### Check for updates

### OPEN ACCESS

EDITED BY Juan Pablo de Rivero Vaccari, University of Miami, United States

### REVIEWED BY Fumitaka Shimizu, Yamaguchi University, Japan Nadine Ahmed Kerr,

CORRESPONDENCE

Siming Yang ysm0117@126.com Xiaobing Fu fuxiaobing@vip.sina.com

<sup>†</sup>These authors have contributed equally to this work

#### SPECIALTY SECTION

This article was submitted to Brain Disease Mechanisms, a section of the journal Frontiers in Molecular Neuroscience

RECEIVED 08 August 2022 ACCEPTED 20 September 2022 PUBLISHED 24 October 2022

#### CITATION

Yang J, Ran M, Li H, Lin Y, Ma K, Yang Y, Fu X and Yang S (2022) New insight into neurological degeneration: Inflammatory cytokines and blood-brain barrier. *Front. Mol. Neurosci.* 15:1013933. doi: 10.3389/fnmol.2022.1013933

#### COPYRIGHT

© 2022 Yang, Ran, Li, Lin, Ma, Yang, Fu and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# New insight into neurological degeneration: Inflammatory cytokines and blood-brain barrier

# Jie Yang<sup>1,2†</sup>, Mingzi Ran<sup>1,3†</sup>, Hongyu Li<sup>1,2†</sup>, Ye Lin<sup>4</sup>, Kui Ma<sup>1</sup>, Yuguang Yang<sup>2</sup>, Xiaobing Fu<sup>1\*</sup> and Siming Yang<sup>1,2\*</sup>

<sup>1</sup>Research Centre for Tissue Repair and Regeneration Affiliated to the Medical Innovation Research Department, PLA General Hospital, PLA Medical College, Beijing, China, <sup>2</sup>Department of Dermatology, 4th Medical Centre, PLA General Hospital, Beijing, China, <sup>3</sup>Department of Anaesthesiology, 4th Medical Centre, PLA General Hospital, Beijing, China, <sup>4</sup>Department of Neurology, The First Medical Centre, PLA General Hospital, Beijing, China

Neurological degeneration after neuroinflammation, such as that resulting from Alzheimer's disease (AD), stroke, multiple sclerosis (MS), and posttraumatic brain injury (TBI), is typically associated with high mortality and morbidity and with permanent cognitive dysfunction, which places a heavy economic burden on families and society. Diagnosing and curing these diseases in their early stages remains a challenge for clinical investigation and treatment. Recent insight into the onset and progression of these diseases highlights the permeability of the blood-brain barrier (BBB). The primary factor that influences BBB structure and function is inflammation, especially the main cytokines including IL-1 $\beta$ , TNF $\alpha$ , and IL-6, the mechanism on the disruption of which are critical component of the aforementioned diseases. Surprisingly, the main cytokines from systematic inflammation can also induce as much worse as from neurological diseases or injuries do. In this review, we will therefore discuss the physiological structure of BBB, the main cytokines including IL-1 $\beta$ , TNF $\alpha$ , IL-6, and their mechanism on the disruption of BBB and recent research about the main cytokines from systematic inflammation inducing the disruption of BBB and cognitive impairment, and we will eventually discuss the need to prevent the disruption of BBB.

### KEYWORDS

inflammatory cytokines, blood-brain barrier, neurological degeneration, TNF  $\!\alpha$ , IL-1  $\!\beta$ , IL-6

# Introduction

Aging body is vulnerable to chronic inflammatory conditions that cause neurological degeneration in the central nervous system (CNS), such as Alzheimer's disease (AD) (Ju and Tam, 2022). These diseases are frequently associated with high mortality and morbidity and with permanent cognitive dysfunction, which are commonly considered

a significant economic burden on society and on the families of patients. The difficulties faced in resolving these clinical problems are related to the inability of the damaged and degenerated nerve cells to repair themselves (Ren et al., 2018). Although recent research indicates that necrotic foci can be replaced by proliferative neural stem cells (NSCs) (Kuhn and Svendsen, 1999), the number and scale of the NSCs cannot make up for the entire injury to the CNS, and strategies for curing neurological diseases in the clinic remain far off. Interestingly, recent research finds that these diseases share a key pathological feature, namely, the disruption of the blood-brain barrier (BBB) (Sweeney et al., 2019; Gold et al., 2021). The disruption of BBB can be detected before the onset of neurodegeneration, and the repairing of BBB after neurodegeneration can be beneficial for the disease (Liebner et al., 2018). Therefore, studying the biological and pathological features of BBB may be a useful target for future diagnosis and treatment.

A primary cause to disrupt the BBB is uncontrolled inflammation after injuries or diseases. Under this condition, the abnormal increasing pro-inflammatory cytokines such as TNF $\alpha$ (Chen et al., 2019), interleukin-1beta (IL-1 $\beta$ ) (Hauptmann et al., 2020), interleukin-6 (IL-6) (Yang et al., 2017, Yang J. et al., 2020), interferon- $\gamma$  (INF- $\gamma$ ) (Bonney et al., 2019), and inducible nitric oxide synthase (iNOS) (Smith et al., 2018) can compromise BBB permeability and induce or deteriorate neurological disorders which make the regulation of inflammation more difficult (Ren et al., 2020). In addition, although BBB structure becomes compromised with aging, leaving patients vulnerable to neurological diseases such as AD (Huang et al., 2020), the main inducer to disrupt the BBB is the abnormal increasing inflammatory cytokines after injuries or diseases.

Moreover, most research focus only on the primary diseases or injuries of the brain inducing the dysfunction of BBB and thus CNS impairment, but few on the peripheral injuries leading to that. Our previous studies indicated that peripheral burns and traumatic surgical wound could induce the dysfunction of BBB and thus cognitive impairment through IL-6 and IL-1 $\beta$  (Yang et al., 2017, Yang J. et al., 2020). As a result, regulating inflammatory factors from assaulting the BBB is crucial for curing neurological diseases. In this review, we will systematically discuss how these pro-inflammatory cytokines assaulting on BBB.

# The structure and biomechanisms of blood-brain barrier

In contrast to the peripheral vasculature, the vessels in the brain possess a highly selectively permeable barrier, BBB, that protects the CNS from potential toxins, pathogens, and so on. Only small lipid-soluble molecules <400 Da and with fewer than nine hydrogen bonds can independently cross the BBB *via* lipidmediated diffusion (Padden et al., 2007; Segarra et al., 2021). BBB comprises a tightly sealed monolayer of brain endothelial capillaries (Ohtsuki and Terasaki, 2007) containing cell types such as brain microvascular endothelial cells (BMECs), pericytes (PCs), astrocytes (ACs) end feet, microglia, and neurons (Figures 1A,B; Bazzoni and Dejana, 2004). Molecules between the blood circulation and the brain parenchyma through BBB by two ways include transcellular vesicular transport (transcytosis) and paracellular pathway. BMECs are connected to one another by tight junctions (TJs) and adherens junctions (AJs) (Hawkins and Davis, 2005), which regulate the paracellular permeability of BBB (Bazzoni and Dejana, 2004). TJs include the endothelialspecific claudin family member claudin-5 (Cldn5) and occludin (Ocln), which are linked to the cytoskeleton by members of the zonula occludens family (ZO-1, ZO-2, and ZO-3) (Brown and Davis, 2002). The interactions among these proteins are the primary factors regulating the paracellular barrier of BMECs (Sandoval and Witt, 2008; Figure 1A). AJs, which are formed by homophilic interactions between cadherins, such as vascular endothelial (VE)-cadherin (Cdh5, CD144) and N-cadherin, are considered a prerequisite for the establishment of TJs (Figure 1A). In BMECs, AJs intermingle with TJs to form junctional complexes that contribute to BBB stability (Dejana and Orsenigo, 2013). Another way, transcytosis, is mainly regulated by PCs, which are embedded along with vascular mural cells in the basement membrane of brain microvessels and abnormal PC function may lead to BBB dysfunction and neuroinflammation (Daneman et al., 2010; Dohgu and Banks, 2013; Ben-Zvi et al., 2014). Furthermore, recent research indicates that an important membrane protein, major facilitator superfamily domain containing 2a (Mfsd2a), on BMECs also plays an important role on transcytosis, and its expression may be regulated by PCs (Ben-Zvi et al., 2014) (Figures 1A,B). The reduced Mfsd2a after CNS diseases or injuries can increase the levels of transcellular vesicles including Cav-1, Nrf-2, and HO-1 on BMECs which may contain toxins, pathogens, or cytokines in brain, while upregulation of Mfsd2a will protect BBB from decreasing levels of the vesicles (Eser Ocak et al., 2020; Figure 1C).

Moreover, ACs are also the principal components of the BBB. ACs, whose end feet cover nearly the entire surface of BECs, have a critical effect on the BBB (Cheslow and Alvarez, 2016). Various AC proteins such as aquaporin-4 (Aqp4) and the Kir4.1 potassium channel localize to the end-foot membrane to regulate water homeostasis (Engelhardt and Liebner, 2014; Figure 1A). ACs function in BBB maintenance and induce the barrier properties of BMECs through various pathways such as Wnt (Guérit et al., 2021), Shh (Xing et al., 2020), and VE-PTPdependent restoration of Tie2 signaling (Gurnik et al., 2016). The interaction between ACs and BMECs not only improves AC differentiation via the secretion of leukemia inhibitory factor 1 (LIF1) by BMECs but also maintains the development of the BBB through VE-PTP-dependent restoration of Tie2 signaling (Liebner et al., 2018). However, the relationship between PCs and ACs remains undetermined (Figure 1A).



Structure and biomechanisms of blood-brain barrier during physical or inflammation. (A) The structure of BBB from longitudinal section during physical condition. PCs control the transcytosis and paracellular pathway of BBB by regulating the expression of Mfsd2a and TJs. The structure of potassium channel of Kir4.1 potassium channel and Aqp-4 on ACs regulates the homeostasis of CNS. However, the relationship between ACs and PCs is still unknown. (B) The structure of BBB from cross-section during physical condition. (C) The structure of BBB from cross-section during inflammation. During inflammation, the expression of Mfsd2a is decreasing, which is negatively correlated with transcytosis of BBB. The level of TJs is decreasing, which means the paracellular pathway of BBB is increasing. Moreover, PCs are abnormal contracted and impairment. In addition, ACs are activated to secrete pro-inflammatory cytokines and the level of Aqp-4 is increasing, which is correlated with the deterioration of CNS diseases or injuries.

# Effects of inflammation on blood-brain barrier

During neurological diseases or injuries, immune cells inside the brain secrete pro-inflammatory cytokines, and the structure of BBB tends to be loosened, including the disruption of PCs, which regulate the transcytosis, and TJs, which regulate the paracellular pathway (Figure 1C). Immune cells from outside the brain, such as T cells, specifically target adhesion molecules on BBB, particularly intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1) (Carrano et al., 2012). Subsequently, immune cells, especially CD4 + T cells, pass through BMECs and into the CNS via the transcytosis, mediated by caveolae cluster adhesion molecules, and through the matrix membrane via matrix metalloproteins (MMPs) expression by neutrophils in the early stage. The cytokines secreted by immune cells disrupt TJs to open the paracellular pathway in the late stage, which allows more immune cells to cross the BBB, especially Th1 and Th17 cells (Liebner et al., 2018). Immune cell reactivity and cytokine/chemokine secretion, therefore, play critical roles in the functional and anatomical structure of BBB and brain during neuroinflammation. However, our previous work suggested

that diseases or traumatic injuries outside the brain may also induce BBB disruption and eventually lead to neurological degeneration through immune cells and pro-inflammatory cytokines (Yang et al., 2017, Yang J. et al., 2020). Both excessive activities can lead to BBB permeability and neurodegeneration, thereby worsening the progression of neurological diseases, such as AD. Given the ongoing debate regarding how immune cells gain entry into the CNS and induce BBB impairment and eventually neurological degeneration, we will systematically discuss the major cytokines (e.g.,  $TNF\alpha$ , IL-1 $\beta$ , and IL-6) that are secreted by immune cells and their impact on BBB and CNS (Figure 2).

### The effects of TNF $\alpha$ on blood-brain barrier and neurological degeneration

TNFα, the main homotrimeric transmembrane protein belonging to the TNF/TNFR ligand/receptor superfamily, plays an important role in neuroinflammation and BBB permeability (Table 1 and Figure 3). TNFa primarily acts by binding receptors on the cell surface as membrane-bound precursor (tmTNF $\alpha$ ) and can also be secreted as a 51-kDa soluble



trimer soluble TNFa (sTNFa) via proteolytic cleavage by the TNFα-converting enzyme (TACE) (Tang et al., 1996). During neuroinflammation, TNFa is secreted by neurons (Janelsins et al., 2008), astrocytes (Lieberman et al., 1989), and microglia (Janelsins et al., 2005) and interacts with its cognate receptors, TNF receptor type-1 (TNFR-1, also known as CD120a, p55/60) and TNF receptor type-2 (TNFR-2, also known as CD120b, p75/80). TNFR-1, which is expressed by almost all cells in CNS and mainly integrated with sTNFa, contains an intracellular death domain (DD) that plays an important role in TNFamediated apoptosis and cytotoxicity (Locksley et al., 2001; Sedger and McDermott, 2014). At the onset of neurological diseases or injuries, transient TNFa-induced JNK pathway plays a cytoprotective role through TAK1-dependent phosphorylation (Sato et al., 2005), while continuous TNFa-induced JNK signal leads to caspase-dependent apoptosis through JNK phosphorylation by apoptosis signal-regulating kinase 1 (ASK1) (Tobiume et al., 2001). In contrast, TNFR-2 plays an easy biological role compared to TNFR-1 and is mainly expressed by BMECs. Recent research has found that TNFa integrates with TNFR-2 can have effects on both pro-survival pathway and pro-inflammation through the activation of the cellular inhibitor of apoptosis proteins 1 and 2 and nuclear factor  $\kappa B$ 

(NF-κB) (Rothe et al., 1995). Moreover, TNFR-2 can enhance the association with TNFR-1 and sTNFa to perform proinflammatory role in CNS (Tartaglia et al., 1993). Recent studies have found that  $TNF\alpha$  inhibition TNFR-1-IgG administration was able to control the effects of experimental autoimmune encephalomyelitis (EAE) by reducing activated immune cells, including inflammatory leukocytes and microglia (Korner et al., 1997a). Korner et al. reported that mice lacking TNFa also developed EAE to the same extent as wild-type (WT) mice, because  $TNF^{-/-}$  mice were unable to form the typical mature perivascular cuffs observed in WT mice (Korner et al., 1997b). Neutralizing or knocking out TNFa has varying effects because the two types of TNFa receptor have different functions. It is the key that sustained sTNFα integrated with TNFR-1 plays a detrimental role in neurological diseases. To demonstrate this point, Suvannavejh et al. (2000) used TNFR-1/TNFR- $2^{-/-}$  and TNFR- $2^{-/-}$  mice and found that TNFR- $1^{-/-}$  mice were able to control EAE progression by reducing Th1 cells and immune cell infiltration while  $\text{TNFR-}2^{-/-}$  mice have exacerbated the progression of EAE and the infiltration of immune cells (Suvannavejh et al., 2000). Sophie et al. found that a nanobody-based selective inhibitor of human TNFR1, TROS, was able to effectively delay disease onset, ameliorate symptoms,

and prevent further disease development in a murine model of MS, in which the researchers generated mice expressing human TNFR1 in a mouse TNFR1-knockout background.

Under normal inflammatory condition, TNFa can promote BMEC remodeling through PC proliferation and differentiation from  $\alpha 1$  to  $\alpha 2$  integrin (Tigges et al., 2013). When neuroinflammation cannot be controlled, sustained TNFa can alter BBB permeability and make the neurological diseases or injuries worsen. During neurological diseases or injuries, microglia-induced TNFa interacts with TNFR-1 disrupts TJs and leads to BMECs necroptosis, thereby allowing entry of toxins and pathogens into CNS (Chen et al., 2019). Aslam et al. (2012) found that TNFa can disrupt the TJ structure by reducing Cldn5 via activation of the NF-KB signaling pathway. Ni et al. (2017) used an immortalized human cerebral endothelial cell line (hCMEC/D3) to demonstrate that TNFa can disrupt TJs on BMECs, which can be largely inhibited by SB202190, an inhibitor of p38MAPK, and partly by U0126, a MEK1/2-ERK1/2 inhibitor, thus implicating both of these signaling pathways in BBB disruption. From the outside of CNS, during systemic inflammation, peripheral TNFa can directly cause BBB dysfunction through decreasing transcellular electrical resistance, cellular polarity, and activate BMECs and ACs to secrete pro-inflammatory chemokines, such as MCP-1 and IP-10, which are responsible for the recruitment of leukocyte. The situation can further disrupt TJs and induce an inflammatory milieu, as well as the adhesion molecule ICAM-1 and VCAM-1, which recruit leukocytes by increasing the transcellular permeability of the BBB (O'Carroll et al., 2015; Mantle and Lee, 2018). In addition, Ding et al. suggested that TNFα can upregulate the expression of MMP-9, a proteinase to disrupt TJs and basal membrane of BBB, through activation of Ca/CAMK II/ERK/NF-κB signaling pathway (Ding et al., 2019). Other potential mechanism about peripheral TNFa-induced BBB dysfunction includes cyclooxygenase (COX) pathway and iNOS released. By activation of COX pathway, TNFa can upregulate COX-2 to increase the levels of MMP-2, MMP-3, and MMP-9 and change the cytoskeletal structure of BMECs to disrupt the TJs. Another pathway is TNFa-induced iNOS release which can increase the BBB permeability by impairing BMECs, but the mechanism remains unknown (Liu et al., 2013). At last, sustained TNFa in blood can lead to AC dysfunction. In other situations, hypoxia, recent studies indicate that TNFa will inhibit the expression of erythropoietin (EPO), which is secreted by ACs and has an important neuroprotective role on CNS and BBB, thereby exacerbating the outcome of neurological injury in hypoxia (Nagaya et al., 2014). Kralingen et al. found that  $\text{TNF}\alpha$  generates an inflammatory milieu that will induce a reduction of ACs (van Kralingen et al., 2013). In contrast, although TNFa can induce BBB disruption and TNFa inhibition can reduce CNS inflammation, representing a potentially promising target for treating neuroinflammation, TNFα may play a neuroprotective role in diseases, such as AD,

having been shown to reduce cellular prion protein (PrPC) and A $\beta$  protein levels in BMECs of mouse. The effects of TNF $\alpha$  on the BBB and CNS are well known (Yasutaka et al., 2015). After all, the underlying intercellular signaling pathways, as well as strategies for using these effects to save patients from diseases, such as neurodegeneration, are promising targets for future study.

# The effects of IL-1 $\beta$ on blood-brain barrier and neurological degeneration

IL-1β, a member of the IL-1 family which includes seven agonist signaling ligands, three receptor antagonists, and IL-37, is largely responsible for the acute response and angiogenesis. During the initial inflammation, the inflammasome is synthesized in the cytoplasm which would assemble proteins made up mainly through the nucleotide binding and oligomerization domain (NOD)-like receptors (NLRs). This assembled inflammasome is specific for a set of cell damage-associated molecular patterns (DAMPs) or pathogen-associated microbial patterns (PAMP) according to the type of damage of body recognized by different NLR subtypes. Moreover, the inactive procaspase-1 molecules are recruited to the inflammasome complex to be activated (Nasti and Timares, 2012). IL-1β, primarily, occurs as a 31kDa inactive precursor and would then be cleaved by the caspase-1 to yield the 17-kDa bioactive IL-1β (Netea et al., 2010). The low level of IL-1 $\beta$  in CNS performs an important biological and physiological role which controls sleep, appetite, hypothalamus-pituitary axis, neuroendocrine function (Liu and Quan, 2018), neurogenesis, and synaptic plasticity (Vezzani and Viviani, 2015). Furthermore, Goshen et al. found that blocking IL-1ß signaling induced impaired hippocampal-independent memory and intact performance in adult mice (Cibelli et al., 2010). However, during neuroinflammation, sustained IL-1β integrated with toll-like receptor-4 (TLR-4) or IL-1 receptor (IL-1R) can promote the progression of diseases or injuries by activating ACs to secrete pro-inflammatory cytokines (IL-6, TNF), colony-stimulating factors, chemokines (CCL2, CXCL2, etc.), phospholipase A2, COX-2, prostaglandins and iNO via NF-KB and JNK pathways, and activator protein 1 (AP-1) (Blanco et al., 2005; Kitazawa et al., 2011; Musella et al., 2020). The activation of ACs can initiate microglia response to neuroinflammation by ACs-derived ATP as well (Bianco et al., 2005).

The ability of IL-1 $\beta$  to increase BBB permeability after CNS injuries and neurological degeneration is well established (**Table 1** and **Figure 4**). IL-1 $\beta$  disrupts BBB in two ways: First, IL-1 $\beta$  can activate ACs to disrupt BBB and exacerbate the progression of the neurological diseases or injuries. Recent work has shown that IL-1 $\beta$  induces the expression of hypoxia-inducible factor-1 (HIF-1) and its gene target, vascular

Inflammatory cytokines	Туре	Effects on BBB
ΤΝFα	TNFR-1, Pro-inflammatory effects	Disrupting TJs, especially Claudin-5 Chen et al., 2019
		Altered BBB morphology Ni et al., 2017
		Increased secretion of other pro-inflammatory cytokines Aslam et al., 2012
		Recruitment of leukocytes O'Carroll et al., 2015; Mantle and Lee, 2018
		Disrupting astrocytes van Kralingen et al., 2013
	TNFR-2, anti-inflammatory effects	Reduced recruitment of other pro-inflammatory cytokines Suvannavejh et al., 2000
		Promoting BBB recovery Madsen et al., 2020
IL-1β	Pro-inflammatory cytokine	Disrupting astrocyte
		Decreasing TEEB on the BMECs Argaw et al., 2006
		Increased secretion of other pro-inflammatory cytokines Labus et al., 2014
IL-6	sIL-6R, pro-inflammatory effects	Disrupting TJs Rosejohn and Heinrich, 1994
		Decreased $\beta$ -catenin Yang et al., 2017
	mIL-6R, anti-inflammatory effects	Reduced recruitment of other pro-inflammatory cytokines
		Promoting BBB recovery Jones and Jenkins, 2018
HMGB1	Pro-inflammatory cytokine	Activating neuroinflammation
		BBB disruption O'Connor et al., 2003; Liesz et al., 2015
IL-10	Anti-inflammatory cytokine	Reduced BBB impairment
		Reduced down-regulation of Claudin-5 Couper et al., 2008; Lin et al., 2018

### TABLE 1 Interaction between inflammatory cytokines and the BBB.

endothelial growth factor-A (VEGF-A), in human astrocytes, which in turn induces the breakdown of BBB and exacerbates the CNS degeneration, such as in MS (Argaw et al., 2006). Moynagh et al. (1994) suggested that cerebral IL-1 $\beta$  can activate NF-kB and increase the levels of VCAM-1 and ICAM-1 in ACs, which can recruit the leukocytes into CNS. Second, IL-1ß promotes the secretion of other pro-inflammatory cytokines (IL-6 and TNF $\alpha$ ) to disrupt the paracellular BBB pathway. Labus et al. used a transfected human brain microvascular endothelial cell (THBMEC)-based in vitro BBB model and found that IL-1β decreased TEER and induced the secretion of other proinflammatory cytokines, including IL-6 and TNFa, to increase the paracellular permeability of BBB. The increasing paracellular permeability will be more vulnerable to leukocytes and thus exacerbate neuroinflammation (Labus et al., 2014). Some studies have shown that increasing IL-1 $\beta$  or TNF $\alpha$  has similar effects on the secretion of inflammatory cytokines and the induction of ACs death (Netea et al., 2010). Ni et al. found that IL-1 $\beta$  may be a downstream target of  $TNF\alpha$ , which can change the structure of Ocln, thereby changing BBB permeability, a smaller effect than that of TNFa through p38 MAPK and ERK1/2 in hCMEC/D3 cells (Ni et al., 2017). However, these effects are quite different from the effects of TNFa, as demonstrated by O'Carroll et al. who observed clear differences between the effects of TNFa and IL-16 on BBB permeability. IL-16 preferentially induces the cytokines sICAM-1, IL-6, sTNFRI, sTNFRII, GCSF, and GMCSF and the chemokine IP10, whereas TNFa is more likely to induce RANTES and IL-8 secretion (O'Carroll et al., 2015). As a result, the sustained IL-1 $\beta$  will impair the BBB and then induce neurological degeneration. The fact makes the inhibition

of IL-1 $\beta$  a therapeutic target for attenuating neuroinflammatory and neurodegeneration (Cibelli et al., 2010).

Despite the evidence of IL-1 $\beta$  on disruption of BBB and then inducing the progression of neurological diseases discussed above, there are supplements of the role of IL-1 $\beta$  induced by systematic inflammation. Mario et al. (2010) have found that mice undergoing surgery of the tibia under general anesthesia have an extreme increasement of IL-1 $\beta$  in hippocampal and then inducing memory and cognitive impairment (Mario et al., 2010). Our research indicated that burn disrupted BBB both increasing the paracellular pathway and transcytosis through increasing the peripheral level of IL-1 $\beta$  and IL-6 (Yang J. et al., 2020). However, the potential mechanism of how it infiltrates into CNS is still unclear.

# The effects of IL-6 on blood-brain barrier and neurological degeneration

The common receptor to bind IL-6 is receptor  $\beta$ -subunit, membrane glycoprotein 130 (gp130; also known as IL-6R $\beta$ ), which works with either a non-signaling receptor  $\alpha$ -subunit or a signaling receptor  $\beta$ -subunit like gp130 to exert pleiotropy function (Heinrich et al., 2003). In addition to classical signaling pathway binding with gp130 on membrane, there is a soluble receptor mainly associated with IL-6(sIL-6R) which can bind circulating half-life IL-6 and expand its bioactivity in those cells without gp130 on their membrane. This signaling pathway is called trans-signaling pathway and is involved in numerous chronic inflammation and diseases



Effects of TNF $\alpha$  on BBB and neurological degeneration. (A) The mechanism of TNF $\alpha$  on CNS. The tmTNF $\alpha$  integrated with TNFR2 has two functions including promoting the expression of the cellular inhibitor of apoptosis proteins 1,2 to function as cryoprotection and integrating IkB to induce apoptosis and pro-inflammation. The sTNF $\alpha$  integrated with TNFR1 and TRADD can promote the expression of P38 to disrupt TJs and integrate with IKK $\alpha$  and  $\beta$  and further IkB to exert apoptosis and pro-inflammation. Moreover, the complex of sTNF $\alpha$  integrated with TNFR1 and TRADD can integrate TAK1 to exert cryoprotection through JNK in early stage, while the complex can integrate ASK1 to exert apoptosis through JNK in late stage. (B) The mechanism of TNF $\alpha$  on BBB from inside and outside of CNS. The sTNF $\alpha$  from outside CNS integrated with TNFR1 promotes the expression of ICAM-1/VCAM-1 on the membrane of BMECs and the secretion of pro-inflammatory cytokines and chemokines, which can disrupt the structure of TJs. Moreover, the complex of sTNF $\alpha$  induces COX-2 to promote the expression of MMP-2,3,9 which can further disrupt the TJs and basement membrane. TNF $\alpha$  secreted mainly by ACs and microglia from inside CNS promotes necessary.

(Rosejohn and Heinrich, 1994) (Table 1; Figure 5). During neuroinflammation, normal level and biological function of IL-6 interacted with sIL-6R via trans-signaling pathway can induce repopulating microglia which has an important role on neurogenesis specifically by augmenting the survival of newborn neurons that directly support cognitive function (Willis et al., 2020). The level of IL-6 increases by the activation of ACs induced by TNF and IL-1β via NFκB signaling pathway, while IL-6 is negatively regulated by Wnt signaling pathway (Edara et al., 2020). However, other research indicates that sustained IL-6 can lead to neuroinflammation and pain post-injury via signal transducer and activator of transcription 3 (STAT3) (Hu et al., 2020). Through STAT3, IL-6 can induce the demethylation of NeuroD1 (neurogenic differentiation 1) in neural stem cells (NSCs) to promote the neurogenesis in AD as well (Kong et al., 2019). Moreover, Escrig et al. found in mouse AD model that IL-6 trans-signaling induced increasing levels of amyloid

plaques and neurofibrillary tangles and cognitive dysfunction (Escrig et al., 2019).

Recent research indicates that the peripheral levels of IL-6 are higher compared with those healthy people in those patients with neurological diseases or injuries (Robelin and Gonzalez De Aguilar, 2014; Yang et al., 2017, Yang J. et al., 2020). The disruption of BBB is mainly by the integration of complexes of sIL-6R and gp130 on the membrane of BMECs by trans-signaling pathway. The pathway can directly disrupt the TJs of BBB in one respect and indirectly increase the ICAM-1 and VCAM-1 to recruit neutrophils infiltrating into the inflammatory site in another. Rosejohn and Heinrich (1994) have found that blockage of the trans-signaling pathway can effectively reduce the leakage of BBB and then relieve the progression of neurological degeneration. As mentioned of TNFa and IL-1β, IL-6 can also induce the released pro-inflammatory cytokines/chemokines to deteriorate the progression in several neurological diseases or



injuries (Mantle and Lee, 2018). Zhang et al. (2015) found that peripheral IL-6 levels were increased during hypoxic-ischemic brain injury, thus disrupting BBB permeability, and that using neutralizing anti-IL-6 antibodies (anti-IL-6AB) decreased the disruption of TJs and controlled BBB permeability 24 h after ischemic injury. Emery et al. suggested that using an inhibitor of IL-6R can effectively control the progression of autoimmune diseases, such as amyotrophic lateral sclerosis (ALS), in a phasethree clinical trial (Emery et al., 2008).

Above all, most researches focus on the evidence that the inflammation caused by CNS diseases or injuries can induce neurodegeneration. However, peripheral IL-6 can also induce the BBB dysfunction. Our work using a murine surgery model revealed that surgical wounds can induce age-associated BBB dysfunction, and elevated serum IL-6 levels led to decreased TJs but had no effects on AJs (Yang et al., 2017). In addition, we used the mouse burn model which suggested that burn can increase the peripheral level of IL-6 and IL-1 $\beta$  and the level of MMP-9 to disrupt the integrity of BBB via paracellular pathway by decreasing TJs, and transcytosis by decreasing Mfsd2a (Yang J. et al., 2020). Other research indicates that the increasing peripheral level of IL-6 can disrupt the TJs to disrupt the BBB and cause neuroinflammation on one side (Furutama et al., 2020) and induce the increasing level of COX-2 to disrupt the BMECs (Eskilsson et al., 2014). Our previous study also found that increased peripheral IL-6 levels can induce decreased  $\beta$ catenin levels, thereby activating the Wnt pathway, suggesting that the ability of IL-6 overexpression to disrupt the BBB may be related to Wnt signaling pathway (Yang et al., 2017). The study will provide a new insight how we should prevent the CNS injuries from the other site injuries or diseases.

# The effects of other potential cytokines on the blood-brain barrier and neurological degeneration

Inflammation is controlled by a complex network of various pro-inflammatory cytokines and receptors (**Table 1**; Patin et al., 2016). However, the exact cytokines and receptors that participate in the neuroinflammation and how they produce their effects *via* signaling pathways remain unclear. In this section, we will try to elucidate the current possible factors and signaling pathways involved in BBB disruption. High-mobility group box 1 protein (HMGB1), which belongs to the damage-associated molecular pattern (DAMP) family (Chaudhry et al., 2018), is released from stressed and dying brain cells and is a potent neuroinflammatory mediator that is mainly regulated by post-translational redox modification. HMGB1 is a nuclear DNA-binding protein that contains two DNA-binding domains



(boxes A and B) and a highly repetitive, negatively charged C-terminal tail (Thundyil and Lim, 2015). HMGB1 can disrupt the BBB and induce neurodegeneration by promoting the secretion of pro-inflammatory cytokines, such as TNF $\alpha$ , IL-1 $\beta$ , and IL-6, through the NF- $\kappa$ B signaling pathway and by activating the RAGE axis (O'Connor et al., 2003; Liesz et al., 2015). Moreover, intentionally blocking HMGB1 may improve the outcomes of neurological diseases and injuries such as stroke and TBI and protect the BBB from disruption (Zhang et al., 2011; Okuma et al., 2014).

In contrast, some cytokines play an important role on antiinflammation and protect the permeability of BBB, including interleukin-10 (IL-10), which is an anti-inflammatory cytokine that is expressed by various leukocytes. The two main IL-10 receptors are IL-10Ra, which is specific to IL-10, and IL-10RB (Sanjabi et al., 2009). IL-10 mainly exerts its function by activating the STAT3 signaling pathway. The IL-10 gene promoter contains a STAT3 binding site, resulting in a feedback loop for this pathway. Interestingly, STAT3 plays an anti-inflammatory role when it is activated by IL-10 but plays a pro-inflammatory role with IL-6 (Mosser and Zhang, 2008). Regarding its effects on the BBB and CNS, Lin et al. suggested that IL-10 effectively reduced BMEC apoptosis induced by IL-6-activated STAT3 signaling during neuroinflammation and ameliorate the downregulation of Cldn5 expression in the BBB in a rat model, while in vitro, IL-10 protected BBB integrity against TNF $\alpha$  (Lin et al., 2018). Overall, IL-10 plays an important role in immune system homeostasis. Reduced IL-10 expression increases susceptibility to autoimmune inflammation, whereas IL-10 overexpression will impede the clearance of pathogens and induce a chronic inflammatory state (Couper et al., 2008). As a result, how to use IL-10 for clinical treatment requires further study.

In addition to major cytokines, such as TNF $\alpha$ , IL-1 $\beta$ , IL-6, and their receptors, other cytokines also play an important role in neuroinflammation and neurodegeneration and may provide some promising new targets for clinical application to treat neurological disease and injury. However, determining how these factors interact with each other and how the mechanisms of these effects change with age is of critical importance.

# The effects of inflammatory cytokines on blood-brain barrier and central nervous system from primary neurological diseases or systematic inflammation

In primary neurological diseases including AD, MS, stroke, and TBI, inflammatory cytokines play an important role on

the promotion of the diseases healing or deteriorating the progression of the diseases in early or late stage, respectively. Rational regulation of inflammation will be the protection for the primary CNS diseases (Jayaraj et al., 2019). However, in neurological diseases, damage to the CNS may lead to a decrease in the control of inflammation, which will lead to inflammatory dysregulation (Lambertsen et al., 2019; Ren et al., 2020). Under this condition, the aberrant expression of inflammatory cytokines will induce the deterioration of neurological diseases and thus neurodegeneration, which may provide an effective target for future clinical treatment as well. Neurological diseases like AD, the secondary pro-inflammatory insults, trigger delirium and can accelerate cognitive decline. Lopez-Rodriguez et al. (2021) found that  $A\beta$  was surrounded with NLRP3-integrated inflammasome which recruited microglia to be primed and facilitated to secrete IL-1β using APP/PS1 mice. In addition, activated ACs were primed to pro-inflammatory chemokine responses induced by IL-1β. A CNS trauma like TBI (Kuwar et al., 2019) also found that the complexes of NLRP3integrated inflamma some induced the expression of IL-1 $\beta$  which will subsequently induce the expression of TNFa and IL-6. On the other side, those pro-inflammatory cytokines secreted by the microglia and ACs can further induce the disruption of BBB through the pathway mentioned before. Furthermore, the expression of ICAM-1 and VCAM-1 and the disruption of TJs will subsequently recruit the peripheral immune cells and cytokines into CNS and thus accelerate the progression of primary diseases (Moynagh et al., 1994; Argaw et al., 2006; Labus et al., 2014; O'Carroll et al., 2015; Zhang et al., 2015; Mantle and Lee, 2018).

Nowadays, the CNS injury caused by peripheral trauma or surgery has gradually become a research hotspot which could also trigger delirium and accelerate cognitive decline on the basis of pre-existing diseases or with higher underlying risks of neurological diseases (Yang T. et al., 2020). Our research focused on the disruption of BBB and cognitive impairment induced by peripheral traumatic injuries. We used traumatic surgical model and burns model in mice and found that peripheral inflammatory cytokines, including IL-6 and IL- $1\beta$ , were main factor leading to the disruption of BBB and thus cognitive impairment. In surgical model, we found that BBB was disrupted by the increasing level of IL-6 and thus induced cognitive impairment, especially in old mice with higher serum level of IL-6 than young mice with that (Yang et al., 2017). To fully understand the mechanism about the disruption of BBB induced by systematic inflammation, we further used burns model which had more serious inflammatory response than our previous study used surgical model. We found that the increasing permeability of BBB not only included paracellular pathway but also transcytosis induced by the increasing serum level of IL-1β and IL-6 (Yang J. et al., 2020). Unlike primary CNS disease, there are few inflammatorymediated factors to specifically recruit the immune cells or

inflammatory cytokines into CNS. From our studies, the potential mechanism about the disruption of BBB and thus CNS impairment induced by systematic inflammation may be caused by high amount and functional degeneration of immune cells and pro-inflammatory cytokines to identify the receptors in BMECs, the immune situation of which is more likely occur in aging or chronic diseases population (Sendama, 2020). However, new research associated with AD mentioned a potential mechanism that peripheral surgery inducing cognitive impairment may be through the disruption of nasal epithelium and olfactory receptor neurons by the increasing level of IL-6 after surgery (Zhang et al., 2022), but the sequence of disruption of nasal epithelium, olfactory receptor neurons, and cognitive impairment was not strictly discovered, which indicated that peripheral trauma inducing cognitive impairment through the disruption of BBB caused by systematic inflammation may still be the most probable theory in the field. Cognitive impairment and then neurological degeneration induced by systematic inflammation are our novel insight, which will need to pay attention to the protection of BBB in clinical treatment of peripheral diseases in future. However, the potential mechanism needs to be further studied.

### Conclusion

The primary factors that disrupt the BBB to induce neurological degeneration are three inflammatory cytokines: TNF $\alpha$ , IL-1 $\beta$ , and IL-6. The basal functions of these cytokines are elucidated above. As an inflammatory network, these cytokines integrated with their special receptors can similarly disrupt TJs and TEER of the BBB without any impairment of AJs (Aslam et al., 2012; Labus et al., 2014), promote ACs apoptosis (Argaw et al., 2006; Nagaya et al., 2014), and recruit leukocytes into BBB (O'Carroll et al., 2015), thereby directly or indirectly inducing neurological degeneration. Other cytokines, including HMGB1 and IL-10, also play an important role in the BBB, as mentioned above. Cytokines such as TNFa and IL-6 bind their respective receptors and exert different functions, as discussed above (Locksley et al., 2001; McCoy and Tansey, 2008; Murakami and Hirano, 2012; Sedger and McDermott, 2014; Wolf et al., 2014). Currently, the pathways that may be involved in these processes include NF-KB (Bianco et al., 2005; Aslam et al., 2012; Edara et al., 2020), p38MAPK, and MEK1/2-ERK1/2 signaling pathway (Ni et al., 2017). However, the potential mechanisms by which cytokines attack the BBB are still unclear. Recent work has shown that blocking proinflammatory cytokines, including TNF $\alpha$ , IL-1 $\beta$ , and IL-6, can reduce the permeability of BBB and improve the outcomes of neurological diseases and injuries to a great extent. However, in some conditions, studies have found that cytokines such as TNFa, IL-1β, and IL-6 can also improve the outcome of diseases such as ADs and that neutralizing or knocking out

10.3389/fnmol.2022.1013933

these cytokines can eventually attenuate the progression of neurological diseases or injuries (Goshen et al., 2007; Yasutaka et al., 2015; Han et al., 2016; Patin et al., 2016). Overall, neurological diseases or injuries present a substantial challenge to the medical field because they are difficult to diagnose and cure and are a social burden for families. Almost all neurological diseases and injuries have an important relationship with the BBB and inflammatory cytokines, which represent a new target for treating these diseases and injuries. As a result, the effects of these cytokines on the BBB and neurodegeneration must be investigated to develop new clinical treatments.

The most recent research has focused on local CNS diseases or injuries such as TBI, AD, and MS attacking BBB and inducing neurodegeneration (Owen et al., 2010; Vázquez-Rosa et al., 2020; Puthenparampil et al., 2021). However, our previous research has confirmed that systematic inflammation will also induce neurological degeneration. For example, after peripheral surgery, the concentration of IL-6 increases in peripheral blood and the structure of the BBB breakdown, thereby inducing cognitive impairment in mice. Older individuals (18-month-old mice) have a higher risk of such impairment. Using IL-6-neutralizing antibodies or gene IL-6 knockout protected the BBB from disruption (Yang et al., 2017). Furthermore, we use the mouse burn model to indicate that peripheral burn can also increase the level of IL-6 and IL-1 $\beta$  in serum which disrupt the integrity of BBB through paracellular pathway and transcytosis (Yang J. et al., 2020). Similar results have been observed in humans. Reichenberg et al. (2001) found that humans with Salmonella abortus equi endotoxin (0.8 ng/kg) presented with abnormal behavior compared with healthy individuals. The results revealed that even a small amount of inflammation can impair the BBB and induce cognitive impairments. As a result, with advancing age, people are more vulnerable to various diseases, and their body experiences chronic inflammation. This effect may continue to produce pro-inflammatory cytokines that disrupt the BBB and induce neurological degeneration. However, demonstrating these effects and illuminating the detailed mechanisms require substantial future research.

Current treatments for neurological degeneration are minimally effective. To date, most studies have found that inhibiting the overexpression of inflammatory cytokines is effective at improving the outcomes of neurological diseases and injuries and delaying the onset of neurolegeneration. However, the timing and mode of intervention are not ideal. As in our research, using anti-IL-6 antibody 18 h before surgery effectively reduced BBB disruption and cognitive impairment, whereas post-surgery treatment has no effect. In clinical settings, we cannot readily use protective interventions before injuries or diseases. Additional research has also found that mesenchymal stem cells (MSCs) effectively protect BBB integrity and that MSC intracranial transplantation may be a promising method for future treatment (Si et al., 2011). Tang et al. found that MSC intracranial transplantation protected BBB integrity, inhibited ACs apoptosis, and decreased Aqp4 expression, which reduced brain edema and impairment (Tang et al., 2014). Menge et al. (2012) suggested that the injection of MSCs can increase the level of tissue inhibitor of matrix metalloproteinase-3 (TIMP3) which can inhibit VEGF-A to protect the BBB from disruption. As mentioned before, injecting MSCs requires cerebral transplantation, which involves substantial risk, and the exact mechanism of how MSCs influence the BBB is poorly understood. Our previous study suggested that MSCs injecting through the tail of mice can effectively protect the integrity of BBB by decreasing peripheral inflammatory cytokines including IL-1 $\beta$  and IL-6, the level of MMP-9, and increasing the level of Mfsd2a (Yang J. et al., 2020). However, the mechanism by which MSCs prevent the BBB from proinflammatory cytokines attacking is still under discussed. As a result, additional studies are needed before this approach can be used in clinically.

Individuals with acute or chronic non-cerebral diseases account for a large proportion of patients in the world, and few treatments are available to protect the BBB. Especially, aging makes BBB more vulnerable to the disruption by inflammatory cytokines. The impairment of BBB is easy to recruit more immune cells and cytokines into brain parenchyma and then induce neurological degeneration (Boland et al., 2018). With an aging society coming, the medical care of aging people with neurological diseases is commonly considered a significant economic burden on society and on families (Chen, 2010; Gonneaud and Chételat, 2018; Fang et al., 2020; Pandya and Patani, 2021). As a result, whether the prevention of the disruption of BBB from non-cerebral injuries and diseases will delay the outset and improve the progression of neurological degeneration and this question requires urgent and massive future research to approve. As such, future studies should focus on the mechanisms and treatment of peripheral inflammationinduced BBB disruption. Regardless of cerebral or non-cerebral inflammation, protecting the BBB is a key to preventing neurological degeneration.

# Author contributions

SY, JY, MR, YL, and XF conceived and designed the review. JY, YL, KM, and YY prepared the figures. JY, HL, SY, and XF wrote the manuscript. All authors contributed to the article and approved the submitted version.

# Funding

This study was supported in part by the National Nature Science Foundation of China (81801909, 81830064, and

81721092) and the Military Medical Research and Development Projects (18-JCJQ-QT-020 and AWS17J005, 2019-126).

# **Conflict of interest**

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# References

Argaw, A. T., Zhang, Y., Snyder, B. J., Zhao, M. L., Kopp, N., Lee, S. C., et al. (2006). IL-1beta regulates blood-brain barrier permeability via reactivation of the hypoxia-angiogenesis program. *J. Immunol.* 177, 5574–5584. doi: 10.4049/jimmunol.177.8.5574

Aslam, M., Ahmad, N., Srivastava, R., and Hemmer, B. (2012). TNF-alpha induced NFkappaB signaling and p65 (RelA) overexpression repress Cldn5 promoter in mouse brain endothelial cells. *Cytokine* 57, 269–275. doi: 10.1016/ j.cyto.2011.10.016

Bazzoni, G., and Dejana, E. (2004). Endothelial cell-to-cell junctions: Molecular organization and role in vascular homeostasis. *Physiol. Rev.* 84, 869–901. doi: 10.1152/physrev.00035.2003

Ben-Zvi, A., Lacoste, B., Kur, E., Andreone, B. J., Mayshar, Y., Yan, H., et al. (2014). Mfsd2a is critical for the formation and function of the blood-brain barrier. *Nature* 509, 507–511. doi: 10.1038/nature13324

Bianco, F., Pravettoni, E., Colombo, A., Schenk, U., Möller, T., Matteoli, M., et al. (2005). Astrocyte-derived ATP induces vesicle shedding and IL-1 beta release from microglia. *J. Immunol.* 174, 7268–7277. doi: 10.4049/jimmunol.174.11.7268

Blanco, A., Vallés, S., Pascual, M., and Guerri, C. (2005). Involvement of TLR4/type I IL-1 receptor signaling in the induction of inflammatory mediators and cell death induced by ethanol in cultured astrocytes. *J. Immunol.* 175, 6893–6899. doi: 10.4049/jimmunol.175.10.6893

Boland, B., Yu, W., Corti, O., Mollereau, B., Henriques, A., Bezard, E., et al. (2018). Promoting the clearance of neurotoxic proteins in neurodegenerative disorders of ageing. *Nat. Rev. Drug Discov.* 17, 660–688. doi: 10.1038/nrd.2018.109

Bonney, S., Seitz, S., Ryan, C., Jones, K., Clarke, P., Tyler, K., et al. (2019). Gamma interferon alters junctional integrity via rho kinase, resulting in bloodbrain barrier leakage in experimental viral encephalitis. *mBio* 10:e01675-19. doi: 10.1128/mBio.01675-19

Brown, R., and Davis, T. (2002). Calcium modulation of adherens and tight junction function: A potential mechanism for blood-brain barrier disruption after stroke. *Stroke* 33, 1706–1711. doi: 10.1161/01.STR.0000016405.06729.83

Carrano, A., Hoozemans, J. J., van der Vies, S. M., van Horssen, J., de Vries, H. E., and Rozemuller, A. J. (2012). Neuroinflammation and blood-brain barrier changes in capillary amyloid angiopathy. *Neurodegener. Dis.* 10, 329–331. doi: 10.1159/000334916

Chaudhry, S. R., Hafez, A., Rezai Jahromi, B., Kinfe, T. M., Lamprecht, A., Niemela, M., et al. (2018). Role of damage associated molecular pattern molecules (DAMPs) in aneurysmal subarachnoid hemorrhage (aSAH). *Int. J. Mol. Sci.* 19:2035. doi: 10.3390/ijms19072035

Chen, A., Fang, Z., Chen, X., Yang, S., Zhou, Y., Mao, L., et al. (2019). Microgliaderived TNF- $\alpha$  mediates endothelial necroptosis aggravating blood brain-barrier disruption after ischemic stroke. *Cell Death Dis.* 10:487. doi: 10.1038/s41419-019-1716-9

Chen, L. (2010). Population ageing is a global phenomenon, which affects both Taiwan and China. Preface. *Ageing Res. Rev.* 9(Suppl. 1):S1. doi: 10.1016/j.arr. 2010.04.006

Cheslow, L., and Alvarez, J. I. (2016). Glial-endothelial crosstalk regulates bloodbrain barrier function. *Curr. Opin. Pharmacol.* 26, 39–46. doi: 10.1016/j.coph. 2015.09.010

Cibelli, M., Fidalgo, A. R., Terrando, N., Ma, D., Monaco, C., Feldmann, M., et al. (2010). Role of interleukin-1beta in postoperative cognitive dysfunction. *Ann. Neurol.* 68, 360–368. doi: 10.1002/ana.22082

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Couper, K. N., Blount, D. G., and Riley, E. M. (2008). IL-10: The master regulator of immunity to infection. *J. Immunol.* 180, 5771–5777. doi: 10.4049/jimmunol. 180.9.5771

Daneman, R., Zhou, L., Kebede, A. A., and Barres, B. A. (2010). Pericytes are required for blood-brain barrier integrity during embryogenesis. *Nature* 468, 562–566. doi: 10.1038/nature09513

Dejana, E., and Orsenigo, F. (2013). Endothelial adherens junctions at a glance. J. Cell Sci. 126(Pt 12), 2545–2549. doi: 10.1242/jcs.124529

Ding, X., Sun, X., Shen, X., Lu, Y., Wang, J., Sun, Z., et al. (2019). Propofol attenuates TNF- $\alpha$ -induced MMP-9 expression in human cerebral microvascular endothelial cells by inhibiting Ca/CAMK II/ERK/NF- $\kappa$ B signaling pathway. *Acta Pharmacol. Sin.* 40, 1303–1313. doi: 10.1038/s41401-019-0258-0

Dohgu, S., and Banks, W. (2013). Brain pericytes increase the lipopolysaccharide-enhanced transcytosis of HIV-1 free virus across the in vitro blood-brain barrier: Evidence for cytokine-mediated pericyte-endothelial cell crosstalk. *Fluids Barriers CNS* 10:23. doi: 10.1186/2045-8118-10-23

Edara, V., Nooka, S., Proulx, J., Stacy, S., Ghorpade, A., and Borgmann, K. (2020).  $\beta$ -catenin regulates wound healing and IL-6 expression in activated human astrocytes. *Biomedicines* 8:479. doi: 10.3390/biomedicines8110479

Emery, P., Keystone, E., Tony, H. P., Cantagrel, A., van Vollenhoven, R., Sanchez, A., et al. (2008). IL-6 receptor inhibition with tocilizumab improves treatment outcomes in patients with rheumatoid arthritis refractory to anti-tumour necrosis factor biologicals: Results from a 24-week multicentre randomised placebo-controlled trial. *Ann. Rheum. Dis.* 67, 1516–1523. doi: 10. 1136/ard.2008.092932

Engelhardt, B., and Liebner, S. (2014). Novel insights into the development and maintenance of the blood-brain barrier. *Cell Tissue Res.* 355, 687–699. doi: 10.1007/s00441-014-1811-2

Escrig, A., Canal, C., Sanchis, P., Fernández-Gayol, O., Montilla, A., Comes, G., et al. (2019). IL-6 trans-signaling in the brain influences the behavioral and physiopathological phenotype of the Tg2576 and 3xTgAD mouse models of Alzheimer's disease. *Brain Behav. Immun.* 82, 145–159. doi: 10.1016/j.bbi.2019.08.005

Eser Ocak, P., Ocak, U., Sherchan, P., Gamdzyk, M., Tang, J., and Zhang, J. H. (2020). Overexpression of Mfsd2a attenuates blood brain barrier dysfunction via Cav-1/Keap-1/Nrf-2/HO-1 pathway in a rat model of surgical brain injury. *Exp. Neurol.* 326:113203. doi: 10.1016/j.expneurol.2020.113203

Eskilsson, A., Mirrasekhian, E., Dufour, S., Schwaninger, M., Engblom, D., and Blomqvist, A. (2014). Immune-induced fever is mediated by IL-6 receptors on brain endothelial cells coupled to STAT3-dependent induction of brain endothelial prostaglandin synthesis. *J. Neurosci.* 34, 15957–15961. doi: 10.1523/JNEUROSCI. 3520-14.2014

Fang, E., Xie, C., Schenkel, J., Wu, C., Long, Q., Cui, H., et al. (2020). A research agenda for ageing in China in the 21st century (2nd edition): Focusing on basic and translational research, long-term care, policy and social networks. *Ageing Res. Rev.* 64:101174. doi: 10.1016/j.arr.2020.101174

Furutama, D., Matsuda, S., Yamawaki, Y., Hatano, S., Okanobu, A., Memida, T., et al. (2020). IL-6 induced by periodontal inflammation causes neuroinflammation and disrupts the blood-brain barrier. *Brain Sci.* 10:679. doi: 10.3390/ brainsci10100679

Gold, B., Shao, X., Sudduth, T., Jicha, G., Wilcock, D., Seago, E., et al. (2021). Water exchange rate across the blood-brain barrier is associated with CSF amyloid- $\beta$  42 in healthy older adults. *Alzheimers Dement.* 17, 2020–2029. doi: 10.1002/alz.12357

Gonneaud, J., and Chételat, G. (2018). Which is to blame for cognitive decline in ageing: Amyloid deposition, neurodegeneration or both? *Brain* 141, 2237–2241. doi: 10.1093/brain/awy174

Goshen, I., Kreisel, T., Ounallah-Saad, H., Renbaum, P., Zalzstein, Y., Ben-Hur, T., et al. (2007). A dual role for interleukin-1 in hippocampal-dependent memory processes. *Psychoneuroendocrinology* 32, 1106–1115. doi: 10.1016/j.psyneuen. 2007.09.004

Guérit, S., Fidan, E., Macas, J., Czupalla, C., Figueiredo, R., Vijikumar, A., et al. (2021). Astrocyte-derived Wnt growth factors are required for endothelial bloodbrain barrier maintenance. *Prog. Neurobiol.* 199:101937. doi: 10.1016/j.pneurobio. 2020.101937

Gurnik, S., Devraj, K., Macas, J., Yamaji, M., Starke, J., Scholz, A., et al. (2016). Angiopoietin-2-induced blood-brain barrier compromise and increased stroke size are rescued by VE-PTP-dependent restoration of Tie2 signaling. *Acta Neuropathol.* 131, 753–773. doi: 10.1007/s00401-016-1551-3

Han, Y., Ripley, B., Serada, S., Naka, T., and Fujimoto, M. (2016). Interleukin-6 deficiency does not affect motor neuron disease caused by superoxide dismutase 1 mutation. *PLoS One* 11:e0153399. doi: 10.1371/journal.pone.0153399

Hauptmann, J., Johann, L., Marini, F., Kitic, M., Colombo, E., Mufazalov, I., et al. (2020). Interleukin-1 promotes autoimmune neuroinflammation by suppressing endothelial heme oxygenase-1 at the blood-brain barrier. *Acta Neuropathol.* 140, 549–567. doi: 10.1007/s00401-020-02187-x

Hawkins, B. T., and Davis, T. P. (2005). The blood-brain barrier/neurovascular unit in health and disease. *Pharmacol. Rev.* 57, 173–185. doi: 10.1124/pr.57.2.4

Heinrich, P. C., Behrmann, I. S., Hermanns, H. M., Muller-Newen, G., and Schaper, F. (2003). Principles of interleukin (IL)-6-type cytokine signalling and its regulation [Review]. *Biochem. J.* 374(Pt 1), 1–20. doi: 10.1042/bj2003 0407

Hu, Z., Deng, N., Liu, K., Zhou, N., Sun, Y., and Zeng, W. (2020). CNTF-STAT3-IL-6 axis mediates neuroinflammatory cascade across Schwann cell-neuronmicroglia. *Cell Rep.* 31:107657. doi: 10.1016/j.celrep.2020.107657

Huang, Z., Wong, L., Su, Y., Huang, X., Wang, N., Chen, H., et al. (2020). Blood-brain barrier integrity in the pathogenesis of Alzheimer's disease. *Front. Neuroendocrinol.* 59:100857. doi: 10.1016/j.yfrne.2020.100857

Janelsins, M. C., Mastrangelo, M. A., Oddo, S., LaFerla, F. M., Federoff, H. J., and Bowers, W. J. (2005). Early correlation of microglial activation with enhanced tumor necrosis factor-alpha and monocyte chemoattractant protein-1 expression specifically within the entorhinal cortex of triple transgenic Alzheimer's disease mice. J. Neuroinflammation 2:23. doi: 10.1186/1742-2094-2-23

Janelsins, M. C., Mastrangelo, M. A., Park, K. M., Sudol, K. L., Narrow, W. C., Oddo, S., et al. (2008). Chronic neuron-specific tumor necrosis factor-alpha expression enhances the local inflammatory environment ultimately leading to neuronal death in 3xTg-AD mice. *Am. J. Pathol.* 173, 1768–1782. doi: 10.2353/ ajpath.2008.080528

Jayaraj, R. L., Azimullah, S., Beiram, R., Jalal, F. Y., and Rosenberg, G. A. (2019). Neuroinflammation: Friend and foe for ischemic stroke. *J. Neuroinflammation* 16:142. doi: 10.1186/s12974-019-1516-2

Jones, S. A., and Jenkins, B. J. (2018). Recent insights into targeting the IL-6 cytokine family in inflammatory diseases and cancer. *Nat. Rev. Immunol.* 18, 773–789. doi: 10.1038/s41577-018-0066-7

Ju, Y., and Tam, K. Y. (2022). Pathological mechanisms and therapeutic strategies for Alzheimer's disease. *Neural Regen. Res.* 17, 543–549. doi: 10.4103/1673-5374.320970

Kitazawa, M., Cheng, D., Tsukamoto, M., Koike, M., Wes, P., Vasilevko, V., et al. (2011). Blocking IL-1 signaling rescues cognition, attenuates tau pathology, and restores neuronal  $\beta$ -catenin pathway function in an Alzheimer's disease model. *J. Immunol.* 187, 6539–6549. doi: 10.4049/jimmunol.1100620

Kong, X., Gong, Z., Zhang, L., Sun, X., Ou, Z., Xu, B., et al. (2019). JAK2/STAT3 signaling mediates IL-6-inhibited neurogenesis of neural stem cells through DNA demethylation/methylation. *Brain Behav. Immun.* 79, 159–173. doi: 10.1016/j.bbi. 2019.01.027

Korner, H., Lemckert, F. A., Chaudhri, G., Etteldorf, S., and Sedgwick, J. D. (1997a). Tumor necrosis factor blockade in actively induced experimental autoimmune encephalomyelitis prevents clinical disease despite activated T cell infiltration to the central nervous system. *Eur. J. Immunol.* 27, 1973–1981. doi: 10.1002/eji.1830270822

Korner, H., Riminton, D. S., Strickland, D. H., Lemckert, F. A., Pollard, J. D., and Sedgwick, J. D. (1997b). Critical points of tumor necrosis factor action in central nervous system autoimmune inflammation defined by gene targeting. *J. Exp. Med.* 186, 1585–1590. doi: 10.1084/jem.186.9.1585

Kuhn, H. G., and Svendsen, C. N. (1999). Origins, functions, and potential of adult neural stem cells. *BioEssays* 21, 625–630. doi: 10.1002/(SICI)1521-1878(199908)21:8<625::AID-BIES1>3.0.CO;2-6

Kuwar, R., Rolfe, A., Di, L., Xu, H., He, L., Jiang, Y., et al. (2019). A novel small molecular NLRP3 inflammasome inhibitor alleviates neuroinflammatory response following traumatic brain injury. *J. Neuroinflammation* 16:81. doi: 10. 1186/s12974-019-1471-y

Labus, J., Hackel, S., Lucka, L., and Danker, K. (2014). Interleukin-1beta induces an inflammatory response and the breakdown of the endothelial cell layer in an improved human THBMEC-based in vitro blood-brain barrier model. *J. Neurosci. Methods* 228, 35–45. doi: 10.1016/j.jneumeth.2014.03.002

Lambertsen, K., Finsen, B., and Clausen, B. (2019). Post-stroke inflammationtarget or tool for therapy? *Acta Neuropathol.* 137, 693–714. doi: 10.1007/s00401-018-1930-z

Lieberman, A. P., Pitha, P. M., Shin, H. S., and Shin, M. L. (1989). Production of tumor necrosis factor and other cytokines by astrocytes stimulated with lipopolysaccharide or a neurotropic virus. *Proc. Natl. Acad. Sci. U.S.A.* 86, 6348–6352. doi: 10.1073/pnas.86.16.6348

Liebner, S., Dijkhuizen, R. M., Reiss, Y., Plate, K. H., Agalliu, D., and Constantin, G. (2018). Functional morphology of the blood-brain barrier in health and disease. *Acta Neuropathol.* 135, 311–336.

Liesz, A., Dalpke, A., Mracsko, E., Antoine, D. J., Roth, S., Zhou, W., et al. (2015). DAMP signaling is a key pathway inducing immune modulation after brain injury. *J. Neurosci.* 35, 583–598. doi: 10.1523/JNEUROSCI.2439-14.2015

Lin, R., Chen, F., Wen, S., Teng, T., Pan, Y., and Huang, H. (2018). Interleukin-10 attenuates impairment of the blood-brain barrier in a severe acute pancreatitis rat model. *J. Inflammation* 15:4. doi: 10.1186/s12950-018-0180-0

Liu, H., Luiten, P., Eisel, U., Dejongste, M., and Schoemaker, R. (2013). Depression after myocardial infarction:  $TNF-\alpha$ -induced alterations of the bloodbrain barrier and its putative therapeutic implications. *Neurosci. Biobehav. Rev.* 37, 561–572. doi: 10.1016/j.neubiorev.2013.02.004

Liu, X., and Quan, N. (2018). Microglia and CNS interleukin-1: Beyond immunological concepts. *Front. Neurol.* 9:8. doi: 10.3389/fneur.2018.00008

Locksley, R. M., Killeen, N., and Lenardo, M. J. (2001). The TNF and TNF receptor superfamilies: Integrating mammalian biology. *Cell* 104, 487–501. doi: 10.1016/S0092-8674(01)00237-9

Lopez-Rodriguez, A., Hennessy, E., Murray, C., Nazmi, A., Delaney, H., Healy, D., et al. (2021). Acute systemic inflammation exacerbates neuroinflammation in Alzheimer's disease: IL-1 $\beta$  drives amplified responses in primed astrocytes and neuronal network dysfunction. *Alzheimers Dement.* 17, 1735–1755. doi: 10.1002/alz.12341

Madsen, P., Desu, H., de Rivero Vaccari, J., Florimon, Y., Ellman, D., Keane, R., et al. (2020). Oligodendrocytes modulate the immune-inflammatory response in EAE via TNFR2 signaling. *Brain Behav. Immun.* 84, 132–146. doi: 10.1016/j.bbi. 2019.11.017

Mantle, J., and Lee, K. (2018). A differentiating neural stem cell-derived astrocytic population mitigates the inflammatory effects of TNF- $\alpha$  and IL-6 in an iPSC-based blood-brain barrier model. *Neurobiol. Dis.* 119, 113–120. doi: 10.1016/j.nbd.2018.07.030

Mario, C., Antonio Rei, F., Niccolò, T., Daqing, M., Claudia, M., Marc, F., et al. (2010). Role of interleukin-1beta in postoperative cognitive dysfunction. *Ann. Neurol.* 68, 360–368.

McCoy, M. K., and Tansey, M. G. (2008). TNF signaling inhibition in the CNS: Implications for normal brain function and neurodegenerative disease. *J. Neuroinflammation* 5:45. doi: 10.1186/1742-2094-5-45

Menge, T., Zhao, Y., Zhao, J., Wataha, K., Gerber, M., Zhang, J., et al. (2012). Mesenchymal stem cells regulate blood-brain barrier integrity through TIMP3 release after traumatic brain injury. *Sci. Transl. Med.* 4:161ra150. doi: 10.1126/ scitranslmed.3004660

Mosser, D. M., and Zhang, X. (2008). Interleukin-10: New perspectives on an old cytokine. *Immunol. Rev.* 226, 205–218. doi: 10.1111/j.1600-065X.2008.00706.x

Moynagh, P., Williams, D., and O'Neill, L. (1994). Activation of NF-kappa B and induction of vascular cell adhesion molecule-1 and intracellular adhesion molecule-1 expression in human glial cells by IL-1. Modulation by antioxidants. *J. Immunol.* 153, 2681–2690.

Murakami, M., and Hirano, T. (2012). The pathological and physiological roles of IL-6 amplifier activation. *Int. J. Biol. Sci.* 8, 1267–1280. doi: 10.7150/ijbs.4828

Musella, A., Fresegna, D., Rizzo, F., Gentile, A., De Vito, F., Caioli, S., et al. (2020). 'Prototypical' proinflammatory cytokine (IL-1) in multiple sclerosis: Role in pathogenesis and therapeutic targeting. *Expert Opin. Ther. Targets* 24, 37–46. doi: 10.1080/14728222.2020.1709823

Nagaya, Y., Aoyama, M., Tamura, T., Kakita, H., Kato, S., Hida, H., et al. (2014). Inflammatory cytokine tumor necrosis factor alpha suppresses neuroprotective endogenous erythropoietin from astrocytes mediated by hypoxia-inducible factor-2alpha. *Eur. J. Neurosci.* 40, 3620–3626. doi: 10.1111/ejn.12747 Nasti, T. H., and Timares, L. (2012). Inflammasome activation of IL-1 family mediators in response to cutaneous photodamage. *Photochem. Photobiol.* 88, 1111–1125. doi: 10.1111/j.1751-1097.2012.01182.x

Netea, M. G., Simon, A., van de Veerdonk, F., Kullberg, B. J., Van der Meer, J. W., and Joosten, L. A. (2010). IL-1beta processing in host defense: Beyond the inflammasomes. *PLoS Pathog.* 6:e1000661. doi: 10.1371/journal.ppat.1000661

Ni, Y., Teng, T., Li, R., Simonyi, A., Sun, G. Y., and Lee, J. C. (2017). TNFalpha alters occludin and cerebral endothelial permeability: Role of p38MAPK. *PLoS One* 12:e0170346. doi: 10.1371/journal.pone.0170346

O'Carroll, S. J., Kho, D. T., Wiltshire, R., Nelson, V., Rotimi, O., Johnson, R., et al. (2015). Pro-inflammatory TNFalpha and IL-1beta differentially regulate the inflammatory phenotype of brain microvascular endothelial cells. *J. Neuroinflammation* 12:131. doi: 10.1186/s12974-015-0346-0

O'Connor, K. A., Hansen, M. K., Rachal Pugh, C., Deak, M. M., Biedenkapp, J. C., Milligan, E. D., et al. (2003). Further characterization of high mobility group box 1 (HMGB1) as a proinflammatory cytokine: Central nervous system effects. *Cytokine* 24, 254–265. doi: 10.1016/j.cyto.2003.08.001

Ohtsuki, S., and Terasaki, T. (2007). Contribution of carrier-mediated transport systems to the blood-brain barrier as a supporting and protecting interface for the brain; importance for CNS drug discovery and development. *Pharm. Res.* 24, 1745–1758. doi: 10.1007/s11095-007-9374-5

Okuma, Y., Date, I., and Nishibori, M. (2014). [Anti-high mobility group box-1 antibody therapy for traumatic brain injury]. *Yakugaku Zasshi* 134, 701–705. doi: 10.1248/yakushi.13-00255-2

Owen, J., Sultana, R., Aluise, C., Erickson, M., Price, T., Bu, G., et al. (2010). Oxidative modification to LDL receptor-related protein 1 in hippocampus from subjects with Alzheimer disease: Implications for A $\beta$  accumulation in AD brain. *Free Radic. Biol. Med.* 49, 1798–1803. doi: 10.1016/j.freeradbiomed.2010. 09.013

Padden, M., Leech, S., Craig, B., Kirk, J., Brankin, B., and McQuaid, S. (2007). Differences in expression of junctional adhesion molecule-A and betacatenin in multiple sclerosis brain tissue: Increasing evidence for the role of tight junction pathology. *Acta Neuropathol.* 113, 177–186. doi: 10.1007/s00401-006-0 145-x

Pandya, V., and Patani, R. (2021). Region-specific vulnerability in neurodegeneration: Lessons from normal ageing. *Ageing Res. Rev.* 67:101311. doi: 10.1016/j.arr.2021.101311

Patin, F., Baranek, T., Vourc'h, P., Nadal-Desbarats, L., Goossens, J. F., Marouillat, S., et al. (2016). Combined metabolomics and transcriptomics approaches to assess the IL-6 blockade as a therapeutic of ALS: Deleterious alteration of lipid metabolism. *Neurotherapeutics* 13, 905–917. doi: 10.1007/s13311-016-0461-3

Puthenparampil, M., Tomas-Ojer, P., Hornemann, T., Lutterotti, A., Jelcic, I., Ziegler, M., et al. (2021). Altered CSF albumin quotient links peripheral inflammation and brain damage in MS. *Neurol. Neuroimmunol. Neuroinflamm.* 8:e951. doi: 10.1212/NXI.000000000000951

Reichenberg, A., Yirmiya, R., Schuld, A., Kraus, T., Haack, M., Morag, A., et al. (2001). Cytokine-associated emotional and cognitive disturbances in humans. *Arch. Gen. Psychiatry* 58, 445–452. doi: 10.1001/archpsyc.58.5.445

Ren, C., Yao, R. Q., Zhang, H., Feng, Y. W., and Yao, Y. M. (2020). Sepsis-associated encephalopathy: A vicious cycle of immunosuppression. *J. Neuroinflammation* 17:14. doi: 10.1186/s12974-020-1701-3

Ren, C., Yin, P., Ren, N., Wang, Z., Wang, J., Zhang, C., et al. (2018). Cerebrospinal fluid-stem cell interactions may pave the path for cell-based therapy in neurological diseases. *Stem Cell Res. Ther.* 9:66. doi: 10.1186/s13287-018-0807-3

Robelin, L., and Gonzalez De Aguilar, J. L. (2014). Blood biomarkers for amyotrophic lateral sclerosis: Myth or reality? *Biomed Res. Int.* 2014:525097. doi: 10.1155/2014/525097

Rosejohn, S., and Heinrich, P. C. (1994). Soluble receptors for cytokines and growth factors: Generation and biological function. *Biochem. J.* 300(Pt 2), 281–290. doi: 10.1042/bj3000281

Rothe, M., Pan, M., Henzel, W., Ayres, T., and Goeddel, D. (1995). The TNFR2-TRAF signaling complex contains two novel proteins related to baculoviral inhibitor of apoptosis proteins. *Cell* 83, 1243–1252. doi: 10.1016/0092-8674(95) 90149-3

Sandoval, K., and Witt, K. (2008). Blood-brain barrier tight junction permeability and ischemic stroke. *Neurobiol. Dis.* 32, 200–219. doi: 10.1016/j.nbd. 2008.08.005

Sanjabi, S., Zenewicz, L. A., Kamanaka, M., and Flavell, R. A. (2009). Antiinflammatory and pro-inflammatory roles of TGF-beta, IL-10, and IL-22 in immunity and autoimmunity. *Curr. Opin. Pharmacol.* 9, 447–453. doi: 10.1016/j. coph.2009.04.008 Sato, S., Sanjo, H., Takeda, K., Ninomiya-Tsuji, J., Yamamoto, M., Kawai, T., et al. (2005). Essential function for the kinase TAK1 in innate and adaptive immune responses. *Nat. Immunol.* 6, 1087–1095. doi: 10.1038/ni1255

Sedger, L. M., and McDermott, M. F. (2014). TNF and TNF-receptors: From mediators of cell death and inflammation to therapeutic giants - past, present and future. *Cytokine Growth Factor Rev.* 25, 453–472. doi: 10.1016/j.cytogfr.2014.07. 016

Segarra, M., Aburto, M., and Acker-Palmer, A. (2021). Blood-brain barrier dynamics to maintain brain homeostasis. *Trends Neurosci.* 44, 393-405. doi: 10.1016/j.tins.2020.12.002

Sendama, W. (2020). The effect of ageing on the resolution of inflammation. Ageing Res. Rev. 57:101000. doi: 10.1016/j.arr.2019.101000

Si, Y., Zhao, Y., Hao, H., Fu, X., and Han, W. (2011). MSCs: Biological characteristics, clinical applications and their outstanding concerns. *Ageing Res. Rev.* 10, 93–103. doi: 10.1016/j.arr.2010.08.005

Smith, N., Giacci, M., Gough, A., Bailey, C., McGonigle, T., Black, A., et al. (2018). Inflammation and blood-brain barrier breach remote from the primary injury following neurotrauma. *J. Neuroinflammation* 15:201. doi: 10.1186/s12974-018-1227-0

Suvannavejh, G. C., Lee, H. O., Padilla, J., Dal Canto, M. C., Barrett, T. A., and Miller, S. D. (2000). Divergent roles for p55 and p75 tumor necrosis factor receptors in the pathogenesis of MOG(35-55)-induced experimental autoimmune encephalomyelitis. *Cell. Immunol.* 205, 24–33. doi: 10.1006/cimm.2000.1706

Sweeney, M., Zhao, Z., Montagne, A., Nelson, A., and Zlokovic, B. (2019). Blood-brain barrier: From physiology to disease and back. *Physiol. Rev.* 99, 21–78. doi: 10.1152/physrev.00050.2017

Tang, G., Liu, Y., Zhang, Z., Lu, Y., Wang, Y., Huang, J., et al. (2014). Mesenchymal stem cells maintain blood-brain barrier integrity by inhibiting aquaporin-4 upregulation after cerebral ischemia. *Stem Cells* 32, 3150–3162. doi: 10.1002/stem.1808

Tang, P., Hung, M. C., and Klostergaard, J. (1996). Human pro-tumor necrosis factor is a homotrimer. *Biochemistry* 35, 8216–8225. doi: 10.1021/bi952182t

Tartaglia, L., Pennica, D., and Goeddel, D. (1993). Ligand passing: The 75-kDa tumor necrosis factor (TNF) receptor recruits TNF for signaling by the 55-kDa TNF receptor. *J. Biol. Chem.* 268, 18542–18548. doi: 10.1016/S0021-9258(17) 46661-0

Thundyil, J., and Lim, K. L. (2015). DAMPs and neurodegeneration. Ageing Res. Rev. 24(Pt A), 17–28. doi: 10.1016/j.arr.2014.11.003

Tigges, U., Boroujerdi, A., Welser-Alves, J., and Milner, R. (2013). TNF-α promotes cerebral pericyte remodeling in vitro, via a switch from α1 to α2 integrins. *J. Neuroinflammation* 10:33. doi: 10.1186/1742-2094-10-33

Tobiume, K., Matsuzawa, A., Takahashi, T., Nishitoh, H., Morita, K., Takeda, K., et al. (2001). ASK1 is required for sustained activations of JNK/p38 MAP kinases and apoptosis. *EMBO Rep.* 2, 222–228. doi: 10.1093/embo-reports/kve046

van Kralingen, C., Kho, D. T., Costa, J., Angel, C. E., and Graham, E. S. (2013). Exposure to inflammatory cytokines IL-1beta and TNFalpha induces compromise and death of astrocytes; implications for chronic neuroinflammation. *PLoS One* 8:e84269. doi: 10.1371/journal.pone.0084269

Vázquez-Rosa, E., Shin, M., Dhar, M., Chaubey, K., Cintrón-Pérez, C., Tang, X., et al. (2020). P7C3-A20 treatment one year after TBI in mice repairs the bloodbrain barrier, arrests chronic neurodegeneration, and restores cognition. *Proc. Natl. Acad. Sci. U.S.A.* 117, 27667–27675. doi: 10.1073/pnas.2010430117

Vezzani, A., and Viviani, B. (2015). Neuromodulatory properties of inflammatory cytokines and their impact on neuronal excitability. *Neuropharmacology* 96, 70–82. doi: 10.1016/j.neuropharm.2014.10.027

Willis, E., MacDonald, K., Nguyen, Q., Garrido, A., Gillespie, E., Harley, S., et al. (2020). Repopulating microglia promote brain repair in an IL-6-dependent manner. *Cell* 180, 833–846.e16. doi: 10.1016/j.cell.2020.02.013

Wolf, J., Rose-John, S., and Garbers, C. (2014). Interleukin-6 and its receptors: A highly regulated and dynamic system. *Cytokine* 70, 11–20. doi: 10.1016/j.cyto. 2014.05.024

Xing, G., Zhao, T., Zhang, X., Li, H., Li, X., Cui, P., et al. (2020). Astrocytic sonic hedgehog alleviates intracerebral hemorrhagic brain injury *via* modulation of blood-brain barrier integrity. *Front. Cell. Neurosci.* 14:575690. doi: 10.3389/fncel.2020.575690

Yang, J., Ma, K., Zhang, C., Liu, Y., Liang, F., Hu, W., et al. (2020). Burns impair blood-brain barrier and mesenchymal stem cells can reverse the process in mice. *Front. Immunol.* 11:578879. doi: 10.3389/fimmu.2020.578879

Yang, S., Gu, C., Mandeville, E. T., Dong, Y., Esposito, E., Zhang, Y., et al. (2017). Anesthesia and surgery impair blood-brain barrier and cognitive function in mice. *Front. Immunol.* 8:902. doi: 10.3389/fimmu.2017.00902 Yang, T., Velagapudi, R., and Terrando, N. (2020). Neuroinflammation after surgery: From mechanisms to therapeutic targets. *Nat. Immunol.* 21, 1319–1326. doi: 10.1038/s41590-020-00812-1

Yasutaka, Y., Watanabe, T., Nakashima, A., Matsumoto, J., Futagami, K., Yamauchi, A., et al. (2015). Tumor necrosis factor-alpha reduces beta-amyloid accumulation primarily by lowering cellular prion protein levels in a brain endothelial cell line. *FEBS Lett.* 589, 263–268. doi: 10.1016/j.febslet.2014.12.007

Zhang, C., Han, Y., Liu, X., Tan, H., Dong, Y., Zhang, Y., et al. (2022). Odor enrichment attenuates the anesthesia/surgery-induced cognitive

impairment. Ann. Surg. [Epub ahead of print]. doi: 10.1097/SLA.00000000000 5599

Zhang, J., Sadowska, G. B., Chen, X., Park, S. Y., Kim, J. E., Bodge, C. A., et al. (2015). Anti-IL-6 neutralizing antibody modulates blood-brain barrier function in the ovine fetus. *FASEB J.* 29, 1739–1753. doi: 10.1096/fj.14-258822

Zhang, J., Takahashi, H. K., Liu, K., Wake, H., Liu, R., Maruo, T., et al. (2011). Anti-high mobility group box-1 monoclonal antibody protects the blood-brain barrier from ischemia-induced disruption in rats. *Stroke* 42, 1420–1428. doi: 10.1161/STROKEAHA.110.598334

# Glossary

AD, Alzheimer's disease; MS, multiple sclerosis; TBI, post-traumatic brain injury; BBB, blood-brain barrier; CNS, central nervous system; NSCs, neural stem cells; TNF $\alpha$ , tumor necrosis factor  $\alpha$ ; IL-1 $\beta$ , interleukin-1beta; IL-6, interleukin-6; iNOS, inducible nitric oxide synthase; BECs, brain endothelial cells; PCs, pericytes; ACs, astrocytes; TJs, tight junctions; AJs, adherens junctions; Cldn5, claudin-5; Ocln, occludin; ZO, zonula occludens; VE, vascular endothelial; Aqp4, aquaporin-4; Kir4.1, inwardly rectifying 4.1; shh, sonic hedgehog; VE-PTP, vascular endothelial protein tyrosine phosphatase; Tie2, type I tyrosine kinase receptor 2; ECs, endothelial cells; ICAM-1, intercellular adhesion molecule-1; VCAM-1, vascular cell adhesion molecule-1; MMPs, matrix metalloproteinases; PD, Parkinson's disease; TACE, TNFα-converting enzyme; TNFR, tumor necrosis factor receptor; DD, death domain; NF-κB, nuclear factor κB; EAE, experimental autoimmune encephalomyelitis; WT, wild-type; hCMEC/D3, immortalized human cerebral endothelial cell line; EPO, erythropoietin; PrPC, cellular prion protein; MBEC4, mouse brain microvascular endothelial cell line; IL-37, interleukin-37; HIF-1, hypoxia-inducible factor-1; VEGF-A, vascular endothelial growth factor-A; THBMEC, transfected human brain microvascular endothelial cell; TEER, transendothelial electrical resistance; IL-11,interleukin-11; IL-27, interleukin-27; IL-31, interleukin-31; OSM, oncostatin M; LIF, leukemia inhibitory factor; CNTF, ciliary neurotrophic factor; CT-1, cardiotrophin 1; CLCF1, cardiotrophinlike cytokine factor 1; gp130, membrane glycoprotein 130; CXCL, CXC- chemokine ligand; HMGB1, high-mobility group box 1 protein; DAMP, damage-associated molecular pattern; MHC, major histocompatibility complex; RAGE, receptor for advanced glycation endproducts; STAT3, signal transducers and activators of transcription; MAPK, mitogen-activated protein kinase; IL-10, interleukin-10; MSCs, mesenchymal stem cells.