

# Still a Host of Hosts for *Wolbachia*: Analysis of Recent Data Suggests That 40% of Terrestrial Arthropod Species Are Infected

# Roman Zug\*, Peter Hammerstein

Institute for Theoretical Biology, Humboldt-Universität zu Berlin, Berlin, Germany

#### **Abstract**

Wolbachia are intracellular bacteria that manipulate the reproduction of their arthropod hosts in remarkable ways. They are predominantly transmitted vertically from mother to offspring but also occasionally horizontally between species. In doing so, they infect a huge range of arthropod species worldwide. Recently, a statistical analysis estimated the infection frequency of Wolbachia among arthropod hosts to be 66%. At the same time, the authors of this analysis highlighted some weaknesses of the underlying data and concluded that in order to improve the estimate, a larger number of individuals per species should be assayed and species be chosen more randomly. Here we apply the statistical approach to a more appropriate data set from a recent survey that tested both a broad range of species and a sufficient number of individuals per species. Indeed, we find a substantially different infection frequency: We now estimate the proportion of Wolbachia-infected species to be around 40% which is lower than the previous estimate but still points to a surprisingly high number of arthropods harboring the bacteria. Notwithstanding this difference, we confirm the previous result that, within a given species, typically most or only a few individuals are infected. Moreover, we extend our analysis to include several reproductive parasites other than Wolbachia that were also screened for in the aforementioned empirical survey. For these symbionts we find a large variation in estimated infection frequencies and corroborate the finding that Wolbachia are the most abundant endosymbionts among arthropod species.

Citation: Zug R, Hammerstein P (2012) Still a Host of Hosts for Wolbachia: Analysis of Recent Data Suggests That 40% of Terrestrial Arthropod Species Are Infected. PLoS ONE 7(6): e38544. doi:10.1371/journal.pone.0038544

Editor: Richard Cordaux, University of Poitiers, France

Received February 16, 2012; Accepted May 7, 2012; Published June 7, 2012

**Copyright:** © 2012 Zug, Hammerstein. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was funded by the Deutsche Forschungsgemeinschaft (http://www.dfg.de/en/index.jsp), SFB 618: "Theoretical Biology: Robustness, Modularity and Evolutionary Design of Living Systems." The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

\* E-mail: roman.zug@biologie.hu-berlin.de

### Introduction

Wolbachia are the predominant bacterial endosymbionts of arthropods, infecting a vast number of host species worldwide [1]. Both the proportion of infected individuals within species (prevalence) and the overall percentage of infected species (incidence) are important parameters describing the infection frequency of Wolbachia. In order to estimate these parameters, Hilgenboecker et al. [2] recently presented a meta-analysis that combined the data from 20 Wolbachia screenings with more than 900 arthropod species in total. Using a statistical approach, i.e. a beta-binomial model, they found that prevalences are typically very low or very high, and estimated the incidence of Wolbachia to be around 66%, which is considerably higher than previous estimates of approximately 20% [3,4]. A major reason for such underestimation is the sampling of only one or a few individuals per species. With these one-individual samples, low (and even high) prevalence infections are likely to be overlooked. On the other hand, Hilgenboecker et al. [2] found that samples comprising more than 100 individuals per species tend to be biased towards infected species, e.g. due to prior knowledge of infection. Although they corrected for the latter bias by excluding particularly large samples, many studies used in their metaanalysis still included quite a lot of one-individual samples and

were restricted to specific host taxa (see [2] for details). Therefore, in order to more accurately assess the incidence of *Wolbachia* in arthropod hosts, it is crucial to analyze a data set that comprises a medium number of individuals from randomly chosen species. Here, we apply the approach by Hilgenboecker et al. [2] to data from a recent survey by Duron et al. [5] that meets these requirements more closely. This survey also tested for the presence of several reproductive parasites, which allows us to estimate incidences of other endosymbionts and compare them to that of *Wolbachia*.

# Methods

1

In the survey by Duron et al. [5], 136 species of terrestrial arthropods (2052 individuals in total) were screened for the presence of seven reproductive parasites: Wolbachia, Arsenophonus, Cardinium, Flavobacterium, Rickettsia, Spiroplasma ixodetis and S. poulsonii. Since Flavobacterium was never observed, we excluded it from our analysis. In the survey, not more than 40 individuals were sampled per species, and in only 25 of the 136 species tested, less than 10 individuals were sampled (median: 15 individuals per species; mode: 20 individuals per species). This range of sampled individuals should help to avoid the drawbacks of both one-individual samples and the bias associated with extensive sampling.

Arthropod species tested encompassed 15 orders and three classes (Insecta, Arachnida, Malacostraca), thus representing a wide-spread and sufficiently random collection. Taken together, the data from Duron et al. [5] should satisfy the requirements for an improved data set as outlined above.

We again use the framework of a beta-binomial model to estimate symbiont prevalence q and incidence x. Different species are assumed to exhibit different prevalences, and thus q values follow a probability distribution p(q). The incidence x is then estimated by integrating the prevalence distribution:

$$x = \int_{c}^{1} p(q) dq,$$

where  $\epsilon$  defines a threshold frequency below which species are considered to be uninfected. For a more detailed account of the model, see [2].

#### **Results and Discussion**

The prevalence distribution for *Wolbachia* shows that either most or only few individuals within a species are infected (Figure 1). Based on this distribution, *Wolbachia* incidence is estimated to be x=0.406 for c=0.001 (Table 1). We chose c=0.001 in accordance with Hilgenboecker et al. [2] to facilitate comparisons. Our results confirm the main qualitative findings from the previous meta-analysis, i.e. the 'most-or-few' prevalence pattern and the likely underestimation of incidence in previous *Wolbachia* screenings. However, there is one major difference between the results of the two analyses: In the first study, *Wolbachia* incidence was estimated to be 66% (for c=0.001). Based on the data from Duron et al. [5], we now obtain a lower estimate of the percentage of *Wolbachia*-infected species, i.e. approximately 40%. We think that our current estimate is more reasonable for the following three reasons.

First, the underlying data contain only a very low proportion of species samples in which only a few individuals were tested. Testing only a small number of individuals considerably increases the likelihood of randomly picking some uninfected individuals

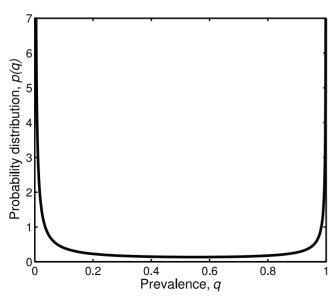


Figure 1. Estimated probability distribution of *Wolbachia* prevalence.

doi:10.1371/iournal.pone.0038544.g001

from an actually infected species, particularly if prevalence levels are low. Indeed, there is evidence that infection frequencies within species are often variable between geographically distinct populations. Such a prevalence variation between populations was found in several species-specific surveys, ranging from 0% to 100% in the cherry fruit fly or from 4% to 100% in two planthoppers [6,7]. In another fruit fly screening that tested 1500 individuals, only extremely low prevalence levels were found among different populations, ranging from 0% to 3% [8]. Moreover, species might also be falsely classified as uninfected because of low-titer infections that are not detected. Recent evidence suggests that such low-titer Wolbachia infections within arthropod hosts are more common than previously thought [7,9]. Taken together, sampling more than just a few individuals – as it was predominantly done by Duron et al. [5] - avoids the pitfalls outlined above and thus significantly improves estimates of Wolbachia infection frequencies in nature.

A second reason why we think our current estimate is more accurate is that the new data set does not include large samples (not more than 40 individuals per species). Large samples are likely to be biased towards infection, probably because respective species were already known to be infected and were sampled extensively to study infection prevalence in more detail. Additionally, large samples will disproportionately often be samples of common species, just because common species are more easily collected in large amounts ([2]; cf. the collecting procedure in [10]). Common species, however, are again prone to have already been tested for infection. These are important issues because large samples inherently have a strong impact on the estimation procedure. Therefore, as was already pointed out by Hilgenboecker et al. [2], omission of large samples will improve estimates of *Wolbachia* incidence.

Thirdly, the fact that Duron et al. [5] sampled a wide range of arthropod species from 15 different orders should render this collection sufficiently diverse in phylogenetic terms. In contrast to the previous meta-analysis which pooled the results from many taxon-specific screenings, analyzing a broad taxon survey ensures that all species are examined with the same detection method. Usually, Wolbachia infections are detected by PCR assays which crucially depend on the sensitivity of commonly used PCR primers. A recent assessment of standard PCR protocols used for Wolbachia detection, however, reveals considerable variation in primer efficiency [11]. To summarize, the data set compiled by Duron et al. [5] is the first one that satisfies our criteria for a reliable estimate (no one-individual samples, no large samples, no restriction to a specific host taxon). In contrast, several Wolbachia screenings that have since been published fail to satisfy the criteria and are not included in our analysis (see, for example, [12–16]). We therefore think that our estimate of Wolbachia incidence within arthropods is more reliable than previous attempts.

Another reason for the difference in incidence estimates between our study and that by Hilgenboecker et al. [2] might be the different sampling range. As described above, infection frequencies within species can differ greatly between geographically distinct populations. Duron et al. [5] pointed out that such geographical variation in prevalence was likely to increase when expanding the sampling range beyond Western Europe, where species were predominantly collected. In contrast, the meta-analysis by Hilgenboecker et al. [2] comprises samples from most continents, covering both temperate and tropical zones. Although speculative, this unequal geographical sampling might partially explain the difference in incidence estimates derived from both data sets.

**Table 1**. Estimates of incidence x of different endosymbionts, depending on threshold infection frequency c.

Threshold frequency c	Incidence x					
	Wolbachia	Arsenophonus	Cardinium	Rickettsia	Spiroplasma ixodetis	S. poulsonii
0.01	0.335	0.059	0.111	0.014	0.148	0.022
0.001	0.406	0.066	0.162	0.061	0.221	0.032
0.0001	0.470	0.072	0.211	0.114	0.289	0.041

The parameter c is the infection frequency above which species are considered infected. doi:10.1371/journal.pone.0038544.t001

Previous broad taxon surveys of Wolbachia infection frequencies among arthropods found approximately 20% of the tested species to be infected [3,4]. In general, previous surveys have estimated incidence by dividing the number of infected species by the overall number of species tested. Adopting the same straightforward approach to the data by Duron et al. [5] yields a very similar estimate (22.8%). However, this is roughly only half of our 40% estimate, although based on the same data set. Therefore, the proportion of Wolbachia-infected species seems to be considerably higher than a first glance would suggest.

In order to compare the infection frequency of Wolbachia to that of other reproductive parasites, we estimate the incidence for five endosymbionts that were also included in the survey by Duron et al. [5]. Since these symbionts were detected only in very few species, the graphic representation of the prevalence distributions is of limited value and therefore not displayed here. Incidence levels range from 0.032 (Spiroplasma poulsonii) to 0.221 (Spiroplasma *ixodetis*; Table 1, all values for c = 0.001). Again, our estimates are consistently higher than those obtained by the straightforward

- Werren JH, Baldo L, Clark ME (2008) Wolbachia: master manipulators of invertebrate biology. Nat Rev Microbiol 6: 741-751.
- Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren IH (2008) How many species are infected with Wolbachia? - a statistical analysis of current data. FEMS Microbiol Lett 281: 215-220.
- Werren JH, Windsor D, Guo L (1995) Distribution of Wolbachia among neotropical arthropods. Proc R Soc Lond B 262: 197-204.
- Werren JH, Windsor DM (2000) Wolbachia infection frequencies in insects: evidence of a global equilibrium? Proc R Soc Lond B 267: 1277-1285.
- Duron O, Bouchon D, Boutin S, Bellamy L, Zhou L, et al. (2008) The diversity of reproductive parasites among arthropods: Wolbachia do not walk alone. BMC Biology 6: 27.
- Arthofer W, Riegler M, Schneider D, Krammer M, Miller WJ, et al. (2009) Hidden Wolbachia diversity in field populations of the European cherry fruit fly, Rhagoletis cerasi (Diptera, Tephritidae). Mol Ecol 18: 3816-3830
- 7. Hughes GL, Allsopp PG, Brumbley SM, Woolfit M, McGraw EA, et al. (2011) Variable infection frequency and high diversity of multiple strains of Wolbachia pipientis in Perkinsiella planthoppers. Appl Environ Microbiol 77: 2165-2168.
- Sun X, Cui L, Li Z (2007) Diversity and phylogeny of Wolbachia infecting Bactrocera dorsalis (Diptera: Tephritidae) populations from China. Environ Entomol 36: 1283-1289.
- Arthofer W, Riegler M, Avtzis DN, Stauffer C (2009) Evidence for low-titre infections in insect symbiosis: Wolbachia in the bark beetle Pityogenes chalcographus (Coleoptera, Scolytinae). Environ Microbiol 11: 1923-1933.

approach (see [5]). Yet, even S. ixodetis as the most common of these other symbionts does not match Wolbachia in terms of incidence, which corroborates the status of Wolbachia as the most abundant reproductive parasite of arthropod hosts. Considering the species richness of the global arthropod community [17], our estimate implies that more than a million species are infected with Wolbachia. Thus, although lower than estimated by Hilgenboecker et al. [2], the number of species harboring Wolbachia is still remarkably high and justifies further efforts to investigate interactions between these endosymbionts and their arthropod

# Acknowledgments

We are grateful to John H. Werren, Kirsten Hilgenböcker, and two anonymous reviewers for helpful comments on the manuscript.

## **Author Contributions**

Analyzed the data: RZ. Wrote the paper: RZ PH.

- 10. Weinert LA, Tinsley MC, Temperley M, Jiggins FM (2007) Are we underestimating the diversity and incidence of insect bacterial symbionts? A case study in ladybird beetles. Biol Lett 3: 678-681.
- 11. Simões PM, Mialdea G, Reiss D, Sagot M-F, Charlat S (2011) Wolbachia detection: an assessment of standard PCR protocols. Mol Ecol Resour 11: 567-579
- 12. Wiwatanaratanabutr I, Kittayapong P, Caubet Y, Bouchon D (2009) Molecular phylogeny of Wolbachia strains in arthropod hosts based on groE-homologous rene sequences, Zool Sci 26: 171–177.
- 13. Kondo NI, Tuda M, Toquenaga Y, Lan Y-C, Buranapanichpan S, et al. (2011) Wolbachia infections in world populations of bean beetles (Coleoptera: Chrysomelidae: Bruchinae) infesting cultivated and wild legumes. Zool Sci 28: 501 - 508
- 14. Yun Y, Peng Y, Liu FX, Lei C (2011) Wolbachia screening in spiders and assessment of horizontal transmission between predator and prey. Neotrop Entomol 40: 164-169.
- Doudoumis V. Tsiamis G. Wamwiri F. Brelsfoard C. Alam U. et al. (2012) Detection and characterization of Wolbachia infections in laboratory and natural populations of different species of tsetse flies (genus Glossina). BMC Microbiol
- Evison SEF, Roberts KE, Laurenson L, Pietravalle S, Hui J, et al. (2012) Pervasiveness of parasites in pollinators. PLoS ONE 7: e30641
- 17. Hamilton AJ, Basset Y, Benke KK, Grimbacher PS, Miller SE, et al. (2010) Quantifying uncertainty in estimation of tropical arthropod species richness. Am Nat 176: 90-95.

References