

Therapeutic Drug Monitoring of Ceftazidime-Avibactam Concentrations in Carbapenem-Resistant *K. pneumoniae*-Infected Patients With Different Kidney Statuses

Xin-Qi Teng^{1†}, Qiang Qu^{2,3†}, Yue Luo⁴, Wen-Ming Long^{1,5}, Hai-Hui Zhuang¹, Jiao-Hua Xu^{1,6}, Yu-Xin Wen^{1,7}, Hui-Lin Zhang^{1,8} and Jian Qu^{1*}

OPEN ACCESS

Edited by:

Alvaro Francisco Lopes Sousa, University of São Paulo, Brazil

Reviewed by:

Hasan Ejaz, Al Jouf University, Saudi Arabia Joao Massud, Independent researcher, Sao Paulo, Brazil

*Correspondence:

Jian Qu qujianstanley@csu.edu.cn

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Drugs Outcomes Research and Policies, a section of the journal Frontiers in Pharmacology

Received: 23 September 2021 Accepted: 05 May 2022 Published: 22 June 2022

Citation:

Teng X-Q, Qu Q, Luo Y, Long W-M, Zhuang H-H, Xu J-H, Wen Y-X, Zhang H-L and Qu J (2022) Therapeutic Drug Monitoring of Ceftazidime-Avibactam Concentrations in Carbapenem-Resistant K. pneumoniae-Infected Patients With Different Kidney Statuses. Front. Pharmacol. 13:780991. doi: 10.3389/fphar.2022.780991 ¹Department of Pharmacy, The Second Xiangya Hospital, Central South University, Institute of Clinical Pharmacy, Central South University, Changsha, China, ²Department of Pharmacy, Xiangya Hospital, Central South University, Changsha, China, ³National Clinical Research Center for Geriatric Disorders, Xiangya Hospital, Central South University, Changsha, China, ⁴Department of Pharmacy, The People's Hospital of Liuyang, Liuyang, China, ⁵Department of Pharmacy, Second People's Hospital of Huaihua, Huaihua, China, ⁶Department of Pharmacy, The Fourth People's Hospital of Yiyang, Yiyang, China, ⁷Department of Pharmacy, Lixian People's Hospital, Lixian, China, ⁸Department of Pharmacy, Lixian Hospital of Traditional Chinese Medicine, Changde, China

Aims: Carbapenem-resistant *K. pneumoniae* (CRKP) is the most common carbapenemresistant *Enterobacteriaceae* with high mortality. Ceftazidime-avibactam (CAZ-AVI) has exhibited excellent *in vitro* activity *in vivo* against CRKP. However, the efficacy of CAZ-AVI in KPC-producing CRKP-infected patients with different kidney statuses varies, such as renal insufficiency, normal renal function, and augmented renal clearance (ARC). We explored the use of therapeutic drug monitoring (TDM) to evaluate the concentration and efficacy of CAZ-AVI in CRKP-infected patients with different kidney statuses.

Methods: Serum concentrations for CAZ and AVI were determined by the high-performance liquid chromatography method. Bacterial identification, routine susceptibility testing, renal function index, and others were performed in standard protocols in the hospital's clinical laboratories.

Results: In the two patients with ARC, in case 1, CAZ-AVI 2.5g q6h was used with good efficacy, and the concentrations were up to the pharmacokinetics/pharmacodynamics targets. In Case 2, 2.5 g q8h was used with invalid effectiveness, and AVI C_{min} was only 0.797 mg/l, which is lower than the PK/PD target. Case 3 was renal insufficiency using CAZ-AVI 1.25 q8h, and case 4 was normal renal function using 2.5 g q8h. Their concentrations were both up to the PK/PD targets.

Conclusion: TDM results demonstrated that CAZ-AVI steady-state plasma concentration varies among patients with different kidney statuses, providing evidence for the utility of TDM of CAZ-AVI in individualized drug dose adjustment. ARC patients may need more CAZ-AVI daily doses than the standard dose.

Keywords: carbapenem-resistant K. pneumonia, therapeutic drug monitoring, KPC, ceftazidime-avibactam, CAZ-AVI

INTRODUCTION

World Health Organization lists carbapenem-resistant Enterobacteriaceae (CRE) as the "critical" group of bacteria that pose the greatest threat to human health (WHO, 2017). Especially, carbapenem-resistant *K. pneumoniae* (CRKP) is also the most common CRE with high mortality (Zhang et al., 2018).

Carbapenem resistance within Enterobacteriales derives from two main mechanisms, including the acquisition of carbapenemase genes that encode enzymes capable of hydrolyzing carbapenems and a decrease in the uptake of antibiotics by a qualitative or/and quantitative deficiency of porin expression in association with overexpression of βlactamases that possess a very weak affinity for carbapenems (Nordmann et al., 2012). Carbapenem-hydrolyzing β -lactamases are of three categories: 1) class A serine enzymes, such as KPCtype enzymes; 2) class B metal enzymes, such as NDM, VIM, and IMP metallo-β-lactamases, and 3) class D serine enzymes, such as OXA-48-type enzymes, which confer resistance to most βlactams, including carbapenems (Usman Qamar et al., 2020; Findlay et al., 2021). KPC-Kp exhibits extensive drug-resistant phenotypes, and there has been an ongoing debate on the best strategies for managing KPC-Kp infections.

Avibactam is a novel non- β -lactam β -lactamase inhibitor that restores the activity of ceftazidime against those producing KPC and OXA-48 *carbapenemases* (Chen et al., 2021). Ceftazidime/ avibactam (CAZ-AVI) has exhibited excellent *in vitro* activity *in vivo* against KPC-producing *Enterobacteriaceae* and superior efficacy against these isolates compared with other established treatment regimens (van Duin and Bonomo, 2016; van Duin et al., 2018; Tumbarello et al., 2021).

However, the efficacy of CAZ-AVI in KPC-producing CRKPinfected patients with different kidney statuses varies. The kidneys predominantly clear CAZ and AVI with similar pharmacokinetics (PK) profiles and protein binding (Li et al., 2020). Therefore, patients with different renal function state, such as renal insufficiency, normal renal function, and augmented renal clearance (ARC), have different degrees of drug clearance (Li et al., 2020).

We report a case series about KPC-producing CRKP-infected patients with different kidney statuses treated with CAZ-AVI. Therapeutic drug monitoring (TDM) evaluated the adequacy of therapy and guided the dose selection and dose optimization for CAZ-AVI.

MATERIALS AND METHODS

Sample Collection and Therapeutic Drug Monitoring

Serum samples were planned at the following intervals 3 days after CAZ-AVI treatment: 0.5 h immediately before the next dose and 5 min after intravenous drip. Blood samples were collected and were then immediately sent to the central laboratory for centrifuging and serum separation. Serum samples were stored in transport tubes at -80° C until they were assayed. The high-performance liquid chromatography (HPLC) method

determined serum concentrations for CAZ and AVI referring to relevant literature studies (Wang et al., 2019; Deng Yang et al., 2020). Plasma concentrations were determined by HPLC on CAPCELL PAK C18 column (4.6 mm \times 250 mm, 5.0 µm) with mobile phase A of 50 mmol L⁻¹ potassium dihydrogen phosphate solution, mobile phase B of the mixed organic phase (acetonitrile/ methanol/water = 7:2:1), A/B (V/V, 93:7), a flow rate of 1.0 ML min⁻¹, and a wavelength is 254 nm.

Antimicrobial Susceptibility Testing

Bacterial identification and routine susceptibility testing were performed in standard protocols in the hospital's clinical laboratories. Susceptibilities were retested using the VITEKVR 2 system (bioMe'rieux, Marcy-l'E' toile, France). The MICs of CZA for each strain were evaluated using the trace broth double dilution method. A fixed avibactam concentration of 4 µg/ml in conjunction with two-fold dilutions of ceftazidime was used in these tests according to Clinical and Laboratory Standards Institute (CLSI) guidelines (Institute, 2020). Results were interpreted by CLSI guidelines (Institute, 2020). The modified Hodge test (MHT) with EDTA disk test was used to detect carbapenemase-producing *Enterobacteriaceae* phenotypically (Yan et al., 2014).

CASE PRESENTATION

Case 1

A 56-year-old man was admitted to the Neurological Intensive Care Unit (NICU) due to fever for 3 weeks, and abnormal mental behavior for 2 weeks. The patient was admitted with a temperature of 39.3°, accompanied by chills, fatigue, and head pain. Head MRI showed abnormal signals in the left hippocampus, and the CT scan of the lungs showed lung infection. Considering the possibility of viral meningoencephalitis and pneumonia, meropenem 1g q8h, acyclovir, and foscarnet sodium were used for antiviral and antibacterial purposes. The patient was lethargic. Peripherally inserted central catheter and ventilator were used in the patient. The creatinine clearance rate (CrCl) was 145.32 ml/min/1.73 m². On the third day in the hospital, bilateral blood cultures showed ESBL (-), carbapenem-sensitive pneumonia. The sputum culture showed stenotrophomonas maltophilia and hence SMZco was added. On day 11, bilateral blood cultures turn negative. While on day 15, the urine culture revealed the CRKP (800 cfu/ml) urinary tract infection, only sensitive to polymyxin B (PMB) (MIC ≤ 1) and CAZ-AVI(MIC = 2/4). On day 21, CRKP strains were isolated from sputum and bronchoalveolar fluid (BALF) cultures, only sensitive to PMB (MIC \leq 1) and CAZ-AVI (MIC = 2/4). Then, the antibiotic regimens were changed to meropenem 2g q8h plus fosfomycin 4g q8h. Subsequently, CRKP was repeatedly detected in sputum, alveolar lavage fluid, and urine. While the clinical efficacy was not satisfactory, considering that the CrCl was 194.98 ml/min/1.73 m², CAZ-AVI 2.5g q6h was substituted for fosfomycin on day 25. Three days later, TDM of the concentration of CAZ-AVI showed the following: CAZ Cmax 149.21 mg/L and Cmin 14.33 mg/L; AVI Cmax 37.76 mg/L, and

 C_{min} 3.16 mg/L. After 10 days of CAZ-AVI treatment, the sputum and urine cultures turned negative. Five more days later, the patient's bacterial infection condition improved, and the antibiotic regimens were discontinued. On day 58, the patient's intracranial condition was stable and he was transferred to a local hospital for rehabilitation.

Case 2

A 32-year-old man with cerebellar infarction and bodyweight of 80 kg was transferred to NICU after the venous embolus, posterior fossa decompression, dilated dural repair, and pulmonary infection on May 18, 2021. Upon admission, the patient had a temperature of 39°C, a pulse of 140 beats/min, and was delirious, with a white blood cell count of 16.25×10^9 /L, a neutrophil ratio of 91.60%, the C-reactive protein of 278.00 mg/L, the procalcitonin of 0.321 ng/ml, and t interleukin-6 of 6137.00 pg/ml. The initial anti-infection regimen was meropenem (2g q8h) combined with linezolid (600mg q12h). On day 16, CRKP was isolated from the cerebrospinal fluid and sputum, both sensitive to PMB (MIC \leq 1) and CAZ-AVI (MIC = 2/4). PMB was administered intravenously (75mg q12h) with intrathecal administration (3.3mg qd). Twelve days later, CRKP strains were still found positive in sputum and BALF culture but clean in cerebrospinal fluid. On day 35, the regiments for CRKP were changed to CAZ-AVI (2.5g q6h on the first day, then 2.5g g8h) and meropenem (1g g6h). The patient's CrCl was 295.49 ml/ min/1.73 m² on day 35. Three days later, the TDM of CAZ-AVI results showed the following: CAZ Cmax 57.13 mg/L and Cmin 4.96 mg/L, and AVI C_{max} 21.267 mg/L and C_{min} 0.797 mg/L. On day 42, the BALF culture results showed that CRAB strains instead of CRKP strains were found after 7 days of CAZ-AVI treatment. So, tigecycline (TGC) (100mg q12h) combined with meropenem was used. However, CRKP was detected in BALF several times later. The patient's infection failed to improve and he was transferred to a rehabilitation hospital for further treatment.

Case 3

A 77-year-old man was hospitalized in NICU with progressive limb weakness and a temperature of 38.6°C on day 1. After admission, systemic examinations revealed pulmonary infection, hypertension, and chronic renal insufficiency. The initial anti-infective therapy was piperacillin/tazobactam. After 2 days, the patient was delirious, unresponsive, had low oxygen saturation, and was intubated. The inflammatory markers were high (PCT4.48 ng/ml, CRP 241 mg/L), and the antibacterial regiment was switched to meropenem 1g q8h. On day 23, the sputum and BALF culture results showed CRKP infection, which was just sensitive to CAZ-AVI (MIC = 4/4) and PMB (MIC ≤ 1). The patient had renal insufficiency, and the CrCl was 39.78 ml/ min/1.73 m². CAZ-AVI (first day 2.5g q8h, later 1.25g q8h) combined with meropenem was used to anti-CRKP. TDM showed that CAZ C_{max} 77.22 mg/L, C_{min} 37.83 mg/L; AVI C_{max} 20.24 mg/L, C_{min} 8.67 mg/L. After 12 days of treatment with CAZ-AVI, the CRKP strains were clean in sputum and BALF culture, while the CRAB strains were isolated. The CAZ-AVI was replaced by PMB (75 mg loading dose with 50mg q12h maintenance dose) on day 40. On day 44, the patient died of gastrointestinal bleeding and lowered blood pressure.

Case 4

A 35-year-old man had a poor appetite, fatigued for more than a month, and in coma for 20 days. On April 15, 2021, this patient was diagnosed with chronic and acute liver failure and hepatic encephalopathy. The patient was admitted to the department of liver transplantation on May 10, 2021. Under general anesthesia, the patient underwent orthotopic classical liver transplantation plus intestinal adhesiolysis. After surgery, the inflammatory markers were high (PCT 3.35 ng/ml, CRP 39.26 mg/L), and the patient was given the anti-infection treatment of meropenem (1g q8h), PMB (75mg q12h), teicoplanin (400mg qd), and caspofungin (50mg qd). On the 4th day of admission, CRKP (PMB MIC \leq 1, TGC MIC \leq 1, CAZ-AVI MIC = 8/4) and CRAB were isolated from the sputum culture. Meropenem and teicoplanin were discontinued, and TGC (100mg q12h) combined with PMB was replaced to treat negative bacterial infection. Later, CRKP (PMB MIC \leq 1, CAZ-AVI MIC = 8/4) was isolated from wound secretion, blood, and stool cultures. However, multiple sputum cultures, blood, and abdominal drainage fluid cultures indicated CRKP. On day 11 (2021.05.21), the CrCl of the patient was 170.44 ml/min/ 1.73 m² and so TGC was replaced with CAZ-AVI 2.5g q8h. Three days later, the patient's clinical infection symptoms and inflammatory indicators did not improve; considering ARC, CAZ-AVI was changed to q6h. Two days later, the TDM of CAZ-AVI results showed the following: CAZ C_{max} 111.18 mg/ L and Cmin 28.81 mg/L; and AVI Cmax 32.95 mg/L and Cmin 7.73 mg/L, while the CrCl decreased to 103.06 ml/min/1.73 m². PMB was discontinued on day 17. On day 29, the patient's PCT value decreased to 0.415 ng/ml and the CrCl was 100.29 ml/min/ 1.73 m², while CRKP was still isolated from sputum. CAZ-AVI was given with 2.5g q8h to continue until day 39. Although the CRKP strains were still isolated from sputum on days 45 and 47, the patient had no fever with a slight cough and no sputum, and inflammatory markers tended to be normal. After 50 days of treatment, the patient was discharged.

Summary of Cases

The basic information of the four cases is shown in **Table 1**. Patients were all men with ages ranging from 35 to 77 years. Three patients were admitted to NICU with neurological symptoms, and one patient was admitted to the liver transplantation department because of hepatic failure. All patients had HAP caused by CRKP during hospitalization. Case 1 also had UTI; Case 2 also had intracranial infection; Case 4 had BSI and ABSSSI, and all were infected with CRKP. MER + CAZ-AVI regimens were used for three patients used and CAZ-AVI + TGC regimens were used for one patient. Two patients showed good responses to the combined regimen of CAZ-AVI, and one patient died because of gastrointestinal bleeding and lowered blood pressure. The anti-infection effect of CAZ-AVI was not good in Case 2.

All the CRKP strains produced KPC enzymes and were isolated from urine, sputum, BALF, cerebrospinal fluid, wound

TDM of CAZ-AVI Concentrations in CRKP Infection

Parameters	Case 1	Case 2	Case 3	Case 4
Age (years)	56	32	77	35
Sex	Male	Male	Male	Male
Admission department	NICU	NICU	NICU	Liver Transplantation Department
Underlying disease	Hypertension, secondary epilepsy, liver insufficiency	Hypertension, arrhythmology, pneumonia	Hypertension, pneumonia, gout, old tuberculosis, hemifacial spasm	Liver failure, hepatic encephalopathy, chronic viral hepatitis B, spontaneous peritonitis, pneumonia
The highest temperature (°C)	39.5	39.3	39	40
WBC (normal 4–10, × 10 ⁹ /L)	10.15	18.22	11.35	10.95
Neutrophil ratio (%)	82.2	94.5	82.5	90.4
CRP (mg/L)	44.9	278	241	168.4
PCT (ng/ml)	0.179	0.321	4.48	39.6
Infection type	HAP, UTI	Intracranial infection, HAP	HAP	BSI, HAP, ABSSSI
Treatment regimens for CRKP (days)	MER + CAZ-AVI (15)	MER + PMB; MER + CAZ-AVI (9)	MER + CAZ-AVI (12)	MER + PMB; MER + TGC; CAZ-AVI + TGC (28)
Outcomes	Effective	Ineffective	Dead	Effective

TABLE 1 | Clinical characteristics of patients.

UTI, urinary tract infection; HAP, hospital acquired pneumonia; ABSSSI, acute bacterial skin and skin-structure infection; CAZ-AVI, ceftazidime-avibactam; TGC, tigecycline; MER, meropenem: CRKP, carbapenem-resistant K, pneumoniae.

TABLE 2	In vitro susceptibilit	v of CRKP to	the tested antibiotics	(with MICs in µa/mL aiven in	parentheses) and bacterial clearance

Cases	Sources of CRKP isolated specimens	Carbapenemase	Bacterial clearance (days)	Antibiotics*				
				CAZ-AVI	РМВ	TGC	CAZ	
Case 1	Urine	KPC	Yes (8)	S (2/4)	S (≤1)	I (4)	R (>32)	
	Sputum/BALF	KPC	Yes (9)	S (2/4)	S (≤1)	S (1), I (4)	R (>32)	
Case 2	Cerebrospinal fluid	KPC	Yes (12)	S (2/4)	S (≤1)	I (4)	R (>32)	
	Sputum	KPC	Yes (7)	S (2/4)	S (≤1)	I (4)	R (>32)	
Case 3	Sputum/BALF	KPC	Yes (9)	S (4/4)	S (≤1)	I (4)	R (>32)	
Case 4	Sputum	KPC	Not cleared	S (8/4)	S (≤1)	S (≤1)	R (>32)	
	Wound secretion	KPC	No repeated testing	S (8/4)	S (≤1)	I (4)	R (>32)	
	Blood	KPC	Yes (4)	S (8/4)	S (≤1)	I (4)	R (>32)	
	Stool	KPC	No repeated testing	S (8/4)	S (≤1)	S (2)	R (>32)	

CAZ-AVI, ceftazidime-avibactam; CAZ, ceftazidime; KPC, K. pneumoniae carbapenemase; PMB, polymyxin B; S, sensitive; I, intermediate; BALF, bronchoalveolar fluid. * Resistant to all other antibiotics.

secretion, blood, and stool (**Table 2**). All strains are sensitive to PMB (MIC \leq 1) and CAZ-AVI (MIC 2/4, 4/4, and 8/4). Most strains were intermediate sensitive to TGC, and all strains were resistant to CAZ (MIC > 32). After CAZ-AVI-based regimen treatment, CRKP was clean in Cases 1–3, and the blood of case 4.

Cases 1 and 2 were ARC patients with CRCL 194.98 and 295.49 ml/min/1.73 m². The AVI C_{min} was 3.16 mg/L when Case 1 was given CAZ-AVI 2.5g q6h. Moreover, in Case 2, after CAZ-AVI 2.5g q6h was given on the first day and 2.5q8h was given subsequently, the AVI C_{min} was only 0.797 mg/L, and the CAZ C_{min} was only 4.96 mg/L. Case 3 had renal insufficiency with CRCL 39.78 ml/min/1.73 m², and TDM showed that the C_{min} of CAZ and AVI were higher than the MIC value after CAZ-AVI 2.5g q8h was given on the first day and 1.25 q8h was given subsequently. In case 4, the renal function was normal, and the plasma concentration of CAZ-AVI was higher than the MIC value after CAZ-AVI 2.5g q8h was given for 3 days and then 2.5g q6h for 2 days (**Table 3**).

DISCUSSION

This report described the therapeutic drug monitoring of CAZ-AVI concentrations in *K. pneumoniae* carbapenemase (KPC)producing CRKP-infected patients with different kidney statuses, such as augmented renal clearance (ARC), normal renal function, and renal insufficiency. TDM results demonstrated that CAZ-AVI steady-state plasma concentration varied among patients with different kidney statuses, providing evidence for the utility of TDM of CAZ-AVI in individualized drug dose adjustment.

CAZ and AVI have similar pharmacokinetics and both are eliminated mainly through kidneys (Merdjan et al., 2015). Therefore, the renal function status significantly affects the PK/PD of CAZ-AV, thus influencing the clinical and microbiological efficacy (Merdjan et al., 2017).

Augmented renal clearance (ARC) is a pathophysiological condition defined as the occurrence of a measured CrCl \geq 130 ml/min/1.73 m² in males and \geq 120 in females coupled

TABLE 3	Renal function and	blood concentration	n of CAZ-AVI detected	bv TDM
				~,

Cases	CAZ-AVI regimen	CrCl (ml/min/1.73m ²)	CAZ C _{max} (mg/L)	CAZ C _{min} (mg/L)	AVI C _{max} (mg/L)	AVI C _{min} (mg/L)
Case 1	2.5g q6h	194.98	149.21	14.33	37.76	3.16
Case 2	2.5g q6h first day, then 2.5q8h	295.49	57.13	4.96	21.267	0.797
Case 3 Case 4	2.5g q8h first day, then 1.25 q8h 2.5g q8h for 3 days, then 2.5g q6h for 2 days to TDM	39.78 Before CAZ-AVI use: 170.44, TDM day: 103.06	77.22 111.18	37.83 28.81	20.25 32.95	8.67 7.73

CrCl, creatinine clearance; CAZ, ceftazidime; AVI, avibactam; TDM, therapeutic drug monitoring.

with a normal serum creatinine value (0.6-1.4 mg/dl) (Hobbs et al., 2015; Cook and Hatton-Kolpek, 2019). ARC occurred in patients with subarachnoid hemorrhage, trauma, burn, and sepsis patients. A subgroup analysis of the REPROVE trial about CAZ-AVI showed that in ARC patients, the standard dosage of 2.5g q8h over 2 h ensured >95% probability of target attainment of a more conservative target of 50% fT > MIC, while a 35% decrease in drug exposure was observed compared to patients with normal renal function (mean AUC) (Das et al., 2019; Li et al., 2020). Realworld experience is limited to CAZ-AVI's PK/PD analysis in two ARC critically ill patients. Treatment with 2.5g q8h over 2 h allowed the achievement of the conservative target of 50% fT >MIC against pathogens with a MIC up to 16 mg/L (Stein et al., 2019). For multidrug-resistant bacterial infection or severe infection, the higher the time-dependent antibacterial agents % fT > MIC, the better the efficacy is, and even the % fT > $4-5 \times$ MIC is better (Stein et al., 2019). Furthermore, antibiotic dosing regimens required to suppress the emergence of resistance should be focused on achieving the PK/PD target of 100% fT > 4–8 \times MIC (Guilhaumou et al., 2019). Previous studies demonstrated that the PK/PD targets for CAZ-AVI (CAZ 50% fT > MIC 8 mg/L and AVI 50% fT > C_T of 1 mg/L) were therefore deemed appropriate for the probability of target attainment (PTA) analyses to select ceftazidime-avibactam dosage (Coleman et al., 2014; Berkhout et al., 2016; Nichols et al., 2018; Das et al., 2019). A study used a 10,000-patient Monte Carlo simulation (MCS) to calculate the PTA and the cumulative fraction of response (CFR) for different CAZ-AVI dosage regimens to evaluate their efficacies and optimize the best initial dosage regimen found that for patients with CrCl values of 121-190 ml/min, and the standard dose (2.5g iv q8h) failed to reach 90% CFR against some Enterobacteriaceae members and P. aeruginosa (Dai et al., 2021). In the two patients with ARC, case 1 used CAZ-AVI 2.5g q6h, and AVI C_{min} was 3.16 mg/L. Case 2 used 2.5q8h (first day 2.5g q6h), and AVI C_{min} was only 0.797 mg/L. It implies that ARC patients may need more CAZ-AVI daily doses than the standard dose.

Due to the predominantly renal excretion of both CAZ and AVI, the concentrations of both drugs increase with increasing severity of renal impairment (Merdjan et al., 2017; Das et al., 2019). CAZ-AVI dosage adjustments are required according to different severities of renal insufficiency to minimize the risk of overexposure in patients with a CrCl of \leq 50 ml/min (Li et al., 2020). The modified dosage regimens are now included in product labels, which advise close monitoring of CrCl in patients with renal impairment (Das et al., 2019). A study also

discussed that patients with rapidly improving renal function without a timely change in dosage were underdosed (Li et al., 2020). PTAs for the CAZ-AVI original dosage regimens given to patients with moderate renal impairment in the event of renal function improvement to normal renal function were well below the targeted range (Li et al., 2020). Hence, if the renal insufficiency is caused by infection, the dose of CAZ-AVI can be appropriately increased according to the adjustment of the manual to control the infection better, and the dose can be adjusted timely with the improvement of renal function. Therefore, despite renal insufficiency after infection in patient 3, the creatinine clearance rate was 39.78 ml/min/1.73 m². The dose of CAZ-AVI given was standard on the first day and was adjusted to 1.25g q8h later. TDM results showed that the Cmin values of CAZ and AVI were higher than the target concentration, and the PK/ PD target rate was 100%.

In recent years, the extensive use of carbapenems increased carbapenem resistance in K. pneumoniae. The primary mechanism is the production of a carbapenemase classified into the Ambler class A (K. pneumoniae carbapenemase, KPC), B (the metalloenzymes NDM, VIM, and IMP), and D (oxacillin enzyme, OXA) (Logan and Weinstein, 2017). A previous study found that KPC-2 was the most frequently detected gene in CRKP strains in Southwestern China (Tian et al., 2019). The coding gene of KPC is *blaKPC*, which can be transferred by the plasmid among K. pneumoniae strains through Tn3-based transposon Tn4401 (Logan and Weinstein, 2017). NDM-1 K. pneumoniae is a newly emerged highly resistant bacteria that can produce NDM-1, which is capable of breaking down beta-lactam antibiotics, including avibactam (Kumarasamy et al., 2010). To date, NDM-1 K. pneumoniae has also been reported worldwide, including in China (Tian et al., 2019).

We explored the use of TDM to evaluate the concentration and efficacy of CAZ-AVI in CRKP-infected patients with different kidney statuses. The limited clinical samples limit the results of this study. Large-scale clinical sample experiments are needed to confirm the formulation of an individualized administration scheme of CAZ-AVI based on different renal functions.

CONCLUSION

In conclusion, our study demonstrated that CAZ-AVI steadystate plasma concentration varies among patients with different kidney statuses, including ARC, normal renal function, and renal insufficiency, providing evidence for the utility of TDM of CAZ-AVI in individualized drug dose adjustment. ARC patients may need more CAZ-AVI daily doses than the standard dose.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material; further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by The study protocol was approved by the Ethics Committees of the Second Xiangya Hospital of Central South University (LYF-2020021) in Changsha, China. Informed

REFERENCES

- Berkhout, J., Melchers, M. J., van Mil, A. C., Seyedmousavi, S., Lagarde, C. M., Schuck, V. J., et al. (2016). Pharmacodynamics of Ceftazidime and Avibactam in Neutropenic Mice with Thigh or Lung Infection. *Antimicrob. Agents Chemother.* 60 (1), 368–375. doi:10.1128/AAC.01269-15
- Chen, H. Y., Jean, S. S., Lee, Y. L., Lu, M. C., Ko, W. C., Liu, P. Y., et al. (2021). Carbapenem-Resistant Enterobacterales in Long-Term Care Facilities: A Global and Narrative Review. *Front. Cell Infect. Microbiol.* 11, 601968. doi:10.3389/ fcimb.2021.601968
- Coleman, K., Levasseur, P., Girard, A. M., Borgonovi, M., Miossec, C., Merdjan, H., et al. (2014). Activities of Ceftazidime and Avibactam against β-lactamaseproducing Enterobacteriaceae in a Hollow-Fiber Pharmacodynamic Model. *Antimicrob. Agents Chemother.* 58 (6), 3366–3372. doi:10.1128/AAC.00080-14
- Cook, A. M., and Hatton-Kolpek, J. (2019). Augmented Renal Clearance. Pharmacotherapy 39 (3), 346–354. doi:10.1002/phar.2231
- Dai, Y., Chang, W., Zhou, X., Yu, W., Huang, C., Chen, Y., et al. (2021). Evaluation of Ceftazidime/Avibactam Administration in Enterobacteriaceae and *Pseudomonas aeruginosa* Bloodstream Infections by Monte Carlo Simulation. Drug Des. Devel Ther. 15, 2899–2905. doi:10.2147/DDDT.S309825
- Das, S., Li, J., Riccobene, T., Carrothers, T. J., Newell, P., Melnick, D., et al. (2019). Dose Selection and Validation for Ceftazidime-Avibactam in Adults with Complicated Intra-abdominal Infections, Complicated Urinary Tract Infections, and Nosocomial Pneumonia. *Antimicrob. Agents Chemother.* 63 (4), e02187–18. doi:10.1128/AAC.02187-18
- Deng Yang, X. B., Li, X., Qi, Q., Zhou, G-Q., Guo, S-W., and Luo, X-L. (2020). Determination of Cefepime and Ceftazidine in Human Plasma by HPLC Method and its Establishment of Clinical Sampling Process. *Chin. J. Hosp. Pharm.* 40 (20), 2120–2125. doi:10.13286/j.1001-5213.2020.20.06
- Findlay, J., Poirel, L., Juhas, M., and Nordmann, P. (2021). KPC-mediated Resistance to Ceftazidime-Avibactam and Collateral Effects in *Klebsiella pneumoniae. Antimicrob. Agents Chemother.* 65 (9), e0089021. doi:10.1128/ AAC.00890-21
- Guilhaumou, R., Benaboud, S., Bennis, Y., Dahyot-Fizelier, C., Dailly, E., Gandia, P., et al. (2019). Optimization of the treatment with beta-lactam antibiotics in critically ill patients-guidelines from the French Society of Pharmacology and Therapeutics (Société Française de Pharmacologie et Thérapeutique-SFPT) and the French Society of Anaesthesia and Intensive Care Medicine (Société Française d'Anesthésie et Réanimation-SFAR). *Crit. Care* 23 (1), 104. doi:10. 1186/s13054-019-2378-9
- Hobbs, A. L., Shea, K. M., Roberts, K. M., and Daley, M. J. (2015). Implications of Augmented Renal Clearance on Drug Dosing in Critically Ill Patients: A Focus on Antibiotics. *Pharmacotherapy* 35 (11), 1063–1075. doi:10.1002/phar.1653

consent was obtained from patients and guardians. The patients/participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

JQ contributed to the design of the article, and X-QT, QQ, and YL contributed to the drafting of the article. W-ML and H-HZ reviewed and modified the article. J-HX, Y-XW, and H-LZ contributed to the sample collection.

ACKNOWLEDGMENTS

We thank the patients who participated in our study.

- Institute, C. a. L. S. (20202020). Performance Standards for Antimicrobial Susceptibility Testing. thirty edition. Wayne, PA: CLSI.
- Kumarasamy, K. K., Toleman, M. A., Walsh, T. R., Bagaria, J., Butt, F., Balakrishnan, R., et al. (2010). Emergence of a New Antibiotic Resistance Mechanism in India, Pakistan, and the UK: a Molecular, Biological, and Epidemiological Study. *Lancet Infect. Dis.* 10 (9), 597–602. doi:10.1016/ S1473-3099(10)70143-2
- Li, J., Lovern, M., Riccobene, T., Carrothers, T. J., Newell, P., Das, S., et al. (2020). Considerations in the Selection of Renal Dosage Adjustments for Patients with Serious Infections and Lessons Learned from the Development of Ceftazidime-Avibactam. Antimicrob. Agents Chemother. 64 (4), e02105–19. doi:10.1128/ AAC.02105-19
- Logan, L. K., and Weinstein, R. A. (2017). The Epidemiology of Carbapenem-Resistant Enterobacteriaceae: The Impact and Evolution of a Global Menace. J. Infect. Dis. 215 (Suppl. l_1), S28–S36. doi:10.1093/infdis/jiw282
- Merdjan, H., Rangaraju, M., and Tarral, A. (2015). Safety and Pharmacokinetics of Single and Multiple Ascending Doses of Avibactam Alone and in Combination with Ceftazidime in Healthy Male Volunteers: Results of Two Randomized, Placebo-Controlled Studies. *Clin. Drug Investig.* 35 (5), 307–317. doi:10.1007/ s40261-015-0283-9
- Merdjan, H., Tarral, A., Das, S., and Li, J. (2017). Phase 1 Study Assessing the Pharmacokinetic Profile and Safety of Avibactam in Patients with Renal Impairment. J. Clin. Pharmacol. 57 (2), 211–218. doi:10.1002/jcph.793
- Nichols, W. W., Newell, P., Critchley, I. A., Riccobene, T., and Das, S. (2018). Avibactam Pharmacokinetic/Pharmacodynamic Targets. Antimicrob. Agents Chemother. 62 (6). e02446-17. doi:10.1128/AAC.02446-17
- Nordmann, P., Dortet, L., and Poirel, L. (2012). Carbapenem Resistance in Enterobacteriaceae: Here Is the Storm!. *Trends Mol. Med.* 18 (5), 263–272. doi:10.1016/j.molmed.2012.03.003
- Stein, G. E., Smith, C. L., Scharmen, A., Kidd, J. M., Cooper, C., Kuti, J., et al. (2019). Pharmacokinetic and Pharmacodynamic Analysis of Ceftazidime/Avibactam in Critically Ill Patients. Surg. Infect. (Larchmt) 20 (1), 55–61. doi:10.1089/sur. 2018.141
- Tian, X., Huang, C., Ye, X., Jiang, H., Zhang, R., Hu, X., et al. (2019). Molecular Epidemiology of and Risk Factors for Extensively Drug-Resistant *Klebsiella pneumoniae* Infections in Southwestern China: A Retrospective Study. Front. Pharmacol. 10, 1307. doi:10.3389/fphar.2019.01307
- Tumbarello, M., Raffaelli, F., Giannella, M., Mantengoli, E., Mularoni, A., Venditti, M., et al. (2021). Ceftazidime-Avibactam Use for *Klebsiella pneumoniae* Carbapenemase-Producing *K. pneumoniae* Infections: A Retrospective Observational Multicenter Study. *Clin. Infect. Dis.* 73, 1664–1676. doi:10. 1093/cid/ciab176
- Usman Qamar, M., S Lopes, B., Hassan, B., Khurshid, M., Shafique, M., Atif Nisar, M., et al. (2020). The Present Danger of New Delhi Metallo-β-Lactamase: a

Threat to Public Health. Future Microbiol. 15, 1759–1778. doi:10.2217/fmb-2020-0069

- van Duin, D., and Bonomo, R. A. (2016). Ceftazidime/Avibactam and Ceftolozane/ Tazobactam: Second-Generation β-Lactam/β-Lactamase Inhibitor Combinations. Clin. Infect. Dis. 63 (2), 234–241. doi:10.1093/cid/ciw243
- van Duin, D., Lok, J. J., Earley, M., Cober, E., Richter, S. S., Perez, F., et al. (2018). Colistin versus Ceftazidime-Avibactam in the Treatment of Infections Due to Carbapenem-Resistant Enterobacteriaceae. *Clin. Infect. Dis.* 66 (2), 163–171. doi:10.1093/cid/cix783
- Wang, Y., Yao, S.-C., Zhang, P-P., Zou, W-B., Jin, L., Xu, M-Z., et al. (2019). Simultaneous Determination of Compound Preparation of Ceftazidime-Avibactam for Injection by HPLC-DAD. *Chin. J. New Drugs* 28 (5), 541–546.
- WHO (2017). Global Priority List of Antibiotic-Resistant Bacteria to Guide Research, Discovery, and Development of New Antibiotics. WHO, 348–365.
- Yan, Y., Yang, H., Pan, L., Sun, K., Fan, H., Lu, Y., et al. (2014). Improving the Efficiency of the Modified Hodge Test in KPC-Producing *Klebsiella pneumoniae* Isolates by Incorporating an EDTA Disk. *Curr. Microbiol.* 69 (1), 47–52. doi:10.1007/s00284-014-0552-5
- Zhang, Y., Wang, Q., Yin, Y., Chen, H., Jin, L., Gu, B., et al. (2018). Epidemiology of Carbapenem-Resistant Enterobacteriaceae Infections: Report from the China

CRE Network. Antimicrob. Agents Chemother. 62 (2). doi:10.1128/AAC. 01882-17

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Teng, Qu, Luo, Long, Zhuang, Xu, Wen, Zhang and Qu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.