Tatiana Helena Rech^{1,2}, Geisiane Custódio¹, Leonardo Viliano Kroth³, Sabrina Frighetto Henrich⁴, Édison Moraes Rodrigues Filho^{2,5}, Daisy Crispim^{1,6}, Cristiane Bauermann Leitão^{1,6}

 Postgraduate Program in Medical Sciences: Endocrinology, Universidade Federal do Rio Grande do Sul - Porto Alegre (RS), Brazil.
Intensive Care Unit, Hospital de Clínicas de Porto Alegre, Universidade Federal do Rio Grande do Sul - Porto Alegre (RS), Brazil.
Renal Transplant Unit, Hospital São Lucas -Porto Alegre (RS), Brazil.
Intensive Care Unit, Hospital São Vicente de Paulo - Passo Fundo (RS), Brazil.
Intensive Care Unit, Hospital Dom Vicente Scherer - Porto Alegre (RS), Brazil.
Endocrine Division, Hospital de Clínicas de Pato Alegre Universidade Federal do Rio Grande

Porto Alegre, Universidade Federal do Rio Grande do Sul - Porto Alegre (RS). Brazil.

Conflicts of interest: None.

Submitted on June 19, 2018 Accepted on December 10, 2018

Corresponding author:

Tatiana Helena Rech Hospital de Clínicas de Porto Alegre Universidade Federal do Rio Grande do Sul Rua Ramiro Barcelos, 2.350, prédio 12, 4º andar Zip code: 90035-903 - Porto Alegre (RS), Brazil E-mail: threch@hcpa.edu.br

Responsible editor: Glauco Adrieno Westphal

DOI: 10.5935/0103-507X.20190009

Brain death-induced cytokine release is not associated with primary graft dysfunction: a cohort study

A liberação de citocinas induzida pela morte cerebral não está associada à disfunção primária do enxerto: um estudo de coorte

ABSTRACT

Objective: To examine the association between donor plasma cytokine levels and the development of primary graft dysfunction of organs transplanted from deceased donors.

Methods: Seventeen deceased donors and the respective 47 transplant recipients were prospectively included in the study. Recipients were divided into two groups: group 1, patients who developed primary graft dysfunction; and group 2, patients who did not develop primary graft dysfunction. Donor plasma levels of TNF, IL-6, IL- 1β , and IFN- γ assessed by ELISA were compared between groups.

Results: Sixty-nine organs were retrieved, and 48 transplants were performed. Donor plasma cytokine levels did not differ between groups (in pg/mL): TNF, group 1: 10.8 (4.3 - 30.8) versus group 2: 8.7 (4.1 - 33.1), p = 0.63; IL-6, group 1: 1617.8 (106.7 - 5361.7) versus group 2: 922.9 (161.7 - 5361.7), p = 0.56; IL-1β, group 1: 0.1 (0.1 -126.1) versus group 2: 0.1 (0.1 - 243.6), p = 0.60; and IFN-γ, group 1: 0.03 (0.02 - 0.2) versus group 2: 0.03 (0.02 - 0.1), p = 0.93). Similar findings were obtained when kidney transplants were analyzed separately.

Conclusion: In this sample of transplant recipients, deceased donor plasma cytokines TNF, IL-6, IL-1 β , and IFN- γ were not associated with the development of primary graft dysfunction.

Keywords: Brain death; Inflammation; Cytokines; Primary graft dysfunction; Deceased donor; Transplantation

۲

INTRODUCTION

Brain death (BD) leads to an inflammatory condition associated with adverse outcomes in organ transplantation in experimental^(1,2) and clinical settings.⁽³⁾ Kidney grafts from HLA-mismatched living donors are known to perform better than kidneys from deceased donors.⁽³⁾ Brain death-induced inflammatory activity is characterized by the upregulation of plasma cytokines, as demonstrated in previous studies by our group⁽⁴⁻⁶⁾ and, together with other important factors, plays a role in the development of primary graft dysfunction (PGD). This inflammatory trigger adversely affects organ function and is one of the possible pathways associated with clinical outcomes of transplants.^(7,8) Kusaka et al. compared rat models of BD to controls and showed a dense

inflammatory infiltrate in the glomerular tubules of brain-dead animals.⁽⁹⁾ In line with this finding, Contreras et al. demonstrated that BD-induced inflammatory activity had an adverse impact on islet function in rats, increasing apoptosis in beta cells.⁽¹⁰⁾

Primary graft dysfunction is a common complication of deceased donor organ transplantation. This dysfunction is associated with increased risk of graft loss in the first 36 months of follow-up, as well with increased length of hospital stay⁽¹¹⁾ and increased costs.⁽¹²⁾ The association between BD and PGD, however, is not fully understood. It is supposed that by activating the inflammatory cascade, BD can be a key component of ischemia-reperfusion injury, an effect that might be even more pronounced in organs from expanded criteria donors. Interestingly, there is an association for the development of PGD across different organs transplanted from the same donor.⁽¹³⁾

The present study was designed to examine the association between donor plasma tumour necrosis factor (TNF), interleukin-6 (IL-6), interleukin-1 β (IL-1 β), and interferon-gamma (IFN- γ) and the development of PGD of organs transplanted from deceased donors.

METHODS

Deceased donors and transplant recipients

The study protocol was approved by the Ethics Committee (number 13-060) at Hospital de Clínicas de Porto Alegre, in the city of Porto Alegre, and three other participating transplant centers: Hospital São Lucas and Hospital Dom Vicente Scherer, both in Porto Alegre, and Hospital São Vicente de Paulo, in Passo Fundo, in agreement with the Helsinki declaration of 1975. All institutions are in Rio Grande do Sul, the southernmost state of Brazil. Informed consent was obtained from the patients' legal representatives. Brain death was assessed independently by two physicians and was based on the following criteria: coma with complete unresponsiveness, absence of brain stem reflexes, apnea test, and confirmatory test with absence of cerebral blood flow according to the Brazilian law.⁽¹⁴⁾ From November 2010 to December 2011, brain-dead patients older than 18 years admitted to the intensive care unit at Hospital de Clínicas de Porto Alegre were prospectively included in the study after the first clinical examination consistent with BD. Blood

samples were collected at study entry. Organ recipients were identified from a crossover list provided by the regional organ distribution center. Clinical and laboratory data were recorded for brain-dead donors and transplant recipients.

Allograft dysfunction was defined as follows: (1) renal: requirement for dialysis during the first week after transplant;⁽¹⁵⁾ (2) liver: primary nonfunction during the first week after transplant leading to retransplantation or death or initial poor function characterized by aspartate aminotransferase > 2,000IU/L, serum bilirubin > 10mg/ dL, or prothrombin time >16 seconds within 2 - 7 days after transplant;⁽¹⁶⁾ (3) lung: development of severe hypoxemia, lung edema and radiographic opacities compatible with acute respiratory distress syndrome during the first 3 days after transplant;⁽¹⁷⁾ and (4) heart: need for mechanical support, such as external ventricular assist device, aortic counterpulsation pump, and extracorporeal membrane oxygenation, in the first 3 days after transplant or retransplantation/death during the first 30 days.⁽¹⁸⁾

Plasma TNF, IL-6, IL-1 β , and IFN- γ quantification

A 20mL whole blood sample was collected in a siliconecoated tube (Vacutainer[®]) for each brain-dead donor and centrifuged at 2,500g for 10 minutes at 4°C. Plasma was separated and immediately stored at -80°C until analysis. Circulating levels of TNF, IL-6, IL-1 β , and IFN- γ were assessed by enzyme-linked immunosorbent assay (ELISA) using commercially available kits with primary polyclonal antibodies following the manufacturer's instructions (detection levels: TNF, 0.7 - 518pg/mL; IL-6: 20 -5000pg/mL; IL-1 β : 0.35 - 1166pg/mL; and IFN- γ : 0.03 - 30pg/mL; Biosource Europe S.A., Nivelles, Belgium).

Statistical analysis

Categorical variables were expressed as percentages. Quantitative data were expressed as the mean and standard deviation (SD) if normally distributed. Variables with skewed distributions were log-transformed before analysis and expressed as median and minimum-maximum. Groups were compared using Student's t test, chi-square test or Fisher's exact test, as appropriate. Spearman's rank correlation was used to assess correlations between different quantitative variables. A sample size of 32 organ transplant recipients (16 patients with PGD and 16 patients without PGD) was required to detect a difference of at least one SD in TNF log,⁽⁴⁾ considering a power of 80% and an alpha-error of 5%. Values were considered to be statistically significant if p < 0.05. All statistical analyses were performed using Statistical Package for Social Science (SPSS), version 18.0 (Chicago, IL).

RESULTS

A total of 69 organs were retrieved from 17 deceased donors (a mean of 4.05 organs per donor): 34 kidneys, 17 pancreases, 13 livers, four lungs, and one heart. All pancreases were used for research purposes, and one liver and two lungs were discarded due to technical problems. Forty-eight transplants were performed in 47 patients (one patient underwent a simultaneous liver-kidney transplant). The characteristics of 38 transplant recipients (data were not available for nine patients) and 17 deceased donors included in the study are summarized in table 1. Briefly, 70% of donors were men with a mean \pm SD age of 54 \pm 11 years; stroke was the leading cause of BD (76.5%) followed by anoxic encephalopathy (23.5%). All deceased donors required vasopressor support, and sepsis was present in 41%.

Primary graft dysfunction occurred in 52.6% of the transplant recipients. To analyze donor characteristics potentially related to PGD, we divided recipients into two groups: group 1, patients who developed PGD; and group 2, patients who did not develop PGD. Blood samples were obtained from donors with a median of 12 hours (10 - 18 hours), and the plasma cytokine values were then compared between the two groups of transplant recipients to evaluate the effects of BD-induced inflammatory activity on the outcome of transplants. The results were as follows: TNF, group 1: 10.8 (4.3 - 30.8) versus group 2: 8.7 (4.1 - 33.1) pg/mL, p = 0.63; IL-6, group 1: 1617.8 (106.7 - 5361.7) versus group 2: 922.9 (161.7 - 5361.7) pg/mL, p = 0.56; IL-1 β , group 1: 0.1 (0.1 - 126.1) versus group 2: 0.1 (0.1 - 243.6) pg/mL, p = 0.60; and IFN-y, group 1: 0.03 (0.02 - 0.2) versus group 2: 0.03 (0.02 - 0.1) pg/mL, p = 0.93). Donor plasma cytokine values did not differ between groups (Figure 1).

When kidney transplants were analyzed separately (n=28), donor age (group 1: 58 ± 9.6 *versus* group 2: 48

 \pm 13.1 years, p = 0.035), duration of ventilatory support (group 1: 96 [48 - 240] versus group 2: 48 [9 - 114] hours, p = 0.003), and length of hospital stay before BD diagnosis (group 1: 5.5 [2 - 14] versus group 2: 2 [1 - 9] hours, p = 0.019) were different between the two groups, but plasma sodium (group 1: 156 ± 8.4 versus group 2: 158 ± 12.4 mEq/L, p = 0.6) and final creatinine (group 1: 1.3 ± 0.6 versus group 2: 1.3 ± 0.4 mg/dL, p = 0.6) were not different. Additionally, plasma cytokine levels were not different between groups: TNF, group 1: 10.8 (4.3-30.8) versus group 2: 15.5 (5.2 - 33.0) pg/mL, p = 0.32; IL-6, group 1: 2312.7 (106-5361.7) versus group 2: 922.9 $(161.7 - 5000) \text{ pg/mL}, \text{ p} = 0.63; \text{IL-1}\beta, \text{ group } 1: 0.1 (0.1)$ -126.1) versus group 2: 0.1 (0.1 - 243.6) pg/mL, p = 0.70; and IFN-y, group 1: 0.04 (0.02 - 0.21) versus group 2: 0.06 (0.02 - 0.11) pg/mL, p = 0.29. The rate of delayed graft function (DGF) in kidney transplant recipients was 60.7%.

Subsequently, we divided patients into two groups, above and below cytokine median values (median TNF = 9.8pg/mL; median IL-6 = 923pg/mL; median IL-1 β = 0.1pg/mL; and median IFN- γ = 0.04pg/mL) and we tested the association with PGD development. However, no association was detected (TNF, p = 0.63; IL-6, p = 0.59; IL-1 β , p = 0.50; and IFN- γ , p = 0.85). Additionally, we used 193pg/mL as a cut-off point for IL-6 but found no differences in PGD development (p = 0.62).

In a logistic regression model with PGD as the dependent factor and donor age, duration of ventilatory support, cold ischemia time, TNF, and IL-6 as cofactors, cold ischemia time was the only variable associated with PGD development (odds ratio - OR = 0.85, 95% confidence interval - 95%CI 0.74 - 0.98, p = 0.032).

We also tested correlations between donor plasma cytokines and clinical variables. There was a moderate positive correlation between TNF, IL-6, and IL-1 β and donor age (TNF: r = 0.35 p = 0.021; IL-6: r = 0.45, p = 0.002; and IL-1 β : r = 0.41, p = 0.006). Additionally, a moderate positive correlation was found between plasma sodium levels and TNF (r = 0.36, p = 0.018), but there was no correlation with the other cytokines (IL-6: r = 0.28, p = 0.07; IL-1 β : r = -0.16, p = 0.29; and IFN- γ : r = 0.12, p = 0.1).

Characteristics	All transplant recipients (n = 38)	With graft dysfunction $(n = 20)$	Without graft dysfunction $(n = 18)$	p value
Donor characteristics				
Age (years)	54 ± 11	56 ± 11.2	50 ± 11.4	0.09
Plasma sodium (mEq/L)	158 ± 9.7	156 ± 8.2	158 ± 12.1	0.54
Final creatinine (mg/dL)	1.2 ± 0.5	1.2 ± 0.6	1.2 ± 0.4	0.92
Duration of ventilatory support (days)	3 (1 - 10)	4 (1 - 10)	2 (1 - 10)	0.07
LOS before BD diagnosis (days)	4 (1 - 14)	5.5 (1 - 14)	2.5 (1 - 13)	0.09
Recipient characteristics				
Age (years)	53 ± 14	56 ± 10	49 ± 17	0.15
Male	30 (79)	17 (44.7)	13 (34.3)	0.43
Transplanted organ	28 kidneys, 9 livers, 1 heart	17 kidneys, 3 livers, 0 heart	11 kidneys, 6 livers, 1 heart	0.14*
Cold ischemia time (hours)	17 ± 7.6	19 ± 6.8	14 ± 8.1	0.07
LOS after transplantation (days)	23 (1 - 92)	25 (1 - 92)	22 (1 - 86)	0.74
Graft survival at 12 months	30 (81)	16 (44)	14 (37)	0.69
Patient survival at 12 months	25 (67.5)	11 (29.7)	14 (37.8)	0.29
Total mortality	6 (16.2)	3 (8.1)	3 (8.1)	1.0

Table 1 - Baseline characteristics of deceased donors and organ transplant recipients

LOS - length of stay; BD - brain death. p values refer to patients with graft dysfunction vs patients without graft dysfunction. * p value refers to kidney transplants. Results are presented as mean ± standard deviation, median and minimum - maximum, or n (%).

DISCUSSION

In this sample of recipients of organs from deceased donors, BD-induced plasma cytokine release was not associated with PGD, as donor plasma TNF, IL-6, IL-1 β , and IFN- γ levels did not differ between transplant recipients who developed PGD and those who did not.

Primary graft dysfunction is a serious complication of transplantation that results from ischemia-reperfusion injury, a process triggered by the systemic inflammatory state of the donor.⁽²⁰⁾ Patients who died within 30 days of lung transplant had elevated levels of IL-6 in biopsies prior to implantation.⁽²¹⁾ Likewise, myocardial TNF mRNA expression during organ retrieval predicted right ventricular dysfunction in heart recipients.⁽⁸⁾ Moreover, liver biopsies from deceased donors showed higher CD4 and CD8 infiltration than biopsies from living donors.⁽²²⁾ Taken together, these studies suggest an association between increased organ tissue inflammation and PGD. However, our study did not show an association between plasma cytokine levels and PGD. One possible explanation for the lack of association between systemic cytokine levels and PGD, in contrast with the findings reported for tissue levels, is that the degree of tissue inflammation is higher

than that measured in plasma, suggesting that tissue inflammation levels might be a better predictor of PGD than plasma inflammation levels. Furthermore, the high rate of DGF in kidney recipients (60.7%) observed in our sample of patients, which is consistent with that reported in another Brazilian study,⁽¹¹⁾ might have prevented us from detecting a difference between groups.

Murugan et al. recruited 30 deceased donors and analyzed the outcomes of the respective 78 transplant recipients. In this cohort of patients, higher plasma IL-6 levels but not TNF and IL-10 levels before organ procurement correlated with a trend to lower 6-month hospital-free survival in recipients.⁽¹⁹⁾ Therefore, we used the cut-off value of IL-6 suggested in their study (193pg/ mL), but no association with PGD was found for values above or below this threshold, suggesting that plasma IL-6 may not be a good predictor of early outcomes, such as PGD. In the short term, cold ischemia time and donor age appeared to be better predictors of outcome, as demonstrated previously^(11,23,24) and supported by our findings.

In a previous study, we demonstrated an upregulation of plasma TNF and IL-6 in deceased donors compared to controls.⁽⁴⁾ Interestingly, in the study by Murugan



Figure 1 - Deceased donor plasma cytokine levels determined by ELISA in transplant recipients with and without primary graft dysfunction. (A) Tumour necrosis factor (pg/mL). (B) Interleukin-6 (pg/mL). (C) Interleukin-1β (pg/mL). (D) Interferon-gamma (pg/mL). A *t* test was used for statistical analysis. Graphs represent median and interquartile range. Dots and asterisks represent outliers. TNF - tumour necrosis factor; PGD - primary graft dysfunction; IL-6 - interleukin-1β; INF - interferon.

et al., plasma concentrations of IL-6 immediately before organ procurement were lower in donors treated with corticosteroids than in untreated donors.⁽¹⁹⁾ Likewise, Kotsch et al., in a randomized controlled trial, showed that methylprednisolone therapy in deceased donors reduces inflammation in the donor liver and improves outcome after liver transplantation.⁽²⁵⁾ However, we did not find an association between plasma cytokine levels and liver PGD.

Elevated plasma IL-6 levels have been associated with a poorer prognosis in a variety of critical care settings⁽²⁶⁻²⁸⁾ and with lower organ yield in transplantation settings.⁽¹⁹⁾ However, to the best of our knowledge, this study is the first to evaluate donor plasma TNF, IL-6, IL-1 β , and IFN- γ levels as predictors of PGD development in organs transplanted from brain-dead donors. This study had several limitations. First, the sample size was calculated to detect a difference of one SD in TNF log between brain-dead and control patients without brain-dead in a previous study⁽⁴⁾ and, in fact, may be underpowered to detect differences in PDG development between groups of transplant recipients. However, when estimating the sample size required to detect a difference in the present study, a sample size of at least 770 organ transplant

recipients would be necessary. Second, we measured plasma cytokines only at the time of organ procurement, which may have been late in the inflammatory process, as cytokines peak earlier after BD. Third, we believe that a time course with earlier time points, such as 1, 2 and 6 hours, and up to 12 hours, might provide more consistent information about inflammation in brain-dead donors and its association with PGD development.

CONCLUSION

Primary graft dysfunction is a predictor of worse shortand long-term outcomes after transplantation. In this respect, detecting clinical and laboratory variables that can accurately predict the development of primary graft dysfunction would be clinically relevant. Plasma cytokines can be easily and quickly measured, but the small sample size and the single time point measurement of plasma cytokines in our study preclude a conclusion regarding the association of donor plasma TNF, IL-6, IL-1 β , and IFN- γ values with development of primary graft dysfunction. Then, the role of inflammatory cytokines, as a possible pathway associated with primary graft dysfunction development, should be the focus of investigation of larger studies.

RESUMO

Objetivo: Examinar a associação entre os níveis de citocinas no plasma do doador e o desenvolvimento de disfunção primária do enxerto de órgãos transplantados a partir de doadores falecidos.

Métodos: Foram incluídos no estudo de forma prospectiva 17 doadores falecidos e os respectivos 47 pacientes receptores de transplante. Os receptores foram divididos em dois grupos: grupo 1, de pacientes que desenvolveram disfunção primária do enxerto, e grupo 2, de pacientes que não desenvolveram disfunção primária do enxerto. Os níveis de TNF, IL-6, IL-1 β , e IFN- γ , avaliados por meio de ELISA, foram comparados entre os grupos.

Resultados: Obtiveram-se 69 órgãos, sendo realizados 48 transplantes. Os níveis plasmáticos de citocinas nos doadores

Authors' contributions

TH Rech participated in study conception and design, data acquisition, analysis and interpretation of data, statistical analysis, drafting and revision of the manuscript. G Custódio, LV Kroth, S Henrich, and EM Rodrigues Filho participated in data acquisition. D Crispim participated in study conception and revised the manuscript. CB Leitão participated in study conception and design, interpretation of data, statistical analysis and revised the manuscript. TH Rech is the guarantor of this work and, as such, has full access to all data and takes responsibility for the integrity of the data and the accuracy of the data analyses.

ACKNOWLEDGMENTS

This work was supported by *Fundo de Incentivo à Pesquisa e Ensino* of *Hospital de Clínicas de Porto Alegre* and by *Fundação de Amparo à Pesquisa do Estado do Rio Grande do Sul -* FAPERGS (Edital FAPERGS/CNPq 12/2014 - PRONEX).

Daisy Crispim and Cristiane Bauermann Leitão are recipients of fellowships from *Conselho Nacional de Desenvolvimento Científico e Tecnológico* (CNPq PQ-1D).

We thank the regional organ distribution center for providing the donor-recipient crossover list.

não diferiram entre os grupos (em pg/mL): TNF no grupo 1, com 10,8 (4,3 - 30,8) *versus* no grupo 2, com 8,7 (4,1 - 33,1), com valor de p = 0,63; IL-6 no grupo 1: 1.617,8 (106,7 -5.361,7) *versus* no grupo 2: 922,9 (161,7 - 5.361,7), com p = 0,56; IL-1 β , no grupo 1: 0,1 (0,1 - 126,1) *versus* no grupo 2: 0,1 (0,1 - 243,6), com p = 0,60; e IFN- γ , no grupo 1: 0,03 (0,02 - 0,2) *versus* no grupo 2: 0,03 (0,02 - 0,1), p = 0,93). Obtivemos resultados similares ao examinar separadamente os casos de transplante renal.

Conclusão: Nesta amostra de receptores de transplante, os níveis plasmáticos das citocinas TNF, IL-6, IL-1β e IFN-γ nos doadores não se associaram com o desenvolvimento de disfunção primária do enxerto.

Descritores: Morte encefálica; Inflamação; Citocinas; Disfunção primária do enxerto; Cadáver; Transplantes

REFERENCES

- Pratschke J, Wilhelm MJ, Kusaka M, Laskowski I, Tilney NL. A model of gradual onset brain death for transplant-associated studies in rats. Transplantation. 2000;69(3):427-30.
- Li S, Korkmaz S, Loganathan S, Radovits T, Hegedüs P, Karck M, et al. Shortand long-term effects of brain death on post-transplant graft function in a rodent model. Interact Cardiovasc Thorac Surg. 2015;20(3):379-86.
- Terasaki PI, Cecka JM, Gjertson DW, Takemoto S. High survival rates of kidney transplants from spousal and living unrelated donors. N Engl J Med. 1995;333(6):333-6.
- Rech TH, Crispim D, Rheinheimer J, Barkan SS, Osvaldt AB, Grezzana Filho TJ, et al. Brain death-induced inflammatory activity in human pancreatic tissue: a case-control study. Transplantation. 2014;97(2):212-9.
- Schwarz P, Custódio G, Rheinheimer J, Crispim D, Leitão CB, Rech TH. Brain death-induced inflammatory activity is similar to sepsis-induced cytokine release. Cell Transplant. 2018;27(10):1417-24.
- Custódio G, Schwarz P, Crispim D, Moraes RB, Czepielewski M, Leitão CB, et al. Association between vitamin D levels and inflammatory activity in brain death: A prospective study. Transpl Immunol. 2018;48:65-9.
- Fisher AJ, Donnelly SC, Hirani N, Haslett C, Strieter RM, Dark JH, et al. Elevated levels of interleukin-8 in donor lungs is associated with early graft failure after lung transplantation. Am J Respir Crit Care Med. 2001;163(1):259-65.
- Birks EJ, Owen VJ, Burton PB, Bishop AE, Banner NR, Khaghani A, et al. Tumor necrosis factor-alpha is expressed in donor heart and predicts right ventricular failure after human heart transplantation. Circulation. 2000;102(3):326-31.
- Kusaka M, Pratschke J, Wilhelm MJ, Ziai F, Zandi-Nejad K, Mackenzie HS, et al. Activation of inflammatory mediators in rat renal isografts by donor brain death. Transplantation. 2000;69(3):405-10.
- Contreras JL, Eckstein C, Smyth CA, Sellers MT, Vilatoba M, Bilbao G, et al. Brain death significantly reduces isolated pancreatic islet yields and functionality in vitro and in vivo after transplantation in rats. Diabetes. 2003;52(12):2935-42.
- Helfer MS, Vicari AR, Spuldaro F, Gonçalves LF, Manfro RC. Incidence, risk factors, and outcomes of delayed graft function in deceased donor kidney transplantation in a Brazilian center. Transplant Proc. 2014;46(6):1727-9.
- Croome KP, Hernandez-Alejandro R, Chandok N. Early allograft dysfunction is associated with excess resource utilization after liver transplantation. Transplant Proc. 2013;45(1):259-64.
- Oto T, Excell L, Griffiths AP, Levvey BJ, Bailey M, Marasco S, et al. Association between primary graft dysfunction among lung, kidney and heart recipients from the same multiorgan donor. Am J Transplant. 2008;8(10):2132-9.
- 14. Brasil. Presidência da República. Casa Civil. Subchefia para Assuntos Jurídicos. Lei nº 9434, de 4 de fevereiro de 1997. Dispõe sobre a remoção de órgãos, tecidos e partes do corpo humano para fins de transplante e tratamento e dá outras providências. Brasília (DF). 1997.

- Saidi RF, Elias N, Kawai T, Hertl M, Farrell ML, Goes N, et al. Outcome of kidney transplantation using expanded criteria donors and donation after cardiac death kidneys: realities and costs. Am J Transplant. 2007;7(12):2769-74.
- Coelho MP, Afonso RC, Hidalgo R, Felga G, Almeida MD, Della-Guardia B, et al. Results of retransplantation for primary nonfunction in a single center. Transplant Proc. 2011;43(1):174-6.
- 17. Lee JC, Christie JD. Primary graft dysfunction. Clin Chest Med. 2011;32(2):279-93.
- Yusen RD, Edwards LB, Kucheryavaya AY, Benden C, Dipchand AI, Goldfarb SB, et al. The Registry of the International Society for Heart and Lung Transplantation: Thirty-second Official Adult Lung and Heart-Lung Transplantation Report--2015; Focus Theme: Early Graft Failure. J Heart Lung Transplant. 2015;34(10):1264-77.
- Murugan R, Venkataraman R, Wahed AS, Elder M, Hergenroeder G, Carter M, Madden NJ, Powner D, Kellum JA; HIDonOR Study Investigators. Increased plasma interleukin-6 in donors is associated with lower recipient hospital-free survival after cadaveric organ transplantation. Crit Care Med. 2008;36(6):1810-6.
- Avlonitis VS, Wigfield CH, Golledge HD, Kirby JA, Dark JH. Early hemodynamic injury during donor brain death determines the severity of primary graft dysfunction after lung transplantation. Am J Transplant. 2007;7(1):83-90.
- Kaneda H, Gutierrez C, Perrot M, Yamane M, Quadri S, Arenovich T, et al. Pre-implantation multiple citokine mRNA expression analysis in donor lung grafts predicts survival after lung transplantation in humans. Journal Heart Lung Transplant. 2004;23(2 Suppl):S49-50.
- Jassem W, Koo DD, Cerundolo L, Rela M, Heaton ND, Fuggle SV. Leukocyte infiltration and inflammatory antigen expression in cadaveric and livingdonor livers before transplant. Transplantation. 2003;75(12):2001-7.
- Debout A, Foucher Y, Trébern-Launay K, Legendre C, Kreis H, Mourad G, et al. Each additional hour of cold ischemia time significantly increases the risk of graft failure and mortality following renal transplantation. Kidney Int. 2015;87(2):343-9.
- 24. Sert I, Colak H, Tugmen C, Dogan SM, Karaca C. The effect of cold ischemia time on delayed graft function and acute rejection in kidney transplantation. Saudi J Kidney Dis Transpl. 2014;25(5):960-6.
- 25. Kotsch K, Ulrich F, Reutzel-Selke A, Pascher A, Faber W, Warnick P, et al. Methylprednisolone therapy in deceased donors reduces inflammation in the donor liver and improves outcome after liver transplantation: a prospective randomized controlled trial. Ann Surg. 2008;248(6):1042-50.
- Corrêa TD, Pereira AJ, Brandt S, Vuda M, Djafarzadeh S, Takala J, et al. Time course of blood lactate levels, inflammation, and mitochondrial function in experimental sepsis. Crit Care. 2017;21(1):105.
- Calfee CS, Janz DR, Bernard GR, May AK, Kangelaris KN, Matthay MA, et al. Distinct molecular phenotypes of direct vs indirect ARDS in singlecenter and multicenter studies. Chest. 2015;147(6):1539-48.
- Pfeiffer D, Roßmanith E, Lang I, Falkenhagen D. miR-146a, miR-146b, and miR-155 increase expression of IL-6 and IL-8 and support HSP10 in an in vitro sepsis model. PLoS One. 2017;12(6):e0179850.