

G OPEN ACCESS

Citation: Zhao X, Li Y, Zheng M, Bian X, Liu M, Sun Y, et al. (2015) Comparative Analysis of Growth and Photosynthetic Characteristics of (*Populus simonii* × *P. nigra*) × (*P. nigra* × *P. simonii*) Hybrid Clones of Different Ploidides. PLoS ONE 10(4): e0119259. doi:10.1371/journal.pone.0119259

Academic Editor: Rongling Wu, Pennsylvania State University, UNITED STATES

Received: September 2, 2014

Accepted: January 11, 2015

Published: April 13, 2015

Copyright: © 2015 Zhao et al. This is an open access article distributed under the terms of the <u>Creative Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

Funding: The role of the Innovation Project of State Key Laboratory of Tree Genetics and Breeding (Northeast Forestry University) (No. 2013B09) was data collection, analysis, and the role of Fundamental Research Funds for the Central Universities (2572014CA16) support for the research was decision to publish. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. **RESEARCH ARTICLE**

Comparative Analysis of Growth and Photosynthetic Characteristics of (*Populus simonii* × *P. nigra*) × (*P. nigra* × *P. simonii*) Hybrid Clones of Different Ploidides

Xiyang Zhao^{1,3°}, Ying Li^{1°}, Mi Zheng^{1°}, Xiuyan Bian¹, Mengran Liu¹, Yanshuang Sun¹, Jing Jiang¹, Fuwei Wang², Shuchun Li², Yonghong Cui³, Guifeng Liu¹*, Chuanping Yang¹*

1 State Key Laboratory of Tree Genetics and Breeding (Northeast Forestry University), Northeast Forestry University, Harbin, 150040, China, **2** Tree Seedling Management Station, Forestry Department of Jilin Province, Changchun, 130000, China, **3** Seed Orchard of Siping, Siping, 136000, China

• These authors contributed equally to this work.

* liuguifeng@126.com (GFL); yangcp@nefu.edu.cn (CPY)

Abstract

To evaluate differences among poplar clones of various ploidies, 12 hybrid poplar clones (P. simonii × P. nigra) × (P. nigra × P. simonii) with different ploidies were used to study phenotypic variation in growth traits and photosynthetic characteristics. Analysis of variance showed remarkable differences for each of the investigated traits among these clones (P < 0.01). Coefficients of phenotypic variation (PCV) ranged from 2.38% to 56.71%, and repeatability ranged from 0.656 to 0.987. The Pn (photosynthetic rate) photosynthetic photon flux density (PPFD) curves of the 12 clones were S-shaped, but the Pn-ambient CO₂ (Ca) curves were shaped like an inverted "V". The stomatal conductance (Gs)-PPFD and transpiration rate (Tr)-PPFD curves had an upward tendency; however, with increasing PFFD, the intercellular CO₂ concentration (Ci)-PPFD curves had a downward tendency in all of the clones. The Pn-PPFD and Pn-Ca curves followed the pattern of a quadratic equation. The average light saturation point and light compensation point of the triploid clones were the highest and lowest, respectively, among the three types of clones. For Pn-Ca curves, diploid clones had a higher average CO₂ saturation point and average CO₂ compensation point compared with triploid and tetraploid clones. Correlation analyses indicated that all investigated traits were strongly correlated with each other. In future studies, molecular methods should be used to analyze poplar clones of different ploidies to improve our understanding of the growth and development mechanisms of polyploidy.



Competing Interests: The authors have declared that no competing interests exist.

Introduction

Poplars (*Populus* spp.) are some of the most important economic tree species in the temperate regions of the world $[\underline{1}, \underline{2}]$. With the publication of the *P. trichocarpa* genome $[\underline{3}]$, poplar has become a model organism for the study of trees and is the most intensively studied tree genus. In 1959, the Chinese Academy of Forestry (CAF) conducted a large-scale crossing experiment on poplar. After 20 years, hybridized combinations of *P. simonii* × *P. nigra* and *P. nigra* × *P. simonii* were selected as excellent materials for afforestation in Northern China because of the rapid growth, excellent wood properties, high cold resistance and drought endurance of their offspring $[\underline{4}, \underline{5}]$. In recent years, many studies have been conducted using these two families as materials, which have mainly focused on growth traits $[\underline{6}, \underline{7}]$, resistance $[\underline{8}]$, physiology $[\underline{9}]$, and molecular research [10, 11].

Polyploidy is a ubiquitous phenomenon in higher plants. It is estimated that polyploidy has occurred in 50-70% of flowering species [12, 13], most of which have experienced one or more polyploidization events during their evolution [14]. The increase in chromosomes has resulted in increased gene dosages and cell volumes [15]. Therefore, polyploid plants usually have larger leaves, greater height and diameter, and an increased ability to adapt to their environment [16, 17]. The first natural European aspen polyploid (Populus tremula) was discovered by Nilsson-Ehle [18] in Sweden. Since then, breeders have paid close attention to different ploidy level (diploid, triploid and tetraploid) breeding in forest trees because of the huge growth of forestry. Many natural triploid poplars were found in the Soviet Union, Sweden, Finland and other countries [19-22]. In research on different types of polyploids, triploid poplars are often characterized as having fast growth, large leaves, vigor and low fertility compared with their diploid counterparts [23]. However, fewer comparative studies on poplars with different ploidies, including tetraploids, have been reported [24]. In this study, with the aim of obtaining high growth rate and highly resistant offspring, *P. simonii* × *P. nigra* and *P. nigra* × *P. simonii* were selected as parents. Colchicine was used in the crossing experiment and many offspring were obtained with different ploidies. Twelve offspring with different ploidies were used as materials in this experiment. Our primary objectives were to explore the variation in growth, photosynthesis and chlorophyll fluorescence traits among hybrid clones with different ploidies, and to provide a theoretical basis for polyploid poplar clone selection.

Materials and Methods

Plant materials

Female *Populus simonii* × *P. nigra* and male *P. nigra* × *P. simonii* plants (both of the parents were diploids) were selected as parents, and artificially controlled pollination was performed in 2006. During male and female flower development, a 0.5% colchicine solution was injected into flower buds at the beginning of reduction mitosis to obtain reduplicated pollen and metrocytes. The crossing experiment was carried with reduplicated pollen and reduplicated metrocytes to obtain polar seeds with different ploidies. After sowing, the DNA contents in the leaves were evaluated by flow cytometry using the method of Zhang [25]. Twelve hybrid clones with different ploidies [(*Populus simonii* × *P. nigra*) × (*P. nigra* × *P. simonii*)] were used for this study, including three tetraploid clones (sn4.1, sn4.2 and sn4.3), five triploid clones (sn3.1, sn3.2, sn3.4, sn3.8 and sn3.9) and four diploid clones (sn, sn2.1, sn2.2, sn2.3). The total DNA contents of each clone were measured and the percentages of the cells that showed varied DNA content were processed by the DPAC software. The results of sn2.1, sn3.8 and sn4.1 are shown in Fig 1. In 2013, 50 hardwood cuttings (15 cm long each) of each clone were collected from 1-year-old seedlings in a greenhouse at the Northeast Forestry University. All cuttings were



Fig 1. DNA content of sn2.1 (a: the main peak at channel 100), sn3.8 (b: the main peak at channel 150) and sn4.1 (c: the main peak at channel 200). The ordinate (count) represents the relative value of the cell population, the abscissa (FL1) represents the passageway value of fluorescence. The abscissa location which peak point corresponding were the ploidy of test sample.

moistened and stored in plastic bags at 4°C for about 2 weeks and then planted in 1.5-L plastic pots containing garden soil mixed with peat at a ratio of 3:1 (v/v). The cuttings were grown in artificial climate chambers of Northeast Forestry University under a cycle of 1000 μ mol m⁻² s⁻¹ light for 16 h from 8:00 to 24:00 and dark for 8 h. The temperature and humidity were set at 27°C and 60%, respectively.

Growth traits observation

After growth for 3 months, 30 healthy and uniform cuttings of each clone were used as materials to measure tree height (H), basal diameter (BD) and leaf number (LN) of each plant. Nine leaves (4–12) from a shoot tip per cutting were taken to measure the leaf area (LA). Each leaf and stem (the whole stem) was weighed for the leaf fresh weight (LFW) and stem fresh weight (SFW). Leaves and stems were then dried in an oven (90°C) to a constant weight and measured for leaf dry weight (LDW) and stem dry weight (SDW).

Photosynthetic analysis

Nine uniform plants of each clone were selected to measure the photosynthetic parameters. Net photosynthesis rate (Pn), intercellular CO_2 concentration (Ci), stomatal conductance (Gs) and transpiration rate (Tr) were measured from 8:30 a.m. to 11:30 a.m. using a Lico-6400 portable photosynthesis measuring system (Li-Cor Inc., Lincoln, NE, USA) on the third to fifth fully expanded leaf of each plant. The conditions for photosynthetic trait measurements were: leaf temperature 28°C, photosynthetic photon flux density (PPFD) 1400 µmol m⁻² s⁻¹, relative air humidity (RH) 60% and ambient CO_2 concentration (Ca) 400 µmol mol⁻¹.

Light and CO₂ response measurements

Pn, Gs, Ci and Tr responses to PPFD and Ca were also measured. Photosynthesis traits-PPFD curves were produced under a Ca concentration of 400 µmol mol⁻¹, leaf temperature around

28°C and relative humidity about 60%. The light intensities varied with 0, 20, 50, 80, 100, 200, 400, 600, 800, 1000, 1200, 1400, 1600, 1800 and 2000 μ mol m⁻² s⁻¹. Response curves were modeled by a quadratic equation (Eq 1) according to Zhao [26, 27]. Light saturation point (LSP) and light compensation point (LCP) were evaluated by fitting data to the model function. Pn-Ca curves were measured under saturating light intensity (1400 μ mol m⁻² s⁻¹), and a temperature and humidity around 28°C and 60%. The CO₂ concentrations in the leaf chamber were set at 0, 50, 80, 100, 200, 300, 400, 600, 800, 1000, 1200, 1400 and 1600 μ mol mol⁻¹. Model 1 was used to create response curves and to evaluate the carbon dioxide saturation point (CSP) and carbon dioxide compensation point (CCP).

$$Y = b_0 + b_1 X + b_2 X^2 \tag{1}$$

Where *Y* was the Pn value, *X* was the PPFD (Ca), b_0 was constant, and b_1 and b_2 were coefficients, respectively.

Chlorophyll fluorescence parameter measurements

Chlorophyll fluorescence parameters were measured using a pulse amplitude modulation chlorophyll fluorometer MINI-PAM2500 (Walz, Effeltrich, Germany). Minimal fluorescence, F₀, was measured in 30-min dark-adapted leaves using weak modulated light of < 0.15 μ mol m⁻² s⁻¹. Maximal fluorescence, Fm, was measured after a 0.8-s saturating white light pulse (6000 μ mol m⁻² s⁻¹) in the same leaves. Maximal variable fluorescence (F_v = F_m-F₀) and the photochemical efficiency of PSII (Fv/Fm) for dark-adapted leaves were calculated.

Data analysis

Statistical analyses were carried out using the Statistical Product and Service Solutions (SPSS 19.0) software. All the growth traits, instantaneous photosynthetic and chlorophyll fluorescence parameters were compared using analysis of variance; the significance of fixed effects was tested with *F*-tests.

The coefficient of phenotypic variation (PCV) of all the investigated traits was calculated using the following formula [28]:

$$PCV = \frac{SD \times 100}{\overline{X}} \tag{2}$$

where \bar{X} is the phenotypic mean of the trait and SD is the standard deviation of the trait.

The repeatability (*R*) of all the investigated traits was calculated as follows [29]:

$$R = \frac{\sigma_c^2}{\sigma_c^2 + \sigma_e^2} \tag{3}$$

where σ_c^2 is the genetic variance component between clones and σ_e^2 is the error variance component.

1

The phenotype correlation $r_A(xy)$ of traits x and y was calculated as follows [30]:

$$r_A(xy) = \frac{\sigma_{a(xy)}}{\sqrt{\sigma_{a(x)}^2 \cdot \sigma_{a(y)}^2}}$$
(4)

where $\sigma_{a(x)}^2$ is the clone variance component for the trait x, $\sigma_{a(y)}^2$ is the clone variance component for the trait y and $\sigma_{a(xy)}$ is the clone covariance component.

Traits	Variation source	SS	df	MS	F	P value	PCV (%)	R
н	Ploidies	1058.400	2	529.200	41.422	0.000**		
	Clones	1197.333	11	108.848	9.242	0.000**	18.76	0.892
BD	Ploidies	2.896	2	1.448	12.221	0.000**		
	Clones	4.058	11	0.369	3.221	0.008**	12.60	0.690
LN	Ploidies	179.706	2	89.853	14.886	0.000**		
	Clones	272.889	11	24.808	5.617	0.000**	18.86	0.822
LA	Ploidies	1365.937	2	682.969	17.530	0.000**		
	Clones	1867.071	11	169.734	5.192	0.000**	27.89	0.807
SFW	Ploidies	77.325	2	38.663	31.385	0.000**		
	Clones	104.961	11	9.542	17.593	0.000**	43.36	0.943
SDW	Ploidies	3.054	2	1.527	10.839	0.000**		
	Clones	6.310	11	0.574	9.878	0.000**	42.08	0.899
LFW	Ploidies	.568	2	.284	11.716	0.000**		
	Clones	0.782	11	0.071	2.907	0.014*	22.73	0.656
LDW	Ploidies	.079	2	.040	15.886	0.000**		
	Clones	0.101	11	0.009	3.613	0.004**	18.76 12.60 18.86 27.89 43.36 42.08 22.73 27.01	0.723

Table 1. ANOVA analyses, PCV and R of different traits among different ploidies and clones.

**(*) indicated variance is significant at the 0.01(0.05) level

doi:10.1371/journal.pone.0119259.t001

PLOS ONE

Results

Variation among growth traits

The results of ANOVA for all growth traits are presented in <u>Table 1</u>. There were significant differences among the clones with different ploidies (P < 0.01) and clones, except for LFW among clones (P = 0.014) based on overall *F*-tests. The PCVs of different growth traits ranged from 12.60% to 43.36%, with the value of BD being the lowest and that of SFW being the highest, respectively. The repeatabilities (R) of the different traits are also presented in <u>Table 1</u>. The R values for all growth traits were higher than 0.60. High R and PCV are favorable for excellent clone selection.

Average growth traits for different ploidy poplar clones

The average growth traits of hybrid poplar clones with different ploidies are summarized in Table 2. The overall average tree height and average base diameter were 35.54 cm and 3.49 mm, respectively, for all plants. The average values of all growth traits in the triploids were higher than those in the tetraploid and diploid hybrid clones, and the diploid hybrid clones had the lowest growth traits.

Pn-PPFD curves

The Pn-PPFD curves of different poplar clones are shown in Fig 2. The Pn-PPFD curves of all 12 clones were S-shaped. The Pn values of all clones increased as PPFD increased in low light conditions and reached a plateau when PPFD reached 1200–1400 μ mol m⁻² s⁻¹. The triploid hybrid clones showed higher Pn values than the diploid and tetraploid hybrid clones. In particular, sn3.8 had the highest Pn values across all light intensities.



Ploidy	clone	H (m)	BD (mm)	LN	LA (cm ²)	SFW (g)	SDW (g)	LFW (g)	LDW (g)
Tetraploid	4.1	33.00±2.65	3.51±0.18	15.67±2.08	27.44±1.64	3.88±0.16	0.95±0.18	0.78±0.16	0.21±0.04
	4.2	35.00±2.68	3.54±0.24	16.67±2.08	30.00±1.73	3.92±0.80	0.99±0.14	0.88±0.19	0.27±0.06
	4.3	31.00±2.44	3.52±0.12	15.67±2.08	28.11±2.03	3.85±0.46	0.90±0.19	0.79±0.10	0.21±0.05
	average	33.00±2.87	3.52±0.16	16.00±1.87	28.52±2.61	3.88±0.47	0.95±0.19	0.82±0.14	0.23±0.05
Triploid	3.1	39.33±2.08	3.77±0.18	20.67±2.08	37.00±6.09	5.46±0.31	1.46±0.45	0.94±0.12	0.34±0.04
	3.2	41.33±3.15	3.81±0.18	23.33±2.31	39.33±6.37	6.52±0.50	1.61±0.10	1.07±0.20	0.31±0.03
	3.4	38.67±2.03	3.55±0.39	18.00±2.65	31.67±7.64	4.71±0.49	1.03±0.28	0.92±0.20	0.33±0.04
	3.8	45.00±4.58	4.17±0.30	21.33±2.89	47.00±3.67	7.56±0.64	2.21±0.33	1.19±0.11	0.29±0.02
	3.9	39.33±3.21	3.62±0.50	17.00±1.73	36.33±4.73	5.01±0.89	0.96±0.22	0.94±0.10	0.25±0.03
	Average	40.73±3.83	3.78±0.35	20.07±3.10	38.27±8.27	5.85±1.36	1.45±0.53	1.01±0.17	0.31±0.06
Diploid	2.1	30.33±4.93	3.40±0.42	15.67±2.31	27.67±5.49	3.71±0.88	0.87±0.19	0.75±0.23	0.20±0.05
	2.2	25.67±1.15	2.96±0.62	14.00±1.80	23.67±3.79	1.11±0.28	0.61±0.02	0.74±0.20	0.20±0.04
	2.3	28.33±5.03	2.92±0.39	14.67±2.08	19.78±1.58	2.16±0.10	0.85±0.09	0.64±0.10	0.20±0.03
	2.4	29.00±2.65	3.21±0.18	16.67±1.53	26.56±5.87	2.94±0.20	0.94±0.26	0.78±0.07	0.21±0.02
	average	28.33±3.70	3.13±0.42	15.25±1.82	24.42±5.00	2.48±1.08	0.82±0.20	0.73±0.15	0.20±0.04

Table 2. Average growth traits for different hybrid clones of different ploidies.

Gs-PPFD, Ci-PPFD and Tr-PPFD curves

Gs-PPFD, Ci-PPFD and Tr-PPFD curves are shown in Figs 3-5. Gs and Tr values had an upward tendency in all clones as PPFD increased. Clones sn3.8, sn3.2 and sn3.1 had higher Gs values than the other clones at different light intensities: as PPFD increased, these differences became larger. Clones sn2.3, sn2.2 and sn had obviously lower Gs values than the other clones. By contrast, the Ci-PPFD curves showed a downward tendency, and the Ci values of all clones declined as PPFD increased. This indicated that as Pn values increased, more CO₂ was needed for photosynthesis, and the concentration of CO₂ used between the cell spaces decreased. In







Fig 3. Gs-Par curves of poplar clones of different ploidies.

the Ci-PPFD curves, the Ci values of sn2.3, sn2.2 and sn were higher than those of the other clones, while the Ci values of sn3.8, sn3.2 and sn3.1 were much lower.

Pn-PPFD curve model

To understand the change in Pn values at different levels of PPFD, quadratic functions were used to simulate the Pn-PPFD relationship. Two simulation curves for sn3.8 and sn2.3 are presented in Fig 6. The parameters used for the simulation curves are listed in Table 3. Parameter R^2 was higher than 0.9, which indicated that the model was effective. The parameters used for LSP and point LCP are also presented in Table 3. The average LSP of the triploid poplar clones









Fig 5. Tr-Par curves of poplar clones of different ploidies.

exceeded that of the tetraploid and diploid clones. The LCP of the triploid poplar clones was lower than that of the diploid and tetraploid clones, which suggested that the triploid poplar clones had greater adaptability to different light intensities. When the PPFD reached the LCP, different clones expressed Pn differently. Pn varied from 15.90 μ mol m⁻² s⁻¹ (sn2.3) to 25.01 μ mol m⁻² s⁻¹ (sn3.8) and the average Pn of the triploid clones was higher than that of the tetraploids and diploids.

Pn-Ca curves

The Pn-Ca curves of different poplar clones are shown in Fig 7. The Pn-Ca curves were shaped like an inverted "V". As Ca increased, Pn increased initially and then decreased. The pattern was the same as the Pn-PPFD curves; triploid clones showed higher values than diploid and tetraploid clones. In particular, sn3.2 and sn3.8 showed clearly higher values at different CO_2 concentrations. The Pn-Ca curves were also simulated using quadratic models; the simulated curves for sn3.8 and sn2.3 are shown in Fig 8 and the model parameters are presented in





Ploidy	clone	а	b	с	LSP	LCP	Pn	R ²
Tetraploid	sn4.1	-7.007E-06	0.0210	1.710	1496.91	7.07	17.41	0.955
	sn4.2	-7.452E-06	0.0213	2.354	1428.14	5.17	17.55	0.947
	sn4.3	-8.096E-06	0.0237	2.244	1465.79	4.86	19.64	0.951
	average				1463.61	5.70	18.20	
Triploid	sn3.1	-9.301E-06	0.0273	1.930	1467.55	8.74	21.96	0.960
	sn3.2	-9.488E-06	0.0285	2.406	1502.98	3.85	23.84	0.966
	sn3.4	-7.815E-06	0.0233	2.383	1489.67	5.67	19.72	0.957
	sn3.8	-9.745E-06	0.0295	2.744	1511.49	2.91	25.01	0.965
	sn3.9	-9.193E-06	0.0260	2.603	1415.46	3.06	21.02	0.956
	average				1477.43	4.84	22.31	
Diploid	sn	-7.681E-06	0.0225	1.420	1465.54	23.96	17.92	0.953
	sn2.1	-8.445E-06	0.0232	2.290	1371.51	10.05	18.17	0.926
	sn2.2	-7.681E-06	0.0210	1.966	1365.92	13.22	16.30	0.929
	sn2.3	-6.923E-06	0.0192	2.556	1388.24	10.71	15.90	0.922
	average				1397.80	14.48	17.07	

Table 3. Parameters for each Pn-PPFD simulated curve.

Parameters a, b and c are coefficients of the quadratic function. R^2 is the criterion for evaluating the effectiveness of the model. The units for LSP, LCP and Pn are µmol m⁻² s⁻¹.

doi:10.1371/journal.pone.0119259.t003

<u>Table 4</u>. The R^2 values of all curve models were higher than 0.9, which indicated that all models were highly efficient. The diploid poplar clones showed higher average CSP and CCP values than the triploids and tetraploids. The average Pn value of the CO₂ concentration at the CSP of diploid clones was lower than in other ploidy level poplar clones. Clone sn3.8 still showed a higher Pn at the CSP, but sn had a lower value.







Fig 8. Pn-Ca simulated curve of clone sn3.8 (left) and sn2.3 (right). Solid lines are the simulated curves and white circles are the observed data.

Instantaneous photosynthetic and chlorophyll fluorescence factors

ANOVA analyses of photosynthetic and chlorophyll fluorescence factors among different clones are shown in <u>Table 5</u>. All factors showed significant differences (P < 0.01) among clones or ploidies levels. PCV varied from 2.38% (Fv/Fm) to 56.71% (Gs) and repeatability ranged from 0.745 (Ci) to 0.987 (Fv/Fm). The average values for each photosynthetic factor under conditions of leaf temperature 28°C, PPFD 1400 µmol m⁻² s⁻¹, RH 60% and Ca 400 µmol mol⁻¹ are shown in <u>Table 6</u>. The triploids showed high Pn, Gs and Tr, but low Ci values. As with the photosynthetic factors, the triploid poplar clones had higher Fv/Fm values than the diploid and tetraploid clones.

Ploidy	Clone	а	b	С	CSP	CCP	Pn	R ²
Tetraploid	sn4.1	-2.573E-05	0.045	0.339	865.363	11.341	19.605	0.935
	sn4.2	-2.484E-05	0.047	-0.415	945.794	24.341	21.807	0.975
	sn4.3	-2.485E-05	0.048	-1.334	958.039	18.604	21.475	0.988
	Average				923.066	18.096	20.962	
Triploid	sn3.1	-2.876E-05	0.053	-0.600	919.055	21.177	23.689	0.971
	sn3.2	-3.031E-05	0.057	-0.591	937.974	27.083	26.079	0.967
	sn3.4	-2.950E-05	0.055	-0.565	927.981	24.179	24.836	0.969
	sn3.8	-2.980E-05	0.056	-0.446	940.023	27.024	25.884	0.965
	sn3.9	-2.912E-05	0.053	-0.623	914.070	21.667	23.709	0.973
	Average				927.821	24.226	24.839	
Diploid	Sn	-1.166E-05	0.028	-1.555	1216.240	33.422	15.694	0.971
	sn2.1	-2.030E-05	0.038	-0.530	928.700	30.049	16.979	0.980
	sn2.2	-1.174E-05	0.029	-1.599	1214.183	34.145	15.705	0.971
	sn2.3	-1.572E-05	0.035	-0.746	1114.362	8.706	18.775	0.991
	Average				1118.371	26.580	16.788	

Table 4. Coefficients and variation parameters for each Pn-Ca simulated curve.

Parameters a, b and c are coefficients of the quadratic function. R^2 is the criterion for evaluating the effectiveness of the model. The unit for CSP and CCP is µmol mol⁻¹ and the unit for Pn is µmol m⁻² s⁻¹.

doi:10.1371/journal.pone.0119259.t004

PLOS ONE



Traits	Variation source	SS	df	MS	F	P value	PCV (%)	R
Pn	Ploidies	197.450	2	98.725	30.582	0.000**		
	Clones	279.979	11	25.453	25.453	0.000**	16.29	0.961
Gs	Ploidies	2.472	2	1.236	26.497	0.000**		
	Clones	3.899	11	0.354	76.298	0.000**	56.71	0.987
Ci	Ploidies	9426.351	2	4713.176	6.113	0.006**		
	Clones	14620.579	11	1329.144	3.922	0.002**	10.68	0.745
Tr	Ploidies	78.277	2	39.138	33.372	0.000**		
	Clones	108.312	11	9.847	27.266	0.000**	31.77	0.963
Fv/Fm	Ploidies	0.010	2	0.005	62.893	0.000**		
	Clones	0.013	11	0.001	79.311	0.000**	2.38	0.987

Table 5. ANOVA analyses for PCV and R of clones of different ploidies and clones in photosynthetic and chlorophyll fluorescence traits.

** indicates that the variance is significant at the 0.01 level.

doi:10.1371/journal.pone.0119259.t005

Correlation coefficients

The correlation coefficients of different traits are shown in Table 7. All pair-wise correlation coefficients were significant (P < 0.01). The correlation coefficients among growth traits (H, BD, LN, LA, LFW, SFW, LDW and SDW) ranged from 0.512 to 0.857. There were also significant positive correlations between Pn and Gs, Pn and Tr, and Gs and Tr, but Ci was negatively correlated with the other photosynthetic factors. All growth traits were positively correlated with Pn, Gs and Tr, and negatively correlated with Ci. Fv/Fm was positively correlated with growth factors, Fv/Fm was positively correlated with Pn, Gs and Tr, and negatively correlated with Ci.

Ploidy	clone	Pn	Gs	Ci	Tr	Fv/Fm
Tetraploid	4.1	15.90	0.40	311.00	5.19	0.813
	4.2	16.30	0.63	300.00	5.41	0.815
	4.3	18.00	0.53	310.00	6.23	0.800
	Average	16.73	0.52	307.00	5.61	0.809
Triploid	3.1	20.33	1.21	276.00	8.63	0.828
	3.2	22.50	0.93	274.67	7.65	0.838
	3.4	18.20	0.58	293.67	5.84	0.815
	3.8	23.80	1.21	259.67	8.63	0.846
	3.9	19.30	0.52	300.00	5.86	0.827
	Average	20.83	0.89	280.80	7.32	0.831
Diploid	2.1	16.80	0.37	307.00	4.82	0.792
	2.2	14.80	0.26	321.00	3.27	0.775
	2.3	14.63	0.22	324.00	3.39	0.786
	2.4	16.57	0.30	316.00	4.13	0.798
	Average	15.70	0.29	317.00	3.90	0.788

Table 6. Average photosynthetic and chlorophyll fluorescence traits of hybrid clones of different ploidies.

The units for Pn, Gs, Ci and Tr are µmol m⁻² s⁻¹, µmol m⁻² s⁻¹, µmol mol⁻¹ and mol m⁻² s⁻¹, respectively.

Table 7.	Pair-wise	correlation	coefficients	of d	ifferent traits.
----------	-----------	-------------	--------------	------	------------------

Trait	BD	LN	LA	SFW	SDW	LFW	LDW	Pn	Gs	Ci	Tr	Fv/Fm
н	0.657**	0.692**	0.786**	0.840**	0.657**	0.726**	0.638**	0.755**	0.756**	-0.489**	0.748**	0.858**
BD		0.535**	0.567**	0.681**	0.512**	0.618**	0.584**	0.640**	0.673**	-0.460**	0.674**	0.748**
LN			0.690**	0.716**	0.678**	0.536**	0.655**	0.705**	0.727**	-0.518**	0.663**	0.758**
LA				0.769**	0.725**	0.717**	0.632**	0.794**	0.710**	-0.575**	0.739**	0.820**
SFW					0.857**	0.764**	0.646**	0.842**	0.810**	-0.598**	0.822**	0.887**
SDW						0.689**	0.594**	0.768**	0.803**	-0.573**	0.722**	0.770**
LFW							0.586**	0.658**	0.626**	-0.557**	0.607**	0.739**
LDW								0.533**	0.633**	-0.502**	0.593**	0.633**
Pn									0.858**	-0.449**	0.920**	0.856**
Gs										-0.469**	0.938**	0.839**
Ci											-0.457**	-0.660**
Tr												0.845**

**Correlation is significant at the 0.01 level (2-tailed).

doi:10.1371/journal.pone.0119259.t007

Discussion

Polyploidy is an important force in plant evolution [31]. In general, because of the increased heterozygosity and genic and biochemical flexibility provided by the additional alleles, polyploid plants are often larger and have larger cells. Polyploids are considered better able to colonize a wider range of habitats and survive in harsh, unstable environments [23], [31]. The major cytological mechanisms of polyploidy are the union of unreduced gametes, somatic doubling and polyspermy [32]. The union of unreduced gametes used in this research is one of the most important mechanisms for obtaining polyploids [32]. In terms of growth traits at different ploidies, Zhu [24] found that there were significant differences between diploid and tetraploid *P. davidiana* Dode hybrid clones in terms of tree height, basal diameter and chlorophyll content. In 1995, Zhu found that triploid P. tomentosa clones grew faster than diploid clones [33]. In this study, triploid poplar clones had clearly higher values for all growth traits compared with diploid and tetraploid poplar clones. Tetraploid poplar clones did not show superiority to diploid poplar clones in growth traits, which agreed with the results of Comai [34] who indicated that because of the ploidy increase, increased numbers of chromosomes led to increased numbers of cells and nuclei, thereby increasing the possibility of producing aneuploidy in meiosis and mitosis, resulting in epigenetic instability.

The most common parameter used to describe the extent of variability in a breeding population is the average PCV. In this study, the average PCVs of the traits ranged from 2.38% (Fv/ Fm) to 56.71% (Gs). Except for Fv/Fm, the PCV of all traits exceeded 10%. The repeatability value indicates the reliability with which a genotype can be recognized by its phenotypic expression. In this study, the estimates of repeatability for all traits at the clone mean level ranged from 0.656 (LFW) to 0.987 (Fv/Fm), which is in general agreement with research by Kien [35]. The high PCV and high repeatability indicated that the variation of each trait in poplar hybrid clones with different ploidies is not significantly influenced by environmental factors and that selection and evaluation of these clones will be effective [36].

Light can affect the enzyme activity and stomatal aperture of plant leaves. Without light, there is no photosynthesis. The light compensation point is a critical value; at this point, the amounts of CO_2 used in photosynthesis and created by respiration are equal. The light saturation point is a limiting value; over this value, the photosynthetic rate does not increase, and

may decrease [37]. The LSP in plants is important under high light conditions, as the dark reaction follows the light reaction in photosynthesis and then limits the instantaneous photosynthetic rate [38]. Poplars are shade-avoiding species, and our results showed that they had a high photosynthetic rate ($15.90-25.01 \mu mol m^{-2} s^{-1}$) when the light radiation reached the LSP. The five triploid clones had higher LSPs and lower LCPs, indicating that triploid clones can adapt to extensive light illumination intensities. These results were similar to those of Fu [39] for rice and Qi [16] for poplar. At the LSP, the Pn values of the triploid clones were higher than those of the tetraploid and diploid clones. There were no significant variations between the tetraploid and diploid clones. These variations might reflect differences in leaf structure, composition, transcriptional level, methylation of DNA or nucleolus dominance in clones of different ploidies, which should be explored in future research [16]. As the light illumination intensity increased, Gs and Tr showed the same regular pattern as Pn, indicating that the photosynthetic indices interacted with each other during photosynthesis. These results are in agreement with research carried out on *P. tomentosa* [26] and *Catalpa bungei* [27].

 CO_2 is the most important substrate in photosynthesis. Within a certain concentration range, enhanced CO_2 concentration can promote the instantaneous photosynthetic rate [40]. There is strong evidence that plants have already responded to the increase in atmospheric CO_2 concentration [41]. Atmospheric CO_2 concentrations are projected to double from the current concentration of 350 µmol mol⁻¹ to 700 µmol mol⁻¹, which will further stimulate plant growth and ecosystem changes within the next 80 years [37]. CO_2 concentration is the main limiting factor of photosynthesis for plants when the temperature and humidity are moderate and the light radiation is saturated. In this study, as the CO_2 concentration increased, the Pn values of poplar clones with different ploidies initially increased and then decreased. The CSP values of the diploids were obviously higher than those of the tetraploid and triploid poplar clones. However, at the CSP, the Pn values of the triploids were higher than those of the diploids and tetraploids. These results suggested that after 80 years, when the CO_2 concentration is higher CSP.

Chlorophyll fluorescence is a measure of photosynthetic performance and is widely used by plant physiologists and ecophysiologists [42]. The ratio Fv/Fm is the most frequently used parameter in ecophysiological research to determine the maximum photochemical efficiency of PSII [43, 44]. This ratio, typically ranging between 0.75 and 0.85, is proportional to the effectiveness of light energy utilization under standard conditions of CO_2 fixation and to the quantum yield of photochemical processes [45]. In this study, Fv/Fm values ranged from 0.775 to 0.846, which indicated that the plants were not under stress and that the greenhouse conditions were appropriate for poplar growth. However, the Fv/Fm values were higher in the triploid and tetraploid clones than in the diploid clones. Polyploid poplar clones might have higher pre-resistance than diploid poplar clones [46].

Ceulemans [47] and Ma [48] indicated that the Pn and Gs of *Populus* clones were positively correlated with growth traits; Pn and Gs are tightly coupled [49]. In this research, strong positive correlations between growth traits and Pn, Gs, Tr and Fv/Fm were observed, and there were also significant correlations among all photosynthetic parameters, indicating that all traits influence and restrict each other, ultimately controlling plant growth.

Conclusion

In conclusion, there were significant variations in growth, photosynthetic and chlorophyll fluorescence (Fv/Fm) traits between different clones with ploidies, and there were also significant correlations between different traits, which suggested that all traits collaborated to promote plant growth. Triploid clones showed higher growth traits and higher instantaneous photosynthetic and chlorophyll fluorescence factors than diploid and tetraploid clones, which indicated that triploid clones were preferable at the seedling stage. Clone sn3.8 showed high growth traits, Pn and Fv/Fm values, and should be investigated further in future studies. Additional investigations are also needed to determine the transcriptomic, proteomic, metabolomic and structural differences between poplars with different ploidies, to explain the variation in chromosome polyploidy, and to improve the genetic manipulation of poplar.

Acknowledgments

We acknowledge the Innovation Project of State Key Laboratory of Tree Genetics and Breeding (Northeast Forestry University) (No. 2013B09) and Fundamental Research Funds for the Central Universities (2572014CA16) support for the research. We are also grateful to Dr. Wei Hou for critical reading of the manuscript.

Author Contributions

Conceived and designed the experiments: XZ GL CY. Performed the experiments: XZ MZ XB ML. Analyzed the data: YS SL CY JJ. Contributed reagents/materials/analysis tools: CY JJ FW SL YC XB. Wrote the paper: XZ YL. Critical reading of the manuscript: YL.

References

- 1. Zhao XY, Zheng HQ, Li SW, Yang CP, Jiang J, Liu GF. The rooting of poplar cutting: a review. New Forests. 2013; doi: 10.1007/s11056-013-9389-1
- Sixto H, Salvia J, Barrio M, Ciria M, Canellas C. Genetic variation and genotype- environment interactions in short rotation Populus plantations in southern Europe. New Forests. 2011; 42:163–177
- Tuskan GA, Difazio S, Jansson S, Bohlmann J, Grigoriev I, Hellsten U, et al. The genome of black cottonwood, *Populus trichocarpa*. Science. 2006; 313:1596–1604 PMID: <u>16973872</u>
- Li XX, Wang JJ, Pang ZW, Zhao XJ, Li YC. Evaluation of growth and management density for Yingchun poplar. Journal of Northeast Forestry University. 1998; 26: 24–27
- 5. Jiang ZH, Fan SH, Feng HX, Zhang Q, Liu GL, Zong YC. Biomass and distribution patterns of *Populus×xiaohei* plantation in sandy land of north china. Scientia Silvae Sinicae. 2007; 43:15–20
- 6. Fan SH, Liu GL, Zhang Q, Feng HX, Zong YC, Ren HQ. A study on biomass and productivity of *Populus×xiaohei* plantation on study land in north china. Forest Research. 2010; 23:71–76
- Li ZX, Zhao XY, Yang CJ, Wang GY, Wang FS, Zhang LF, et al. Variation and growth adaptability analysis of transgenic *populous simonii*×*P.nigra*. lines carrying *TaLEA* gene. Journal of Beijing Forestry University. 2013; 35: 57–62
- Wang L, Zhou BR, Wu LL, Lv CY, Qu YJ, Zheng W, et al. Cloning and expression analysis of a ring zinc-finger gene in *populous simonii×P.nigra*. Plant Physiology Communications. 2009; 45(12):1160–1166
- 9. Wang S, Liu MR, Huang HJ, Mu HZ, Li ZX, Liu GF. Selection of cold resistant strains from *TaLEA* gene transferred *populous simoniixP.nigra*. Journal of Northeast Forestry University. 2011; 39: 5–7,16
- Yang CP, Liu GF, Liang HW, Zhang H. Study on the transformation of *Populous simonii×P.nigra* with salt resistance gene BET-A. Scientia Silvae Sinicae. 2001; 37: 34–38
- Jiang J, Chang YG, Dong JX, Wang ZY, Liu GF. Study on two insecticidal transgenic genes in *populous* simonii xP. nigra. Plant Physiology Communications. 2004; 40: 669–672
- Masterson J. Stomatal size in fossil plants: evidence for polyploid in majority of angiosperms. Science. 1994; 264:421–424 PMID: <u>17836906</u>
- 13. Wendel JF. Genome evolution in polyploids. Plant Mol Biol. 2000; 42:225–249 PMID: 10688139
- Li A, Hu BQ, Xue ZY, Chen L, Wang WX, Song WQ, et al. DNA methylation in genomes of several annual herbaceous and woody perennial plants of varying ploidy as detected by MSAP. Plant Mol Biol Rep. 2011; 29:784–793 PMID: <u>24415835</u>
- Kang XY. Advances in researches on polyploid breeding of forest trees. Journal of Beijing Forestry University. 2003; 25: 70–74

- Qi CL, Jin CL, Li KL, Li ZX, Zhao H. Comparison of photosynthetic characteristics and leaf anatomy structure of different ploidy *Populus ussuriensis* Kom. Plant Physiology Journal. 2010; 46:917–922
- Xing XT, Zhang ZY, Zhang WJ. Variations in wood thermal properties of triploid clones of *Populus* tomentosa. Journal of Beijing Forestry University. 2000; 22: 21–23
- Nilsson EH. über eine in der nature gefundene gigasform von Populus tremula. Hereditas 1936; 21:379–382
- Sylven N. Annual report on the work of the Association for Forest Tree Breeding during the year. Svensk PappTidn. 1943; 47: 38
- 20. Sarvas R. Two triploid aspen two triploid Birches. Commun Inst For Fenn. 1958; 49: 25
- 21. Johnsson H. Cytological studies of diploid and triploid *Populus tremula* and of crosses between them. Hereditas Lund. 1940; 26: 321–352
- Buijtenen J P, Joranson P N, Einspahr D W. Diploid versus triploid aspen as pulpwood sources with reference to growth, chemical, physical and pulping differences. Tappi. 1958; 41: 170–175
- Xi XJ, Li D, Xu WT, Guo LQ, Zhang JF, Li BL. 2n egg formation in populous euramericana (Dode) Guinier. Tree Genetics & Genomes.2012; 8: 1237–1245
- Ewald D, Ulrich K, Naujoks G, Schroder MB. Induction of tetraploid poplar and black locus plants using colchicines: chloroplast number as an early marker for selecting polyploids in vitro. Plant Cell Tiss Organ Cult.2009; 99: 353–357
- Zhang JE, Guo WW, Deng XX. Relationship between ploidy variation of citrus calli and competence for somatic embryogenesis. Acta Genetica Sinica. 2006; 33: 647–654 PMID: <u>16875323</u>
- Zhao XY, Ma KF, Zhang M, Bian JL, Jiao WY, Zhang ZY.Comparative analysis of the photosynthetic characteristics of three-year-old *Populus tomentosa* clones. Forest Research. 2011; 24:370–378
- Zhao XY, Wang JH, Zhang JF, Zhang SG, Zhao K, Jiao YD. Photosynthetic characteristics of Catalpa bungei clones. Acta Bot. Boreal. 2011; 31:1665–1671
- Hai PH, Jansson G, Harwood C, Thinh B, Huy H. Genetic variation in growth, stem straightness and branch thickness in clonal trials of Acacia auriculiformis at three contrasting sites in Vietnam. For Ecol Manag.2008; 255:156–167
- Hansen JK, Roulund H.Genetic parameters for spiral grain, stem form, Pilodyn and growth in 13 years old clones of Sitka Spruce (*Picea sitchensis* (Bong.) Carr.). Silvae Genet. 1996; 46: 107–113
- Pliura A, Zhang SY, Mackay J, Mackay J, Bousquet J. Genotypic variation in wood density and growth traits of poplar hybrids at four clonal trials. For Ecol Manag. 2007; 238:92–106
- Maherali H, Walden AE, Husband BC. Genome duplication and the evolution of physiological responses to water stress. New Phytol. 2009; 184: 721–731 doi: <u>10.1111/j.1469-8137.2009.02997.x</u> PMID: <u>19703115</u>
- Ramsey J. Polyploidy and ecological adaptation in wild yarrow. Proc Natl Acad Sci. 2011; 108: 7096–7101 doi: <u>10.1073/pnas.1016631108</u> PMID: <u>21402904</u>
- Zhu ZT, Lin HB, Kang XY. Studies on allotriploid breeding of *Populus tomentosa* B301 clones. Sci Silvae Sin. 1995; 31:499–505
- Comai L.The advantages and disadvantages of being polyploidy. Nature reviews genetics. 2005; 6: 836–846 PMID: <u>16304599</u>
- 35. Kien ND, Jansson G, Harwood C, Almqvist C, Thinh HH. Genetic variation in wood basic density and pilodyn penetration and their relationships with growth, stem straightness and branch size for *Eucalyptus urophylla* S. T. Blake in Northern Vietnam. NZ J For Sci. 2008; 38: 160–175
- **36.** Maniee M, Kahrizi D, Mohammadi R. Genetic variability of some morphophysiological traits in durum wheat (*Triticum turgidum* var. durum). Journal of Applied Sciences. 2009; 9: 1383–1387
- Zhou YM, Yang CP, Wang SJ, Wu YL, Wang WZ, Han SJ. A study on photosynthetic characteristics of Betula platyphylla. Journal of Forestry Research. 2002; 13: 209–212
- Surabhi G, Reddy R, Singh S. Photosynthesis, fluorescence shoot biomass and seed weight responses of three cowpea (*Vigna unguiculata* (L.) Walp) cultivars with contrasting sensitivity to UV-B radiation. Environmental and Experimental Botany. 2009; 66:160–171
- Fu YP, Yan HL, Li LF, Yu YH, Si HM, Hu GC, et al. Photosynthesis—related characteristics of different ploidy rice plants. Chinese J Rice Sci. 1999; 13:157–160
- 40. Ward JK, Strain BR. Elevated CO2 studies: past, present and future. Tree Physiol. 1999; 19:211–220 PMID: <u>12651563</u>
- Dippery JK, Tissue DT, Thomas RB, Strain BR. Effects of low and elevated CO2 and C3 and C4 annuals. I. growth and biomass allocation. Oecologia, 1995; 101:13–20

- **42.** Cowley R, Luckett D. Chlorophyll fluorescence as a method to detect moisture limiting stress in canola. 17th Australian Research Assembly on Brassicas (ARAB).2011.
- 43. Zhao XY, Wang JH, Zhang JF, Zhang SG, Zhang ZS, Ma JW, et al. Variation analysis on chlorophyll fluorescence and growth traits of *Catalpa bungei* clones. Journal of Beijing University. 2012; 34: 41–47 doi: <u>10.1093/jncimonographs/lgs001</u> PMID: <u>22623594</u>
- 44. Krause GH, Weis E. Chlorophyll fluorescence and photosynthesis: The basics. Annual Review of Plant Physiology and Plant Molecular Biology. 1991; 42: 313–349
- 45. Demming B, Bjorkman O. Comparison of the effect of excessive light on chlorophyll fluorescence (77K) and photon yield of O2 evolution in leaves of higher plants. Planta. 1987; 171:171–184 doi: <u>10.1007/</u> <u>BF00391092</u> PMID: 24227324
- 46. Zheng HQ, Lin SZ, Zhang Q, Lei Y, Hou L, Zhang ZY. Functional identification and regulation of the PtDrl02 gene promoter from triploid white poplar. Plant Cell Rep. 2010; 29: 449–460 doi: <u>10.1007/</u> s00299-010-0834-8 PMID: 20179934
- Ceulemans R, Impens U, Steenackers V. Variations in photosynthetic, anatomical, and enzymatic leaf traits and correlations with growth in recently selected *Populus* hybrids. Can J For Res. 1987; 17:273–283
- Ma KF, Song YP, Jiang XB, Zhang ZY, Li BL, Zhang DQ. Photosynthetic response to genome methylation affects the growth of Chinese white poplar. Tree Genetics & Genomes. 2012; 8: 1407–1421
- **49.** Bassman JH, Zwier JC. Gas exchange characteristics of *Populus trichocarpa*, *Populus deltoides* and *Populus trichocarpa* × *P*. *deltoides* clones. Tree Physiol.1991; 2:145–159 PMID: <u>14972886</u>