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ORIGINAL RESEARCH Mechanism of Qiguiyin Decoction Sensitizing Levofloxacin Against Multidrug-Resistant Pseudomonas aeruginosa Infection Based on PK-PD and Antibody Chip Technology

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Background: Qiguiyin decoction (QGYD) is a traditional Chinese medicine (TCM) and its combined application with levofloxacin (LVFX) has been confirmed effective in the clinical treatment of multidrug-resistant Pseudomonas aeruginosa (MDR PA) infection. This study investigated the therapeutic effect and possible mechanism of QGYD in sensitizing LVFX against MDR PA infection.

Materials and Methods: Pulmonary infections were induced in rats by MDR PA. The changes in pharmacokineticspharmacodynamics (PK-PD) parameters of LVFX after combined with QGYD were investigated in MDR PA-induced rats. Subsequently, the correlation between PK and PD was analyzed and PK-PD models were established to elucidate the relationship between QGYD-induced alterations in LVFX metabolism and its sensitization to LVFX. Antibody chip technology was used to detect the levels of inflammatory factors, suggesting the relationship between the beneficial effect of immune regulation and the sensitization of QGYD

Results: QGYD significantly enhanced the therapeutic efficacy of LVFX against MDR PA infection. The combination of QGYD changed the PK parameters of LVFX such as T_{max}, t_{1/2}, MRT, V_d/F, CL/F and PD parameters such as MIC, AUC_{0-24h}/MIC. Predicted results from PK-PD models demonstrated that the antibacterial effect of LVFX was significantly enhanced with the combination of QGYD, consistent with experimental findings. Antibody chip results revealed that the combination of QGYD made IL-1 β, IL-6, TNF- α , IL-10, and MCP-1 levels more akin to those of the blank group.

Conclusion: These findings indicated that QGYD could change the PK-PD behaviors of LVFX and help the body restore immune balance faster. This implied that a potential drug interaction might occur between QGYD and LVFX, leading to improved clinical efficacy when combined.

Keywords: multidrug-resistant Pseudomonas aeruginosa infection, combination therapy, PK/PD model, Qiguiyin decoction, levofloxacin

Introduction

The overuse of levofloxacin (LVFX) has resulted in a shift in the susceptibility of *Pseudomonas aeruginosa* (PA) to LVFX, contributing to the rise of multidrug-resistant (MDR) PA strains, posing a significant threat to patient health.¹⁻⁴ Guo⁵ analyzed drug sensitivity data from pulmonary tuberculosis patients with PA infection between 2016 to 2020, revealing a notable increase in PA resistance to LVFX in 2020. This underscores the challenge of achieving optimal efficacy with LVFX monotherapy, prompting the adoption of combination therapy strategies for the treatment of MDR PA infections.⁶

Traditional Chinese medicine (TCM) has played an important role in preventing and treating infected diseases since ancient times.⁷ In the clinical treatment of MDR PA infection, numerous Chinese medicinal treatments, along with their

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concurrent use with antibacterial drugs, have demonstrated efficacy.^{8,9} Qiguiyin decoction (QGYD) has a good anti-infection effect combined with antibacterial drugs in clinical practice,¹⁰ which consists of five Chinese medicines including *Astragalus membranaceus* (Fisch.) Bge, *Lonicera japonica* Thunb., *Angelica sinensis* (Oliv.) Diels, *Artemisia annua* L., and *Polygonum cuspidatum* Sieb. et Zucc. Our previous studies have shown that QGYD could delay or reverse PA resistance to antibacterial drugs, regulate inflammation and immune dysregulation resulting from PA infection, and enhance therapeutic outcomes when combined with LVFX.^{11,12} Nonetheless, the impact of QGYD on the pharmacokinetic-pharmacodynamic (PK-PD) profile of LVFX in PA-infected rats, as well as the mechanism underlying QGYD's sensitization of LVFX, remains obscure.

Following PA infection, the body often experiences a disruption in inflammatory factors. MDR PA infections are more prone to inducing or exacerbating inflammatory immune imbalances compared to typical bacterial infections.¹³ Some studies have shown that TCM can improve the body's immunity.¹⁴ Consequently, we hypothesize that the co-administration of QGYD influences the PK-PD parameters of LVFX and immune alterations in vivo, thereby facilitating the sensitization of LVFX against MDR PA.

We undertook three main tasks in this study. Firstly, we investigated the PK parameters of LVFX administered alone or in conjunction with QGYD in pneumonia rat models induced by MDR PA, employing ultra-high-performance liquid chromatography-triple quadrupole mass spectrometry (UPLC-TQ/MS). Subsequently, the bacteriostatic activity of drug-containing serum in pneumonia rats was examined in vitro. Furthermore, PK/PD models were established to assess the efficacy of LVFX as monotherapy or in combination with QGYD against MDR PA, offering insights into the integration of Chinese and Western medicine in clinical practice. Additionally, we employed antibody chips to analyze alterations in serum inflammatory factors in pneumonia rats following treatment with LVFX alone or in combination with QGYD, aiming to elucidate the mechanism by which QGYD sensitizes LVFX from an immunological perspective and present a novel clinical option for enhancing treatment against MDR PA resistance.

Materials and Methods

Materials

Drugs and Chemicals

Levofloxacin (Batch No: Y02N7C23971, purity > 99%), and ciprofloxacin hydrochloride (Batch No: SM0325YG13, purity > 98%) were purchased from Shanghai Yuan-ye Biological Technology Co., Ltd. (Shanghai, China). Levofloxacin tablets (Batch No: BY008A1) were purchased from Daiichi Sankyo Pharmaceutical Co., Ltd. (Beijing, China). QGYD is composed of five traditional Chinese medicines including *Astragalus membranaceus* (Fisch.) Bunge. (Huangqi), *Angelica sinensis* (Oliv.) Diels. (Danggui), *Lonicera japonica* Thunb. (Jinyinhua), *Polygonum cuspidatum* Sieb. et Zucc. (Huzhang), and *Artemisia annua* L. (Qinghao), which were purchased from Beijing Sanhe Pharmaceutical Co., Ltd. (Beijing, China) and authenticated by Professor Yaojun Yang (School of Chinese Materia Medica, Beijing University of Chinese Medicine). LC-MS grade methanol, acetonitrile, and formic acid were purchased from Thermo Fisher Scientific (New Jersey, USA). Ultra-pure water was purified by a Milli-Q water system (MA, USA). Cytokine Antibody Chip Kit was from RayBiotech, Inc. (Atlanta, USA).

Animals and Bacterial Suspension

Forty healthy male Wistar rats, (4–5 months old, SPF grade), weighing around 220–260 g, were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd. (Beijing, China), license number: SCXK (jing) 2021–0011. All rats were adaptively fed with a standard diet and water for 3 days before the experiments. The environmental conditions were maintained at a temperature of 23 ± 2 °C, with a relative humidity of $60 \pm 5\%$, and a 12-hour light-dark cycle. This study adhered to the Guide for the Care and Use of Laboratory Animals (National Institute) and received approval from the Animal Experimental Medical Ethics Review Committee of Beijing University of Chinese Medicine (No: BUCM-4-2022030401-1033).

The MDR PA strain (No.1912109) was sourced from the Department of Clinical Laboratory of Dongzhimen Hospital, affiliated with Beijing University of Chinese Medicine. The strain was cultured on Mueller-Hinton Agar (MHA) medium at 37°C for 16–20 hours. Subsequently, the MDR PA suspension was adjusted to a concentration of 6×10^8 CFU/mL using a turbidimetric method with sterile saline, with an absorbance value at 625 nm (OD₆₂₅) ranging at 0.3–0.4.

Preparation of QGYD Extract

The extraction method for QGYD had been previously established and its stability had been confirmed through fingerprint analysis. Thus the extraction method was directly applied in this experiment. QGYD is comprised of five traditional Chinese medicines, including Huangqi, Danggui, Jinyinhua, Huzhang, and Qinghao, with the respective compounds mixed in proportions of 12:2:3:2:3. The preparation process involved extracting QGYD three times using deionized water at ratios of 12, 10, and 10 folds (v/w), each extraction lasting 1 hour. The combined solutions were then concentrated to a concentration of 1.0 g/mL, followed by the addition of ethanol to achieve a concentration of 60%. The mixture was left to stand for 24 hours at 4 °C. The supernatant was collected after suction filtration, dried under vacuum at 60 °C, resulting in the QGYD extract (yield: 26.13%).

Animal Experimental Design

Forty rats were randomly assigned to four groups: control group, model group, LVFX group, and QGYD-LVFX group. The rat infection model was established following a previously report.¹² The MDR PA-induced pneumonia rat model was established by endotracheal intubation. Each rat received 100 μ L of MDR PA suspension at a concentration of 6×10⁸ CFU/mL, while the control group received an equal volume of sterile saline. Following modeling, the control group was housed separately from the MDR PA-infected groups. The rat dosage was scaled to be equivalent to 6.25 times that for a 60 kg adult. An hour after infection, rats in the LVFX group received oral administration of LVFX (0.078 g/kg/d), while rats in the QGYD-LVFX group received co-administration of QGYD (11.5 g/kg/d) and LVFX (0.078 g/kg/d). The control and model groups received equivalent amounts of distilled water.

The general condition of the rats was monitored daily after modeling. The rats fasted for 12 hours before the final administration but drank water freely. After five days of continuous gavage, blood samples were collected from the orbital venous plexus at 0 h, 0.25 h, 0.5 h, 1 h, 1.5 h, 2h, 4 h, 8 h, 12 h, and 24 h after the last administration. The collected blood samples were placed in EP tubes containing a coagulant and centrifuged at 3500 rpm at 4 °C for 15 min following 1 h at room temperature. The supernatant was collected and stored at -80 °C for later use.

Determination of Levofloxacin Concentrations in Rat Serum Samples

A validated ultra-performance liquid chromatography-tandem mass spectrometry (UPLC-MS/MS) method was employed to quantify LVFX concentrations from serum samples. Frozen plasma samples were thawed at 4 °C before use. The 5 μ L of internal standard (IS) solution (20 μ g/mL CPFX) was added to a 20 μ L plasma sample, followed by the addition of 55 μ L methanol to precipitate proteins. The mixture was vortexed for 5 min and then centrifuged at 14,000 rpm at 4 °C for 15 min. The supernatant was collected and dried under a nitrogen stream. Subsequently, the residues were redissolved in 1 mL of initial proportion of the mobile phase, vortexed for 30s, and then, centrifuged at 14,000 rpm for 15 min at 4 °C. The final supernatant was injected into UPLC-TQ/MS for analysis.

The chromatographic separation was achieved on ultra-high performance liquid chromatography (UPLC) with an ACQUITY UPLC BEH C_{18} column (2.1×100 mm, 1.7 µm; Waters Corp., USA) at a column temperature of 40 °C. The temperature of the sample chamber was 10 °C. The mobile phase was composed of 0.1% formic acid aqueous solution (A) and acetonitrile (B). The gradient was set as follows: 0–0.1 min, 90–85% A; 0.1–3.0 min, 85–20% A; 3.0–3.5 min, 20–90% A. The flow rate was 0.30 mL/min, and the injection volume was 1 µL.

For mass spectrum acquisition, a triple quadrupole tandem mass spectrometry (TQ/MS) (Thermo Fisher, USA) was used in the positive ion (ESI+) mode by multiple reaction monitoring (MRM) of the transition of *m/z* 362.2395-*m/z* 261.1767 for LVFX and *m/z* 332.1651-*m/z* 245.1919 for CPFX (internal standard). The optimal MS parameters were as follows: the capillary voltage was set at 2.5 kV; desolvation gas flow rate was set at 650 L/h; source and desolvation temperatures maintained at 150 °C and 350 °C, respectively; the optimized cone voltage and collision energy were set at 44 V and 22 eV for LVFX, and 42 V and 20 eV for CPFX, respectively. Data acquisition and processing were performed using MassLynx 4.2 software (Waters Corp., USA).

The lower limit of quantification for levofloxacin was 2 ng/mL, and the standard curve exhibited linearity within the range of 2–400 ng/mL, with a coefficient of determination (R^2) of 0.9997.

Pharmacokinetic Analysis

PK parameters were determined by DAS 2.0 software. Based on the actual serum concentration-time data of rats in each group, PK parameters were calculated using the non-compartmental model, such as time to peak time (T_{max}), peak concentration (C_{max}), area under the concentration-time curve (AUC), mean residence time (MRT), half life ($t_{1/2}$), volume of distribution adjusted for bioavailability (V_d /F) and clearance adjusted for bioavailability (CL/F). The akaike information criterion (AIC) and R² were used to determine the optimal atrioventricular model.

Susceptibility Testing

Determination of Minimum Inhibitory Concentration of Levofloxacin and Qiguiyin Decoction

The minimum inhibitory concentration (MIC) of LVEX and QGYD against MDR PA (No.1912109) were determined using the broth microdilution method according to CLSI guidelines. Twofold serial dilution of LVFX (1024 ug/mL) or QGYD (2 g/mL) in Mueller-Hinton Broth (MHB) was prepared in 24 -well microtitre plate and the volume in each well was 1 mL. Besides, the positive and negative control wells were set. 10 μ L of the prepared MDR PA bacterial solution was added to each well (except the negative control well) with a concentration of 1.5×10⁸ CFU/mL, and the 24-well plate was placed in the incubator at 37 °C for 24 h. Record the value of MIC.

Determination of Fractional Inhibitory Concentration Index of Levofloxacin and Qiguiyin Decoction Using Broth Microdilution Method

According to the checkerboard method design, the combined bacteriostatic effect of LVFX and QGYD with different concentrations (1/16MIC-2MIC) on the MDR PA strain was determined by the broth microdilution method. 50 uL of twofold serial dilution of LVFX (64 ug/mL) and QGYD (2 g/mL) in MHB was prepared in a 96-well microtitre plate. There are also positive and negative control wells. 100 μ L of this suspension with a concentration of 3×10⁶ CFU/mL was inoculated into the broths except for the negative control wells. The final volume in each well was 200 uL After being incubated at 37 °C for 24 h in the incubator, the fractional inhibitory concentration index (FICI) was calculated using the following formula:

FICI = [MIC(LVFX in combination with QGYD)/MIC(LVFX alone)] + [MIC(QGYD in combination with LVFX)/MIC(QGYD alone)]

Interpretation criteria: $FICI \le 0.5$ denotes "synergistic"; $0.5 < FICI \le 1$ denotes "additivity"; $1 < FICI \le 4$ denotes "no interaction"; FICI > 4 denotes "antagonism".

Determination of Antibacterial Activity in the Serum of Rats Administered with Levofloxacin Alone and in Combination with Qiguiyin Decoction

The antibacterial activity in rat serum of all groups was determined by adding 50 uL of serum at various time points to the 96well plate containing 130 uL of MHB. Then 20 μ L of bacterial solution (1.5 × 10⁷ CFU/mL) was added. In addition, positive and negative control holes containing blank serum were set for parallel control. After being incubated in a 37 °C constant temperature incubator for 18–24 hours, the result was detected at 600 nm by the microplate reader, and the antibacterial rate of drug-containing serum at different time points was calculated according to the OD value.

Fitting and Analyzing of PK-PD Model

The dynamic studies between the semi-body parameter AUC_{24h}/MIC and the bacteriostatic effect were modeled and analyzed using E_{max} Model in Phoenix WinNonlin software. The standard error (Stderr) and coefficient of variation (CV %) were used to evaluate the accuracy of fit of the model and the AIC value was used to evaluate the model's goodness of fit. The optimal model was comprehensively compared and determined.

Determination of Serum Inflammatory Cytokines

The QAM-INF-1 Kit was used to simultaneously detect 5 common inflammatory cytokines including IL-1 β , IL-6, IL-10, MCP-1, and TNF- α in rat serum in each group. The operation steps are as follows: 100 μ L of sample diluent was added and incubated in a shaking table at room temperature for 1 h to close the chip and then discarded. And 100 μ L of sample solution or standard solution was added and incubated in a shaking table at 4 °C. The chips were washed by washing solutions I and II after being

diluted with deionized water. After adding 80 mL test antibodies, it was incubated in a 37 °C shaker for 2 hours. After being washed as above, 80 uL of Cy3-Streptavidin was added and then incubated in a 37 °C shaker for 1 hour. After being washed and dried at room temperature, the chips were detected at 532 nm with Innoscan 300 scanner. QAM-INF-1 software was used to analyze the changes of the above inflammatory cytokines and search for differential inflammatory factors in different groups of rats.

Statistical Analysis

The data were expressed as mean \pm standard deviation (mean \pm SD) in figures and tables. The differences of PK parameters and levels of inflammatory factors were analyzed by one-way ANOVA with SPSS 21.0 software. P < 0.05 was considered to be statistically significant. GraphPad Prism 8.0 software was used for making charts.

Results

Susceptibility Testing

The MICs of LVEX and QGYD against MDR PA were determined using the broth microdilution method. The results showed that the combination of QGYD reduced the MIC of LVFX from 8 μ g/mL to 4 μ g/mL, and the MIC of QGYD decreased from 250 mg/mL to 125 mg/mL compared to QGYD alone. Compared to the single treatment group, the MIC of LVFX and OGYD decreased in the combined treatment group, indicating an increased bacteriostasis rate following the combination of these two drugs. Furthermore, the FICI of QGYD and LVFX in combination against MDR PA-induced pneumonia rats was calculated as 1, indicating an additive effect against the MDR PA strain.

Effects of Qiguiyin Decoction on Pharmacokinetics of Levofloxacin in Serum of Pneumonia Rats

The PK parameters of LVFX group and LVFX-QGYD group in the MDR PA-infected rat are shown in Table 1. The PK of LVFX in pneumonia rats followed a primary absorption rate and a two-compartment model. The model-fitting results are shown in Figure 1. The results basically corresponded with those reported in the previous literature,^{15,16} indicating the determination of LVFX concentration in rat serum was accurate and reliable.

Parameter	Unit	LVFX Group	LVFX-QGYD Group	
AUC ₀₋₂₄	ug/mL*h	36.48±5.00	32.00±0.62	
AUC _{0-∞}	ug/mL*h	47.50±8.52	34.03±2.43*	
MRT ₀₋₂₄	h	6.18±0.35	7.59±0.17**	
MRT _{0-∞}	h	17.14±7.16	11.86±1.00	
t _{1/2}	h	15.60±8.27	5.65±1.41*	
T _{max}	h	0.46±0.10	I.42±0.38**	
V _d /F	mL/kg	35.88±14.61	18.53±3.12*	
CL/F	mL/h/kg	1.70±0.38	2.31±0.15**	
C _{max}	ug/mL	10.32±0.71	4.45±0.75**	

Table	L	The	PΚ	Parameters	of LVFX	Group	and LVF	X-QGYD
Group	in	the	MD	R PA-Infect	ed Rats.	(n=6, M	lean±SD)	

Notes: ∞ , the complete elimination of the drug from the body; *P < 0.05, versus the LVFX group; **P < 0.01, versus the LVFX group.

Abbreviations: AUC₀₋₂₄, area under the concentration-time curve from 0 to 24 hours; AUC_{0-∞}, area under the concentration-time curve from 0 to[∞]; MRT₀₋₂₄, mean residence time from 0 to 24 hours; MRT_{0-∞}, mean residence timefrom 0 to[∞]; t_{1/2}, biological half life; T_{max}, peak time; V_a/F, volume of distribution adjusted for bioavailability; CL/F, clearance adjusted for bioavailability; C_{max}, peak concentration.



Figure I The model-fitting results of LVFX group (A) and LVFX-QGYD group (B). (AIC, akaike information criterion; R², determination coefficient).

Plasma concentration-time profiles of the LVFX group and LVFX-QGYD group in MDR PA-infected rats are shown in Figure 2. The results showed that the combination of QGYD resulted in prolonged MRT and T_{max} of LVFX in pneumonia rats. Moreover, the combination of QGYD reduced V_d/F and $t_{1/2}$, and increased CL/F in pneumonia rats. These results suggested that the combination of QGYD might affect the absorption, distribution, metabolism, and excretion process of LVFX in vivo by changing its PK parameters.

Effects of Qiguiyin Decoction on Pharmacodynamics of Levofloxacin in Serum of Pneumonia Rats

The average bacteriostatic effect-time curves of the LVFX group and LVFX-QGYD group are shown in Figure 3, revealing a higher initial bacteriostatic rate and longer T_{max} following the combination of QGYD. Furthermore, the serum bacteriostatic rate of the combination group remained consistently higher than that of LVFX alone 1 h after administration, indicating a significant enhancement in the inhibition of LVFX on MDR PA-induced rats by the combination with QGYD.

Establishment of PK/PD Modelings

The semi-body parameter AUC_{24h}/MIC was selected as PK/PD parameter to describe the antibacterial effect of LVFX alone or in combination with QGYD on MDR PA-induced rats. Compared with the LVFX single group, the combination of QGYD



Figure 2 Plasma concentration versus time profiles of LVFX group and LVFX-QGYD group.



Figure 3 The average bacteriostatic effect versus time curves of the LVFX group and LVFX-QGYD group.

significantly increased the AUC_{0-24h}/MIC of LVFX from 4.56±0.63 to 7.99±0.16. The PK-PD parameters of LVFX alone and in combination with QGYD are presented in Table 2 and Table 3. It was concluded that the PK/PD model of LVFX singly was $E=(81.25 \times C^{0.93})/(0.75^{0.93}+C^{0.93})$, while the PK/PD model of LVFX-QGYD was $E= 38.30+(46.80 \times C^{2.06})/(2.94^{2.06}+C^{2.06})$. Therefore, the antibacterial effect of LVFX singly was (68.34±1.36) %, and LVFX-QGYD was (79.81±0.19) % (P < 0.01).

Effects of Qiguiyin Decoction Combined with Levofloxacin on Inflammatory Cytokines in Serum of Pneumonia Rats

It is well established that inflammatory cytokines play a pivotal role in promoting the occurrence and development of inflammatory immunity in the body. Dynamic changes in inflammatory factors during the modeling and administration process were analyzed using antibody chip. As shown in Figure 4, our study revealed that the levels of 5 inflammatory

Parameter	Estimate	Stderr	CV %	2.5% CI	97.5% CI
EC ₅₀	0.75	0.23	30.42	0.19	1.31
N	0.93	0.25	27.09	0.31	1.55
E _{max}	81.25	8.94	11.01	59.37	103.13

Table 2 The PK-PD Parameters of LVFX Group

 $\label{eq:solution} \begin{array}{l} \textbf{Abbreviations:} \ EC_{50}, \ integration \ concentration \ value at 50\% \ of the maximum efficacy; \\ N=Gamma, \ the shape \ factor \ of \ an \ S-shaped \ curve; \ E_{max}, \ maximum \ effect \ of \ the \ drug; \\ Stderr, \ standard \ deviation; \ CV\%, \ coefficient \ of \ variation; \ CI, \ confidence \ interval. \end{array}$

Table 3	The PK-PD	Parameters	of LVFX-QGYD	Group
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Parameter	Estimate	Stderr	CV%	2.5% CI	97.5% CI
EC ₅₀	2.94	0.25	8.55	2.3	3.59
N	2.06	0.46	22.22	0.88	3.24
Eo	38.3	2.41	6.3	32.1	44.51
E _{max}	46.8	6.55	13.99	29.97	63.64

Abbreviations: EC₅₀, integration concentration value at 50% of the maximum efficacy; N=Gamma, the shape factor of an S-shaped curve; E₀, basic pharmacological effect value; E_{max}, maximum effect of the drug; Stderr, standard deviation; CV%, coefficient of variation; Cl, confidence interval.



Figure 4 Expressions of serum inflammatory cytokines in each group. (Bars represent mean \pm SD (N = 4, per group). *P < 0.05, versus the control group; **P < 0.01, versus the control group; $^{\#P}$ < 0.05, versus the model group; $^{\#P}$ < 0.01, versus the model group; $^{\Delta P}$ < 0.01, versus the LVFX group).

factors in the model group were elevated compared to those in the control group. These findings suggested that MDR PA infection could significantly enhance the expressions of IL-1 β , IL-6, TNF- α , IL-10, and MCP-1, leading to a disruption in the body's immune homeostasis. Following intervention with LVFX, a significant decrease in the expressions of IL-1 β and TNF- α was observed, and the expressions of IL-6, IL-10, and MCP-1 showed a slight reduction compared to the model group. Notably, the combined treatment of LVFX with QGYD resulted in expressions of IL-1 β , IL-6, TNF- α , IL-10, and MCP-1 that were closer to those in the control group than LVFX administered alone. Consequently, QGYD-LVFX treatment demonstrated a significant regulatory effect on the expressions of inflammatory factors in MDR PA-induced rats, facilitating the restoration of immune balance.

Discussion

Traditionally, PK-PD studies are conducted using healthy laboratory animals.¹⁷ However, disparities exist in the PK and PD parameters of antibacterial drugs between normal and infected animal models.¹⁸ The PK-PD parameters of LVFX in combination with QGYD in infected models hold greater significance and clinical relevance compared to those in normal models. Therefore, we established a pneumonia model induced by MDR PA in rats and further investigated the PK-PD of LVFX alone or in combination with QGYD, providing a robust foundation for the combined use of Chinese and Western medicines in the treatment of MDR PA infections.

PK experiments revealed that the combination of QGYD slowed down the release of LVFX in rats. And the distribution of LVFX in various tissues was reduced, its access into the vascular ventricle was increased, and the clearance rate was enhanced. Moreover, the AUC_{0-24h}/MIC ratio of LVFX significantly increased after combining with QGYD, suggesting a potential enhancement in the pharmacological activity of LVFX. Model-predicted results revealed that the serum bacteriostatic effect was significantly augmented in the combined group compared to the LVFX group, indicating the combination of QGYD could enhance the efficacy of LVFX against MDR PA infection in rats These findings were also consistent with previous researches.^{10,19}

Furthermore, we used the antibody chip to investigate the mechanism of QGYD restoring LVFX sensitivity to MDR PA. Pneumonia infection caused by PA involves multiple inflammatory pathways. The results demonstrated a significant increase in the serum inflammatory cytokine IL-1 β , IL-6, TNF- α , IL-10, and MCP-1 levels in the model group, consistent with previous studies.^{12,20} Compared to LVFX alone, the QGYD-LVFX combination treatment normalized some proinflammatory factors and anti-inflammatory factors towards levels observed in the control group, suggesting that QGYD-LVFX therapy could inhibit the further expansion of inflammatory response and help the body recover to normal immune balance more quickly. TCM has also been proven to promote inflammation recovery.²¹ Consequently, the combination of QGYD and LVFX not only improved antibacterial efficacy by influencing pharmacokinetics but also enhances overall immunity. Therefore, our study demonstrated that the combination of QGYD significantly enhanced the antibacterial effect of LVFX in pneumonia rats, potentially through herb-drug interactions that improved anti-infective efficacy.

Conclusion

In conclusion, the combination of QGYD changed the PK/PD parameters of LVFX in pneumonia rats, enhancing its activity against MDR PA. PK/PD model fitting further confirmed the superior antibacterial effects of the LVFX-QGYD combination over LVFX alone, providing additional evidence for the efficacy of QGYD in combination with LVFX against MDR PA. From the perspective of immunity, compared to LVFX alone, the combination of QGYD could significantly regulate the expressions of IL-1 β , TNF- α , IL-6, and IL-10 caused by MDR PA infection. This modulation promoted the restoration of immune balance in pneumonia rats and enhanced the sensitizing effect of LVFX.

Abbreviations

AIC, Akaike information criterion; AUC, area under the concentration-time curve; C_{max} , peak concentration; CL/F, clearance adjusted for bioavailability; CV%, coefficient of variation; FICI, fractional inhibitory concentration index; IS, internal standard; LVFX, levofloxacin; MDR PA, multidrug-resistant Pseudomonas aeruginosa; MHB, Mueller-Hinton Broth; MIC, minimum inhibitory concentration; MRT, mean residence time; PK-PD, pharmacokinetics-pharmacodynamics; QGYD, Qiguiyin decoction; Stderr, standard error; SPF, specific pathogen free; $t_{1/2}$, half life; T_{max} , peak time; TCM, traditional Chinese medicine; UPLC-MS/MS, ultra-performance liquid chromatography-tandem mass spectrometry; UPLC-TQ/MS, ultra-high-performance liquid chromatography-triple quadrupole mass spectrometry; V_d/F , volume of distribution adjusted for bioavailability.

Data Sharing Statement

The data used to support the findings of this study are available from the corresponding author upon request.

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Disclosure

The authors report no conflicts of interest in this work.

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