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IncRNA RPSAP52 induced the development of tongue squamous cell carcinomas via miR-423-5p/MYBL2

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Abstract

Growing IncRNAs have been noted to involve in the initiation and development of several tumours including tongue squamous cell carcinomas (TSCCs). However, the biological role and mechanism of IncRNA RPSAP52 were not well-explained. We indicated that RPSAP52 was higher in TSCC samples compared with that in control samples. The higher expression of RPSAP52 was positively correlated with higher T stage and TNM stage. Ectopic expression of RPSAP52 induced TSCC cell growth and cycle and induced cytokine secretion including IFN- γ , IL-1 β and IL-6, IL-8, IL-10 and TGF- β . We found that the overexpression of RPSAP52 suppressed miR-423-5p expression in SCC-4 cell. miR-423-5p was lower in TSCC samples compared with that in control samples, and miR-423-5p level was negatively correlated with higher T stage and TNM stage. Pearson's correlation indicated that miR-423-5p was negatively associated with that of RPSAP52 in TSCC tissues. Furthermore, MYBL2 was one direct gene of miR-423-5p and elevated expression of miR-423-5p suppressed MYBL2 expression and ectopic expression of RPSAP52 increased MYBL2 expression in SCC-4 cell. Finally, we illustrated that RPSAP52 overexpression promoted TSCC cell growth and cycle and induced cytokine secretion including IFN- γ , IL-1 β and IL-6, IL-8, IL-10 and TGF- β via modulating MYBL2. These data provided new insight into RPSAP52, which may be one potential treatment target for TSCC.

KEYWORDS

miR-423-5p, MYBL2, RPSAP52, tongue squamous cell carcinomas

1 | INTRODUCTION

Oral squamous cell carcinomas (OSCCs) are understudied and undertreated diseases.¹⁻⁵ SCC of tongue (TSCC) is the most frequent carcinoma of OSCC, with aspects of rapid metastatic spread and local invasion.⁶⁻⁸ The incidence of these diseases is really grown in middle and young age populations.⁹⁻¹¹ Although treatment attempts such as surgery, radiotherapy and chemotherapy have been tried, the prognosis of TSCC remains unsatisfactory.¹²⁻¹⁵ It suggested that it is one of the major health issues and it needs the understanding of molecular mechanisms of progression or carcinogenesis of this disease. It will be helpful for improving prevention, therapy and diagnosis of TSCC.

IncRNAs were loosely regarded as new RNA molecules, which were more than 200 nts and lack protein-coding ability. $^{16-20}$ Deregulated

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expression of IncRNAs is associated with several diseases including respiratory, neurodegenerative and cardiovascular diseases, and scoliosis.²¹⁻²⁴ Numerous references illuminated that IncRNAs participate in critical cell processes including cell metabolism, invasion, apoptosis, proliferation and differentiation.²⁵⁻²⁸ Increasing studies also noted that IncRNAs are involved in the development and progression of TSCC.^{29,30} Recently, a new IncRNA RPSAP52 has been identified and revealed to participate in several tumours such as glioblastoma, pituitary tumours and pancreatic cancer.³¹⁻³⁴ However, the biological role and mechanism of IncRNA RPSAP52 were not well-explained.

2 | MATERIALS AND METHODS

2.1 | Patient's samples and cell culture and transfection

TSCC and their morphological control specimens were collected from 30 TSCC cases undergoing surgery at Jinan Stomatological Hospital. Specimens were immediately frozen in liquid nitrogen until RNA was used. All protocols were agreed with the Ethics Committee of Jinan Stomatological Hospital, and all patients signed the informed consent. TSCC lines (UM1, SCC-4, SCC-1 and Cal27) and NHOK were obtained from ATCC. Cells were cultured in the DMEM supplemented with FBS, penicillin and streptomycin. pcDNA-RPSAP52 and MYBL2 siRNA vector, miRNA mimic and control plasmids were synthesized by GenePharma and then transfected into cells using Lipofectamine 3000 (Invitrogen). Total RNA from TSCC tissues and cells was extracted using TRIzol Kit (Invitrogen) following the instructions. This primer was designed as follows: RPSAP52: F, 5-'GAG CAA ACA CAT CGG AGACA-3', and R, 5-'AAT TGG ATT CCC ACTG CAAG-3'; GAPDH: F, 5-'GACCTG ACC TGC CGT CTA G-3', and R, 5-'AGG AGT GGG TGT CGC TGT-3'; U6: F, 5'-CTCGCT TCGGCA GCACA-3', and R, 5'-AACGCT TCA CGAATT TGCGT-3'; MYBL2: F, 5-'GAGGG ATAGC AAGTG CAAGGT-3', and R, 5-'TTCCA GTCCT GCTGTC CAAA-3'; and miR-423-5p: F, 5-'TGAGG GGCAG AGCGA GACTTT-3', and R, 5-'GTGCA GGGTCC GAGGTGG GCAGAG CGAGACTTT-3'. qRT-PCR analysis was conducted with SYBR Premix I (Takara) on the ABI 7900HT qRT-PCR System (Bio-Rad). The relative value of the miRNA, IncRNA and mRNA expression was analysed using $2^{-\Delta\DeltaCt}$ method. U6 and GAPDH acted as control genes for miRNA and IncRNA and mRNA, respectively.

2.3 | Cell proliferation and ELISA

Different groups of TSCC cells were plated in 96-well plates. Cell growth was determined by CCK-8 method, and 10 μ L CCK-8 reagent was added into each well until visual colour was changed. The absorbance was determined by the microplate reader at 450 nM. Cytokine level was detected using ELISA reagents (R&D Systems). The absorbance at 450 nm was recorded by the microplate reader.



FIGURE 1 RPSAP52 was overexpressed in TSCC samples. (A) RTqPCR data suggested that RPSAP52 was higher in TSCC samples compared with that in control samples. (B) RPSAP52 was overexpressed in 25 TSCC cases (25/30, 83.3%) compared with control no-tumour samples. (C) The higher expression of RPSAP52 was positively correlated with higher T stage. (D) The higher expression of RPSAP52 was positively correlated with TNM stage. GAPDH was used as internal control. *P < .05 and **P < .01 WILEY

2.4 | Luciferase reporter gene assay

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Cells were cultured in 96-well dish. The mutant sequence of MYBL2 (pmirGLO-MYBL2-Mut) and its wild sequence (pmirGLO-MYBL2-wt) were cloned into luciferase reporter and then named as pmirGLO-MYBL2-Mut and pmirGLO-MYBL2-WT, respectively. Cells were transfected with mimic and scramble and pmirGLO-MYBL2-Mut and pmirGLO-MYBL2-WT by Lipofectamine 3000 (Invitrogen). Fortyeight hours later, cells were harvested and luciferase value was detected by dual-luciferase analysis (Promega) following the protocol.

2.5 | Statistical analysis

The statistical assay was conducted by SPSS 21.0 software, and results were presented as mean \pm SD. The difference between 2 groups was evaluated by *t* test, and *P* value < .05 was considered to be significant. The correlation between miR-423-5p and RPSAP52 was analysed by Pearson's correlation.

3 | RESULTS

3.1 | RPSAP52 was overexpressed in TSCC samples

RT-qPCR data suggested that RPSAP52 was higher in TSCC samples compared with that in control samples (Figure 1A). RPSAP52 was overexpressed in 25 TSCC cases (25/30, 83.3%) compared with control no-tumour samples (Figure 1B). The higher expression of RPSAP52 was positively correlated with higher T stage (Figure 1C) and TNM stage (Figure 1D).

3.2 | miR-423-5p was down-regulated in TSCC specimens

RT-qPCR assay showed that miR-423-5p was lower in TSCC samples compared with that in control samples (Figure 2A). miR-423-5p was down-regulated in 24 TSCC cases (24/30, 80%) compared with control no-tumour samples (Figure 2B). The miR-423-5p level was





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FIGURE 3 Ectopic expression of RPSAP52 induced TSCC cell growth. (A) RPSAP52 level was overexpressed in four TSCC lines (UM1, SCC-4, SCC-1 and Cal-27) compared with NHOK cell. (B) The expression of RPSAP52 was detected by qRT-PCR. (C) Elevated expression of RPSAP52 increased cells at S stage and decreased cells at G0-G1 stage. (D) Overexpression of RPSAP52 induced ki-67 expression in SCC-4 cell. (E) The expression of CDK2 was measured by gRT-PCR analysis. (F) Ectopic RPSAP52 expression increased cell growth in SCC-4 cell. *P < .05, **P < .01 and ***P < .001



FIGURE 4 Elevated expression of RPSAP52 induced cytokine secretion in TSCC cell. (A) The expression of IFN-y was determined by ELISA. (B) The expression of IL-1 β was detected using ELISA. (C) The expression of IL-6 was detected using ELISA. (D) The expression of IL-8 was determined by ELISA. (E) The expression of IL-10 was detected using ELISA. (F) The expression of TGF- β was detected using ELISA. ***P < .001

negatively correlated with higher T stage (Figure 2C) and TNM stage (Figure 2D). Pearson's correlation indicated that miR-423-5p was negatively associated with that of RPSAP52 in TSCC tissues (Figure 2E).

3.3 | Ectopic expression of RPSAP52 induced TSCC cell growth

RPSAP52 level was overexpressed in four TSCC lines (UM1, SCC-4, SCC-1 and Cal-27) compared with NHOK cell (Figure 3A). RPSAP52 level was strikingly up-regulated in SCC-4 after transfected with pcDNA-RPSAP52 (Figure 3B). Elevated expression of RPSAP52 increased cells at S stage and decreased cells at G0-G1 stage (Figure 3C). Overexpression of RPSAP52 induced ki-67 (Figure 3D) and CDK2 (Figure 3E) expression in SCC-4 cell. Ectopic RPSAP52 expression increased cell growth in SCC-4 cell (Figure 3F).

3.4 | Elevated expression of RPSAP52 induced cytokine secretion in TSCC cell

The cytokine concentrations of IFN- γ (Figure 4A), IL-1 β (Figure 4B) and IL-6 (Figure 4C) were up-regulated in SCC-4 cell after treated with pcDNA-RPSAP52. Ectopic expression of RPSAP52 promoted cytokine concentration excretion including IL-8 (Figure 4D), IL-10 (Figure 4E) and TGF- β (Figure 4F) expression.

3.5 | MYBL2 was one direct gene of miR-423-5p

One bioinformatic TargetScan assay (http://www.targetscan. org/vert_72/) noted that MYBL2 contained target sites of miR-423-5p as indicated in Figure 5A. The level of miR-423-5p was strikingly overexpressed in SCC-4 cell after transfection with miR-423-5p mimic (Figure 5B). Luciferase reporter assay suggested that

A Position 116-122 of MYBL2 3' UTR 5'-AGCCUUCUGCCACCAGCCCCUCC-3' ||||||| hsa-miR-423-5p 3'-UUUCAGAGCGAGAGACGGGGAGU-5' MYBL2 mut 3' UTR 5'-AGCCUUCUGCCACCACGGGGAGC-3'



FIGURE 5 MYBL2 was one direct gene of miR-423-5p. (A) Bioinformatic TargetScan assay noted that MYBL2 contained target sites of miR-423-5p. (B) The level of miR-423-5p was determined by qRT-PCR. (C) Luciferase reporter assay suggested the ectopic expression of miR-423-5p attenuated luciferase value of MYBL2-WT reporter vector, but not that of MYBL2-mut reporter vector in SCC-4 cell. (D) Elevated expression of miR-423-5p suppressed MYBL2 expression in SCC-4 cell. (E) Overexpression of RPSAP52 suppressed miR-423-5p expression in SCC-4 cell. (F) Ectopic expression of RPSAP52 increased MYBL2 expression in SCC-4 cell. *P < .01

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ectopic expression of miR-423-5p attenuated luciferase value of MYBL2-WT reporter vector, but not that of MYBL2-mut reporter vector in SCC-4 cell (Figure 5C). Elevated expression of miR-423-5p suppressed MYBL2 expression in SCC-4 cell (Figure 5D). Overexpression of RPSAP52 suppressed miR-423-5p expression in SCC-4 cell (Figure 5E). Ectopic expression of RPSAP52 increased MYBL2 expression in SCC-4 cell (Figure 5F).

3.6 | Ectopic expression of RPSAP52 increased cell growth via modulating MYBL2 in TSCC cell

miR-423-5p level was down-regulated in four TSCC lines (UM1, SCC-4, SCC-1 and Cal-27) compared with NHOK cell (Figure 6A). The expression of MYBL2 was down-regulated in SCC-4 cell after transfected with MYBL2 siRNA (Figure 6B). Knockdown expression of MYBL2 suppressed ki-67 (Figure 6C) and CDK2 (Figure 6D) expression in RPSAP52-overexpressing SCC-4 cell. Down-regulation of MYBL2 suppressed cells at S stage and increased cells at G0-G1 stage in RPSAP52-overexpressing SCC-4 cell (Figure 6E).

Knockdown expression of MYBL2 inhibited cell proliferation in RPSAP52-overexpressing SCC-4 cell (Figure 6F).

3.7 | RPSAP52 induced cytokine secretion through regulating MYBL2 in TSCC cell

The cytokine concentrations of IFN- γ (Figure 7A), IL-1 β (Figure 7B) and IL-6 (Figure 7C) were decreased in SCC-4 cell after treatment with MYBL2 siRNA in RPSAP52-overexpressing SCC-4 cell. Knockdown expression of MYBL2 suppressed cytokine concentration excretion including IL-8 (Figure 7D), IL-10 (Figure 7E) and TGF- β (Figure 7F) expression.

4 | DISCUSSION

Growing IncRNAs have been noted to involve in the initiation and development of several tumours including TSCC. For instance, Zhang et al³⁵ indicated that TUG1 level was overexpressed in TSCC



FIGURE 6 Ectopic expression of RPSAP52 increased cell growth via modulating MYBL2 in TSCC cell. (A) miR-423-5p level was downregulated in four TSCC lines (UM1, SCC-4, SCC-1 and Cal-27) compared with NHOK cell. (B) The expression of MYBL2 was determined by qRT-PCR analysis. (C) The expression of ki-67 was studied by qRT-PCR analysis. (D) The expression of CDK2 was studied using qRT-PCR analysis. (E) Down-regulation of MYBL2 suppressed cells at S stage and increased cells at G0-G1 stage in RPSAP52-overexpressing SCC-4 cell. (F) Knockdown expression of MYBL2 inhibited cell proliferation in RPSAP52-overexpressing SCC-4 cell. **P* < .05 and ***P* < .01



FIGURE 7 RPSAP52 induced cytokine secretion through regulating MYBL2 in TSCC cell. (A) The expression of IFN- γ was determined by ELISA. (B) The expression of IL-1 β was detected using ELISA. (C) The expression of IL-6 was detected using ELISA. (D) The expression of IL-8 was determined by ELISA. (E) The expression of IL-10 was detected using ELISA. (F) The expression of TGF- β was detected using ELISA. *P < .05, **P < .01 and ***P < .001

cells and tissues of cisplatin resistance. Knockdown of TUG1 suppressed cisplatin resistance to CAL27/CDDP and SCC25/CDDP cells via modulating CXCR4 and miR-133b. Lin et al³⁶ noted that PRNCR1 played as one oncogenic IncRNA gene in the development of TSCC via regulating HOXB5/ miR-944. Yan et al³⁷ suggested that knockdown of PCAT-1 suppressed metastasis of proliferation of TSCC cell through up-regulating P21. Liu and workmates found that knockdown of SNHG17 suppressed TSCC cell migration, invasion and proliferation through regulating miR-876/SP1.³⁸ Recently, a new IncRNA RPSAP52 has been identified and revealed to participate in several tumours such as glioblastoma, pituitary tumours and pancreatic cancer.³¹⁻³⁴ Wang et al³³ showed that RPSAP52 predicted postoperative survival and promoted cell stemness in glioblastoma through regulating TGF- β 1. D'Angelo et al³¹ found that RPSAP52 was up-regulated in pituitary cancers and induced cell growth through sponging HMGA. However, the role of RPSAP52 remains unknown. In our study, we indicated that RPSAP52 was higher in TSCC samples compared with that in control samples. The higher expression of RPSAP52 was positively correlated with higher T stage and TNM stage. Ectopic expression of RPSAP52 induced TSCC cell growth and cycle and induced cytokine secretion including IFN- γ , IL-1 β and IL-6, IL-8, IL-10 and TGF- β .

Emerging references illustrated that IncRNAs acted as 'sponges' of miRNAs to involve in several cellular processes. For example, Qiao et al³⁹ showed that KCNQ1OT1 regulated cisplatin resistance of TSCC via TRIM14/miR-124-3p axis. Li et al⁴⁰ illustrated that ADAMTS9-AS2 enhanced TSCC migration, EMT and growth through EZH2/miR-600 axis. Ma et al⁴¹ suggested that GIHCG induced TSCC progression via modulating miR-429. Zuo et al⁴² proved that CASC15 induced TSCC development via sponging miR-33a-5p. Kou et al⁴³ noted that H19 facilitated TSCC invasion and migration through regulating miR-let-7. Recently, Chen et al³⁴ found that RPSAP52 inhibited hypoxia-influenced epithelial cell apoptosis of renal proximal tubular via GSTM1/miR-423-5p axis. We also found that the overexpression of RPSAP52 suppressed miR-423-5p expression in SCC-4 cell. miR-423-5p was lower in TSCC samples compared with that in control samples, and miR-423-5p level was negatively correlated with higher T stage and TNM stage. Pearson's correlation indicated that miR-423-5p was negatively associated with that of RPSAP52 in TSCC tissues. Furthermore, MYBL2 was one direct gene of miR-423-5p and elevated expression of miR-423-5p suppressed MYBL2 expression and ectopic expression of RPSAP52 increased MYBL2 expression in SCC-4 cell. A previous study indicated that MYBL2 acted as one

oncogene in tumour development.^{44,45} Finally, we illustrated that RPSAP52 overexpression promoted TSCC cell growth and cycle and induced cytokine secretion including IFN- γ , IL-1 β and IL-6, IL-8, IL-10 and TGF- β via modulating MYBL2.

In summary, we identified that RPSAP52 was up-regulated in TSCC samples and positively correlated with higher T stage and TNM stage. Ectopic expression of RPSAP52 induced TSCC cell growth and cycle and induced cytokine secretion including IFN- γ , IL-1 β and IL-6, IL-8, IL-10 and TGF- β through regulating miR-423-5p/ MYBL2 axis. These data provided new insight into RPSAP52, which may be one potential treatment target for TSCC.

CONFLICT OF INTEREST

There is no conflict of interest.

AUTHOR CONTRIBUTIONS

Xiaozhen Wu: Conceptualization (equal); Data curation (equal); Writing-original draft (equal); Writing-review & editing (equal). Zuode Gong: Conceptualization (equal); Investigation (equal); Resources (equal); Writing-original draft (equal); Writing-review & editing (equal). Long Ma: Conceptualization (equal); Data curation (equal); Investigation (equal); Writing-original draft (equal). Qibao Wang: Conceptualization (equal); Data curation (equal); Funding acquisition (equal); Project administration (equal); Writing-original draft (equal); Writing-review & editing (equal).

DATA AVAILABILITY STATEMENT

The data will be made available after being required upon request from the corresponding author.

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REFERENCES

- Wu XZ, Gong ZD, Sun LY, Ma L, Wang QB. MicroRNA-802 plays a tumour suppressive role in tongue squamous cell carcinoma through directly targeting MAP2K4. *Cell Prolif.* 2017;50(3):e12336.
- Sun LY, Liang J, Wang QB, Li ZY, Du Y, Xu X. MicroRNA-137 suppresses tongue squamous carcinoma cell proliferation, migration and invasion. *Cell Prolif.* 2016;49:628-635.
- Tseng SH, Yang CC, Yu EH, et al. K14-EGFP-miR-31 transgenic mice have high susceptibility to chemical-induced squamous cell tumorigenesis that is associating with Ku80 repression. *Int J Cancer*. 2015;136:1263-1275.
- Cao J, Guo T, Dong Q, Zhang J, Li Y. miR-26b is downregulated in human tongue squamous cell carcinoma and regulates cell proliferation and metastasis through a COX-2-dependent mechanism. *Oncol Rep.* 2015;33:974-980.
- Manikandan M, Deva Magendhra Rao AK, Munirajan AK. Altered levels of miR-21, miR-125b-2*, miR-134, miR-155, miR-184, and miR-205 in oral squamous cell carcinoma and association with clinicopathological characteristics. *J Oral Pathol Med*. 2014;44(10):792-800.
- Zeng S, Yang J, Zhao J, et al. Silencing dicer expression enhances cellular proliferative and invasive capacities in human tongue squamous cell carcinoma. *Oncol Rep.* 2014;31:867-873.

- Song KB, Liu WJ, Jia SS. miR-219 inhibits the growth and metastasis of TSCC cells by targeting PRKCI. Int J Clin Exp Med. 2014;7:2957-2965.
- Ren W, Wang X, Gao L, et al. MiR-21 modulates chemosensitivity of tongue squamous cell carcinoma cells to cisplatin by targeting PDCD4. *Mol Cell Biochem*. 2014;390:253-262.
- Lin Z, Sun L, Chen W, et al. miR-639 regulates transforming growth factor beta-induced epithelial-mesenchymal transition in human tongue cancer cells by targeting FOXC1. *Cancer Sci.* 2014;105:1288-1298.
- Kai Y, Peng W, Ling W, Jiebing H, Zhuan B. Reciprocal effects between microRNA-140-5p and ADAM10 suppress migration and invasion of human tongue cancer cells. *Biochem Biophys Res Comm.* 2014;448:308-314.
- Jia LF, Wei SB, Gan YH, et al. Expression, regulation and roles of miR-26a and MEG3 in tongue squamous cell carcinoma. *Int J Cancer*. 2014;135:2282-2293.
- Xu JY, Yang LL, Ma C, Huang YL, Zhu GX, Chen QL. MiR-25-3p attenuates the proliferation of tongue squamous cell carcinoma cell line Tca8113. Asian Pac J Trop Med. 2013;6:743-747.
- Ratovitski EA. Phospho-DeltaNp63alpha-dependent microR-NAs modulate chemoresistance of squamous cell carcinoma cells to cisplatin: at the crossroads of cell life and death. FEBS Lett. 2013;587:2536-2541.
- Liu M, Wang J, Huang H, Hou J, Zhang B, Wang A. miR-181a-Twist1 pathway in the chemoresistance of tongue squamous cell carcinoma. *Biochem Biophys Res Comm.* 2013;441:364-370.
- Sun L, Yao Y, Liu B, et al. MiR-200b and miR-15b regulate chemotherapy-induced epithelial-mesenchymal transition in human tongue cancer cells by targeting BMI1. Oncogene. 2012;31:432-445.
- Zhu SB, Fu W, Zhang LY, et al. LINC00473 antagonizes the tumour suppressor miR-195 to mediate the pathogenesis of Wilms tumour via IKK. *Cell Prolif.* 2018;51(1):e12416.
- Zhao J, Gao Z, Zhang C, Wu H, Gu R, Jiang R. Long non-coding RNA ASBEL promotes osteosarcoma cell proliferation, migration and invasion by regulating microRNA-21. J Cell Biochem. 2018;119(8):6461-6469.
- Zhang JM, Yin MN, Peng G, Zhao Y. CRNDE: an important oncogenic long non-coding RNA in human cancers. *Cell prolif*; 2018:51(3):e12440.
- Yu X, Zheng HY, Tse G, Zhang L, Wu WKK. CASC2: an emerging tumour-suppressing long noncoding RNA in human cancers and melanoma. *Cell Prolif.* 2018;51(6):e12506.
- Xu RD, Feng F, Yu XS, Liu ZD, Lao LF. LncRNA SNHG4 promotes tumour growth by sponging miR-224-3p and predicts poor survival and recurrence in human osteosarcoma. *Cell Prolif.* 2018;51(6):e12515.
- 21. Li Z, Li X, She J, Zhang L, Chan M T V, Wu William K K. Emerging roles of non-coding RNAs in scoliosis. *Cell Prolif.* 2020;53:e12736.
- Cao C, Xu Y, Du K, et al. LINC01303 functions as a competing endogenous RNA to regulate EZH2 expression by sponging miR-101-3p in gastric cancer. J Cell Mol Med. 2019;23(11):7342-7348.
- Chen C, Tan H, Bi J, et al. LncRNA-SULT1C2A regulates Foxo4 in congenital scoliosis by targeting rno-miR-466c-5p through PI3K-ATK signalling. J Cell Mol Med. 2019;23:4582-4591.
- 24. Zou YF, Zhong YT, Wu JJ, et al. Long non-coding PANDAR as a novel biomarker in human cancer: a systematic review. *Cell Prolif.* 2018;51(1):e12422.
- Xiong W-C, Han N, Nan W, et al. Interplay between long noncoding RNA ZEB1-AS1 and miR-101/ZEB1 axis regulates proliferation and migration of colorectal cancer cells. *Am J Transl Res.* 2018;10:605-617.
- She KL, Yan H, Huang J, Zhou HP, He JX. miR-193b availability is antagonized by LncRNA-SNHG7 for FAIM2-induced tumour progression in non-small cell lung cancer. *Cell Prolif.* 2018;51(1):e12406.

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- Chen Q, Huang X, Li R. IncRNA MALAT1/miR-205-5p axis regulates MPP-induced cell apoptosis in MN9D cells by directly targeting LRRK2. Am J Transl Res. 2018;10:563-572.
- Pan Y, Wu YJ, Hu JL, et al. Long noncoding RNA HOTAIR promotes renal cell carcinoma malignancy through alpha-2, 8-sialyltransferase 4 by sponging microRNA-124. *Cell Prolif.* 2018;51(6):e12507.
- Ding JH, Yang CX, Yang S. LINC00511 interacts with miR-765 and modulates tongue squamous cell carcinoma progression by targeting LAMC2. J Oral Pathol Med. 2018;47:468-476.
- Lin ZY, Sun LJ, Xie SL, et al. Chemotherapy-induced long non-coding RNA 1 promotes metastasis and chemo-resistance of TSCC via the Wnt/beta-catenin signaling pathway. *Mol Ther.* 2018;26:1494-1508.
- D'Angelo D, Mussnich P, Sepe R, et al. RPSAP52 IncRNA is overexpressed in pituitary tumors and promotes cell proliferation by acting as miRNA sponge for HMGA proteins. J Mol Med. 2019;97:1019-1032.
- Oliveira-Mateos C, Sanchez-Castillo A, Soler M, et al. The transcribed pseudogene RPSAP52 enhances the oncofetal HMGA2-IGF2BP2-RAS axis through LIN28B-dependent and independent let-7 inhibition. *Nat Commun.* 2019;10(1):3979.
- Wang SW, Guo XR, Lv WY, et al. LncRNA RPSAP52 upregulates TGFbeta 1 to increase cancer cell sternness and predict postoperative survival in glioblastoma. *Cancer Manag Res.* 2020;12:2541-2547.
- Chen J, Zheng Y, Li LP. LncRNA RPSAP52 regulates miR-423-5p/ GSTM1 axis to suppress hypoxia-induced renal proximal tubular epithelial cell apoptosis. Arch Physiol Biochem. 2020;1-5. Online ahead of print.
- Zhang K, Zhou H, Yan B, Cao X. TUG1/miR-133b/CXCR4 axis regulates cisplatin resistance in human tongue squamous cell carcinoma. *Cancer Cell Int.* 2020;20:148.
- Lin C, Zou Y, Li R, Liu D. Long non-coding RNA PRNCR1 exerts oncogenic effects in tongue squamous cell carcinoma in vitro and in vivo by sponging microRNA-944 and thereby increasing HOXB5 expression. Int J Mol Med. 2020;46(1):119-130.
- 37. Yan M, Zhao T, Lingling F, Tian R, Li D, Da Y. Knockdown of long non-coding RNA prostate cancer-associated transcript 1 inhibits the proliferation and metastasis of tongue squamous cell carcinoma cells by upregulating p21. Oncol lett. 2020;19:2839-2845.

- Liu X, Zhang B, Jia Y, Fu M. SNHG17 enhances the malignant characteristics of tongue squamous cell carcinoma by acting as a competing endogenous RNA on microRNA-876 and thereby increasing specificity protein 1 expression. *Cell cycle*. 2020;19:711-725.
- Qiao CY, Qiao TY, Jin H, Liu LL, Zheng MD, Wang ZL. LncRNA KCNQ10T1 contributes to the cisplatin resistance of tongue cancer through the KCNQ10T1/miR-124-3p/TRIM14 axis. Eur Rev Med Pharmacol Sci. 2020;24:200-212.
- Li YR, Wan Q, Wang WW, et al. LncRNA ADAMTS9-AS2 promotes tongue squamous cell carcinoma proliferation, migration and EMT via the miR-600/EZH2 axis. *Biomed Pharmacother*. 2019;112:108719.
- Ma L, Wang QB, Gong ZD, Xue LD, Zuo ZB. Long noncoding RNA GIHCG enhanced tongue squamous cell carcinoma progression through regulating miR-429. J Cell Biochem. 2018;119:9064-9071.
- 42. Zuo ZB, Ma L, Gong ZD, Xue LD, Wang QB. Long non-coding RNA CASC15 promotes tongue squamous carcinoma progression through targeting miR-33a-5p. *Environ Sci Pollut R*. 2018;25:22205-22212.
- Kou N, Liu S, Li XJ, et al. H19 Facilitates Tongue Squamous Cell Carcinoma Migration and Invasion via Sponging miR-let-7. Oncol Res. 2019;27:173-182.
- 44. Xiong Y, Wang J, Cheng Y, Zhang X, Ye X. Overexpression of MYBL2 promotes proliferation and migration of non-small-cell lung cancer via upregulating NCAPH. *Mol Cell Biochem.* 2020;468:185-193.
- Li M, Liu Y, Liu J, et al. Circ_0006332 promotes growth and progression of bladder cancer by modulating MYBL2 expression via miR-143. Aging. 2019;11:10626-10643.

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