



# Whole-Genome Sequences of Two Manganese(II)-Oxidizing Bacteria, *Bosea* sp. Strain BIWAKO-01 and Alphaproteobacterium Strain U9-1i

## Kunihiro Okano,<sup>a</sup> Seiko Furuta,<sup>b</sup> Satoshi Ichise,<sup>b</sup> Naoyuki Miyata<sup>a</sup>

Department of Biological Environment, Faculty of Bioresource Sciences, Akita Prefectural University, Shimoshinjo-Nakano, Akita City, Akita, Japan<sup>a</sup>; Lake Biwa Environmental Research Institute, Otsu, Shiga, Japan<sup>b</sup>

This report describes the whole-genome sequences of two Mn(II)-oxidizing bacteria, filamentous Mn oxide microparticleforming *Bosea* sp. strain BIWAKO-01 and alphaproteobacterium strain U9-1i.

Received 27 September 2016 Accepted 7 October 2016 Published 23 November 2016

Citation Okano K, Furuta S, Ichise S, Miyata N. 2016. Whole-genome sequences of two manganese(II)-oxidizing bacteria, *Bosea* sp. strain BIWAKO-01 and alphaproteobacterium strain U9-1i. Genome Announc 4(6):e01309-16. doi:10.1128/genomeA.01309-16.

Copyright © 2016 Okano et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Naoyuki Miyata, nmiyata@akita-pu.ac.jp.

Microbial Mn(II) oxidation that yields insoluble Mn(III, IV) oxides is a fundamental biogeochemical process in terrestrial and aquatic environments (1–3). Bacterial Mn(II) oxidizers are widespread, yet their ecology remains an enigma (4, 5). For instance, filamentous Mn-rich microparticles that occur in stratified lake and marine environments are considered bacteriogenic, but this has not been clarified (5). Recently, an Mn(II)-oxidizing alphaproteobacterium, *Bosea* sp. strain BIWAKO-01 was found to produce filamentous Mn microparticles under laboratory conditions that are similar to those that occur naturally (6).

Strain U9-1i, isolated from a laboratory Mn(II)-oxidizing enrichment culture (7), is also an alphaproteobacterium, but it deposits formless Mn oxide phases, suggesting that physiological features of the two alphaproteobacteria are different. We temporarily named this alphaproteobacterium strain U9-1i, which is closely related to the *Caulobacteraceae* bacterium OTSz\_A\_272 (DDBJ/EMBL/GenBank accession no. CP013244) based on 16S rRNA gene sequence. Here, we report the whole-genome sequences of strains BIWAKO-01 und U9-1i.

Whole-genome sequencing of strain BIWAKO-01 was carried out using a PacBio RS II (Pacific Biosciences, Menlo Park, CA, USA) with a SMRTbell library (20 kbp). PacBio reads were assembled *de novo* using FALCON version 0.4.0, and draft contig errors were corrected by SMRTAnalysis version 2.3.0.140936.p4.150482.

Whole-genome sequencing of strain U9-1i was carried out using an Illumina HiSeq 1000 system (Illumina, San Diego, CA, USA), with a paired-end library (400 bp), and a Roche/454 PE Genome Sequencer FLX (454 Life Sciences, Branford, CT, USA), with a mate-pair library (8 kb). HiSeq reads were assembled *de novo* using Velvet version 1.2.08 and combined into a hybrid assembly with the 454 reads using GS De Novo Assembler version 2.8 (454 Life Sciences). Gaps between the 14 resulting contigs were closed using NESONI version 0.118 and *Platanus* version 1.2.1. The whole genomes were annotated using the RAST server.

The genomes of strains BIWAKO-01 and U9-1i were represented by two and five contigs (one scaffold), respectively. The strain BIWAKO-01 genome had a total length of 7,326,961 bp and a G+C content of 64.80%. It included 6,988 protein-coding sequences and 67 RNA-coding genes (i.e., four sets of rRNA genes and 55 tRNA genes). The annotation revealed that 4,938 coding sequences (CDSs) exhibited homology to genes with known functions, and the remaining 2,050 genes were identified as encoding hypothetical proteins of unknown function. The strain U9-1i genome had a total length of 4,053,247 bp and a G+C content of 63.30%. It included 4,196 protein-coding sequences and 48 RNAcoding genes (i.e., one set of rRNA genes and 45 tRNA genes). A total of 2,587 CDSs exhibited homology to genes with known functions.

Strain BIWAKO-01 had a multicopper oxidase (MCO) (locus\_tag BIWAKO\_02306 on accession no. BCQA01000001) with 78% sequence identity to MoxA, an Mn(II) oxidase of *Pedomicrobium* sp. (8). Our two strains also possessed proteins similar to MopA, a heme peroxidase-type Mn(II) oxidase (9–11): locus\_tag BIWAKO\_06830 of strain BIWAKO-01 and U91I\_01921 of strain U9-1i, which were related to MopA (accession no. ABY99245) of *Pseudomonas putida* GB-1 (11) with 71% and 54% sequence identities, respectively. Strains BIWAKO-01 and U9-1i may use these enzymes for Mn(II) oxidation.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession numbers BCQA00000000 and BBSY00000000 and refers to the first versions that are described in this paper.

### ACKNOWLEDGMENTS

This work was supported by the President Project of Akita Prefectural University and JSPS KAKENHI grant no. 26281042.

## FUNDING INFORMATION

This work, including the efforts of Kunihiro Okano, was funded by the President Project of Akita Prefectural University. This work, including the efforts of Naoyuki Miyata, was funded by JSPS KAKENHI (26281042).

## REFERENCES

1. Tebo BM, Bargar JR, Clement BG, Dick GJ, Murray KJ, Parker D, Verity R, Webb SM. 2004. Biogenic manganese oxides: properties and mechanisms of formation. Annu Rev Earth Planet Sci **32:**287–328. http://dx.doi.org/10.1146/annurev.earth.32.101802.120213.

- Miyata N, Tani Y, Sakata M, Iwahori K. 2007. Microbial manganese oxide formation and interaction with toxic metal ions. J Biosci Bioeng 104:1–8. http://dx.doi.org/10.1263/jbb.104.1.
- Spiro TG, Bargar JR, Sposito G, Tebo BM. 2010. Bacteriogenic manganese oxides. Acc Chem Res 43:2–9. http://dx.doi.org/10.1021/ar800232a.
- Tebo BM, Johnson HA, McCarthy JK, Templeton AS. 2005. Geomicrobiology of manganese(II) oxidation. Trends Microbiol 13:421–428. http://dx.doi.org/10.1016/j.tim.2005.07.009.
- Ehrlich HL, Newman DK. 2009. Geomicrobiology of manganese, p 347–420. In Ehrlich HL, Newman DK (ed), Geomicrobiology, 5th ed. CRC Press, Boca Raton, FL.
- Furuta S, Ikegaya H, Hashimoto H, Ichise S, Kohno T, Miyata N, Takada J. 2015. Formation of filamentous Mn oxide particles by the alphaproteobacterium *Bosea* sp. strain BIWAKO-01. Geomicrobiol J 32: 666–676. http://dx.doi.org/10.1080/01490451.2014.982837.
- Miyata N, Sugiyama D, Tani Y, Tsuno H, Seyama H, Sakata M, Iwahori K. 2007. Production of biogenic manganese oxides by repeated-batch cul-

tures of laboratory microcosms. J Biosci Bioeng 103:432–439. http://dx.doi.org/10.1263/jbb.103.432.

- Ridge JP, Lin M, Larsen EI, Fegan M, McEwan AG, Sly LI. 2007. A multicopper oxidase is essential for manganese oxidation and laccase-like activity in *Pedomicrobium* sp. ACM3067. Environ Microbiol 9:944–953. http://dx.doi.org/10.1111/j.1462-2920.2006.01216.x.
- Anderson CR, Johnson HA, Caputo N, Davis RE, Torpey JW, Tebo BM. 2009. Mn(II) oxidation is catalyzed by heme peroxidases in "Aurantimonas manganoxydans" strain SI85-9A1 and Erythrobacter sp. strain SD-21. Appl Environ Microbiol 75:4130–4138. http://dx.doi.org/10.1128/ AEM.02890-08.
- Andeer PF, Learman DR, McIlvin M, Dunn JA, Hansel CM. 2015. Extracellular heme peroxidases mediate Mn(II) oxidation in a marine *Roseobacter* bacterium via superoxide production. Environ Microbiol 17: 3925–3936. http://dx.doi.org/10.1111/1462-2920.12893.
- Geszvain K, Smesrud L, Tebo BM. 2016. Identification of a third Mn(II) oxidase enzyme in *Pseudomonas putida* GB-1. Appl Environ Microbiol 82:3774–3782. http://dx.doi.org/10.1128/AEM.00046-16.