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ICAM-1 K469E polymorphism is a **DEN** genetic determinant for the clinical risk factors of T2D subjects with retinopathy in Indians: a populationbased case-control study

Kumari Vinita,¹ Sarangapani Sripriya,¹ Krishnamurthy Prathiba,¹ Kulothungan Vaitheeswaran,² Ravichandran Sathyabaarathi,³ Mahendran Rajesh,² John Amali,² Vetrivel Umashankar,³ Govindasamy Kumaramanickavel,² Swakshyar Saumya Pal, 4 Rajiv Raman, 2 Tarun Sharma, 4 SNDREAMS project

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For numbered affiliations see end of article

Correspondence to

Dr Sarangapani Sripriya, drss@snmail.org, vatsanpriya@gmail.com

ABSTRACT

Objective: Elevated levels of intercellular adhesion molecule-1 (ICAM-1) are demonstrated in diabetes complications. The current study aims to understand association of K469E (rs5498) in ICAM-1 gene, in type 2 diabetic (T2D) subjects with retinopathy.

Design: Case-control study.

Setting: Sankara Nethralaya Diabetic Retinopathy Epidemiology and Molecular Genetic Study, an epidemiology study (on prevalence of diabetic retinopathy in T2D subjects (T2DR) from south India) and outpatient department of Sankara Nethralaya, a tertiary care hospital, in Chennai, India.

Participants: A total of 356 T2D subjects of >15 years of diabetes duration, with (n=199) and without (n=157) retinopathy.

Methods: The rs5498 polymorphism was genotyped by direct sequencing. Multivariate analysis for various clinical covariates was done using SPSS V.14. Comparative assessment of structure stability, folding rate of the variants were assessed using bioinformatics tools like STRIDE, MuPro, ModellerV97, fold rate server, etc.

Results: The AA genotype of rs5498 was seen at a higher frequency in the retinopathy group (p=0.012). The risk for diabetic retinopathy (DR) increased in the presence of AA genotype (OR=1.89-4.82) after the sequential addition of various clinical covariates. Multivariate logistic regression analysis showed 8.26 times high risk for developing DR in the AG genotype (p=0.003). Structural superimposition of ICAM-1 wild type (K469) and variant (E469) showed 0.943 Å of backbone root mean square deviation as calculated by PYMOL software. A difference in the fold rate time was also observed between the wild type (5.4/s) and variant (3.3/s).

Conclusions: This study shows that allele A of rs5498 in ICAM-1 is a putative risk predisposing allele for T2D retinopathy and its clinical covariates in Indian population. The folding rate of the protein decreases for the A allele implicating a potential effect on the structure and function of ICAM-1.

ARTICLE SUMMARY

Article focus

- Intercellular adhesion molecule-1 (ICAM-1) is a biomarker for inflammation and endothelial cell dysfunction and increased levels have been implicated in diabetic retinopathy (DR) pathology both in human and animal models. Strong heritability and genetic control on ICAM-1 expression have been observed by genome-wide association studies.
- The present study investigates the association of K469E (rs5498) polymorphism with DR in type 2 diabetic patients with ≥15 years duration of diabetes from south Indian population.

Key messages

- The AA genotype of rs5498 is a putative risk predisposing genotype for DR (OR=1.89-4.82) when adjusted for clinical covariates.
- The single-nucleotide polymorphism (SNP) imposes a change in the folding rate of the ICAM-1 protein that has potential functional implications.
- These results indicate antigenicity incobotulinumtoxinA.

Strengths and limitations of this study

- The detailed clinical evaluation and homogeneity of the study subjects, representing the southern part of India, is the main strength of the current study.
- The statistical approach that has accounted for the clinical confounding factors for DR has proved the strong correlation of the rs5498 with DR.
- With the limited available literature on the possible effect of rs5498 on ICAM-1 structure and function, the bioinformatics results provided an additional knowledge on the putative effect on the protein folding by the SNP through structure superimposition analysis that has potential futuristic research implications.
- Functional characterisation is the potential limitation of the study that could have further helped in proving the positive association observed for the AA genotype. The lack of correlation of serum ICAM-1 levels and sample size are other limitations.

INTRODUCTION

Diabetic retinopathy (DR) is reaching an alarming proportion in developing countries. In India DR has been reported as the sixth cause of blindness with an age-adjusted prevalence of 18% in diabetes subjects from rural and urban populations.¹

DR is characterised by retinal vessels basement membrane thickening, loss of pericytes and endothelial cells, blood-retinal barrier breakdown, capillary non-perfusion, microaneurysms, haemorrhages and neovascularisation.³ Several molecules and biochemical pathways like polyol pathway, activation of protein kinase C, formation of advanced glycation end products (AGEs), oxidative stress, upregulation of growth factors, adhesion molecules, etc have been implicated in the pathogenesis of DR. Recent research insights describe DR as a retinal disease associated with vascular neuroinflammation.3 Molecular and functional characterisation of the diabetic retina from animals, humans and cell culture studies have shown an increase in leukostasis, cytokines and growth factors resulting in breakdown of the blood-retinal barrier, thus implying the role of inflammation in DR.³⁻⁵ In lieu of such inferences, many anti-inflammatory molecules are being tested in recent years as a target for a possible remedy in DR.

Intercellular adhesion molecule-1 (ICAM-1) is a biomarker for endothelial cell dysfunction and inflammation that mediates leucocyte influx and persistent retinal leukostasis, retinal vascular leakage, capillary non-perfusion and endothelial cell injury and death subsequently resulting from Fas/FasL-mediated apoptosis.⁶ Its levels are upregulated along with the integrin ligands in patients with DR and retina of animal models.⁶ A decrease in the adherent retinal leucocytes have also been observed in ICAM-1 knock out animal models⁵ demonstrating the role of ICAM-1-mediated inflammation in DR pathogenesis.

Genetic variants in *ICAM-1* gene have been shown to regulate the expression level and have been widely studied for possible genetic association with a range of degenerative and inflammatory diseases including diabetic retinopathy.⁷ 8–15 The K469E (rs5498) polymorphism in exon 6 of *ICAM-1* gene has been shown to influence the binding of ICAM-1 on endothelial cells and LFA-1 and Mac-1 on leucocytes, mediating leukostasis and its migration in an inflammatory environment.¹⁶ This domain is essential for the structure and function of ICAM-1.¹⁷ Recent genomewide association studies have demonstrated a strong correlation between rs5498 and sICAM-1 levels.¹⁸ In the present study, we have investigated the association of the K469E polymorphism with retinopathy in type 2 diabetes (T2D) subjects from south India.

MATERIALS AND METHODS Sample collection

The patients were recruited prospectively from SNDREAMS (Sankara Nethralaya Diabetic Retinopathy Epidemiology and Molecular Biology Study), an epidemiology study to understand the prevalence of DR in south

Indian population¹ and from the outpatient departments of Sankara Nethralaya, a tertiary eye care hospital in Chennai.

The study subjects were enrolled between the years 2003 and 2010. The study protocol adhered to the Declaration of Helsinki and approved by institutional review board. After an informed consent all the subjects underwent detailed history, physical examination and pedigree analysis. Ocular examination included 45° fundus photograph using four-field stereoscopic digital photography that were graded by two independent observers in a masked fashion and the grading agreement showed a high κ value of 0.83. The diagnosis of DR was based on the modified Klein classification of the Early Treatment Diabetic Retinopathy Study scale. The methodology of sample selection were as described earlier. 19 20 The inclusion criteria for the study participants were T2D, south Indian origin and ≥40 years of age. The duration of diabetes varied between cases (≥10 years) and controls (>15 years). Subjects with sight threatening diabetic retinopathy (STDR, inclusive of severe nonproliferative diabetic retinopathy (NPDR), proliferative diabetic retinopathy (PDR) or clinically significant macular oedema (CSMO)) constitute the case group (DR+, n=199) and those without any signs of DR were included as the controls (DR-, n=157). Age-related macular degeneration (AMD), other hereditary retinal disease and non-south Indian origin were the exclusion criteria.

Genotyping

DNA was extracted from the peripheral blood samples by $method^{21}$ conventional phenol chloroform NucleoSpin Blood XL maxi kit method (Macherey-Nagel, Duren, Germany) according to the manufacturer's instructions. The genomic region flanking the K469E (rs5498; accession no. NT_011295) polymorphism in exon 6 of ICAM-1 was amplified with forward (5'-CTTGAGGGCACCTACCTCTG-3') and reverse (5'-CATTATGACTGCGGCTGCTA-3') using the following protocol: initial denaturation at 94°C for 5 min followed by 35 cycles of denaturation (94°C/30 s) primer annealing (60°C/1 min) and extension (72°C/1 min) followed by final extension at 72°C for 7 min. Genotype scoring was performed by direct sequencing in ABI PRISM 3100 Avant genetic analyser (Applied Biosystems, Foster City, California, USA).

Statistical analysis

Deviation from Hardy-Weinberg equilibrium(HWE) for genotypes was analysed. Statistical analyses were performed using SPSS software (for Windows V.14.0; SPSS Science, Chicago, Illinois, USA). The results were expressed as mean SD if the variables were continuous and as percentage if the variables were categorical. The Student's t test and χ^2 test were performed to compare continuous variables and proportions among groups, respectively. Distribution of genotypes and alleles

between the case and the control groups were compared using χ^2 test. To assess the specific effect of the genotypes on the various clinical factors, multivariate analysis was performed in the DR+ group. Multivariate analysis with stepwise sequential addition of various clinical variables was performed in the DR+ group with the three genotypes GG, AG and AA as the dependent variable. OR and 95% CIs were calculated and p<0.05 was considered significant.

Bioinformatic analysis

Sequence alignment analysis and the effect of polymorphism on structure and function of the protein are performed with BLAST (basic local alignment search tool), Polyphen-2 (polymorphism phenotyping v2) and SIFT (sorting intolerant from tolerant). Conservatonal analysis using ConSurf 9 was done with Swiss-Prot Accession ID: P05362 as the reference sequence.²² Three-dimensional structural co-ordinates of the ICAM-1 having the natural variant of rs5498, elucidated using x-ray crystallography at resolution of 2.70 Å was retrieved from Protein Data bank (PDBID: 2OZ4). The structure for K469 variant elucidated using Modeller9V723 and validated for quality by checking the stereo chemical and energetic aspects. The structural stability of the wild and variant was analysed through potential energy of the molecule. The effect of these variants at the cell adhesion was also studied through protein dimerisation and interaction with integrin α-M and β-2 using MUpro.²⁴

RESULTS

In the current study we have analysed the frequency distribution of the K469E (A>G, rs5498) polymorphism in

T2D subjects from south Indian population and analysed for the putative association of the same with DR. The demographic details of the study participants are given in table 1.

The genotypes were in HWE (p>0.05) The observed and expected frequencies for the homozygous, heterozgous variant did not deviate and were found to be consistent with HWE (p=0.313 and 0.316 for cases and controls, respectively. The AA genotype showed a higher frequency of distribution in the DR+ group when compared with the DR- (p=0.012); OR=1.94 (95% CI 1.06 to 3.55; table 2).

Results of multivariate analysis adjusted for the various clinical risk factors for DR are given in table 3. A negative association was observed for age/body mass index and age/high-density lipoprotein (HDL) in genotypes GG and AG, respectively. A positive association was observed for insulin usage and HbA1c in all the genotypes in addition to microalbuminuria which showed 8.26 times high risk for developing DR in the AG genotype.

Table 4 shows the multivariate logistic analysis performed with the genotypes (GG vs AG, GG vs AA and AG vs AA) as the independent variables and DR status as the dependent variables. Unadjusted analysis was performed initially, followed by sequential adjustment of the various clinical factors (covariates) mentioned in table 4. Significant value of p<0.05 and OR>1.0 was observed for the AA genotype when compared with the other genotypes {unadjusted p=0.923, OR=1.02, 95% CI 0.62 to 1.70 for GG vs AG; p=0.032, OR=1.94, 95% of CI 1.06 to 3.55 for GG vs AA; and p=0.019, OR=1.89, 95% of CI 1.11 to 3.22 for AG vs AA}. After adjusted for all the covariates, maximum OR with statistical significance was observed

Table 1 Baseline characteristics of the	study participants		
Variables	DR- control (n=157)	DR+ case (n=199)	р
Age (years)*	64.32±9.01	58.81±8.63	<0.0001
Male gender (n, %)	98 (62.4)	128 (64.3)	0.711
Duration of T2DM (years)*	18.44±6.18	17.74±5.45	0.261
User of insulin	34 (21.7)	83 (41.7)	< 0.0001
Age at diabetes onset (years)*	44.88±9.19	41.06±9.25	<0.0001
HbA1c (%)*	7.55±1.97	9.23±2.69	< 0.0001
HbA1c (mmol/mol)*	58.97±21.51	77.39±29.37	
Systolic BP (mm Hg)*	133.06±18.24	135.82±20.07	0.180
Diastolic BP (mm Hg)*	77.04±9.38	81.52±10.66	<0.0001
BMI* (kg/m ²)	25.33±7.78	23.79±5.36	0.031
History of hypertension	66 (42.0)	78 (39.2)	0.587
Smokers	23 (14.6)	37 (18.6)	0.321
Total cholesterol (mmol/l)*	4.29±1.10	4.05±1.03	0.069
HDL cholesterol (mmol/l)*	1.06±0.31	1.09±1.06	0.763
Triglycerides (mmol/l)*	1.17±0.61	1.12±0.54	0.541
Microalbuminuria	29 (19.9)	86 (58.5)	< 0.0001
Macroalbuminuria	3 (2.1)	14 (9.5)	0.010
*B			

*Data are M±SD. p<0.05: statistically significant.

BMI, basal metabolic index; BP, blood pressure; DR+, T2D subjects with retinopathy; DR-, T2D subjects without retinopathy; HbA1C, glycosylated haemoglobin; HDL, high-density lipoprotein; n, total subjects; T2DM, type 2 diabetes mellitus.

Table 2 Distribution of ICAM-1 rs5498 genotype and allele frequencies in DR+ and DR- groups

Genotype	Groups		p Values			
•	DR- controls (n=157)	DR+ case (n=199)		Dominant AA+AG=GG	Recessive AA=AG+GG	
GG	44 (28.0)	47 (23.6)	0.344	0.4	0.01*	
AG	84 (53.5)	92 (46.2)	0.173			
AA	29 (18.5)	60 (30.2)	0.012*			
Alleles						
G	172 (54.8)	186 (46.7)				
Α	142 (45.2)	212 (53.3)	0.033*			

*p<0.05—significant p value.

DR+, T2D subjects with retinopathy; DR-, T2D subjects without retinopathy; ICAM-1, intercellular adhesion molecule-1.

for AA (p=0.004; OR=4.82; 95% CI 1.64 to 14.14) when compared against AG genotype.

Bioinformatics analysis

Conservational analysis using ConSurf 9²² predicted the amino acid at position 469 to be of highly variable nature. Further, multiple sequence alignment of *ICAM-1* in different species showed the E469 genotype as a conservative variant. Possible pathogenic effects inferred using Polyphen and SIFT, predicted the variant to be benign and tolerable. Moreover, sequence-based stability analysis using MUpro²⁴ showed an increase in stability for the natural variant (E469) when compared with the wild (K469) with a confidence score of 0.10666991. Folding rate (using Fold rate server) was predicted to be 5.54/s and 3.3/s for the wild and variant proteins, respectively.

Three-dimensional structure of K469 ICAM-1 variant was modelled using the natural variant (PDBID: 2OZ4) as a template. The generated structure was loop refined using loop.py module of Modeller9v7. The steric clashes and bad contacts were removed using What-If server. Structural quality of the protein was assessed by checking the Ramachandran plot and ProQ server and tabulated (table 5). Moreover, the structure of wild and variant were energy optimised using optimised potential for liquid simulations force field for 1000 runs of steepest descent. Backbone superimposition of wild and variant forms of ICAM-1 showed RMSD (root mean square deviation) deviation of 0.943 Å. Since structural superimposition studies showed mild (figure 1), secondary structural analysis were performed using STRIDE²⁵ a software tool for secondary structure assignment from atomic resolution protein structures,

Table 3 Multivariate analysis between DR+ and DR- group for ICAM-1 rs5498 genotypes and the clinical covariates with the DR status as the dependable variable

GG		AG		AA			
Characteristics	DR+		DR+		DR+		
Number	OR (95% CI)	р	OR (95% CI)	р	OR (95% CI)	р	
Age (years)	0.91 (0.84 to 0.98)	0.019*	0.95 (0.91 to 0.99)	0.016*	0.92 (0.84 to 1.01)	0.077	
Male sex	3.28 (0.86 to 12.44)	0.081	0.94 (0.43 to 2.05)	0.880	1.82 (0.46 to 7.10)	0.391	
Duration of DM (years)	0.93 (0.82 to 1.06)	0.277	0.99 (0.94 to 1.06)	0.880	0.95 (0.82 to 1.06)	0.404	
User of insulin	4.35 (1.09 to 17.32)	0.037*	2.66 (1.24 to 5.69)	0.012*	5.44 (1.03 to 28.90)	0.047*	
Age at diabetes onset (years)	0.95 (0.89 to 1.01)	0.091	0.96 (0.93 to 1.00)	0.054	0.97 (0.90 to 1.04)	0.356	
HbA1c (DCCT) (%)	1.47 (1.14 to 1.89)	0.003*	1.32 (1.11 to 1.57)	0.002*	1.48 (1.07 to 2.04)	0.017*	
HbA1c (IFCC) mmol/mol)	1.04 (1.01 to 1.06)		1.03 (1.01 to 1.04)		1.04 (1.01 to 1.07)	33	
Systolic BP (mm Hg)	0.99 (0.96 to 1.03)	0.693	1.01 (0.98 to 1.02)	0.664	1.03 (0.98 to 1.07)	0.234	
Diastolic BP (mm Hg)	1.08 (0.99 to 1.18)	0.065	1.04 (1.00 to 1.08)	0.048*	1.08 (0.99 to 1.17)	0.069	
BMI	0.83 (0.70 to 0.97)	0.020*	0.93 (0.85 to 1.01)	0.102	0.84 (0.71 to 1.01)	0.059	
History of hypertension	1.70 (0.48 to 5.97)	0.407	0.69 (0.32 to 1.46)	0.332	1.26 (0.34 to 4.74)	0.731	
Smokers	1.17 (0.22 to 6.29)	0.857	1.10 (0.42 to 2.86)	0.846	3.72 (0.45 to 31.02)	0.225	
Total cholesterol (mmol/l)	0.29 (0.08 to 1.05)	0.060	0.99 (0.48 to 2.04)	0.972	0.13 (0.01 to 1.39)	0.092	
HDL cholesterol (mmol/l)	0.55 (0.01 to 55.94)	0.798	0.03 (0.00 to 0.80)	0.037*	6.83 (0.04 to 127.59)	0.471	
Triglycerides (mmol/l)	0.40 (0.03 to 5.75)	0.504	0.41 (0.07 to 2.49)	0.337	3.34 (0.07 to 15.72)	0.540	
Microalbuminuria	2.06 (0.21 to 20.26)	0.537	8.26 (2.06 to 33.11)	0.003*	3.59 (0.29 to 43.63)	0.316	

*p<0.05—significant p value.

BMI, basal metabolic index; BP, blood pressure; DCCT, Diabetes Control and Complications Trial; DM, diabetes mellitus; DR+, T2D subjects with retinopathy; DR-, T2D subjects without retinopathy; HbA1C, glycosylated haemoglobin; HDL, high-density lipoprotein; ICAM-1, intercellular adhesion molecule-1; IFCC, International Federation of Clinical Chemistry and Laboratory Medicine.

Table 4 Multivariate logistic analysis in DR+ group with sequential addition of clinical covariates with ICAM-1 rs5498 genotypes as the dependable variables

	DR+					
Characteristics	GG vs AG		GG vs AA		AG vs AA	
Number	OR (95% CI)	Р	OR (95% CI)	р	OR (95% CI)	р
Unadjusted	1.02 (0.62 to 1.70)	0.923	1.94 (1.06 to 3.55)	0.032*	1.89 (1.11 to 3.22)	0.019
Age	1.02 (0.60 to 1.74)	0.931	1.97 (1.05 to 3.69)	0.035*	1.92 (1.10 to 3.34)	0.022
Age+gender	1.02 (0.60 to 1.74)	0.932	2.00 (1.06 to 3.77)	0.031*	1.93 (1.10 to 3.36)	0.021
Age+gender+DD	1.01 (0.59 to 1.73)	0.958	2.00 (1.06 to 3.77)	0.031*	1.94 (1.11 to 3.39)	0.020
Age+gender+DD+insulin	1.04 (0.61 to 1.80)	0.873	2.28 (1.19 to 4.36)	0.012*	2.12 (1.20 to 3.75)	0.010
Age+gender+DD+insulin+HbA1c	1.31 (0.73 to 2.34)	0.368	1.83 (1.25 to 2.69)	0.002*	2.12 (1.16 to 3.85)	0.014
Age+gender+DD+insulin+HbA1c+systolic BP	1.26 (0.70 to 2.28)	0.439	1.79 (1.22 to 2.65)	0.003*	2.15 (1.17 to 3.93)	0.013
Age+gender+DD+insulin+HbA1c+systolic BP+diastolic BP	1.28 (0.70 to 2.32)	0.425	1.81 (1.21 to 2.70)	0.004*	2.12 (1.15 to 3.91)	0.016
Age+gender+DD+insulin+HbA1c+systolic BP+diastolic BP+BMI	1.31 (0.71 to 2.42)	0.377	1.97 (1.29 to 3.01)	0.002	2.21 (1.18 to 4.13)	0.013
Age+gender+DD+insulin+HbA1c+systolic BP+diastolic BP+BMI+SMK	1.31 (0.71 to 2.41)	0.392	1.99 (1.29 to 3.06)	0.002*	2.22 (1.18 to 4.16)	0.013
Age+gender+DD+insulin+HbA1c+systolic BP+diastolic BP+BMI+SMK+microalbumin	1.25 (0.60 to 2.59)	0.552	2.04 (1.26 to 3.32)	0.004*	3.17 (1.41 to 7.08)	0.005
Age+gender+DD+insulin+HbA1c+systolic BP+diastolic BP+BMI+SMK+microalbumin+triglyceride	0.68 (0.27 to 1.75)	0.430	1.77 (0.98 to 3.21)	0.059	3.75 (1.28 to 10.98)	0.016
Age+gender+DD+insulin+HbA1c+systolic BP+diastolic BP+BMI+SMK+microalbumin+triglyceride+HDL	0.74 (0.27 to 1.97)	0.543	1.75 (0.97 to 3.18)	0.064	4.17 (1.39 to 12.53)	0.011
Age+gender+DD+insulin+HbA1c+systolic BP+diastolic BP+BMI+SMK+microalbumin+triglyceride+HDL+cholesterol	0.67 (0.25 to 1.84)	0.439	1.80 (0.98 to 3.34)	0.060	4.07 (1.34 to 12.35)	0.013
Age+gender+DD+insulin+HbA1c+BMI+cholesterol+ HDL+SMK+triglyceride+microalbumin+HOHT	0.63 (0.23 to 1.72)	0.365	1.72 (0.98 to 3.02)	0.061	3.56 (1.23 to 10.03)	0.020
Gender+onset of diabetes+insulin+HbA1c+systolic BP+diastolic BP+BMI+SMK+cholesterol+HDL+triglyceride+microalbumin	0.67 (0.26 to 1.76)	0.417	1.89 (1.03 to 3.49)	0.041*	4.82 (1.64 to 14.14)	0.004

^{*}Significant p value.

BMI, basal metabolic index; BP, blood pressure; DD, duration of diabetes; DM, diabetes mellitus; DR+, T2D subjects with retinopathy; DR-, T2D subjects without retinopathy; HbA1C, glycosylated haemoglobin; HDL, high-density lipoprotein; HOHT, history of hypertension; ICAM-1, intercellular adhesion molecule-1; SMK, smoking.

Table 5 Comparison of the structural properties of the wild (KK) and variant (EE) proteins for SNP rs5498 of ICAM-1 gene

Experimental type	Wild Homology modelling	Variant PDBID: 20Z4
		Resolution:
		2.70 Å
Residues in most	95.5%	88.8%
favoured regions		
Residues in additional	4.0%	10.8%
allowed region		
Residues in generously	0.4%	0.4%
allowed regions		
Residues in disallowed	0.0%	0.0%
regions		
G-factor	-0.04	0.13
Bond lengths		
Main chain	99.8%	100%
Within limits	0.2%	0.0%
Bond angles		
Main chain	:93.9%	100%
Within limits	6.1%	0.0%
Planarity		
Planar groups	100%	100%
Within limits	0.0%	0.0%
Energy minimisation	-2.9554834e	-3.7942434e
using Gromacs for	+04	+04
1000 runs of steepest		
descent		
ICAM-1, intercellular adhesi	on molecule-1; SNF	P, single nucleotide

that, however, did not show any significant change

DISCUSSION

polymorphism.

between the wild and the variant.

In the present study, the frequency distribution of K469E (rs5498) polymorphism in the *ICAM-I* gene was analysed for its possible association with T2D retinopathy from south India. A higher frequency of AA genotype is being observed in the DR+ when compared with the DR- group (OR=1.94 (95% CI 1.06 to 3.55; p, 0.012)). The frequency of rs5498 polymorphism observed in the current study

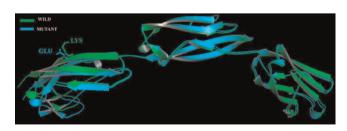


Figure 1 Structural superimposition of intercellular adhesion molecule-1 wild type K469 (green) and variant E469 (blue) with a 0.943 Å of backbone root mean square deviation, as calculated by PYMOL. The variant residues are shown in stick representation.

simulates the other reports from India on ICAM-1 gene single nucleotide polymorphisms (SNPs). ²⁶ ²⁷

The association of rs5498 AA genotype with DR identified in the current study simulates the Japanese and Chinese study on PDR populations. The A allele of rs5498 was also observed to confer disease susceptibility in type 1 diabetes (T1D) patients with nephropathy of Swedish Caucasian origin. A high heterozygous index was observed in the current study is similar to that observed in GoKinD study. However, another similar study from Caucasian cohort shows significant association with GG genotype suggesting the population difference per se. 15

Our results also differ from another similar study in India, by Balasubbu *et al.*²⁹ The possible reasons for the observed differences between our study and the other reports from India could be attributed to the area of sampling and the types of DR included in the study. The present study represents a more homogenous population from the same geographical area of southern India against that of Balasubbu *et al* which represents a hospital-based population. We have included STDR patients with PDR, NPDR or CSMO while the ARAVIND study includes only PDR similar to the Caucasian study.

Strong heritability factor has been observed for circulating levels of sICAM-1 by bivariate quantitative genetic analyses³⁰ and upregulated expressions are being reported in animal models and PDR patients.⁶ Such elevated levels are reported to be influenced by glycaemic control, disturbances in lipid metabolism, obesity and insulin resistance, etc which are also important clinical determinants of DR. Hence we performed a multivariate logistic regression analysis for the different genotypes between DR+ and DR–groups after adjusting these parameters.

In the current study, we observed an OR of 8.0 (95% CI 2.06 to 33.11) for the heterozygous genotype (AG) in the DR group after adjusting for microalbuminuria (table 3) and a genotype-dependent risk was observed after the sequential addition of the gender (table 4). Plasma levels of ICAM-1, vascular cell adhesion molecule 1 were increased in T2D patients with microalbuminuria thus suggesting a significant correlation between the same. In GoKinD population allele G has been detected to confer a decreased risk susceptibility to the development of diabetic nephropathy in female T1D patients.

ICAM-1 expression has been reported to share a common genetic modulation with traits related to obesity, insulin resistance and HDL3 cholesterol. We observed significant p value with OR>1.0 for the AA genotype after the sequential addition of lipid biomarkers (table 4). An abnormal endothelial activation after an oral lipid meal, coupled with an increased oxidative stress is being observed in patients with familial history of T2D. High-fat meals are shown to increase ICAM-1 and other adhesion molecules in normal and diabetic subjects. Similarly, a recent study on the coeffects of inflammation and endothelial dysfunction and insulin resistance on hypertension in a large Asian population reports elevated levels of biomarkers for inflammation and endothelial cell dysfunction including

sICAM-1.³⁴ The ICAM-1 GG (E469E) genotype was reported as an independent contribution factor of plasma fibrinogen level as well as HDL cholesterol and urinary albumin excretion in atherosclerosis.¹¹ However, we did not identify any statistically significant association for the GG genotype with these two parameters in the current study.

Collectively, multivariate analysis after sequential addition of various DR risk factors reveal a high susceptibility for DR (OR>1, table 4) in the AA genotype of rs5498 for all the DR covariates. Interestingly, these parameters are reported to have a significant effect on ICAM-1 expression. 32–34 Estimation of ICAM-1 levels in individuals with E469K polymorphism has been reported to be associated with many inflammatory and infectious diseases. However, population differences are seen. 35–37 These differences could be due to the dietary influence/epigenetic silencing of ICAM-1 expression by hypermethylation as reported in animal and human models. 38 39

Any disease-associated polymorphism can possibly mediate the effect either through altered expression or function of the protein. The rs5498 is located at threebase position upstream of the splice donor site that produces an alternatively spliced short isoform (ICAM-1-S) that has no transmembrane or intracellular domain and speculated to influence the ICAM-1 signal transduction and cell-cell contact including Fas-FasL interaction.⁴⁰ Thus, correlated decrease in Fas-associated death domain-like interleukin-1-β-converting enzyme-inhibitory protein long form (FLIPL) mRNA expression and apoptosis suggests a putative role of the polymorphism in regulating apoptosis by modifying the inflammatory immune responses. 40 Comparison of the RNA splicing patterns in cells expressing G/G (469E) and A/A (469K) genotypes showed a comparatively higher expression of ICAM-1-S mRNA in A/A cells. 40

The polymorphism rs5498 results in a non-conservative change from lysine to glutamic acid in the fifth immunoglobulin-like domain of ICAM-114 that is essential for dimerisation, surface presentation and solubilisation of the protein.¹⁷ We therefore performed a bioinformatic analysis to study the putative effect of this SNP on the ICAM-1 structure and thus its influence on the expression. As per the sequence analysis, the variant K469E was shown as benign without any significant secondary structural change. The x-ray structure of ICAM-1 consists of Ig-like C2-type domain 3, 4 and 5 that consist of four intradisulphide bridges as per Swiss-Prot annotation and it includes 237-290, 332-371, 403-419 and 431-457 (http://www. uniprot.org/uniprot/P05362). It could therefore be inferred that the variant K469E does not affect the disulphide bridges. Interestingly, structure superimposition of the two variants (figure 1), revealed a 0.943 Å deviation of backbone RMSD as calculated by the software PYMOL thus suggesting a structural effect of the SNP. The difference in the fold rate time observed between the KK (5.4/s) when compared with the EE (3.3/s) variant highlights the need of further dimerisation studies.

Our results indicate that the AA genotype of *ICAM-1* (rs5498) gene increases the risk predisposition for retinopathy in T2D patients in south Indian population. Clinical covariates such as microalbuminuria, lipid biomarkers, etc show a genotype-dependent (AG/AA) increase in the risk for DR among T2D patients. Bioinformatics analysis of rs5498 showed a deviation in the structure and folding rate of the ICAM-1 protein. These observations emphasise the need for further studies to identify the molecular mechanism connecting the SNP, expression, protein structure and function.

Author affiliations

¹SN ONGC Department of Genetics and Molecular Biology, Vision Research Foundation, Chennai, Tamil Nadu, India

²Sankara Nethralaya Diabetic Retinopathy Project, Sankara Nethralaya, Chennai, Tamil Nadu, India

³Centre for Bioinformatics, Vision Research Foundation, Chennai, Tamil Nadu, India

⁴Shri Bhagwan Mahavir Vitreoretinal Services, Medical Research Foundation, Sankara Nethralaya, Chennai, Tamil Nadu, India

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REFERENCES

- Rani PK, Raman R, Sharma V, et al. Analysis of a comprehensive diabetic retinopathy screening model for rural and urban diabetics in developing countries. Br J Ophthalmol 2007;91:1425–9.
- Rani PK, Raman R, Agarwal S, et al. Diabetic retinopathy screening model for rural population: awareness and screening methodology. Rural Remote Health 2005;5:350.
- Liou GI. Diabetic retinopathy: role of inflammation and potential therapies for anti-inflammation. World J Diabetes 2010;1:12–18.
- Kern TS. Contributions of inflammatory processes to the development of the early stages of diabetic retinopathy. Exp Diabetes Res 2007;2007:1–14.
- Joussen AM, Poulaki V, Le ML, et al. A central role for inflammation in the pathogenesis of diabetic retinopathy. FASEB J 2004; 18:1450–2.
- Swenarchuk LE, Whetter LE, Adamis AP. The role of inflammation in the pathophysiology of diabetic retinopathy. *Contemp Diabetes* 2008; 2:303–31.
- Papa A, Danese S, Urgesi R, et al. Intercellular adhesion molecule 1 gene polymorphisms in inflammatory bowel disease. Eur Rev Med Pharmacol Sci 2004;8:187–91.
- Ji YN, Wang Q, Zhan P. Intercellular adhesion molecule 1 gene K469E polymorphism is associated with coronary heart disease risk: a meta-analysis involving 12 studies. Mol Biol Rep 2011 2012;39:6043–8.
- Milutinovic A, Petrovic D. The K469E polymorphism of the intracellular adhesion molecule 1 (ICAM-1) gene is not associated with myocardial infarction in Caucasians with type 2 diabetes. Folia Biol (Praha) 2006;52:79–80.

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- Nejentsev S, Laaksonen M, Tienari PJ, et al. Intercellular adhesion molecule-1 K469E polymorphism: study of association with multiple sclerosis. Hum Immunol 2003;64:345–9.
- Yokoyama H, Tahara H, Emoto M, et al. The K469E polymorphism of the intercellular adhesion molecule-1 gene is associated with plasma fibrinogen level in type 2 diabetes. Metabolism 2005;54:381–6.
- Ma J, Mollsten A, Prazny M, et al. Genetic influences of the intercellular adhesion molecule 1 (ICAM-1) gene polymorphisms in development of type 1 diabetes and diabetic nephropathy. Diabetes Med 2006;23:1093–9
- Kamiuchi K, Hasegawa G, Obayashi H, et al. Intercellular adhesion molecule-1 (ICAM-1) polymorphism is associated with diabetic retinopathy in type 2 diabetes mellitus. *Diabetes Med* 2002;19: 371–6
- Liu L, Yu Q, Wang H, et al. Association of intercellular adhesion molecule 1 polymorphisms with retinopathy in Chinese patients with type 2 diabetes. Diabet Med 2006;23:643

 –8.
- Petrovic MG, Osredkar J, Saraga-Babic M, et al. K469E polymorphism of the intracellular adhesion molecule 1 gene is associated with proliferative diabetic retinopathy in Caucasians with type 2 diabetes. Clin Exp Ophthalmol 2008;36:468–72.
- Gundel RH, Letts LG. Adhesion molecules and the modulation of mucosal inflammation (chapter 3). In: Goldie R, ed., Immunopharmacology of epithelial barriers. London: Elsevier, 1994; 273n
- Miller J, Knorr R, Ferrone M, et al. Intercellular adhesion molecule-1 dimerization and its consequences for adhesion mediated by lymphocyte function associated-1. J Exp Med 1995;182:1231–41.
- Pare G, Ridker PM, Rose L, et al. Genome-wide association analysis of soluble ICAM-1 concentration reveals novel associations at the NFKBIK, PNPLA3, RELA, and SH2B3 loci. PLoS Genet 2011;7:e1001374.
- Agarwal S, Raman R, Paul PG, et al. Sankara Nethralaya Diabetic Retinopathy Epidemiology and Molecular Genetic Study (SNDREAMS1): study design and research methodology. Ophthalmic Epidemiol 2005;12:143–53.
- Pal SS, Raman R, Ganesan S, et al. Sankara Nethralaya Diabetic Retinopathy Epidemiology and Molecular Genetic Study (SN– DREAMS III): study design and research methodology. BMC Ophthalmol 2011;11:7
- Wolff R, Gemmil R. Purifying and analyzing genomic DNA. In: Birren B, Green ED, Hieter P, et al., eds. Genome analysis: a laboratory manual. Vol. 1. New York: Cold Spring Harbor Laboratory Press. 1997:1–82
- Goldenberg O, Erez E, Nimrod G, et al. The ConSurf-DB: pre-calculated evolutionary conservation profiles of protein structures. Nucleic Acids Res 2009;37(Database issue):D323–7. http://conseq.tau.ac.il (accessed 15 June 2011).
- Sali A, Matsumoto R, McNeil HP, et al. Three-dimensional models of four mouse mast cell chymases. Identification of proteoglycan-binding regions and protease-specific antigenic epitopes. J Biol Chem 1993:268:9023–34.
- Cheng J, Randall A, Baldi P. Prediction of protein stability changes for single-site mutations using support vector machines. *Proteins* 2006;62:1125–32. http://www.igb.uci.edu/servers/servers.html (accessed 15 June 2011).

- Heinig M, Frishman D. STRIDE: a web server for secondary structure assignment from known atomic coordinates of proteins. *Nucleic Acids Res* 2004;32(Web Server issue): W500–2. http:// webclu.bio.wzw.tum.de/stride/ (accessed 26 nov 2011).
- Sinha S, Qidwai T, Kanchan K, et al. Variations in host genes encoding adhesion molecules and susceptibility to falciparum malaria in India. Malar J 2008;7:250.
- Sengupta S, Farheen S, Mukherjee N, et al. DNA sequence variation and haplotype structure of the ICAM1 and TNF genes in 12 ethnic groups of India reveal patterns of importance in designing association studies. Ann Hum Genet 2004;68:574–87.
- Ma J, Zhang D, Brismar K, et al. Evaluation of the association between the common E469K polymorphism in the ICAM-1 gene and diabetic nephropathy among type 1 diabetic patients in GoKinD population. BMC Med Genet 2008;9:47.
- Balasubbu S, Sundaresan P, Rajendran A, et al. Association analysis of nine candidate gene polymorphisms in Indian patients with type 2 diabetic retinopathy. BMC Med Genet 2010;11:158.
- Bielinski SJ, Pankow JS, Foster CL, et al. Circulating soluble ICAM-1 levels shows linkage to ICAM gene cluster region on Chromosome 19: the NHLBI Family Heart Study follow-up examination. Atherosclerosis 2008;199:172–8.
- Bruno CM, Valenti M, Bertino G, et al. Plasma ICAM-1 and VCAM-1 levels in type 2 diabetic patients with and without microalbuminuria. Minerva Med 2008;99:1–5.
- Kent JW, Comuzzie AG, Mahaney MC, et al. Intercellular adhesion molecule-1 concentration is genetically correlated with insulin resistance, obesity, and HDL concentration in Mexican Americans. *Diabetes* 2004;53:2691–5.
- Madec S, Corretti V, Santini E, et al. Effect of a fatty meal on inflammatory markers in healthy volunteers with a family history of type 2 diabetes. Br J Nutr 2011;106:364–8.
- Li H, Zhu X, Wang A, et al. Co-effect of insulin resistance and biomarkers of inflammation and endothelial dysfunction on hypertension. Hypertens Res 2012;35:513–17.
- Hwang SJ, Ballantyne CM, Sharrett RA, et al. Circulating adhesion molecules VCAM-1, ICAM-1, and E-selectin in carotid atherosclerosis and incident coronary heart disease cases. The atherosclerosis risk in communities (ARIC) study. Circulation 1997:96:4219–25.
- Lu FH, Shang Q, Wen PE, et al. A study on K469E polymorphism of ICAM1 gene and ICAM1 plasma level in patients with coronary heart disease. Zhonghua Yi Xue Yi Chuan Xue Za Zhi 2006;23:195–7.
- Sanadgol N, Nikravesh A, Motalleb G, et al. Evaluation of the association between ICAM-1 gene polymorphisms and sICAM-1 serum levels in multiple sclerosis (MS) patients in Southeast Iran. Int J Genet Mol Biol 2011;3:81–6.
- Brake DK, Smith EO, Mersmann H, et al. Robker. ICAM-1 expression in adipose tissue: effects of diet-induced obesity in mice. Am J Physiol Cell Physiol 2006;29: C1232–9.
- Hellebrekers DM, Castermans K, Vire E, et al. Epigenetic regulation of tumor endothelial cell anergy: silencing of intercellular adhesion molecule-1 by histone modifications. Cancer Res 2006:66:10770-7.
- Iwao M, Morisaki H, Morisaki T. Single-nucleotide polymorphism g.1548G > A (E469K) in human ICAM-1 gene affects mRNA splicing pattern and TPA- induced apoptosis. *Biochem Biophys Res* Commun 2004;317:729–35.