

Draft Genome Sequence of the Bacteriophage vB_Eco_slurp01

Pavelas Sazinas,^a Carrie Smith,^b Aqilah Suhaimi,^b Jon L. Hobman,^c Christine E. R. Dodd,^c Andrew D. Millard^a

Microbiology and Infection Unit, Warwick Medical School, University of Warwick, Coventry, United Kingdom^a; School of Life Sciences, University of Warwick, Coventry, United Kingdom^b; School of Biosciences, University of Nottingham, Sutton Bonington Campus, Sutton Bonington, United Kingdom^c

Bacteriophage vB_Eco_slurp01 was isolated from porcine feces using *Escherichia coli* MG1655 as a host. With a genome size of 348 kb, vB_Eco_slurp01 is one of the largest bacteriophages isolated to date.

Received 22 August 2016 Accepted 29 September 2016 Published 17 November 2016

Citation Sazinas P, Smith C, Suhaimi A, Hobman JL, Dodd CER, Millard AD. 2016. Draft genome sequence of the bacteriophage vB_Eco_slurp01. *Genome Announc* 4(6):e01111-16. doi:10.1128/genomeA.01111-16.

Copyright © 2016 Sazinas et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Andrew D. Millard, a.d.millard@warwick.ac.uk.

There are approximately 4.4 million pigs in the United Kingdom, with an estimated £212 million export value (1). Yet there are relatively few studies on the virome of pigs. This study aimed to begin investigating the diversity of bacteriophages in porcine feces. Here, we isolated and sequenced the bacteriophage vB_Eco_slurp01, which is capable of infecting *Escherichia coli* MG1655.

Bacteriophage genomic DNA was prepared from cultures using a phenol:chloroform extraction method (2). One nanogram of genomic DNA was prepared using the Nextera XT DNA sample preparation kit (Illumina) prior to sequencing on the MiSeq platform using V2 (2 × 250-bp) chemistry. The resulting FASTQ files were assembled with SPAdes version 3-7 with the “-careful” option (3). The genome was sequenced to an average depth of 381×. The resultant single contig was annotated with Prokka version 1.11 using a custom database constructed from all complete viral genomes within the European Nucleotide Archive (4). ROARY was used to identify core genes between the phages vB_Eco_slurp01, PBECO4, and 121Q (5). Bacteriophage vB_Eco_slurp01 had a genome size of 348 kb with 588 coding sequences, seven tRNAs, and a GC content of 34.05%. There are currently only eight bacteriophage genomes that are larger than 300 kb in size; these infect a range of bacteria, including *Bacillus*, *Aureococcus*, *Cronobacter*, *Enterobacter*, *Escherichia*, and *Pseudomonas* spp. Thus, vB_Eco_slurp01 adds to this small number of “jumbo” phages. At the nucleotide level, vB_Eco_slurp01 is similar to the coliphages PBECO4 (KC295538.1) (6) and 121Q (KM507819.1) with an average nucleotide identity of 98.4% and 96.46%, respectively, across the genome. This high level of identity is surprising given that these phages were isolated from geographically distant regions: Gwacheon, South Korea (PBECO4), Quebec, Canada (121Q), and Nottinghamshire, United Kingdom (vB_Eco_slurp01).

Furthermore, these phages all share a high degree of synteny across their genomes. Comparison of gene content between the three strains revealed a core gene set of 405 genes. While conserved between isolates, the majority of these genes encode hypothetical proteins. As with previous jumbo phages, a number of bacterial host homologue genes were detected, including

genes coding for a σ^{54} modulation protein, RpoD, GyrA, GyrB, and ribonucleoside-diphosphate reductase subunits. Although homologous to *E. coli* genes, many of the coding sequences had greater similarity to genes from other bacteria; for example, *gyrB* had higher similarity to *Bacteriovorax* sp. DB6_IX than *E. coli*. In addition, a gene encoding a putative tellurite resistance protein (TelA) was also observed. This is a feature that is also common to the phages PBECO4 and 121Q. Intriguingly, the resistance protein is associated with Gram-positive bacteria (7) and is not part of the tellurite resistance operon found in *E. coli* (8).

The genome of vB_Eco_slurp01 provides further insights into the small number of bacteriophages that have genomes greater than 300 kb in size. Furthermore, this study demonstrates that double-stranded DNA bacteriophages from distant geographical regions have highly conserved genomes.

Accession number(s). The draft genome sequence of bacteriophage vB_Eco_slurp01 has been deposited in DDBJ/ENA/GenBank under the accession number [LT603033](https://www.ncbi.nlm.nih.gov/nuclink/MT603033).

FUNDING INFORMATION

Data analysis was carried out on CLIMB infrastructure, funded by the Medical Research Council (MRC) (MR/L015080/1).

REFERENCES

1. Department for Environment, Food and Rural Affairs. 2014. Livestock populations at 1 December 2013, United Kingdom. https://www.gov.uk/government/uploads/system/uploads/attachment_data/file/293717/structure-dec2013-uk-19mar14.pdf.
2. Rihtman B, Meaden S, Clokie MR, Koskella B, Millard AD. 2016. Assessing Illumina technology for the high-throughput sequencing of bacteriophage genomes. *PeerJ* 4:e2055. <http://dx.doi.org/10.7717/peerj.2055>.
3. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
4. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
5. Page AJ, Cummins CA, Hunt M, Wong VK, Reuter S, Holden MT,

- Fookes M, Falush D, Keane JA, Parkhill J. 2015. Roary: rapid large-scale prokaryote pan genome analysis. *Bioinformatics* 31:3691–3693. <http://dx.doi.org/10.1093/bioinformatics/btv421>.
6. Kim MS, Hong SS, Park K, Myung H. 2013. Genomic analysis of bacteriophage PBECO4 infecting *Escherichia coli* O157:H7. *Arch Virol* 158:2399–2403. <http://dx.doi.org/10.1007/s00705-013-1718-3>.
7. Collins B, Joyce S, Hill C, Cotter PD, Ross RP. 2010. TelA contributes to the innate resistance of *Listeria monocytogenes* to nisin and other cell wall-acting antibiotics. *Antimicrob Agents Chemother* 54:4658–4663. <http://dx.doi.org/10.1128/AAC.00290-10>.
8. Orth D, Grif K, Dierich MP, Würzner R. 2007. Variability in tellurite resistance and the *ter* gene cluster among Shiga toxin-producing *Escherichia coli* isolated from humans, animals and food. *Res Microbiol* 158:105–111. <http://dx.doi.org/10.1016/j.resmic.2006.10.007>.