

Review

ω -3 Fatty Acids and Cardiovascular Diseases: Effects, Mechanisms and Dietary Relevance

Hanne K. Maehre *, Ida-Johanne Jensen, Edel O. Elvevoll and Karl-Erik Eilertsen

Norwegian College of Fishery Science Faculty of Biosciences, Fisheries and Economics,
UIT The Arctic University of Norway, N-9037 Tromsø, Norway;

E-Mails: ida-johanne.jensen@uit.no (I.-J.J.); edel.elvevoll@uit.no (E.O.E.);

karl-erik.eilertsen@uit.no (K.-E.E.)

* Author to whom correspondence should be addressed; E-Mail: hanne.maehre@uit.no;
Tel.: +47-7764-6793; Fax: +47-7764-6020.

Academic Editor: Charles Brennan

Received: 30 June 2015 / Accepted: 9 September 2015 / Published: 18 September 2015

Abstract: ω -3 fatty acids (*n*-3 FA) have, since the 1970s, been associated with beneficial health effects. They are, however, prone to lipid peroxidation due to their many double bonds. Lipid peroxidation is a process that may lead to increased oxidative stress, a condition associated with adverse health effects. Recently, conflicting evidence regarding the health benefits of intake of *n*-3 from seafood or *n*-3 supplements has emerged. The aim of this review was thus to examine recent literature regarding health aspects of *n*-3 FA intake from fish or *n*-3 supplements, and to discuss possible reasons for the conflicting findings. There is a broad consensus that fish and seafood are the optimal sources of *n*-3 FA and consumption of approximately 2–3 servings per week is recommended. The scientific evidence of benefits from *n*-3 supplementation has diminished over time, probably due to a general increase in seafood consumption and better pharmacological intervention and acute treatment of patients with cardiovascular diseases (CVD).

Keywords: *n*-3 fatty acids; eicosapentaenoic acids (EPA); docosahexaenoic acid (DHA); lipid peroxidation; cardiovascular diseases; seafood; supplements

1. Introduction

The story of ω -3 fatty acids (*n*-3 FA) and their impact on health and diseases began in the 1970s when it was discovered that the Inuit in Greenland had a markedly lower incidence of cardiovascular diseases (CVD) than the Danish population. The Danish researchers Bang and Dyerberg conducted several studies comparing plasma lipid profiles and food habits between the two populations. Their main conclusion was that there was a correlation between diet compositions in the two populations and the frequency of CVD. A large difference in intake of the long-chain polyunsaturated fatty acids (PUFA) eicosapentaenoic acids (EPA, C20:5, *n*-3) and docosahexaenoic acid (DHA, C22:6, *n*-3) was concluded to be the main explanatory factor for this phenomenon [1–4]. Recently, several papers have been published raising questions about the conduction of Bang and Dyerberg's research. The main issues in these papers are that the systems for health monitoring and cause of death registration in Greenland at the time were inadequate due to the dispersed settlements [5,6].

Nevertheless, numerous studies on the relationship between seafood or EPA + DHA supplements and several diseases have been performed and positive effects of a high intake have been documented for a range of different medical conditions, in particular conditions related to inflammatory processes.

However, there is a possible drawback associated with intake of refined and concentrated *n*-3 PUFA. Due to their high content of double bonds, they are especially prone to lipid peroxidation. Metabolites from lipid peroxidation have been under suspicion for negatively affecting biological processes. A high intake of *n*-3 PUFA could thus be a double-edged sword.

Seafood consumption has traditionally been a robust measure in epidemiological studies, showing a clear correlation between a high seafood intake and reduced risk of CVD. Various fish oil supplements have been tested in clinical trials, giving partly conflicting results compared to the epidemiological studies. In this review, recent literature regarding different health aspects of *n*-3 PUFA intake from different sources will be discussed and possible reasons for conflicting findings between them will be addressed.

2. Cardiovascular Diseases

Cardiovascular disease is a collective term comprising a group of disorders of the heart and blood vessels. These diseases are the largest cause of morbidity and premature death worldwide and accounted for 17.5 million deaths, or 31% of all deaths, in 2012 [7]. The two most frequent disorders are coronary heart disease (CHD) and cerebrovascular disease (stroke), making up 42% and 38% of CVD deaths, respectively. There are several risk factors associated with the development of CVD, among them dyslipidemia, hypertension, tobacco smoking, obesity and diabetes mellitus [8]. All of these risk factors are associated with oxidative stress, which is a condition that arises when there is an imbalance between pro-oxidants (such as reactive oxygen species, ROS) and antioxidants (such as glutathione) in the body. A combination of multiple factors increases the risk considerably, the rule of thumb being the greater the level of each risk factor, the greater the total risk [9].

One of the major underlying causes for CVD is atherosclerosis, which is a complex, multifactorial and progressive inflammatory condition affecting the arteries. Intima, the inner layer of arteries, consists of endothelial cells (EC) with a vast range of metabolic and regulatory functions. These include transport of metabolic substances, regulation of vascular tone, defense against inflammation, angiogenesis and

regulation of hemostasis and coagulation [10]. A key substance in these regulatory processes is the vasodilator nitric oxide (NO) and reduced bioavailability of this compound, for instance by increased oxidative stress, may lead to an activation of the EC [11], which over time may evolve into a condition called endothelial dysfunction. Activation of EC leads to an inflammatory response involving the production of a cascade of chemokines (monocyte chemoattractant protein-1 (MCP-1), macrophage colony-stimulating factor (M-CSF), interleukin-8 (IL-8) *etc.*), adhesion factors (vascular cell adhesion molecule-1 (VCAM-1), intercellular adhesion molecule-1 (ICAM-1), selectins (endothelial and platelet selectins (E-selectin and P-selectin) *etc.*) and integrins (very late antigen 1 and 4 (VLA-1 and VLA-4) *etc.*) produced by both circulating blood and endothelial cells. These released substances are involved in the concurrent processes of further recruitment of monocytes to the endothelial surface, followed by their adherence and transmigration into the vessel wall intima. The influx of monocytes is often accompanied by influx of other inflammation cells, such as T-cells, dendritic cells and mast cells. Once placed in the intima, monocytes may differentiate into macrophages influenced by pro-inflammatory cytokines (IFN- γ , IL-1 β , TNF α , *etc.*). Macrophages are phagocytic cells expressing scavenger receptors for uptake of modified low density lipoprotein (LDL). In these activated macrophages, programmed to protect our bodies from danger, the normal processes for cholesterol handling and transport is impaired and accumulation of cholesteryl esters will eventually lead to the formation of foam cells, also known as fatty streaks [12]. Continued inflammatory responses may further accelerate the progression of atherosclerotic lesions. Stimulation of proliferation and migration of smooth muscle cells (SMC) may build up a large plaque inside the intima. Protease secretion by macrophages degrade extracellular matrix, such as collagen, and a fibrous cap is formed around the excess lipids. Expression of collagen degrading enzymes, such as matrix metalloproteinases (MMP) can gradually weaken the fibrous cap leading to plaque rupture and release of intracellular content into the arteries, thrombus formation and this may eventually result in myocardial infarction (MI) [13].

3. ω -3 Fatty Acids and Their Metabolites

3.1. General

The common chemical structure of FA is an aliphatic hydrocarbon chain of with a carboxylic acid group at one end and a methyl group at the other end. The length of the carbon chain normally ranges between 4 and 22 carbon atoms and they are covalently bound together with only single bonds, with one carbon-carbon double bond or with up to six double bonds. The respective fatty acids are classified as saturated, monounsaturated or polyunsaturated FA, depending on the number of double bonds in the chain. Unsaturated FA are further classified by the so-called “ ω ” denotation, which indicates the placement of the first double bond in the chain, counted from the methyl end of the C-chain.

Fatty acids are present in a range of different structures in the body and they also have a range of different properties. Bound in triacylglycerols (TAG) they serve as energy storage, and bound in phospholipids (PL) they serve as integral parts of cellular membranes. They are also origin for a range of compounds with specific biological functions and some of these are of particular interest when it comes to health issues and development of different medical conditions/chronic diseases.

3.2. Eicosanoids and Specialized Pro-Resolving Mediators (SPM)

A wide range of compounds may be enzymatically synthesized from FA containing 20 or 22 C-atoms. Several of these metabolites, such as eicosanoids and so-called specialized pro-resolving mediators (SPM), have impact on acute and chronic inflammatory responses.

Eicosanoids are hormone-like substances originating from FA containing 20 C-atoms, namely dihomo- γ -linolenic acid (DGLA, C20:3, *n*-6), arachidonic acid (AA, C20:4, *n*-6) and EPA. Eicosanoids include prostaglandins (PG), prostacyclins (PI), thromboxanes (TX) and leukotrienes (LT). All subclasses of eicosanoids may be synthesized from all of the three mentioned FA; however, nomenclature, properties and potencies depend on which FA is the origin [14]. Nomenclature of the eicosanoids are related to the number of double bonds in the structure and hence, prostaglandins derived from AA are categorized as 2-series PG, while those derived from EPA are categorized as 3-series PG. Leukotrienes from the two FA are categorized as 4-series LT and 5-series LT, respectively.

The SPM comprise lipoxins, resolvins, maresins and protectins, all inflammation-resolving compounds which can be derived from AA (lipoxins), EPA (resolvins) and DHA (resolvins, maresins and protectins) [15].

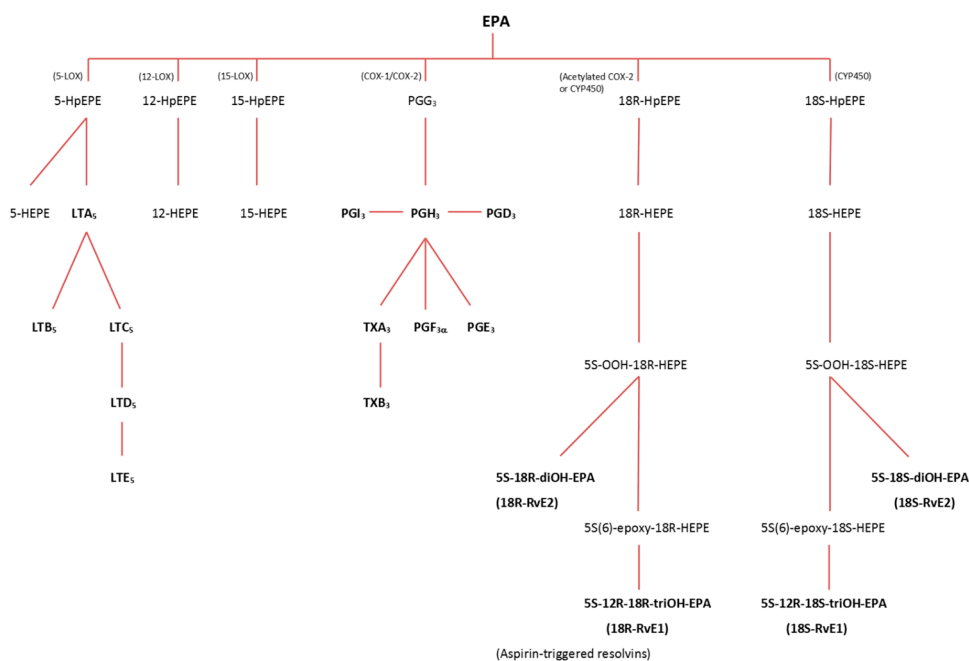


Figure 1. Overview of the pathways involved in the production of eicosanoids and specialized pro-resolving mediators from eicosapentaenoic acid (EPA). The figure is adapted from [16,17]. Products derived through the lipoxygenase (LOX) pathway are 5-, 12- and 15-HEPE, along with the leukotrienes A to E (LTA, LTB, LTC, LTD and LTE). Through the cyclooxygenase (COX) pathway, the prostaglandins D to I (PGD, PGE, PGF, PGG, PGH and PGI) and thromboxanes A and B (TXA and TXB) are produced. Aspirin-acetylated COX-2 catalyzes the production of aspirin-triggered resolvins E1 (18R-Rv1) and E2 (18R-Rv2), while the regular resolvins E1 (18S-Rv1) and E2 (18S-Rv2) are catalyzed by cytochrome P-450 (CYP-450). HpEPE: hydroperoxyeicosapentaenoic acids; HEPE: hydroxyeicosapentaenoic acids.

Synthesis of all of these compounds from AA, EPA and DHA starts with the liberation of the FA from the membrane phospholipids catalysed by phospholipase A₂. Several enzymes are involved in the following synthesis pathway (Figures 1 and 2), the main classes being lipoxygenases (LOX), cyclooxygenases (COX) and cytochrome P450 (CYP) [14]. The metabolic pathways for formation of the eicosanoids and SPM are similar independent of the originating FA, and in this review, only the intermediary products of EPA and DHA will be presented.

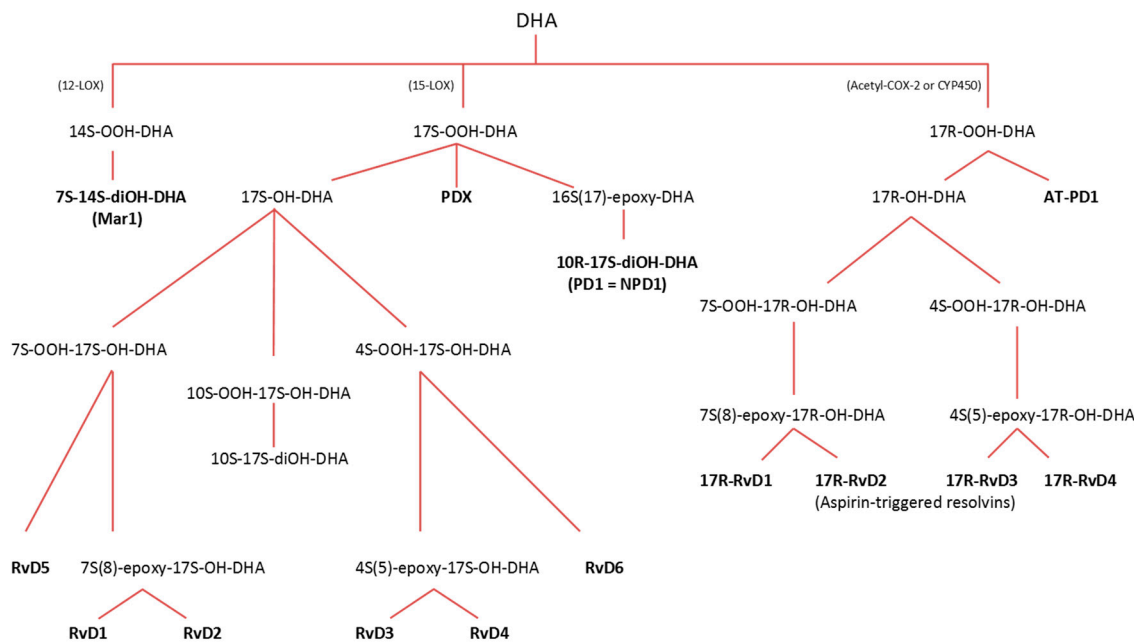


Figure 2. Overview of the pathways involved in the production of specialized pro-resolving mediators from docosahexaenoic acid (DHA). The figure is adapted from [17]. Through the lipoxygenase (LOX) pathway, LOX-12 catalyzes the production of maresin-1 (Mar-1), while 15-LOX catalyzes the formation of neuroprotectin D1 (NPD1), protectin DX (PDX) and resolvins D1 (17S-RvD1) to D6 (17S-RvD6). Aspirin-acetylated cyclooxygenase-2 (COX-2) and/or cytochrome P-450 (CYP-450) catalyzes the formation of aspirin-triggered resolvins D1 (17R-RvD1) to D4 (17R-RvD4), along with aspirin-triggered protectin D1 (AT-PD1).

3.2.1. Lipoxygenase (LOX) Pathway

There are three major lipoxygenases (5-, 12- and 15-LOX) involved in the formation of eicosanoids and SPM [18]. These enzymes catalyse the insertion of a hydroperoxy group in the C-chain of the FA. The numbering of the LOX corresponds to the C-atom on which the insertion takes place. The resulting compounds are hydroperoxyeicosapentaenoic acids (HpEPE) which is further rapidly reduced to hydroxyeicosapentaenoic acids (HEPE).

The most common products derived from EPA via the LOX-pathways are 5-series LTs. Their formation is catalysed by 5-lipoxygenase (5-LOX), which is activated by 5-lipoxygenase activating protein (FLAP) and converts EPA to LTA₅ (Figure 3) via dehydration of the unstable 5-hydroperoxy-eicosapentaenoic acid (5-HpEPE). Several enzymatic reactions in different cells thereafter convert LTA₅ to other LT, namely LTB₅, LTC₅, LTD₅ and LTE₅ [19].

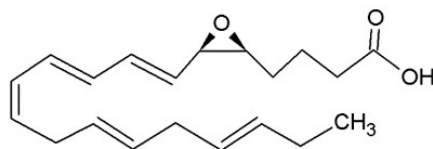


Figure 3. Leukotriene A₅.

Also, DHA may act as a substrate for different LOX pathways, forming resolvins of the D-series, along with maresin 1 and protectin D1. Maresin 1 is formed via the 12-LOX pathway, while D-series resolvins (D1–D6) and protectin D1 are formed via the LOX-15 pathway [15,17].

3.2.2. Cyclooxygenase (COX) Pathway

Prostaglandins, prostacyclins and thromboxanes are formed by enzymatic conversion of EPA by prostaglandin-endoperoxidase synthase (PTGS), more commonly known as cyclooxygenase (COX). This enzyme has two main isoforms involved in the synthesis of prostaglandins, COX-1 and COX-2, having both distinct and common properties [20,21]. Also, a third COX-isoform (COX-3) has been discovered, but this isoform seems to have low or limited impact on the eicosanoid synthesis [22].

The eicosanoid synthesis starts with the insertion of two oxygen molecules to the EPA forming a cyclopentane ring structure between C8 and C12 and an oxygen bridge between C9 and C11. In addition, a hydroperoxy group is inserted in the *S*-chirality at C15. This intermediate compound is called prostaglandin PGG₃ (Figure 4) and is rapidly reduced to PGH₃ by peroxidase. Tissue-specific enzymes thereafter convert PGH₃ to the other PG, PI and TX.

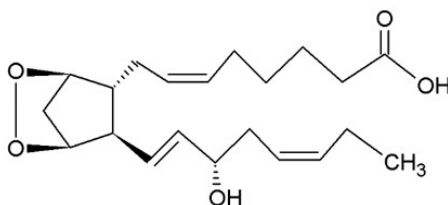


Figure 4. Prostaglandin G₃.

Both isoforms of the COX-enzyme may be acetylated by aspirin, leaving COX-1 deactivated. Aspirin-acetylated COX-2, on the other hand, may catalyse the insertion of a hydroperoxy group in the *R*-chirality at C18 in EPA resulting in the formation of 18*R*-hydroperoxy-eicosapentaenoic acid (18*R*-HpEPE). This peroxide is then converted into aspirin-triggered resolvins E1 and E2 by several other enzymatic reactions involving peroxidases, 5-LOX and epoxidases [17]. Aspirin-acetylated COX-2 may also trigger a similar process in DHA, resulting in the formation of aspirin-triggered resolvins D1-D4 or protectin PD1 [18].

3.2.3. Cytochrome P-450 (CYP) Pathway

The third enzymatic pathway important in the metabolism of LC-PUFAs is the cytochrome P-450 pathway. Here, CYP monooxidases inserts oxygen into the AA, EPA or DHA to yield either

hydroxylated or epoxidized FA. Hydroxylation takes place at C19 or C20 in EPA and C21 or C22 in DHA, while epoxidation may take place at each of the double bonds in the FA [23,24].

This pathway may also promote the formation of non-aspirin-triggered resolvins from EPA, via insertion of a hydroperoxy group in the *S* chirality of C18, forming 18*S*-HpEPE. In a process similar to the one described for aspirin-triggered resolvins, peroxidases, 5-LOX and epoxidases further convert this compound into resolvins [17].

3.2.4. Eicosanoids, SPM and Cellular Effects on Atherosclerosis

Eicosanoids are involved in a vast range of biological processes and the subspecies most relevant for the development of CVD are PGI, TXA and LTB. These compounds exert their effects on EC, monocytes, blood platelets and vascular smooth muscle cells (VSMC) [25], all of which are involved in the atherosclerotic process. Thromboxane A₂ affects aggregation of platelets and is a potent vasoconstrictor, while LTB₄ induces inflammation, leukocyte chemotaxis and adherence. Although their EPA-derived counterparts affect many of the same mechanisms, their potency is much lower and the adverse effects are not as pronounced. The third subspecies, PGI, have opposite effects as they serve as active vasodilators and inhibit platelet aggregation [26].

Only very few studies have been published showing the direct effects or mechanisms of the different eicosanoids on inflammation cascade factors and processes [27–29]. In contrast, as reviewed by Yates *et al.* [30], multiple cell-based studies have shown indirect effects where addition of *n*-3 FA reduces the inflammatory responses effectively by downregulating or inhibiting expression of these cascade compounds. In some comparative studies, DHA has been shown to be more effective than EPA in attenuating the effects of these compounds [31,32].

The anti-inflammatory mechanisms of the SPM vary between the different compounds, but some common features are reduction of migration of activated immune cells to the injured endothelium and stimulation of phagocytic ability of macrophages. Comprehensive overviews of the anti-inflammatory and inflammation resolving properties of the SPM are presented in several reviews [19,33].

3.3. Lipid Peroxidation and Its Products

Polyunsaturated fatty acids are, due to unstable double bonds, prone to oxidation. The incidence of oxidation increases with the number of double bonds present and hence, the long chain *n*-3 FA are especially exposed. Lipid oxidation may occur either in foods or endogenously and is a three-phased chain reaction divided into initiation, propagation and termination (Figure 5). The initiation phase, triggered by energy (as for instance oxygen, light energy and radiation), ROS and transition metals, starts with the abstraction of the unstable H-atom between the double bonds of the PUFA resulting in the formation of an alkyl radical (L•) and a hydrogen radical (H•). Delocalisation of the remaining double bonds result in the formation of conjugated double bonds, either in *cis* or *trans* configuration. Due to increased stability, the *trans* configuration dominates. In the propagation phase, oxygen is added to the alkyl radical, forming a peroxy radical (LOO•). The peroxy radical reacts with another unstable H-atom in a PUFA, forming a peroxide (LOOH) and a new alkyl radical and the chain reaction has commenced. Peroxides are known as primary oxidation products. They are unstable and easily degraded to alkoxy radicals (LO•) and hydroxyl radicals (•OH). The alkoxy radical can either affect the initiation

phase, forming a hydroxylated FA (LOH) or it can be decomposed into smaller molecules, such as alcohols, aldehydes and ketones. The amount of radicals, peroxides and oxidation products increase exponentially up until the radicals start reacting with each other, forming stable polymeric products. This phase is known as the termination phase. All unsaturated FA may undergo this process. The products formed depend on the original FA and each of them can be the source of multiple products. The possible range of lipid peroxidation products is therefore enormous. Some of the most studied lipid peroxidation products are reactive aldehydes, such as malondialdehyde (MDA), and 4-hydroxy-2-alkenals [34–36]. While MDA can be derived from all FA with three or more double bonds, 4-hydroxy-2-nonenal (HNE) is derived from *n*-6 PUFA and 4-hydroxy-2-hexenal (HHE) is derived from *n*-3 PUFA.

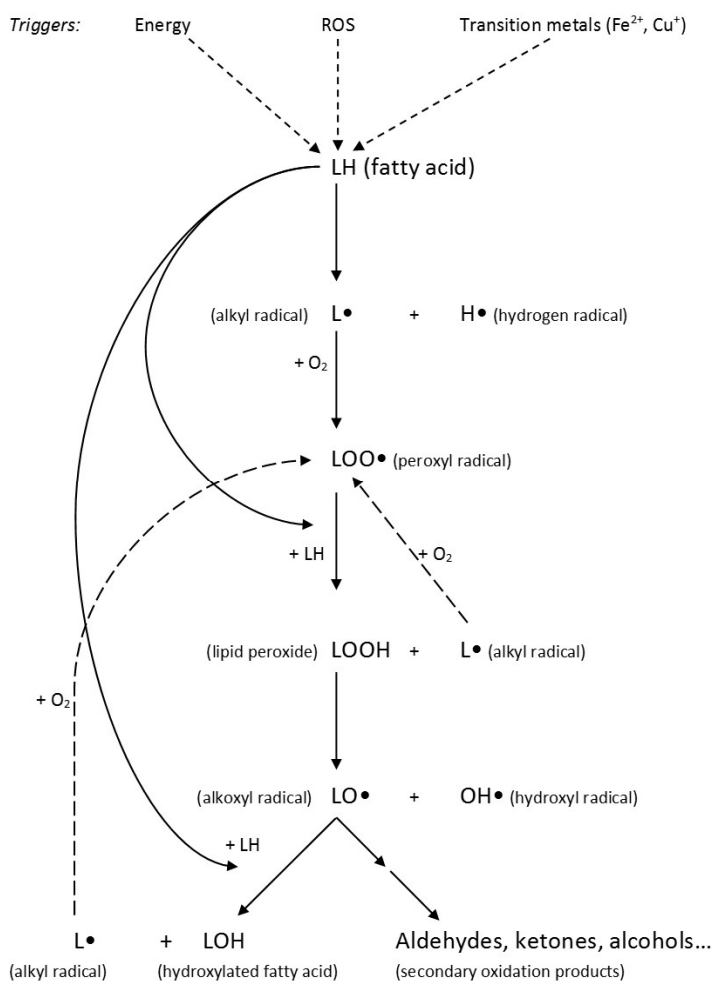


Figure 5. Illustration of the lipid peroxidation process. Triggered by energy, reactive oxygen species (ROS) and/or transition metals, an unstable hydrogen atom is abstracted from a fatty acid (LH), forming an alkyl radical ($\text{L}\cdot$) and a hydrogen radical ($\text{H}\cdot$). The alkyl radical reacts with oxygen, forming a peroxy radical ($\text{LOO}\cdot$) that further reacts with another fatty acid, forming a lipid peroxide (LOOH) and a new alkyl radical. The lipid peroxide is easily degraded into an alkoxy radical ($\text{LO}\cdot$) and a hydroxyl radical ($\text{OH}\cdot$). The alkoxy radical may either react with a new fatty acid, forming a hydroxylated fatty acid (LOH), or decompose into smaller, volatile compounds, such as aldehydes, ketones or alcohols.

Lipid Peroxidation Products and Cellular Effects on Atherosclerosis

The free radicals formed during lipid oxidation may contribute to oxidative stress, a condition which stimulates the activation of nuclear factor κ B (NF κ B) and subsequently the production of inflammatory chemokines, adhesion factors and cytokines [37]. In addition, both the free radicals and the primary lipid oxidation products (lipid peroxides) may accelerate further lipid peroxidation [38]. The presence of primary oxidation products in foods is not easily detected, as they are tasteless and odourless. The secondary oxidation products, however, are formed rapidly and are more easily detected in oxidized foods because of their volatility, which causes a rancid taste and odour.

In toxicology studies, HNE and HHE showed equal cytotoxic properties on rat cortical neurons and also on the depletion of the endogenous antioxidant glutathione [39]. Activation of NF κ B, which stimulates the production of the previously described chemokines, adhesion factors and cytokines, has been shown for HHE [40]. In contrast, in low concentrations, 4-HHE has been shown to induce the antioxidant enzyme heme oxygenase-1 (HO-1) through activation of the nuclear factor-erythroid-2-related factor-2 (Nrf2) and thereby protect vascular cells from cytotoxicity induced by oxidative stress [41].

A property of reactive aldehydes especially relevant regarding CVD are their capability of forming covalent bonds to DNA, proteins and phospholipids [36]. After binding to the protein constituent of LDL (apoB), they contribute to oxidative modification of this particle. Oxidized LDL are triggers of endothelial dysfunction, which is crucial for the onset of atherosclerosis. They are also easily digested by macrophages and hence contribute to the development of foam cells [42] and later to fatty streaks and atherosclerotic lesions. Protein-bound HHE has also been detected in atherosclerotic lesions [43].

3.4. Clinical Effects of *n*-3 Intake on Atherosclerosis

An increased intake of *n*-3 FA has been shown to increase the phospholipid content of these FAs in both plasma membranes and red blood cells at the expense of AA. The relative proportion of EPA + DHA in the red blood cell membranes (as percentage of total FA), is termed the ω -3-index. This value has been shown to be dose-dependent on the *n*-3 FA intake [44,45] and a low index was first recognized as an independent risk marker and later as an actual risk factor for CHD death [46–48]. As EPA competes with AA in the COX and LOX-mediated eicosanoid production, a consequence of this is that the eicosanoid production also is shifted towards a higher level of 3-series prostaglandins and 5-series of leukotrienes at the cost of the 2-series prostaglandins and 4-series leukotrienes [49–51]. Also, the CYP-derived metabolites of PUFA are shifted when increasing the *n*-3 intake, indicating that EPA and DHA are preferred over AA as substrates in this metabolic pathway [52,53]. However, in subjects with high basal seafood consumption, additional supplementation with fish oil did not alter the FA composition of phospholipids [54].

In addition, biochemical markers of inflammation have been shown to be affected by increased intake of *n*-3 FA. Plasma levels of the cytokines IL-6 and TNF α , along with the acute phase inflammatory protein C-reactive protein (CRP), have been shown to decrease with increased *n*-3 intake in both healthy people [55–57] and in patients with CVD [58–60].

As described in Section 2, dyslipidemia is recognized as a risk factor for development of atherosclerosis. Most clinical studies show that an increased intake of *n*-3, either as fatty fish or as

supplements, decrease the plasma content of TAG [61–64]. Trends regarding cholesterol changes are not as conclusive. Several studies have shown a slight increase in LDL cholesterol levels following fish oil or *n*-3 supplementation [65,66]. It has been suggested that DHA, and not EPA, could be responsible for this phenomenon [67]. Other studies find no change in the LDL cholesterol content [61,63,64,68], yet others report a decreased LDL cholesterol level following increased intake of *n*-3 FA [69,70].

The effects of increased *n*-3 intake on blood pressure reduction are also inconclusive. Although some studies report an attenuation of blood pressure related to *n*-3 intake [71,72], others find no effect [73]. Systematic reviews on fish oil supplements and hypertension by Campbell *et al.* [74] and Miller *et al.* [75] show a modest improvement on blood pressure in hypertensive patients, but little or no effect on normotensive subjects.

Dietary Intake of Oxidized Lipids

Most of the existing literature on cardiovascular impact of intake of oxidized lipids is based on animal trials. Feeding mice with either oxidized *n*-3-rich PL or oxidised *n*-3-rich TAG resulted in increased plasma concentrations of 4-HHE and the inflammation markers IL-6 and MCP-1 compared to control groups receiving un-oxidised PL or TAG [76]. Other animal feeding trials with oxidized oils have shown reduced growth and increased production of inflammatory markers, and these effects were reduced with concomitant intake of antioxidants [77–79]. Yet other studies have reported a significant reduction of atherosclerotic plaques in animal models with simultaneous intake of antioxidants and marine oils [80,81].

Concerning the potential risks of human consumption of oxidized *n*-3, the available literature is scarce. Ottestad *et al.* [82] published a study showing no adverse effect of short-term intake of lightly oxidized oils in healthy volunteers. García-Hernandez *et al.* [83] came to the same conclusions for intake of lightly oxidized oils, while the conclusion for highly oxidized oils was that they have adverse effects on cholesterol levels. The duration of both these studies were, however, relatively short and the number of included subjects were low. Due to the suspicion of adverse biological effects of oxidized lipids, studies over a longer time-span and with more participants would be considered un-ethical and hence, such studies would be impossible to conduct.

4. Seafood or *n*-3 Supplements?

Fish and other seafood are the most important dietary sources of *n*-3 FA. Moreover, most marine species are low in *n*-6 FA, which make the *n*-6/*n*-3 ratio very low. As most seafood are subjected to some kind of household preparation prior to consumption, a range of studies focusing on retention of *n*-3 FA as a result of preparation have been performed. Most of them conclude that the absolute changes in *n*-3 content are low for most preparation techniques. However, when fat is added in the process, for instance in pan frying or deep frying, the total lipid content and the *n*-6/*n*-3 ratio is increased [84–87]. In addition to the *n*-3 FA, fish and seafood contain a wide range of other important nutrients, such as high-quality proteins, vitamin A, D, B₁₂ and several minerals [88]. Seafood is also rich in the sulfur-containing free amino acid taurine, which has been shown to hold a range of cardioprotective properties [61,89]. Also other endogenous compounds, such as anti-oxidative peptides protecting the *n*-3 PUFA from

peroxidation are embedded in the seafood [90]. Combinatory effects between *n*-3 FA and other beneficial compounds should therefore not be ruled out [91].

Crude fish oils are rich in *n*-3 FA, but they also contain other substances such as free fatty acids, phospholipids, pigments, sterols, metals and persistent organic pollutants (POP). Some of these are detrimental for health (such as POP) or for the quality of the oil. The presence of transition metals will for instance initiate lipid peroxidation. The removal of unwanted compounds by refining processes is thus necessary. Traditional refining involves several processes, including degumming, neutralization, bleaching and deodorization (steam distillation) and some of these are performed at high temperatures. During the refining process, also some desirable compounds, such as natural antioxidants, are partly removed or destroyed, reducing the overall cardioprotective effect of the fish oil. A gentler refining process, based on short-path distillation, has been developed [92] and this process has been shown to effectively remove unwanted compounds, but at the same time protect the desirables [93].

Fish oils will, however, still be prone to oxidation and different protective measures are therefore applied in order to inhibit, reduce or delay this process. Addition of antioxidants is one of these measures and this is done routinely in most, if not all, *n*-3 supplements. This certainly reduces and delays the peroxidation process, however, it will not be completely inhibited [94,95]. Encapsulation of the oils is another measure that has been shown to increase stability. Here, the fish oils are embedded in materials such as polydextrins and different proteins [96,97]. However, some encapsulation materials have been shown to mask odour from volatile oxidation products and this may reduce the ability to detect early stages of oxidation in these products [98]. Using high quality oils, with low lipid oxidation levels, as source material is therefore very important.

A wide range of *n*-3 supplements are on the market today. Most of these are sold as either traditional liquid fish oils, encapsulated fish oils or concentrated supplements. In liquid fish oils and some encapsulated *n*-3 supplements, the lipids are present as TAG and the content of EPA + DHA is normally around 30%. In the refining process of the oils, TAG may be trans-esterified with ethanol making FA ethyl esters (FAEE). The mixture of FAEE may be fractionated and up-concentrated, increasing the concentration of the targeted FA (EPA + DHA) up to around 90% [99]. This increase in *n*-3 PUFA in the supplements will in itself lead to higher risk of oxidation. In addition, in a recent study, FAEE oils have been shown to oxidize more rapidly than TAG oils [100].

5. Epidemiological Studies on Consumption of Fish and *n*-3 Supplements

Numerous studies have been performed in order to investigate beneficial effects following consumption of these FA, as fish oils, capsules or seafood [101–108]. Fish and other seafood are the most important dietary sources of *n*-3 FA and has traditionally been target for most of these studies (Table 1). Most such studies conclude that increased seafood consumption is beneficial and may contribute to a prevention of development of CVD, particularly in the reduction of fatal CVD events [102,106,108]. Another aspect of high seafood consumption is that it displaces other nutrient sources, with possible adverse effects on cardiac health, in the diet. For instance, Bernstein *et al.* [109] showed that one serving a day of fish instead of red meat was associated with 24% lower risk of CHD.

Table 1. Recent systematic reviews and meta-analyses evaluating the association between the consumption of fish and risk of cardiovascular disease.

Reference	Year	Main Conclusions
Zheng <i>et al.</i> [108]	2012	Included 315,812 participants; average follow up period 15.9 years. Low (1 serving/week) or moderate fish consumption (2–4 servings/week) had a prevented of CHD mortality significantly. Difficult to conclude for high fish consumption (>5 servings/week) due to the limited amount of studies of high fish consumption.
Djousse <i>et al.</i> [110]	2012	Included 176,441 participants; average follow up period 13.3 years. The authors concluded that there was a linear and inverse association between consumption of fish, as well as marine ω -3 fatty acids, and the risk of heart failure (HF).
Chowdhury <i>et al.</i> [111]	2012	Included 794,000 participants from 26 prospective cohort-studies and 12 randomized controlled trials; Higher (especially fatty) fish consumption (1 fish meal per week vs. 2 or more weekly meals) is moderately but significantly associated with a reduced risk of cerebrovascular disease. No association with marine n -3 fatty acids and cerebrovascular disease. Effect attributed to other components in fish?
Li <i>et al.</i> [112]	2013	Included 170,231 participants; average follow up period 9.7 years. Reported a dose-dependent inverse relationship between fish consumption and incidence of HF. Weekly fish consumption (once or more) could reduce HF incidence.
Xun <i>et al.</i> [113]	2012	Included 402,127 individuals with an average 12.8 years of follow-up. Observed a modest inverse association between fish intake and ischemic stroke.
Yinko <i>et al.</i> [114]	2014	Included 408,305 participants from 11 prospective cohort and 8 case-control studies and reported an inverse, dose-related association between fish consumption and the risk of acute coronary syndrome.

In recent years substantial efforts have been made to evaluate the accumulated effects on cardiovascular disease of fish consumption and of marine n -3 supplementation using systematical reviews and meta-analyses. A summary of these efforts is presented in Table 2.

Table 2. Recent systematic reviews and meta-analyses evaluating the association between intake of marine n -3 supplements and risk of cardiovascular disease.

Reference	Year	Main Conclusions
Musa-Veloso <i>et al.</i> [115]	2011	Included 214,426 subjects and concluded that a daily supplementation with 250 mg EPA and DHA (or more) significantly reduced the risk of sudden cardiac death, whereas the risks of total fatal coronary events and non-fatal myocardial infarction was not significantly reduced.
Delgado-Lista <i>et al.</i> [116]	2012	Included 46,737 subjects with high cardiovascular risk. The marine fatty acids EPA and DHA decreased the risk of cardiovascular events, cardiac death and coronary events, especially in patients with high cardiovascular risk, but no reduced risk of all-cause mortality.
Rizos <i>et al.</i> [117]	2013	Included 68,680 patients. The authors concluded that there was no association between EPA and DHA supplementation and lowered risk of all-cause mortality, cardiac death, myocardial infarction and stroke.

Table 2. Cont.

Reference	Year	Main Conclusions
Casula <i>et al.</i> [118]	2013	Including 15,348 patients with a history of cardiovascular disease. Long-term supplementation of high dose <i>n</i> -3 PUFA may be beneficial for the onset of cardiac death, sudden death and myocardial infarction among patients with a history of CVD.
Zheng <i>et al.</i> [119]	2014	Included 24,788 patients with impaired glucose metabolism (IGM). EPA and DHA had no protective effect on cardiovascular mortality, major cardiovascular events or all-cause mortality in IGM patients.
Wen <i>et al.</i> [120]	2014	Included 32,656 patients with coronary heart disease (CHD). Supplement of <i>n</i> -3 PUFA in patients with CHD is associated with a reduction in death from cardiac causes, sudden cardiac death and death from all causes, whereas no association where found on major cardiovascular events.
Enns <i>et al.</i> [121]	2014	Included 396 patients with peripheral arterial disease. Not sufficient evidence to indicate a beneficial effect of <i>n</i> -3 PUFA supplementation on the incidence of cardiovascular events and related serous complications in adults with peripheral arterial disease.

Over the last few decades, *n*-3 FA has been generally considered effective means for lowering the risk of CHD and for secondary prevention of myocardial infarction (MI), effects that have been at least partially, ascribed to the previously discussed TAG-lowering effect. The DART randomized controlled trial (RCT) was the first RCT demonstrating that intervention with *n*-3 FA supplementation or increased intake of fish (advised to take fish oil capsules containing 500–900 mg *n*-3 FA/day or to eat 300 mg fatty fish per week), reduced the all-cause mortality in patients who had recovered from an earlier MI [122]. A few years later, the Italian GISSI-Prevenzione study consolidated these results in patients who recently had an MI. 11,324 patients were included and those receiving 885 mg EPA + DHA for up to 3.5 years had a 15% reduced combined risk for all-cause death and non-fatal cardiovascular events [123]. Also in the more recent, very large Japanese JELIS-study, a significant reduction was observed for the risk of coronary events in hypercholesterolemic patients receiving a daily dose of 1800 mg EPA in combination with 5 mg/day simvastatin compared to patients receiving statin only [107].

The apparent beneficial effect of *n*-3 PUFA supplementation on CVD was still evident a few years ago when the effects of marine *n*-3 supplementation was evaluated in patients with high cardiovascular risk [116], as EPA and DHA intake as food or supplement effectively prevented cardiovascular events, cardiac deaths and coronary events in patients with high cardiovascular risk [116]. Similar results were also indicated in the general population a few years ago, where those consuming more than 250 mg/day *n*-3 PUFA had a significant reduction in the risk of sudden cardiac death compared to subjects with a daily intake of less than 250 mg *n*-3 FA. On the contrary, Rizos *et al.* [117] presented a highly debated paper containing a cumulative meta-analysis of studies from 1995 to 2012 on *n*-3 supplementation and all-cause mortality. In this paper it was pointed out that the positive effect of *n*-3 supplementation has declined over the years, and that there no longer is a significant positive effect of *n*-3 supplementation on major cardiovascular outcomes. Their overall conclusion was that there are no longer grounds for recommending supplementary *n*-3 intake for protection against development of CVD. There are several

possible explanations to why the positive effects of *n*-3 supplementation seem to decline and some of these issues will be discussed in the following.

The most frequently used experimental method in these studies, RCT, is traditionally recognized as the “gold standard” of epidemiological studies. In such studies participants are randomly divided into groups, usually one control group and one or more treatment groups. However, when conducting RCT certain assumptions have to be made, for instance that the basal diets of the participants in each group are similar. Over time these basal conditions may change and a meta-analysis comparing RCT over a long time-span may be affected by these changes. For instance, the seafood consumption has more than doubled throughout the world during the last decades, from an average of 9.9 kg·capita⁻¹·year⁻¹ in the 1960s to 17.0 kg in the 2000s and 18.9 kg in 2010. Preliminary estimates for 2012 are pointing towards further growth to 19.2 kg·capita⁻¹·year⁻¹ [124]. This fact alone could have reduced the potential benefits of supplementary *n*-3 [54,125].

Over the years the use of medical treatment for control of risk factors, such as statins for hypercholesterolemia and ACE-inhibitors for hypertension, has increased, and such medications may also diminish the effects of supplementary *n*-3. An interesting example of this was observed in the recent Italian randomized controlled clinical trial examining the effect of *n*-3 FA (EPA and DHA ethyl esters in a daily dose of 850 mg) on non-fatal MI, non-fatal stroke and death [104]. The observed incidents of these primary outcomes was much lower than anticipated, and after a year the study group had to change the outcome to hospitalization or death from cardiovascular causes. There may be several explanations for this observation, and this is obviously associated with the improved pharmacological interventions (such as the statin effect) in combination with a substantial improved acute treatment of acute coronary events ultimately resulting in a lowered death rate from cardiovascular causes.

Choosing the right control, or placebo, group is one of the most challenging factors of performing such studies. In several of the studies included in Rizos *et al.* [117], olive oil has been chosen as placebo [104]. Olive oil is rich in the monounsaturated FA oleic acid (C18:1, *n*-9), along with a range of polyphenolic compounds. Olive oil is therefore recognized as an effective anti-oxidant and may be protective against development of CVD in itself [126] and is hence, not the best choice as placebo when examining cardioprotective effects.

Most studies involving *n*-3 supplements are currently not reporting the degree of oxidation of the supplements used in the trials [127]. As discussed in Section 4, the quality and stability of *n*-3 supplements may be important factors when evaluating their cardioprotective effects. Recently a range of studies on the oxidative status on over-the-counter *n*-3 supplements have been published showing that the contents of *n*-3 are lower than declared and that the limits for oxidation products are exceeded in a large number of products [128–131]. Inferior quality of the supplements may therefore be another explanatory factor to the lack of evidence for positive effects of *n*-3 supplementation. Several recent meta-analyses and systematic reviews have presented results using different patient populations without finding any significant protective effects from EPA and DHA on major cardiovascular events, MI, cardiac death or all-cause mortality [119,121]. However, in patients with a history of CVD or CHD, *n*-3 PUFA supplementation may still lower the risk for MI and sudden cardiac death [118,120].

6. Dietary Recommendations

In a study by Mozaffarian and Rimm [125], the correlation between *n*-3 intake and protective effect was reviewed. Their conclusion was that a modest intake of fish, *i.e.*, 1–2 servings per week corresponding to approximately 250 mg EPA + DHA per day, reduced the risk of coronary death by 36% and risk of total death by 17%. A higher seafood intake did not reduce the risk further.

Another focus regarding protective effects of *n*-3 FA is the relationship or ratio between *n*-6 and *n*-3 FA in the diet. In historic times, the ratio between these two FA was around 1:1, while in present times this ratio has shifted markedly in favor of *n*-6. Current ratios are 4:1 in Japan, 15–17:1 in the Western societies (Europe and USA) and 38–50:1 in urban areas of India [132]. The imbalance between these FA as seen worldwide today may promote the development of several lifestyle-related diseases such as CVD [132–135]. Increasing seafood intake is considered to be a good strategy in order to achieve an improved ratio between these FA.

In 2010 and 2014, two large analyses of the risks and benefits of consuming seafood were performed, one by FAO/WHO [136] and one by the Norwegian Scientific Committee for Food Safety [91]. The conclusion in both of these studies was that adults consuming less than one serving of seafood per week may miss the beneficial effects of seafood on CVD.

Based on the conclusions from the epidemiological studies on seafood consumption and the risk and benefit analyses, several organizations and authorities have published dietary recommendations regarding intake of marine *n*-3 FA and/or fish (Table 3).

Table 3. Overview of dietary recommendations regarding fish or marine *n*-3 fatty acids.

Authority/Organization	Country/Region	Year	Recommendation	Reference
The American Heart Association	USA	2015	A variety of (preferably fatty) fish at least twice a week	[137]
The Norwegian Directorate of Health/VKM	Norway	2014	Fish as dinner at least 2–3 times per week	[91]
Food and Agricultural Organization of the United Nations (FAO)/World Health Organization (WHO)	World	2011	At least 1–2 100 g servings of fatty fish per week	[136]
European Food Safety Association (EFSA)	Europe	2010	250 mg EPA + DHA daily	[138]
Scientific Advisory Committee for Nutrition (SACN)	UK	2004	450 mg EPA + DHA daily	[139]
International Society for the Study of Fatty Acids and Lipids (ISSFAL)	UK/Europe	2004	500 mg EPA + DHA daily	[140]

Table 3 shows the recommendations given to general, healthy populations. Several authorities/organizations specify that people with documented health issues, such as CVD, may need more *n*-3 than the general population. For instance, AHA recommends two weekly seafood meals for people without documented CHD, 1 g EPA + DHA for people with documented CHD and 2–4 g EPA + DHA for people in need for lowering TAG [137].

The more recent official advice focus on intake of fish and seafood as sources of *n*-3 FA. This is partly because uptake and incorporation of *n*-3 from seafood has been shown to be superior to that from *n*-3 supplements [141–143]. Another reason for recommending seafood intake over *n*-3 supplements, as

stated in Section 4, is that seafood, in addition to its *n*-3 content, is also a rich source of other nutrients that may provide synergy effects towards health benefits.

7. Conclusions

Fish and seafood are the optimal sources of *n*-3 FA and there is a broad consensus that the general consumption of seafood should be approximately 2–3 servings per week in order to obtain the maximal protective effect towards development of CVD. There are conflicting findings between no effects vs. beneficial effects in recent RCT and meta-analyses for *n*-3 supplements. The scientific evidence for benefits from *n*-3 supplementation has diminished over time, probably due to a general increase in seafood consumption and better pharmacological intervention and acute treatment of patients with CVD.

Acknowledgments

This work was supported by the Publication Fund of UIT The Arctic University of Norway.

Author Contributions

Hanne K. Maehre, Ida-Johanne Jensen, Edel O. Elvevoll and Karl-Erik Eilertsen contributed equally to the idea of this review; Hanne K. Maehre wrote the manuscript; Ida-Johanne Jensen, Edel O. Elvevoll and Karl-Erik Eilertsen contributed with critical comments and suggestions during the writing process. All authors read and approved the final manuscript.

Conflicts of Interest

The authors declare no conflict of interest

References

- 1 Bang, H.O.; Dyerberg, J.; Hjorne, N. Composition of food consumed by greenland eskimos. *Acta Med. Scand.* **1976**, *200*, 69–73.
- 2 Bang, H.O.; Dyerberg, J.; Nielsen, A.B. Plasma lipid and lipoprotein pattern in Greenlandic West-coast Eskimos. *Lancet* **1971**, *1*, 1143–1145.
- 3 Dyerberg, J.; Bang, H.O. Hemostatic function and platelet poly-unsaturated fatty acids in eskimos. *Lancet* **1979**, *2*, 433–435.
- 4 Dyerberg, J.; Bang, H.O.; Stoffersen, E.; Moncada, S.; Vane, J.R. Eicosapentaenoic acid and prevention of thrombosis and atherosclerosis? *Lancet* **1978**, *2*, 117–119.
- 5 Bjerregaard, P.; Young, T.K.; Hegele, R.A. Low incidence of cardiovascular disease among the inuit-what is the evidence? *Atherosclerosis* **2003**, *166*, 351–357.
- 6 Fodor, J.G.; Helis, E.; Yazdekhasi, N.; Vohnout, B. “Fishing” for the origins of the “Eskimos and heart disease” Story: Facts or wishful thinking? *Can. J. Cardiol.* **2014**, *30*, 864–868.
- 7 Cardiovascular diseases (CVDs): Fact sheet no. 317; World Health Organization: Geneva, Switzerland. Available online: <http://www.who.int/mediacentre/factsheets/fs317/en/index.html> (accessed on 15 August 2015).

- 8 Mendis, S.; Puska, P.; Norrving, B. *Global Atlas on Cardiovascular Disease Prevention and Control*; Mendis, S., Puska, P., Eds.; World Health Organization: Geneva, Switzerland, 2011.
- 9 Yusuf, H.R.; Giles, W.H.; Croft, J.B.; Anda, R.F.; Casper, M.L. Impact of multiple risk factor profiles on determining cardiovascular disease risk. *Prev. Med.* **1998**, *27*, 1–9.
- 10 Galley, H.F.; Webster, N.R. Physiology of the endothelium. *Br. J. Anaesth.* **2004**, *93*, 105–113.
- 11 Bonetti, P.O.; Lerman, L.O.; Lerman, A. Endothelial dysfunction: A marker of atherosclerotic risk. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, 168–175.
- 12 McLaren, J.E.; Michael, D.R.; Ashlin, T.G.; Ramji, D.P. Cytokines, macrophage lipid metabolism and foam cells: Implications for cardiovascular disease therapy. *Prog. Lipid Res.* **2011**, *50*, 331–347.
- 13 Szmítko, P.E.; Wang, C.H.; Weisel, R.D.; de Almeida, J.R.; Anderson, T.J.; Verma, S. New markers of inflammation and endothelial cell activation: Part I. *Circulation* **2003**, *108*, 1917–1923.
- 14 Smith, W.L.; Murphy, R.C. The eicosanoids: Cyclooxygenase, lipoxygenase, and epoxygenase pathways. In *Biochemistry of Lipids, Lipoproteins and Membranes*, 5th ed.; Vance, D.E., Vance, J.E., Eds.; Elsevier B.V.: Amsterdam, The Netherlands, 2008.
- 15 Bannenberg, G.; Serhan, C.N. Specialized pro-resolving lipid mediators in the inflammatory response: An update. *Biochim. Biophys. Acta* **2010**, *1801*, 1260–1273.
- 16 Larsson, S.C.; Kumlin, M.; Ingelman-Sundberg, M.; Wolk, A. Dietary long-chain *n*-3 fatty acids for the prevention of cancer: A review of potential mechanisms. *Am. J. Clin. Nutr.* **2004**, *79*, 935–945.
- 17 Calder, P.C. Marine ω -3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance. *Biochim. Biophys. Acta* **2015**, *1851*, 469–484.
- 18 Serhan, C.N. Resolution phase of inflammation: Novel endogenous anti-inflammatory and proresolving lipid mediators and pathways. *Annu. Rev. Immunol.* **2007**, *25*, 101–137.
- 19 Stables, M.J.; Gilroy, D.W. Old and new generation lipid mediators in acute inflammation and resolution. *Prog. Lipid Res.* **2011**, *50*, 35–51.
- 20 Morita, I. Distinct functions of COX-1 and COX-2. *Prostaglandins Other Lipid Mediat.* **2002**, *68–69*, 165–175.
- 21 Smith, W.L.; Langenbach, R. Why there are two cyclooxygenase isozymes. *J. Clin. Investig.* **2001**, *107*, 1491–1495.
- 22 Simmons, D.L. Variants of cyclooxygenase-1 and their roles in medicine. *Thromb. Res.* **2003**, *110*, 265–268.
- 23 Barbosa-Sicard, E.; Markovic, M.; Honeck, H.; Christ, B.; Muller, D.N.; Schunck, W.H. Eicosapentaenoic acid metabolism by cytochrome P450 enzymes of the CYP2C subfamily. *Biochem. Biophys. Res. Commun.* **2005**, *329*, 1275–1281.
- 24 Arnold, C.; Konkel, A.; Fischer, R.; Schunck, W.H. Cytochrome P450-dependent metabolism of ω -6 and ω -3 long-chain polyunsaturated fatty acids. *Pharmacol. Rep.* **2010**, *62*, 536–547.
- 25 Pratico, D.; Dogne, J.M. Vascular biology of eicosanoids and atherogenesis. *Expert Rev. Cardiovasc. Ther.* **2009**, *7*, 1079–1089.
- 26 Simopoulos, A.P. The importance of the ω -6/ ω -3 fatty acid ratio in cardiovascular disease and other chronic diseases. *Exp. Biol. Med.* **2008**, *233*, 674–688.

- 27 Friedrich, E.B.; Tager, A.M.; Liu, E.; Pettersson, A.; Owman, C.; Munn, L.; Luster, A.D.; Gerszten, R.E. Mechanisms of leukotriene B-4-triggered monocyte adhesion. *Arterioscler. Thromb. Vasc. Biol.* **2003**, *23*, 1761–1767.
- 28 Huang, L.; Zhao, A.; Wong, F.; Ayala, J.M.; Struthers, M.; Ujjainwalla, F.; Wright, S.D.; Springer, M.S.; Evans, J.; Cui, J.S. Leukotriene B-4 strongly increases monocyte chemoattractant protein-1 in human monocytes. *Arterioscler. Thromb. Vasc. Biol.* **2004**, *24*, 1783–1788.
- 29 Moreno, J.J. Differential effects of arachidonic and eicosapentaenoic acid-derived eicosanoids on polymorphonuclear transmigration across endothelial cell cultures. *J. Pharmacol. Exp. Ther.* **2009**, *331*, 1111–1117.
- 30 Yates, C.M.; Calder, P.C.; Rainer, G.E. Pharmacology and therapeutics of ω -3 polyunsaturated fatty acids in chronic inflammatory disease. *Pharmacol. Ther.* **2014**, *141*, 272–282.
- 31 Oliver, E.; McGillicuddy, F.C.; Harford, K.A.; Reynolds, C.M.; Phillips, C.M.; Ferguson, J.F.; Roche, H.M. Docosahexaenoic acid attenuates macrophage-induced inflammation and improves insulin sensitivity in adipocytes-specific differential effects between LC *n*-3 PUFA. *J. Nutr. Biochem.* **2012**, *23*, 1192–1200.
- 32 Yagi, S.; Aihara, K.; Fukuda, D.; Takashima, A.; Hara, T.; Hotchi, J.; Ise, T.; Yamaguchi, K.; Tobiume, T.; Iwase, T.; *et al.* Effects of docosahexaenoic acid on the endothelial function in patients with coronary artery disease. *J. Atheroscler. Thromb.* **2015**, *22*, 447–454.
- 33 Qu, Q.; Xuan, W.; Fan, G.H. Roles of resolvins in the resolution of acute inflammation. *Cell Biol. Int.* **2015**, *39*, 3–22.
- 34 Ayala, A.; Munoz, M.F.; Arguelles, S. Lipid peroxidation: Production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. *Oxid. Med. Cell. Longev.* **2014**, *2014*, doi:10.1155/2014/360438.
- 35 Esterbauer, H.; Schaur, R.J.; Zollner, H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic. Biol. Med.* **1991**, *11*, 81–128.
- 36 Riahi, Y.; Cohen, G.; Shamni, O.; Sasson, S. Signaling and cytotoxic functions of 4-hydroxyalkenals. *Am. J. Physiol. Endocrinol. Metab.* **2010**, *299*, E879–E886.
- 37 Van den Berg, R.; Haenen, G.R.M.M.; van den Berg, H.; Bast, A. Transcription factor NF- κ B as a potential biomarker for oxidative stress. *Br. J. Nutr.* **2001**, *86*, S121–S127.
- 38 Esterbauer, H. Cytotoxicity and genotoxicity of lipid-oxidation products. *Am. J. Clin. Nutr.* **1993**, *57*, 779–786.
- 39 Long, E.K.; Murphy, T.C.; Leiphon, L.J.; Watt, J.; Morrow, J.D.; Milne, G.L.; Howard, J.R.; Picklo, M.J., Sr. *Trans*-4-hydroxy-2-hexenal is a neurotoxic product of docosahexaenoic (22:6; *n*-3) acid oxidation. *J. Neurochem.* **2008**, *105*, 714–724.
- 40 Je, J.H.; Lee, J.Y.; Jung, K.J.; Sung, B.; Go, E.K.; Yu, B.P.; Chung, H.Y. NF- κ B activation mechanism of 4-hydroxyhexenal via NIK/IKK and p38 MAPK pathway. *FEBS Lett.* **2004**, *566*, 183–189.
- 41 Ishikado, A.; Nishio, Y.; Morino, K.; Ugi, S.; Kondo, H.; Makino, T.; Kashiwagi, A.; Maegawa, H. Low concentration of 4-hydroxy hexenal increases heme oxygenase-1 expression through activation of Nrf2 and antioxidative activity in vascular endothelial cells. *Biochem. Biophys. Res. Commun.* **2010**, *402*, 99–104.

- 42 Uchida, K. Role of reactive aldehyde in cardiovascular diseases. *Free Radic. Biol. Med.* **2000**, *28*, 1685–1696.
- 43 Yamada, S.; Funada, T.; Shibata, N.; Kobayashi, M.; Kawai, Y.; Tatsuda, E.; Furuhashi, A.; Uchida, K. Protein-bound 4-hydroxy-2-hexenal as a marker of oxidized *n*-3 polyunsaturated fatty acids. *J. Lipid Res.* **2004**, *45*, 626–634.
- 44 Flock, M.R.; Skulas-Ray, A.C.; Harris, W.S.; Etherton, T.D.; Fleming, J.A.; Kris-Etherton, P.M. Determinants of erythrocyte ω -3 fatty acid content in response to fish oil supplementation: A dose-response randomized controlled trial. *J. Am. Heart Assoc.* **2013**, *2*, e000513.
- 45 Keenan, A.H.; Pedersen, T.L.; Fillaus, K.; Larson, M.K.; Shearer, G.C.; Newman, J.W. Basal ω -3 fatty acid status affects fatty acid and oxylipin responses to high-dose *n*-3-HUFA in healthy volunteers. *J. Lipid Res.* **2012**, *53*, 1662–1669.
- 46 Harris, W.S.; von Schacky, C. The ω -3 index: A new risk factor for death from coronary heart disease? *Prev. Med.* **2004**, *39*, 212–220.
- 47 Von Schacky, C. ω -3 index and cardiovascular health. *Nutrients* **2014**, *6*, 799–814.
- 48 Harris, W.S. The ω -3 index: From biomarker to risk marker to risk factor. *Curr. Atheroscler. Rep.* **2009**, *11*, 411–417.
- 49 Maaloe, T.; Schmidt, E.B.; Svensson, M.; Aardestrup, I.V.; Christensen, J.H. The effect of *n*-3 polyunsaturated fatty acids on leukotriene B₄ and leukotriene B₅ production from stimulated neutrophil granulocytes in patients with chronic kidney disease. *Prostaglandins Leukot. Essent. Fatty Acids* **2011**, *85*, 37–41.
- 50 Raatz, S.K.; Young, L.R.; Picklo, M.J., Sr.; Sauter, E.R.; Qin, W.; Kurzer, M.S. Total dietary fat and fatty acid content modifies plasma phospholipid fatty acids, desaturase activity indices, and urinary prostaglandin E in women. *Nutr. Res.* **2012**, *32*, 1–7.
- 51 Stanke-Labesque, F.; Moliere, P.; Bessard, J.; Laville, M.; Vericel, E.; Lagarde, M. Effect of dietary supplementation with increasing doses of docosahexaenoic acid on neutrophil lipid composition and leukotriene production in human healthy volunteers. *Br. J. Nutr.* **2008**, *100*, 829–833.
- 52 Fischer, R.; Konkel, A.; Mehling, H.; Blosssey, K.; Gapelyuk, A.; Wessel, N.; von Schacky, C.; Dechend, R.; Muller, D.N.; Rothe, M.; *et al.* Dietary ω -3 fatty acids modulate the eicosanoid profile in man primarily via the CYP-epoxygenase pathway. *J. Lipid Res.* **2014**, *55*, 1150–1164.
- 53 Arnold, C.; Markovic, M.; Blosssey, K.; Wallukat, G.; Fischer, R.; Dechend, R.; Konkel, A.; von Schacky, C.; Luft, F.C.; Muller, D.N.; *et al.* Arachidonic acid-metabolizing cytochrome P450 enzymes are targets of ω 3 fatty acids. *J. Biol. Chem.* **2010**, *285*, 32720–32733.
- 54 Watanabe, N.; Watanabe, Y.; Kumagai, M.; Fujimoto, K. Administration of dietary fish oil capsules in healthy middle-aged Japanese men with a high level of fish consumption. *Int. J. Food Sci. Nutr.* **2009**, *60* (Suppl. 5), 136–142.
- 55 Ciubotaru, I.; Lee, Y.S.; Wander, R.C. Dietary fish oil decreases C-reactive protein, interleukin-6, and triacylglycerol to HDL-cholesterol ratio in postmenopausal women on HRT. *J. Nutr. Biochem.* **2003**, *14*, 513–521.
- 56 Kiecolt-Glaser, J.K.; Belury, M.A.; Andridge, R.; Malarkey, W.B.; Glaser, R. ω -3 supplementation lowers inflammation and anxiety in medical students: A randomized controlled trial. *Brain Behav. Immun.* **2011**, *25*, 1725–1734.

- 57 Kiecolt-Glaser, J.K.; Belury, M.A.; Andridge, R.; Malarkey, W.B.; Hwang, B.S.; Glaser, R. ω -3 supplementation lowers inflammation in healthy middle-aged and older adults: A randomized controlled trial. *Brain Behav. Immun.* **2012**, *26*, 988–995.
- 58 Moertl, D.; Hammer, A.; Steiner, S.; Hutuleac, R.; Vonbank, K.; Berger, R. Dose-dependent effects of ω -3-polyunsaturated fatty acids on systolic left ventricular function, endothelial function, and markers of inflammation in chronic heart failure of nonischemic origin: A double-blind, placebo-controlled, 3-arm study. *Am. Heart J.* **2011**, *161*, doi:10.1016/j.ahj.2011.02.011.
- 59 Nodari, S.; Triggiani, M.; Campia, U.; Manerba, A.; Milesi, G.; Cesana, B.M.; Gheorghide, M.; Cas, L.D. Effects of *n*-3 polyunsaturated fatty acids on left ventricular function and functional capacity in patients with dilated cardiomyopathy. *J. Am. Coll. Cardiol.* **2011**, *57*, 870–879.
- 60 Zhao, Y.T.; Shao, L.; Teng, L.L.; Hu, B.; Luo, Y.; Yu, X.; Zhang, D.F.; Zhang, H. Effects of *n*-3 polyunsaturated fatty acid therapy on plasma inflammatory markers and N-terminal pro-brain natriuretic peptide in elderly patients with chronic heart failure. *J. Int. Med. Res.* **2009**, *37*, 1831–1841.
- 61 Elvevoll, E.O.; Eilertsen, K.E.; Brox, J.; Dragnes, B.T.; Falkenberg, P.; Olsen, J.O.; Kirkhus, B.; Lamglait, A.; Osterud, B. Seafood diets: Hypolipidemic and antiatherogenic effects of taurine and *n*-3 fatty acids. *Atherosclerosis* **2008**, *200*, 396–402.
- 62 Juarez-Lopez, C.; Klunder-Klunder, M.; Madrigal-Azcarate, A.; Flores-Huerta, S. ω -3 polyunsaturated fatty acids reduce insulin resistance and triglycerides in obese children and adolescents. *Pediatr. Diabetes* **2013**, *14*, doi:10.1111/pedi.12024.
- 63 Zhang, J.; Wang, C.; Li, L.; Man, Q.; Meng, L.; Song, P.; Froyland, L.; Du, Z.Y. Dietary inclusion of salmon, herring and pompano as oily fish reduces CVD risk markers in dyslipidaemic middle-aged and elderly Chinese women. *Br. J. Nutr.* **2012**, *108*, 1455–1465.
- 64 Zhang, J.; Wang, C.; Li, L.; Man, Q.; Song, P.; Meng, L.; Du, Z.Y.; Froyland, L. Inclusion of atlantic salmon in the Chinese diet reduces cardiovascular disease risk markers in dyslipidemic adult men. *Nutr. Res.* **2010**, *30*, 447–454.
- 65 Baldassarre, D.; Amato, M.; Eligini, S.; Barbieri, S.S.; Mussoni, L.; Frigerio, B.; Kozakova, M.; Tremoli, E.; Sirtori, C.R.; Colli, S. Effect of *n*-3 fatty acids on carotid atherosclerosis and haemostasis in patients with combined hyperlipoproteinemia: A double-blind pilot study in primary prevention. *Ann. Med.* **2006**, *38*, 367–375.
- 66 Ramprasath, V.R.; Eyal, I.; Zchut, S.; Jones, P.J. Enhanced increase of ω -3 index in healthy individuals with response to 4-week *n*-3 fatty acid supplementation from krill oil vs. fish oil. *Lipids Health Dis.* **2013**, *12*, doi:10.1186/1476-511X-12-178.
- 67 Ishida, T.; Ohta, M.; Nakakuki, M.; Kami, H.; Uchiyama, R.; Kawano, H.; Notsu, T.; Imada, K.; Shimano, H. Distinct regulation of plasma LDL cholesterol by eicosapentaenoic acid and docosahexaenoic acid in high fat diet-fed hamsters: Participation of cholesterol ester transfer protein and LDL receptor. *Prostaglandins Leukot. Essent. Fatty Acids* **2013**, *88*, 281–288.
- 68 Root, M.; Collier, S.R.; Zwetsloot, K.A.; West, K.L.; McGinn, M.C. A randomized trial of fish oil ω -3 fatty acids on arterial health, inflammation, and metabolic syndrome in a young healthy population. *Nutr. J.* **2013**, *12*, doi:10.1186/1475-2891-12-40.

- 69 Garaiova, I.; Muchova, J.; Nagyova, Z.; Mislanova, C.; Oravec, S.; Dukat, A.; Wang, D.; Plummer, S.F.; Durackova, Z. Effect of a plant sterol, fish oil and B vitamin combination on cardiovascular risk factors in hypercholesterolemic children and adolescents: A pilot study. *Nutr. J.* **2013**, *12*, doi:10.1186/1475-2891-12-7.
- 70 Paoli, A.; Moro, T.; Bosco, G.; Bianco, A.; Grimaldi, K.A.; Camporesi, E.; Mangar, D. Effects of *n*-3 polyunsaturated fatty acids (ω -3) supplementation on some cardiovascular risk factors with a ketogenic Mediterranean diet. *Mar. Drugs* **2015**, *13*, 996–1009.
- 71 Iketani, T.; Takazawa, K.; Yamashina, A. Effect of eicosapentaenoic acid on central systolic blood pressure. *Prostaglandins Leukot. Essent. Fatty Acids* **2013**, *88*, 191–195.
- 72 Ramel, A.; Martinez, J.A.; Kiely, M.; Bandarra, N.M.; Thorsdottir, I. Moderate consumption of fatty fish reduces diastolic blood pressure in overweight and obese European young adults during energy restriction. *Nutrition* **2010**, *26*, 168–174.
- 73 Rasmussen, B.M.; Vessby, B.; Uusitupa, M.; Berglund, L.; Pedersen, E.; Riccardi, G.; Rivellese, A.A.; Tapsell, L.; Hermansen, K.; Group, K.S. Effects of dietary saturated, monounsaturated, and *n*-3 fatty acids on blood pressure in healthy subjects. *Am. J. Clin. Nutr.* **2006**, *83*, 221–226.
- 74 Campbell, F.; Dickinson, H.O.; Critchley, J.A.; Ford, G.A.; Bradburn, M. A systematic review of fish-oil supplements for the prevention and treatment of hypertension. *Eur. J. Prev. Cardiol.* **2013**, *20*, 107–120.
- 75 Miller, P.E.; van Elswyk, M.; Alexander, D.D. Long-chain ω -3 fatty acids eicosapentaenoic acid and docosahexaenoic acid and blood pressure: A meta-analysis of randomized controlled trials. *Am. J. Hypertens.* **2014**, *27*, 885–896.
- 76 Awada, M.; Soulage, C.O.; Meynier, A.; Debard, C.; Plaisancie, P.; Benoit, B.; Picard, G.; Loizon, E.; Chauvin, M.A.; Estienne, M.; *et al.* Dietary oxidized *n*-3 PUFA induce oxidative stress and inflammation: Role of intestinal absorption of 4-HHE and reactivity in intestinal cells. *J. Lipid Res.* **2012**, *53*, 2069–2080.
- 77 Lu, T.; Harper, A.F.; Zhao, J.; Dalloul, R.A. Effects of a dietary antioxidant blend and vitamin E on growth performance, oxidative status, and meat quality in broiler chickens fed a diet high in oxidants. *Poult. Sci.* **2014**, *93*, 1649–1657.
- 78 Lu, T.; Harper, A.F.; Zhao, J.; Estienne, M.J.; Dalloul, R.A. Supplementing antioxidants to pigs fed diets high in oxidants: I. Effects on growth performance, liver function, and oxidative status. *J. Anim. Sci.* **2014**, *92*, 5455–5463.
- 79 Yang, S.P.; Liu, H.L.; Wang, C.G.; Yang, P.; Sun, C.B.; Chan, S.M. Effect of oxidized fish oil on growth performance and oxidative stress of *Litopenaeus vannamei*. *Aquac. Nutr.* **2015**, *21*, 121–127.
- 80 Eilertsen, K.E.; Maehre, H.K.; Cludts, K.; Olsen, J.O.; Hoylaerts, M.F. Dietary enrichment of apolipoprotein E-deficient mice with extra virgin olive oil in combination with seal oil inhibits atherogenesis. *Lipids Health Dis.* **2011**, *10*, 1–8.
- 81 Eilertsen, K.E.; Maehre, H.K.; Jensen, I.J.; Devold, H.; Olsen, J.O.; Lie, R.K.; Brox, J.; Berg, V.; Elvevoll, E.O.; Osterud, B. A wax ester and astaxanthin-rich extract from the marine copepod *Calanus finmarchicus* attenuates atherogenesis in female apolipoprotein E-deficient mice. *J. Nutr.* **2012**, *142*, 508–512.

- 82 Ottestad, I.; Vogt, G.; Retterstol, K.; Myhrstad, M.C.; Haugen, J.E.; Nilsson, A.; Ravn-Haren, G.; Nordvi, B.; Bronner, K.W.; Andersen, L.F.; *et al.* Oxidised fish oil does not influence established markers of oxidative stress in healthy human subjects: A randomised controlled trial. *Br. J. Nutr.* **2012**, *108*, 315–326.
- 83 Garcia-Hernandez, V.M.; Gallar, M.; Sanchez-Soriano, J.; Micol, V.; Roche, E.; Garcia-Garcia, E. Effect of ω -3 dietary supplements with different oxidation levels in the lipidic profile of women: A randomized controlled trial. *Int. J. Food Sci. Nutr.* **2013**, *64*, 993–1000.
- 84 Asghari, L.; Zeynali, F.; Sahari, M.A. Effects of boiling, deep-frying, and microwave treatment on the proximate composition of rainbow trout fillets: Changes in fatty acids, total protein, and minerals. *J. Appl. Ichthyol.* **2013**, *29*, 847–853.
- 85 Mierke-Klemeyer, S.; Larsen, R.; Oehlenschlager, J.; Maehre, H.; Elvevoll, E.O.; Bandarra, N.M.; Parreira, R.; Andrade, A.M.; Nunes, M.L.; Schram, E.; *et al.* Retention of health-related beneficial components during household preparation of selenium-enriched African catfish (*Clarias gariepinus*) fillets. *Eur. Food Res. Technol.* **2008**, *227*, 827–833.
- 86 Perez-Palacios, T.; Petisca, C.; Casal, S.; Ferreira, I.M.P.L.V.O. Changes in chemical composition of frozen coated fish products during deep-frying. *Int. J. Food Sci. Nutr.* **2014**, *65*, 212–218.
- 87 Pirini, M.; Testi, S.; Ventrella, V.; Pagliarani, A.; Badiani, A. Blue-back fish: Fatty acid profile in selected seasons and retention upon baking. *Food Chem.* **2010**, *123*, 306–314.
- 88 Weichselbaum, E.; Coe, S.; Buttriss, J.; Stanner, S. Fish in the diet: A review. *Nutr. Bull.* **2013**, *38*, 128–177.
- 89 Larsen, R.; Eilertsen, K.E.; Maehre, H.K.; Jensen, I.J.; Elvevoll, E.O. Taurine content in marine foods—Beneficial health effects. In *Bioactive Compounds from Marine Foods: Plant and Animal Sources*; Hernández-Ledesma, B., Herrero, M., Eds.; Wiley-Blackwell Oxford: Chichester, UK, 2014; pp. 249–268.
- 90 Jensen, I.J.; Abrahamsen, H.; Maehre, H.K.; Elvevoll, E.O. Changes in antioxidative capacity of saithe (*Pollachius virens*) and Shrimp (*Pandalus borealis*) during *in vitro* digestion. *J. Agric. Food Chem.* **2009**, *57*, 10928–10932.
- 91 Skåre, J.U.; Brantsæter, A.L.; Frøyland, L.; Hemre, G.L.; Knutsen, H.K.; Lillegaard, I.T.L.; Torstensen, B. Benefit-risk assessment of fish and fish products in the Norwegian diet—An update. *Eur. J. Nutr. Food Saf.* **2015**, *5*, 260–266.
- 92 Oterhals, A.; Kvamme, B.; Berntssen, M.H. Modeling of a short-path distillation process to remove persistent organic pollutants in fish oil based on process parameters and quantitative structure properties relationships. *Chemosphere* **2010**, *80*, 83–92.
- 93 Oterhals, A.; Berntssen, M.H. Effects of refining and removal of persistent organic pollutants by short-path distillation on nutritional quality and oxidative stability of fish oil. *J. Agric. Food Chem.* **2010**, *58*, 12250–12259.
- 94 Wang, H.; Liu, F.; Yang, L.; Zu, Y.G.; Wang, H.; Qu, S.Z.; Zhang, Y. Oxidative stability of fish oil supplemented with carnosic acid compared with synthetic antioxidants during long-term storage. *Food Chem.* **2011**, *128*, 93–99.
- 95 Zuta, P.C.; Simpson, B.K.; Zhao, X.; Leclerc, L. The effect of α -tocopherol on the oxidation of mackerel oil. *Food Chem.* **2007**, *100*, 800–807.

- 96 Ahn, J.H.; Kim, Y.P.; Seo, E.M.; Choi, Y.K.; Kim, H.S. Antioxidant effect of natural plant extracts on the microencapsulated high oleic sunflower oil. *J. Food Eng.* **2008**, *84*, 327–334.
- 97 Kagami, Y.; Sugimura, S.; Fujishima, N.; Matsuda, K.; Kometani, T.; Matsumura, Y. Oxidative stability, structure, and physical characteristics of microcapsules formed by spray drying of fish oil with protein and dextrin wall materials. *J. Food Sci.* **2003**, *68*, 2248–2255.
- 98 Bottcher, S.; Steinhäuser, U.; Drusch, S. Off-flavour masking of secondary lipid oxidation products by pea dextrin. *Food Chem.* **2015**, *169*, 492–498.
- 99 Kralovec, J.A.; Zhang, S.C.; Zhang, W.; Barrow, C.J. A review of the progress in enzymatic concentration and microencapsulation of ω -3 rich oil from fish and microbial sources. *Food Chem.* **2012**, *131*, 639–644.
- 100 Ritter, J.C.S.; Budge, S.M.; Jovica, F.; Reid, A.J.M. Oxidation rates of triacylglycerol and ethyl ester fish oils. *J. Am. Oil Chem. Soc.* **2015**, *92*, 561–569.
- 101 Bosch, J.; Gerstein, H.C.; Dagenais, G.R.; Diaz, R.; Dyal, L.; Jung, H.; Maggiono, A.P.; Probstfield, J.; Ramachandran, A.; Riddle, M.C.; *et al.* *n*-3 fatty acids and cardiovascular outcomes in patients with dysglycemia. *N. Engl. J. Med.* **2012**, *367*, 309–318.
- 102 Chrysohoou, C.; Panagiotakos, D.B.; Pitsavos, C.; Skoumas, J.; Krinos, X.; Chloptsios, Y.; Nikolaou, V.; Stefanadis, C. Long-term fish consumption is associated with protection against arrhythmia in healthy persons in a Mediterranean region—The ATTICA study. *Am. J. Clin. Nutr.* **2007**, *85*, 1385–1391.
- 103 Mozaffarian, D.; Marchioli, R.; Macchia, A.; Sillelta, M.G.; Ferrazzi, P.; Gardner, T.J.; Latini, R.; Libby, P.; Lombardi, F.; O’Gara, P.T.; *et al.* Fish oil and postoperative atrial fibrillation the ω -3 fatty acids for prevention of post-operative atrial fibrillation (OPERA) randomized trial. *JAMA* **2012**, *308*, 2001–2011.
- 104 Roncaglioni, M.C.; Tombesi, M.; Avanzini, F.; Barlera, S.; Caimi, V.; Longoni, P.; Marzona, I.; Milani, V.; Sillelta, M.G.; Tognoni, G.; *et al.* *n*-3 fatty acids in patients with multiple cardiovascular risk factors. *N. Engl. J. Med.* **2013**, *368*, 1800–1808.
- 105 Tavazzi, L.; Maggioni, A.P.; Marchioli, R.; Barlera, S.; Franzosi, M.G.; Latini, R.; Lucci, D.; Nicolosi, G.L.; Porcu, M.; Tognoni, G.; *et al.* Effect of *n*-3 polyunsaturated fatty acids in patients with chronic heart failure (the GISSI-HF trial): A randomised, double-blind, placebo-controlled trial. *Lancet* **2008**, *372*, 1223–1230.
- 106 Virtanen, J.K.; Mozaffarian, D.; Chiuve, S.E.; Rimm, E.B. Fish consumption and risk of major chronic disease in men. *Am. J. Clin. Nutr.* **2008**, *88*, 1618–1625.
- 107 Yokoyama, M.; Origasa, H.; Matsuzaki, M.; Matsuzawa, Y.; Saito, Y.; Ishikawa, Y.; Oikawa, S.; Sasaki, J.; Hishida, H.; Itakura, H.; *et al.* Effects of eicosapentaenoic acid on major coronary events in hypercholesterolaemic patients (JELIS): A randomised openlabel, blinded endpoint analysis. *Lancet* **2007**, *369*, 1090–1098.
- 108 Zheng, J.; Huang, T.; Yu, Y.; Hu, X.; Yang, B.; Li, D. Fish consumption and CHD mortality: An updated meta-analysis of seventeen cohort studies. *Public. Health Nutr.* **2012**, *15*, 725–737.
- 109 Bernstein, A.M.; Sun, Q.; Hu, F.B.; Stampfer, M.J.; Manson, J.E.; Willett, W.C. Major dietary protein sources and risk of coronary heart disease in women. *Circulation* **2010**, *122*, 876–883.
- 110 Djousse, L.; Akinkuolie, A.O.; Wu, J.H.Y.; Ding, E.L.; Gaziano, J.M. Fish consumption, ω -3 fatty acids and risk of heart failure: A meta-analysis. *Clin. Nutr.* **2012**, *31*, 846–853.

- 111 Chowdhury, R.; Stevens, S.; Gorman, D.; Pan, A.; Warnakula, S.; Chowdhury, S.; Ward, H.; Johnson, L.; Crowe, F.; Hu, F.B.; *et al.* Association between fish consumption, long chain ω -3 fatty acids, and risk of cerebrovascular disease: Systematic review and meta-analysis. *BMJ* **2012**, *345*, e6698.
- 112 Li, Y.H.; Zhou, C.H.; Pei, H.J.; Zhou, X.L.; Li, L.H.; Wu, Y.J.; Hui, R.T. Fish consumption and incidence of heart failure: A meta-analysis of prospective cohort studies. *Chin. Med. J.* **2013**, *126*, 942–948.
- 113 Xun, P.; Qin, B.; Song, Y.; Nakamura, Y.; Kurth, T.; Yaemsiri, S.; Djousse, L.; He, K. Fish consumption and risk of stroke and its subtypes: Accumulative evidence from a meta-analysis of prospective cohort studies. *Eur. J. Clin. Nutr.* **2012**, *66*, 1199–1207.
- 114 Yinko, S.S.L.L.; Stark, K.D.; Thanassoulis, G.; Pilote, L. Fish consumption and acute coronary syndrome: A meta-analysis. *Am. J. Med.* **2014**, *127*, 848–852.
- 115 Musa-Veloso, K.; Binns, M.A.; Kocenas, A.; Chung, C.; Rice, H.; Oppedal-Olsen, H.; Lloyd, H.; Lemke, S. Impact of low *v.* moderate intakes of long-chain *n*-3 fatty acids on risk of coronary heart disease. *Br. J. Nutr.* **2011**, *106*, 1129–1141.
- 116 Delgado-Lista, J.; Perez-Martinez, P.; Lopez-Miranda, J.; Perez-Jimenez, F. Long chain ω -3 fatty acids and cardiovascular disease: A systematic review. *Br. J. Nutr.* **2012**, *107* (Suppl. 2), S201–S213.
- 117 Rizos, E.C.; Ntzani, E.E.; Bika, E.; Kostapanos, M.S.; Elisaf, M.S. Association between ω -3 fatty acid supplementation and risk of major cardiovascular disease events: A systematic review and meta-analysis. *JAMA* **2012**, *308*, 1024–1033.
- 118 Casula, M.; Soranna, D.; Catapano, A.L.; Corrao, G. Long-term effect of high dose ω -3 fatty acid supplementation for secondary prevention of cardiovascular outcomes: A meta-analysis of randomized, placebo controlled trials. *Atheroscler. Suppl.* **2013**, *14*, 243–251.
- 119 Zheng, T.; Zhao, J.; Wang, Y.; Liu, W.; Wang, Z.; Shang, Y.; Zhang, W.; Zhang, Y.; Zhong, M. The limited effect of ω -3 polyunsaturated fatty acids on cardiovascular risk in patients with impaired glucose metabolism: A meta-analysis. *Clin. Biochem.* **2014**, *47*, 369–377.
- 120 Wen, Y.T.; Dai, J.H.; Gao, Q. Effects of ω -3 fatty acid on major cardiovascular events and mortality in patients with coronary heart disease: A meta-analysis of randomized controlled trials. *Nutr. Metab. Cardiovasc. Dis.* **2014**, *24*, 470–475.
- 121 Enns, J.E.; Yeganeh, A.; Zarychanski, R.; Abou-Setta, A.M.; Friesen, C.; Zahradka, P.; Taylor, C.G. The impact of ω -3 polyunsaturated fatty acid supplementation on the incidence of cardiovascular events and complications in peripheral arterial disease: A systematic review and meta-analysis. *BMC Cardiovasc. Disord.* **2014**, *14*, doi:10.1186/1471-2261-14-70.
- 122 Burr, M.L.; Gilbert, J.F.; Holliday, R.M.; Elwood, P.C.; Fehily, A.M.; Rogers, S.; Sweetnam, P.M.; Deadman, N.M. Effects of changes in fat, fish, and fiber intakes on death and myocardial reinfarction—Diet and reinfarction trial (DART). *Lancet* **1989**, *2*, 757–761.
- 123 Valagussa, F.; Franzosi, M.G.; Geraci, E.; Mininni, N.; Nicolosi, G.L.; Santini, M.; Tavazzi, L.; Vecchio, C.; Marchioli, R.; Bomba, E.; *et al.* Dietary supplementation with *n*-3 polyunsaturated fatty acids and vitamin E after myocardial infarction: Results of the GISSI-Prevenzione trial. *Lancet* **1999**, *354*, 447–455.

- 124 FAO (2014); The state of world fisheries and aquaculture 2014 (Online). The Food and Agriculture Organization of the United Nations, Rome, Italy. Available online: <http://www.fao.org/3/a-i3720e.pdf> (accessed on 10 August 2015).
- 125 Mozaffarian, D.; Rimm, E.B. Fish intake, contaminants, and human health—Evaluating the risks and the benefits. *JAMA* **2006**, *296*, 1885–1899.
- 126 Hohmann, C.D.; Cramer, H.; Michalsen, A.; Kessler, C.; Steckhan, N.; Choi, K.; Dobos, G. Effects of high phenolic olive oil on cardiovascular risk factors: A systematic review and meta-analysis. *Phytomedicine* **2015**, *22*, 631–640.
- 127 Albert, B.B.; Cameron-Smith, D.; Hofman, P.L.; Cutfield, W.S. Oxidation of marine ω -3 supplements and human health. *Biomed. Res. Int.* **2013**, *2013*, doi:10.1155/2013/464921.
- 128 Albert, B.B.; Derraik, J.G.B.; Cameron-Smith, D.; Hofman, P.L.; Tumanov, S.; Villas-Boas, S.G.; Garg, M.L.; Cutfield, W.S. Fish oil supplements in New Zealand are highly oxidised and do not meet label content of n -3 PUFA. *Sci. Rep.* **2015**, *5*, doi:10.1038/srep07928.
- 129 Halvorsen, B.L.; Blomhoff, R. Determination of lipid oxidation products in vegetable oils and marine ω -3 supplements. *Food Nutr. Res.* **2011**, *55*, doi:10.3402/fnr.v55i0.5792.
- 130 Opperman, M.; Benade, S. Analysis of the ω -3 fatty acid content of South African fish oil supplements: A follow-up study. *Cardiovasc. J. Afr.* **2013**, *24*, 297–302.
- 131 Kleiner, A.C.; Cladis, D.P.; Santerre, C.R. A comparison of actual vs. stated label amounts of EPA and DHA in commercial ω -3 dietary supplements in the United States. *J. Sci. Food Agric.* **2015**, *95*, 1260–1267.
- 132 Simopoulos, A.P. Evolutionary aspects of diet: The ω -6/ ω -3 ratio and the brain. *Mol. Neurobiol.* **2011**, *44*, 203–215.
- 133 Simopoulos, A.P. The importance of the ratio of ω -6/ ω -3 essential fatty acids. *Biomed. Pharmacother.* **2002**, *56*, 365–379.
- 134 Calder, P.C. n -3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. *Am. J. Clin. Nutr.* **2006**, *83*, 1505S–1519S.
- 135 Patterson, E.; Wall, R.; Fitzgerald, G.F.; Ross, R.P.; Stanton, C. Health implications of high dietary ω -6 polyunsaturated Fatty acids. *J. Nutr. Metab.* **2012**, *2012*, doi:10.1155/2012/539426.
- 136 FAO/WHO. *Report of the Joint FAO/WHO Expert Consultation on the Risks and Benefits of Fish Consumption*. Food and Agriculture Organization of the United Nations; World Health Organization: Rome, Italy; Geneva, Switzerland, 2011.
- 137 AHA (2015); AHA recommendation (Online). The American Heart Association, Dallas, TX, USA. Available online: http://www.heart.org/HEARTORG/GettingHealthy/NutritionCenter/Fish-101_UCM_305986_Article.jsp#aha_recommendation (accessed on 10 August 2015).
- 138 EFSA (2010); EFSA sets European dietary reference values for nutrient intakes (Online). European Food Safety Authority, Parma, Italy. Available online: <http://www.efsa.europa.eu/en/press/news/nda100326.htm> (accessed on 15 August 2015).
- 139 UK SACN. Advice on Fish Consumption: Benefits and Risks; In *EDID Collection*, Proceedings of the Scientific Advisory Committee on Nutrition, Norwich, UK, 18 June 2004.

- 140 ISSFAL (2004); Recommendations for Intake of Polyunsaturated Fatty Acids in Healthy Adults (Online). Proceedings of the International Society for the Study of Fatty Acids and Lipids, Brighton, UK. Available online: <http://www.issfal.org/statements/pufa-recommendations/statement-3> (accessed on 10 August 2015).
- 141 Elvevoll, E.O.; Barstad, H.; Breimo, E.S.; Brox, J.; Eilertsen, K.E.; Lund, T.; Olsen, J.O.; Osterud, B. Enhanced incorporation of *n*-3 fatty acids from fish compared with fish oils. *Lipids* **2006**, *41*, 1109–1114.
- 142 Galli, C.; Marangoni, F. *n*-3 fatty acids in the Mediterranean diet. *Prostaglandins Leukot. Essent. Fatty Acids* **2006**, *75*, 129–133.
- 143 Visioli, F.; Rise, P.; Barassi, M.C.; Marangoni, F.; Galli, C. Dietary intake of fish vs. formulations leads to higher plasma concentrations of *n*-3 fatty acids. *Lipids* **2003**, *38*, 415–418.

© 2015 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/4.0/>).