

# Potential role of hydrogen sulfide in central nervous system tumors: a narrative review

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## Abstract

Central nervous system tumors are classified as diseases of special clinical significance with high disability and high mortality. In addition to cerebrovascular diseases and craniocerebral injuries, tumors are the most common diseases of the central nervous system. Hydrogen sulfide, the third endogenous gas signaling molecule discovered in humans besides nitric oxide and carbon monoxide, plays an important role in the pathophysiology of human diseases. It is reported that hydrogen sulfide not only exerts a wide range of biological effects, but also develops a certain relationship with tumor development and neovascularization. A variety of studies have shown that hydrogen sulfide acts as a vasodilator and angiogenic factor to facilitate growth, proliferation, migration and invasion of cancer cells. In this review, the pathological mechanisms and the effect of hydrogen sulfide on the central nervous system tumors are introduced.

**Key words:** central nervous system; clinical research; experimental research; glioblastoma multiforme; hydrogen sulfide; hypoxia inducible factor-1 $\alpha$ ; p38 mitogen-activated protein kinase; pituitary tumor; therapeutic implications

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## INTRODUCTION

Central nervous system (CNS) tumors are classified as nervous system diseases characterized by high disability and mortality.<sup>1</sup> They are the most common diseases of the CNS except for cerebrovascular diseases and craniocerebral injuries, including meningiomas, gliomas, pituitary tumors and so on.<sup>2</sup> Among them, glioblastoma multiforme (GBM) is the most malignant brain tumor, with a strong ability to spread and invade the surrounding parenchyma. The number of treatment options is increasing; however, the prognosis is still poor.<sup>3,4</sup> Pituitary tumors are a group of tumors originated from the residual cells of anterior and posterior pituitary and craniopharyngeal epithelium, accounting for 10–20% of intracranial tumors, most of which are benign.<sup>5,6</sup>

Hydrogen sulfide (H<sub>2</sub>S) is usually considered an environmental pollutant with its smell of rotten eggs. However, with continuously furthering research in recent years, H<sub>2</sub>S has been discovered as a gaseous chemical messenger with regulatory roles in neurotransmission, cardiovascular regulation, cell metabolism and other physiological processes in mammals.<sup>7-9</sup> Recently, it has been reported that H<sub>2</sub>S plays a role in a wide range of physiological and pathophysiological properties on cancer progress.<sup>10,11</sup> Previous results have evidenced that H<sub>2</sub>S may act as a vascular relaxant and angiogenic factor to promote the growth, proliferation, migration and invasion of cancer cells.<sup>12,13</sup> H<sub>2</sub>S is extremely fat-soluble, can freely pass through cell membranes, widely presenting in various parts of the human body, with a variety of production and transformation methods.<sup>14</sup> Generally speaking, on the one hand, H<sub>2</sub>S can be synthesized by cystathionine  $\beta$ -synthetase and cystathionine

c-lyase extracted with sulfur-containing amino acids as substrates, in pyridoxal-50-phosphate-(PLP)-dependent reactions.<sup>15-17</sup> On the other hand, it can produce through nonenzymatic reduction of elemental sulfur in the blood using reduction equivalents provided by glycolysis, or in the form of sodium bisulfide.<sup>18</sup> Changes in H<sub>2</sub>S metabolism and/or signal transduction are closely related to human diseases, especially cancer. An increasing number of studies have reported that cystathionine  $\beta$ -synthetase, cystathionine c-lyase and 3-mercaptopyruvate sulfurtransferase are overexpressed in tumor cell lines and tumor tissues, with colorectal cancer, ovarian cancer, breast cancer, and glioma involved.<sup>11,16,19-21</sup> In this review, the potential role of endogenous and exogenous H<sub>2</sub>S in CNS tumors is described and the implications of H<sub>2</sub>S on future treatment strategies are discussed.

## EXPERIMENTAL FINDING ABOUT THE ROLES OF HYDROGEN SULFIDE IN CENTRAL NERVOUS SYSTEM TUMORS

The mechanisms underlying the roles of H<sub>2</sub>S in the CNS tumors have not been fully elucidated. As is known to all, a therapeutic method must be verified by a large amount of basic experiments before being applied to clinic. However, not all studies can reach the same result. We collected several experiments related to CNS tumors and H<sub>2</sub>S and summarized the outcomes (**Table 1**), in which the researchers detected the effects of H<sub>2</sub>S on brain tumors and explored the potential mechanisms by which this gas can promote tumor cells growth.

Li et al.<sup>22</sup> reported that H<sub>2</sub>S serves as a stimulator in the development of rat glioblastoma and exogenous H<sub>2</sub>S strongly promotes the tumor growth. It was observed that the H<sub>2</sub>S

**Table 1: Experimental studies regarding hydrogen sulfide in brain tumor**

Study	Year	Cell	Cancer model	Results
Li et al. <sup>22</sup>	2012	C6 glioma cell	Glioblastoma	Hydrogen sulfide serves as a stimulator in the development of rat glioblastoma by the increase of hypoxia-inducible factor-1 $\alpha$ expression and neovascular formation.
Zhen et al. <sup>23</sup>	2015	C6 glioma cell	Glioblastoma	Exogenous hydrogen sulfide promotes C6 glioma cell growth through activation of the p38 mitogen-activated protein kinase/extracellular signal-regulated protein kinase1/2-cyclooxygenase-2 pathways.
Zhao et al. <sup>24</sup>	2015	C6 glioma cell	Glioblastoma	Exogenous hydrogen sulfide effectively reduces cell number of C6 cells by triggering the apoptosis via Caspase-dependent pathway.
Sitdikova et al. <sup>25</sup>	2010	Rat pituitary tumor cell GH3	Pituitary tumor	Hydrogen sulfide increases calcium-activated potassium channel activity of rat pituitary tumor cells in a concentration-dependent manner.
Mustafina et al. <sup>26</sup>	2015	Rat pituitary tumor cell GH3	Pituitary tumor	Hydrogen sulfide induces hyperpolarization of GH3 cells with dose-dependent, resulting in a decrease of hormone release.

content in the tumor group was significantly higher than that in the control group. Compared with normal rats, endogenous H<sub>2</sub>S production in the brain of tumor-bearing mice increased, and with the growth of tumors, endogenous H<sub>2</sub>S production further increased. Furthermore, exogenous H<sub>2</sub>S promoted the expression of hypoxia inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ) in GBM tissues, while increased HIF-1 $\alpha$  further stimulates tumor growth. HIF-1 $\alpha$  can be activated in a variety of factors (including hypoxia, inflammation, adenosine triphosphate levels, and isocitrate dehydrogenase (IDH) mutation status), as well as in gliomas and immune cell infiltrating tumors.<sup>27</sup> HIF-1 $\alpha$  increases the expression of inducible nitric oxide synthase in cancer cells, which impairs the recruitment of tumor immune cells and produces immunosuppression.<sup>28</sup> Similarly, HIF-1 $\alpha$ 's increased activity promotes the production of vascular endothelial growth produced by endothelial cells, which can modulate chemotactic properties of immune cell-mediated tumor infiltration.<sup>29</sup> Vascular endothelial growth factor binds to its receptor neuropilin-1 to attract regulatory T cells to the tumor site, and malignant cells secrete transforming growth factor- $\beta$  to attract regulatory T cells to GBM, preventing the killing of cancer cells.<sup>27,30</sup> There is increasing evidence that IDH1, closely related to the carcinogenesis mediated by HIF-1 $\alpha$ , is the most common mutated gene in GBM.<sup>31-33</sup> The mutated IDH1 increases its level by protecting HIF-1 $\alpha$  from degradation. In the midst of IDH1 mutant gliomas, the transcriptional activity of HIF-1 $\alpha$  increases, intensifying the growth of gliomas.<sup>34,35</sup> Besides, under normal oxygen conditions, the degradation of HIF-1 $\alpha$  is promoted by prolyl hydroxylase domain-containing proteins with its transcriptional activity being inhibited by HIF-1 $\alpha$  inhibitor protein (encoded by Hif-1 $\alpha$ ).<sup>36,37</sup> A typical feature of HIF-1 $\alpha$ -dependent signaling is hypoxia, a condition in which tissue oxygen supply is in reduction.<sup>37,38</sup> Since hypoxic conditions are characteristic of the tumor microenvironment, the effects of both are inhibited. Therefore, HIF-1 $\alpha$  is often upregulated in solid malignant tumors (such as GBM).<sup>38</sup> Lactic acid accumulation caused by hypoxia can lead to an acidic tumor microenvironment and induction local inflammation. Lactic acid can promote the polarization of macrophages' expression of immunosuppressive arginase 1 through HIF-1 $\alpha$ -mediated mechanism and promotion of tumor growth.<sup>27,39</sup> In addition, the tumor-specific T cell effector functions can be impacted by HIF-1 $\alpha$  via an increase

of glycolysis and promotion of T cell differentiation.<sup>40,41</sup> This study provides evidence for the angiogenic effect of H<sub>2</sub>S mediated by promoting and stabilizing HIF-1 $\alpha$  protein expression under hypoxic conditions.

In 2015, Zhen et al.<sup>23</sup> found that H<sub>2</sub>S induced C6 glioma cell proliferation through utilizing its two-fold cytoprotective and anti-apoptosis functions via decrease of the expression of caspase-3. Within the range of physiological dose of H<sub>2</sub>S (0.2–1 mM), different doses of NaHS (100–1600  $\mu$ M) promoted cell proliferation. The optimal concentration of NaHS for inducing maximum proliferation and markedly diminished cell apoptosis was 400  $\mu$ M. On the other hand, Zhao et al.<sup>24</sup> reported NaHS failed to act as a mitogen to promote proliferation of C6 cells but succeeded in functioning as a stimulus to activate the apoptosis of C6 cells. They showed that the application of NaHS significantly increased the phospho-p38/p38 protein expression rate in C6 cells, indicating that p38 mitogen-activated protein kinase (MAPK) activation is linked to NaHS-mediated apoptosis of C6 cells. Different research results are related to different conditions, such as time and concentration of cell treatment. The latter treated C6 cells with NaHS (100–1000  $\mu$ M) for 48 hours. MAPK, a serine threonine protein kinase, evolutionarily conserved in all eucaryotes that can be activated by different extracellular stimuli, such as cytokines, neurotransmitters, hormones, cell stress and cell adhesion,<sup>42-44</sup> and plays significant roles in gene expression regulation and cytoplasmic function activities. Moreover, MAPK takes part in the regulation of many important cell physiology/pathology processes such as cell growth and proliferation, differentiation, adaption to the environment, and inflammation response.<sup>45</sup> Researches have founded that MAPKs (p38 MAPK and extracellular signal-regulated protein kinase 1/2) also activate cyclooxygenase-2 signal pathway and other signaling cascades in various tumor cell types.<sup>46-49</sup> Cyclooxygenase-2 is a kind of inducible enzyme whose expression is enhanced under the conditions of tissue damage and inflammation, closely related to the growth of tumor as an important molecule in the process of tumor proliferation, angiogenesis, anti-apoptosis and invasion.<sup>50-53</sup> Besides, previous studies have emphasized that the activation of cyclooxygenase-2 in the glioma depends on the extracellular signal-regulated protein kinase 1/2 pathway.<sup>54-56</sup> p38 MAPK and extracellular signal-regulated protein kinase 1/2 pathway can be activated by NaHS to induce proliferation



and anti-apoptosis.<sup>23</sup>

In addition, some studies have discovered the potential role of H<sub>2</sub>S in pituitary tumor cells. The one major discover of these studies is that the NaHS induces a dose-dependent hyperpolarization and truncation of spontaneous action potentials in rat pituitary GH3 cells via activating of adenosine triphosphate-sensitive potassium channels as H<sub>2</sub>S donor, resulting in decrease of secretion.<sup>26</sup> NaHS increases the magnitude of adenosine triphosphate-sensitive potassium currents, as well as whether in GH3 cells at rest or in a depolarization reaction, NaHS can reduce the exocytosis of secreted particles.<sup>26</sup> This effect is exerted in a dose-dependent manner.<sup>26</sup> Sitdikova et al.<sup>25</sup> found H<sub>2</sub>S promotes the activation of calcium-activated potassium (BK) channels in rat pituitary tumor (GH3) cells, which is probably related to the reduction of sulfhydryl groups of the channel protein. BK channels mediates or regulates many physiological functions and pathophysiological conditions.<sup>57</sup> As is known to all, the activity of BK channels in the channels protein is regulated by the redox state of the key sulfhydryl group, with the exchange of free mercaptan and disulfide involved.<sup>58,59</sup> Under oxidation conditions, activity of BK channels is enhanced in both reduction and inhibition.<sup>60</sup> H<sub>2</sub>S as a reducing agent may increase the channel open probability by redox regulation of cysteine or other residues of channel protein.<sup>25</sup>

It is concluded that H<sub>2</sub>S is fully involved in the pathological process of CNS tumors, especially in that of gliomas and pituitary tumors, indicating that the clinical application of H<sub>2</sub>S has great prospects, which can be further examined as a potential neuroprotective gas for CNS tumors, according to experimental results above.

## CLINICAL FINDINGS ABOUT THE ROLES OF CENTRAL NERVOUS SYSTEM TUMORS

Currently, research on H<sub>2</sub>S in CNS tumors is still at the cells experimental stage and has not yet been applied clinically. However, there is ample evidence to show that *in vivo*, H<sub>2</sub>S is produced mainly by two key enzymes, cystathionine β-synthetase and cystathionine c-lyase, which are primarily present in the CNS. Although there is no direct evidence indicating that H<sub>2</sub>S has neuroprotective effect in clinical trials, the close touch with brain injury caused by CNS tumors is no doubt, which needs us to make great efforts to the study.

## PERSPECTIVES

According to the data of the current research, H<sub>2</sub>S plays a role in promoting growth of brain tumors. Although some studies have found that the role of H<sub>2</sub>S is somewhat inconsistent with those mentioned above, which may be caused by differences in dose, mode of action, and type of disease. Although we have not fully understood its mechanism, we will continue to do a lot of research in the future. Finally, we believe that H<sub>2</sub>S will open up a new path into the treatment of the CNS tumors.

### Author contributions

WP and MLZ were responsible for writing the manuscript. JZ was responsible for its revision. JZ and GC were responsible for its drafting and revision. All authors read and approved the final version of the manuscript for publication.

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## REFERENCES

- Leece R, Xu J, Ostrom QT, Chen Y, Kruchko C, Barnholtz-Sloan JS. Global incidence of malignant brain and other central nervous system tumors by histology, 2003-2007. *Neuro Oncol.* 2017;19:1553-1564.
- McNeill KA. Epidemiology of brain tumors. *Neurol Clin.* 2016;34:981-998.
- Porcù E, Maule F, Boso D, et al. BMP9 counteracts the tumorigenic and pro-angiogenic potential of glioblastoma. *Cell Death Differ.* 2018;25:1808-1822.
- Blumenthal DT, Goria T, Gilbert MR, et al. Is more better? The impact of extended adjuvant temozolomide in newly diagnosed glioblastoma: a secondary analysis of EORTC and NRG Oncology/RTOG. *Neuro Oncol.* 2017;19:1119-1126.
- Karimian-Jazi K. Pituitary gland tumors. *Radiologe.* 2019;59:982-991.
- Laws ER, Jr, Penn DL, Repetti CS. Advances and controversies in the classification and grading of pituitary tumors. *J Endocrinol Invest.* 2019;42:129-135.
- Zhang ML, Peng W, Ni JQ, Chen G. Recent advances in the protective role of hydrogen sulfide in myocardial ischemia/reperfusion injury: a narrative review. *Med Gas Res.* 2021;11:83-87.
- Mao YG, Chen X, Zhang Y, Chen G. Hydrogen sulfide therapy: a narrative overview of current research and possible therapeutic implications in future. *Med Gas Res.* 2020;10:185-188.
- Zhang J, Shi C, Wang H, et al. Hydrogen sulfide protects against cell damage through modulation of PI3K/Akt/Nrf2 signaling. *Int J Biochem Cell Biol.* 2019;117:105636.
- Ye M, Yu M, Yang D, et al. Exogenous hydrogen sulfide donor NaHS alleviates nickel-induced epithelial-mesenchymal transition and the migration of A549 cells by regulating TGF-β1/Smad2/Smad3 signaling. *Ecotoxicol Environ Saf.* 2020;195:110464.
- Cao X, Ding L, Xie ZZ, et al. A review of hydrogen sulfide synthesis, metabolism, and measurement: is modulation of hydrogen sulfide a novel therapeutic for cancer? *Antioxid Redox Signal.* 2019;31:1-38.
- Zhang S, Bian H, Li X, et al. Hydrogen sulfide promotes cell proliferation of oral cancer through activation of the COX2/AKT/ERK1/2 axis. *Oncol Rep.* 2016;35:2825-2832.
- Akbari M, Sogutdelen E, Juriasingani S, Sener A. Hydrogen sulfide: emerging role in bladder, kidney, and prostate malignancies. *Oxid Med Cell Longev.* 2019;2019:2360945.
- Kashfi K. The role of hydrogen sulfide in health and disease. *Biochem Pharmacol.* 2018;149:1-4.
- Kimura H. Signaling by hydrogen sulfide (H<sub>2</sub>S) and polysulfides (H<sub>2</sub>S(n)) in the central nervous system. *Neurochem Int.* 2019;126:118-125.
- Giuffrè A, Tomé CS, Fernandes DGF, Zuhra K, Vicente JB. Hydrogen sulfide metabolism and signaling in the tumor microenvironment. *Adv Exp Med Biol.* 2020;1219:335-353.
- Wang XY, Yang HW. Upregulation of CBS/H<sub>2</sub>S system contributes to asymmetric dimethylarginine-triggered protection against the neurotoxicity of glutamate to PC12 cells by inhibiting NOS/NO pathway. *Exp Cell Res.* 2016;346:111-118.
- di Masi A, Ascenzi P. H<sub>2</sub>S: a "double face" molecule in health and disease. *Biofactors.* 2013;39:186-196.
- Bhattacharyya S, Saha S, Giri K, et al. Cystathionine beta-synthase (CBS) contributes to advanced ovarian cancer progression and drug resistance. *PLoS One.* 2013;8:e79167.





20. Szabo C, Coletta C, Chao C, et al. Tumor-derived hydrogen sulfide, produced by cystathionine- $\beta$ -synthase, stimulates bioenergetics, cell proliferation, and angiogenesis in colon cancer. *Proc Natl Acad Sci U S A*. 2013;110:12474-12479.
21. Hellmich MR, Coletta C, Chao C, Szabo C. The therapeutic potential of cystathionine  $\beta$ -synthetase/hydrogen sulfide inhibition in cancer. *Antioxid Redox Signal*. 2015;22:424-448.
22. Li ZY, Liu SC, Xu PJ, Yang Z, Zhang T. Hydrogen sulfide stimulates the development of rat glioblastoma. *Zhonghua Zhong Liu Za Zhi*. 2012;34:254-258.
23. Zhen Y, Zhang W, Liu C, et al. Exogenous hydrogen sulfide promotes C6 glioma cell growth through activation of the p38 MAPK/ERK1/2-COX-2 pathways. *Oncol Rep*. 2015;34:2413-2422.
24. Zhao L, Wang Y, Yan Q, Lv W, Zhang Y, He S. Exogenous hydrogen sulfide exhibits anti-cancer effects through p38 MAPK signaling pathway in C6 glioma cells. *Biol Chem*. 2015;396:1247-1253.
25. Sitdikova GF, Weiger TM, Hermann A. Hydrogen sulfide increases calcium-activated potassium (BK) channel activity of rat pituitary tumor cells. *Pflugers Arch*. 2010;459:389-397.
26. Mustafina AN, Yakovlev AV, Gaifullina A, Weiger TM, Hermann A, Sitdikova GF. Hydrogen sulfide induces hyperpolarization and decreases the exocytosis of secretory granules of rat GH3 pituitary tumor cells. *Biochem Biophys Res Commun*. 2015;465:825-831.
27. Gabriely G, Wheeler MA, Takenaka MC, Quintana FJ. Role of AHR and HIF-1 $\alpha$  in glioblastoma metabolism. *Trends Endocrinol Metab*. 2017;28:428-436.
28. Peyssonnaud C, Datta V, Cramer T, et al. HIF-1 $\alpha$  expression regulates the bactericidal capacity of phagocytes. *J Clin Invest*. 2005;115:1806-1815.
29. Gerri C, Marin-Juez R, Marass M, Marks A, Maischein HM, Stainier DYR. Hif-1 $\alpha$  regulates macrophage-endothelial interactions during blood vessel development in zebrafish. *Nat Commun*. 2017;8:15492.
30. Atzori MG, Tentori L, Ruffini F, et al. The anti-vascular endothelial growth factor receptor-1 monoclonal antibody D16F7 inhibits invasiveness of human glioblastoma and glioblastoma stem cells. *J Exp Clin Cancer Res*. 2017;36:106.
31. Chen JB, Lu YY, Xu KC. A narrative review of hydrogen oncology: from real world survey to real world evidence. *Med Gas Res*. 2020;10:130-133.
32. Kim GH, Choi SY, Oh TI, et al. IDH1(R132H) causes resistance to HDAC inhibitors by increasing NANOG in glioblastoma cells. *Int J Mol Sci*. 2019;20:2679.
33. Rahman M, Kresak J, Yang C, et al. Analysis of immunobiologic markers in primary and recurrent glioblastoma. *J Neurooncol*. 2018;137:249-257.
34. Xu Q, Ahmed AK, Zhu Y, et al. Oncogenic MicroRNA-20a is downregulated by the HIF-1 $\alpha$ /c-MYC pathway in IDH1 R132H-mutant glioma. *Biochem Biophys Res Commun*. 2018;499:882-888.
35. Viswanath P, Radoul M, Izquierdo-Garcia JL, et al. Mutant IDH1 gliomas downregulate phosphocholine and phosphoethanolamine synthesis in a 2-hydroxyglutarate-dependent manner. *Cancer Metab*. 2018;6:3.
36. Oguz H, Yildizgoren MT. Ozone therapy for the treatment of recurrent pigmented villonodular synovitis of the knee. *Med Gas Res*. 2020;10:142-143.
37. Womeldorff M, Gillespie D, Jensen RL. Hypoxia-inducible factor-1 and associated upstream and downstream proteins in the pathophysiology and management of glioblastoma. *Neurosurg Focus*. 2014;37:E8.
38. Balamurugan K. HIF-1 at the crossroads of hypoxia, inflammation, and cancer. *Int J Cancer*. 2016;138:1058-1066.
39. Colegio OR, Chu NQ, Szabo AL, et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. *Nature*. 2014;513:559-563.
40. Miska J, Lee-Chang C, Rashidi A, et al. HIF-1 $\alpha$  is a metabolic switch between glycolytic-driven migration and oxidative phosphorylation-driven immunosuppression of Tregs in glioblastoma. *Cell Rep*. 2019;27:226-237.e4.
41. Minami T, Matsumura N, Sugimoto K, et al. Hypoxia-inducing factor (HIF)-1 $\alpha$ -derived peptide capable of inducing cancer-reactive cytotoxic T lymphocytes from HLA-A24(+) patients with renal cell carcinoma. *Int Immunopharmacol*. 2017;44:197-202.
42. Schaeffer HJ, Weber MJ. Mitogen-activated protein kinases: specific messages from ubiquitous messengers. *Mol Cell Biol*. 1999;19:2435-2444.
43. Kiefer F, Tibbles LA, Lassam N, Zanke B, Iscove N, Woodgett JR. Novel components of mammalian stress-activated protein kinase cascades. *Biochem Soc Trans*. 1997;25:491-498.
44. Chang L, Karin M. Mammalian MAP kinase signalling cascades. *Nature*. 2001;410:37-40.
45. Roux PP, Blenis J. ERK and p38 MAPK-activated protein kinases: a family of protein kinases with diverse biological functions. *Microbiol Mol Biol Rev*. 2004;68:320-344.
46. Wagner EF, Nebreda AR. Signal integration by JNK and p38 MAPK pathways in cancer development. *Nat Rev Cancer*. 2009;9:537-549.
47. Kuang W, Deng Q, Deng C, Li W, Shu S, Zhou M. Hepatocyte growth factor induces breast cancer cell invasion via the PI3K/Akt and p38 MAPK signaling pathways to up-regulate the expression of COX2. *Am J Transl Res*. 2017;9:3816-3826.
48. Limami Y, Pinon A, Leger DY, et al. The P2Y2/Src/p38/COX-2 pathway is involved in the resistance to urolic acid-induced apoptosis in colorectal and prostate cancer cells. *Biochimie*. 2012;94:1754-1763.
49. Gauthier ML, Pickering CR, Miller CJ, et al. p38 regulates cyclooxygenase-2 in human mammary epithelial cells and is activated in premalignant tissue. *Cancer Res*. 2005;65:1792-1799.
50. Hashemi Goradel N, Najafi M, Salehi E, Farhood B, Mortezaee K. Cyclooxygenase-2 in cancer: a review. *J Cell Physiol*. 2019;234:5683-5699.
51. Möbius C, Stein HJ, Spiess C, et al. COX2 expression, angiogenesis, proliferation and survival in Barrett's cancer. *Eur J Surg Oncol*. 2005;31:755-759.
52. Wu T, Han C, Lunz JG 3rd, Michalopoulos G, Shelhamer JH, Demetris AJ. Involvement of 85-kd cytosolic phospholipase A(2) and cyclooxygenase-2 in the proliferation of human cholangiocarcinoma cells. *Hepatology*. 2002;36:363-373.
53. Liao Z, Mason KA, Milas L. Cyclo-oxygenase-2 and its inhibition in cancer: is there a role? *Drugs*. 2007;67:821-845.
54. Najafi M, Ahmadi A, Mortezaee K. Extracellular-signal-regulated kinase/mitogen-activated protein kinase signaling as a target for cancer therapy: an updated review. *Cell Biol Int*. 2019;43:1206-1222.
55. Wu M, Guan J, Li C, et al. Aberrantly activated Cox-2 and Wnt signaling interact to maintain cancer stem cells in glioblastoma. *Oncotarget*. 2017;8:82217-82230.
56. Cook PJ, Thomas R, Kingsley PJ, et al. Cox-2-derived PGE2 induces Id1-dependent radiation resistance and self-renewal in experimental glioblastoma. *Neuro Oncol*. 2016;18:1379-1389.
57. Wang R, Wu L. The chemical modification of KCa channels by carbon monoxide in vascular smooth muscle cells. *J Biol Chem*. 1997;272:8222-8226.
58. DiChiara TJ, Reinhart PH. Redox modulation of hslco Ca<sup>2+</sup>-activated K<sup>+</sup> channels. *J Neurosci*. 1997;17:4942-4955.
59. Wang ZW, Nara M, Wang YX, Kotlikoff MI. Redox regulation of large conductance Ca(2+)-activated K<sup>+</sup> channels in smooth muscle cells. *J Gen Physiol*. 1997;110:35-44.
60. Tang XD, Daggett H, Hanner M, et al. Oxidative regulation of large conductance calcium-activated potassium channels. *J Gen Physiol*. 2001;117:253-274.

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