

Anisakis pegreffii Larvae in Sea Eels (*Astroconger myriaster*) from the South Sea, Republic of Korea

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Abstract: *Anisakis simplex* sensu stricto (s.s.), *Anisakis pegreffii*, *Anisakis berlandi* (= *A. simplex* sp. C), and *Anisakis typica* are the 4 major species of *Anisakis* type I larvae. In the Republic of Korea (Korea), *A. pegreffii*, *A. berlandi*, and *A. typica* larvae in fish hosts has seldom been documented. In this study, molecular analysis was performed on *Anisakis* larvae from the sea eels (*Astroconger myriaster*), the major source of human anisakiasis in Korea, collected from Tongyeong City, a southern coastal area of Korea. All 20 sea eels examined were infected with *Anisakis* type I larvae (160 larvae; 8 per fish). Their species were analyzed using PCR-RFLP patterns and nucleotide sequences of internal transcribed spacers (ITS1, 5.8 subunit gene, and ITS2) and mitochondrial cytochrome c oxidase 2 (*cox2*). Most (86.8%; 112/129) of the *Anisakis* type I larvae were *A. pegreffii*, and 7.8% (10/129) were *A. typica*. The remaining 5.4% (7/129) was not identified. Thus, *A. pegreffii* is the major species of anisakid larvae in sea eels of the southern coast of Korea.

Key words: *Anisakis pegreffii*, sea eel, PCR-RFLP, ITS1, *cox2*

Human anisakiasis is caused mainly by the larvae of the genera *Anisakis* and *Pseudoterranova* [1]. The 2 most important species are *Anisakis simplex* sensu stricto (s.s.) and *Pseudoterranova decipiens* [1,2]. However, since 1999, a few human infections with *Anisakis pegreffii* larvae (a sibling species of *A. simplex* s.s.) have been reported by molecular analysis in Italy [3-6] and Japan [7,8]. Subsequently, human infections with *A. pegreffii* larvae were confirmed in 2015 in 15 of 16 clinical patients in the Republic of Korea (Korea), from whom the larvae were extracted by gastroduodenoscopy [9]. The number of human *A. pegreffii* cases may be much higher than hitherto considered.

The sources of human anisakiasis are marine fish and squids [1,2]. Many studies have reported on the taxonomy and classification of anisakid larvae recovered in these hosts. Several morphological types were described, including *Anisakis* type I, *Anisakis* type II, *Terranova* type A, *Terranova* type B, *Contracaecum* type A, and *Contracaecum* type B [1,10]. However, since the development of molecular techniques, it became apparent

that even within a type several species or strains may be included. For example, *Anisakis* type I larvae include at least 4 different species: *A. simplex* s.s., *A. pegreffii*, and *Anisakis berlandi* (*A. simplex* sp. C), and *Anisakis typica* [11].

In Korea, studies have been performed on anisakid larvae in various marine fish species [12-19] including the sea eel (*Astroconger myriaster*), which is one of the most important sources of human anisakiasis in Korea [1]. In the sea eel, most studies reported the presence of *Anisakis* type I larvae (some authors interpreted them as *A. simplex* s.s. larvae), and the presence of *A. pegreffii* larvae has seldom been documented with the exception of a recent report that detected *A. pegreffii* larvae by molecular analysis of the gene [20]. In other fish species and squids, only 2 reports [18,19] described the presence of *A. pegreffii* larvae in fish and squids caught in Korea. Thus, the present study aimed to investigate the status of *A. pegreffii* larvae infection in the sea eels caught from the South Sea, Korea.

Twenty sea eels (*A. myriaster*) were purchased in March 2013 at Guri Agricultural and Marine Products Market, Gyeonggi-do, Korea. The sea eels were stated to have been caught near Tongyeong city in the south coast of Korea. In total, 160 *Anisakis* type I third-stage larvae (L3) were collected; 129 of these 160 larvae were molecularly analyzed.

The total genomic DNA (gDNA) was extracted by DNeasy Blood and Tissue Kit (Qiagen, Hilden, Germany) according to

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the manufacturer's instructions. Identification to the species level was carried out by PCR-RFLP and sequencing of a 274-900 bp fragment of internal transcribed spacer (ITS1, 5.8 subunit rRNA, and ITS2) gene and a 582-629 bp fragment of the mitochondrial cytochrome *c* oxidase 2 (*cox2*) gene. A region of the nuclear ribosomal DNA (rDNA) was amplified using the ITS A (5'-GTC GAA TTC GTA GGT GAA CCT GCG GAA GGA TCA-3') and B (5'-GCC GGA TCC GAA TCC TGG TTA GTT TCT TTT CCT-3') (21) with Smart 2× PCR premix Taq (Solgent, Daejeon, Korea), containing 10 pmol of each primer and 30 ng of the total DNA. The mixture undergone initial denaturation at 95°C for 10 min, followed by 30 cycles of denaturation at 95°C for 30 sec, annealing at 55°C for 30 sec, extension at 72°C for 1.15 min with a final extension at 72°C for 7 min. The *cox2* gene was amplified using the primers 211F 5'-TTT TCT AGT TAT ATA GAT TGR TTY AT-3' and 210R 5'-CAC CAA CTC TTA AAA TTA TC-3' (22). The mixture was denatured at 94°C for 3 min, followed by 34 cycles at 94°C for 30 sec, 46°C for 1 min, and 72°C for 1.5 min, followed by post-amplification at 72°C for 10 min. The ITS PCR products (17 µl) were digested with 10 units of restriction endonuclease *Hinf*I (1 µl) (Enzynomics, Daejeon, Korea) and 10× EZ-one buffer (2 µl) (Enzynomics) in a final volume of 20 µl at 37°C for 1 hr. The digested products were then separated by electrophoresis on 3% agarose gels containing 1 µg/ml ethidium bromide and visualized under ultraviolet light. The nucleotide sequences of ITS and *cox2* regions were obtained from 3-6 larval specimens and aligned using the program Geneious v.6.0.3.

All of the sea eels examined were found to be infected with

Anisakis larvae (20/20, 100%), and a total of 160 *Anisakis* type I larvae (8.0 per fish) were collected. The larvae had a boring tooth, a ventriculus, and a mucron. The anterior portion showed a prominent boring tooth in the cephalic region. In the digestive tract, the ventriculus level showed the simply connected esophagus, ventriculus, and intestine. The posterior portion showed a mucron in the caudal region.

Out of the 129 *Anisakis* larvae identified by PCR-RFLP analysis, an approximately 900 bp fragment was produced after amplification of the rDNA region (ITS1, 5.8 subunit, and ITS2). The PCR products were processed to identify the species with restriction enzyme *Hinf*I. *Hinf*I is the most appropriate and the best known enzyme for anisakid larvae identification [7,23]. RFLP produced 2 patterns. One revealed 3 different fragments approximately between 250 and 500 bp (*A. pegreffii*) and the other displayed 2 different fragments between 250 bp and 1,000 bp (*A. typica*). Most (86.8%; 112/129) larvae were diagnosed as *A. pegreffii*, whereas 7.8% (10/129) were diagnosed as *A. typica*. The remaining 5.4% (7/129) could not be diagnosed (Fig. 1).

The final confirmation of the species was made after sequencing of the ITS and *cox2*. Three randomly selected *Anisakis* type I larvae were undergone sequencing of the ITS region (274 bp), and all of the samples revealed 100% identity with *A. pegreffii* (GenBank no. AB277823) and 99.2% identity with *A. simplex* s.s. (no. AB277822) (Table 1). In addition, 6 randomly selected samples underwent *cox2* sequencing of the 582-629 bp region. Results indicated that the larvae were identical (97.8-100%) with *A. pegreffii* (no. EU413958) rather than

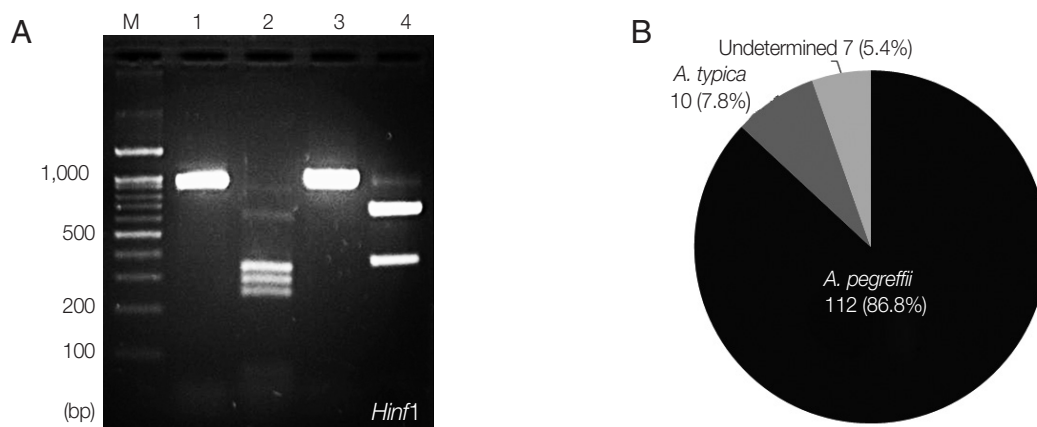


Fig. 1. (A) PCR-RFLP patterns of the rDNA region spanning the ITS-1, 5.8S, and ITS-2, for *A. pegreffii* and *A. typica* from the sea eels. M, 100-bp ladder. Lanes 1 and 3, gDNA from *Anisakis* spp.; lane 2, *A. pegreffii* (370, 300, and 250 bp); lane 4, *A. typica* (350 and 620 bp). (B) The infection status of the sea eels with different species of *Anisakis* larvae (n = 129).

Table 1. Nucleotide identity (%) of our specimens with the known *Anisakis pegreffii*, *A. simplex* s.s. and *A. typica* sequences in GenBank for the ITS region and *cox2*

	ITS region			<i>cox2</i>		
	<i>A. pegreffii</i> (EU413958)	<i>A. simplex</i> s.s (AB517560)	<i>A. typica</i> (AB517571)	<i>A. pegreffii</i> (AB277823)	<i>A. simplex</i> s.s. (AB277822)	<i>A. typica</i> (AY826724)
Sample 1	100.0	99.2	63.4	98.0	95.2	86.9
Sample 2	100.0	99.2	63.4	99.6	94.9	87.4
Sample 3	100.0	99.2	63.4	98.0	95.5	86.9
Sample 4	-	-	-	97.8	94.9	86.8
Sample 5	-	-	-	99.6	94.6	87.1
Sample 6	-	-	-	100.0	94.9	87.1

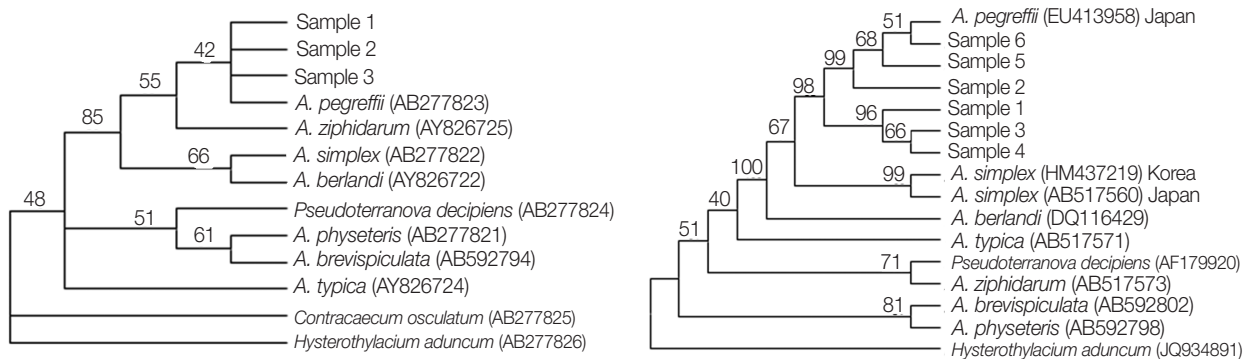


Fig. 2. Neighbor-Joining trees of *Anisakis* spp. based on (A) ITS region (274 bp) and (B) *cox2* gene (511 bp) exploring the relationships among *Anisakis* spp. available in GenBank and *Anisakis* larvae collected from the sea eels in Korea.

with *A. simplex* s.s. (AB517560) (94.9-95.5%) (Table 1). Based on this information, phylogenetic trees of anisakid nematodes based on ITS and *cox2* sequences were constructed using the neighbor-joining method (Free Software Foundation, Boston, Massachusetts, USA) (Fig. 2).

Human anisakiasis is an important fish-borne parasitic zoonosis. The infection is caused by the ingestion of raw or improperly cooked fish infected by the larvae [1,2]. The overall infection rate of *Anisakis* larvae in the sea eels was 100%, with the infection density of 8 larvae per fish. As seen from its infection status, the sea eel is suspected as one of the most important paratenic fish hosts that may be related with human anisakiasis in Korea [1].

A. pegreffii was first reported from a Mediterranean monk seal in 1955 [24]. *A. pegreffii* was subsequently repeatedly reported in marine fish, squids, but rarely humans, in various localities including Europe, Australia, America, and Asia (including Japan, Chia, and Korea) [7,9,25]. Because the morphology of *A. pegreffii* larvae is almost indistinguishable from *A. simplex* s.s. larvae, electrophoretic analysis was first used for

the differentiation [26]. Later, other molecular techniques, including PCR-RFLP and nucleotide sequencing, were developed, and these techniques are most popularly used [3-9].

Human *A. pegreffii* infection was first reported in Italy using PCR-RFLP [3]. The second was from Japan, in which 1 of 85 anisakid larvae extracted from human anisakiasis patients in Kyushu was identified as *A. pegreffii* by PCR-RFLP of the ITS region [7]. Thereafter, 11 human cases have been described in Italy [4-6]. Recently, 15 human infections with *A. pegreffii* were confirmed in Korea by nucleotide sequencing of the ITS region [9]. Therefore, at present, 28 molecularly confirmed human *A. pegreffii* infections have been reported in the literature.

With regard to the *Anisakis* larvae from fish in Korea, most previous reports designated the larvae simply as *Anisakis* type I [12-15], and sometimes *A. simplex* larvae (this means *A. simplex* complex) were used to designate them [16,17]. According to a recent report, the prevalence of *Anisakis* larvae was high; 52.3% (45 of 86) of fish from the East Sea, 76.6% (131 of 171) of fish from the South Sea, and 40.2% (37 of 92) of fish from the Yellow Sea [17]. In Busan, a port city on the South

Sea, 2,537 fish and cephalopods were collected, and the overall infection rate of *A. simplex* larvae (i.e., *A. simplex* complex) was 34.3%, and the intensity was 17.1 and 5.9 larvae per fish and cephalopod, respectively, with the highest abundance in the spring season [16].

The diagnosis of *A. pegreffii* for the larvae from fish in Korea was first given to those collected from marine fish (*Scomber japonicus* and *Trichiurus lepturus*) and squids (*Tadarodes pacificus*) purchased from wholesale and retail markets in 2009 based on PCR-RFLP patterns of the ITS region [18]. It is noteworthy that most of the larvae were *A. pegreffii* (47/60 larvae; 78.3%) and *A. typica* (10/60; 16.7%), while only 1 (from a squid) of 60 larvae was *A. simplex* s.s. [18]. In contrast, a study of *Anisakis* larvae from the chum salmon (*Oncorhynchus keta*) caught in an eastern coastal area of Korea revealed that all 48 larvae analyzed by PCR-RFLP of mitochondrial DNA *cox2* region were consistent with *A. simplex* s.s. [19]. Another study using PCR-RFLP and sequencing of ITS and mitochondrial *cox1* regions reported that most of the larvae collected from 10 fish species from the Yellow Sea, South Sea, and East Sea were *A. pegreffii*, whereas very few were *A. simplex* s.s. [20]. Our study is consistent with 2 recent studies in Korea [18,20]. The significance of detecting *A. typica* larvae in sea eels remains to be further studied.

The geographical distribution of *A. pegreffii* larvae in Korea and Japan is interesting. In Japan, *A. pegreffii* larvae were more frequently detected than *A. simplex* s.s. in the western part (the East and South Seas of Korea), whereas *A. simplex* s.s. larvae dominated in the eastern part (the Pacific Ocean) [27]. In Korea, most larvae were *A. pegreffii* in various kinds of marine fish and squids [18,20]. In the present study, 86.8% (112/129) of the larvae were *A. pegreffii* and 7.8% (10/129 larvae) were *A. typica*. The predominance of *A. pegreffii* in the sea eels and other marine fish and cephalopods seems to be closely related to the predominance of human *A. pegreffii* infection (15 of 16 human cases) in Korea [9]. Molecular studies on the larvae extracted from human infections are needed in Korea.

CONFLICT OF INTEREST

We have no conflict of interest related to this work.

REFERENCES

- Sohn WM, Chai JY. Anisakiosis (Anisakidosis). In Palmer SR, Soulsby L, Torgerson PR, Brown WG, eds, Oxford Textbook of Zoonoses. 2nd ed. Oxford, UK: Oxford University Press; 2011. pp 774-786.
- Chai JY, Murrell KD, Lymbery AJ. Fish-borne parasitic zoonoses: status and issues. *Int J Parasitol* 2005; 35: 1233-1254.
- D'Amelio S, Mathiopoulou KD, Brandonisio O, Lucarelli G, Doro F, Paggi L. Diagnosis of a case of gastric anisakidosis by PCR-based restriction fragment length polymorphism analysis. *Parassitologia* 1999; 41: 591-593.
- Fumarola L, Monno R, Ierardi E, Rizzo G, Giannelli G, Lalle M, Pozio E. *Anisakis pegreffii* etiological agent of gastric infections in two Italian women. *Foodborne Pathog Dis* 2009; 6: 1157-1159.
- Mattiucci S, Paoletti M, Borrini F, Palumbo M, Palmieri RM, Gomes V, Casati A, Nascetti G. First molecular identification of the zoonotic parasite *Anisakis pegreffii* (Nematoda: Anisakidae) in a paraffin-embedded granuloma taken from a case of human intestinal anisakiasis in Italy. *BMC Infect Dis* 2011; 11: 82 (1-6).
- Mattiucci S, Fazii P, Rosa AD, Paoletti M, Megna AS, Glielmo A, de Angelis M, Costa A, Meucci C, Calvaruso V, Sorrentini I, Palma G, Bruschi F, Nascetti G. Anisakiasis and gastroallergic reactions associated with *Anisakis pegreffii* infection, Italy. *Emerg Infect Dis* 2013; 19: 496-499.
- Umehara A, Kawakami Y, Araki J, Uchida A. Molecular identification of the etiological agent of the human anisakiasis in Japan. *Parasitol Int* 2007; 56: 211-215.
- Arizono N, Yamada M, Tegoshi T, Yoshikawa M. *Anisakis simplex* sensu stricto and *Anisakis pegreffii*: biological characteristics and pathogenetic potential in human anisakiasis. *Foodborne Pathog Dis* 2012; 9: 517-521.
- Lim H, Jung BK, Cho J, Yooyen T, Shin EH, Chai JY. Molecular diagnosis of cause of anisakiasis in humans, South Korea. *Emerg Infect Dis* 2015; 21: 342-344.
- Chai JY, Cho SR, Kook J, Lee SH. Infection status of the sea eel (*Astroconger myriaster*) purchased from the Noryangjin fish market with anisakid larvae. *Korean J Parasitol*. 1992;30:157-162.
- Mattiucci S, Cipriani P, Webb SC, Paoletti M, Marcer F, Bellisario B, Gibson DI, Nascetti G. Genetic and morphological approaches distinguish the three sibling species of the *Anisakis simplex* species complex, with a species designation as *Anisakis berlandi* n. sp. for *A. simplex* sp. C (Nematoda: Anisakidae). *J Parasitol* 2014; 100: 199-214.
- Chai JY, Cho YM, Sohn WM, Lee SH. Larval anisakids collected from the yellow corvine in Korea. *Korean J Parasitol* 1986; 24: 1-11.
- Kim KH, Joo KH, Rim HJ. A study about infection state of anisakis larvae and parasitic helminths in salmon (*Oncorhynchus keta*) and sea trout (*Oncorhynchus masou*) which were caught from Taepo Port, Kangwon Do. *Korean J Rural Med* 1990; 15: 27-32.
- Song SB, Hwang EG. Infection status of larval anisakids in *Astroconger myriaster* collected from the Southern Sea near Pusan. *Korean J Parasitol* 1992; 30: 263-267.
- Song SB, Lee SR, Chung HH, Han NS. Infection status of ani-

- sakid larvae in anchovies purchased from local fishery market near southern and eastern sea in Korea. Korean J Parasitol 1995; 33: 95-99 (in Korean).
16. Choi SH, Kim J, Jo JO, Cho MK, Yu HS, Cha HJ, Ock MS. *Anisakis simplex* larvae: infection status in marine fish and cephalopods purchased from the Cooperative Fish Market in Busan, Korea. Korean J Parasitol 2011; 49: 39-44.
 17. Cho SH, Lee SE, Park OH, Na BK, Sohn WM. Larval anisakid infections in marine fish from three sea areas of the Republic of Korea. Korean J Parasitol 2012; 50: 295-299.
 18. Lee MH, Cheon DS, Choi C. Molecular genotyping of *Anisakis* species from Korean sea fish by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP). Food Control 2009; 20: 623-626.
 19. Setyobudi E, Jeon CH, Lee CH, Seong KB and Kim JH. Occurrence and identification of *Anisakis* spp. (Nematoda: Anisakidae) isolated from chum salmon (*Oncorhynchus keta*) in Korea. Parasitol Res 2011; 108: 585-592.
 20. Sohn WM, Kang JM, Na BK. Molecular analysis of *Anisakis* type I larvae in marine fish from three different sea areas in Korea. Korean J Parasitol 2014; 52: 383-389.
 21. D'Amelio S, Mathiopoulos KD, Santos CP, Pugachev ON, Webb SC, Picanço M, Paggi L. Genetic markers in ribosomal DNA for the identification of members of the genus *Anisakis* (Nematoda: ascaridoidea) defined by polymerase-chain-reaction-based restriction fragment length polymorphism. Int J Parasitol 2000; 30: 223-226.
 22. Nadler SA, Hudspeth DSS. Phylogeny of the Ascaridoidea (Nematoda: Ascaridida) based on three genes and morphology: hypotheses of structural and sequence evolution. J Parasitol 2000; 86: 380-393.
 23. Umehara A, Kawakami Y, Matsui T, Araki J, Uchida A. Molecular identification of *Anisakis simplex* sensu stricto and *Anisakis pegreffii* (Nematoda: Anisakidae) from fish and cetacean in Japanese waters. Parasitol Int 2006; 55: 267-271.
 24. Campana-Rouget PY, Biocca E. Une Nouvelle Espèce D'*Anisakis* Chez un Phoque Mediterranean. Ann de Parasitol 1955; 30: 477-480.
 25. Zhang L, Hu M, Shamsi S, Beveridge I, Li H, Xu Z, Li L, Cantacessi C, Gasser RB. The specific identification of anisakid larvae from fishes from the Yellow Sea, China, using mutation scanning-coupled sequence analysis of nuclear ribosomal DNA. Mol Cell Probes 2007; 21: 386-390.
 26. Nascetti G, Paggi L, Orecchia P, Smith JW, Mattiucci S, Bullini L. Electrophoretic studies on the *Anisakis simplex* complex (Ascaridida: Anisakidae) from the Mediterranean and North-East Atlantic. Int J Parasitol 1986; 16: 633-640.
 27. Quiazon KMA, Yoshinaga T, Ogawa K. Distribution of *Anisakis* species larvae from fishes of the Japanese waters. Parasitol Int 2011; 60: 223-226.

