

Pharmacodynamic Consequences of Administration of VLA-4 Antagonist CDP323 to Multiple Sclerosis Subjects: A Randomized, Double-Blind Phase 1/2 Study

Christian Wolf¹, Jagdev Sidhu², Christian Otoul³, Dexter L. Morris^{4*}, Jennifer Cnops⁵, Jorg Taubel⁶, Barbara Bennett⁷

1 Clinical Development, Lycalis s.p.r.l., Brussels, Belgium, **2** Clinical Pharmacology/Early Development, CSL Ltd., Melbourne, Australia, **3** Exploratory Statistics, UCB Pharma, Brabant, Belgium, **4** Drug Development, UCB Biosciences Inc., Raleigh, North Carolina, United States of America, **5** Department of Cellular and Molecular Immunology, Vrije Universiteit Brussel, Brussels, Belgium, **6** Richmond Pharmacology, London, United Kingdom, **7** BABennett Consulting, Marietta, Georgia, United States of America

Abstract

Background: Lymphocyte inhibition by antagonism of α_4 integrins is a validated therapeutic approach for relapsing multiple sclerosis (RMS).

Objective: Investigate the effect of CDP323, an oral α_4 -integrin inhibitor, on lymphocyte biomarkers in RMS.

Methods: Seventy-one RMS subjects aged 18–65 years with Expanded Disability Status Scale scores ≤ 6.5 were randomized to 28-day treatment with CDP323 100 mg twice daily (bid), 500 mg bid, 1000 mg once daily (qd), 1000 mg bid, or placebo.

Results: Relative to placebo, all dosages of CDP323 significantly decreased the capacity of lymphocytes to bind vascular adhesion molecule-1 (VCAM-1) and the expression of α_4 -integrin on VCAM-1-binding cells. All but the 100-mg bid dosage significantly increased total lymphocytes and naive B cells, memory B cells, and T cells in peripheral blood compared with placebo, and the dose-response relationship was shown to be linear. Marked increases were also observed in natural killer cells and hematopoietic progenitor cells, but only with the 500-mg bid and 1000-mg bid dosages. There were no significant changes in monocytes. The number of samples for regulator and inflammatory T cells was too small to draw any definitive conclusions.

Conclusions: CDP323 at daily doses of 1000 or 2000 mg induced significant increases in total lymphocyte count and suppressed VCAM-1 binding by reducing unbound very late antigen-4 expression on lymphocytes.

Trial Registration: ClinicalTrials.gov NCT00726648.

Citation: Wolf C, Sidhu J, Otoul C, Morris DL, Cnops J, et al. (2013) Pharmacodynamic Consequences of Administration of VLA-4 Antagonist CDP323 to Multiple Sclerosis Subjects: A Randomized, Double-Blind Phase 1/2 Study. PLoS ONE 8(3): e58438. doi:10.1371/journal.pone.0058438

Editor: Robyn Klein, Washington University, United States of America

Received: July 6, 2012; **Accepted:** February 5, 2013; **Published:** March 5, 2013

Copyright: © 2013 Wolf et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was sponsored by UCB Pharma and Biogen Idec Inc. The funders, in consultation with the authors, were included in discussions relating to study design and provided assistance with data analysis. They had no role in data collection or the decision to publish, and contributions to manuscript preparation were restricted to authors employed by the funders and financing of editorial support provided by Infusion Communications.

Competing Interests: This study was funded by UCB Pharma and Biogen Idec, which were jointly developing CDP323 as a potential treatment for multiple sclerosis. Dr. Morris has been an employee of and holds stock in GlaxoSmithKline and is currently employed by and has received stock options from UCB Biosciences Inc. Among my coauthors, Dr. Wolf was a salaried employee of UCB Pharma S.A. during the time the study was conducted and has been a consultant for Novartis and Synthon. Dr. Sidhu was a salaried employee of and received stock in UCB Pharma during the time the study was conducted. Dr. Otoul is an employee of UCB Pharma. Dr. Taubel is employed by Richmond Pharmacology, which received financial remuneration for their clinical involvement in this study. Dr. Bennett was a salaried employee of UCB Biosciences Inc. during the time the study was conducted and is now an independent consultant. Dr. Cnops reports no conflicts of interest. This does not alter the authors' adherence to all of the PLOS ONE policies on sharing data and materials.

* E-mail: tony.morris@ucb.com

Introduction

Multiple sclerosis (MS) is a chronic, disabling autoimmune disease of the central nervous system (CNS) characterized by inflammation, demyelination, and axonal destruction [1]. Relapsing MS (RMS) has a characteristic pathology of intermittent development of multifocal inflammatory lesions in the brain and spinal cord thought to result from activated lymphocytes breaching the blood-brain barrier [2–4]. Inflammatory mediators such as tumor necrosis factor- α , which is produced during

inflammatory responses, induce vascular endothelial cells to express vascular adhesion molecule-1 (VCAM-1) [5]. The diapedesis of lymphocytes into the CNS is dependent on lymphocyte adhesion to vascular endothelial cells. This adhesion is mediated by the binding of integrins such as very late antigen-4 (VLA-4 or $\alpha_4\beta_1$ integrin), expressed on activated lymphocytes, to VCAM-1 [5–8].

Inhibition of lymphocyte trafficking by antagonism of α_4 integrins is a validated therapeutic approach for inflammatory diseases such as MS [9]. Clinical studies with natalizumab, a

recombinant humanized monoclonal antibody that targets $\alpha_4\beta_1$ integrin, have confirmed that this integrin is an effective therapeutic target for relapsing forms of MS [10]. Through selective inhibition of $\alpha_4\beta_1$ integrin, natalizumab effectively disrupts migration of immune cells and reduces subsequent CNS inflammation [6,11,12]. Therapeutically, this results in reduced numbers of inflammatory brain lesions, lower exacerbation rates, and delayed progression of MS disability [10,12,13].

Small-molecule α_4 antagonists such as CDP323 (UCB Pharma, Brussels, Belgium, and Biogen Idec, Weston, MA, USA), a phenylalanine enamide mixed α_4 antagonist [9], have been considered potential alternatives to therapeutic antibodies because of their increased selectivity for specific α_4 -containing integrin complexes, their inhibition of different affinity states, and the increased patient convenience of oral dosing [14–16]. In addition, their short half-lives compared with those of antibodies permits relatively fast elimination from the body in the event of serious side effects that necessitate termination of therapy.

Preclinical investigations have shown that CDP323 possesses anti-inflammatory properties [6,11], and phase 1 data in human volunteers have suggested that CDP323 is a potent and effective α_4 inhibitor that is well tolerated at oral doses up to 1000 mg twice daily (bid) [17]. The 1000-mg bid CDP323 dosage appears to have a similar effect on peripheral lymphocyte counts in healthy subjects as natalizumab does at doses of 3 and 6 mg/kg in subjects with MS [18]. However, no data have been gathered on the effects of CDP323 on peripheral lymphocyte counts. The magnitude of increased total lymphocyte counts induced by an α_4 antagonist may be indicative of its potential to inhibit lymphocyte trafficking and, therefore, predictive of its therapeutic potential, though this has not been formally validated.

When the current study was designed, a 24-week serial MRI phase 2 study investigating two dosages of CDP323 in RMS subjects was ongoing [19]. The study reported herein was intended to complement data on the exposure-response relationship (ie, the 24-week study) by using CDP323 in a wider dosage range and examining different lymphocyte subsets over a 4-week period in subjects with RMS. The objectives of the present study were to identify and support a minimally effective dosage of CDP323 in RMS therapy (as assessed by biomarker changes) and to collect additional information on the biomarker effects and safety of a previously unused higher dosage (1000 mg bid).

In addition, this study investigated CDP323 1000 mg given once daily (qd) in order to characterize how a partial recovery of VCAM-1 binding during a 24-hour dosing interval would affect the trafficking of lymphocytes. A working hypothesis was that a partial recovery might offer safety advantages.

Methods

This was a hybrid phase 1/phase 2, double-blind, randomized, parallel-group, placebo-controlled study conducted at a single center in London (UK). Male and female subjects aged 18 to 65 years with a diagnosis of RMS, an Expanded Disability Status Scale (EDSS) score of ≤ 6.5 , one relapse within the last 24 months, and no other conditions that could potentially compromise the immune response were recruited to the study. Subjects were excluded if, prior to study screening, they had received immunomodulating or immunosuppressive drugs within 30 days (for interferons, glatiramer acetate, corticosteroids, adrenocorticotropic hormone, or recombinant cytokines), 6 months (for azathioprine, cyclosporine A, or human antibodies), or 12 months (for natalizumab, rituximab, or mitoxantrone); if they had ever been treated with cyclophosphamide; or if they had received total

lymphoid irradiation, any antilymphocyte monoclonal antibody therapy, or any inoculation with attenuated live vaccines in the 30 days prior to screening. Subjects were not permitted to take any form of concomitant immunosuppressive or immunomodulatory therapy during the study or for a period of 30 days following completion or withdrawal from the study.

Subjects were randomized to receive placebo or CDP323 at dosages of 100, 500, or 1000 mg qd or 1000 mg bid for a period of 28 consecutive days (day 1 to day 28; last dose given as a morning dose). Treatment was supplied as hard-gelatin capsules containing 50 mg or 250 mg of CDP323 or matched placebo.

Subjects were hospitalized for the first 5 days following initiation of treatment and for the last 2 nights of the treatment period. Pharmacodynamic (PD) assessments were conducted during the predose phase (screening; day -1 at 08:00, 09:00, 10:00, 12:00, 16:00, and 20:00 hours), during the first day of study medication administration (day 1: predose and 1, 2, 4, 8, 12, and 24 hours post-dose), on day 15 (predose), and after the final dose on day 28 (predose and 1, 2, 4, 8, 12, 24, 48, 72, 96, and 168 hours post-dose). Assessments consisted of measurement of absolute counts of total lymphocytes, B cells, T cells, monocytes, hematopoietic progenitor cells, natural killer (NK) cells, NK T cells, regulator T cells (T_{reg}), and inflammatory T cells. Expression of α_4 was also recorded for each subset of lymphocytes except for inflammatory T cells. In addition, the effect of CDP323 on VCAM-1 inhibition was investigated as well.

The protocol for this trial and supporting CONSORT checklist are available as supporting information; see Checklist S1 and Protocol S1.

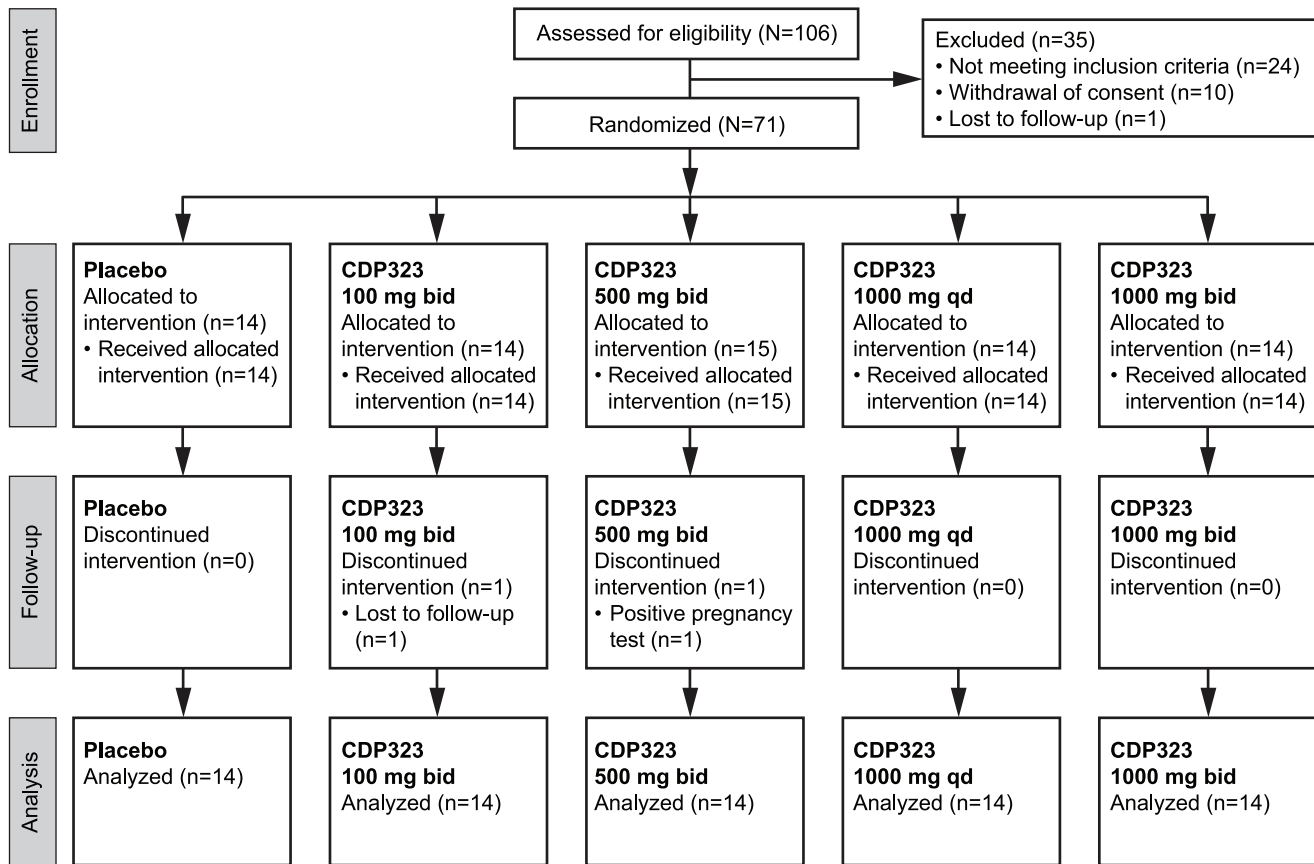
Flow Cytometry Methodology

All samples were analyzed by Esoterix Clinical Trial Services (Mechelen, Belgium). White blood cell and differential white blood cell counts were obtained from each whole blood sample using a hematological analyzer before staining and analysis. A 100- μ L sample of sodium heparin anticoagulated whole blood from each patient was mixed with appropriate amounts of fluorescent monoclonal antibody cocktails, CD3 peridinin chlorophyll protein (perCP), or CD19 perCP antibody complex and incubated in the dark for 20 minutes. To each tube, 4 mL of a warm lysing solution was added. Samples were mixed and allowed to stand in the dark for a further 5 minutes. After washing, pellets were fixated with 500 μ L of 1% paraformaldehyde. Data were acquired with a FACSCaliburTM (BD Biosciences, Erembodegem, Belgium) and analyzed with CellQuestTM software (BD Biosciences). Fluorescence markers for each subject were optimized to assess nonspecific staining.

Statistical Analysis

Because of the exploratory nature of this study, no formal sample size computation was performed. A sample of 15 subjects per group was considered adequate to describe the effects of CDP323 upon biomarkers across the three tested dosage levels.

Statistical analysis was performed using SAS[®] software version 9.1 (SAS Institute, Cary, NC, USA). The analysis was performed on the per protocol (PP) population, a subset of subjects without any major protocol deviations affecting the PD evaluation defined prior to database lock and unblinding. Missing values were not imputed. Each leukocyte subset (raw absolute count and percentage change from baseline [defined as measurements collected on day -1]) was analyzed separately and summarized by treatment group and time-point measurements or by day (mean over 24 hours duration) using graphical displays and summary statistics (mean and standard deviation [SD]).

**Figure 1. Subject disposition.**

doi:10.1371/journal.pone.0058438.g001

Table 1. Subject baseline characteristics.

Characteristic	Placebo (n = 14)	CDP323 100 mg bid (n = 14)	CDP323 500 mg bid (n = 15)	CDP323 1000 mg qd (n = 14)	CDP323 1000 mg bid (n = 14)	Overall (N = 71)
Age, mean, y (range)	45.5 (25.5–60.1)	46.4 (36.3–62.3)	46.0 (27.4–59.0)	45.4 (24.8–59.7)	47.2 (41.3–53.9)	46.1 (24.8–62.3)
Female, n (%)	6 (42.9)	7 (50.0)	12 (80.0)	8 (57.1)	11 (78.6)	44 (62.0)
Caucasian, n (%)	13 (92.9)	14 (100)	13 (86.7)	12 (85.7)	14 (100)	66 (93.0)
BMI, mean (range)	27.7 (18.1–38.0)	28.4 (21.2–43.3)	28.1 (21.6–39.9)	27.9 (20.4–45.2)	28.5 (18.3–45.0)	28.1 (18.1–45.2)
Creatinine clearance, mean, mL/min (range) ^a	134.1 (79–212)	114.5 (79–227)	113.9 (86–192)	126.6 (78–222)	121.9 (75–231)	122.2 (75–231)
Type of MS, n (%)						
Relapsing remitting	11 (78.6)	10 (71.4)	11 (73.3)	8 (57.1)	10 (71.4)	50 (70.4)
Secondary progressive	3 (21.4)	4 (28.6)	4 (26.7)	6 (42.9)	4 (28.6)	21 (29.6)
EDSS						
Mean (SD)	5.5 (1.5)	4.3 (1.7)	4.9 (1.6)	4.8 (1.8)	5.2 (1.7)	4.9 (1.7)
Median (range)	6.0 (1.5–6.5)	4.8 (2.0–6.5)	6.0 (2.0–6.5)	5.8 (2.0–6.5)	6.0 (1.0–6.5)	6.0 (1.0–6.5)
Relapses in last 12 months						
Mean (SD)	1.6 (1.9)	1.1 (0.9)	1.1 (0.8)	1.6 (1.0)	1.1 (0.9)	1.3 (1.2)
Median (range) ^b	1.0 (0–8)	1.0 (0–3)	1.0 (0–3)	1.0 (0–3)	1.0 (0–3)	1.0 (0–8)

^aDetermined by Cockcroft formula: males = $[(140 - \text{age}) \times \text{body weight}] / [72 \times \text{serum creatinine (mg/dL)}]$; females = $[(140 - \text{age}) \times \text{body weight}] / [72 \times \text{serum creatinine (mg/dL)}] \times 0.85$.

^bAlthough 12 subjects had not experienced a relapse in the previous 12 months (4 in the 1000-mg bid group, 3 in each of the 100-mg bid and 500-mg bid groups, and 1 in each of the 1000-mg qd and placebo groups), they all fulfilled the inclusion criteria of having had at least one relapse in the previous 24 months.

bid = twice daily; BMI = body mass index; EDSS = Expanded Disability Status Scale; qd = once daily; SD = standard deviation.

doi:10.1371/journal.pone.0058438.t001

Table 2. Summary statistics on absolute counts of lymphocyte subsets at baseline, day 1, and day 28 after treatment.

Lymphocyte markers, mean (SD), 10 ³ cells/ μ L ^a	Day	Placebo (n = 14)	CDP323 100 mg bid (n = 14)	CDP323 500 mg bid (n = 14)	CDP323 1000 mg qd (n = 14)	CDP323 1000 mg bid (n = 14)
Total lymphocytes (CD45+)	Day -1 (baseline)	2.756 (1.246)	2.353 (0.587)	2.421 (0.619)	2.758 (0.779)	2.344 (0.449)
	Day 1	2.564 (0.841)	2.635 (0.597)	3.109 (0.754)	3.510 (0.985)	3.316 (0.897)
	Day 28	2.629 (0.812)	2.570 (0.656)	3.409 (0.981)	3.512 (1.155)	3.455 (0.802)
Naive B cells (CD19+/CD20+)	Day -1 (baseline)	0.444 (0.324)	0.291 (0.146)	0.319 (0.133)	0.313 (0.125)	0.285 (0.087)
	Day 1	0.422 (0.234)	0.354 (0.138)	0.448 (0.167)	0.478 (0.177)	0.496 (0.162)
	Day 28	0.415 (0.286)	0.347 (0.198)	0.551 (0.286)	0.522 (0.245)	0.557 (0.245)
Memory B cells (CD19+/CD20+/CD27+)	Day -1 (baseline)	0.097 (0.064)	0.067 (0.026)	0.086 (0.051)	0.092 (0.065)	0.083 (0.059)
	Day 1	0.088 (0.046)	0.089 (0.035)	0.149 (0.096)	0.147 (0.095)	0.160 (0.100)
	Day 28	0.090 (0.046)	0.083 (0.039)	0.190 (0.132)	0.182 (0.150)	0.194 (0.114)
T cells (CD3+)	Day -1 (baseline)	1.934 (0.872)	1.732 (0.411)	1.692 (0.524)	2.088 (0.618)	1.689 (0.332)
	Day 1	1.769 (0.621)	1.841 (0.434)	2.137 (0.629)	2.527 (0.798)	2.282 (0.654)
	Day 28	1.874 (0.553)	1.860 (0.407)	2.310 (0.665)	2.537 (0.841)	2.363 (0.494)
NK T cells (CD3+/CD56+)	Day -1 (baseline)	0.075 (0.068)	0.069 (0.053)	0.068 (0.074)	0.078 (0.063)	0.065 (0.039)
	Day 1	0.077 (0.077)	0.089 (0.095)	0.097 (0.103)	0.104 (0.092)	0.096 (0.069)
	Day 28	0.089 (0.076)	0.073 (0.062)	0.084 (0.081)	0.107 (0.108)	0.080 (0.051)
NK cells (CD3-/CD56+)	Day -1 (baseline)	0.208 (0.101)	0.197 (0.099)	0.248 (0.197)	0.215 (0.137)	0.200 (0.082)
	Day 1	0.222 (0.111)	0.290 (0.136)	0.327 (0.262)	0.286 (0.157)	0.314 (0.149)
	Day 28	0.195 (0.065)	0.229 (0.109)	0.333 (0.303)	0.272 (0.169)	0.300 (0.103)
Monocytes (CD14+)	Day -1 (baseline)	0.390 (0.128)	0.295 (0.092)	0.329 (0.147)	0.321 (0.071)	0.391 (0.137)
	Day 1	0.348 (0.157)	0.323 (0.100)	0.391 (0.176)	0.375 (0.114)	0.437 (0.166)
	Day 28	0.369 (0.060)	0.322 (0.103)	0.404 (0.203)	0.412 (0.237)	0.432 (0.131)
Hematopoietic progenitor cells (CD34+)	Day -1 (baseline)	0.003 (0.001)	0.003 (0.002)	0.003 (0.002)	0.003 (0.001)	0.003 (0.001)
	Day 1	0.003 (0.001)	0.004 (0.002)	0.006 (0.003)	0.006 (0.003)	0.006 (0.002)
	Day 28	0.003 (0.001)	0.004 (0.003)	0.007 (0.003)	0.005 (0.001)	0.006 (0.002)

^aMean value over 24 hours.

bid = twice a day; NK = natural killer; qd = once daily; SD = standard deviation.

doi:10.1371/journal.pone.0058438.t002

Analyses were conducted for the percentage change from baseline (time-matched day -1) observed at the end of treatment (day 28) in absolute lymphocyte and lymphocyte subset cell counts, VCAM-1 inhibition, and α_4 expression. These data were analyzed using a mixed-effect model repeated-measures analysis of variance (ANOVA; PROC MIXED in SAS) with the subject as the random effect, treatment and treatment by time-point interaction as fixed effects, and baseline values as a covariate. The treatment by time-point interaction was inspected. The dose-response relationship (using the dosages of 0, 200 mg/day, 1000 mg/day [as 1000 mg qd or 500 mg bid], or 2000 mg/day) was investigated by means of a linear trend test contrast [20] for unequally spaced doses in the SAS PROC MIXED model with a type-I error of 0.05. (Negative quadratic contrast was added to check for any plateau effect.)

In addition, five pairwise comparisons to the placebo group were evaluated (with the higher dosages emphasized: 1000 mg bid, 1000 mg qd, and 500 mg bid), and the 500-mg bid and 1000-mg qd regimens were also compared (to assess any potential regimen effect). These comparisons were applied using SAS contrasts of the PROC MIXED procedure. To take into account multiple comparisons (five pairwise comparisons), the type I error of 0.05 was adjusted by Bonferroni correction for these pairwise comparisons. Thus, differences were considered statistically significant when $P < 0.01$.

Ethics Statement

The study was approved by the London-Surrey Borders Research Ethics Committee and overseen by an independent safety advisory board (see Acknowledgments for board members). It was conducted in accordance with the ethical principles of the Declaration of Helsinki, the International Conference on Harmonisation Good Clinical Practice guidelines, and UK laws. Written consent was obtained from all subjects. The study was registered at the National Institutes of Health ClinicalTrials.gov website (identifier: NCT00726648).

Results

Subject Baseline Characteristics

A total of 106 subjects were screened, of whom 71 were randomized to treatment. The majority of subjects (97.2%; $n = 69$) completed the study; one subject in the CDP323 100-mg bid group was lost to follow-up and one subject in the CDP323 500-mg bid group discontinued because of a positive pregnancy test at day 27. The latter subject, who did not receive study drug on day 28, was the only exclusion from the PP population ($n = 70$). Subject disposition in each of the treatment groups is given in Figure 1.

Baseline demographics and disease characteristics are presented in Table 1. There were no clinically relevant differences between the treatment groups with respect to age, race, body mass index, or

Table 3. Percentage change from baseline in absolute lymphocyte counts at day 28 following treatment with CDP323 for 28 days (per protocol population).

Lymphocyte type	Treatment group ^a	Mean change from baseline, % ^b	Mean change vs placebo, % (99% CI) ^c	P value
Total lymphocytes (CD45+)	Linear trend test			<0.0001
	Quadratic trend test			0.0669
	Placebo	12.1	–	–
	CDP323 100 mg bid	8.1	–4.0 (–28.7, 20.6)	0.6678
	CDP323 500 mg bid	43.7	31.6 (6.9, 56.3)	0.0012
	CDP323 1000 mg qd	39.7	27.6 (2.9, 52.3)	0.0042
	CDP323 1000 mg bid	46.2	34.2 (9.5, 58.8)	0.0005
Naive B cells (CD19+/CD20+)	Linear trend test			<0.0001
	Quadratic trend test			0.7857
	Placebo	33.2	–	–
	CDP323 100 mg bid	13.0	–20.3 (–78.9, 38.4)	0.3637
	CDP323 500 mg bid	74.8	41.6 (–17.1, 100.3)	0.0650
	CDP323 1000 mg qd	69.4	36.1 (–22.8, 95.0)	0.1091
	CDP323 1000 mg bid	101.8	68.6 (9.9, 127.2)	0.0028
Memory B cells (CD19+/CD20+/CD27+)	Linear trend test			<0.0001
	Quadratic trend test			0.1080
	Placebo	31.6	–	–
	CDP323 100 mg bid	18.2	–13.4 (–87.1, 60.3)	0.6343
	CDP323 500 mg bid	138.0	106.4 (33.2, 179.7)	0.0002
	CDP323 1000 mg qd	126.3	94.7 (21.3, 168.2)	0.0010
	CDP323 1000 mg bid	166.8	135.2 (62.3, 208.1)	<0.0001
T cells (CD3+)	Linear trend test			0.0019
	Quadratic trend test			0.0542
	Placebo	15.5	–	–
	CDP323 100 mg bid	9.1	–6.4 (–33.1, 20.3)	0.5277
	CDP323 500 mg bid	41.4	25.9 (–0.9, 52.7)	0.0128
	CDP323 1000 mg qd	41.1	25.6 (–1.3, 52.4)	0.0140
	CDP323 1000 mg bid	38.4	22.9 (–3.8, 49.5)	0.0261
NK T cells (CD3+/CD56+)	Linear trend test			0.0900
	Quadratic trend test			0.0146
	Placebo	13.3	–	–
	CDP323 100 mg bid	14.8	1.5 (–31.3, 34.3)	0.9043
	CDP323 500 mg bid	50.1	36.8 (4.1, 69.5)	0.0039
	CDP323 1000 mg qd	36.6	23.2 (–10.0, 56.5)	0.0693
	CDP323 1000 mg bid	28.8	15.5 (–17.3, 48.3)	0.2171
NK cells (CD3+/CD56+)	Linear trend test			<0.0001
	Quadratic trend test			0.0836
	Placebo	5.5	–	–
	CDP323 100 mg bid	23.5	18.0 (–9.0, 45.1)	0.0836
	CDP323 500 mg bid	47.6	42.1 (14.9, 69.2)	<0.0001
	CDP323 1000 mg qd	40.5	35.0 (7.8, 62.2)	0.0010
	CDP323 1000 mg bid	57.7	52.2 (25.2, 79.2)	<0.0001
Monocytes (CD14+)	Linear trend test			0.0991
	Quadratic trend test			0.8544
	Placebo	21.5	–	–
	CDP323 100 mg bid	8.0	–13.5 (–54.3, 27.4)	NR
	CDP323 500 mg bid	27.6	6.1 (–34.6, 46.8)	NR
	CDP323 1000 mg qd	27.9	6.4 (–34.6, 47.3)	NR

Table 3. Cont.

Lymphocyte type	Treatment group ^a	Mean change from baseline, % ^b	Mean change vs placebo, % (99% CI) ^c	P value
Hematopoietic progenitor cells (CD34+)	CDP323 1000 mg bid	37.2	15.7 (–24.7, 56.1)	NR
	Linear trend test			0.0592
	Quadratic trend test			0.2727
	Placebo	18.6	–	–
	CDP323 100 mg bid	65.1	46.5 (–126.7, 219.6)	NR
	CDP323 500 mg bid	185.7	167.1 (–11.5, 345.6)	NR
	CDP323 1000 mg qd	75.4	56.8 (–110.6, 224.2)	NR
	CDP323 1000 mg bid	140.4	121.8 (–39.7, 283.2)	NR

^an = 14 per group except for CD34+ cells, where n = 5 for placebo and CDP323 1000 mg bid and n = 4 for CDP323 100 mg bid, CDP323 500 mg bid, and CDP323 1000 mg qd.

^bLeast squares mean from analysis of variance (ANOVA).

^cStatistical comparisons were made using univariate ANOVA.

bid = twice daily; CI = confidence interval; NK = natural killer; NR = not reported (linear trend test for dose-response relationship was not statistically significant); qd = once daily.

Note: For all lymphocyte types, differences between the 500-mg bid and 1000-mg qd dosages were not statistically significant and are not displayed in this table. doi:10.1371/journal.pone.0058438.t003

EDSS score. Overall, 62% of the subjects were female, although the percentage of female subjects in each group varied from 43% to 80%.

The majority of subjects had a diagnosis of relapsing-remitting MS; a higher percentage of subjects had a diagnosis of relapsing secondary progressive MS in the CDP323 1000-mg qd group (43%) than in the other groups (21% to 29%). The mean annualized relapse rate determined over the 12 months prior to the study was 1.3.

PD Effects of CDP323

Summary data for absolute counts of lymphocytes and lymphocyte subsets at baseline, day 1, and day 28 are presented in Table 2. Table 3 presents the percentage change from baseline, based on absolute counts, in total peripheral blood lymphocytes

and lymphocyte subsets following treatment with CDP323 for 28 days.

Dose-response analysis showed a statistically significant linear trend increase for total lymphocytes, naive B cells, memory B cells, T cells, and NK cells. For NK T cells, the significant negative quadratic trend highlighted a rise of the NK T cell count from 0 mg (placebo) to 1000 mg/day (500 mg bid and 1000 mg qd), corresponding to the daily dose at which maximum increase occurred (mean of 43.4%), followed by a decline (28.8%) at 2000 mg/day.

Mean increases in the total lymphocyte count of 43.7% and 46.2% were found with CDP323 dosages of 500 mg bid and 1000 mg bid, respectively. Smaller mean increases were seen with the 1000-mg qd (39.7%) and 100-mg bid (8.1%) dosages as well as in the placebo group (12.1%). Increases in total lymphocytes were

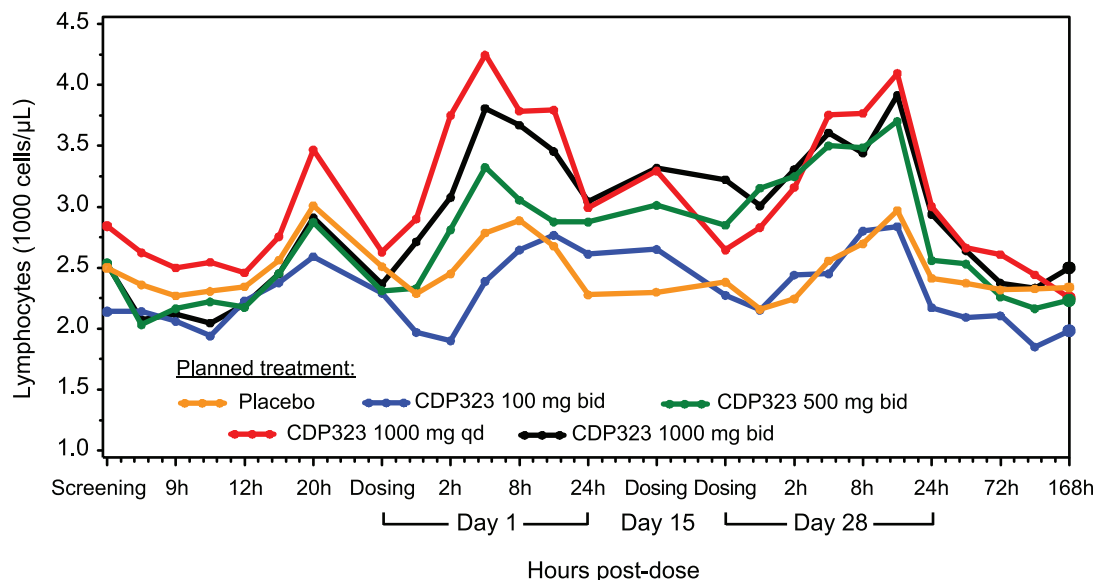


Figure 2. Geometric mean time profile for total lymphocyte count by administered treatment (per protocol population). bid = twice daily; qd = once daily.

doi:10.1371/journal.pone.0058438.g002

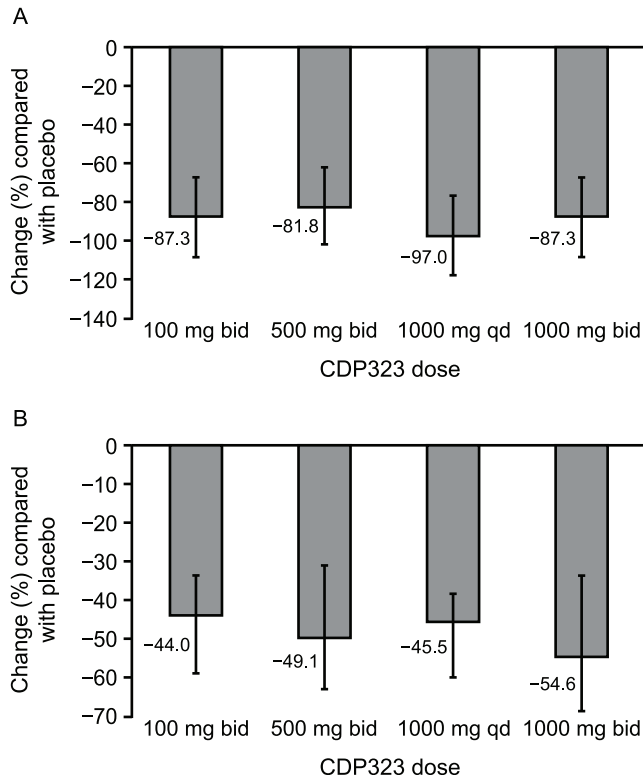


Figure 3. Mean change (95% CI) in (A) lymphocyte VCAM-1 binding and (B) VCAM-1-binding cell α_4 integrin expression following CDP323 administration. bid = twice daily; CI = confidence interval; qd = once daily; VCAM-1 = vascular adhesion molecule-1. doi:10.1371/journal.pone.0058438.g003

statistically significantly greater with all dosages of CDP323 except the lowest (100-mg bid) dosage than with placebo (Table 2). On day 28, the peak percentage of total lymphocytes was generally observed 8 to 12 hours following dosing of CDP323, and it returned to pretreatment baseline values approximately 24 hours after dosing.

Compared with placebo, the 500-mg bid, 1000-mg qd, and 1000-mg bid dosages of CDP323 generally resulted in statistically significant increases in naive B, memory B, and NK cells (Table 3). Increases in NK T cells observed with CDP323 compared with placebo reached statistical significance only with the 500-mg bid dosage. Although the mean percentage increase from baseline in hematopoietic progenitor cells was substantially greater following administration of CDP323 (65.1%–185.7%) compared with placebo (18.6%), the considerable intersubject variability occluded any statistically significant findings for the 100-mg bid and 1000-mg bid dosages. With the exception of the 100-mg bid dosage, CDP323 resulted in increases in monocytes compared with placebo, although none was statistically significant. Similarly, no statistically significant differences were observed for any of the lymphocyte subsets with CDP323 100 mg bid compared with placebo. Results for changes in T_{reg} cells and inflammatory T cells were inconclusive because the number of samples was insufficient for meaningful analysis.

In general, each of the PD parameters assessed returned to predose values within 24 to 96 hours after the last dose of CDP323 on day 28. Figure 2 shows the time profile of geometric mean values for total lymphocytes from all dosage groups.

Patients in the placebo group showed mean increases in lymphocyte binding to VCAM-1 of 41.7% and expression of α_4 -integrin on VCAM-1-binding cells of 33.6%, which may be the result of normal variations observed in lymphocyte counts [21]. In contrast, administration of CDP323 (at all dosages) caused statistically significant reductions compared with placebo in lymphocyte binding to VCAM-1 (mean reduction of 81.8%–97.0%; $P \leq 0.003$; Figure 3A) and in the expression of α_4 -integrin

Table 4. Summary of adverse events reported by ≥ 2 subjects in any treatment group (safety population).

Adverse event, n (%)	Placebo (n = 14)	CDP323 100 mg bid (n = 14)	CDP323 500 mg bid (n = 15)	CDP323 1000 mg qd (n = 14)	CDP323 1000 mg bid (n = 14)
Subjects with at least one AE	13 (92.9)	7 (50.0)	13 (86.7)	10 (71.4)	13 (92.9)
Headache	3 (21.4)	2 (14.3)	0	3 (21.4)	5 (35.7)
Fatigue	3 (21.4)	0	3 (20.0)	1 (7.1)	4 (28.6)
Diarrhea	3 (21.4)	0	2 (13.3)	1 (7.1)	0
Hypoesthesia	1 (7.1)	0	1 (6.7)	1 (7.1)	2 (14.3)
Muscle spasms	1 (7.1)	1 (7.1)	1 (6.7)	0	2 (14.3)
Vomiting	2 (14.3)	0	1 (6.7)	0	2 (14.3)
Back pain	0	0	1 (6.7)	2 (14.3)	1 (7.1)
Palpitations	0	3 (21.4)	1 (6.7)	0	0
Paresthesia	2 (14.3)	0	2 (13.3)	0	0
Muscular weakness	0	0	0	1 (7.1)	2 (14.3)
Dysphonia	0	0	0	2 (14.3)	0
Fall	0	0	0	0	2 (14.3)
Herpes simplex	0	0	2 (13.3)	0	0
Pain in extremity	0	0	2 (13.3)	0	0
Pharyngolarungeal pain	0	0	0	0	2 (14.3)

AE = adverse event; bid = twice daily; qd = once daily. doi:10.1371/journal.pone.0058438.t004

on VCAM-1-binding cells (mean reduction of 44.0%–54.6%; $P \leq 0.0001$; Figure 3B).

CDP323 was well tolerated in this study, with headache being the most frequently reported adverse event (AE) across the treatment groups (Table 4). The majority of AEs were mild or moderate in severity, and there were no deaths during the study. Four subjects reported a total of five serious AEs: hypoglycemia (placebo), unintended pregnancy and abortion induced (CDP323 500 mg bid), gamma-glutamyltransferase increase (CDP323 1000 mg bid), and hallucinations mixed (CDP323 1000 mg qd). Only the pregnancy event resulted in discontinuation from the study. Four subjects treated with CDP323 had at least one elevated liver function test of grade 3 or 4 according to the Common Terminology Criteria for Adverse Events (version 3; 9 August 2006). There were no consistent changes in any safety laboratory or vital signs parameters. No cases of opportunistic infection were identified during the study.

Discussion

This hybrid phase 1/2 study assessed the effect of different dosages of CDP323, a small-molecule α_4 -integrin antagonist, on changes in total lymphocytes and lymphocyte subsets in subjects with RMS. In general, the numbers of circulating T, B, NK, NK T, and hematopoietic progenitor cells were significantly greater in subjects who had received CDP323 dosages of 1000 mg qd (smallest increases), 500 mg bid, or 1000 mg bid (largest increases) than in those who had received placebo. In total lymphocytes and most subtypes, the dose-response relationship was shown to be linear.

The observed effect of CDP323 on the total peripheral blood lymphocyte count and the number of circulating mature and immature B cells was similar to the effects previously reported with the humanized monoclonal $\alpha_4\beta_1$ -integrin antibody natalizumab [22–24]. Unlike natalizumab, CDP323 had no significant effect on the number of circulating monocytes [23], nor on T_{reg} and inflammatory T-cell counts. However, the sample size for the T_{reg} and inflammatory T-cell results was too small to permit any definitive conclusions to be drawn.

The PD effects of CDP323 on lymphocytes and lymphocyte subsets are readily reversible, with cell numbers generally returning to predose levels within 96 hours after the last administered dose. However, given the short duration of this

study, it is not possible to predict treatment reversibility after long-term CDP323 administration.

Functional inactivation of the α_4 integrin has been observed following mobilization of CD34+ hematopoietic stem cells [25], and it is known that these cells are mobilized by natalizumab when it is used in the treatment of patients with MS [26–29]. CDP323 appears to have similar effects on these cells and was observed to significantly decrease the capacity of lymphocytes to bind VCAM-1 and α_4 integrin on VCAM-1-binding cells compared with placebo.

The differences in the effects of CDP323 and natalizumab on circulating lymphocytes demonstrate that despite targeting the same molecule, albeit via different delivery pathways, these compounds may induce distinct changes in leukocyte production and, possibly, functionality. Though the significance of these findings in relation to α_4 antagonism and changes in circulating lymphocyte levels remain unknown, the limited efficacy of CDP323 on brain lesions in a serial MRI phase 2 study [19] as compared with the substantial efficacy of natalizumab may indicate that the stable antagonism achieved by a long-acting antibody has clinical significance.

Supporting Information

Checklist S1 CONSORT Checklist.
(DOC)

Protocol S1 Trial Protocol.
(PDF)

Acknowledgments

The authors thank Ans Valgaeren and Bella Ertik (UCB S.A.) for their expert operational assistance in the analyses of the samples from this study and thank Ralph Bloomfield (UCB Celltech) for independent statistical advice to the safety Advisory Board. Medical writing support was provided by Mark Hughes and editorial support was provided by Joshua Safran, both of Infusion Communications. **Independent Safety Advisory Board:** Gavin Giovannoni (chair; London, UK), Tarek Youstry (London, UK), Mark Van Ranst (Leuven, Belgium).

Author Contributions

Conceived and designed the experiments: CW JS CO JT. Performed the experiments: JT. Analyzed the data: CW JS CO DLM JC JT BB. Wrote the paper: CW JS CO DLM JC JT BB.

References

- Stadelmann C, Wegner C, Brück W (2011) Inflammation, demyelination, and degeneration—recent insights from MS pathology. *Biochim Biophys Acta* 1812: 275–282.
- Ferguson TA, Mizutani H, Kupper TS (1991) Two integrin-binding peptides abrogate T cell-mediated immune responses in vivo. *Proc Natl Acad Sci U S A* 88: 8072–8076.
- Chan PY, Aruffo A (1993) VLA-4 integrin mediates lymphocyte migration on the inducible endothelial cell ligand VCAM-1 and the extracellular matrix ligand fibronectin. *J Biol Chem* 268: 24655–24664.
- D'hooghe MB, Nagels G, Bissay V, De Keyser J (2010) Modifiable factors influencing relapses and disability in multiple sclerosis. *Mult Scler* 16: 773–785.
- Müller WA (2011) Mechanisms of leukocyte transendothelial migration. *Annu Rev Pathol* 6: 323–344.
- Yednock TA, Cannon C, Fritz LC, Sanchez-Madrid F, Steinman L, et al. (1992) Prevention of experimental autoimmune encephalomyelitis by antibodies against alpha 4 beta 1 integrin. *Nature* 356: 63–66.
- Soilu-Hanninen M, Roytta M, Salmi A, Salonen R (1997) Therapy with antibody against leukocyte integrin VLA-4 (CD49d) is effective and safe in virus-facilitated experimental allergic encephalomyelitis. *J Neuroimmunol* 72: 95–105.
- Lee SJ, Benveniste EN (1999) Adhesion molecule expression and regulation on cells of the central nervous system. *J Neuroimmunol* 98: 77–88.
- Davenport RJ, Munday JR (2007) Alpha4-integrin antagonism—an effective approach for the treatment of inflammatory diseases? *Drug Discov Today* 12: 569–576.
- Polman CH, O'Connor PW, Havrdová E, Hutchinson M, Kappos L, et al. (2006) A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med* 354: 899–910.
- Kent SJ, Karlik SJ, Cannon C, Hines DK, Yednock TA, et al. (1995) A monoclonal antibody to alpha 4 integrin suppresses and reverses active experimental allergic encephalomyelitis. *J Neuroimmunol* 58: 1–10.
- Rudick RA, Sandrock A (2004) Natalizumab: alpha 4-integrin antagonist selective adhesion molecule inhibitors for MS. *Expert Rev Neurother* 4: 571–580.
- Radue EW, Stuart WH, Calabresi PA, Confavreux C, Galetta SL, et al. (2010) Natalizumab plus interferon beta-1a reduces lesion formation in relapsing multiple sclerosis. *J Neurol Sci* 292: 28–35.
- Muro F, Iimura S, Yoneda Y, Chiba J, Watanabe T, et al. (2008) Identification of 4-[1-[3-chloro-4-[N'-(5-fluoro-2-methylphenyl)ureido]phenylacetyl]-(4S)-fluoro-(2S)-pyrrolidinylmethoxy]benzoic acid as a potent, orally active VLA-4 antagonist. *Bioorg Med Chem* 16: 9991–10000.
- Kenyon NJ, Liu R, O'Roark EM, Huang W, Peng L, et al. (2009) An alpha4beta1 integrin antagonist decreases airway inflammation in ovalbumin-exposed mice. *Eur J Pharmacol* 603: 138–146.
- Vanderslice P, Woodside DG, Caivano AR, Decker ER, Munsch CL, et al. (2010) Potent in vivo suppression of inflammation by selectively targeting the high affinity conformation of integrin alpha4beta1. *Biochem Biophys Res Commun* 400: 619–624.

17. Baker M, Shock A, Parton T, Hales P, Parker G (2006) Pharmacokinetic and pharmacodynamic properties of the VLA-4 inhibitor CDP323. *Mult Scler* 12: S106.
18. Miller D, Khan O, Sheremata W, Blumhardt L, Rice G, et al. (2001) Results of a double-blind, randomized, placebo-controlled, phase II trial of Antegren (natalizumab) in subjects with relapsing multiple sclerosis (MS). *Mult Scler* 7: S16.
19. Polman C, Bowen J, Barkhof F, Bates D, Wynn D, et al. (2010) Double-blind, placebo-controlled, randomized phase II serial MRI, safety and tolerability study of two doses of CDP323 in subjects with relapsing forms of multiple sclerosis over 24 weeks. Presented at the 62nd Annual Meeting of the American Academy of Neurology, Toronto, Canada.
20. Kong L, Koch G, Liu T, Wang H (2005) Performance of some multiple testing procedures to compare three doses of a test drug and placebo. *Pharmaceut. Statist.* 4: 25–35.
21. Kovarik JM, Schmouder R, Barilla D, Riviere GJ, Wang Y, et al. (2004) Multiple-dose FTY720: tolerability, pharmacokinetics, and lymphocyte responses in healthy subjects. *J Clin Pharmacol* 44: 532–537.
22. Niino M, Bodner C, Simard ML, Alatab S, Gano D, et al. (2006) Natalizumab effects on immune cell responses in multiple sclerosis. *Ann Neurol* 59: 748–754.
23. Krumbholz M, Meinl I, Kumpfel T, Hohlfeld R, Meinl E (2008) Natalizumab disproportionately increases circulating pre-B and B cells in multiple sclerosis. *Neurology* 71: 1350–1354.
24. Putzki N, Baranwal MK, Tettenborn B, Limmroth V, Kreuzfelder E (2010) Effects of natalizumab on circulating B cells, T regulatory cells and natural killer cells. *Eur Neurol* 63: 311–317.
25. Lichterfeld M, Martin S, Burkly L, Haas R, Kronenwett R (2000) Mobilization of CD34+ haematopoietic stem cells is associated with a functional inactivation of the integrin very late antigen 4. *Br J Haematol* 110: 71–81.
26. Zohren F, Toutzaris D, Klarner V, Hartung HP, Kieseier B, et al. (2008) The monoclonal anti-VLA-4 antibody natalizumab mobilizes CD34+ hematopoietic progenitor cells in humans. *Blood* 111: 3893–3895.
27. Bonig H, Wundes A, Chang KH, Lucas S, Papayannopoulou T (2008) Increased numbers of circulating hematopoietic stem/progenitor cells are chronically maintained in patients treated with the CD49d blocking antibody natalizumab. *Blood* 111: 3439–3441.
28. Jing D, Oelschlaegel U, Ordemann R, Holig K, Ehninger G, et al. (2010) CD49d blockade by natalizumab in patients with multiple sclerosis affects steady-state hematopoiesis and mobilizes progenitors with a distinct phenotype and function. *Bone Marrow Transplant* 45: 1489–1496.
29. Neumann F, Zohren F, Haas R (2009) The role of natalizumab in hematopoietic stem cell mobilization. *Expert Opin Biol Ther* 9: 1099–1106.