



# Article Dietary-Derived Essential Nutrients and Amyotrophic Lateral Sclerosis: A Two-Sample Mendelian Randomization Study

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Abstract: Previous studies have suggested a close but inconsistent relationship between essential nutrients and the risk of amyotrophic lateral sclerosis (ALS), and whether this association is causal remains unknown. We aimed to investigate the potential causal relation between essential nutrients (essential amino acids, essential fatty acids, essential minerals, and essential vitamins) and the risk of ALS using Mendelian randomization (MR) analysis. Large-scale European-based genomewide association studies' (GWASs) summary data related to ALS (assembling 27,205 ALS patients and 110,881 controls) and essential nutrient concentrations were separately obtained. MR analysis was performed using the inverse variance-weighted (IVW) method, and sensitivity analysis was conducted by the weighted median method, simple median method, MR-Egger method and MR-PRESSO method. We found a causal association between genetically predicted linoleic acid (LA) and the risk of ALS (OR: 1.066; 95% CI: 1.011–1.125; p = 0.019). An inverse association with ALS risk was noted for vitamin D (OR: 0.899; 95% CI: 0.819–0.987; p = 0.025) and for vitamin E (OR: 0.461; 95% CI: 0.340–0.626;  $p = 6.25 \times 10^{-7}$ ). The sensitivity analyses illustrated similar trends. No causal effect was observed between essential amino acids and minerals on ALS. Our study profiled the effects of diet-derived circulating nutrients on the risk of ALS and demonstrated that vitamin D and vitamin E are protective against the risk of ALS, and LA is a suggested risk factor for ALS.

Keywords: amyotrophic lateral sclerosis; nutrition; Mendelian randomization; genes; risk factor

## 1. Introduction

Amyotrophic lateral sclerosis (ALS) is a fatal neurodegenerative disease associated with muscle weakness and atrophy, and patients usually die as a result of respiratory failure within 3–5 years of symptom onset [1,2]. No treatment is currently effective in reversing the progression of ALS, with the exception of riluzole, which probably prolongs median survival by 2–3 months [3]. The etiology of ALS is complex, largely unknown, and involves genetic and environmental factors [4]. Given the accumulated studies investigating risk factors for ALS, the implication of such findings has been a focus. Targeted lifestyle and dietary changes have been shown to be beneficial in the early stages of ALS [5,6].

Essential nutrients accessible from the diet include essential amino acids, essential fatty acids, vitamins, and minerals. Supplementing essential nutrients is considered a promising preventive intervention for many diseases due to its ability to maintain metabolic homeostasis and to mitigate oxidative stress [7–9]. With efforts made to delineate the influence



Citation: Xia, K.; Wang, Y.; Zhang, L.; Tang, L.; Zhang, G.; Huang, T.; Huang, N.; Fan, D. Dietary-Derived Essential Nutrients and Amyotrophic Lateral Sclerosis: A Two-Sample Mendelian Randomization Study. *Nutrients* **2022**, *14*, 920. https:// doi.org/10.3390/nu14050920

Academic Editor: Susanna C. Larsson

Received: 30 January 2022 Accepted: 18 February 2022 Published: 22 February 2022

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**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). of essential nutrients on ALS, different conclusions have been drawn. For instance, a large, pooled prospective study reported that long-term vitamin E supplementation was inversely associated with the risk of ALS [10], while null results were found in two other case–control studies [11,12]. Similarly, controversial results were observed for docosahexaenoic acid (DHA) [13,14], iron [15,16], vitamin A, and vitamin C [11,17]. This inconsistency is largely because most of this evidence came from observational studies, which are plagued by selection bias and unmeasured confounders. In addition, we previously found that most ALS patients showed a loss of appetite [18] and the presence of metabolic disorders [19], which may affect the absorption and consumption of dietary nutrients, making the true relationship between essential nutrients and ALS elusive. Randomized controlled trials (RCTs) are recognized to overcome the limitations of observational studies and provide the highest level of evidence [20]. However, no previous RCT has been conducted to identify the risk/protective effect of essential nutrients on ALS.

Mendelian randomization (MR) analysis is a genetic statistical approach to estimate the causal association between an exposure and an outcome using single nucleotide polymorphisms as instrumental variables (IVs), which mimic the random assignment to groups of RCTs by naturally allocating alleles and can lessen residual confounding [21].

In this study, we performed a two-sample MR approach to systematically decipher the causal association between nutrition status and ALS.

#### 2. Materials and Methods

#### 2.1. Data Collection and Selection of Instrumental Variables (IVs)

Essential amino acids, essential fatty acids, essential minerals, and essential vitamins were considered essential nutrients. We searched PubMed to extract identified exposure-related SNPs from the most recent and largest genome-wide association studies (GWASs) based on the European population.

SNPs related to amino acids were extracted from seven datasets, namely, isoleucine, leucine, lysine, methionine, phenylalanine, tryptophan, and valine [22,23]. Arachidonic acid (AA), alpha-linolenic acid (ALA), DHA, docosapentaenoic acid (DPA), dihomogamma-linolenic acid (DGLA), eicosapentaenoic acid (EPA), gamma linolenic acid (GLA), and linoleic acid (LA) [22,24,25] were included in the essential fatty acid category. For the essential minerals, GWASs for calcium [26], copper, iron [27], magnesium [28], phosphorus [29], selenium [30], and zinc [31] were searched separately. In addition, we sought out updated calcium GWAS data in a preprint paper based on the UK Biobank with a fivefold enlarged sample size [32], and these data were recorded as a supplement to calcium, named "Calcium (UKB)". To comprehensively assess the effects of vitamins and provitamins on disease, we examined both the absolute levels of vitamins or provitamins in blood and the relative concentrations of corresponding circulating metabolites quantified by a high-throughput platform. For absolute vitamin or provitamin levels, absolute vitamin A (retinol) [33], absolute beta-carotene [34], absolute vitamin B (vitamin B-6 and vitamin B-12) [35,36], absolute vitamin C (ascorbic acid) [37], absolute vitamin D (25-hydroxyvitamin D (25OHD)) [38], and absolute vitamin E (alpha-tocopherol) [39] were included, and for the relative concentrations of corresponding metabolites, relative retinol [40], relative vitamin C (ascorbate) [23], and relative vitamin E (alpha-tocopherol and gamma-tocopherol) [23] were used.

We obtained large-scale European-based ALS GWAS summary data from a recently published study, which included 27,205 ALS patients and 110,881 controls [41]. All patients were diagnosed according to the (revised) El Escorial criteria by specialized neurologists.

We set strict selection criteria for eligible IVs: (1) independent SNPs ( $r^2 = 0.01$ , KB = 5000) with genome-wide association levels ( $p < 5 \times 10^{-8}$ ) for exposure events; for exposures without genome-wide association significant IVs, suggestive level genome-wide association significant ( $p < 1 \times 10^{-5}$ ) IVs were employed as proxies; (2) nonrare SNPs (MAF  $\geq 0.01$ ); and (3) proxies in strong linkage disequilibrium ( $r^2 > 0.8$ ) were employed to replace SNPs that failed to provide corresponding information for ALS. Proxy loci not

included in the ALS dataset were excluded. The LDlink online tool [42] was employed in searching proxies. In total, we derived 1–168 genetic instruments for 35 essential nutritional biomarkers. Due to methodology limitations, exposures with at least two IVs were ultimately included in the downstream analysis. Lysine, methionine, ALA, absolute lycopene, absolute beta-carotene, and absolute vitamin B-6 were removed.

To measure the phenotypic variation explained by each SNP,  $R^2 = 2 \times beta^2 \times (1 - EAF) \times EAF/SD^2$ , where EAF = effect allele frequency and beta = effect of each SNP on exposures [43]. We similarly calculated F statistics for individual SNPs to indicate the strength of IVs (F =  $R^2 \times (N - 2)/((1 - R^2))$ ) [44]. Detailed information on SNPs is displayed in Supplementary Table S1.

### 2.2. Mendelian Randomization

Aiming to ensure the validity of the IVs, heterogeneity and directional pleiotropy were tested using Cochran's Q test and the MR–PRESSO method. For Cochran's Q test, nonsignificant heterogeneity (p > 0.05) was indicated by analogical estimates for each IV. For the MR–PRESSO method [45], (1) the overall horizontal pleiotropy was tested with the "global test"; (2) the "MR–PRESSO outlier test" was adopted to remove possible outliers in IVs and to generate corrected results; and (3) the "MR–PRESSO distortion test" was used to determine whether there was distortion between pre- and post-correction results. Similarly, the MR–Egger regression intercept was calculated as an additional means to examine horizontal pleiotropy. When there was no obvious distance from the intercept to the origin, it was regarded as having no influence on pleiotropy.

In this MR framework, the multiplicative random effect inverse variance–weighted method (IVW) was utilized as the main approach to analyze causality, which regresses the effect of SNPs on the outcome by the effect of the SNP on the exposure [46]. To ensure the robustness of the IVW results, additional sensitivity analyses were performed, including the simple median method, weighted median method and MR–Egger method. The weighted median method enables satisfactory estimates when at least 50% of the IVs are valid instruments [47]. The MR–Egger method can model terms to detect and correct the bias caused by pleiotropy, obtaining a consistent estimate of the causal effect [48]. We also implemented the "leave-one-out" (LOO) method to assess the influence of an individual SNP on the overall causal findings by excluding each genetic variant in turn and recomputing the MR–IVW estimates. The MR–Steiger method was employed to determine whether the direction of causality was true. Our flow chart was visualized in Figure 1.



**Figure 1.** Flow chart showing the process for the Mendelian randomization analyses. The number in the line indicates 3 key assumptions for MR. Assumption 1: The selected genetic variants are not related to other confounders. Assumption 2: The selected genetic variants are significantly related to the exposure. Assumption 3: The selected genetic variants are significantly related to the risk of the outcome only through the pathway from exposure. IVs, instrumental variables; GWASs, genome-wide association studies; MR, Mendelian randomization; IVW, inverse variance–weighted; MR–PRESSO, Mendelian randomization pleiotropy residual sum and outlier.

*p* values were adjusted according to the strict Bonferroni correction. *p* < 0.05/k was considered a significant level (where k is the number of exposures), and a *p* value between 0.05/k and 0.05 was suggestive of a positive association. All analyses were performed in R software version 4.1.0 by the "TwoSampleMR" package (version 0.5.6) [49] and the "MR–PRESSO" package (version 1.0) [45].

## 3. Results

The 29 dietary-derived potential risk factors for ALS that we evaluated involved five traits related to amino acids, seven to fatty acids, eight to minerals, and nine to vitamins, including 2–168 SNPs as genetic proxies for each of them. All associations analyzed by IVW random-effects models are shown in Table 1.

Table 1. Summary of the causal effects of each trait on ALS via different MR methods.

Exposures	Methods	No. SNPs	OR (95%CI)	p Value			
Essential amino acids							
Isoleucine	IVW	2	1.003 (0.666, 1.510)	0.988			
Leucine	IVW	3	0.961 (0.770, 1.198)	0.723			
Leucine	MR–Egger	3	0.370 (0.131, 1.045)	0.312			
Leucine	Simple median	3	0.927 (0.708, 1.214)	0.583			
Leucine	Weighted median	3	0.899 (0.745, 1.084)	0.264			
Phenylalanine	IVW	3	1.035 (0.800, 1.340)	0.792			
Phenylalanine	MR-Egger	3	0.352 (0.045, 2.722)	0.500			
Phenylalanine	Simple median	3	1.085 (0.844, 1.395)	0.523			
Phenylalanine	Weighted median	3	1.111 (0.887, 1.392)	0.359			
Tryptophan	IVW	20	1.848 (0.383, 8.916)	0.444			
Tryptophan	MR-Egger	20	609.49 (0, 534, 720, 778, 694.025)	0.549			
Tryptophan	Simple median	20	1.096 (0.248, 4.851)	0.904			
Tryptophan	Weighted median	20	1.358 (0.302, 6.117)	0.690			
Valine	IVW	5	0.949(0.840, 1.071)	0.397			
Valine	MR-Egger	5	0.688(0.340, 1.391)	0.374			
Valine	Simple median	5	0.94 (0.790, 1.120)	0.492			
Valine	Weighted median	5	0.932(0.791, 1.099)	0.404			
	Essential polyu	nsaturated fatty	acids				
Arachidonic acid (AA)	IVW	2	0.995 (0.984, 1.007)	0.415			
Dihomo-gamma-linolenic	17/14/	2	1 031 (1 017 1 045)	$1.70 \times 10^{-5}$			
acid (DGLA)	1 V VV	2	1.051 (1.017, 1.045)	$1.70 \times 10$			
Docosahexaenoic acid (DHA)	IVW	5	1.048 (0.924, 1.190)	0.464			
Docosahexaenoic acid (DHA)	MR–Egger	5	1.013 (0.427, 2.403)	0.978			
Docosahexaenoic acid (DHA)	Simple median	5	1.147 (0.971, 1.355)	0.106			
Docosahexaenoic acid (DHA)	Weighted median	5	1.008 (0.889, 1.143)	0.902			
Docosapentaenoic acid (DPA)	IVW	3	0.874 (0.607, 1.257)	0.466			
Docosapentaenoic acid (DPA)	MR–Egger	3	0.660 (0.276, 1.576)	0.521			
Docosapentaenoic acid (DPA)	Simple median	3	0.885 (0.635, 1.233)	0.469			
Docosapentaenoic acid (DPA)	Weighted median	3	0.840 (0.645, 1.095)	0.198			
Eicosapentaenoic acid (EPA)	IVW	2	0.872 (0.647, 1.175)	0.367			
Gamma linolenic acid (GLA)	IVW	2	0.724 (0.184, 2.844)	0.643			
Linoleic acid (LA)	IVW	18	1.066 (1.011, 1.125)	0.019			
Linoleic acid (LA)	MR–Egger	18	1.162 (1.029, 1.312)	0.028			
Linoleic acid (LA)	Simple median	18	1.047 (0.972, 1.128)	0.226			
Linoleic acid (LA)	Weighted median	18	1.049 (0.976, 1.128)	0.196			
	Ū N	linerals					
Calcium	IVW	7	1.088 (0.775, 1.527)	0.626			
Calcium	MR–Egger	7	0.928 (0.484, 1.781)	0.831			
Calcium	Simple median	7	1.04 (0.582, 1.856)	0.895			
Calcium	Weighted median	7	0.98 (0.668, 1.437)	0.918			
Calcium (UKB)	ĬVW	168	1.027 (0.952, 1.107)	0.495			
Calcium (UKB)	MR–Egger	168	1.011 (0.868, 1.178)	0.883			
Calcium (UKB)	Simple median	168	1.009 (0.891, 1.143)	0.887			
Calcium (UKB)	Weighted median	168	0.989 (0.858, 1.139)	0.878			

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Exposures	Methods	No. SNPs	OR (95%CI)	<i>p</i> Value
Copper	IVW	2	1.007 (0.940, 1.079)	0.837
Iron	IVW	3	1.033 (0.890, 1.199)	0.670
Iron	MR–Egger	3	0.664 (0.468, 0.942)	0.262
Iron	Simple median	3	1.098 (0.982, 1.229)	0.100
Iron	Weighted median	3	1.080 (0.973, 1.200)	0.147
Magnesium	U IVW	6	0.450 (0.057, 3.532)	0.448
Magnesium	MR–Egger	6	0.058 (0, 44.465)	0.448
Magnesium	Simple median	6	2.813 (0.294, 26.959)	0.370
Magnesium	Weighted median	6	0.592 (0.085, 4.110)	0.596
Phosphorus	Ŭ IVW	5	1.196 (0.946, 1.512)	0.134
Phosphorus	MR-Egger	5	0.743 (0.160, 3.443)	0.730
Phosphorus	Simple median	5	1.185 (0.785, 1.788)	0.420
Phosphorus	Weighted median	5	1.123 (0.781, 1.615)	0.532
Selenium	IVW	2	1.036 (0.885, 1.213)	0.658
Zinc	IVW	2	1.032 (0.961, 1.109)	0.387
2	V	itamins	1.002 (0.001) 1.100))	0.007
absolute retinol	IVW	2	1.502 (0.718, 3.142)	0.280
absolute vitamin B-12	IVW	6	1.003 (0.919, 1.095)	0.942
absolute vitamin B-12	MR-Egger	6	0.949 (0.732, 1.230)	0.714
absolute vitamin B-12	Simple median	6	1.018 (0.885, 1.170)	0.804
absolute vitamin B-12	Weighted median	6	0.986 (0.879, 1.107)	0.812
absolute ascorbate	IVW	9	1.013 (0.889, 1.153)	0.848
absolute ascorbate	MR-Egger	9	1.098 (0.895, 1.346)	0.401
absolute ascorbate	Simple median	9	1.050 (0.862, 1.278)	0.631
absolute ascorbate	Weighted median	9	1.060 (0.912, 1.232)	0.445
absolute 250HD	IVW	71	0.899 (0.819, 0.987)	0.025
absolute 250HD	MR-Egger	71	0.876 (0.774, 0.993)	0.042
absolute 250HD	Simple median	71	0.943(0.779, 1.142)	0.548
absolute 250HD	Weighted median	71	0.901(0.800, 1.014)	0.084
absolute alpha-tocopherol	IVW	3	1 192 (0 599, 2 372)	0.616
absolute alpha-tocopherol	MR-Egger	3	8 432 (0 018 4012 598)	0.621
absolute alpha-tocopherol	Simple median	3	1661(0.784, 3.518)	0.185
absolute alpha-tocopherol	Weighted median	3	1 364 (0 673, 2 766)	0.100
relative retinol	IVW	22	0.980(0.949, 1.012)	0.221
relative retinol	MR_Egger	22	$0.963 (0.884 \ 1.049)$	0.395
relative retinol	Simple median	22	0.989(0.949, 1.049)	0.579
relative retinol	Weighted median	22	0.986(0.946, 1.028)	0.515
relative ascorbate	IVW	12	1 005 (0.893, 1.130)	0.937
relative ascorbate	MR Eggor	12	1.003 (0.693, 1.130) 1.008 (0.687, 1.478)	0.970
relative ascorbate	Simple median	12	1.000(0.007, 1.470) 1.050(0.886, 1.245)	0.573
relative ascorbate	Weighted median	12	1.050(0.000, 1.245) 1.086(0.921, 1.280)	0.378
relative alpha tocophoral		12	0.461 (0.321, 1.200)	$6.52 \times 10^{-7}$
relative alpha-tocopherol	MB Egger	11	0.401(0.340, 0.020) 0.480(0.220, 1.084)	0.52 × 10
relative alpha-tocopherol	Simple median	11	0.409(0.220, 1.004) 0.617(0.255, 1.072)	0.112
relative alpha to conherol	Weighted median	11 11	0.017 (0.333, 1.072) = 0.536 (0.224, 0.886)	0.007
relative appra-tocopherol		11	0.000 (0.024, 0.000)	0.010
relative gamma-tocopherol	IV VV	12	0.002 (0.552, 1.470)	0.918
relative gamma-tocopherol	MK-Egger	12	0.902 (0.553, 1.470)	0.688
relative gamma-tocopherol	Simple median	12	1.018 (0.738, 1.404)	0.915
relative gamma-tocopherol	Weighted median	12	0.950 (0.718, 1.259)	0.723

Table 1. Cont.

In our restricted analysis of amino acid profiles and ALS risk, no association was noted between them (Tables 1 and 2, Figure 2, and Supplementary Figure S1). However, the IVW estimated for isoleucine and tryptophan may be vulnerable to the heterogeneity of SNPs (isoleucine: Cochran's Q test *p* value = 0.037; tryptophan: Cochran's Q test *p* value = 0). Due to the limited number of IVs in isoleucine, we cannot conduct further analysis. After removing the tryptophan outliers, we obtained analyzable results (*p*-outlier value = 0.893).

Exposures	F Statistics *	R2	MR-E	gger	Cochr	an's Q	Q MR-PRESSO MR-Steiger			-Steiger		
			Intercept	р	Q	р	RS Sobs	P- Global Test	P- Outlier	P- Distortion	Correct Causal Direction	р
					Es	ssential ami	no acids					
Isoleucine	32.07	0.003	NA	NA	4.333	0.037	NA	NA	NA	NA	TRUE	$4.74 imes10^{-23}$
Leucine	32.532	0.004	0.09	0.32	4.049	0.132	NA	NA	NA	NA	TRUE	$3.27 imes10^{-38}$
Phenylalanine	26.032	0.004	0.083	0.487	3.901	0.142	NA	NA	NA	NA	TRUE	$7.23  imes 10^{-31}$
Tryptophan	16.831	0.043	-0.031	0.587	52.831	0	58.666	$<\!\!2  imes 10^{-4}$	0.893	0.074	TRUE	$4.08 imes10^{-134}$
Valine	29.951	0.006	0.031	0.429	3.423	0.49	4.651	0.587	NA	NA	TRUE	$7.73  imes 10^{-61}$
					Poly	unsaturated	fatty acids					
Arachidonic acid (AA)	1302.627	0.233	NA	NA	0.725	0.395	NA	NA	NA	NA	TRUE	$2.57  imes 10^{-198}$
Docosahexaenoic acid (DHA)	19.211	0.007	0.004	0.943	5.774	0.217	9.826	0.26	NA	NA	TRUE	$2.22  imes 10^{-49}$
Docosapentaenoic acid (DPA)	129.675	0.044	0.016	0.603	3.878	0.144	NA	NA	NA	NA	TRUE	$5.75  imes 10^{-192}$
Eicosapentaenoic acid (EPA)	68.305	0.016	NA	NA	1.321	0.25	NA	NA	NA	NA	TRUE	$8.54 imes10^{-63}$
Gamma linolenic acid (GLA)	77.298	0.018	NA	NA	0.993	0.319	NA	NA	NA	NA	TRUE	$8.70  imes 10^{-76}$
						Minera	ls					
Calcium	22.182	0.003	0.006	0.591	6.118	0.41	8.54	0.486	NA	NA	TRUE	$7.48 imes10^{-85}$
Calcium (UKB)	25.391	0.014	0	0.827	186.865	0.139	188.91	0.152	NA	NA	TRUE	0
Copper	18.08	0.014	NA	NA	0.835	0.361	NA	NA	NA	NA	TRUE	$3.71 \times 10^{-28}$
Iron	159.741	0.01	0.095	0.238	6.747	0.034	NA	NA	NA	NA	TRUE	$9.72  imes 10^{-127}$
Magnesium	20.988	0.005	0.015	0.556	11.06	0.05	19.824	0.1	NA	NA	TRUE	$4.97 imes10^{-73}$
Phosphorus	18.29	0.004	0.022	0.58	2.733	0.603	4.43	0.633	NA	NA	TRUE	$2.82  imes 10^{-50}$
Selenium	39.218	0.008	NA	NA	2.291	0.13	NA	NA	NA	NA	TRUE	$8.59 \times 10^{-42}$
Zinc	23.67	0.018	NA	NA	0.966	0.326	NA	NA	NA	NA	TRUE	$4.77 \times 10^{-28}$
						Vitamiı	ns					
absolute retinol	24.943	0.01	NA	NA	2	0.157	NA	NA	NA	NA	TRUE	$7.89 \times 10^{-25}$
absolute vitamin B-12	75.036	0.012	0.008	0.676	5.109	0.403	0.774	0.366	NA	NA	TRUE	$3.62 \times 10^{-212}$
absolute ascorbate	20.303	0.004	-0.007	0.35	8.331	0.402	9.643	0.506	NA	NA	TRUE	$9.77  imes 10^{-120}$
absolute alpha-tocopherol	3.121	0.001	-0.065	0.643	4.022	0.134	NA	NA	NA	NA	TRUE	$1.04  imes 10^{-24}$
relative retinol	4.158	0.047	0.005	0.665	28.945	0.115	31.643	0.13	NA	NA	TRUE	$1.59  imes 10^{-118}$
relative ascorbate	83.86	0.04	-0.0002	0.988	9.901	0.539	11.769	0.55	NA	NA	TRUE	$1.17 imes10^{-62}$
relative gamma-tocopherol	5.881	0.012	0.004	0.684	14.503	0.206	17.795	0.214	NA	NA	TRUE	$2.71 \times 10^{-52}$

**Table 2.** Summary of the additional MR analysis for the effect of each non ALS-associated trait on ALS; \* the F statistics are the mean F value for each eligible instrumental variable.

NA, Due to the limitation of the number of SNPs, no valid value can be obtained. True, The assumed causal relationship is in the right direction.

With regard to the effect of fatty acids on ALS, the results are summarized in Figure 3. We found a suggestive significant association between LA and the risk of ALS, which was elevated by 6.6% for a one standard deviation unit increase in the LA concentration (IVW-OR: 1.066, 95% CI: 1.011–1.125, p value = 0.019). The robustness of this result was supported by the MR–Egger approach (OR = 1.162, 95% CI: 1.029–1.312, p value = 0.028), and estimates using the weighted median method (OR = 1.049, 95% CI: 0.976-1.128, p value = 0.196) and simple median method (OR = 1.047, 95% CI: 0.972–1.128, *p* value = 0.226) also showed similar trends. No heterogeneity was detected in the individual SNPs associated with LA and the risk of ALS (Cochran's Q test p value = 0.253). No indication of horizontal pleiotropy was found based on the MR–Egger intercept test (intercept = -0.014, p value = 0.146), and the MR–PRESSO analysis showed no outlying SNPs (Table 3). Scatter plots indicated the estimated effect of LA on ALS by each SNP (Figure 4A). In addition, we noted a positive significant association between DGLA and the risk of ALS (OR: 1.031, 95% CI: 1.017–1.045, *p* value =  $1.70 \times 10^{-5}$ ) without the influence of heterogeneity. Due to only two proxy SNPs being available for DGLA, sensitivity analyses could not be performed. The estimated effect of DGLA on ALS by each SNP is displayed in Figure 4B. Genetically predicted AA, DHA, DPA, EPA, and GLA were not associated with the risk of ALS (Tables 1 and 2 and Supplementary Figure S2).

Essential amino acids	No. of SNPs		OR (95% CI)	<i>p</i> -value
Isoleucine			· · ·	
IVW	2		1.003 (0.666, 1.510)	0.988
Leucine				
IVW	3	-	0.961 (0.770, 1.198)	0.723
MR Egger	3		0.370 (0.131, 1.045)	0.312
Simple median	3		0.927 (0.708, 1.214)	0.585
Weighted median	3	-	0.899 (0.745, 1.084)	0.264
(egger-intercept)			(0.090)	0.320
Phenylalanine				
IVW	3		1.035 (0.800, 1.340)	0.792
MR Egger	3	<b>—</b>	0.352 (0.045, 2.722)	0.500
Simple median	3		1.085 (0.844, 1.395)	0.523
Weighted median	3		11.111 (0.887, 1.392)	0.359
(egger-intercept)			(0.083)	0.487
Tryotophan				
IVW	20		1.848 (0.383, 8.916)	0.444
MR Egger	20		609.49 (0, 534720778694.025)	0.549
Simple median	20	$\longrightarrow$	1.096 (0.248, 4.851)	0.904
Weighted median	20	<b>∎</b> →	1.358 (0.302, 6.117)	0.690
(egger-intercept)			(-0.031)	0.587
Valine				
IVW	5	+	0.949 (0.840, 1.071)	0.397
MR Egger	5		0.688 (0.340, 1.391)	0.374
Simple median	5	+	0.940 (0.790, 1.120)	0.492
Weighted median	5	-	0.932 (0.791, 1.099)	0.404
(egger-intercept)			(0.031)	0.429
(egger mercept)			(0.051)	0.420

0 0.5 1 1.5 OR (95% CI) for Amyotrophic Lateral Sclerosis

**Figure 2.** Associations of genetically predicted essential amino acids with risk of ALS using main and sensitivity MR analyses. ALS, amyotrophic lateral sclerosis; IVW, inverse variance–weighted; MR, Mendelian randomization; OR, odds ratio; 95% CI, 95% confidence interval; SNPs, single nucleotide polymorphisms.

We observed no causal relationship between circulating essential minerals and the risk of ALS (Tables 1 and 2, Figure 5, and Supplementary Figure S3). The MR–Egger regression did not identify evidence of horizontal pleiotropy for any of the mineral-related traits. Heterogeneity in the SNPs for iron was inferred (Cochran's Q test p value = 0.034). However, because of the removal of SNPs with a null association with the overall iron status, this calculation was only based on three SNPs.

Methods		Dihomo-Gamma- Linolenic Acid (DGLA)	Linoleic Acid (LA)	Absolute 25-OHD	Relative Alpha-Tocopherol
mean F statistics		218.419	17.009	89.901	4.07
R2		0.049	0.023	0.014	0.006
MR–Egger	intercept	NA	-0.014	0.001	-0.065
00	<i>p</i> value	NA	0.146	0.548	0.643
Cochran's Q	Q	0.062	20.419	84.75	6.652
	<i>p</i> value	0.804	0.253	0.11	0.758
MR-PRESSO	RS Sobs	NA	23.102	85.808	7.907
	P-global test	NA	0.258	0.144	0.807
	P-outlier	NA	NA	NA	NA
	P-distortion	NA	NA	NA	NA
MR–Steiger	correct causal direction	TRUE	TRUE	TRUE	TRUE
	<i>p</i> value	$4.37\times10^{-207}$	$2.11\times10^{-207}$	0	$1.98  imes 10^{-45}$

Table 3. Summary of the additional analysis for the causal effects of each ALS-associated trait on ALS.

NA, Due to the limitation of the number of SNPs, no valid value can be obtained. True, The assumed causal relationship is in the right direction.

Essential polyunsaturated fatty acids	No. of SNPs		OR (95% CI)	<i>p</i> -value
Arachidonic acid (AA)				
IVW	2		0.995 (0.984, 1.007)	0.415
Dihomo–gamma–linolenic acid (DGLA)				
IVW	2		1.031 (1.017, 1.045)	1.70×10 <sup></sup>
Docosahexaenoic acid (DHA)				
IVW	5	-	1.048 (0.924, 1.190)	0.464
MR Egger	5	← ● →	1.013 (0.427, 2.403)	0.978
Simple median	5		1.147 (0.971, 1.355)	0.106
Weighted median	5	-	1.008 (0.889, 1.143)	0.902
(egger-intercept)			(0.004)	0.943
Docosapentaenoic acid (DPA)				
IVW	3		0.874 (0.607, 1.257)	0.466
MR Egger	3	←■	0.660 (0.276, 1.576)	0.521
Simple median	3		0.885 (0.635, 1.223)	0.469
Weighted median	3		0.840 (0.645, 1.095)	0.198
(egger-intercept)			(0.016)	0.603
Eicosapentaenoic acid (EPA)				
IVW	2		0.872 (0.647, 1.175)	0.367
Gamma linolenic acid (GLA)				
IVW	2	←∎→	0.724 (0.184, 2.844)	0.643
Linoleic acid (LA)				
IVW	18		1.066 (1.011, 1.125)	0.019
MR Egger	18		1.162 (1.029, 1.312)	0.028
Simple median	18	+	1.047 (0.972, 1.128)	0.226
Weighted median	18	+	1.049 (0.976, 1.128)	0.196
(egger-intercept)			(-0.014)	0.146



**Figure 3.** Associations of genetically predicted essential polyunsaturated fatty acids with risk of ALS using main and sensitivity MR analyses. ALS, amyotrophic lateral sclerosis; IVW, inverse variance–weighted; MR, Mendelian randomization; OR, odds ratio; 95% CI, 95% confidence interval; SNPs, single nucleotide polymorphisms.

When information about IVs related to circulating vitamins was considered, the results generated by IVW analysis indicated that absolute 25OHD and relative alpha-tocopherol had strongly protective effects on ALS risk (absolute 25OHD, OR: 0.899, 95% CI: 0.819–0.987, *p* value = 0.025; and relative alpha-tocopherol, OR: 0.461; 95% CI: 0.340–0.626; *p* value =  $6.25 \times 10^{-7}$ ) (Tables 1 and 3 and Figure 6). Both associations were broadly concordant in various sensitivity analyses. The results of the MR–Egger method (OR = 0.876, 95% CI: 0.774–0.993, *p* value = 0.042) and weighted median method (OR = 0.901, 95% CI: 0.800–1.014, *p* value = 0.084) underpinned the causal effect of 25OHD on ALS, while the estimates of the simple median method (OR = 0.617, 95% CI: 0.324–0.886, *p* value = 0.042) and weighted median method, *p* value = 0.087) and weighted median method (OR = 0.536, 95% CI: 0.324–0.886, *p* value = 0.015) were aligned with the significant relationship between relative alpha-tocopherol. No hetero-

geneity was detected using Cochran's Q test separately (absolute 25OHD: *p* value = 0.110; relative alpha-tocopherol: *p* value = 0.758). The MR–Egger intercept test showed no evidence of directional pleiotropy (absolute 25OHD: intercept = 0.001, *p* value = 0.548; relative alpha-tocopherol: intercept = -0.065, *p* value = 0.643), and no outlying SNP was found. The scatter plots indicated the SNP-based causal estimates of absolute 25OHD on ALS (Figure 4C) and relative alpha-tocopherol on ALS (Figure 4D). The LOO analysis showed that SNPs rs577185477 and rs11723621 at a known risk locus for absolute 25OHD influenced the causal estimate for ALS (Supplementary Figure S4F). No effect of individual relative alpha-tocopherol-related SNPs on ALS risk was observed (Supplementary Figure S4H). A null effect of genetically predicted increases in circulating levels of absolute retinol, absolute vitamin B-12, absolute ascorbate, absolute alpha-tocopherol, relative retinol, relative ascorbate, and relative gamma-tocopherol on the risk of ALS was found (Tables 1 and 2, Figure 6, and Supplementary Figure S4).



**Figure 4.** Scattered plots of the casual effect of essential nutrients on ALS. The x-axis plots the previously published  $\beta$ -estimate for the association between each SNP and essential nutrients. The y-axis plots the  $\beta$ -estimate for the association between each SNP and risk of ALS. The slope of each line corresponds to the estimated MR effect per method. The examined essential nutrients included (**A**) linoleic acid (LA), (**B**) Dihomo–gamma–linoleic acid (DGLA), (**C**) absolute 25OHD, and (**D**) relative alpha-tocopherol. ALS, amyotrophic lateral sclerosis; IVW, inverse variance–weighted; MR, Mendelian randomization; OR, odds ratio; 95% CI, 95% confidence interval; SNPs, single nucleotide polymorphisms.

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123(0.781, 1.615)	0.532
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OR (95% CI) for Amyotrophic Lateral Sclerosis

**Figure 5.** Associations of genetically predicted essential minerals with risk of ALS using main and sensitivity MR analyses. ALS, amyotrophic lateral sclerosis; IVW, inverse variance–weighted; MR, Mendelian randomization; OR, odds ratio; 95% CI, 95% confidence interval; SNPs, single nucleotide polymorphisms.

Vitamins	No. of SNPs		OR (95% CI)	<i>p</i> -value
absolute retinol				-
IVW	2		1.502 (0.718, 3.142)	0.280
absolute vitamin B12				
IVW	6	+	1.003 (0.919, 1.095)	0.942
MR Egger	6		0.949 (0.732, 1.230)	0.714
Simple median	6		1.018 (0.885, 1.170)	0.804
Weighted median	6	-	0.986 (0.879, 1.107)	0.812
(egger-intercept)			(0.008)	0.676
absolute ascorbate				
IVW	9		1.013 (0.889, 1.153)	0.848
MR Egger	9		1.098 (0.895, 1.346)	0.401
Simple median	9		1.050 (0.862, 1.278)	0.631
Weighted median	9		1.060 (0.912, 1.232)	0.445
(egger-intercept)			(-0.007)	0.350
absolute 25OHD				
IVW	71		0.899 (0.819, 0.987)	0.025
MR Egger	71		0.876 (0.774, 0.993)	0.042
Simple median	71		0.943 (0.779, 1.142)	0.548
Weighted median	71		0.901 (0.800, 1.014)	0.084
(egger-intercept)			(0.001)	0.548
absolute alpha-tocopherol				
IVW	3		1.192 (0.599, 2.372)	0.616
MR Egger	3	· · · · · ·	8.432 (0.018, 4013)	0.462
Simple median	3		1.661 (0.784, 3.518)	0.185
Weighted median	3		1.364 (0.673, 2.766)	0.389
(egger-intercept)			(-0.065)	0.643
relative retinol				
IVW	22	-	0.980 (0.949, 1.012)	0.221
MR Egger	22	-	0.963 (0.884, 1.049)	0.395
Simple median	22	•	0.989 (0.949, 1.030)	0.579
Weighted median	22	•	0.986 (0.946, 1.028)	0.515
(egger-intercept)			(0.005)	0.665
relative ascorbate				
IVW	12		1.005 (0.893, 1.130)	0.937
MR Egger	12		1.008 (0.687, 1.478)	0.970
Simple median	12		1.050 (0.886, 1.245)	0.573
Weighted median	12		1.086 (0.921, 1.280)	0.328
(egger-intercept)			(-0.0002)	0.988
relative alpha-tocopherol				7
IVW	11	÷	0.461 (0.340, 0.626)	6.52×10 <sup>-7</sup>
MR Egger	11	•	0.489 (0.220, 1.084)	0.112
Simple median	11	+ <b>-</b>	0.617 (0.355, 1.072)	0.087
Weighted median	11		0.536 (0.324, 0.886)	0.015
(egger-intercept)			(-0.065)	0.643
relative gamma-tocopherol	10	1	0.000 (0.001 1.015)	0.040
IVW	12		0.988 (0.784, 1.245)	0.918
MK Egger	12		0.902 (0.553, 1.470)	0.688
Simple median	12		1.018 (0.738, 1.404)	0.915
weighted median	12		0.950 (0.718, 1.259)	0.723
(egger-intercept)			(0.004)	0.684
		0.5 1 1.5		
	OR (95%	6 CI) for Amyotrophic Lateral	Sclerosis	

**Figure 6.** Associations of genetically predicted essential vitamins with risk of ALS using main and sensitivity MR analyses. ALS, amyotrophic lateral sclerosis; IVW, inverse variance–weighted; MR, Mendelian randomization; OR, odds ratio; 95% CI, 95% confidence interval; SNPs, single nucleotide polymorphisms.

## 4. Discussion

In the present MR study, we classified and profiled the potential causal role of essential dietary nutritiona6l modifiers on ALS risk using GWAS summary statistics. Higher genetically predicted levels of 25OHD and relative alpha-tocopherol were calculated to be associated with a reduced risk of ALS, and increased LA levels showed a suggestive positive association with ALS risk. Our findings implied that the abnormal levels of these nutrients were not only a symptom of ALS but also independent risk/protective factors for ALS. Among the essential amino acids and minerals, no evidence supports that additional supplementation has an influence on the risk of ALS.

Essential fatty acids are important components of cholesterol and phospholipids and are involved in the maintenance of membrane fluidity and beta-oxidation [50]. Our results showed that LA was a major risk factor for the effects of essential fatty acids on ALS, preliminarily suggesting that dietary supplementation with LA is not beneficial or is even harmful to ALS. As secondary evidence, we found that DGLA, which is derived from LA and is capable of anti-inflammatory and antiproliferative effects, had a strong risk effect on ALS [51,52]. LA is an n-6 polyunsaturated fatty acid. Because of the deficiency of desaturases that insert double bonds beyond C9 in humans, LA is accessible only from diet [53]. Although there is no direct clinical evidence for the neurotoxic effect of LA, a longitudinal pre-diagnostic study revealed that circulating AA, an N-6 polyunsaturated fatty acid derived from LA, was positively associated with the risk of ALS [13]. Due to the extremely low concentrations of polyunsaturated fatty acids in plasma and the limited sample sizes of that study, the measurement error cannot be neglected. Our study adopted the MR framework and involved large-scale genetic data, ensuring reliable statistical power. Studies have shown that LA-rich food increases oxidative stress and inflammation [54,55], both of which contribute to ALS [56]. LA enhances tumor necrosis factor (TNF)-mediated oxidative stress [57], while TNF- $\alpha$ , a potent inflammatory cytokine, has been found to induce apoptosis and to contribute to oxidative stress by activating microglia in an SOD1-G93A mouse model [58]. In addition, LA is considered to limit the synthesis of longchain n-3 PUFAs from alpha-linolenic acid [59,60], while the intake of n-3 PUFA-enriched foods can prevent or delay the onset of ALS by reducing inflammation and providing neuroprotection [14,61]. Therefore, excessive LA may create a potentially inflammatory environment that increases the risk of ALS [62]. However, no relationship was found between n-3 PUFAs and ALS in our research, which may be due to the limited IV numbers of n-3 PUFAs and thereby the finite proportion of phenotypic variance explanation. In the future, more experiments are needed to explore the exact mechanism underlying LA and ALS.

Among the vitamins, we found an inversely causal association between two fat-soluble vitamins, namely, vitamin D and vitamin E, and ALS, while water-soluble vitamins showed negative results. Vitamin D is a steroid molecule that is produced in the skin by exposure to the ultraviolet spectrum of sunlight or is obtained from food and biological dietary supplements [63]. It has been reported that severe vitamin D deficiency is independently associated with significantly accelerated ALS progression and reduced life expectancy [64,65]. Since overactivation of glutamate receptors leads to continuously increased cytoplasmic calcium concentrations and subsequent ALS burden [66,67], vitamin D can exert neuroprotective effects by upregulating calcium-binding proteins and reducing glutamate-induced caspase-3 activity [68,69]. In addition, vitamin D can assist neurotrophic factors in protecting motoneurons [64]. However, contrary to our findings, Susanna C Larsson et al. reported a null causal association between serum 25OHD concentrations and ALS risk [70], which may be due to the different selections of GWAS data. The 25OHD GWAS [71] utilized in their study included 79,366 Europeans and yielded only two more new loci (SEC23A and AMDHD1) than previous 25OHD GWAS findings and explained approximately 7.5% of the heritability of 25OHD. Due to the gene prediction level of 25OHD depending on a medium polygenic structure, the statistical power of the 25OHD GWAS used was insufficient. To compensate for this shortcoming, we enrolled the 25OHD GWAS with the largest

sample size. Similarly, a larger ALS GWAS and various sensitivity analyses were employed in our study to ensure convincing and robust estimates.

Vitamin E is another common essential vitamin [72], which mainly provides antioxidative protection against lipid peroxidation, reactive oxygen species (ROS), and reactive nitrogen species (RNS) [73]. The results of several large prospective cohort studies implied that the regular use of vitamin E supplements has a time-dependent protective effect on ALS [10,74]. In addition, vitamin E was suggested to have a synergistic effect with n-3 polyunsaturated fatty acids on reducing the risk of ALS [75]. In this category, we adopted two respective sets of instruments to measure the absolute and relative concentrations of vitamins, aiming to generate robust results. Although both sets for retinol and ascorbate yield similar conclusions, a significant association between the relative concentration of alpha-tocopherol and the risk of ALS was not found for absolute alpha-tocopherol. Owing to the high overlapping IVs of absolute alpha-tocopherol and lipid metabolism [76], its protective effect may be offset by the strong risk that lipids pose in ALS. Moreover, the number of IVs is too limited to develop MR–PRESSO analysis; therefore, pleiotropic bias cannot be ruled out, which violates the MR-InSIDE hypothesis. The IVs adopted for relative alpha-tocopherol have allowed for circumvention of these hurdles. Hence, our findings, in combination with previous literature, indicate that vitamin E supplementation may be a preventive agent for ALS. However, vitamin E and fat always coexist in the diet, making it necessary to comprehensively consider the complex effects of the overall diet on ALS, where using vitamin supplements may be advisable.

This pioneering study provided evidence for the primary prevention of ALS from a dietary perspective. We found that ALS-associated essential nutrients were concentrated in polyunsaturated fatty acids and fat-soluble vitamins, providing supportive evidence of the importance of lipid metabolism in ALS. No evidence has shown that diet-derived circulating essential amino acids and minerals influence the risk of ALS in the general population, even though some of them (e.g., branched-chain amino acids [77], iron, zinc, and selenium [78]) have attracted widespread attention. However, our research has certain limitations. First, since MR analyses assume linearity, potential nonlinear associations may bias the results, whereas we were unable to confirm whether the relationships between these essential nutrients and ALS risk are non-U-shaped or are other shapes. Second, some vitamins have a variety of compounds. A previous study proposed that the plasma alphatocopherol concentration was not an ideal biomarker for vitamin E status [79]. The survival of circulating alpha-tocopherol was longer in individuals with high lipid levels, possibly related to reduced lipoprotein catabolism and tissue absorption. As the best biomarkers for vitamin status cannot be determined via MR approaches, we still leveraged currently widely recognized active forms of vitamins as their proxies, which have mature measurement techniques and biological mechanisms, allowing for more clinically applicable conclusions than those for the unusual forms. It is tempting to select other naturally occurring vitamin forms to verify our findings. Third, our results suggested a potential influence of dietary nutrient supplementation on ALS prevention. The estimates according to genetic variance reflect the effects of lifetime accumulation exposure to a biomarker. Therefore, the shortterm efficacy of nutrient supplementation for diseases should be interpreted with caution. Fourth, given that different nutrients play specific roles in biological processes, their protective effect on the overall ALS population may be driven by a strong effect on a certain subgroup of diseases. For example, vitamin D can maintain calcium homeostasis, which may be extremely beneficial for those with a severe calcium imbalance. However, the lack of individual-level ALS GWAS data makes it hard to validate this hypothesis accurately. Perhaps subsequent experiments could pay attention to this issue. Moreover, when genetic data become available for ALS patients with clinical phenotypes, future similar studies should be performed to assess whether dietary nutrients provide modifying effects on ALS progression and not just consider disease prevention.

## 5. Conclusions

Evidence from our study demonstrated that vitamin D and vitamin E are protective against the risk of ALS, and LA is suggested to be positively associated with ALS risk. It is unlikely that a clinical benefit for the primary prevention of ALS will be achieved through supplementing essential amino acids and minerals. Our study classified and profiled the effects of specific diet-derived circulating nutrients on the risk of ALS, providing evidence for medical interventions for ALS in the general population from the perspective of gene–environmental interactions.

**Supplementary Materials:** The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/nu14050920/s1, Figure S1: Scatter plot and forest plot of essential amino acids and amyotrophic lateral sclerosis; Figure S2: Scatter plot and forest plot of essential fatty acids and amyotrophic lateral sclerosis; Figure S3: Scatter plot and forest plot of essential minerals and amyotrophic lateral sclerosis; Figure S4: Scatterplot and forest plot of essential vitamins and amyotrophic lateral sclerosis; Table S1: Characteristics of instrumental variables (IVs).

**Author Contributions:** Formal analysis, K.X.; data curation, Y.W.; writing—original draft preparation, Y.W. and K.X.; writing—review and editing, L.T., K.X., and Y.W.; methodology, T.H. and N.H.; validation, L.Z. and G.Z.; supervision, D.F. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the National Natural Science Foundation of China, grant numbers 81873784 and 82071426, and the Clinical Cohort Construction Program of Peking University Third Hospital, grant number BYSYDL2019002.

**Institutional Review Board Statement:** Ethical review and approval were waived for this study because no individual patients were directly involved in the overall process of our study. Our study was based only on publicly available GWAS data.

Informed Consent Statement: Not applicable.

**Data Availability Statement:** All data generated or analyzed during this study are included in this published article and its Supplementary Materials. Codes generated or used during the study are available from the corresponding author by request.

**Acknowledgments:** We would like to thank the GWASs for making summary data publicly available, and we are grateful to all participants who contributed to those studies. We also thank the National Natural Science Foundation of China and Clinical Cohort Construction Program of Peking University Third Hospital for the support.

Conflicts of Interest: The authors declare no conflict of interest.

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