


# Gut microbiota crosstalk mechanisms are key in pulmonary hypertension: The involvement of melatonin is instrumental too

Steve Jacobs<sup>1</sup> | Carmen Payne<sup>1</sup> | Sara Shaboodien<sup>1</sup> | Thato Kgatla<sup>1</sup> |  
Amy Pretorius<sup>1</sup> | Chrisstoffel Jumaar<sup>1</sup> | Olakunle Sanni<sup>1</sup> |  
Ghazwan Butrous<sup>2</sup> | Gerald Maarman<sup>1</sup> 

<sup>1</sup>CARMA: Centre for Cardio-Metabolic Research in Africa, Division of Medical Physiology, Department of Biomedical Sciences, Faculty of Medicine & Health Sciences, Stellenbosch University, Cape Town, South Africa

<sup>2</sup>School of Pharmacy, Imperial College of London, University of Kent, Canterbury, UK

## Correspondence

Gerald Maarman, CARMA: Centre for Cardio-Metabolic Research in Africa, Division of Medical Physiology, Department of Biomedical Sciences, Faculty of Medicine & Health Sciences, Stellenbosch University, 3rd floor, BMRI Bldg, Room 3044, Francie van Zijl Ave, Tygerberg, Cape Town 8000, South Africa.

Email: [gmaarman@sun.ac.za](mailto:gmaarman@sun.ac.za)

## Funding information

Faculty of Medicine & Health Sciences, Stellenbosch University, Grant/Award Number: K2443; South African Medical Research Council, Grant/Award Number: 49692; National Research Foundation of South Africa, Grant/Award Number: 35671; Harry Crossley Foundation, Grant/Award Number: K2443; South African Rooibos Council, Grant/Award Number: 56221

## Abstract

The microbiota refers to a plethora of microorganisms with a gene pool of approximately three million, which inhabits the human gastrointestinal tract or gut. The latter, not only promotes the transport of nutrients, ions, and fluids from the lumen to the internal environment but is linked with the development of diseases including coronary artery disease, heart failure, and lung diseases. The exact mechanism of how the microbiota achieves crosstalk between itself and distant organs/tissues is not clear, but factors released to other organs may play a role, like inflammatory and genetic factors, and now we highlight melatonin as a novel mediator of the gut-lung crosstalk. Melatonin is present in high concentrations in the gut and the lung and has recently been linked to the pathogenesis of pulmonary hypertension (PH). In this comprehensive review of the literature, we suggest that melatonin is an important link between the gut microbiota and the development of PH (where suppressed melatonin-crosstalk between the gut and lungs could promote the development of PH). More studies are needed to investigate the link between the gut microbiota, melatonin and PH. Studies could also investigate whether microbiota genes play a role in the epigenetic aspects of PH. This is relevant because, for example, dysbiosis (caused by epigenetic factors) could reduce melatonin signaling between the gut and lungs, reduce subcellular melatonin concentrations in the gut/lungs, or reduce melatonin serum levels secondary to epigenetic factors. This area of research is largely unexplored and further studies are warranted.

## KEYWORDS

gut microbiota, inflammation, melatonin, pulmonary hypertension, signaling pathways

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.

© 2023 The Authors. *Pulmonary Circulation* published by John Wiley & Sons Ltd on behalf of Pulmonary Vascular Research Institute.

## INTRODUCTION

The microbiota comprises myriad microorganisms including protozoans, archaea, fungi, and bacteria<sup>1</sup> that inhabit the human gastrointestinal tract or gut.<sup>2</sup> Gut microbiota contains microorganisms with a gene pool of approximately three million genes.<sup>3,4</sup> Moreover, gut microbiota contains digestive enzymes that help the body metabolize animal and plant carbohydrates,<sup>5</sup> and it can produce short-chain fatty acids that support beta-oxidation.<sup>6</sup> These features of the microbiota allow it to code for thousands of physiological factors that are kept in homeostatic balance through microbiome-host interactions<sup>7</sup> whereas an imbalance can also contribute to some diseases.<sup>8</sup> Gut microbiota can release these factors (which could be inflammatory, genetic, or metabolic) into the circulation where they can reach and impact<sup>3,4</sup> other parts of the body<sup>9</sup> describe in this paper as crosstalk.<sup>10</sup> This is observed for example, when short-chain fatty acids (butyrate, propionate, and acetate) are produced by bacterial fermentation of indigestible polysaccharides and triggers the release of the satietogenic hormone glucagon-like peptide-1 from enteroendocrine L-cells.<sup>11</sup> These hormones travel through the body via blood circulation to the hypothalamus where they can regulate ingestive behavior.<sup>11</sup> On the other hand, propionate, butyrate, and acetate modulate the metabolism and function of these tissues.<sup>12</sup> The gut microbiota is instrumental in the homeostasis of nutrient metabolism and energy production, by supporting oxidative metabolic pathways via this crosstalk action.<sup>13</sup> Needless to

say, a homeostatic balance exists between “good and bad” organisms, to promote good health.<sup>14</sup> Herein lies two important concepts, eubiosis,<sup>15</sup> which occurs when good organisms prevail over the bad, and dysbiosis, which is the disbalance in the microbiota that predisposes to sickness.<sup>15</sup> Dysbiosis may occur due to disturbances in the microbiome caused by various lifestyle and environmental factors.<sup>16–19</sup> Also with regard to dysbiosis,<sup>20,21</sup> the literature supports the notion that it can contribute to the development and progression of heart and lung diseases.<sup>19,22</sup>

## THE ROLE OF GUT MICROBIOTA IN MAINTAINING IN HEALTH

The gut microbiota plays a major role in human health through its ability to influence immune and metabolic health, neuro- and cardiovascular-related functions.<sup>23</sup> Organisms that form part of the gut microbiota<sup>24</sup> are too many to discuss in this paper, but we present an abridged list in Table 1. Some of the most well-known species include *Clostridium spp.*, *Lactobacillus reuteri*, *Enterococcus faecium*, *Bifidobacterium bifidum*, and *Escherichia coli*.<sup>25</sup> These organisms are believed to enter the human host at birth, via the amniotic and maternal fluids.<sup>26</sup> A study by Putignani et al.<sup>27</sup> demonstrated that the normal structure, composition and thus functioning of the gut microbiota promotes overall health. This suggests the importance of maintaining a balance in microbiota composition to ensure overall good health. *Clostridium*

**TABLE 1** A brief overview of phyla, classes, genus, and species of microorganisms that form part of the gut microbiota.

Phylum	Class	Genus	Species
Firmicutes	Bacilli	Lactobacillus	<i>Lactobacillus reuteri</i>
		Staphylococcus	<i>Staphylococcus leei</i>
		Enterococcus	<i>Enterococci faecium</i>
	Clostridia	Clostridium	<i>Clostridium spp.</i>
		Ruminococcus	<i>Ruminococcus faecis</i>
Bacteroidetes	Bacteroidia	Bacteroides	<i>Bacteroides vulgatis</i>
		Prevotella	<i>Prevotella spp.</i>
Actinobacteria	Actinobacteria	Bifidobacterium	<i>Bifidobacterium bifidum</i> <i>Bifidobacterium longum</i>
Proteobacteria	Delta proteobacteria	Desulfovibrio	<i>Desulfovibrio intestinalis</i>
	Gamma proteobacteria	Escherichia	<i>Escherichia coli</i>
Fusobacteria	Fusobacteriia	Fusobacterium	<i>Fusobacterium nucleatum</i>
Verrucomicrobia	Verrucomicrobiae	Akkermansia	<i>Akkermansia muciniphila</i>

spp, one predominant part of the gut microbiota, has been reported to be essential in maintaining good health. They have been associated with therapeutic effects on intestinal and systemic health, probiotic ability, and the ability to counteract the effects of inflammation and other dysfunctions of the gut.<sup>28</sup> *Lactobacillus* spp, known for their lactic acid-producing ability are commonly used as probiotics and can be found in different parts of the body including the gut, female reproductive system, and so forth.<sup>29</sup> The effects of these microorganisms include colonization resistance against potential disease-causing bacteria.<sup>30</sup> A study by Mu et al.<sup>31</sup> outlined several benefits of *L. reuteri* and these included antimicrobial activity, inhibition of pathogenic activity, anti-inflammatory effects, and so forth. *Bifidobacteria* spp, are also an important group of the gut microflora and have been associated with immune system stimulation and the production of short-chain fatty acids.<sup>32</sup> The examples of some species that make up the gut microflora share similar effects, showing their ability to influence chemical processes that occur and molecules the function within the body, immunity, pathogen-colonization resistance and thus, the normal functioning of the body.<sup>29</sup>

## THE ROLE OF GUT MICROBIOTA IN CARDIOVASCULAR DISEASES

Gut microbiota is essential for maintaining the uptake of essential nutrients and immune responses, and it can influence how the host responds to pathologic conditions.<sup>16,33–35</sup> Alterations in gut microbiota can have detrimental effects on human health and studies have reported several of these effects including dysfunctions in the intestinal barrier and systemic health, cognitive impairment, COVID-19 severity, and heart disease.<sup>36–39</sup> Cardiovascular diseases (CVDs) are a leading cause of death worldwide and has high morbidity and mortality rates<sup>19,35,40,41</sup> Multiple risk factors contribute to the development of CVDs such as obesity, diabetes, hypertension, and dyslipidaemia that result in permanent damage to the cardiovascular system.<sup>35,40,42</sup> Among these detrimental risk factors, abnormal immune regulation, and metabolic disorders play an important role in the progression of CVDs.<sup>19,40</sup> In recent years, studies have found that alterations in the gut microbiome have been linked to CVDs.<sup>16,33,34</sup> Atherosclerosis is a chronic inflammatory disease that is considered a key factor in the onset and progression of CVD.<sup>33,34,40,43</sup> Several studies have highlighted the presence of an altered gut microbiota in patients with atherosclerosis and coronary artery disease.<sup>42,44,45</sup> Bacterial DNA present in

atherosclerotic plaques correlates with the presence of number of white blood cells in atherosclerosis plaques<sup>43</sup> and microbial species differ significantly between atherosclerotic patients and healthy individuals.<sup>46,47</sup>

Given the instrumental role of gut microbiota in CVDs, the opposite is also true, which is that it could serve as a potential target for antihypertensive therapies.<sup>19</sup> Here, the importance of a particular metabolite produced by gut microbiota, trimethylamine-N-oxide (TMAO) has been highlighted.<sup>40,43</sup> TMAO is produced through the digestion of choline, phosphatidylcholine, and carnitine, found in most animal products and a few plant products.<sup>19,34,48,49</sup> The increase of TMAO strongly influences multiple risk factors for CVD, such as hypercholesterolemia and hypertension.<sup>19,34,42,49</sup> Increased TMAO also reduces cholesterol clearance from the body and associates with the production of atherosclerotic plaques<sup>19,50</sup> (Table 2). Similarly, a strong association has been found between hypertension and TMAO production, data from a meta-analysis demonstrate that individuals with higher TMAO are more likely to develop hypertension<sup>51,52</sup> (Table 2). Therefore, TMAO levels have considerable biomarker potential CVDs.<sup>40,53</sup> However, much remain to be better understood regarding the full extent of gut microbiota's involvement in the development of treatment of CVDs.<sup>54</sup>

## THE ROLE OF GUT MICROBIOTA IN PULMONARY HYPERTENSION (PH)

PH is defined as a mean pulmonary artery pressure  $\geq 20$  mmHg, when diagnosed with right heart catheterization.<sup>60,61</sup> Characteristic features include sustained vasoconstriction, vascular remodeling, and thrombosis formation.<sup>62</sup> Pathologic triggers are believed to prime cells in the pulmonary arterioles to excessive proliferation that leads to narrowing and obliteration of the vessel lumen, and thus, elevated pulmonary artery pressure.<sup>63</sup> There is a growing body of evidence from both preclinical and clinical studies demonstrating that gut microbiota and its by-products play a role in the development of PH.<sup>11,64,65</sup> The full process is complex and poorly understood, but an important finding is that microorganisms of the gut are also present in the lungs,<sup>10</sup> and these include *Bacteroidetes* and *Firmicutes*, *Bacteroidetes*, *Firmicutes*, and *Proteobacteria*.<sup>66</sup>

### The *Firmicutes* to *Bacteroidetes* (F/B) ratio in PH

Anaerobic *Firmicutes* and *Bacteroidetes* make up >90% of the total bacterial species in healthy individuals.<sup>56</sup>

**TABLE 2** A summary of key studies that highlights gut microbiota signatures in the context of cardiovascular disease and PH.

Gut microbiota signature	Model	Findings	References
Cardiovascular diseases			
	<i>Humans</i>	The increase in TMAO strongly influences multiple risk factors for cardiovascular diseases, such as hypercholesterolemia and hypertension	[19, 34, 42, 49]
		Increased TMAO also reduces cholesterol clearance from the body and associates with the production of atherosclerotic plaques	[19, 50]
		A strong association has been found between hypertension and TMAO production, data from a meta-analysis demonstrate that individuals with higher TMAO are more likely to develop hypertension	[51, 52]
PH			
	<i>Monocrotaline-induced rats</i>	Elevated <i>Firmicutes-Bacteroidetes</i>	[55]
	<i>Hypoxia plus SU5416 treated rats</i>	Three-fold increase in <i>Firmicutes-to-Bacteroidetes</i> ratio. Acetate and acetate-producing bacteria to be reduced in the serum of PH rats.	[56]
	<i>ACE2 knock-in PH mice</i>	Imbalances between beneficial bacteria such as Bacteroids and SCFAs-producing bacteria, and potential pathogenic bacteria such as TMA/TMAO-associated bacteria, are in parallel with PAH, with functional changes of microbiome	[55]
	<i>Humans</i>	Increased <i>Firmicutes-Bacteroidetes</i> ratio in PH is due to a decrease in <i>Bacteroidetes</i> as well as <i>Firmicutes</i> enrichment	[57]
		No change demonstrated in alpha-microbiota diversity	[55, 56]
	<i>Humans</i>	Reduced alpha-diversity	[57–59]
	<i>Humans</i>	Reduction in genes responsible for generating the SCFA metabolite, propionate	[59]

Abbreviations: SCFA, short chain fatty-acids; TMAO, trimethylamine-N-oxide.

Characteristic findings of dysbiosis is an increased *Firmicutes-Bacteroidetes* ratio, reduction in short chain fatty-acids (SCFA) producing bacteria and decreases in acetate- and butyrate producing bacteria as well as increase in lactate producing bacteria<sup>56</sup> (Table 2). An elevated *Firmicutes-Bacteroidetes* ratio, especially due to reduced *Bacteroidetes* have been shown in both pre-clinical and clinical studies to be a feature of altered gut microbiota in PH. These findings were echoed by Sharma et al.<sup>55</sup> who also demonstrated that an increased *Firmicutes-Bacteroidetes* ratio in monocrotaline induced PH (Table 2). Callejo and colleagues demonstrated in a hypoxia plus SU5416 treated rats an almost four-fold decrease in the *Bacteroidetes* population in PH induced rats (Table 2). Zhang et al.<sup>57</sup> demonstrated that an increased *Firmicutes-Bacteroidetes* ratio in PH patients is due to a decrease in *Bacteroidetes* as well as *Firmicutes* enrichment (Table 2). Furthermore, although there is no change demonstrated in alpha-microbiota diversity in PH animal models,<sup>55,56</sup> both Li et al.<sup>58</sup> Kim et al.<sup>59</sup> and Zhang

et al.<sup>57</sup> demonstrated a reduced alpha-diversity in PH patients.

### The reduction in SCFA producing bacteria and TMAO in PH

Reduction in SCFA producing bacteria contributes to increased gut leakiness and inflammation through the release and circulation of for example, acetate.<sup>67</sup> Cajello et al.<sup>56</sup> found acetate and acetate-producing bacteria to be reduced in the serum of PH rats (Table 2). Kim et al.<sup>59</sup> also found a significant reduction in genes responsible for generating the SCFA metabolite, propionate, in patients with PH, thereby confirming the potential role of gut microbiota in the pathogenesis of PH in humans (Table 2). On the other hand, various bacterial communities are associated with TMA/TMAO production, including *Coriobacteriales* which is significantly increased in PH<sup>6</sup> (Table 2).<sup>7</sup> TMAO promotes the

proliferation and migration of pulmonary artery smooth muscle cells, by upregulating inflammatory factors.<sup>11,68</sup> Endothelial dysfunction has also been shown to be influenced by TMAO via inflammatory pathways that lead to endothelial hyperpermeability.<sup>58,69</sup> Furthermore, TMAO promotes the production of reactive oxygen species that adds to the impairment of endothelial function.<sup>58,70</sup> Poor prognosis and increased severity of disease were associated with increased TMAO in PH (Li and colleagues).

### Streptococcus as part of the microbiota, and its role in PH

Kim et al.<sup>59</sup> have demonstrated an association between *Streptococcus* and PH in humans, while animal studies have shown depleted *Streptococcus* levels.<sup>55</sup> Interestingly, *Streptococcus* has been implicated in the activation of a pro-survival kinase pathway mediated via ERK and PI3K.<sup>71</sup> These kinases are involved in the pathogenesis of PH through the induction and proliferation of pulmonary smooth muscle cells.<sup>72</sup> *Streptococcus* also promote serotonin production<sup>73</sup> which has previously been implicated in the smooth muscle cell contraction and proliferation contribution to PH pathogenesis.

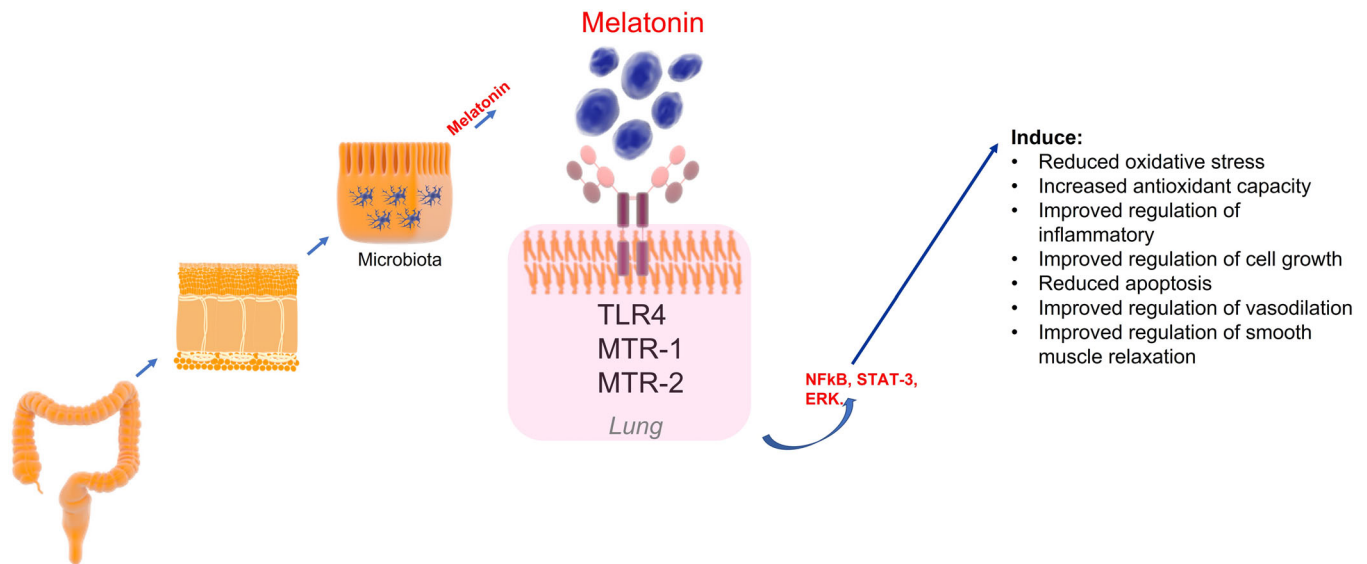
### CROSSTALK BETWEEN MICROBIOTA AND LUNGS IN PH VIA INFLAMMATORY MEDIATORS

Crosstalk between the lung and gut, also termed the *gut-lung axis*, comprises a key mechanism through which the gut microbiota influences the pathogenesis of respiratory diseases.<sup>66,74,75</sup> The gut microbiota plays an important role in the regulation of the immune system,<sup>76,77</sup> specifically, systemic and lung immunity, via the regulation of T-cell differentiation, the migration and apoptosis of immune cells, and the activation of toll-like receptor signaling.<sup>78</sup> The microbiota play a role in modulating immune function in the prevention and progression of chronic respiratory diseases.<sup>79,80</sup> Therefore, there is a crosstalk between the lungs and gut microbiota. This cross-talk is important as it can influence the pathogenesis and treatment of lung diseases,<sup>79–83</sup> and furthermore, the amount and function of immune cells activated through the gut-lung crosstalk, influences the pharmacokinetics and bioavailability of therapeutics for lung diseases.<sup>84</sup> Also important is that when there is dysbiosis of the gut microbiota, clearance of lung macrophages becomes compromised in diseases

like for example, lung tuberculosis, leading to an imbalance in cytokine production, and the activation and migration of T and B cells become impaired.<sup>83</sup> When the gut microbiota is in a state of equilibrium, there is immune function homeostasis which equates to homeostatic cytokine production, correct clearance of lung macrophages, and the proper activation and migration of T and B cells.<sup>83</sup> The bulk of studies therefore confirm the crosstalk between gut and lungs, and the instrumental roles of gut microbiota in the pathogenesis of lung diseases.<sup>80</sup>

Although the mature respiratory and gastrointestinal tracts are different in several ways, one must recall that they originate from the same embryonic structure and thus, they do also display similarities in structure and microbial colonization.<sup>85,86</sup> Both derive from the endoderm and consist of columnar epithelial cells with projections (microvilli in the gut and cilia in the respiratory tract) that serve as a physical barrier as well as forming part of the immune system in collaboration with lymphoid tissue, allowing both organs to influence each other's immune responses and creating somewhat of an immunological relationship.<sup>64</sup> However, impaired gut permeability and systemic migration of gut microbiota is believed to be associated with systemic inflammatory responses that may act as a trigger factor for PH pathogenesis.<sup>87</sup> The gut-lung crosstalk<sup>87</sup> occurs in both directions (to and fro), and is achieved when cytokines, endotoxins, hormones, and metabolites are circulated between the gut and the lungs.<sup>88</sup> This happens, for example, during lung inflammation, when inflammatory mediators (interleukin-6, interferon-gamma, tumor necrosis factor-alpha) are circulated to the gut, where lymphocytes are activated and transported back to the lungs.<sup>89</sup> Thus, the gut microbiota is closely linked with the pulmonary system to provide protection in the presence of lung pathology.

However, one must keep in mind that the crosstalk is a homeostatic process, and an imbalance (abnormal inflammatory processes)<sup>22,66</sup> may also contribute to PH. Furthermore, inflammatory, and immunological processes are key to the pathogenesis of PH.<sup>16,90,91</sup> In experimental PH, rats display dysbiosis, which excessively increases their *Firmicutes*-to-*Bacteroidetes* ratio and reduces their acetate-producing bacterial genera.<sup>92</sup> This likely happens when gut dysbiosis increases gut permeability and allows the transport of gut bacteria and bacterial endotoxin to the pulmonary vasculature where it could trigger PH<sup>92,93</sup> (Figure 1). Some investigators have suggested that these abnormal inflammatory processes may contribute to the development of PH,<sup>92</sup> and this interaction between



**FIGURE 1** Depicts how the gut microbiota produce and release certain factors (inflammatory, genetic, and melatonin) as crosstalk between the gut and the lungs.

the gut and lungs occur via lung-gut crosstalk (Figure 1). The crosstalk is further mediated by bacterial lipopolysaccharides and interleukin-6 that are circulated from the gut to the lungs.<sup>92</sup>

### CROSSTALK BETWEEN MICROBIOTA AND LUNGS IN PH VIA GENETIC FACTORS

Abnormal gut microbiota early in life, is linked to the development of lung diseases later in life.<sup>94–96</sup> This has been ascribed to the insufficient maturation of gut microbiota, the excessive use of antibiotics<sup>87,97,98</sup> a genetically inherent risk.<sup>94–96</sup> For example, susceptibility to autoimmune diseases is under strong genetic control by class-II human leukocyte antigen (HLA) allele combinations<sup>99</sup> Core microbiome and beta diversity (i.e., the ratio between regional and local species diversity)<sup>100</sup> differs with HLA risk group and genotype, and in addition, there is also the presence of inherent protective HLA haplotypes that play a role.<sup>99</sup> These are important factors to consider, as an inherent genetic risk can play a role in determining which patients are more susceptible to diseases. This may have relevance to PH too, HLA haplotypes (HLA risk) have been linked with the pathogenesis of PH in patients, and based on the work of Russel and colleagues,<sup>100</sup> gut microbiota are involved in different HLA risk groups (Figure 1). This is an important form of crosstalk between gut and lungs, and surely plays a role in PH pathogenesis.

### CROSSTALK BETWEEN MICROBIOTA AND LUNGS IN PH VIA MELATONIN

#### Melatonin

Melatonin (N-acetyl-5-methoxytryptamine) is a hormone produced by the pineal gland in the vertebrate brain, but also occurs in fruits and vegetables.<sup>101,102</sup> It is present in all bodily cells and play a role in regulation the circadian rhythm, and can induce anti-inflammatory, anticancer, antioxidant, and vasodilatory effects<sup>103</sup> In the brain, melatonin is mainly produced at night in response to changes in environmental light which leads to serotonin-N-acetyltransferase activity increasing significantly.<sup>104–107</sup> Serotonin-N-acetyltransferase is responsible for converting 5-hydroxytryptamine into N-acetylserotonin, is methylated by acetylserotonin-O-methyltransferase to form melatonin.<sup>108</sup> These moieties are also modulated by noradrenergic and neuropeptidergic projections to the pineal gland as well as light information that is transmitted from the retina to the pineal gland via the suprachiasmatic nucleus in the hypothalamus. In humans, melatonin secretion begins shortly after sunset and peaks in the middle of the night. Although melatonin is the main hormone produced by the pineal gland, extra-pineal sources include the retina, bone marrow cells, platelets, skin, lymphocytes, cerebellum, and especially the gut, among other.<sup>109</sup> The concentration of melatonin in gut can be as high as 10–100 higher than in plasma and up to 400 times higher than in the pineal gland.<sup>108,110,111</sup> The most well-known molecular pathways through which

melatonin acts, is the activation of G-protein coupled membrane receptors, MT1 (or Mel1a or MTNR1A) and MT2 (or Mel1b or MTNR1B).<sup>108,112,113</sup> MT1 melatonin receptor activation inhibits cAMP in target cells and is associated with reproductive and metabolic functions, as well as vasoconstriction.<sup>104,114</sup> On the other hand, MT2 receptor activation leads to cAMP and cGMP inhibition, plays a role in regulating circadian rhythms and dopamine release in the retina, and also contributes to vasodilation.<sup>113</sup> Melatonin interact with the cardiopulmonary system via various signaling pathways.<sup>115–118</sup> It does so by directly exerting its function over a receptor dependant pathway including membrane receptors type 1 (MT1) and type 2 (MT2) as well as the retinoid-related orphan nuclear receptors RZR and ROR $\alpha$ .<sup>116,119</sup> It also exerts its beneficial effects indirectly as a free radical scavenger by protecting cells against oxidative stress, reducing the generation of free radical and reactive oxygen species.<sup>120–122</sup> Melatonin regulates blood pressure by centrally modulating the baroreflex set point, and the sympathetic and parasympathetic vascular tone.<sup>123</sup>

## The involvement of melatonin in PH

In patients with PH, levels of melatonin are lower than in healthy patients and this negatively correlates with an increase in cytokine levels.<sup>62</sup> Also, lower levels of melatonin correlate with a worse long-term survival rate of PAH patients.<sup>124</sup> Several animal studies have provided evidence advocating the beneficial role of endogenous melatonin. Hung et al.<sup>125</sup> outlined that melatonin can improve vascular resistance and oxidative injury in hypoxia-induced PH. By using a rat model, they demonstrated how right ventricular systolic pressure, vascular remodeling, markers of oxidative stress and inflammation, TNF-alpha, and malondialdehyde, were significantly decreased when chronic hypoxic rats were given pharmacological doses of melatonin. They also observed that the nitric oxide bioavailability has increased.<sup>125</sup> Torres et al.<sup>126</sup> reported similar results, including enhanced pulmonary vascular function in new-born sheep with PH. An in vitro model showed attenuated proliferation and inflammation in pulmonary arterial muscle cells in the presence of melatonin while a rat model showed improvements in right ventricular systolic pressure and pulmonary vascular remodeling.<sup>127</sup> Zhang and colleagues demonstrate how melatonin significantly ameliorated hypoxia-induced thickness of the pulmonary artery wall and improved the remodeling of blood vessels. They also demonstrated that PH is associated with decreased serum melatonin levels.<sup>62</sup> While decreased levels of melatonin were linked to poor

prognosis and increased mortality, Cai et al.<sup>124</sup> reported that at the time of diagnosis, serum melatonin levels can be used to anticipate the clinical outcome of PH. Maarman and colleagues in their monocrotaline rat model demonstrated how melatonin therapy at both therapeutic concentration and at concentration as found in food protects against PH by reducing oxidative stress, reduced right ventricular hypertrophy, improving right ventricular function and cardiac interstitial fibrosis. They however noted that the dose of melatonin required for an equivalent effect in humans would be substantially higher than what was the dose used in clinical studies at the time.<sup>128</sup> In concert, these studies demonstrate the key role of melatonin in PH. However, there is a shortage of literature that describes melatonin as a key player in gut-lung crosstalk, and how it can potentially modulate the pathogenesis or treatment of PH.

## The roles of melatonin in promoting gut microbiota reprogramming

Melatonin contributes to improved metabolism of lipids<sup>1</sup> and remodeling of the gut microbiota composition in both animals and humans.<sup>1,2</sup> Yin et al.<sup>129</sup> demonstrated in a gut microbiota transplant mouse model that was subjected to a high fat diet, displayed an accumulation of intestinal lipids as well as dysbiosis. In this study, melatonin reduced lipid concentration and the reversal of dysbiosis. The *Firmicutes-Bacteroidetes* ratio was also significantly increased and improved synthesis of acetic acid upon supplementation with melatonin. These findings therefore support the important role melatonin plays in reprogramming of the gut microbiota.<sup>129</sup> In their study, Lui et al.<sup>130</sup> investigated the role of melatonin in alleviating dysbiosis caused by aflatoxin B1 and its effects on the intestinal/liver axis, they found that the alterations due to aflatoxin B1 exposure to be corrected by melatonin administration. Melatonin reduced the relative abundance of *Firmicutes* and the *Firmicutes-Bacteroides* ratio and elevated that of *Bacteroides* at the phylum level of the taxonomic units, while also decreasing the relative abundance of *Clostridiales* and *Lactobacillales*. Aflatoxin B1 exposure resulted in elevations in the relative abundance of *Desulfovibrio*, *Clostridium-XIVa*, and *Lactobacillus* at the genus level, but upon exposure to melatonin these changes were reversed. It was concluded that the dysbiosis in the gut microbiota due to AFB1 was successfully reprogrammed by melatonin.<sup>130</sup>

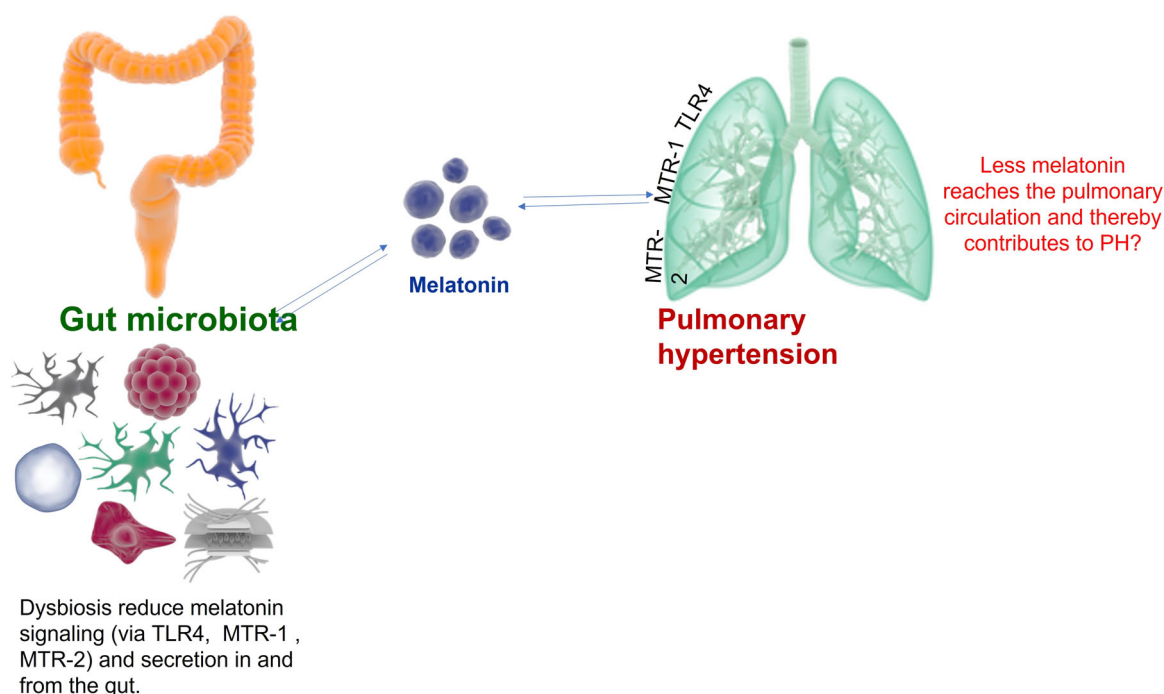
Moreover, the gut microbiota can produce and release serotonin<sup>131</sup> and tryptophan,<sup>132</sup> which are precursors for the formation of melatonin. Melatonin concentrations are 400 times higher in the gut than in the pineal gland.<sup>133,134</sup>

The gut has toll-like receptors, which are molecular patterns of antigens whose activation can stimulate innate immunity to destroy pathogens.<sup>135</sup> During the early postnatal stage, the body reduces the expression of toll-like receptors in the gut, to promote healthy gut microbiota.<sup>136</sup> In turn, the gut microbiota can trigger toll-like receptor signaling to suppress inflammatory responses that make it possible for these good microorganisms to survive in the gut.<sup>136,137</sup> This is important, because the gut expresses several receptors on its surface, including the toll-like receptors<sup>137</sup> and melatonin receptors 1, 2, and 3.<sup>138</sup> These receptors can bind melatonin<sup>139</sup> that resides within the gut and the lungs.<sup>133,134</sup> To this end, Kim et al.<sup>140</sup> showed that improved sensing of bad bacteria through toll-like receptor-4 and the regulation of bacteria through altered goblet cells and antimicrobial peptides in microbiota, is involved in the anti-colic effects of melatonin. Melatonin-induced activation of these receptors stimulates the expression and release of NFkB, STAT-3 and ERK<sup>141</sup> which are known role players in PH pathogenesis (Figure 1).

## GUT-LUNG CROSTALK IN PH VIA MELATONIN: THE MAIN HYPOTHESIS

Melatonin levels can be reduced by stress,<sup>142</sup> smoking,<sup>143</sup> or shift work,<sup>144</sup> where reduced melatonin are considered pathologic. Furthermore, low serum levels of melatonin

associates with PH. Therefore, it is likely that dysbiosis from psychological stress or other factors, may reduce levels of melatonin and contribute to the development of PH (Figure 2). Thus, dysbiosis reduces melatonin signaling (via TLR4, MTR-1, MTR-2) and secretion in and from the gut, and less melatonin reaches the lung, this limits its protective effects and predisposes to the development of PH. Considering that exogenous melatonin can improve experimental PH,<sup>60,128</sup> it is of interest to delineate the roles of melatonin as a crosstalk agent between the gut microbiota and the lungs. Patients and animal models of PH display impaired gut microbiota,<sup>59,145</sup> and it makes one wonder whether this is also associated with impaired melatonin crosstalk<sup>92</sup> Furthermore, would the latter contribute to a predisposition to develop PH? Further investigations are needed to better understand the roles and involvement of melatonin in the crosstalk between gut and lungs. Future studies could investigate the levels of melatonin in the gut and lungs, in appropriate models of PH,<sup>146</sup> as this may provide useful information regarding its involvement in the crosstalk between the gut and lungs. Using radioactive labeling of melatonin (<sup>101</sup>I-2-iodomelatonin)<sup>147</sup> could help to trace the melatonin crosstalk, over time. For example, in a rat model of PH one could measure melatonin levels in the gut, lungs and circulation before PH is induced, and measure it over a period of time. In this instance, one might be able to see how melatonin levels change in the



**FIGURE 2** Depicts how the gut microbiota produces and releases melatonin as a crosstalk between the gut and the lungs. Melatonin can activate and regulate signaling pathways that could induce processes that may underpin the development of PH.



gut, lungs, and circulation over time. A decrease melatonin levels in the gut and an increase in the circulation and lungs, may suggest a crosstalk effect, or movement of melatonin from gut to lungs during PH. This might mean that (1) a lack of melatonin contributes to PH, (2) that increased levels is a compensating effect for the existing lung damage in PH, and (3) may confirm the melatonin crosstalk in PH. However, whether these investigations are performed in animal models or patients, it is important to keep in mind that time of day and light exposure may be confounding factors that influence the levels of melatonin that is measured.<sup>147</sup>

## CONCLUSIONS

It is plausible that melatonin and its receptors in both the gut microbiota and the lungs may provide a relatively novel crosstalk between these two biological sites. Studies can investigate whether microbiota and melatonin genes play a role in the epigenetic aspects of PH, as dysbiosis (caused by epigenetic or environmental factors) could reduce melatonin levels and signaling or sub-cellular concentrations in the gut or reduce melatonin. This may help to explore the role of melatonin in the crosstalk between the gut microbiota and the lungs during PH. This area of research is largely unexplored in the context of PH and may present a good opportunity for the discovery of novel therapeutic targets or pathologic role players in PH pathogenesis.

## AUTHOR CONTRIBUTIONS

All authors meet the criteria to qualify as authors, have contributed sufficiently to the manuscript, and have seen/approved the final version of the manuscript.

## ACKNOWLEDGMENTS

Special thanks and appreciation to the Faculty of Medicine & Health Sciences, Stellenbosch University, National Research Foundation of South Africa, The South African Medical Research Council, the South African Rooibos Council, and the Harry Crossley Foundation.

## CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

## ETHICS STATEMENT

No ethical requirements were relevant for this paper.

## ORCID

Gerald Maarman  <https://orcid.org/0000-0003-3336-5594>

## REFERENCES

1. King CH, Desai H, Sylvetsky AC, LoTempio J, Ayanyan S, Carrie J, Crandall KA, Fochtman BC, Gasparyan L, Gulzar N, Howell P, Issa N, Krampis K, Mishra L, Morizono H, Piseigna JR, Rao S, Ren Y, Simonyan V, Smith K, VedBrat S, Yao MD, Mazumder R. Baseline human gut microbiota profile in healthy people and standard reporting template. *PLoS One*. 2019;14:1–5. <https://doi.org/10.1371/journal.pone.0206484>
2. Bull MJ, Plummer NT. Part 1: the human gut microbiome in health and disease. *Integrative Medicine*. 2014;13:17–22.
3. Goodrich JK, Waters JL, Poole AC, Sutter JL, Koren O, Blekhman R, Beaumont M, Van Treuren W, Knight R, Bell JT, Spector TD, Clark AG, Ley RE. Human genetics shape the gut microbiome. *Cell*. 2014;159:789–99. <https://doi.org/10.1016/j.cell.2014.09.053>
4. Rothschild D, Weissbrod O, Barkan E, Kurilshikov A, Korem T, Zeevi D, Costea PI, Godneva A, Kalka IN, Bar N, Shilo S, Lador D, Vila AV, Zmora N, Pevsner-Fischer M, Israeli D, Kosower N, Malka G, Wolf BC, Avnit-Sagi T, Lotan-Pompan M, Weinberger A, Halpern Z, Carmi S, Fu J, Wijmenga C, Zhernakova A, Elinav E, Segal E. Environment dominates over host genetics in shaping human gut microbiota. *Nature*. 2018;555:210–5. <https://doi.org/10.1038/nature25973>
5. Cantarel BL, Henrissat VL. B. Complex carbohydrate utilization by the healthy human microbiome. *Plos One*. 2012;7(6):1–10.
6. Dalile B, Van Oudenhove L, Vervliet B, Verbeke K. The role of short-chain fatty acids in microbiota-gut-brain communication. *Nat Rev Gastroenterol Hepatol*. 2019;16:461–78. <https://doi.org/10.1038/s41575-019-0157-3>
7. Xu F, Fu Y, Sun T, Jiang Z, Miao Z, Shuai M, Gou W, Ling C, Yang J, Wang J, Chen Y, Zheng JS. The interplay between host genetics and the gut microbiome reveals common and distinct microbiome features for complex human diseases. *Microbiome*. 2020;8:1–14. <https://doi.org/10.1186/s40168-020-00923-9>
8. Witkowski M, Weeks TL, Hazen SL. Gut microbiota and cardiovascular disease. *Circ Res*. 2020;127:553–70. <https://doi.org/10.1161/CIRCRESAHA.120.316242>
9. Rath CM, Dorrestein PC. The bacterial chemical repertoire mediates metabolic exchange within gut microbiomes. *Curr Opin Microbiol*. 2012;15:147–54. <https://doi.org/10.1016/j.mib.2011.12.009>
10. Enaud R, Prevel R, Ciarlo E, Beauflis F, Wieërs G, Guery B, Delhaes L. The gut-lung axis in health and respiratory diseases: a place for inter-organ and inter-kingdom cross-talks. *Front Cell Infect Microbiol*. 2020;10:1–11. <https://doi.org/10.3389/fcimb.2020.00009>
11. Wu P, Zhu T, Tan Z, Chen S, Fang Z. Role of gut microbiota in pulmonary arterial hypertension. *Front Cell Infect Microbiol*. 2022;12:1–12. <https://doi.org/10.3389/fcimb.2022.812303>
12. Schönfeld P, Wojtczak L. Short- and medium-chain fatty acids in energy metabolism: the cellular perspective. *J Lipid Res*. 2016;57(6):943–54. <https://doi.org/10.1194/jlr.R067629>
13. Martin AM, Sun EW, Rogers GB, Keating DJ. The influence of the gut microbiome on host metabolism through the

- regulation of gut hormone release. *Front Physiol.* 2019;10:1–11. <https://doi.org/10.3389/fphys.2019.00428>
14. Ohland CL, Jobin C. Microbial activities, and intestinal homeostasis: a delicate balance between health and disease. *Cell Mol Gastroenterol Hepatol.* 2015;1:28–40. <https://doi.org/10.1016/j.jcmgh.2014.11.004>
  15. Iebba V, Totino V, Gagliardi A, Santangelo F, Cacciotti F, Trancassini M, Mancini C, Cicerone C, Corazziari E, Pantanella F, Schippa S. Eubiosis and dysbiosis: the two sides of the microbiota. *New Microbiol.* 2016;39:1–12.
  16. Nicholson JK, Holmes E, Wilson ID. Gut microorganisms, mammalian metabolism and personalized health care. *Nat Rev Microbiol.* 2005;3:431–8.
  17. Rosenfeld M, Ann Campbell L. Pathogens and atherosclerosis: update on the potential contribution of multiple infectious organisms to the pathogenesis of atherosclerosis. *Thromb Haemost.* 2011;106:858–67.
  18. Battson ML, Lee DM, Weir TL, Gentile CL. The gut microbiota as a novel regulator of cardiovascular function and disease. *J Nutr Biochem.* 2018;56:1–15.
  19. Jin M, Qian Z, Yin J, Xu W, Zhou X. The role of intestinal microbiota in cardiovascular disease. *J Cell Mol Med.* 2019;23:2343–50.
  20. Amabebe E, Robert FO, Agbalalah T, Orubu ESF. Microbial dysbiosis-induced obesity: role of gut microbiota in homeostasis of energy metabolism. *Br J Nutr.* 2020;123:1127–37. <https://doi.org/10.1017/S0007114520000380>
  21. Cheng L, Qi C, Zhuang H, Fu T, Zhang X. gutMDisorder: a comprehensive database for dysbiosis of the gut microbiota in disorders and interventions. *Nucleic Acids Res.* 2020;48:D554–60. <https://doi.org/10.1093/nar/gkz843>
  22. Anand S, Mande SS. Diet, microbiota and gut-lung connection. *Front Microbiol.* 2018;9:1–12. <https://doi.org/10.3389/fmicb.2018.02147>
  23. Li F, Lu J, Wang L, Ma X. CYP3A-mediated generation of aldehyde and hydrazine in atazanavir metabolism. *Drug Metab Dispos.* 2011;39:394–401.
  24. Kho ZY, Lal SK. The human gut microbiome—a potential controller of wellness and disease. *Front Microbiol.* 2018;9:1–23. <https://doi.org/10.3389/fmicb.2018.01835>
  25. Rinninella E, Raoul P, Cintoni M, Franceschi F, Miggiano G, Gasbarrini A, Mele M. What is the healthy gut microbiota composition? A changing ecosystem across age, environment, diet, and diseases. *Microorganisms.* 2019;7:1–22. <https://doi.org/10.3390/microorganisms7010014>
  26. Dunn AB, Jordan S, Baker BJ, Carlson NS. The maternal infant microbiome: considerations for labor and birth. *MCN Am J Matern Child Nurs.* 2017;42:318–25. <https://doi.org/10.1097/NMC.0000000000000373>
  27. Putignani L, Del Chierico F, Petrucca A, Vernocchi P, Dallapiccola B. The human gut microbiota: a dynamic interplay with the host from birth to senescence settled during childhood. *Pediatr Res.* 2014;76:2–10.
  28. Guo P, Zhang K, Ma X, He P. Clostridium species as probiotics: potentials and challenges. *J Anim Sci Biotechnol.* 2020;11(1):1–10.
  29. Walter J. Ecological role of lactobacilli in the gastrointestinal tract: implications for fundamental and biomedical research. *Appl Environ Microbiol.* 2008;74:4985–96.
  30. Orrhage K, Nord CE. Bifidobacteria and lactobacilli in human health. *Drugs Exp Clin Res.* 2000;26:95–111.
  31. Mu Q, Tavella VJ, Luo XM. Role of *Lactobacillus reuteri* in human health and diseases. *Front Microbiol.* 2018;9:1–17.
  32. Arboleya S, Watkins C, Stanton C. Gut bifidobacteria populations in human health and aging. *Front Microbiol.* 2016;7:1–9.
  33. Carding S, Verbeke K, Vipond DT, Corfe BM, Owen LJ. Dysbiosis of the gut microbiota in disease. *Microb Ecol Health Dis.* 2015;26:1–9.
  34. Novakovic M, Rout A, Kingsley T, Kirchoff R, Singh A, Verma V, Kant R, Chaudhary R. Role of gut microbiota in cardiovascular diseases. *World J Cardiol.* 2020;12:110–22.
  35. Rahman MM, Islam F, Or-Rashid MH, Mamun AA, Rahaman MS, Islam MM, Meem AFK, Sutradhar PR, Mitra S, Mimi AA, Emran TB, Fatimawali, Idroes R, Tallei TE, Ahmed M, Cavalu S. The gut microbiota (microbiome) in cardiovascular disease and its therapeutic regulation. *Front Cell Infect Microbiol.* 2022;12:1–22.
  36. Zuo T, Zhang F, Lui GCY, Yeoh YK, Li AYL, Zhan H, Wan Y, Chung ACK, Cheung CP, Chen N, Lai CKC, Chen Z, Tso EYK, Fung KSC, Chan V, Ling L, Joynt G, Hui DSC, Chan FKL, Chan PKS, Ng SC. Alterations in gut microbiota of patients with COVID-19 during time of hospitalization. *Gastroenterology.* 2020;159:944–55.
  37. He L. Alterations of gut microbiota by overnutrition impact gluconeogenic gene expression and insulin signaling. *Int J Mol Sci.* 2021;22:1–13.
  38. Olson CA, Iñiguez AJ, Yang GE, Fang P, Pronovost GN, Jameson KG, Rendon TK, Paramo J, Barlow JT, Ismagilov RF, Hsiao EY. Alterations in the gut microbiota contribute to cognitive impairment induced by the ketogenic diet and hypoxia. *Cell Host Microbe.* 2021;29:1378–92.
  39. Sun W, Du D, Fu T. Alterations of the gut microbiota in patients with severe chronic heart failure. *Front Microbiol.* 2021;12:1–10.
  40. Zhou W, Cheng Y, Zhu P. Implication of gut microbiota in cardiovascular diseases. *Oxid Med Cell Longev.* 2020;1:1–14.
  41. Brown JM, Hazen SL. Microbial modulation of cardiovascular disease. *Nat Rev Microbiol.* 2018;16:171–81.
  42. Kumar D, Mukherjee SS, Chakraborty R, Roy RR, Pandey A, Patra S, Dey S. The emerging role of gut microbiota in cardiovascular diseases. *Indian Heart J.* 2021;73:264–72.
  43. Koren O, Spor A, Felin J, Fåk F, Stombaugh J, Tremaroli V, Behre CJ, Knight R, Fagerberg B, Ley RE, Bäckhed F. Human oral, gut, and plaque microbiota in patients with atherosclerosis. *Proc Natl Acad Sci.* 2011;108:4592–8.
  44. Chan YK, Brar MS, Kirjavainen PV, Chen Y, Peng J, Li D, Leung FC, El-Nezami H. High fat diet induced atherosclerosis is accompanied with low colonic bacterial diversity and altered abundances that correlates with plaque size, plasma A-FABP and cholesterol: a pilot study of high fat diet and its intervention with *Lactobacillus rhamnosus* GG (LGG) or telmisartan in ApoE<sup>-/-</sup> mice. *BMC Microbiol.* 2016;16:1–13.
  45. Cui L, Zhao T, Hu H. Association study of gut flora in coronary heart disease through high-throughput sequencing. *BioMed Res Int.* 2017;1:1–10.
  46. Karlsson FH, Fåk F, Nookaew I, Tremaroli V, Fagerberg B, Petranovic D, Bäckhed F, Nielsen J. Symptomatic atherosclerosis

- is associated with an altered gut metagenome. *Nat Commun.* 2012;3:1–8.
47. Jie Z, Xia H, Zhong SL, Feng Q, Li S, Liang S, Zhong H, Liu Z, Gao Y, Zhao H, Zhang D, Su Z, Fang Z, Lan Z, Li J, Xiao L, Li J, Li R, Li X, Li F, Ren H, Huang Y, Peng Y, Li G, Wen B, Dong B, Chen JY, Geng QS, Zhang ZW, Yang H, Wang J, Wang J, Zhang X, Madsen L, Brix S, Ning G, Xu X, Liu X, Hou Y, Jia H, He K, Kristiansen K. The gut microbiome in atherosclerotic cardiovascular disease. *Nat Commun.* 2017;8:1–12.
  48. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, DuGar B, Feldstein AE, Britt EB, Fu X, Chung YM, Wu Y, Schauer P, Smith JD, Allayee H, Tang WHW, DiDonato JA, Lusis AJ, Hazen SL. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature.* 2011;472:57–63.
  49. Bennett BJ, Vallim TQA, Wang Z, Shih DM, Meng Y, Gregory J, Allayee H, Lee R, Graham M, Croke R, Edwards PA, Hazen SL, Lusis AJ. Trimethylamine-N-oxide, a metabolite associated with atherosclerosis, exhibits complex genetic and dietary regulation. *Cell Metab.* 2013;17:49–60.
  50. Charach G, Rabinovich A, Argov O. The role of bile acid excretion in atherosclerotic coronary artery disease. *Int J Vasc Med.* 2012;2012:949672. <https://doi.org/10.1155/2012/949672>
  51. Ge X, Zheng L, Zhuang R, Yu P, Xu Z, Liu G, Xi X, Zhou X, Fan H. The gut microbial metabolite trimethylamine N-oxide and hypertension risk: a systematic review and dose–response meta-analysis. *Adv Nutr.* 2020;11:66–76.
  52. Zhang WQ, Wang YJ, Zhang A. TMA/TMAO in hypertension: novel horizons and potential therapies. *J Cardiovasc Transl Res.* 2021;2021:1–8.
  53. Lam V, Su J, Koprowski S, Hsu A, Tweddell JS, Rafiee P, Gross GJ, Salzman NH, Baker JE. Intestinal microbiota determine severity of myocardial infarction in rats. *FASEB J.* 2012;26:1727–35.
  54. Kazemian N, Mahmoudi M, Halperin F, Wu JC, Pakpour S. Gut microbiota and cardiovascular disease: opportunities and challenges. *Microbiome.* 2020;8:1–17.
  55. Sharma RK, Oliveira AC, Yang T, Kim S, Zubcevic J, Aquino V, Lobaton GO, Goel R, Richards EM, Raizada MK. Pulmonary arterial hypertension-associated changes in gut pathology and microbiota. *ERJ Open Res.* 2020;6(3):1–10. <https://doi.org/10.1183/23120541.00253-2019>
  56. Callejo M, Mondejar-Parreño G, Barreira B, Izquierdo-Garcia JL, Morales-Cano D, Esquivel-Ruiz S, Moreno L, Cogolludo Á, Duarte J, Perez-Vizcaino F. Pulmonary arterial hypertension affects the rat gut microbiome. *Sci Rep.* 2018;8(1):1–10. <https://doi.org/10.1038/s41598-018-27682-w>
  57. Zhang C, Zhang T, Lu W, Duan X, Luo X, Liu S, Chen Y, Li Y, Chen J, Liao J, Zhou D, Chen X, Feng H, Gu G, Wang T, Tang H, Makino A, Zhong N, Yuan JXJ, Yang K, Wang J. Altered airway microbiota composition in patients with pulmonary hypertension. *Hypertension.* 2020;76(5):1589–99. <https://doi.org/10.1161/HYPERTENSIONAHA.120.15025>
  58. Li J, Zhao F, Wang Y, Chen J, Tao J, Tian G, Wu S, Liu W, Cui Q, Geng B, Zhang W, Weldon R, Auguste K, Yang L, Liu X, Chen L, Yang X, Zhu B, Cai J. Gut microbiota dysbiosis contributes to the development of hypertension. *Microbiome.* 2017;5:1–19.
  59. Kim S, Rigatto K, Gazzana MB, Knorst MM, Richards EM, Pepine CJ, Raizada MK. Altered gut microbiome profile in patients with pulmonary arterial hypertension. *Hypertension.* 2020;75(4):1063–71. <https://doi.org/10.1161/HYPERTENSIONAHA.119.14294>
  60. Maarman GJ, Schulz R, Sliwa K, Schermuly RT, Lecour S. Novel putative pharmacological therapies to protect the right ventricle in pulmonary hypertension: a review of current literature. *Br J Pharmacol.* 2017;174:497–511. <https://doi.org/10.1111/bph.13721>
  61. Maarman GJ, Shaw J, Allwood B. Pulmonary hypertension in majority countries: opportunities amidst challenges. *Curr Opin Pulm Med.* 2020;26:373–83. <https://doi.org/10.1097/MCP.0000000000000702>
  62. Zhang J, Lu X, Liu M, Fan H, Zheng H, Zhang S, Rahman N, Wołczyński S, Kretowski A, Li X. Melatonin inhibits inflammasome-associated activation of endothelium and macrophages attenuating pulmonary arterial hypertension. *Cardiovasc Res.* 2020;116(13):2156–69. <https://doi.org/10.1093/cvr/cvz312>
  63. Maarman GJ. Pulmonary hypertension and the right ventricle: the roles of mitochondrial reactive oxygen species in causing further right ventricular mitochondrial changes. In: Chakraborti S, Dhalla NS, Ganguly NK, editors. *Oxidative stress in heart diseases.* Singapore: Springer; 2019. p. 539–49.
  64. Budden KF, Gellatly SL, Wood DLA, Cooper MA, Morrison M, Hugenholtz P, Hansbro PM. Emerging pathogenic links between microbiota and the gut-lung axis. *Nat Rev Microbiol.* 2017;15(1):55–63. <https://doi.org/10.1038/nrmicro.2016.142>
  65. Moutsoglou DM, Tatah J, Prisco SZ, Prins KW, Staley C, Lopez S, Blake M, Teigen L, Kazmirczak F, Weir EK, Kabage AJ, Guan W, Khoruts A, Thenappan T. Pulmonary arterial hypertension patients have a proinflammatory gut microbiome and altered circulating microbial metabolites. *Am J Respir Crit Care Med.* 2023;207(6):740–56. <https://doi.org/10.1164/rccm.202203-0490OC>
  66. Marsland BJ, A T, Gollwitzer ES. The gut-lung axis in respiratory disease. *Ann Am Thorac Soc.* 2015;12(suppl 2):150–6. <https://doi.org/10.1513/AnnalsATS.201503-133AW>
  67. Kelly CJ, Zheng L, Campbell EL, Saeedi B, Scholz CC, Bayless AJ, Wilson KE, Glover LE, Kominsky DJ, Magnuson A, Weir TL, Ehrentraut SF, Pickel C, Kuhn KA, Lanis JM, Nguyen V, Taylor CT, Colgan SP. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. *Cell Host Microbe.* 2015;17(5):662–71. <https://doi.org/10.1016/j.chom.2015.03.005>
  68. Huang Y, Lin F, Tang R, Bao C, Zhou Q, Ye K, Shen Y, Liu C, Hong C, Yang K, Tang H, Wang J, Lu W, Wang T. Gut microbial metabolite trimethylamine N-oxide aggravates pulmonary hypertension. *Am J Respir Cell Mol Biol.* 2022;66(4):452–60. <https://doi.org/10.1165/rcmb.2021-0414OC>
  69. Mutengo KH, Masenga SK, Mweemba A, Mutale W, Kirabo A. Gut microbiota dependant trimethylamine N-oxide and hypertension. *Front Physiol.* 2023;14:1–13. <https://doi.org/10.3389/fphys.2023.1075641>

70. Brunt VE, Gioscia-Ryan RA, Casso AG, VanDongen NS, Ziemba BP, Sapinsley ZJ, Richey JJ, Zigler MC, Neilson AP, Davy KP, Seals DR. Trimethylamine-N-oxide promotes age-related vascular oxidative stress and endothelial dysfunction in mice and healthy humans. *Hypertension*. 2020;76(1):101–12. <https://doi.org/10.1161/HYPERTENSIONAHA.120.14759>
71. Tsay JCJ, Wu BG, Badri MH, Clemente JC, Shen N, Meyn P, Li Y, Yie TA, Lhakhang T, Olsen E, Murthy V, Michaud G, Sulaiman I, Tsirigos A, Heguy A, Pass H, Weiden MD, Rom WN, Sterman DH, Bonneau R, Blaser MJ, Segal LN. Airway microbiota is associated with upregulation of the PI3K pathway in lung cancer. *Am J Respir Crit Care Med*. 2018;198(9):1188–98. <https://doi.org/10.1164/rccm.201710-2118OC>
72. Zhang Q, Cao Y, Luo Q, Wang P, Shi P, Song C, E M, Ren J, Fu B, Sun H. The transient receptor potential vanilloid-3 regulates hypoxia-mediated pulmonary artery smooth muscle cells proliferation via PI3K/AKT signaling pathway. *Cell Proliferation*. 2018;51(3):1–13. <https://doi.org/10.1111/cpr.12436>
73. MacLean MR. The serotonin hypothesis in pulmonary hypertension revisited: targets for novel therapies (2017 Grover Conference Series). *Pulm Circ*. 2018;8(2):1–9. <https://doi.org/10.1177/2045894018759125>
74. Bingula R, Filaire M, Radosevic-Robin N, Bey M, Berthon JY, Bernalier-Donadille A, Vasson MP, Filaire E. Desired turbulence? Gut-lung axis, immunity, and lung cancer. *J Oncol*. 2017;2017:1–15. <https://doi.org/10.1155/2017/5035371>
75. Zhu W, Wu Y, Liu H, Jiang C, Huo L. Gut–lung axis: microbial crosstalk in pediatric respiratory tract infections. *Front Immunol*. 2021;12:1–7. <https://doi.org/10.3389/fimmu.2021.741233>
76. Cuevas-Ramos G, Petit CR, Marcq I, Boury M, Oswald E, Nougayrède JP. *Escherichia coli* induces DNA damage in vivo and triggers genomic instability in mammalian cells. *Proc Natl Acad Sci*. 2010;107:11537–42.
77. He Z, Gharaibeh RZ, Newsome RC, Pope JL, Dougherty MW, Tomkovich S, Pons B, Mirey G, Vignard J, Hendrixson DR, Jobin C. *Campylobacter jejuni* promotes colorectal tumorigenesis through the action of cytolethal distending toxin. *Gut*. 2019;68:289–300.
78. Samuelson DR, Welsh DA, Shellito JE. Regulation of lung immunity and host defense by the intestinal microbiota. *Front Microbiol*. 2015;6:1–14.
79. O'Dwyer DN, Dickson RP, Moore BB. The lung microbiome, immunity, and the pathogenesis of chronic lung disease. *J Immunol*. 2016;196:4839–47.
80. Gupta N, Kumar R, Agrawal B. New players in immunity to tuberculosis: the host microbiome, lung epithelium, and innate immune cells. *Front Immunol*. 2018;9:1–10.
81. Hand TW. The role of the microbiota in shaping infectious immunity. *Trends Immunol*. 2016;37:647–58.
82. Shukla SD, Budden KF, Neal R, Hansbro PM. Microbiome effects on immunity, health and disease in the lung. *Clin Transl Immunol*. 2017;6:1–12.
83. Liu Y, Wang J, Wu C. Microbiota and tuberculosis: a potential role of probiotics, and postbiotics. *Front Nutr*. 2021;8:1–10.
84. Namasivayam S, Sher A, Glickman MS, Wipperman MF. The microbiome and tuberculosis: early evidence for cross talk. *mBio*. 2018;9:01420–18.
85. Grier A, McDavid A, Wang B, Qiu X, Java J, Bandyopadhyay S, Yang H, Holden-Wiltse J, Kessler HA, Gill AL, Huyck H, Falsey AR, Topham DJ, Scheible KM, Caserta MT, Pryhuber GS, Gill SR. Neonatal gut and respiratory microbiota: coordinated development through time and space. *Microbiome*. 2018;6(1):193. <https://doi.org/10.1186/s40168-018-0566-5>
86. Chunxi L, Haiyue L, Yanxia L, Jianbing P, Jin S. The gut microbiota and respiratory diseases: new evidence. *J Immunol Res*. 2020;2020:1–12. <https://doi.org/10.1155/2020/2340670>
87. Zhang D, Li S, Wang N, Tan HY, Zhang Z, Feng Y. The cross-talk between gut microbiota and lungs in common lung diseases. *Front Microbiol*. 2020;11:301. <https://doi.org/10.3389/fmicb.2020.00301>
88. Keely S, Talley NJ, Hansbro PM. Pulmonary-intestinal cross-talk in mucosal inflammatory disease. *Mucosal Immunol*. 2012;5:7–18. <https://doi.org/10.1038/mi.2011.55>
89. Dumas A, Bernard L, Poquet Y, Lugo-Villarino G, Neyrolles O. The role of the lung microbiota and the gut-lung axis in respiratory infectious diseases. *Cell Microbiol*. 2018;20:1–9. <https://doi.org/10.1111/cmi.12966>
90. Rabinovitch M, Guignabert C, Humbert M, Nicolls MR. Inflammation and immunity in the pathogenesis of pulmonary arterial hypertension. *Circ Res*. 2014;115:165–75. <https://doi.org/10.1161/CIRCRESAHA.113.301141>
91. Frid MG, Thurman JM, Hansen KC, Maron BA, Stenmark KR. Inflammation, immunity, and vascular remodeling in pulmonary hypertension; evidence for complement involvement? *Glob Cardiol Sci Pract*. 2020;2020:1–18. <https://doi.org/10.21542/gcsp.2020.1>
92. Ranchoux B, Bigorgne A, Hautefort A, Girerd B, Sitbon O, Montani D, Humbert M, Tcherakian C, Perros F. Gut-lung connection in pulmonary arterial hypertension. *Am J Respir Cell Mol Biol*. 2017;56:402–5.
93. Thenappan T, Goel A, Marsboom G, Fang YH, Toth PT, Zhang HJ, Kajimoto H, Hong Z, Paul J, Wietholt C, Pogoriler J, Piao L, Rehman J, Archer SL. A central role for CD68(+) macrophages in hepatopulmonary syndrome. *Am J Respir Crit Care Med*. 2011;183:1080–91. <https://doi.org/10.1164/rccm.201008-1303OC>
94. Abrahamsson TR, Jakobsson HE, Andersson AF, Björkstén B, Engstrand L, Jenmalm MC. Low gut microbiota diversity in early infancy precedes asthma at school age. *Clin Exp Allergy*. 2014;44:842–50. <https://doi.org/10.1111/cea.12253>
95. Qian LJ, Kang SM, Xie JL, Huang L, Wen Q, Fan YY, Lu LJ, Jiang L. Early-life gut microbial colonization shapes Th1/Th2 balance in asthma model in BALB/c mice. *BMC Microbiol*. 2017;17:1–8.
96. Stokholm J, Blaser MJ, Thorsen J, Rasmussen MA, Waage J, Vinding RK, Schoos AMM, Kunøe A, Fink NR, Chawes BL, Bønnelykke K, Brejnrod AD, Mortensen MS, Al-Soud WA, Sørensen SJ, Bisgaard H. Maturation of the gut microbiome and risk of asthma in childhood. *Nat Commun*. 2018;9:1–10. <https://doi.org/10.1038/s41467-017-02573-2>

97. McKeever TM, Lewis SA, Smith C, Hubbard R. The importance of prenatal exposures on the development of allergic disease: a birth cohort study using the West Midlands General Practice Database. *Am J Respir Crit Care Med*. 2002;166:827–32. <https://doi.org/10.1164/rccm.200202-158OC>
98. Stiemsma LT, Turvey SE. Asthma and the microbiome: defining the critical window in early life. *Allergy Asthma Clin Immunol*. 2017;13:1–9. <https://doi.org/10.1186/s13223-016-0173-6>
99. Russell JT, Roesch LFW, Ördberg M, Ilonen J, Atkinson MA, Schatz DA, Triplett EW, Ludvigsson J. Genetic risk for autoimmunity is associated with distinct changes in the human gut microbiome. *Nat Commun*. 2019;10:1–12. <https://doi.org/10.1038/s41467-019-11460-x>
100. Wagner BD, Grunwald GK, Zerbe GO, Mikulich-Gilbertson SK, Robertson CE, Zemanick ET, Harris JK. On the use of diversity measures in longitudinal sequencing studies of microbial communities. *Front Microbiol*. 2018;9:1–11. <https://doi.org/10.3389/fmicb.2018.01037>
101. Venegas C, García JA, Escames G, Ortiz F, López A, Doerrier C, García-Corzo L, López LC, Reiter RJ, Acuña-Castroviejo D. Extrapineal melatonin: analysis of its subcellular distribution and daily fluctuations. *J Pineal Res*. 2012;52:217–27. <https://doi.org/10.1111/j.1600-079X.2011.00931.x>
102. Meng X, Li Y, Li S, Zhou Y, Gan RY, Xu DP, Li HB. Dietary sources and bioactivities of melatonin. *Nutrients*. 2017;9:1–64. <https://doi.org/10.3390/nu9040367>
103. Mahmood D. Pleiotropic effects of melatonin. *Drug Res*. 2019;69:65–74. <https://doi.org/10.1055/a-0656-6643>
104. Acuña-Castroviejo D, Escames G, Venegas C, Diaz-Casado ME, Lima-Cabello E, López LC, Rosales-Corral S, Tan DX, Reiter RJ. Extrapineal melatonin: sources, regulation, and potential functions. *Cell Mol Life Sci*. 2014;71(16):2997–3025. <https://doi.org/10.1007/s00018-014-1579-2>
105. Arendt J, Aulinas A. Physiology of the pineal gland and melatonin. In: Feingold KR, Anawalt B, Blackman MR, editors. *Endotext*. MDText.com, Inc.; 2000. <https://www.ncbi.nlm.nih.gov/books/NBK550972/>
106. Brzezinski A. Melatonin in humans. *N Engl J Med*. 1997;336(3):186–95. <https://doi.org/10.1056/NEJM199701163360306>
107. Gastel JA, Roseboom PH, Rinaldi PA, Weller JL, Klein DC. Melatonin production: proteasomal proteolysis in serotonin N-acetyltransferase regulation. *Science*. 1998;279(5355):1358–60. <https://doi.org/10.1126/science.279.5355.1358>
108. Tordjman S, Chokron S, Delorme R, Charrier A, Bellissant E, Jaafari N, Fougere C. Melatonin: pharmacology, functions and therapeutic benefits. *Curr Neuropharmacol*. 2017;15(3):434–43. <https://doi.org/10.2174/1570159X14666161228122115>
109. Slominski A, Wortsman J, Tobin DJ. The cutaneous serotonergic/melatonergic system: securing a place under the sun. *FASEB J*. 2005;19(2):176–94. <https://doi.org/10.1096/fj.04-2079rev>
110. Bubenik GA. Localization, physiological significance and possible clinical implication of gastrointestinal melatonin. *Neurosignals*. 2001;10(6):350–66. <https://doi.org/10.1159/000046903>
111. Konturek SJ, Konturek PC, Brzozowska I, Pawlik M, Sliwowski Z, Cześnikiewicz-Guzik M, Kwiecień S, Brzozowski T, Bubenik GA, Pawlik WW. Localization and biological activities of melatonin in intact and diseased gastrointestinal tract (GIT). *J Physiol Pharmacol*. 2007;58(3):381–405.
112. Morgan PJ, Barrett P, Howell HE, Helliwell R. Melatonin receptors: localization, molecular pharmacology and physiological significance. *Neurochem Int*. 1994;24(2):101–46. [https://doi.org/10.1016/0197-0186\(94\)90100-7](https://doi.org/10.1016/0197-0186(94)90100-7)
113. Dubocovich ML, Markowska M. Functional MT1 and MT2 melatonin receptors in mammals. *Endocrine*. 2005;27(2):101–10. <https://doi.org/10.1385/ENDO:27:2:101>
114. Li D, Smith D, Hardeland R, Yang M, Xu H, Zhang L, Yin H, Zhu Q. Melatonin receptor genes in vertebrates. *Int J Mol Sci*. 2013;14(6):11208–23. <https://doi.org/10.3390/ijms140611208>
115. Genade S, Genis A, Ytrehus K, Huisamen B, Lochner A. Melatonin receptor-mediated protection against myocardial ischaemia/reperfusion injury: role of its anti-adrenergic actions. *J Pineal Res*. 2008;45(4):449–58. <https://doi.org/10.1111/j.1600-079X.2008.00615.x>
116. Paulis L, Simko F, Laudon M. Cardiovascular effects of melatonin receptor agonists. *Expert Opin Invest Drugs*. 2012;21(11):1661–78. <https://doi.org/10.1517/13543784.2012.714771>
117. Lochner A. Cardioprotective effect of melatonin against ischaemia/reperfusion damage. *Front Biosci*. 2013;5(1):305–15. <https://doi.org/10.2741/E617>
118. Yu L, Di W, Dong X, Li Z, Zhang Y, Xue X, Xu Y, Zhang J, Xiao X, Han J, Liu Y, Yang Y, Wang H. Melatonin protects diabetic heart against ischemia-reperfusion injury, role of membrane receptor-dependent cGMP-PKG activation. *Biochim Biophys Acta Mol Basis Dis*. 2018;1864(2):563–78. <https://doi.org/10.1016/j.bbadis.2017.11.023>
119. Hardeland R, Cardinali DP, Srinivasan V, Spence DW, Brown GM, Pandi-Perumal SR. Melatonin—a pleiotropic, orchestrating regulator molecule. *Prog Neurobiol*. 2011;93(3):350–84. <https://doi.org/10.1016/j.pneurobio.2010.12.004>
120. Boutin JA, Saunier C, Guenin SP, Berger S, Moulharat N, Gohier A, Delagrèze P, Cogé F, Ferry G. Studies of the melatonin binding site location onto quinone reductase 2 by directed mutagenesis. *Arch Biochem Biophys*. 2008;477(1):12–9. <https://doi.org/10.1016/j.abb.2008.04.040>
121. Reybier K, Perio P, Ferry G, Bouajila J, Delagrèze P, Boutin JA, Nepveu F. Insights into the redox cycle of human quinone reductase 2. *Free Radic Res*. 2011;45(10):1184–95. <https://doi.org/10.3109/10715762.2011.605788>
122. Tobeiha M, Jafari A, Fadaei S, Mirazimi SMA, Dashti F, Amiri A, Khan H, Asemi Z, Reiter RJ, Hamblin MR, Mirzaei H. Evidence for the benefits of melatonin in cardiovascular disease. *Front Cardiovasc Med*. 2022;9:1–39. <https://doi.org/10.3389/fcvm.2022.888319>
123. Pechanova O, Paulis L, Simko F. Peripheral and central effects of melatonin on blood pressure regulation. *Int J Mol Sci*. 2014;15(10):17920–37. <https://doi.org/10.3390/ijms151017920>

124. Cai Z, Klein T, Geenen LW, Tu L, Tian S, van den Bosch AE, de Rijke YB, Reiss IKM, Boersma E, Duncker DJ, Boomars KA, Guignabert C, Merkus D. Lower plasma melatonin levels predict worse long-term survival in pulmonary arterial hypertension. *J Clin Med.* 2020;9(5):1–13. <https://doi.org/10.3390/jcm9051248>
125. Hung M, Yeung H, Lau C, Poon A, Tipoe G, Fung M. Melatonin attenuates pulmonary hypertension in chronically hypoxic rats. *Int J Mol Sci.* 2017;18(6):1–10. <https://doi.org/10.3390/ijms18061125>
126. Torres F, González-Candia A, Montt C, Ebensperger G, Chubretovic M, Serón-Ferré M, Reyes RV, Llanos AJ, Herrera EA. Melatonin reduces oxidative stress and improves vascular function in pulmonary hypertensive newborn sheep. *J Pineal Res.* 2015;58:362–73. <https://doi.org/10.1111/jpi.12222>
127. Jin H, Wang Y, Zhou L, Liu L, Zhang P, Deng W, Yuan Y. Melatonin attenuates hypoxic pulmonary hypertension by inhibiting the inflammation and the proliferation of pulmonary arterial smooth muscle cells. *J Pineal Res.* 2014;57(4):442–50. <https://doi.org/10.1111/jpi.12184>
128. Maarman G, Blackhurst D, Thienemann F, Blauwet L, Butrous G, Davies N, Sliwa K, Lecour S. Melatonin as a preventive and curative therapy against pulmonary hypertension. *J Pineal Res.* 2015;59:343–53. <https://doi.org/10.1111/jpi.12263>
129. Yin J, Li Y, Han H, Chen S, Gao J, Liu G, Wu X, Deng J, Yu Q, Huang X, Fang R, Li T, Reiter RJ, Zhang D, Zhu C, Zhu G, Ren W, Yin Y. Melatonin reprogramming of gut microbiota improves lipid dysmetabolism in high-fat diet-fed mice. *J Pineal Res.* 2018;65(4):e12524. <https://doi.org/10.1111/jpi.12524>
130. Liu S, Kang W, Mao X, Ge L, Du H, Li J, Hou L, Liu D, Yin Y, Liu Y, Huang K. Melatonin mitigates aflatoxin B1-induced liver injury via modulation of gut microbiota/intestinal FXR/liver TLR4 signaling axis in mice. *J Pineal Res.* 2022;73(2):e12812. <https://doi.org/10.1111/jpi.12812>
131. Strandwitz P. Neurotransmitter modulation by the gut microbiota. *Brain Res.* 2018;1693:128–33. <https://doi.org/10.1016/j.brainres.2018.03.015>
132. Kaur H, Bose C, Mande SS. Tryptophan metabolism by gut microbiome and gut-brain-axis: an in silico analysis. *Front Neurosci.* 2019;13. <https://doi.org/10.3389/fnins.2019.01365>
133. Raikhlin NT, KVETNOY IM, Tolkachev VN. Melatonin may be synthesised in enterochromaffin cells. *Nature.* 1975;255:344–5.
134. Messner M, Huether G, Lorf T, Ramadori G, Schwörer H. Presence of melatonin in the human hepatobiliary-gastrointestinal tract. *Life Sci.* 2001;69:543–51.
135. Lazar V, Ditu LM, Pircalabioru GG, Gheorghie I, Curutiu C, Holban AM, Picu A, Petcu L, Chifiriuc MC. Aspects of gut microbiota and immune system interactions in infectious diseases, immunopathology, and cancer. *Front Immunol.* 2018;9:1830. <https://doi.org/10.3389/fimmu.2018.01830>
136. Francino M. Early development of the gut microbiota and immune health. *Pathogens.* 2014;3:769–90. <https://doi.org/10.3390/pathogens3030769>
137. Hug H, Mohajeri M, La Fata G. Toll-like receptors: regulators of the immune response in the human gut. *Nutrients.* 2018;10:203. <https://doi.org/10.3390/nu10020203>
138. Chen CQ. Distribution, function and physiological role of melatonin in the lower gut. *World J Gastroenterol.* 2011;17:3888–98. <https://doi.org/10.3748/wjg.v17.i34.3888>
139. Maarman GJ, Reiter RJ. Melatonin therapy for blunt trauma and strenuous exercise: a mechanism involving cytokines, NFκB, Akt, MAFBX and MURF-1. *J Sports Sci.* 2018;36:1897–901. <https://doi.org/10.1080/02640414.2018.1424491>
140. Kim SW, Kim S, Son M, Cheon JH, Park YS. Melatonin controls microbiota in colitis by goblet cell differentiation and antimicrobial peptide production through Toll-like receptor 4 signalling. *Sci Rep.* 2020;10:2232. <https://doi.org/10.1038/s41598-020-59314-7>
141. Moniruzzaman M, Ghosal I, Das D, Chakraborty SB. Melatonin ameliorates H2O2-induced oxidative stress through modulation of Erk/Akt/NFκB pathway. *Biol Res.* 2018;51(1):17. <https://doi.org/10.1186/s40659-018-0168-5>
142. Ito Y, Iida T, Yamamura Y, Teramura M, Nakagami Y, Kawai K, Nagamura Y, Teradaira R. Relationships between salivary melatonin levels, quality of sleep, and stress in young Japanese females. *Int J Tryptophan Res.* 2013;6s1:S11760. <https://doi.org/10.4137/ijtr.S11760>
143. Ozguner F, Koyu A, Cesur G. Active smoking causes oxidative stress and decreases blood melatonin levels. *Toxicol Ind Health.* 2005;21:21–6. <https://doi.org/10.1191/0748233705th211oa>
144. Dumont M, Lanctôt V, Cadieux-Viau R, Paquet J. Melatonin production and light exposure of rotating night workers. *Chronobiol Int.* 2012;29:203–10. <https://doi.org/10.3109/07420528.2011.647177>
145. Hong W, Mo Q, Wang L, Peng F, Zhou Y, Zou W, Sun R, Liang C, Zheng M, Li H, Zhao D, Gao M, Pu J, Li B, Ran P, Peng G. Changes in the gut microbiome and metabolome in a rat model of pulmonary arterial hypertension. *Bioengineered.* 2021;12:5173–83. <https://doi.org/10.1080/21655979.2021.1952365>
146. Maarman G, Lecour S, Butrous G, Thienemann F, Sliwa K. A comprehensive review: the evolution of animal models in pulmonary hypertension research; are we there yet? *Pulm Circ.* 2013;3:739–56. <https://doi.org/10.1086/674770>
147. Kennaway DJ. Measuring melatonin by immunoassay. *J Pineal Res.* 2020;69:12657. <https://doi.org/10.1111/jpi.12657>

**How to cite this article:** Jacobs S, Payne C, Shaboodien S, Kgatla T, Pretorius A, Jumaar C, Sanni O, Butrous G, Maarman G. Gut microbiota crosstalk mechanisms are key in pulmonary hypertension: the involvement of melatonin is instrumental too. *Pulm Circ.* 2023;13:e12277. <https://doi.org/10.1002/pul2.12277>