

# KPC-3-Producing *Klebsiella pneumoniae* Sequence Type 392 from a Dog's Clinical Isolate in Portugal

### Joana Moreira da Silva,<sup>a,b</sup> Duliana Menezes,<sup>a,b</sup> Gabriel Mendes,<sup>c</sup> Sofia Santos Costa,<sup>d</sup> Cátia Caneiras,<sup>c,e</sup> Laurent Poirel,<sup>f</sup> Andreia J. Amaral,<sup>a,b</sup> Constança Pomba<sup>a,b,g</sup>

<sup>a</sup>Centre for Interdisciplinary Research in Animal Health (CIISA), Faculty of Veterinary Medicine, University of Lisbon, Lisbon, Portugal <sup>b</sup>Associate Laboratory for Animal and Veterinary Sciences (AL4AnimalS), Lisbon, Portugal

<sup>c</sup>Microbiology Research Laboratory on Environmental Health (EnviHealthMicro Lab), Institute of Environmental Health, Faculty of Medicine (ISAMB), University of Lisbon, Lisbon, Portugal

<sup>d</sup>Global Health and Tropical Medicine (GHTM), Instituto de Higiene e Medicina Tropical (IHMT), Nova University of Lisbon, Lisbon, Portugal

elnstitute of Preventive Medicine and Public Health, Faculty of Medicine, University of Lisbon, Lisbon, Portugal

Microbiology Spectrum

AMERICAN SOCIETY FOR MICROBIOLOGY

fMedical and Molecular Microbiology Unit, Faculty of Science and Medicine, University of Fribourg, Fribourg, Switzerland 9Genevet, Veterinary Molecular Diagnostic Laboratory, Carnaxide, Portugal

## **KEYWORDS** Carbapenemases, companion animals, multidrug-resistant *Klebsiella pneumoniae*

Resistance to carbapenems in Enterobacterales poses a threat to health care systems worldwide since those infections are associated with high mortality and limited treatment options. The latest European reports show that some southern European countries are almost endemic for carbapenemase-producing Enterobacterales (CPEs) (1, 2). Mirroring the European tendency, Portugal has been observing a steady increase in the occurrence of carbapenem-resistant *Klebsiella pneumoniae*, particularly KPC-3-producing *K. pneumoniae* ST147 lineage (Clonal Group 147) strains in health care settings (3, 4).

To assess the relevance of CPEs in veterinary health care, 977 Enterobacterales isolates obtained at the veterinary molecular diagnostic laboratory (Genevet) during 2020 were screened for carbapenem resistance. Phenotypic disc testing with 10  $\mu$ g meropenem, 30  $\mu$ g temocillin, and MAST CAT-ID discs (Mast Group, UK) and genotypic confirmatory tests

**TABLE 1** Antimicrobial MICs and resistance genes identified during WGS analysis on KPC-3producing Klebsiella pneumoniae ST392<sup>a</sup>

Antimicrobials tested	MIC (mg/L)	Susceptibility phenotype <sup>b</sup>	AMR genes
Amikacin	≤8	S	aac(6')-lb-cr
Ampicillin	>16	R	bla <sub>TEM-1B</sub> ; bla <sub>SHV-11</sub> ; bla <sub>CTX-M-15</sub> ; bla <sub>KPC-3</sub>
Amoxicillin-clavulanic acid	>16/8	Rc	bla <sub>CTX-M-15</sub> ; bla <sub>KPC-3</sub>
Aztreonam	>16	R	bla <sub>CTX-M-15;</sub> bla <sub>KPC-3</sub>
Cefotaxime	>32	R	bla <sub>CTX-M-15</sub> ; bla <sub>KPC-3</sub>
Ceftazidime	>16	R	bla <sub>CTX-M-15</sub> ; bla <sub>KPC-3</sub>
Ceftazidime-avibactam	2	S	NA
Ciprofloxacin	>2	R	aac(6')-lb-cr; OqxB/A; qnrB1;
Colistin	≤2	S	NA
Ertapenem	>1	R	bla <sub>KPC-3</sub>
Gentamicin	≤2	S	aac(6')-lb-cr
Imipenem	>8	R	bla <sub>KPC-3</sub>
Meropenem	>8	R	bla <sub>KPC-3</sub>
Tetracycline	>8	R <sup>c</sup>	tet(A)
Trimethoprim/sulfamethoxazole	≤2/73	S <sup>c</sup>	sul2; OqxB/A

<sup>a</sup>S, susceptible; R, resistant; NA, not applicable; AMR, antimicrobial resistance; WGS, whole genome sequencing. <sup>b</sup>Susceptibility phenotype was determined according to EUCAST breakpoint guidelines.

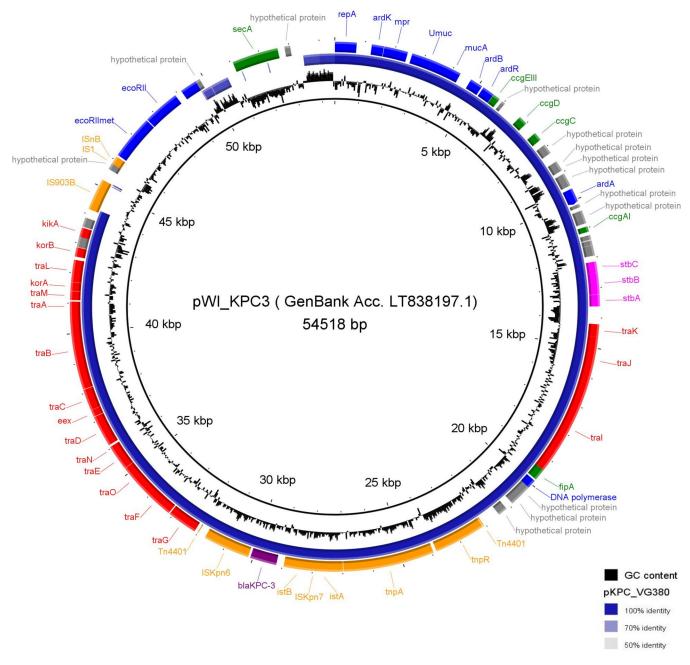
<sup>c</sup>Susceptibility phenotype was determined according to Clinical and Laboratory Standards Institute guidelines (9).

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Address correspondence to Constança Pomba, cpomba@fmv.ulisboa.pt.

The authors declare a conflict of interest. This project was funded by CIISA and FCT Project UIDB/00276/2020 and LA/P/0059/2020 - AL4AnimalS. J.M.D.S. and J.M. were supported by a Fundação para a Ciência e Tecnologia (FCT) PhD fellowship (2020.06540.BD; 2020.07562.BD; respectively).

Published 23 June 2022



**FIG 1** Plasmid alignment comparison between *de novo* assembled contig pKPC\_VG380 (GenBank accession number PRJNA808048) found on the strain KPC-3-producing *Klebsiella pneumoniae* taken from a dog's with an upper respiratory tract infection (in blue) and plasmid pWI\_KPC3 (GenBank accession number LT838197.1), used as backbone plasmid reference, previously described on a French nosocomial isolate (11). Genes are represented by colored blocks: purple, resistance gene; blue, DNA replication, regulation, and restriction systems; red, conjugation-association genes; fuchsia, genes associated with partition and stability systems; orange, transposons, insertion sequences (IS), and transposase genes; green, other genes; gray, hypothetical proteins. Image generated using BRIG 0.95, available at http://brig.sourceforge.net/.

were performed to detect plasmid-mediated AmpC  $\beta$ -lactamase and ESBL- and carbapenemase-producing Enterobacterales (5, 6).

Interestingly, a carbapenem-resistant and carbapenemase-producing *K* pneumoniae isolate (VG380) was recovered, being positive for the  $bla_{\rm KPC-type}$  gene. It was recovered from a nasal swab collected from a 10-year-old dog with an upper respiratory tract infection. Cytology of the nasal exudate displayed an abundance of degenerated neutrophils, bacilli, and cocci. Biopsy of the nasal tissue displayed an ulceration showing signs of mixed inflammation, with an abundance of neutrophils, lymphocytes, macrophages, and dense fibrovascular proliferation, which was compatible with the diagnosis of chronic ulcerative rhinitis. A 1-month

course of treatment with trimethoprim-sulfamethoxazole was followed and the dog made a full recovery.

K. pneumoniae strain VG380 was resistant to ertapenem, meropenem, and imipenem (MIC >1 mg/L for ertapenem and >8 mg/L for imipenem and meropenem; determined by MicroSan NEG44 plates [Beckman Coulter, USA]). It was susceptible to ceftazidime-avibactam (MIC = 2 mg/L; determined by Etest [bioMérieux, France]). Interpretation of MIC determinations followed EUCAST breakpoint tables (https://www.eucast.org/fileadmin/src/media/PDFs/EUCAST\_files/Breakpoint\_tables/v\_12.0\_Breakpoint\_Tables.pdf).

Whole Genome Sequencing (WGS) analysis of strain VG380 was performed using Illumina NovaSeq platform with  $2 \times 150$ -bp paired-end reads. The quality of the resulting raw reads was evaluated using FastQC v0.11.5 (https://www.bioinformatics.babraham.ac.uk/projects/fastqc/). *De novo* genomes were assembled using SPAdes v3.14.1 (7), following two rounds of polishing with Pilon v1.24 (8). The generated assemblies were used to screen for antimicrobial resistance genes and mobile genetic elements resorting to tools available at Centre for Genomic Epidemiology (http://genomicepidemiology.org/), ResFinder 4.1 and Mobile Element Finder v1.0.3, respectively. MLST 2.0 and pMLST 2.0 were also performed.

WGS analysis showed that VG380 possessed a KPC-3-encoding gene and belonged to Sequence Type ST392, a single-locus variant of ST147, also belonging to Clonal Group 147. Therefore, this canine strain was related to the KPC-3-producing *K. pneumoniae* ST147 line-age responsible for the majority of nosocomial infections reported so far in Portugal (3, 4). Virulence factors *traT* and *iutA* were identified, with the former being located on a IncFII(K)-type plasmid. IncFIB(K)- and IncN- type plasmids were also identified. Noteworthy, strain VG380 harbored a series of resistance genes leading to a quite unusual multi-drug resistance profile for a canine isolate (Table 1).

The  $bla_{KPC-3}$  gene was located within transposon Tn4401d (10) on an ~50-kb lncN-type plasmid (pKPC\_VG380, pMLST ST15) (GenBank accession number PRJNA808048) (Fig. 1). When using a previously described lncN plasmid carrying the  $bla_{KPC-3}$  gene in Tn4401 (pWI\_KPC3, pMLST ST15) of clinical human origin (11) as a reference for BLAST, it was possible to demonstrate that one of the contigs for the KPC-3-producing *K. pneumoniae* ST392 VG380 had a large extent of homology with the reference plasmid (Fig. 1).

We report here, to the best of our knowledge, the first characterization of a KPC-3-producing *K. pneumoniae* strain, belonging to the human high-risk clonal group 147 isolated from a dog in Europe. Noteworthy, isolates belonging to the high-risk emerging lineage ST392 from CG147 and carrying the  $bla_{KPC-3}$  gene have been reported in a single Portuguese central hospital during 2020 (4).

To prevent spreading of carbapenem resistance, a one-health surveillance approach is urgent. The study of the epidemiology of plasmid-mediated carbapenem resistance seems to be relevant for the identification of possible pathways for antimicrobial resistance transmission.

### ACKNOWLEDGMENTS

We express our gratitude to the team from Genevet Laboratory. This project was funded by CIISA and Fundação para a Ciência e Tecnologia (FCT) Project UIDB/00276/2020 and LA/P/0059/2020 - AL4AnimalS. J.M.D.S. and J.M. were supported by a FCT PhD fellowship (2020.06540.BD and 2020.07562.BD; respectively).

#### REFERENCES

- European Centre for Disease Prevention and Control. 2019. Carbapenem-resistant Enterobacteriaceae, second update–26 September 2019. ECDC, Stockholm. https://www.ecdc.europa.eu/sites/default/files/documents/carbapenem-resistant -enterobacteriaceae-risk-assessment-rev-2.pdf.
- David S, Reuter S, Harris SR, Glasner C, Feltwell T, Argimon S, Abudahab K, Goater R, Giani T, Errico G, Aspbury M, Sjunnebo S, Feil EJ, Rossolini GM, Aanensen DM, Grundmann H, ESGEM Study Group. 2019. Epidemic of carbapenem-resistant Klebsiella pneumoniae in Europe is driven by nosocomial spread. Nat Microbiol 4:1919–1929. https://doi.org/10.1038/s41564-019-0492-8.
- Aires-de-Sousa M, Ortiz de la Rosa JM, Gonçalves ML, Pereira AL, Nordmann P, Poirel L. 2019. Epidemiology of carbapenemase-producing Klebsiella pneumoniae in a Hospital, Portugal. Emerg Infect Dis 25:1632–1638. https:// doi.org/10.3201/eid2509.190656.
- Guerra AM, Lira A, Lameirão A, Selaru A, Abreu G, Lopes P, Mota M, Novais Â, Peixe L. 2020. Multiplicity of carbapenemase-producers three years after a kpc-3-producing k. pneumoniae st147-k64 hospital outbreak. Antibiotics 9:806–811. https://doi.org/10.3390/antibiotics9110806.
- Marques C, Belas A, Franco A, Aboim C, Gama LT, Pomba C. 2018. Increase in antimicrobial resistance and emergence of major international high-risk clonal lineages in dogs and cats with urinary tract infection: 16 year retrospective study. J Antimicrob Chemother 73:377–384. https://doi.org/10.1093/jac/dkx401.
- Poirel L, Walsh TR, Cuvillier V, Nordmann P. 2011. Multiplex PCR for detection of acquired carbapenemase genes. Diagn Microbiol Infect Dis 70:119–123. https://doi.org/10.1016/j.diagmicrobio.2010.12.002.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N,

Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.

- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal .pone.0112963.
- Clinical and Laboratory Standards Institute (CLSI). 2018. Performance standards for antimicrobial disk and dilution susceptibility testes for

bacteria isolated from animals (4th). CLSI Supplement VET08. CLSI, Wayne, PA.

- Mataseje LF, Boyd DA, Willey BM, Prayitno N, Kreiswirth N, Gelosia A, Poutanen SM, Low DE, Jenkins SG, Katz K, Mulvey MR. 2011. Plasmid comparison and molecular analysis of Klebsiella pneumoniae harbouring blakpc from New York city and Toronto. J Antimicrob Chemother 66:12 73–1277. https://doi.org/10.1093/jac/dkr092.
- Beyrouthy R, Robin F, Lessene A, Lacombat I, Dortet L, Naas T, Ponties V, Bonnet R. 2017. MCR-1 and OXA-48 in vivo acquisition in KPC-producing Escherichia coli after colistin treatment. Antimicrob Agents Chemother 61:1–4.