, Hellewell S, et al. Synthesis and evaluation of optically pure [3H]-(+)-pentazocine, a highly potent and selective radioligand for sigma receptors. FEBS Lett 1989;251: 53–8.
- 601. Meyer DA, Carta M, Partridge LD, et al. Neurosteroids enhance spontaneous glutamate release in hippocampal neurons: possible role of metabotropic sigma1-like receptors. J Biol Chem 2002; 277:28725–32.
- 602. Earley B, Burke M, Leonard B, et al. Evidence for an anti-amnesic effect of JO 1784 in the rat: a potent and selective ligand for the sigma receptor. Brain Res 1991;546:282–6.
- 603. Klink R, Robichaud M, Debonnel G. Gender and gonadal status modulation of dorsal raphe nucleus serotonergic neurons. Part I: effects of gender and pregnancy. Neuropharmacology 2002;43: 1119–28.
- 604. Klink R, Robichaud M, Debonnel G. Gender and gonadal status modulation of dorsal raphe nucleus serotonergic neurons. Part II: regulatory mechanisms. Neuropharmacol 2002;43:1129–38.

- 605. Pepe GJ, Rothchild I. A comparative study of serum progesterone levels in pregnancy and in various types of pseudopregnancy in the rat. Endocrinology 1974;95:275–9.
- Awapara J, Landua A, Fuerst R, Seale B. Free gamma-aminobutyric acid in brain. J Biol Chem 1950;187:35–9.
- Erlander M, Tillakaratne N, Feldblum S, et al. Two genes encode distinct glutamate decarboxylases. Neuron 1991;7:91–100.
- Erlander M, Tobin A. The structural and functional heterogeneity of glutamic acid decarboxylase: a review. Neurochem Res 1991; 16:215–26.
- Krnjevic K. Chemical nature of synaptic transmission in vertebrates. Physiol Rev 1974;54:418–540.
- 610. Otsuka M, Iversen LL, Hall ZW, Kravitz EA. Release of gammaaminobutyric acid from inhibitory nerves of lobster. Proc Natl Acad Sci USA 1966;56:1110–15.
- 611. Roberts E, Frankel S. gamma-Aminobutyric acid in brain: its formation from glutamic acid. J Biol Chem 1950;187:55–63.
- 612. Walters RJ. Excitation and adrenaline: GABA the biopolar neurotransmitter. Cell Sci 2004;1:20–22.
- Roberts E. Pregneolone from Selye to Alzheimer and a model of the pregnenolone sulfate binding site on the GABAA receptor. Biochem Pharmacol 1995;49:1–16.
- 614. Roberts E. GABA: the road to neurotransmitter status. New York: Alan R Liss; 1986.
- 615. Hosie AM, Buckingham SD, Hamon A, Sattelle DB. Replacement of asparagine with arginine at the extracellular end of the second transmembrane (M2) region of insect GABA receptors increases sensitivity to penicillin G. Invertebrate Neurosci 2006;6:75–9.
- 616. Borden L, Smith K, Hartig P, et al. Molecular heterogeneity of the gamma-aminobutyric acid (GABA) transport system: cloning of two novel high affinity GABA transporters from rat brain. J Biol Chem 1992;267:21098–104.
- Guastella J, Nelson N, Nelson H, et al. Cloning and expression of a rat brain GABA transporter. Science (NY) 1990;249:1303–6.
- Barnard E, Darlison M, Fujita N, et al. Molecular biology of the GABAA receptor. Adv Exp Med Biol 1988;236:31–45.
- Olsen RW, Tobin AJ. Molecular biology of GABAA receptors. FASEB J 1990;4:1469–80.
- 620. Schofield PR, Darlison MG, Fujita N, et al. Sequence and functional expression of the GABA A receptor shows a ligand-gated receptor super-family. Nature 1987;328:221–7.
- Bormann J. Electrophysiology of GABAA and GABAB receptor subtypes. Trends Neurosci 1998;11:112–16.
- Chebib M, Johnston G. The 'ABC' of GABA receptors: a brief review. Clin Exp Pharmacol Physiol 1999;26:937–40.
- 623. Darnaudery M, Koehl M, Piazza P, et al. Pregnenolone sulfate increases hippocampal acetylcholine release and spatial recognition. Brain Res 2000;852:173–9.
- 624. Chen G, Trombley P, van den Pol A. GABA receptors precede glutamate receptors in hypothalamic development; differential regulation by astrocytes. J Neurophysiol 1995;74:1473–84.
- 625. Majewska MD, Harrison NL, Schwartz RD, et al. Steroid hormone metabolites are barbiturate-like modulators of the GABA receptor. Science (NY) 1986;232:1004–7.
- 626. Paul SM, Purdy RH. Neuroactive steroids. FASEB J 1992;6: 2311–22.
- 627. Invernizzi R, Pozzi L, Samanin R. Release of dopamine is reduced by diazepam more in the nucleus accumbens than in the caudate nucleus of conscious rats. Neuropharmacology 1991;30:575–8.
- 628. Burt D, Kamatchi G. GABAA receptor subtypes: from pharmacology to molecular biology. FASEB J 1991;5:2916–23.
- Luddens H, Wisden W. Function and pharmacology of multiple GABAA receptor subunits. Trends Pharmacol Sci 1991;12:49–51.
- 630. Pritchett DB, Luddens H, Seeburg PH. Type I and type II GABAA-benzodiazepine receptors produced in transfected cells. Science (NY) 1989;245:1389–92.
- 631. Pritchett DB, Sontheimer H, Gorman CM, et al. Transient expression shows ligand gating and allosteric potentiation of GABAA receptor subunits. Science (NY) 1988;242:1306–8.
- 632. Pritchett DB, Sontheimer H, Shivers BD, et al. Importance of a novel GABAA receptor subunit for benzodiazepine pharmacology. Nature 1989;338:582–5.
- 633. Supplisson S, Bergman C. Control of NMDA receptor activation by a glycine transporter co-expressed in Xenopus oocytes. J Neurosci 1997;17:4580–90.

- 635. Childers S, Pacheco M, Bennett B, et al. Cannabinoid receptors: G-protein-mediated signal transduction mechanisms. Biochem Soc symp 1993;59:27–50.
- 636. Glass M, Dragunow M, Faull R. Cannabinoid receptors in the human brain: a detailed anatomical and quantitative autoradiographic study in the fetal, neonatal and adult human brain. Neuroscience 1997;77:299–318.
- 637. Mackie K. Cannabinoid receptors: where they are and what they do. J Neuroendocrinol 2008;20:10–14.
- 638. Mackie K. Distribution of cannabinoid receptors in the central and peripheral nervous system. HEP 2005;168:299–325.
- 639. Patel KD, Davison JS, Pittman QJ, Sharkey KA. Cannabinoid CB(2) receptors in health and disease. Curr Med Chem 2010;17: 1393–410.
- 640. Nocerino E, Amato M, Izzo AA. Cannabis and cannabinoid receptors. Fitoterapia 2000;71:S6–12.
- 641. Duncan M, Davison J, Sharkey K. Review article: endocannabinoids and their receptors in the enteric nervous system. Aliment Pharmacol Therap 2005;22:667–83.
- 642. Navarrete M, Araque A. Endocannabinoids mediate neuronastrocyte communication. Neuron 2008;57:883–93.
- 643. Mascia MS, Obinu MC, Ledent C, et al. Lack of morphineinduced dopamine release in the nucleus accumbens of cannabinoid CB(1) receptor knockout mice. Eur J Pharmacol 1999;383: R1–2.
- Haj-Dahmane S, Shen R. Modulation of the serotonin system by endocannabinoid signaling. Neuropharmacology 2011;61:414–20.
- 645. Ferreira S, Teixeira F, Garcao P, et al. Presynaptic CB(1) cannabinoid receptors control frontocortical serotonin and glutamate release – species differences. Neurochem Int 2012;61: 219–26.
- 646. Baur R, Gertsch J, Sigel E. The cannabinoid CB1 receptor antagonists rimonabant (SR141716) and AM251 directly potentiate GABA(A) receptors. Br J Pharmacol 2012;165:2479–84.
- 647. Matyas F, Yanovsky Y, Mackie K, et al. Subcellular localization of type 1 cannabinoid receptors in the rat basal ganglia. Neuroscience 2006;137:337–61.
- 648. Takahashi KA, Castillo PE. The CB1 cannabinoid receptor mediates glutamatergic synaptic suppression in the hippocampus. Neuroscience 2006;139:795–802.
- 649. Fernandez-Ruiz J, Garcia C, Sagredo O, et al. The endocannabinoid system as a target for the treatment of neuronal damage. Expert Opin Therap Targets 2010;14:387–404.
- Croxford J. Therapeutic potential of cannabinoids in CNS disease. CNS drugs 2003;17:179–202.
- 651. Lopez-Moreno JA, Gonzalez-Cuevas G, Moreno G, Navarro M. The pharmacology of the endocannabinoid system: functional and structural interactions with other neurotransmitter systems and their repercussions in behavioral addiction. Addiction Biol 2008; 13:160–87.
- 652. Arnold J. Why sigma-1 receptor dysfunction might confer vulnerability to cannabis-induced psychosis. Int J Neuropsychopharmacol 2014;17:1911–13.
- 653. Maurice T, Su TP. The pharmacology of sigma-1 receptors. Pharmacol Therap 2009;124:195–206.
- Wang J, Mack AL, Coop A, Matsumoto RR. Novel sigma (sigma) receptor agonists produce antidepressant-like effects in mice. Eur Neuropsychopharmacol 2007;17:708–16.
- 655. Narita N, Hashimoto K, Tomitaka S, Minabe Y. Interactions of selective serotonin reuptake inhibitors with subtypes of sigma receptors in rat brain. Eur J Pharmacol 1996;307:117–19.
- 656. Graybiel A, Besson M, Weber E. Neuroleptic-sensitive binding sites in the nigrostriatal system: evidence for differential distribution of sigma sites in the substantia nigra, pars compacta of the cat. J Neurosci 1989;9:326–38.
- 657. Eyles D, Stedman T, Pond S. Nonlinear relationship between circulating concentrations of reduced haloperidol and haloperidol: evaluation of possible mechanisms. Psychopharmacology 1994; 116:161–6.
- 658. Villard V, Meunier J, Chevallier N, Maurice T. Pharmacological interaction with the sigma1 (sigma1)-receptor in the acute behavioral effects of antidepressants. J Pharmacol Sci 2011;115: 279–92.

- Itzhak Y, Kassim CO. Clorgyline displays high affinity for sigma binding sites in C57BL/6 mouse brain. Eur J Pharmacol 1990;176: 107–8.
- 660. Schmidt A, Lebel L, Koe BK, et al. Sertraline potently displaces (+)-[3H]3-PPP binding to sigma sites in rat brain. Eur J Pharmacol 1989;165:335-6.
- 661. Fennell A, Swant J, Goodwin S, Khoshbouei H. Involvement of striatal and hippocampal sigma receptors in methamphetamineinduced maladaptation. [Abstract] FASEB J 2011;25:619.12.
- 662. Johnson TD. Modulation of channel function by polyamines. Trends Pharmacol Sci 1996;17:22–7.
- Navarro G, Moreno E, Bonaventura J, et al. Cocaine inhibits dopamine D2 receptor signaling via sigma-1-D2 receptor heteromers. PLoS One 2013;8:e61245.
- 664. Meziane H, Mathis C, Paul SM, Ungerer A. The neurosteroid pregnenolone sulfate reduces learning deficits induced by scopolamine and has promnestic effects in mice performing an appetitive learning task. Psychopharmacology 1996;126:323–30.
- 665. Su TP, Hayashi T, Maurice T, et al. The sigma-1 receptor chaperone as an inter-organelle signaling modulator. Trends Pharmacol Sci 2010;31:557–66.
- 666. Mellado M, Rodriguez-Frade JM, Manes S, Martinez AC. Chemokine signaling and functional responses: the role of receptor dimerization and TK pathway activation. Annu Rev Immunol 2001;19:397–421.
- 667. Maurice T, Phan VL, Urani A, Guillemain I. Differential involvement of the sigma(1) (sigma(1)) receptor in the antiamnesic effect of neuroactive steroids, as demonstrated using an in vivo antisense strategy in the mouse. Br J Pharmacol 2001;134: 1731–41.
- Hayashi M, Morimoto R, Yamamoto A, Moriyama Y. Expression and localization of vesicular glutamate transporters in pancreatic islets, upper gastrointestinal tract, and testis. J Histochem Cytochem 2003;51:1375–90.
- 669. Hayashi M, Yamada H, Uehara S, et al. Secretory granulemediated co-secretion of L-glutamate and glucagon triggers glutamatergic signal transmission in islets of Langerhans. J Biol Chem 2003;278:1966–74.
- 670. Berthois Y, Bourrie B, Galiegue S, et al. SR31747A is a sigma receptor ligand exhibiting antitumoural activity both in vitro and in vivo. Br J Cancer 2003;88:438–46.
- 671. Moriguchi S, Shinoda Y, Yamamoto Y, et al. Stimulation of the sigma-1 receptor by DHEA enhances synaptic efficacy and neurogenesis in the hippocampal dentate gyrus of olfactory bulbectomized mice. PLoS One 2013;8:e60863.
- 672. Hayashi T, Su T. Sigma-1 receptors at galactosylceramideenriched lipid microdomains regulate oligodendrocyte differentiation. Proc Natl Acad Sci USA 2004;101:14949–54.
- 673. Stantchev TS, Markovic I, Telford WG, et al. The tyrosine kinase inhibitor genistein blocks HIV-1 infection in primary human macrophages. Virus Res 2007;123:178–89.
- 674. Vu TH, Weissman AD, London ED. Pharmacological characteristics and distributions of sigma- and phencyclidine receptors in the animal kingdom. J Neurochem 1990;54:598–604.
- 675. Majewska MD, Parameswaran S, Vu T, London ED. Divergent ontogeny of sigma and phencyclidine binding sites in the rat brain. Brain Res Dev Brain Res 1989;47:13–18.
- 676. Wolfe Jr SA, Culp SG, De Souza EB. Sigma-receptors in endocrine organs: identification, characterization, and autoradiographic localization in rat pituitary, adrenal, testis, and ovary. Endocrinology 1989;124:1160–72.
- 677. Wolfe Jr SA, Kulsakdinun C, Battaglia G, et al. Initial identification and characterization of sigma receptors on human peripheral blood leukocytes. J Pharm Exp Therap 1988;247:1114–19.
- Narayanan S, Mesangeau C, Poupaert JH, McCurdy CR. Sigma receptors and cocaine abuse. Curr Topics Med Chem 2011;11: 1128–50.
- 679. Brimson J, Brown C, Safrany S. Antagonists show GTP-sensitive high-affinity binding to the sigma-1 receptor. Br J Pharmacol 2011;164:772–80.
- 680. Holl R, Schepmann D, Frohlich R, et al. Dancing of the second aromatic residue around the 6,8-diazabicyclo[3.2.2]nonane framework: influence on sigma receptor affinity and cytotoxicity. J Med Chem 2009;52:2126–37.

- 380 C. G. Rousseaux & S. F. Greene
- 681. Chien C, Pasternak G. (-)-Pentazocine analgesia in mice: interactions with a sigma receptor system. Eur J Pharmacol 1995;294:303-8.
- Rodvelt KR, Miller DK. Could sigma receptor ligands be a treatment for methamphetamine addiction? Curr Dug Abuse Rev 2010;3:156–62
- 683. Brandes L. Hormetic effects of hormones, antihormones, and antidepressants on cancer cell growth in culture: in vivo correlates. Crit Rev Toxicol 2005;35:587–92.
- Brugmann WB, Firmani MA. Low concentrations of nitric oxide exert a hormetic effect on *Mycobacterium tuberculosis* in vitro. J clin Microbiol 2005;43:4844–6.
- 685. Calabrese E. Paradigm lost, paradigm found: the re-emergence of hormesis as a fundamental dose response model in the Toxicol sci. Environ Pollut (Barking, Essex: 1987) 2005;138:379–411.
- 686. Calabrese E, Blain R. The occurrence of hormetic dose responses in the toxicological literature, the hormesis database: an overview. Toxicol Appl Pharmacol 2005;202:289–301.
- 687. Calabrese E. Defining hormesis. Hum Exp Toxicol 2002;21:91-7.
- 688. Cedergreen N, Ritz C, Streibig J. Improved empirical models describing hormesis. Environ Toxicol Chem 2005;24:3166–172.
- 689. Celik I, Surucu O, Dietz C, et al. Therapeutic efficacy of endostatin exhibits a biphasic dose-response curve. Cancer Res 2005;65:11044–50.
- 690. Chiueh C, Andoh T, Chock P. Induction of thioredoxin and mitochondrial survival proteins mediates preconditioning-induced cardioprotection and neuroprotection. Ann NY Acad Sci 2005; 1042:403–18.
- 691. Dietert R. Commentary on hormetic dose-response relationships in immunology: occurrence, quantitative features of the dose response, mechanistic foundations, and clinical implications. Crit Rev Toxicol 2005;35:305–6.
- 692. Fukushima S, Kinoshita A, Puatanachokchai R, et al. Hormesis and dose-response-mediated mechanisms in carcinogenesis: evidence for a threshold in carcinogenicity of non-genotoxic carcinogenes. Carcinogenesis 2005;26:1835–45.
- 693. Kaiser J. Hormesis. Sipping from a poisoned chalice. Science (NY) 2003;302:376–9.
- 694. Lamming DW, Wood JG, Sinclair DA. Small molecules that regulate lifespan: evidence for xenohormesis. Mol Microbiol 2004;53:1003–9.
- 695. Laughlin Jr RB, Ng J, Guard HE. Hormesis: a response to low environmental concentrations of petroleum hydrocarbons. Science (NY) 1981;211:705–7.
- Lindsay DG. Nutrition, hormetic stress and health. Nutrition Res Rev 2005;18:249–58.
- 697. Liu SZ. Nonlinear dose-response relationship in the immune system following exposure to ionizing radiation: mechanisms and implications. Nonlinear Biol Toxicol Med 2003;1:71–92.
- 698. Puatanachokchai R, Morimura K, Wanibuchi H, et al. Alphabenzene hexachloride exerts hormesis in preneoplastic lesion formation of rat hepatocarcinogenesis with the possible role for hepatic detoxifying enzymes. Cancer Lett 2006;240:102–13.
- 699. Radak Z, Chung HY, Goto S. Exercise and hormesis: oxidative stress-related adaptation for successful aging. Biogerontology 2005;6:71–5.
- Randic M, Estrada E. Order from chaos: observing hormesis at the proteome level. J Proteome Res 2005;4:2133–6.
- 701. Rattan SI. Aging intervention, prevention, and therapy through hormesis. J Gerontol Ser A 2004;59:705–9.
- 702. Rattan SI. Hormetic mechanisms of anti-aging and rejuvenating effects of repeated mild heat stress on human fibroblasts in vitro. Rejuvenat Res 2004;7:40–8.
- 703. Rattan SI. Mechanisms of hormesis through mild heat stress on human cells. Ann NY Acad Sci 2004;1019:554–8.
- Renner R. Hormesis. Nietzsche's toxicology. Sci Am 2003;389: 28–30.
- Shama G, Alderson P. UV hormesis in fruits: a concept ripe for commercialization. Trends Food Sci Technol 2005;16:128–36.
- Sinclair DA. Toward a unified theory of caloric restriction and longevity regulation. Mech Age Dev 2005;126:987–1002.
- 707. Stebbing AR. Hormesis: interpreting the beta-curve using control theory. J Appl Toxicol 2000;20:93–101.
- 708. Liu SZ. Multilevel mechanisms of stimulatory effects of low dose radiation in immunity. In: Sugahara T, Sagan L and Aoyoma T,

eds. Low Dose Irradiation and Biological Defense Mechanisms. Amsterdam: Excerpta Medica, (1992), 225–32.

- 709. Tubiana M. Dose-effect relationship and estimation of the carcinogenic effects of low doses of ionizing radiation: the joint report of the Academie des Sciences (Paris) and of the Academie Nationale de Medecine. Int J Radiat Oncol Biol Phys 2005;63: 317–19.
- Calabrese E. Historical blunders: how toxicology got the doseresponse relationship half right. Cell Mol Biol (Noisy-le-Grand, France) 2005;51:643–54.
- 711. Hueppe F. Principles of bcteriology. Chicago, IL: The Open Court Publishing Company; 1896.
- 712. Schulz H. Uber Hefegifte. Pflugers Archiv Gesamte Physiologie Menschen Tiere 1888;42:517–41.
- 713. Schulz H. Zur Lehre von der Arzneiwirkung. [Virchows] Arch Pathol Anat Physiol Klin Med 1887;108:423–45.
- 714. Cook R, Calabrese E. The importance of hormesis to public health. Ciencia Saude Coletiva 2007;12:955–63.
- Calabrese E. 5-Hydroxytryptamine (serotonin): biphasic dose responses. Crit Rev Toxicol 2001;31:553–61.
- Calabrese E. Adenosine: biphasic dose responses. Crit Rev Toxicol 2001;31:539–51.
- Calabrese E. Adrenergic receptors: biphasic dose responses. Crit Rev Toxicol 2001;31:523–38.
- Calabrese E. Amyloid beta-peptide: biphasic dose responses. Crit Rev Toxicol 2001;31:605–6.
- Calabrese E. Androgens: biphasic dose responses. Crit Rev Toxicol 2001;31:517–22.
- Calabrese E. Apoptosis: biphasic dose responses. Crit Rev Toxicol 2001;31:607–13.
- Calabrese E. Dopamine: biphasic dose responses. Crit Rev Toxicol 2001;31:563–83.
- 722. Calabrese E. Estrogen and related compounds: biphasic dose responses. Crit Rev Toxicol 2001;31:503–15.
- 723. Calabrese E. Hormetic dose-response relationships in immunology: occurrence, quantitative features of the dose response, mechanistic foundations, and clinical implications. Crit Rev Toxicol 2005;35:89–95.
- Calabrese E. Nitric oxide: biphasic dose responses. Crit Rev Toxicol 2001;31:489–501.
- 725. Calabrese E. Opiates: biphasic dose responses. Crit Rev Toxicol 2001;31:585–604.
- Calabrese E. Overcompensation stimulation: a mechanism for hormetic effects. Crit Rev Toxicol 2001;31:425–70.
- Calabrese E. Prostaglandins: biphasic dose responses. Crit Rev Toxicol 2001;31:475–87.
- Calabrese E, Baldwin L. Agonist concentration gradients as a generalizable regulatory implementation strategy. Crit Rev Toxicol 2001;31:471–3.
- Calabrese E, Baldwin L. The frequency of U-shaped dose responses in the toxicological literature. Toxicol Sci 2001;62: 330–8.
- Calabrese E, Baldwin L. Hormesis: a generalizable and unifying hypothesis. Crit Rev Toxicol 2001;31:353–424.
- 731. Calabrese E, Baldwin L. Hormesis: U-shaped dose responses and their centrality in toxicology. Trends Pharmacol Sci 2001;22: 285–91.
- 732. Calabrese E, Staudenmayer J, Stanek III E, Hoffmann G. Hormesis outperforms threshold model in National Cancer Institute antitumor drug screening database. Toxicol Sci 2006; 94:368–78.
- Junien JL, Roman FJ, Brunelle G, Pascaud X. JO1784, a novel sigma ligand, potentiates [3H]acetylcholine release from rat hippocampal slices. Eur J Pharmacol 1991;200:343–5.
- 734. Waterhouse RN, Chapman J, Izard B, et al. Examination of four 123I-labeled piperidine-based sigma receptor ligands as potential melanoma imaging agents: initial studies in mouse tumor models. Nuclear Med Biol 1996;24:587–93.
- 735. van Waarde A, Buursma AR, Hospers GA, et al. Tumor imaging with 2 sigma-receptor ligands, 18F-FE-SA5845 and 11C-SA4503: a feasibility study. J Neuclear Med 2004;45:1939–45.
- 736. Kortekaas R, Maguire RP, van Waarde A, et al. Despite irreversible binding, PET tracer [11C]-SA5845 is suitable for imaging of drug competition at sigma receptors-the cases of ketamine and haloperidol. Neurochem Int 2008;53:45–50.

- 737. Stone JM, Arstad E, Erlandsson K, et al. [123I]TPCNE a novel SPET tracer for the sigma-1 receptor: first human studies and in vivo haloperidol challenge. Synapse (NY) 2006;60:109–17.
- Abel T, Lattal K. Molecular mechanisms of memory acquisition, consolidation and retrieval. Curr Opin Neurobiol 2001;1:180–7.
- 739. Kourrich S, Su TP, Fujimoto M, Bonci A. The sigma-1 receptor: roles in neuronal plasticity and disease. Trends Neurosci 2012;35: 762–71.
- 740. Renaudo A, Watry V, Chassot AA, et al. Inhibition of tumor cell proliferation by sigma ligands is associated with K+ Channel inhibition and p27kip1 accumulation. J Pharm Exp Therap 2004; 311:1105–14.
- 741. Engelhardt W, Friess K, Hartung E, et al. EEG and auditory evoked potential P300 compared with psychometric tests in assessing vigilance after benzodiazepine sedation and antagonism. Br J Anaesth 1992;69:75–80.
- 742. Kulikowski JJ, McGlone FF, Kranda K, Ott H. Are the amplitudes of visual evoked potentials sensitive indices of hangover effects after repeated doses of benzodiazepines? Psychopharmacol Supplement 1984;1:154–64
- 743. Martin F, Siddle DA, Gourley M, et al. P300 and traffic scenes: the effect of temazepam. Biol Psychol 1992;33:225–40.
- 744. Senda T, Mita S, Kaneda K, et al. Effect of SA4503, a novel sigmal receptor agonist, against glutamate neurotoxicity in cultured rat retinal neurons. Eur J Pharmacol 1998;342:105–11.
- 745. Nam Y, Shin EJ, Yang BK, et al. Dextromethorphan-induced psychotoxic behaviors cause sexual dysfunction in male mice via stimulation of sigma-1 receptors. Neurochem Int 2012;61:913–22.
- Kamei H, Noda Y, Nabeshima T. The psychological stress model using motor suppression. Nihon Yakurigaku Zasshi Folia Pharmacol Japonica 1999;113:113–20.
- 747. Roman F, Pascaud X, Vauche D, Junien JL. Evidence for a nonopioid sigma binding site in the guinea-pig myenteric plexus. Life Sci 1988;42:2217–22.
- 748. Whitlock BB, Liu Y, Chang S, et al. Initial characterization and autoradiographic localization of a novel sigma/opioid binding site in immune tissues. J Neuroimmunol 1996;67:83–96.
- 749. Wolfe Jr SA, Ha BK, Whitlock BB, Saini P. Differential localization of three distinct binding sites for sigma receptor ligands in rat spleen. J Neuroimmunol 1997;72:45–58.
- Chorawala M, Oza P, Shah G. Cellular and molecular pharmacology of sigma receptors. Intl J Pharm Sci Rev Res 2011;10: 45–53.
- 751. Novakova M, Bruderova V, Sulova Z, et al. Modulation of expression of the sigma receptors in the heart of rat and mouse in normal and pathological conditions. Gen Physiol Biophys 2007; 26:110–17.
- Stahl SM. Antidepressant treatment of psychotic major depression: potential role of the sigma receptor. CNS Spectr 2005;10: 319–23.
- Maurice T. Improving Alzheimer's disease-related cognitive deficits with sigmal receptor agonists. Drug News Perspect 2002;15:617–25.
- 754. Ujike H, Kuroda S, Otsuki S. sigma Receptor antagonists block the development of sensitization to cocaine. Eur J Pharmacol 1996;296:123–8.
- 755. Vilner BJ, John CS, Bowen WD. Sigma-1 and sigma-2 receptors are expressed in a wide variety of human and rodent tumor cell lines. Cancer Res 1995;55:408–13.
- Zhang H, Cuevas J. Sigma receptor activation blocks potassium channels and depresses neuroexcitability in rat intracardiac neurons. J Pharm Exp Therap 2005;313:1387–96.
- 757. Toyohara J, Sakata M, Ishiwata K. Imaging of sigma1 receptors in the human brain using PET and [11C]SA4503. Central Nerv Syst Agents Med Chem 2009;9:190–6.
- 758. Mori T, Hayashi T, Su TP. Compromising sigma-1 receptors at the endoplasmic reticulum render cytotoxicity to physiologically relevant concentrations of dopamine in a nuclear factor-kappaB/ Bcl-2-dependent mechanism: potential relevance to Parkinson's disease. J Pharm Exp Therap 2012;341:663–71.
- 759. Rybczynska AA, Elsinga PH, Sijbesma JW, et al. Steroid hormones affect binding of the sigma ligand 11C-SA4503 in tumour cells and tumour-bearing rats. Eur J Nuclear Med Mol Imaging 2009;36:1167–75.

- 760. Francardo V, Bez F, Wieloch T, et al. Pharmacological stimulation of sigma-1 receptors has neurorestorative effects in experimental parkinsonism. Brain 2014;137:1998–2014.
- 761. Gundlach A, Largent B, Snyder S. Autoradiographic localization of sigma receptor binding sites in guinea pig and rat central nervous system with (+)3H-3-(3-hydroxyphenyl)-N-(1-propyl)piperidine. J Neurosci 1986;6:1757–70.
- Mash DC, Zabetian CP. Sigma receptors are associated with cortical limbic areas in the primate brain. Synapse (NY) 1992;12: 195–205.
- 763. Largent BL, Gundlach AL, Snyder SH. Pharmacological and autoradiographic discrimination of sigma and phencyclidine receptor binding sites in brain with (+)-[3H]SKF 10,047, (+)-[3H]-3-[3-hydroxyphenyl]-N-(1-propyl)piperidine and [3H]-1-[1-(2-thienyl)cyclohexyl]piperidine. J Pharm Exp Therap 1986; 238:739–48.
- Nakazawa M, Kobayashi T, Matsuno K, Mita S. Possible involvement of a sigma receptor subtype in the neck dystonia in rats. Pharmacol Biochem Behav 1999;62:123–6.
- 765. Bouchard P, Quirion R. [3H]1,3-di(2-tolyl)guanidine and [3H](+)pentazocine binding sites in the rat brain: autoradiographic visualization of the putative sigma1 and sigma2 receptor subtypes. Neuroscience 1997;76:467–77.
- 766. Inoue A, Sugita S, Shoji H, et al. Repeated haloperidol treatment decreases sigma(1) receptor binding but does not affect its mRNA levels in the guinea pig or rat brain. Eur J Pharmacol 2000;401: 307–16.
- 767. Kamei H, Noda Y, Kameyama T, Nabeshima T. Role of (+)-SKF-10,047-sensitive sub-population of sigma 1 receptors in amelioration of conditioned fear stress in rats: association with mesolimbic dopaminergic systems. Eur J Pharmacol 1997;319: 165–72.
- Soby KK, Mikkelsen JD, Meier E, Thomsen C. Lu 28-179 labels a sigma(2)-site in rat and human brain. Neuropharmacology 2002; 43:95–100.
- 769. Blaylock R. Excitotoxins. Santa Fe: Health Press; 1997.
- 770. Greenamyre J, Porter R. Anatomy and physiology of glutamate in the CNS. Neurology 1994;44:S7–13.
- 771. Hayashi T, Su T. Sigma-1 receptor ligands: potential in the treatment of neuropsychiatric disorders. CNS drugs 2004;18: 269–84.
- 772. Palacios G, Muro A, Vela JM, et al. Immunohistochemical localization of the sigma1-receptor in oligodendrocytes in the rat central nervous system. Brain Res 2003;961:92–9.
- 773. Palacios G, Muro A, Verdu E, et al. Immunohistochemical localization of the sigma1 receptor in Schwann cells of rat sciatic nerve. Brain Res 2004;1007:65–70.
- 774. Mavlyutov TA, Epstein ML, Verbny YI, et al. Lack of sigma-1 receptor exacerbates ALS progression in mice. Neuroscience 2013;240:129–34.
- 775. Banister S, Kassiou M. The therapeutic potential of sigma (sigma) receptors for the treatment of central nervous system diseases: evaluation of the evidence. Current Pharm Design 2012;18: 884–901.
- 776. Antonini V, Marrazzo A, Kleiner G, et al. Anti-amnesic and neuroprotective actions of the sigma-1 receptor agonist (–)-MR22 in rats with selective cholinergic lesion and amyloid infusion. J Alzheime Dis 2011;24:569–86.
- 777. Antonini V, Prezzavento O, Coradazzi M, et al. Anti-amnesic properties of (+/-)-PPCC, a novel sigma receptor ligand, on cognitive dysfunction induced by selective cholinergic lesion in rats. J Neurochem 2009;109:744–54.
- 778. Meunier J, Demeilliers B, Celerier A, Maurice T. Compensatory effect by sigma1 (sigma1) receptor stimulation during alcohol withdrawal in mice performing an object recognition task. Behav Brain Res 2006;166:166–76.
- 779. Meunier J, Maurice T. Beneficial effects of the sigmal receptor agonists igmesine and dehydroepiandrosterone against learning impairments in rats prenatally exposed to cocaine. Neurotoxicol Teratol 2004;26:783–97.
- 780. Yang R, Chen L, Wang H, et al. Anti-amnesic effect of neurosteroid PREGS in Abeta25-35-injected mice through sigmal receptor- and alpha7nAChR-mediated neuroprotection. Neuropharmacology 2012;63:1042–50.

- 382 C. G. Rousseaux & S. F. Greene
- Matsuno K. Anti-amnesic effects of sigma (sigma)-receptor agonists. Nihon Yakurigaku Zasshi Folia Pharmacol Japonica 1999;114:25–33.
- 782. Li PK, Rhodes ME, Jagannathan S, Johnson DA. Reversal of scopolamine induced amnesia in rats by the steroid sulfatase inhibitor estrone-3-O-sulfamate. Brain Res Cognitive Brain Res 1995;2:251–4.
- Marrazzo A, Caraci F, Salinaro ET, et al. Neuroprotective effects of sigma-1 receptor agonists against beta-amyloid-induced toxicity. Neuroreport 2005;16:1223–6.
- 784. Maurice T, Martin-Fardon R, Romieu P, Matsumoto RR. Sigma(1) (sigma(1)) receptor antagonists represent a new strategy against cocaine addiction and toxicity. Neurosci Biobehav Rev 2002;26: 499–527.
- Romieu P, Lucas M, Maurice T. Sigma1 receptor ligands and related neuroactive steroids interfere with the cocaine-induced state of memory. Neuropsychopharmacology 2006;31:1431–43.
- 786. Tan F, Guio-Aguilar PL, Downes C, et al. The sigma 1 receptor agonist 4-PPBP elicits ERK1/2 phosphorylation in primary neurons: a possible mechanism of neuroprotective action. Neuropharmacology 2010;59:416–24.
- 787. Leonardo CC, Hall AA, Collier LA, et al. Administration of a Sigma Receptor Agonist Delays MCAO-Induced Neurodegeneration and White Matter Injury. Translational Stroke Research 2010;1:135–45.
- Schetz JA, Perez E, Liu R, et al. A prototypical Sigma-1 receptor antagonist protects against brain ischemia. Brain Res 2007;1181: 1–9.
- Bremner J. Does stress damage the brain? Biol Psychiatry 1999; 45:797–805
- Sheline YI, Gado MH, Kraemer HC. Untreated depression and hippocampal volume loss. Am J Psychiatry 2003;160:1516–18.
- 791. Sheline YI, Sanghavi M, Mintun MA, Gado MH. Depression duration but not age predicts hippocampal volume loss in medically healthy women with recurrent major depression. J Neurosci 1999;19:5034–43.
- Sheline YI, Wang PW, Gado MH, et al. Hippocampal atrophy in recurrent major depression. Proc Natl Acad Sci USA 1996;93: 3908–13.
- Sapolsky RM. The possibility of neurotoxicity in the hippocampus in major depression: a primer on neuron death. Biol Psychiatry 2000;48:755–65.
- Malberg JE, Eisch AJ, Nestler EJ, Duman RS. Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. J Neurosci 2000;20:9104–10.
- Altar C. Neurotrophins and depression. Trends Pharmacol Sci 1999;20:59–61.
- 796. Altar C, Whitehead R, Chen R, et al. Effects of electroconvulsive seizures and antidepressant drugs on brain-derived neurotrophic factor protein in rat brain. Biol Psychiatry 2003;54:703–9.
- 797. Nibuya M, Morinobu S, Duman RS. Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. J Neurosci 1995;15:7539–47.
- 798. Zetterstrom TS, Pei Q, Grahame-Smith DG. Repeated electroconvulsive shock extends the duration of enhanced gene expression for BDNF in rat brain compared with a single administration. Brain Res Molecular Brain Res 1998;57:106–10.
- Colino A, Halliwell J. Differential modulation of three separate Kconductances in hippocampal CA1 neurons by serotonin. Nature 1987;328:73–7.
- Shirayama Y, Chen AC, Nakagawa S, et al. Brain-derived neurotrophic factor produces antidepressant effects in behavioral models of depression. J Neurosci 2002;22:3251–61.
- Siuciak JA, Lewis DR, Wiegand SJ, Lindsay RM. Antidepressantlike effect of brain-derived neurotrophic factor (BDNF). Pharmacol Biochem Behav 1997;56:131–7.
- 802. Ovalle S, Andreu F, Perez MP, et al. Effect of the novel sigmal receptor ligand and putative atypical antipsychotic E-5842 on BDNF mRNA expression in the rat brain. Neuroreport 2002;13: 2345–8.
- 803. Abe K, Saito H. Epidermal growth factor selectively enhances NMDA receptor-mediated increase of intracellular Ca2+ concentration in rat hippocampal neurons. Brain Res 1992;587:102–8.
- 804. Xu QX, Li EM, Zhang YF, et al. Overexpression of sigmal receptor and its positive associations with pathologic TNM

classification in esophageal squamous cell carcinoma. J Histochem Cytochem 2012;60:457–66.

- Noda Y, Kamei H, Nabeshima T. Sigma-receptor ligands and antistress actions. Nihon Yakurigaku Zasshi Folia Pharmacol Japonica 1999;114:43–9.
- Skuza G. Potential antidepressant activity of sigma ligands. Pol J Pharmacol 2003;55:923–34.
- Connolly K, Thase M. Emerging drugs for major depressive disorder. Expert Opinionemerging drugs 2012;17:105–26.
- Hashimoto K, Ishiwata K. Sigma receptor ligands: possible application as therapeutic drugs and as radiopharmaceuticals. Current Pharm Design 2006;12:3857–76.
- Delaunois A, De Ron P, Detrait E, Guyaux M. Inhibitory effects of sigma-1 ligands on handling-induced tachycardia in conscious tethered rats. Fundam Clin Pharmacol 2013;27: 354–63.
- Ito K, Hirooka Y, Matsukawa R, et al. Decreased brain sigma-1 receptor contributes to the relationship between heart failure and depression. Cardiovasc Res 2012;93:33–40.
- Fond G. Using second-generation antidepressants (SGA) in daily practice. Ann Med Psychol 2013;171:198–203.
- Hindmarch I, Hashimoto K. Cognition and depression: the effects of fluvoxamine, a sigma-1 receptor agonist, reconsidered. Human Psychopharmacol 2010;25:193–200.
- 813. Matthews JD, Bottonari KA, Polania LM, et al. An open study of olanzapine and fluoxetine for psychotic major depressive disorder: interim analyses. J Clin Psychiatry 2002;63:1164–70.
- Beck S, Halloran P. Imipramine alters beta-adrenergic, but not serotonergic, mediated responses in rat hippocampal pyramidal cells. Brain Res 1989;504:72–81.
- Shirayama Y, Nishikawa T, Umino A, Takahashi K. p-chlorophenylalanine-reversible reduction of sigma binding sites by chronic imipramine treatment in rat brain. Eur J Pharmacol 1993;237: 117–26.
- Kinsora JJJ, Corbin AE, Snider B, et al. Effects of igmesine in preclinical antidepressant tests [abstract]. Soc Neurocsi 1998;24: 744.
- 817. Ukai M, Maeda H, Nanya Y, et al. Beneficial effects of acute and repeated administrations of sigma receptor agonists on behavioral despair in mice exposed to tail suspension. Pharmacol Biochem Behav 1998;61:247–52.
- Craven R, Grahame-Smith D, Newberry N. WAY-100635 and GR127935: effects on 5-hydroxytryptamine-containing neurones. Eur J Pharmacol 1994;271:R1–3.
- 819. Watanabe Y, Yamada S, Nakai M, et al. The rapid antidepressantlike effect of OPC-14523 in the forced swimming test is independent of action on 5-HT nerve terminals [abstract]. Soc Neurocsi 2000;26:387.16.
- 820. Yamada S, Uwahodo Y, Tottori K, et al. Role of sigma and 5-HT1A receptors in the forced swimming test: supporting the mechanism of action of OPC-14523 [abstract]. Soc Neurocsi 2000;26:2326.
- 821. Tsao LI, Su TP. Naloxone-sensitive, haloperidol-sensitive, [3H](+)SKF-10047-binding protein partially purified from rat liver and rat brain membranes: an opioid/sigma receptor? Synapse (NY) 1997;25:117–24
- Sanchez C, Arnt J, Costall B, et al. The selective sigma2-ligand Lu 28-179 has potent anxiolytic-like effects in rodents. J Pharm Exp Therap 1997;283:1323–32.
- Sanchez C, Papp M. The selective sigma2 ligand Lu 28-179 has an antidepressant-like profile in the rat chronic mild stress model of depression. Behav Pharmacol 2000;11:117–24.
- Skuza G, Rogoz Z. Antidepressant-like effect of PRE-084, a selective sigmal receptor agonist, in Albino Swiss and C57BL/6J mice. Pharmacol Rep 2009;61:1179–83.
- 825. Manji HK, Quiroz JA, Sporn J, et al. Enhancing neuronal plasticity and cellular resilience to develop novel, improved therapeutics for difficult-to-treat depression. Biol Psychiatry 2003; 53:707–2.
- 826. Saeki T, Nakamura M, Hirai N, et al. Localized potentiation of sleep slow-wave activity induced by prefrontal repetitive transcranial magnetic stimulation in patients with a major depressive episode. Brain Stimulation 2013;6:390–6.
- 827. Zhang CL, Feng ZJ, Liu Y, et al. Methylphenidate enhances NMDA-receptor response in medial prefrontal cortex via sigma-1

receptor: a novel mechanism for methylphenidate action. PLoS One 2012;7:e51910.

- 828. Barnes N, Sharp T. A review of central 5-HT receptors and their function. Neuropharmacology 1999;38:1083–152.
- Bhuiyan M, Tagashira H, Fukunaga K. Crucial interactions between selective serotonin uptake inhibitors and sigma-1 receptor in heart failure. J Pharmacol Sci 2013;121:177–84.
- Lopez-Munoz F, Alamo C. Neurobiological background for the development of new drugs in schizophrenia. Clinl Neuropharmacol 2011;34:111–26.
- 831. Uchida N, Ujike H, Nakata K, et al. No association between the sigma receptor type 1 gene and schizophrenia: results of analysis and meta-analysis of case-control studies. BMC Psychiatry 2003; 3:13.
- Johnson J, Horwath E, Weissman MM. The validity of major depression with psychotic features based on a community study. Arch Gen Psychiatry 1991;48:1075–81.
- Thakur M, Hays J, Krishnan KR. Clinical, demographic and social characteristics of psychotic depression. Psychiatry Res 1999;86: 99–106.
- 834. Flint A, Rifat S. The treatment of psychotic depression in later life: a comparison of pharmacotherapy and ECT. Int J Geriatr Psychiatry 1998;13:23–8.
- Jeste DV, Heaton SC, Paulsen JS, et al. Clinical and neuropsychological comparison of psychotic depression with nonpsychotic depression and schizophrenia. Am J Psychiatry 1996;153:490–6.
- Iwanami A, Oyamada S, Shirayama Y, Kamijima K. Algorithms for the pharmacotherapy psychotic depression. Psychiatry Clin Neurosci 1999;53:S45–8.
- Schatzberg AF. New approaches to managing psychotic depression. J Clin Psychiatry 2003;64:19–23.
- Schatzberg AF, Rothschild AJ, Bond TC, Cole JO. The DST in psychotic depression: diagnostic and pathophysiologic implications. Psychopharmacol Bull 1984;20:362–4.
- Vythilingam M, Chen J, Bremner JD, et al. Psychotic depression and mortality. Am J Psychiatry 2003;160:574–6.
- Hamner M, Gold P. Plasma dopamine beta-hydroxylase activity in psychotic and non-psychotic post-traumatic stress disorder. Psychiatry Res 1998;77:175–81.
- Nelson JC, Davis JM. DST studies in psychotic depression: a meta-analysis. Am J Psychiatry 1997;154:1497–503.
- 842. Miodownik C, Lerner V. Risperidone in the treatment of psychotic depression. Clin Neuropharmacol 2000;23:335–7.
- Mjellem N, Lund A, Hole K. Reduction of NMDA-induced behaviour after acute and chronic administration of desipramine in mice. Neuropharmacology 1993;32:591–5.
- Keck Jr PE, McElroy SL, Strakowski SM, Soutullo CA. Antipsychotics in the treatment of mood disorders and risk of tardive dyskinesia. J Clin Psychiatry 2000;61:33–8.
- Hirschfeld R. Efficacy of SSRIs and newer antidepressants in severe depression: comparison with TCAs. J Clin Psychiatry 1999; 60:326–35.
- 846. Masan PS. Atypical antipsychotics in the treatment of affective symptoms: a review. Ann Clin Psychiatry 2004;16:3–13.
- Bel N, Artigas F. Chronic treatment with fluvoxamine increases extracellular serotonin in frontal cortex but not in raphe nuclei. Synapse (NY) 1993;15:243–45.
- Gatti F, Bellini L, Gasperini M, Perez J, et al. Fluvoxamine alone in the treatment of delusional depression. Am J Psychiatry 1996; 153:414–16.
- 849. Zanardi R, Franchini L, Gasperini M, et al. Double-blind controlled trial of sertraline versus paroxetine in the treatment of delusional depression. Am J Psychiatry 1996;153:1631–3.
- 850. Zanardi R, Franchini L, Gasperini M, et al. Long-term treatment of psychotic (delusional) depression with fluvoxamine: an open pilot study. Int Clin Psychopharmacol 1997;12:195–7.
- Zanardi R, Franchini L, Serretti A, et al. Venlafaxine versus fluvoxamine in the treatment of delusional depression: a pilot double-blind controlled study. J Clin Psychiatry 2000;61:26–9.
- Meyers BS, Alexopoulos GS, Kakuma T, et al. Decreased dopamine beta-hydroxylase activity in unipolar geriatric delusional depression. Biol Psychiatry 1999;45:448–52.
- Duval F, Mokrani M, Crocq M, et al. Dopaminergic function and the cortisol response to dexamethasone in psychotic depression. Progr Neuro-psychopharmacol Biol Psychiatry 2000;24:207–25.

- Lykouras L, Markianos M, Hatzimanolis J, et al. Prolactin secretion in response to haloperidol challenge in delusional (psychotic) and non-delusional depression. Eur Psychiatry 2000; 15:480–2.
- Belanoff J, Flores B, Kalezhan M, et al. Rapid reversal of psychotic depression using mifepristone. J Clin Psychopharmacol 2001;21:516–21.
- 856. Chu J, Matthias D, Belanoff J, et al. Successful long-term treatment of refractory Cushing's disease with high-dose mifepristone (RU 486). J clin Endocrinol Metab 2001;86: 3568–73.
- 857. Simpson GM, El Sheshai A, Loza N, et al. An 8-week open-label trial of a 6-day course of mifepristone for the treatment of psychotic depression. J Clin Psychiatry 2005;66:598–602.
- Simpson GM, El Sheshai A, Rady A, et al. Sertraline as monotherapy in the treatment of psychotic and nonpsychotic depression. J Clin Psychiatry 2003;64:959–65.
- Kameyama T, Nagasaka M. Effects of apomorphine and diazepam on a quickly learned conditioned suppression in rats. Pharmacol Biochem Behav 1982;17:59–63.
- Borison R, Diamond B, Dren A. Does sigma receptor antagonism predict clinical antipsychotic efficacy? Psychopharmacol Bull 1991;27:103–6
- 861. Gewirtz G, Gorman J, Volavka J, et al. BMY 14802, a sigma receptor ligand for the treatment of schizophrenia. Neuropsychopharmacology 1994;10:37–40.
- 862. Frieboes R, Murck H, Wiedemann K, et al. Open clinical trial on the sigma ligand panamesine in patients with schizophrenia. Psychopharmacology 1997;132:82–8.
- 863. Modell S, Naber D, Holzbach R. Efficacy and safety of an opiate sigma-receptor antagonist (SL 82.0715) in schizophrenic patients with negative symptoms: an open dose-range study. Pharmacopsychiatry 1996;29:63–6.
- 864. Muller MJ, Grunder G, Wetzel H, et al. Antipsychotic effects and tolerability of the sigma ligand EMD 57445 (panamesine) and its metabolites in acute schizophrenia: an open clinical trial. Psychiatry Res 1999;89:275–80.
- 865. Hirano K, Kimura R, Sugimoto Y, et al. Relationship between brain serotonin transporter binding, plasma concentration and behavioural effect of selective serotonin reuptake inhibitors. Br J Pharmacol 2005;144:695–702.
- Griffin L, Mellon S. Selective serotonin reuptake inhibitors directly alter activity of neurosteroidogenic enzymes. Proc Natl Acad Sci USA 1999;96:13512–17.
- 867. Uzunova V, Sheline Y, Davis JM, et al. Increase in the cerebrospinal fluid content of neurosteroids in patients with unipolar major depression who are receiving fluoxetine or fluoxamine. Proc Natl Acad Sci USA 1998;95:3239–44.
- 868. Iyengar S, Mick S, Dilworth V, et al. Sigma receptors modulate the hypothalamic-pituitary-adrenal (HPA) axis centrally: evidence for a functional interaction with NMDA receptors, in vivo. Neuropharmacology 1990;29:299–303.
- 869. Iyengar S, Wood PL, Mick SJ, et al. (+) 3-[3-hydroxyphenyl-N-(1propyl) piperidine] selectively differentiates effects of sigma ligands on neurochemical pathways modulated by sigma receptors: evidence for subtypes, in vivo. Neuropharmacology 1991;30: 915–22.
- 870. Zamanillo D, Andreu F, Ovalle S, et al. Up-regulation of sigma(1) receptor mRNA in rat brain by a putative atypical antipsychotic and sigma receptor ligand. Neurosci Lett 2000;282:169–72.
- Cervo L, Samanin R. Effects of dopaminergic and glutamatergic receptor antagonists on the acquisition and expression of cocaine conditioning place preference. Brain Res 1995;673:242–50.
- 872. Matsumoto RR. Targeting sigma receptors: novel medication development for drug abuse and addiction. Expert review of clinical pharmacology 2009;2:351–8.
- Gao H, Wang C. Role of intrahippocampal sigma receptor in inhibiting seizure by electroacupuncture in rats. Med Acupunct 2010;22:19–24.
- Borowicz K, Kleinrok Z, Czuczwar S. Influence of 3-PPP, a sigma receptor ligand, on the anticonvulsive action of conventional antiepileptic drugs. Psychopharmacol Res 1999;40:509–16.
- Almansa C, Vela J. Selective sigma-1 receptor antagonists for the treatment of pain. Future Med Chem 2014;6:1179–99.

- 384 C. G. Rousseaux & S. F. Greene
- 876. Corbera J, Vano D, Martinez D, et al. A medicinal-chemistryguided approach to selective and druglike sigma 1 ligands. ChemMedChem 2006;1:140–54.
- 877. Gilmore D, Liu Y, Matsumoto R. Review of the pharmacological and clinical profile of rimcazole. CNS Drug Rev 2004;10:1–22.
- Jaggi AS, Singh N. Therapeutic targets for the management of peripheral nerve injury-induced neuropathic pain. CNS Neurol Disorders Drug Targets 2011;10:589–609.
- 879. Kwon YB, Jeong YC, Kwon JK, et al. The Antinociceptive Effect of Sigma-1 Receptor Antagonist, BD1047, in a Capsaicin Induced Headache Model in Rats. Korean J Physiol Pharmacol 2009;13: 425–9.
- Prezzavento O, Parenti C, Marrazzo A, et al. A new sigma ligand, (+/-)-PPCC, antagonizes kappa opioid receptor-mediated antinociceptive effect. Life Sci 2008;82:549–53.
- Sugai N, Yajima C, Chinzei M, et al. Postoperative pain relief by patient controlled analgesia using intravenous pentazocine. (Masui) Jpn J Anesthesiol 1995;44:216–20.
- 882. Wu HE, Hong JS, Tseng LF. Stereoselective action of (+)-morphine over (-)-morphine in attenuating the (-)-morphineproduced antinociception via the naloxone-sensitive sigma receptor in the mouse. Eur J Pharmacol 1007;571:145–51.
- King MA, Bradshaw S, Chang AH, et al. Potentiation of opioid analgesia in dopamine2 receptor knock-out mice: evidence for a tonically active anti-opioid system. J Neurosci 2001;21:7788–92.
- Cendan C, Pujalte J, Portillo-Salido E, et al. Formalin-induced pain is reduced in sigma(1) receptor knockout mice. Eur J Pharmacol 2005;511:73–4.
- 885. Kest B, Mogil JS, Sternberg WF, et al. 1,3-Di-o-tolylguanidine (DTG) differentially affects acute and tonic formalin pain: antagonism by rimcazole. Pharmacol Biochem Behav 1995;52: 175–8.
- Kest B, Mogil JS, Sternberg WF, et al. Antinociception following 1,3,-di-o-tolylguanidine, a selective sigma receptor ligand. Pharmacol Biochem Behav 1995;50:587–92.
- Pyun K, Son JS, Kwon YB. Chronic activation of sigma-1 receptor evokes nociceptive activation of trigeminal nucleus caudalis in rats. Pharmacol Biochem Behav 2014;124c:278–83.
- Anderson T, Andrew R. Spreading depression: imaging and blockade in the rat neocortical brain slice. J Neurophysiol 2002; 88:2713–25.
- Gonzalez-Cano R, Merlos M, Baeyens J, Cendan C. Sigmal receptors are involved in the visceral pain induced by intracolonic administration of capsaicin in mice. Anesthesiology 2013;118: 691–700.
- Bujalska M. Effect of nonselective and selective opioid receptors antagonists on antinociceptive action of acetaminophen [part III]. Pol J Pharmacol 2004;56:539–45.
- 891. Terashvili M, Wu HE, Moore RM, et al. (+)-Morphine and (-)-morphine stereoselectively attenuate the (-)-morphine-produced tail-flick inhibition via the naloxone-sensitive sigma receptor in the ventral periaqueductal gray of the rat. Eur J Pharmacol 2007;571:1–7.
- Tung AS, Yaksh TL. In vivo evidence for multiple opiate receptors mediating analgesia in the rat spinal cord. Brain Res 1982;247: 75–83.
- 893. Chow L, Huang E, Ho S, Lee T, Tao P. Dextromethorphan potentiates morphine antinociception at the spinal level in rats. Can J Anaest 2004;51:905–10.
- 894. Yoon SY, Roh DH, Seo HS, et al. An increase in spinal dehydroepiandrosterone sulfate (DHEAS) enhances NMDAinduced pain via phosphorylation of the NR1 subunit in mice: involvement of the sigma-1 receptor. Neuropharmacology 2010; 59:460–7.
- 895. Robson MJ, Noorbakhsh B, Seminerio MJ, Matsumoto RR. Sigma-1 receptors: potential targets for the treatment of substance abuse. Current Pharm Design 2012;18:902–19.
- Nuwayhid SJ, Werling LL. Sigma2 (sigma2) receptors as a target for cocaine action in the rat striatum. Eur J Pharmacol 2006;535: 98–103.
- 897. Matsumoto RR, Liu Y, Lerner M, et al. Sigma receptors: potential medications development target for anti-cocaine agents. Eur J Pharmacol 2003;469:1–12.
- 898. Nguyen EC, McCracken KA, Liu Y, et al. Involvement of sigma (sigma) receptors in the acute actions of methamphetamine:

receptor binding and behavioral studies. Neuropharmacology 2005;49:638-45.

- 899. Liu Y, Chen GD, Lerner MR, et al. Cocaine up-regulates Fra-2 and sigma-1 receptor gene and protein expression in brain regions involved in addiction and reward. J Pharm Exp Therap 2005;314: 770–9.
- Romieu P, Martin-Fardon R, Bowen WD, Maurice T. Sigma 1 receptor-related neuroactive steroids modulate cocaine-induced reward. J Neurosci 2003;23:3572–6.
- Romieu P, Martin-Fardon R, Maurice T. Involvement of the sigmal receptor in the cocaine-induced conditioned place preference. Neuroreport 2000;11:2885–8.
- 902. Romieu P, Phan VL, Martin-Fardon R, Maurice T. Involvement of the sigma(1) receptor in cocaine-induced conditioned place preference: possible dependence on dopamine uptake blockade. Neuropsychopharmacology 2002;26:444–55.
- 903. Stefanski R, Justinova Z, Hayashi T, et al. Sigmal receptor upregulation after chronic methamphetamine self-administration in rats: a study with yoked controls. Psychopharmacology 2004; 175:68–75.
- McCracken KA, Bowen WD, Matsumoto RR. Novel sigma receptor ligands attenuate the locomotor stimulatory effects of cocaine. Eur J Pharmacol 1999;365:35–8.
- Maurice T, Romieu P. Involvement of the sigmal receptor in the appetitive effects of cocaine. Pharmacopsychiatry 2001;37: S198–207.
- McDonald JW, Schoepp DD. The metabotropic excitatory amino acid receptor agonist 1S,3R-ACPD selectively potentiates Nmethyl-D-aspartate-induced brain injury. Eur J Pharmacol 1992; 215:353–4.
- 907. Sacaan AI, Schoepp DD. Activation of hippocampal metabotropic excitatory amino acid receptors leads to seizures and neuronal damage. Neurosci Lett 1992;139:77–82.
- Meldrum B. Protection against ischaemic neuronal damage by drugs acting on excitatory neurotransmission. Cerebrovasc Brain Metab Rev 1990;2:27–57.
- 909. Castelli M, Ingianni A, Stefanini E, Gessa G. Distribution of GABA(B) receptor mRNAs in the rat brain and peripheral organs. Life Sci 1999;64:1321–8.
- 910. Dierkes P, Hochstrate P, Schlue W. Distribution and functional properties of glutamate receptors in the leech central nervous system. J Neurophysiol 1996;75:2312–21.
- Spillson AB, Russell JW. Metabotropic glutamate receptor regulation of neuronal cell death. Exp Neurol 2003;184:S97–105.
- 912. Hirata Y, Yamamoto H, Atta M, et al. Chloroquine inhibits glutamate-induced death of a neuronal cell line by reducing reactive oxygen species through sigma-1 receptor. J Neurochem 2011;119:839–47.
- 913. Pal A, Fontanilla D, Gopalakrishnan A, et al. The sigma-1 receptor protects against cellular oxidative stress and activates antioxidant response elements. Eur J Pharmacol 2012;682: 12–20.
- 914. Teichberg VI. Glial glutamate receptors: likely actors in brain signaling. FASEB J 1991;5:3086–91.
- 915. Lai AY, Dhami KS, Todd KG. Moving past the "neuroncentric" perspective: a role for glia in neuropsychiatric disorders. J Psychiatry Neurosci 2009;34:173–4.
- 916. Rosenberg PA, Aizenman E. Hundred-fold increase in neuronal vulnerability to glutamate toxicity in astrocyte-poor cultures of rat cerebral cortex. Neurosci Lett 1989;103:162–8.
- 917. Lopez-Bayghen E, Cruz-Solis I, Corona M, et al. Glutamateinduced octamer DNA binding and transcriptional control in cultured radial glia cells. J Neurochem 2006;98:851–9.
- 918. Kaminska B, Kaczmarek L, Zangenehpour S, Chaudhuri A. Rapid phosphorylation of Elk-1 transcription factor and activation of MAP kinase signal transduction pathways in response to visual stimulation. Mol Cell Neurosci 1999;13:405–14.
- Crottes D, Martial S, Rapetti-Mauss R, et al. Sig1R protein regulates hERG channel expression through a post-translational mechanism in leukemic cells. J Biol Chem 2011;286:27947–58.
- 920. Dobrzynski H, Nikolski V, Sambelashvili A, Greener I, Yamamoto M, Boyett M, Efimov I. Site of origin and molecular substrate of atrioventricular junctional rhythm in the rabbit heart. Circulation Res 2003;93:1102–10.

- 921. Gorza L, Vitadello M. Distribution of conduction system fibers in the developing and adult rabbit heart revealed by an antineurofilament antibody. Circulation Res 1989;65:360–9.
- 922. Widran J, Lev M. The dissection of the atrioventricular node, bundle and bundle branches in the human heart. Circulation 1951; 4:863–7.
- 923. Monassier L, Bousquet P. Sigma receptors: from discovery to highlights of their implications in the cardiovascular system. Fundam Clin Pharmacol 2002;16:1–8.
- 924. Monassier L, Manoury B, Bellocq C, et al. sigma(2)-receptor ligand-mediated inhibition of inwardly rectifying K(+) channels in the heart. J Pharm Exp Therap 2007;322:341–50.
- 925. Bhuiyan M, Tagashira H, Shioda N, Fukunaga K. Targeting sigma-1 receptor with fluvoxamine ameliorates pressure-overload-induced hypertrophy and dysfunctions. Expert Opin Therap Targets 2010;14:1009–22.
- 926. Mueller RW, Gill SS, Pulido OM. The monkey (Macaca fascicularis) heart neural structures and conducting system: an immunochemical study of selected neural biomarkers and glutamate receptors. Toxicol Pathol 2003;31:227–34.
- 927. Ito K, Hirooka Y, Sunagawa K. Brain sigma-1 receptor stimulation improves mental disorder and cardiac function in mice with myocardial infarction. J Cardiovasc Pharmacol 2013;62:222–8.
- Gao X, Yao J, He Y, et al. Sigma-1 receptor agonists directly inhibit Nav1.2/1.4 channels. PLoS One 2012;7:e49384.
- 929. Johannessen M, Fontanilla D, Mavlyutov T, et al. Antagonist action of progesterone at sigma-receptors in the modulation of voltage-gated sodium channels. Am J Physiol Cell Physiol 2011; 300:C328–37.
- 930. Ela C, Barg J, Vogel Z, et al. Sigma receptor ligands modulate contractility, Ca++ influx and beating rate in cultured cardiac myocytes. J Pharm Exp Therap 1994;269:1300–9.
- 931. Ela C, Hasin Y, Eilam Y. Apparent desensitization of a sigma receptor sub-population in neonatal rat cardiac myocytes by pretreatment with sigma receptor ligands. Eur J Pharmacol 1996;295: 275–80.
- 932. Maslov LN, Lishmanov Iu B, Bogomaz SA, et al. [Dependence of the pump function of the isolated rat heart on the functional activity of sigma receptors during reperfusion]. Rossiiskii Fiziologicheskii Zhurnal Imeni IM Sechenova 1999;85:1396–408.
- Bhuiyan M, Fukunaga K. Targeting sigma-1 receptor signaling by endogenous ligands for cardioprotection. Expert Opin Therap Targets 2011;15:145–55.
- 934. Fialova K. Acute and chronic effects of sigma receptor ligands in mammalian myocardium. [PhD Thesis]. Czech Republic: Department of Physiology, Masaryk University; 2010:1–154.
- 935. Lishmanov Yu B, Maslov LN, Naryzhnaya NV, Tam SW. Ligands for opioid and sigma-receptors improve cardiac electrical stability in rat models of post-infarction cardiosclerosis and stress. Life Sci 1999;65:Pl13–17.
- 936. Li DP, Averill DB, Pan HL. Differential roles for glutamate receptor subtypes within commissural NTS in cardiac-sympathetic reflex. Am J Physiol Regulatory Integrative Comparative Physiol 2001;281:R935–43.
- 937. Rennie MJ, Ahmed A, Khogali SE, et al. Glutamine metabolism and transport in skeletal muscle and heart and their clinical relevance. J Nutr 1996;126:1142s–9s.
- 938. Jeanjean AP, Laterre EC, Maloteaux JM. Neuroleptic binding to sigma receptors: possible involvement in neuroleptic-induced acute dystonia. Biol Psychiatry 1997;41:1010–19.
- 939. Hinoi E, Fujimori S, Nakamura Y, Yoneda Y. Group III metabotropic glutamate receptors in rat cultured calvarial osteoblasts. Biochem Biophys Res Commun 2001;281:341–6.
- 940. Laketic-Ljubojevic I, Suva LJ, Maathuis FJ, et al. Functional characterization of N-methyl-D-aspartic acid-gated channels in bone cells. Bone 1999;25:631–7.
- 941. Patton AJ, Genever PG, Birch MA, et al. Expression of an Nmethyl-D-aspartate-type receptor by human and rat osteoblasts and osteoclasts suggests a novel glutamate signaling pathway in bone. Bone 1998;22:645–9.
- 942. Gray C, Marie H, Arora M, et al. Glutamate does not play a major role in controlling bone growth. J Bone Mineral Res 2001;16: 742–9.
- 943. Gu Y, Publicover S. Expression of functional metabotropic glutamate receptors in primary cultured rat osteoblasts.

Cross-talk with N-methyl-D-aspartate receptors. J Biol Chem 2000;275:34252–9.

- 944. Hinoi E, Fujimori S, Takemori A, et al. Demonstration of expression of mRNA for particular AMPA and kainate receptor subunits in immature and mature cultured rat calvarial osteoblasts. Brain Res 2002;943:112–16.
- Espinosa L, Itzstein C, Cheynel H, et al. Active NMDA glutamate receptors are expressed by mammalian osteoclasts. J Physiol 1999; 518:47–53.
- 946. Gu Y, Genever P, Skerry T, Publicover S. The NMDA type glutamate receptors expressed by primary rat osteoblasts have the same electrophysiological characteristics as neuronal receptors. Calcified Tissue Int 2002;70:194–203.
- Hinoi E, Fujimori S, Yoneda Y. Modulation of cellular differentiation by N-methyl-D-aspartate receptors in osteoblasts. FASEB J 2003;17:1532–34.
- 948. Peet NM, Grabowski PS, Laketic-Ljubojevic I, Skerry TM. The glutamate receptor antagonist MK801 modulates bone resorption in vitro by a mechanism predominantly involving osteoclast differentiation. FASEB J 1999;13:2179–85.
- 949. Bhangu P, Genever P, Spencer G, et al. Evidence for targeted vesicular glutamate exocytosis in osteoblasts. Bone 2001;29: 16–23.
- 950. Mason DJ, Suva LJ, Genever PG, et al. Mechanically regulated expression of a neural glutamate transporter in bone: a role for excitatory amino acids as osteotropic agents? Bone 1997;20: 199–205
- 951. Kawamura K, Kubota K, Kobayashi T, et al. Evaluation of [11C]SA5845 and [11C]SA4503 for imaging of sigma receptors in tumors by animal PET. Ann Nuclear Med 2005;19:701–9.
- 952. Bem W, Thomas G, Mamone J, et al. Overexpression of sigma receptors in nonneural human tumors. Cancer Res 1991;51: 6558–62.
- 953. Vilner BJ, de Costa BR, Bowen WD. Cytotoxic effects of sigma ligands: sigma receptor-mediated alterations in cellular morphology and viability. J Neurosci 1995;15:117–34.
- 954. John CS, Bowen WD, Varma VM, et al. Sigma receptors are expressed in human non-small cell lung carcinoma. Life Sci 1995; 56:2385–92.
- Moody TW, Leyton J, John C. Sigma ligands inhibit the growth of small cell lung cancer cells. Life Sci 2000;66:1979–86.
- 956. Mir SU, Ahmed IS, Arnold S, Craven RJ. Elevated progesterone receptor membrane component 1/sigma-2 receptor levels in lung tumors and plasma from lung cancer patients. Int J Cancer 2012; 131:E1–9.
- 957. Allen D, Delohery J, Baker G. Monosodium L-glutamate-induced asthma. J Allergy Clin Immunol 1987;80:530–7.
- 958. Said SI, Berisha HI, Pakbaz H. Excitotoxicity in the lung: Nmethyl-D-aspartate-induced, nitric oxide-dependent, pulmonary edema is attenuated by vasoactive intestinal peptide and by inhibitors of poly(ADP-ribose) polymerase. Proc Natl Acad Sci USA 1996;93:4688–92.
- 959. Purcell WM, Doyle KM, Westgate C, Atterwill CK. Characterisation of a functional polyamine site on rat mast cells: association with a NMDA receptor macrocomplex. J Neuroimmunol 1996;65:49–53.
- 960. Olney JW. Excitotoxins in foods. Neurotoxicology 1994;15: 535–44.
- 961. Brown C, Fezoui M, Selig W, et al. Antitussive activity of sigma-1 receptor agonists in the guinea-pig. Br J Pharmacol 2004;141: 233–40.
- 962. Eaton M, Lookingland K, Moore K. The sigma receptor ligand rimcazole alters secretion of prolactin and alpha-melanocyte stimulating hormone by dopaminergic and non-dopaminergic mechanisms. Eur J Pharmacol 1996;299:171–7.
- 963. Su TP, Wu XZ, Cone EJ, et al. Sigma compounds derived from phencyclidine: identification of PRE-084: a new, selective sigma ligand. J Pharm Exp Therap 1991;259:543–50.
- 964. Eaton M, Lookingland K, Moore K. The sigma ligand rimcazole activates noradrenergic neurons projecting to the paraventricular nucleus and increases corticosterone secretion in rats. Brain Res 1996;733:162–6.
- 965. Le Foll F, Castel H, Soriani O, et al. Gramicidin-perforated patch revealed depolarizing effect of GABA in cultured frog melanotrophs. J Physiol 1998;507:55–69.

- 386 C. G. Rousseaux & S. F. Greene
- 966. Le Foll F, Louiset E, Castel H, et al. Electrophysiological effects of various neuroactive steroids on the GABA(A) receptor in pituitary melanotrope cells. Eur J Pharmacol 1997;331: 303–11.
- 967. Pechnick RN. Effects of opioids on the hypothalamo-pituitaryadrenal axis. Annu Rev Pharmacol Toxicol 1993;33:353–82.
- Peck V, Shenkman L. Haloperidol-induced syndrome of inappropriate secretion of antidiuretic hormone. Clin Pharmacol Therap 1979;26:442–4.
- Inagaki N, Kawasaki H, Nagai H. Characterization of purificationassociated reduction in IgE-dependent histamine release from rat peritoneal mast cells. Inflamm Res 1995;44:541–7.
- 970. Inagaki N, Kuromi H, Gonoi T, et al. Expression and role of ionotropic glutamate receptors in pancreatic islet cells. FASEB J 1995;5:686–91.
- 971. Liu MT, Rothstein JD, Gershon MD, Kirchgessner AL. Glutamatergic enteric neurons. J Neurosci 1997;17:4764–84.
- 972. Tong Q, Ouedraogo R, Kirchgessner AL. Localization and function of group III metabotropic glutamate receptors in rat pancreatic islets. Am J Physio Endocrinol Metab 2002;282: E1324–33.
- 973. Trudeau F, Gagnon S, Massicotte G. Hippocampal synaptic plasticity and glutamate receptor regulation: influences of diabetes mellitus. Eur J Pharmacol 2004;490:177–86.
- 974. Caruso C, Bottino M, Pampillo M, et al. Glutamate induces apoptosis in anterior pituitary cells through group II metabotropic glutamate receptor activation. Endocrinology 2004;145:4677–84.
- 975. Yoneda Y, Ogita K. Localization of [3H]glutamate binding sites in rat adrenal medulla. Brain Res 1986;383:387–91.
- Ronsisvalle G, Marrazzo A, Pasquinucci L, et al. Specific kappa opioid receptor agonists. Farmaco (Societa chimica italiana: 1989) 2001;56:121–5.
- 977. Schaefer M, Habenicht UF, Brautigam M, Gudermann T. Steroidal sigma receptor ligands affect signaling pathways in human spermatozoa. Biol Reprod 2000;63:57–63.
- 978. Lara H, Bastos-Ramos W. Glutamate and kainate effects on the noradrenergic neurons innervating the rat vas deferens. J Neurosci Res 1988;19:239–44.
- 979. Battaglia F. Glutamine and glutamate exchange between the fetal liver and the placenta. J Nutr 2000;130:974s–7s.
- Caveliers V, Everaert H, John C, Lahoutte T, Bossuyt A. Sigma receptor scintigraphy with N-[2-(1'-piperidinyl)ethyl]-3-(123)Iiodo-4-methoxybenzamide of patients with suspected primary breast cancer: first clinical results. J Neuclear Med 2002;43: 1647–9.
- 981. Bowen W. Sigma receptors: recent advances and new clinical potentials. Pharm Acta Helvet 2000;74:211–18.
- 982. Gebreselassie D, Bowen W. Sigma-2 receptors are specifically localized to lipid rafts in rat liver membranes. Eur J Pharmacol 2004;493:19–28.
- Klouz A, Sapena R, Liu J, et al. Evidence for sigma-1-like receptors in isolated rat liver mitochondrial membranes. Br J Pharmacol 2002;135:1607–15.
- 984. Abe T, Sugihara H, Nawa H, et al. Molecular characterization of a novel metabotropic glutamate receptor mGluR5 coupled to inositol phosphate/Ca2+ signal transduction. J Biol Chem 1992; 267:13361–8.
- Lipton SA, Tauck DL. Voltage-dependent conductances of solitary ganglion cells dissociated from the rat retina. J Physiol 1987;385: 361–91.
- 986. Liu LL, Wang L, Zhong YM, Yang XL. Expression of sigma receptor 1 mRNA and protein in rat retina. Neuroscience 2010; 167:1151–9.
- 987. Ola MS, Moore P, El-Sherbeny A, et al. Expression pattern of sigma receptor 1 mRNA and protein in mammalian retina. Brain Res Mol Brain Res 2001;95:86–95.
- Martin PM, Roon P, Van Ells TK, et al. Death of retinal neurons in streptozotocin-induced diabetic mice. Invest Opthamol Vis Sci 2004;45:3330–6.
- 989. Jiang G, Mysona B, Dun Y, et al. Expression, subcellular localization, and regulation of sigma receptor in retinal muller cells. Invest Opthamol Vis Sci 2006;47:5576–82.
- 990. Tuerxun T, Numakawa T, Adachi N, et al. SA4503, a sigma-1 receptor agonist, prevents cultured cortical neurons from oxidative stress-induced cell death via suppression of MAPK pathway

activation and glutamate receptor expression. Neurosci Lett 2010; 469:303–8.

- 991. Jonhede S, Petersen A, Zetterberg M, Karlsson JO. Acute effects of the sigma-2 receptor agonist siramesine on lysosomal and extralysosomal proteolytic systems in lens epithelial cells. Mol Vis 2010;16:819–27.
- 992. Ha Y, Dun Y, Thangaraju M, Duplantier J, et al. Sigma receptor 1 modulates endoplasmic reticulum stress in retinal neurons. Invest Ophthalmol Vis Sci 2011;52:527–40.
- 993. Ha Y, Saul A, Tawfik A, et al. Late-onset inner retinal dysfunction in mice lacking sigma receptor 1 (sigmaR1). Invest Ophthalmol Vis Sci 2011;52:7749–60.
- 994. Wang L, Duncan G. Silencing of sigma-1 receptor induces cell death in human lens cells. Exp Cell Res 2006;312:1439–46.
- 995. Wang L, Prescott AR, Spruce BA, et al. Sigma receptor antagonists inhibit human lens cell growth and induce pigmentation. Invest Opthamol Vis Sci 2005;46:1403–8.
- 996. Li T, Ghishan FK, Bai L. Molecular physiology of vesicular glutamate transporters in the digestive system. World J Gastroenterol 2005;11:1731–6.
- 997. Tsai LH. Function of GABAergic and glutamatergic neurons in the stomach. J Biomed Sci 2005;12:255–66.
- 998. Tsai LH, Lee YJ, Wu J. Effect of excitatory amino acid neurotransmitters on acid secretion in the rat stomach. J Biomed Sci 1999;6:36–44.
- 999. Tsai LH, Tsai W, Wu JY. Effect of L-glutamic acid on acid secretion and immunohistochemical localization of glutamatergic neurons in the rat stomach. J Neurosci Res 1994;38:188–95.
- 1000. Farthing M. Novel agents for the control of secretory diarrhoea. Expert Opin Invest Drugs 2004;13:777–85.
- Collina S, Gaggeri R, Marra A, et al. Sigma receptor modulators: a patent review. Expert Opin Therap patents 2013;23:597–613.
- 1002. Niijima A. Reflex effects of oral, gastrointestinal and hepatoportal glutamate sensors on vagal nerve activity. J Nutr 2000; 130:971s-3s.
- 1003. Raab M, Neuhuber WL. Glutamatergic functions of primary afferent neurons with special emphasis on vagal afferents. Int Rev Cytol 2007;256:223–75.
- 1004. Seagard JL, Dean C, Hopp FA. Role of glutamate receptors in transmission of vagal cardiac input to neurones in the nucleus tractus solitarii in dogs. J Physiol 1999;520:243–53.
- 1005. Hudzik TJ, De Costa BR, McMillan DE. Sigma receptormediated emetic response in pigeons: agonists, antagonists and modifiers. Eur J Pharmacol 1993;236:279–87.
- 1006. Brooks B, Thisted R, Appel S, et al. Treatment of pseudobulbar affect in ALS with dextromethorphan/quinidine: a randomized trial. Neurology 2004;63:1364–70.
- 1007. Junien JL, Gue M, Pascaud X, et al. Selective stimulation of colonic motor response to a meal by sigma ligands in dogs. Gastroenterology 1990;99:684–9.
- 1008. Gue M, Gleizes-Escala C, Del Rio-Lacheze C, Junien J, Bueno L. Reversal of CRF- and dopamine-induced stimulation of colonic motility by CCK and igmesine (JO 1784) in the rat. Br J Pharmacol 1994;111:930–4.
- 1009. Gue M, Junien J, Del Rio C, Bueno L. Neuropeptide Y and sigma ligand (JO 1784) suppress stress-induced colonic motor disturbances in rats through sigma and cholecystokinin receptors. J Pharm Exp Therap 1992;261:850–5.
- 1010. Carr D, De Costa B, Radesca L, Blalock J. Functional assessment and partial characterization of [3H](+)-pentazocine binding sites on cells of the immune system. J Neuroimmunol 1991;35: 153–66.
- 1011. Coccini T, Manzo L, Costa L. 3H-spiperone labels sigma receptors, not dopamine D2 receptors, in rat and human lymphocytes. Immunopharmacology 1991;22:93–105.
- 1012. Iniguez MA, Punzon C, Nieto R, et al. Inhibitory effects of sigma-2 receptor agonists on T lymphocyte activation. Front Pharmacol 2013;4:1–12.
- 1013. Zhou Q, Kwan HY, Chan HC, et al. Blockage of voltage-gated K+ channels inhibits adhesion and proliferation of hepatocarcinoma cells. International J Mol Med 2003;11:261–6.
- 1014. Carayon P, Bouaboula M, Loubet J, et al. The sigma ligand SR 31747 prevents the development of acute graft-versus-host disease in mice by blocking IFN-gamma and GM-CSF mRNA expression. Int J Immunopharmacol 1995;17:753–61.

- Pellegrino T, Bayer BM. In vivo effects of cocaine on immune cell function. J Neuroimmunol 1998;83:139–47.
- 1016. Xu W, Flick T, Mitchel J, et al. Cocaine effects on immunocompetent cells: an observation of in vitro cocaine exposure. Int J Immunopharmacol 1999;21:463–72.
- 1017. Friesland M, Mingorance L, Chung J, et al. Sigma-1 receptor regulates early steps of viral RNA replication at the onset of hepatitis C virus infection. J Virol 2013;87:6377–90.
- 1018. Boehme K, Guglielmi K, Dermody T. Reovirus nonstructural protein sigmals is required for establishment of viremia and systemic dissemination. Proc Natl Acad Sci USA 2009;106: 19986–91.
- 1019. van Waarde A, Rybczynska AA, Ramakrishnan N, et al. Sigma receptors in oncology: therapeutic and diagnostic applications of sigma ligands. Current Pharm Design 2010;16:3519–37.
- 1020. Khansari N, Whitten HD, Chou YK, Fudenberg HH. Effect of polyvinyl pyrrolidone derivative compound on production of interleukin-1 by monocytes. Biomedi Pharmacotherap 1984;38: 308–11.
- 1021. Dornand J, Kamenka J, Bartegi A, Mani J. PCP and analogs prevent the proliferative response of T lymphocytes by lowering IL2 production. An effect related to the blockade of mitogentriggered enhancement of free cytosolic calcium concentration. Biochem Pharmacol 1987;36:3929–36.
- 1022. Megalizzi V, Le Mercier M, Decaestecker C. Sigma receptors and their ligands in cancer biology: overview and new perspectives for cancer therapy. Med Res Rev 2012;32:410–27.
- 1023. Tucci M, Quatraro C, Dammacco F, Silvestris F. Role of active drug transporters in refractory multiple myeloma. Curr Topics Med Chem 2009;9:218–24.
- 1024. Thomas GE, Szucs M, Mamone JY, et al. Sigma and opioid receptors in human brain tumors. Life Sci 1990;46:1279–86.
- 1025. Zamora PO, Moody TW, John CS. Increased binding to sigma sites of N-[1'(2-piperidinyl)ethyl)-4-[I-125]-iodobenzamide (I-125-PAB) with onset of tumor cell proliferation. Life Sci 1998; 63:1611–18.
- 1026. Ryan-Moro J, Chien CC, Standifer KM, Pasternak GW. Sigma binding in a human neuroblastoma cell line. Neurochem Res 1996;21:1309–14.
- 1027. Wheeler KT, Wang LM, Wallen CA, et al. Sigma-2 receptors as a biomarker of proliferation in solid tumours. Br J Cancer 2000;82: 1223–32.
- 1028. Rybczynska AA, Dierckx RA, Ishiwata K, et al. Cytotoxicity of sigma-receptor ligands is associated with major changes of cellular metabolism and complete occupancy of the sigma-2 subpopulation. J Neuclear Med 2008;49:2049–56.
- 1029. Simony-Lafontaine J, Esslimani M, Bribes E, et al. Immunocytochemical assessment of sigma-1 receptor and human sterol isomerase in breast cancer and their relationship with a series of prognostic factors. Br J Cancer 2000;82:1958–66.
- 1030. Mach RH, Smith CR, Al-Nabulsi I, et al. Sigma 2 receptors as potential biomarkers of proliferation in breast cancer. Cancer Res 1997;57:156–61.
- 1031. John CS, Gulden ME, Vilner BJ, Bowen WD. Synthesis, in vitro validation and in vivo pharmacokinetics of [125I]N-[2-(4iodophenyl)ethyl]-N-methyl-2-(1-piperidinyl) ethylamine: a high-affinity ligand for imaging sigma receptor positive tumors. Nuclear Med Biol 1996;23:761–6.
- 1032. John CS, Lim BB, Geyer BC, et al. 99mTc-labeled sigmareceptor-binding complex: synthesis, characterization, and specific binding to human ductal breast carcinoma (T47D) cells. Bioconj Chem 1997;8:304–9.
- 1033. Dittmann H, Coenen H, Zolzer F, et al. In vitro studies on the cellular uptake of melanoma imaging aminoalkyl-iodobenzamide derivatives (ABA). Nuclear Med Biol 1999;26:51–6.
- 1034. Eisenhut M, Hull W, Mohammed A, et al. Radioiodinated N-(2diethylaminoethyl)benzamide derivatives with high melanoma uptake: structure-affinity relationships, metabolic fate, and intracellular localization. J Med Chem 2000;43:3913–22.
- 1035. Everaert H, Flamen P, Franken P, et al. Sigma-receptor imaging by means of I123-IDAB scintigraphy: clinical application in melanoma and non-small cell lung cancer. AntiCancer Res 1997; 17:1577–82.
- 1036. Friebe M, Mahmood A, Bolzati C, et al. [99mTc]oxotechnetium(V) complexes amine-amide-dithiol

chelates with dialkylaminoalkyl substituents as potential diagnostic probes for malignant melanoma. J Med Chem 2001;44: 3132–40.

- 1037. Choi S, Yang B, Plossl K, et al. Development of a Tc-99m labeled sigma-2 receptor-specific ligand as a potential breast tumor imaging agent. Nuclear Med Biol 2001;28:657–66.
- 1038. John CS, Vilner BJ, Geyer BC, et al. Targeting sigma receptorbinding benzamides as in vivo diagnostic and therapeutic agents for human prostate tumors. Cancer Res 1999;59:4578–83.
- 1039. Shiue C, Shiue GG, Benard F, et al. N-(n-Benzylpiperidin-4-yl)-2-[18F]fluorobenzamide: a potential ligand for PET imaging of breast cancer. Nuclear Med Biol 2000;27:763–7.
- 1040. John CS, Gulden ME, Li J, et al. Synthesis, in vitro binding, and tissue distribution of radioiodinated 2-[1251]N-(N-benzylpiperidin-4-yl)-2-iodo benzamide, 2-[1251]BP: a potential sigma receptor marker for human prostate tumors. Nuclear Med Biol 1998;25:189–94.
- 1041. Abate C, Ferorelli S, Contino M, et al. Arylamides hybrids of two high-affinity sigma2 receptor ligands as tools for the development of PET radiotracers. European J Med Chem 2011; 46:4733–41.
- 1042. Zeng C, Vangveravong S, Xu J, et al. Subcellular localization of sigma-2 receptors in breast cancer cells using two-photon and confocal microscopy. Cancer Res 2007;67:6708–16.
- 1043. van Waarde A, Jager PL, Ishiwata K, et al. Comparison of sigmaligands and metabolic PET tracers for differentiating tumor from inflammation. J Neuclear Med 2006;47:150–4.
- 1044. Spirkoski J, Melo FR, Grujic M, et al. Mast cell apoptosis induced by siramesine, a sigma-2 receptor agonist. Biochem Pharmacol 2012;84:1671–80.
- 1045. Brent P, Pang G. Sigma binding site ligands inhibit cell proliferation in mammary and colon carcinoma cell lines and melanoma cells in culture. Eur J Pharmacol 1995;278:151–60.
- 1046. Cho E, Zhang Y, Cai X, Moran C, Wang L, Xia Y. Quantitative analysis of the fate of gold nanocages in vitro and in vivo after uptake by U87-MG tumor cells. Angew Chem 2013;52:1152–5.
- 1047. Wang Y, Xu J, Xia X, et al. SV119-gold nanocage conjugates: a new platform for targeting cancer cells via sigma-2 receptors. Nanoscale 2012;4:421–4.
- 1048. Moebius FF, Reiter RJ, Hanner M, Glossmann H. High affinity of sigma 1-binding sites for sterol isomerization inhibitors: evidence for a pharmacological relationship with the yeast sterol C8-C7 isomerase. Br J Pharmacol 1997;121:1–6.
- 1049. Fraser S, Grimes J, Diss J, et al. Predominant expression of Kv1.3 voltage-gated K+ channel subunit in rat prostate cancer cell lines: electrophysiological, pharmacological and molecular characterisation. Pflugers Archiv 2003;446:559–71.
- 1050. Fraser S, Grimes J, Djamgoz M. Effects of voltage-gated ion channel modulators on rat prostatic cancer cell proliferation: comparison of strongly and weakly metastatic cell lines. The Prostate 2000;44:61–76.
- 1051. Fraser S, Koyuturk M, Djamgoz M. Ion channel activity and cancer cell proliferation: a short review with particular reference to prostate cancer. Trivendrum, India: Research Signpost; 2002.
- 1052. Fraser S, Salvador V, Manning E, et al. Contribution of functional voltage-gated Na+ channel expression to cell behaviors involved in the metastatic cascade in rat prostate cancer: I. Lateral motility. J Cell Physiol 2003;195:479–87.
- 1053. Renaudo A, L'Hoste S, Guizouarn H, et al. Cancer cell cycle modulated by a functional coupling between sigma-1 receptors and Cl-channels. J Biol Chem 2007;282:2259–67.
- 1054. Ouadid-Ahidouch H, Chaussade F, Roudbaraki M, et al. KV1.1 K(+) channels identification in human breast carcinoma cells: involvement in cell proliferation. Biochem Biophys Res Commun 2000;278:272–7.
- 1055. Woodfork KA, Wonderlin WF, Peterson VA, Strobl JS. Inhibition of ATP-sensitive potassium channels causes reversible cell-cycle arrest of human breast cancer cells in tissue culture. J Cell Physiol 1995;162:163–71.
- 1056. Yao X, Kwan HY. Activity of voltage-gated K+ channels is associated with cell proliferation and Ca<sup>2+</sup> influx in carcinoma cells of colon cancer. Life Sci 1999;65:55–62.
- 1057. Wonderlin WF, Strobl JS. Potassium channels, proliferation and G1 progression. J Membrane Biol 1996;154:91–107.

- 388 C. G. Rousseaux & S. F. Greene
- 1058. Klimatcheva E, Wonderlin WF. An ATP-sensitive K(+) current that regulates progression through early G1 phase of the cell cycle in MCF-7 human breast cancer cells. J Membrane Biol 1999;171:35–46.
- 1059. Nilius B, Wohlrab W. Potassium channels and regulation of proliferation of human melanoma cells. J Physiol 1992;445: 537–48.
- 1060. Schwab A, Reinhardt J, Schneider SW, et al. K(+) channeldependent migration of fibroblasts and human melanoma cells. Cell Physiol Biochem 1999;9:126–32.
- 1061. Wang H, Zhang Y, Cao L, et al. HERG K+ channel, a regulator of tumor cell apoptosis and proliferation. Cancer Res 2002;62: 4843–8.
- 1062. Cantiello H, Prat A, Bonventre J, et al. Actin-binding protein contributes to cell volume regulatory ion channel activation in melanoma cells. J Biol Chem 1993;268:4596–9.
- 1063. Shen MR, Chou CY, Hsu KF, et al. Modulation of volumesensitive Cl – channels and cell volume by actin filaments and microtubules in human cervical cancer HT-3 cells. Acta Physiol Scand 1999;167:215–25.
- 1064. Mycielska ME, Fraser SP, Szatkowski M, Djamgoz MB. Contribution of functional voltage-gated Na+ channel expression to cell behaviors involved in the metastatic cascade in rat prostate cancer: II. Secretory membrane activity. J Cell Physiol 2003;195: 461–9.
- 1065. Diss J, Archer S, Hirano J, et al. Expression profiles of voltagegated Na(+) channel alpha-subunit genes in rat and human prostate cancer cell lines. Prostate 2001;48:165–78.
- 1066. Grimes J, Fraser S, Stephens G, et al. Differential expression of voltage-activated Na+ currents in two prostatic tumour cell lines: contribution to invasiveness in vitro. FEBS Lett 1995;369:290–4.
- 1067. Smith P, Rhodes NP, Shortland AP, et al. Sodium channel protein expression enhances the invasiveness of rat and human prostate cancer cells. FEBS Lett 1998;423:19–24.
- 1068. Cairns R, Khokha R, Hill R. Molecular mechanisms of tumor invasion and metastasis: an integrated view. Curr Mol Med 2003; 3:659–71.
- 1069. Pawlak G, Helfman DM. Cytoskeletal changes in cell transformation and tumorigenesis. Curr Opin Genet Dev 2001;11:41–7.
- 1070. Cassano G, Gasparre G, Niso M, et al. F281, synthetic agonist of the sigma-2 receptor, induces Ca2+ efflux from the endoplasmic reticulum and mitochondria in SK-N-SH cells. Cell Calcium 2009;45:340–5.

- 1071. Gardner B, Zhu L, Roth M, et al. Cocaine modulates cytokine and enhances tumor growth through sigma receptors. J Neuroimmunol 2004;147:95–8.
- 1072. Kedjouar B, Daunes S, Vilner BJ, et al. Structural similitudes between cytotoxic antiestrogen-binding site (AEBS) ligands and cytotoxic sigma receptor ligands. Evidence for a relationship between cytotoxicity and affinity for AEBS or sigma-2 receptor but not for sigma-1 receptor. Biochem Pharmacol 1999;58: 1927–39.
- 1073. Megalizzi V, Mathieu V, Mijatovic T, et al. 4-IBP, a sigmal receptor agonist, decreases the migration of human cancer cells, including glioblastoma cells, in vitro and sensitizes them in vitro and in vivo to cytotoxic insults of proapoptotic and proautophagic drugs. Neoplasia (NY) 2007;9:358–69.
- 1074. Aydar E, Onganer P, Perrett R, et al. The expression and functional characterization of sigma (sigma) 1 receptors in breast cancer cell lines. Cancer Lett 2006;242:245–57.
- 1075. Zheng K, Scimemi A, Rusakov DA. Receptor actions of synaptically released glutamate: the role of transporters on the scale from nanometers to microns. Biophys J 2008;95: 4584–96.
- 1076. Nordenberg J, Perlmutter I, Lavie G, et al. Anti-proliferative activity of haloperidol in B16 mouse and human SK-ME L-28 melanoma cell lines. Int J Oncol 2005;27:1097–103.
- 1077. Hornick JR, Spitzer D, Goedegebuure P, et al. Therapeutic targeting of pancreatic cancer utilizing sigma-2 ligands. Surgery 2012;152:S152–6.
- 1078. Kishi T, Yoshimura R, Okochi T, et al. Association analysis of SIGMAR1 with major depressive disorder and SSRI response. Neuropharmacology 2010;58:1168–73.
- 1079. Martin-Fardon R, Strong EM, Weiss F. Effect of sigma(1) receptor antagonism on ethanol and natural reward seeking. Neuroreport 2012;23:809–13.
- 1080. Wegleiter K, Hermann M, Posod A, et al. The sigma-1 receptor agonist 4-phenyl-1-(4-phenylbutyl) piperidine (PPBP) protects against newborn excitotoxic brain injury by stabilizing the mitochondrial membrane potential in vitro and inhibiting microglial activation in vivo. Exp Neurol 2014;261c:501–9.
- 1081. Delgado P, Charney D, Price L, et al. Serotonin function and the mechanism of antidepressant action. Reversal of antidepressantinduced remission by rapid depletion of plasma tryptophan. Arch Gen Psychiatry 1990;47:411–18.