

A new mechanistic approach for the treatment of chronic neuropathic pain with nitrous oxide integrated from a systems biology narrative review

Baptiste Bessière¹, François Iris², Aude Milet¹, Athanasios Beopoulos², Catherine Billoet¹, Géraldine Farjot^{1*}

¹ Air Liquide Santé International, Paris Innovation Campus, Jouy-en-Josas, France

² Bio-Modeling System, Paris, France

*Correspondence to: Géraldine Farjot, PhD, geraldine.farjot@airliquide.com.

orcid: 0000-0003-0945-6365 (Géraldine Farjot)

Abstract

The limitations of the currently available treatments for chronic neuropathic pain highlight the need for safer and more effective alternatives. The authors carried out a focused review using a systems biology approach to integrate the complex mechanisms of nociception and neuropathic pain, and to decipher the effects of nitrous oxide (N₂O) on those pathways, beyond the known effect of N₂O on N-methyl-D-aspartate receptors. This review identified a number of potential mechanisms by which N₂O could impact the processes involved in peripheral and central sensitization. In the ascending pathway, the effects of N₂O include activating TWIK-related K⁺ channel 1 potassium channels on first-order neurons, blocking voltage-dependent calcium channels to attenuate neuronal excitability, attenuating postsynaptic glutamatergic receptor activation, and possibly blocking voltage-dependent sodium channels. In the descending pathway, N₂O induces the release of endogenous opioid ligands and stimulates norepinephrine release. In addition, N₂O may mediate epigenetic changes by inhibiting methionine synthase, a key enzyme involved in DNA and RNA methylation. This could explain why this short-acting analgesic has shown long-lasting anti-pain sensitization effects in animal models of chronic pain. These new hypotheses support the rationale for investigating N₂O, either alone or in combination with other analgesics, for the management of chronic neuropathic pain.

Key words: central nervous system; chronic peripheral neuropathic pain; CPNP; integrative biology; laughing gas; medical gas; N₂O; neuralgia; peripheral nervous system; receptors

doi: 10.4103/2045-9912.310058

How to cite this article: Bessière B, Iris F, Milet A, Beopoulos A, Billoet C, Farjot G. A new mechanistic approach for the treatment of chronic neuropathic pain with nitrous oxide integrated from a systems biology narrative review. *Med Gas Res.* 2021;11(1):34-41.

Funding: This work was supported by Air Liquide Santé International.

INTRODUCTION

The World Health Organization has declared chronic pain, defined as any continuous or recurrent pain that lasts for more than 3 months,¹ to be a leading world health problem.² A subcategory of chronic pain is chronic neuropathic pain, which is chronic pain “*caused by a lesion or disease of the somatosensory nervous system.*”¹ In a large population survey from the USA, the 2017 prevalence of chronic neuropathic pain was estimated to be 15.7% among all reported pain (95% confidence intervals 14.9–16.5),³ which is higher than the 7–8% prevalence of chronic pain with neuropathic characteristics reported by the International Association for the Study of Pain in 2014.⁴

The etiology of chronic neuropathic pain is varied, but common causes include diabetes, multiple sclerosis, shingles, spinal cord injury or spinal surgery, stroke, and trauma.^{1,5} In case of neuropathies, pain arises spontaneously, can be elicited by normally innocuous stimuli (allodynia), is exaggerated and prolonged in response to noxious stimuli (hyperalgesia), and may spread beyond the site of injury (secondary hyperalgesia).¹

The International Association for the Study of Pain recommends three types of treatment for first-line pharmacological management of chronic neuropathic pain: (1) secondary amine tricyclic antidepressants (TCA) or a dual serotonin noradrenaline reuptake inhibitor (SNRI); (2) calcium channel α 2- δ ligands (e.g., gabapentin, pregabalin); and (3) topical lidocaine for localized peripheral pain.⁶ Opioid

analgesics or tramadol are recommended first-line only for patients with acute neuropathic pain, neuropathic pain caused by malignancy, or for episodes of severe pain that occur as exacerbations of the chronic condition.⁶

This paper presents the results of a focused review using an analytical system biology approach to describe the biology of chronic neuropathic pain, taking into account the complex biological systems of pain signal transmission, pain perception, and central sensitization based on an integrative analysis of published data relevant to the mechanistic processes. With this understanding of the mechanical processes, a new theory has been formulated for the potential of nitrous oxide (N₂O) to be used in this indication. Here we summarize the main findings of this approach, focusing on how N₂O specifically may interact with the process of chronic neuropathic pain as a potential treatment.

METHODS

The authors carried out a review using a systems biology approach to integrate the complex mechanisms of nociception and neuropathic pain, and to decipher the effects of N₂O on those pathways, beyond the known effect of N₂O on N-methyl-D-aspartate receptors (NMDAR). The analytical procedure implemented (Computer-Assisted Deductive Integration) associates algorithmics and heuristics (**Additional file 1**). The logic behind this model-building approach does not assume functional linearity within biological systems and the com-



ponents of a model do not incorporate solely what is known. Indeed, since this approach relies upon strict and systematic implementation of negative selection of hypotheses, models arising from this procedure contain elements that have never been described but cannot be refuted by current knowledge and/or available biological data, thereby generating novel understanding. This model-building approach has proven its efficacy in a number of biological research domains, including the discovery of hitherto unsuspected biological mechanisms, pathways, and interactions directly associated with phenotypic transitions *in vivo* (be they pathological or developmental).⁷⁻¹⁵ Further information on the Computer-Assisted Deductive Integration method can be found in a review from Iris et al.¹⁶

CATEGORIES AND MECHANISMS OF CHRONIC NEUROPATHIC PAIN

Neuropathic pain can be described as burning, stabbing, shooting, aching, or like an electric shock, but patients' reports of pain vary in their description, character and intensity, all of which may be used to variables for classification.¹⁶ However, such a classification rarely assists physicians in selecting the most appropriate therapy in chronic neuropathic pain conditions, especially when the underlying etiology of the pain (e.g., diabetes, traumatic nerve lesion) cannot be cured.¹⁷ Instead, classification based on the changes to the peripheral and central nervous systems that lead to the generation of abnormal pain sensitivity can provide mechanistic explanations for many of the temporal, spatial and threshold changes in pain sensibility in acute and chronic clinical pain settings.¹⁷

Primary sensory neurons usually have a high threshold to stimulation so that only noxious stimuli will activate them. However, when tissue is damaged and these neurons are exposed to inflammatory mediators, their excitation threshold is lowered, increasing the responsiveness to nociceptive input.¹⁷ This process is termed "peripheral sensitization" and it contributes to primary hyperalgesia at sites of inflammation.¹⁸ Because the nociception changes occur at the site of tissue injury, ongoing peripheral pathology is required to maintain sensitization.¹⁹ It has been shown that peripheral sensitization is implicated in the pathogenesis of altered temperature sensitivity, but not mechanical sensitivity.²⁰

Central sensitization produces pain hypersensitivity in non-inflamed tissue by changing the sensory response elicited by normal inputs and increasing pain sensitivity long after the initial cause has disappeared or when no cause can be identified.²¹ The molecular mechanisms responsible for central and peripheral sensitization are markedly different.²¹ In central sensitization, neurons and circuits in nociceptive pathways throughout the central nervous system show increased membrane excitability, synaptic efficiency, or reduced inhibition.²¹ These functional changes alter sensory inputs from the periphery causing the central nervous system to generate pain sensations in the absence of any particular peripheral stimuli.

KNOWN MECHANISMS OF NEUROPATHIC PAIN

Pain transmission pathway

The pain sensation occurs when a noxious mechanical, thermal or chemical stimulus is intense or prolonged enough to trigger

an action potential in the usually resting peripheral nociceptors.²² Nociceptor afferents have a unique structure, with both peripheral and central terminals utilizing the same biochemical processes, such that they can send and receive signals from either terminal. In peripheral tissues, the terminal nerve endings have a high threshold and do not transmit signals unless the stimulus presents a threat to physiological homeostasis.²² Triggering the receptor sends an action potential through the afferent neuronal network, composed of unmyelinated C-fibers, and the myelinated A δ and A β -fibers.²² C-fibers respond to mechanical, thermal and chemical stimuli, but transmit signals more slowly than A-fibers, and are responsible for prolonged pain sensations such dull aches or a burning sensation.²² A δ -fibers are activated by noxious mechanical and thermal stimuli, and result in rapid high-frequency firing that is interpreted by the central nervous system as sharp localized pain.²² A β -fibers usually carry input from innocuous stimuli like vibration,²³ but they can also respond to mechanical or thermal stimulation in the noxious range.²² All of these neurons project to specific lamina of the dorsal horn of the spinal cord where they synapse with central projection neurons via second-order neurons.²²

Nociceptive neurons release various excitatory neurotransmitters: C-fibers use neuropeptides (mainly substance P) and are therefore referred to as peptidergic, while glutamate is the key transmitter for A δ -fibers.²⁴ Substance P interacts with G-coupled neurokinin-1 receptors on second-order neurons, activating a complex intracellular signaling cascade that results in nitric oxide production, and activation of both arachidonic acid and NMDAR pathways.²⁴ Glutamate, on the other hand, acts on second-order neurons via the ionotropic α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor.²⁴ Other excitatory neurotransmitters in the dorsal horn include aspartate, vasoactive intestinal peptide, somatostatin, and calcitonin gene-related peptide.²²

Within the dorsal horn, the incoming nociceptive signals are modulated by a complex network of central projection neurons descending from the brainstem,²² and interneurons utilizing a range of endogenous molecules. The interneurons utilize mainly γ -aminobutyric acid (GABA) and glycine as inhibitory neurotransmitters, while the descending pain modulators use norepinephrine, serotonin, and opioid ligands.^{22,24,25} Besides inhibiting excitatory neurons and terminals, noradrenergic and serotonergic fibers excite GABAergic and glycinergic interneurons. Norepinephrine depolarizes GABAergic neurons by activating α 1-adrenoceptors (α 1-AR),²⁶ while serotonin increases the frequency of GABAergic activity by activating 5-hydroxytryptamine 3 (5-HT₃) receptors.²⁷

Peripheral sensitization

As described earlier, peripheral sensitization may occur after nerve damage whereby action potentials in the normally high-threshold peripheral terminals are triggered at a lower stimulus threshold than usual. The increased electrical excitability of these neurons is mediated by upregulation in the expression of voltage-gated sodium channels.²⁸ Clusters of sodium channels accumulate not only at the site of the nerve damage, but also more centrally in the undamaged dorsal root ganglion.²⁸ This hyperactivity is exacerbated by downregulation of potassium

channels such as TWIK-related K⁺ channel 1 (TREK1), which would normally inhibit action potential propagation.²⁹ TREK1 is a potassium channel with a central role in anesthesia, neuroprotection, pain perception and depression. The complex gating properties of TREK1 are modulated by cell volume, actin cytoskeleton dynamics, cellular lipids, membrane-receptor-coupled second messengers and pharmacological agents.³⁰⁻³³ When TREK1 is downregulated, the ongoing afferent traffic initiated by the injury leads to the activation of spinal facilitatory cascades, yielding a greater output to the brain for any given input.

The sympathetic nervous system may also be involved in the generation of pain and other changes associated with pain states following trauma, ischemia, or deep-tissue microvascular vasomotor disturbances (complex regional pain syndrome-type 1). Nerve injury triggers the expression of functional α 1- and α 2-ARs on cutaneous afferent fibers. In addition, the abnormal spontaneous activity at somatosensory nerve terminals triggers the sprouting of sympathetic neurons in the dorsal root ganglion.³⁴ Postganglionic sympathetic neurons may activate receptors on the already sensitized primary afferent nerve terminals and cell bodies, either directly (on synapses via neurotransmitters such as norepinephrine and adenosine triphosphate) or indirectly (through secretion of norepinephrine into the blood); such activation may exacerbate the perception of the neuropathic pain. This magnifies the excitatory input in the dorsal root ganglion,³⁴ and is called sympathetically maintained pain. Sympathetically maintained pain is sensitive to treatments that target norepinephrine-mediated pain such as TCAs or clonidine.³⁵

Central sensitization

Central sensitization develops when the central ascending nociceptive pathway becomes more excitable or there is a reduction in the inhibitory effects of the descending pathway.²¹ Spontaneous activity within dorsal horn neurons develops or increases and the activation threshold of these neurons is reduced.²¹ The result is that patients may experience pain in response to innocuous stimuli as well as noxious ones (allodynia), patients feel pain over a wider spatial area than that normally triggered by a stimulus (secondary hyperalgesia), or pain is persistent even after removal of the initial stimulus (sensation of “afterpain”).²¹

The development and maintenance of central sensitization is dependent on activation of glutamatergic receptors, and notably NMDARs.²¹ Normally, NMDARs are maintained in a quiescent state by the presence of magnesium ions. With sustained nociceptive input, this usual voltage-dependent “brake” on Ca²⁺ influx is removed, and these ions flow into the cell through the NMDAR pore.²¹ This triggers intracellular signaling pathways including activation of Ca²⁺-dependent kinases, such as Ca²⁺/calmodulin-dependent protein kinase II, phosphatases (e.g., calcineurin) and nitric oxide synthase.²¹ In turn, these intracellular processes change the threshold and kinetics of the NMDARs and AMPA receptors, making the neurons more excitable, and promote expression of AMPA receptors on the synaptic surface.²¹ Nitric oxide contributes to the increased excitability and reduces inhibitory activity within the dorsal horn neurons.²¹ The disinhibited neurons

become more susceptible to afferent inputs, including those from the normally non-nociceptive A β -fibers.²¹ The C-fibers in the dorsal horn release the pain-promoting neuropeptide substance P that binds to neurokinin-1 receptor,³⁶ and accelerates the Ca²⁺ influx through the NMDAR. Therefore, an efficient strategy to attenuate or block facilitation may be to reduce signal transduction, Ca²⁺ currents and/or NMDAR activation.

CURRENT TREATMENTS FOR CHRONIC NEUROPATHIC PAIN

Because of the multiple receptors and neuromediators involved with the activation, amplification, and inhibition of nociception, analgesic agents can act at different sites within the pathway (Figure 1).³⁷ Sympathetic blockers (e.g., clonidine) block presynaptic α 1-AR on afferent C-fibers, blocking the release of glutamate and substance P.²⁴ Anticonvulsants (carbamazepine, gabapentin, pregabalin, topiramate) have a range of effects including potentiating the activity of GABA, inhibiting sodium and calcium channels, and inhibiting NMDARs and AMPA receptors.³⁸ Dextromethorphan blocks NMDARs in the spinal cord.²¹ TCAs, dual SNRIs inhibit serotonin and norepinephrine reuptake, enhancing their effect in endogenous pain-inhibitory systems in the brain.³⁵ Tramadol hydrochloride is a relatively weak central opioid analgesic, but it also acts as an inhibitor of serotonin and norepinephrine reuptake.⁶

There is strong evidence for the use of antidepressants

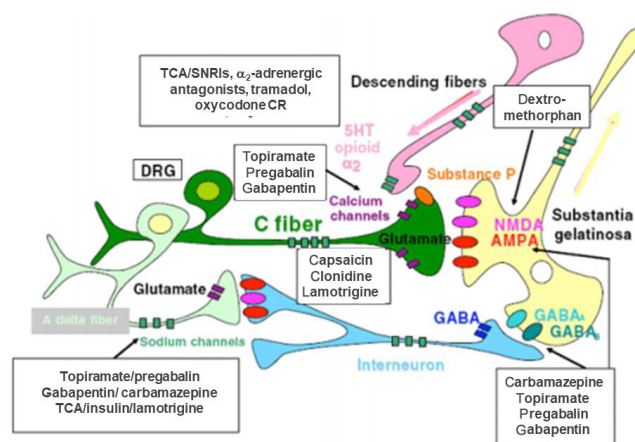


Figure 1: The mechanisms of pharmacologic therapies for chronic neuropathic pain.

Note: 5-HT: 5-Hydroxytryptamine; AMPA: α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid class of glutamate receptor; CR: controlled release; DRG: dorsal root ganglia; GABA: γ -aminobutyric acid; NMDA: N-methyl-D-aspartate class of glutamate receptor; SNRI: serotonin-norepinephrine reuptake inhibitor; TCA: tricyclic antidepressant. Reproduced with permission from Vinik and Mehrabyan.³⁸

in clinical practice, in particular those that act on both norepinephrine and serotonin such as SNRIs and TCAs, for the treatment of neuropathic pain, including post-herpetic neuralgia, painful polyneuropathy, diabetic neuropathy, spinal cord injury, and multiple sclerosis.³⁹⁻⁴¹ However, they have not proven to be particularly effective in human immunodeficiency virus 1 neuropathy, phantom limb pain, or cancer related neuropathic pain.⁶ Since chronic pain and depression often coexist,⁴² the antidepressant effects of these agents may be an additional benefit.⁶



Gabapentin, pregabalin and carbamazepine are anticonvulsants indicated for the treatment of chronic neuropathic pain. Pregabalin is recommended by the National Institute for Health and Care Evidence as first-line treatment for patients with neuropathic pain.⁴³ However, the efficacy of these agents in neuropathic pain conditions is mixed.⁴⁴⁻⁴⁶

The use of opioids for chronic pain conditions of any etiology is increasing,⁴⁷ despite their known potential for adverse events such as paradoxical pain, abuse and addiction.^{48,49} Because of inter-individual variability in response to opioids,⁵⁰ patients may receive higher-than-needed doses for the treatment of pain, increasing the risk of addiction, sleep apnea, respiratory depression and opioid-associated deaths.^{51,52} Therefore, long-term opioid therapy is not suitable for most individuals with chronic neuropathic pain not related to cancer.⁵³ Opioids are only recommended as a second-line option when a combination of first-line agents is not optimally effective or feasible.^{6,53}

The limitations of the currently available treatments for chronic neuropathic pain highlight the need for safer and more effective alternatives of chronic pain control.

NEW HYPOTHETICAL MECHANISMS OF N₂O ON THE PAIN PATHWAY

N₂O is traditionally used as a routine add-on to general anesthesia and for painful procedures.⁵⁴ Research in the last decade has found that N₂O acts on one or more super-families of ligand-gated ion channels⁵⁵ causing multilevel blockade of the ascending and descending pathways, which effectively closes the gate to nociceptive inputs. As an NMDAR antagonist,^{56,57} it produces moderate analgesia at sub-anesthetic concentrations and reduces surgical pain with a substantial neuropathic component.⁵⁸ N₂O also facilitates the release of opioid peptides in the periaqueductal brainstem, activating the descending noradrenergic inhibitory pathways via GABA, thus reducing nociception at the spinal cord level.⁵⁹

In addition to its acute effect, a single exposure facilitates post-injury pain rehabilitation and long-lastingly ameliorates severe neuropathic pain in rats, with the effect lasting for at least a month.⁶⁰ Therefore, the systems biology approach was used to identify possible rational mechanisms of action by which N₂O may be able to interrupt the development of peripheral or central sensitization, and positively impact the course of chronic neuropathic pain.

Effects on the ascending pathway

In common with the inhalational anesthetic agents (chloroform, diethyl ether, halothane and isoflurane), N₂O activates TREK1 (potassium channel subfamily K member 2) in first-order neurons, leading to hyperpolarization of these presynaptic neurons and decreased release of glutamate (**Figure 2**).^{61,62} As described earlier, part of the peripheral sensitization mechanism involves downregulation of TREK1 channels, which dampens the pain inhibitory mechanisms in the dorsal horn lamina.³⁰ N₂O may counteract this mechanism by activating TREK1.

N₂O is also known to block CaV3.2 (T-type) voltage-dependent calcium channels via free radicals.⁶³ This effect allows N₂O to effectively attenuate neuronal excitability in nociceptive pathways,⁶⁴ while concomitantly reinforcing the

effects of TREK1 opening by attenuating pre-synaptic glutamate release (**Figure 2**).⁶⁵

Another effect of N₂O is to attenuate postsynaptic glutamatergic AMPA, kainate and NMDA receptor activation.^{55,56,66} As described earlier, hyperpolarization of postsynaptic receptors removes the voltage-dependent Mg²⁺ block of the NMDAR, causing increased neuronal excitability, but N₂O antagonizes this hyperpolarization, thereby maintaining NMDARs in a non-activated state.⁶⁵

Voltage-dependent sodium channels are responsible for the generation of action potentials and transmission in excitable cells. Blockade of sodium channels leads to the abolition of synaptic transmission and network function.⁶⁷ To date, there have been no reports of N₂O acting on voltage-dependent sodium channels, but an inhibitory effect is likely, given that all major volatile general anesthetic agents (halothane, isoflurane, enflurane, ether and ethanol) have an antagonistic effect upon these channels.⁶⁸ N₂O may affect serotonin-activated sodium/calcium channels by two mechanisms. First, N₂O is reported to activate α 2-ARs⁶⁹⁻⁷¹ while inhibiting β 2-subunit-containing nicotinic acetylcholine channels,⁷² which produces short-term inhibition of nociception mediated by muscarinic acetylcholine receptors,^{73,74} and concurrently potentiating GABA_A and glycine receptors.^{75,76}

Second, N₂O attenuates 5-HT₃ receptor signaling,^{75,77} thus promoting the postsynaptic inhibitory effects of GABA_A receptor activation while minimally affecting GABA and glycine release which is also positively controlled via presynaptic α 2-ARs and muscarinic and nicotinic acetylcholine receptors.⁷⁸⁻⁸⁰

Two types of GABA receptors may be affected by N₂O: GABA_A receptors, which are ligand-gated chloride channels and thus responsible for fast inhibition of neurotransmission,⁸¹ and GABA_B receptors,⁸² which are G-protein coupled receptors that mainly influence potassium and calcium channels through second messenger systems, and result in a slower decrease of neurotransmission than that mediated by GABA_A receptors.⁸² By potentiating GABA_A and glycine receptors,⁷⁶ N₂O appears to allow hyperpolarization (firing inhibition) of second order neurons that would otherwise favor depolarization, while concurrently favoring inhibitory depolarization in primary sensory neurons, thus attenuating nociception at two levels in the ascending pathway (**Figure 2**).⁸³

Effects of N₂O on the descending pathway

Centrally, N₂O exerts an analgesic effect by inducing the release of endogenous ligands of κ opioid receptors.⁸³ Activation of the opioid receptors by these endogenous ligands counteracts the tonic inhibition of descending noradrenergic pathways which is usually maintained by inhibitory GABAergic neurons.^{70,84} In addition, by stimulating norepinephrine release,⁸⁵ N₂O enhances the antinociceptive synergism between the α 2-AR and the δ opioid receptor (DOR) (**Figure 2**).⁸⁶

Epigenetic effects

There is growing evidence that peripheral nerve injury causes epigenetic changes in pain-related pathways, and that these changes are implicated in the pathogenesis of chronic neuropathic pain.^{87,88} By the same logic, inducing epigenetic changes

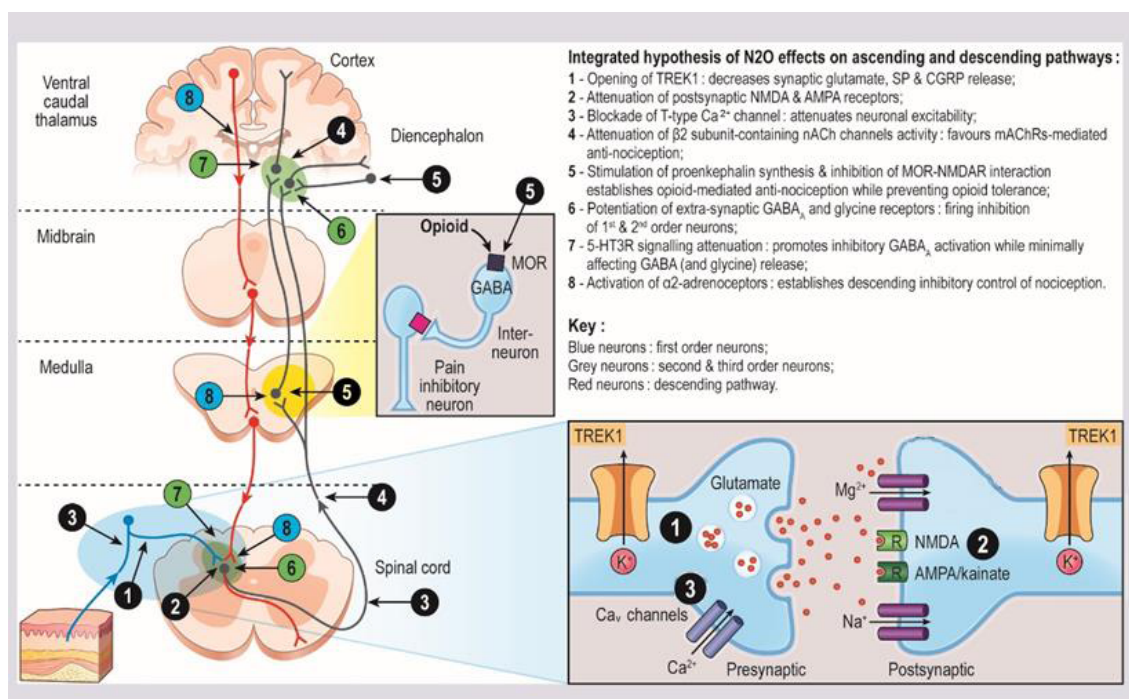


Figure 2: Hypothetical mechanisms of action of nitrous oxide (N₂O) in chronic neuropathic pain.

Note: Potential actions in the ascending pathway are: (1) opening of TWIK-related K⁺ channel 1 (TREK1) in first order neurons, leading to reduced synaptic release of glutamate, substance P (SP), and calcitonin gene-related peptide (CGRP); (2) attenuation of postsynaptic N-methyl-D-aspartate (NMDA) and α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors, causing inhibition of second order neurons; (3) blockade of T-type Ca²⁺ channel in first and second order neurons, which would attenuate neuronal excitability; (4) attenuation of β2 subunit-containing nicotinic acetyl choline (nACh) channel activity in second order neurons, which favors muscarinic acetyl choline receptor (mAChR)-mediated antinociception; (5) stimulation of proenkephalin and inhibition of the interaction between μ-opioid receptors (MORs) and NMDA receptors (NMDARs) to establish opioid-mediated antinociception while preventing tolerance; (6) potentiation of extrasynaptic γ-aminobutyric acid (GABA)_A receptors and glycine receptors in first and second order neurons, which would inhibit the firing of these neurons; (7) attenuation of 5-hydroxytryptamine 3 (5-HT₃) receptor signalling to promote inhibitory GABA_A activation while minimally affecting GABA (and glycine) release. In the descending pathway, the potential actions of N₂O are: (8) activation of α2-adrenoceptors, which would establish descending inhibitory control of nociception, concurrently with stimulation of norepinephrine release (favoring the synergistic antinociceptive effects of δ-opioid receptors and adenosine 2A receptors) and with attenuation of 5-HT₃ signaling (promoting the postsynaptic inhibitory effects of GABA_A and preventing serotonin-mediated facilitatory effects).

may be a mechanism by which short-term treatment could induce long-lasting effects on pain regulation.

Epigenetic mechanisms regulate the transcription of rate-limiting metabolic enzymes, and are mediated by chemical modifications to chromatin, which modulate gene expression without changing the DNA sequence.⁸⁹ N₂O causes the oxidation of vitamin B12 (cobalamin), preventing methyl transfer from the folate cycle,⁹⁰ and inhibiting the B12-dependent enzyme methionine synthase.⁸⁹ This causes a depletion of S-adenosylmethionine (Figure 3).⁸⁸ S-adenosylmethionine is the main cofactor for DNA methylation now considered to be a key mechanism for regulating gene expression in neurons.⁹¹

N₂O may mediate epigenetic changes via the opioid pathway. For example, when S-adenosylmethionine is downregulated, the downstream transcriptional effect is to increase production of δ- and κ-opioid ligands (principally met-enkephalin) at the expense of μ-receptor peptides (β-endorphin) and nociceptin.⁹² A reduction in DNA methylation may also increase the expression of DOR and μ-opioid receptors (MOR), but does not affect κ opioid receptor expression, which is regulated by histone acetylation.⁹³

N₂O-induced epigenetic regulation of μ-peptides and MOR expression should decrease the NMDAR-MOR association and sensitization induced by NMDAR activation.⁹⁴ This is reminiscent of the inhibition of morphine tolerance by administration of NMDAR antagonists, such as ketamine or

dextromethorphan.⁹³ Apart from breaking the opioid tolerogenic MOR-NMDAR interaction, an epigenetically mediated increase in the ratio of DOR ligand expression (e.g., met-enkephalin) would also favour a DOR-α2AR interaction,⁹⁵ which may promote long-lasting anti-nociceptive synergistic effects of N₂O.

DISCUSSION

The results of the current review provide a rational explanation for a potential therapeutic benefit of N₂O in patients with chronic neuropathic pain, and this is supported by data from animal studies suggesting that N₂O has short- and long-lasting analgesic effects.⁹⁶ Usage of N₂O in acute analgesia is well established in clinical practice, however its secondary effects linked to B12 inactivation prevents a daily repeated usage. The long lasting analgesic effect, may overcome this shortage but needs to be proven in clinical trials on established chronic neuropathic pain. However, to date, only one clinical study with N₂O has been undertaken in patients with chronic neuropathic-type low back pain, and this study did not show a significant benefit compared with oxygen in a small cohort of patients (*n* = 70).⁹⁷ One possible explanation is linked to the different etiology of low back pain that has an important inflammatory component, being released by anti-inflammatory medications, contrary to chronic neuropathic pain.

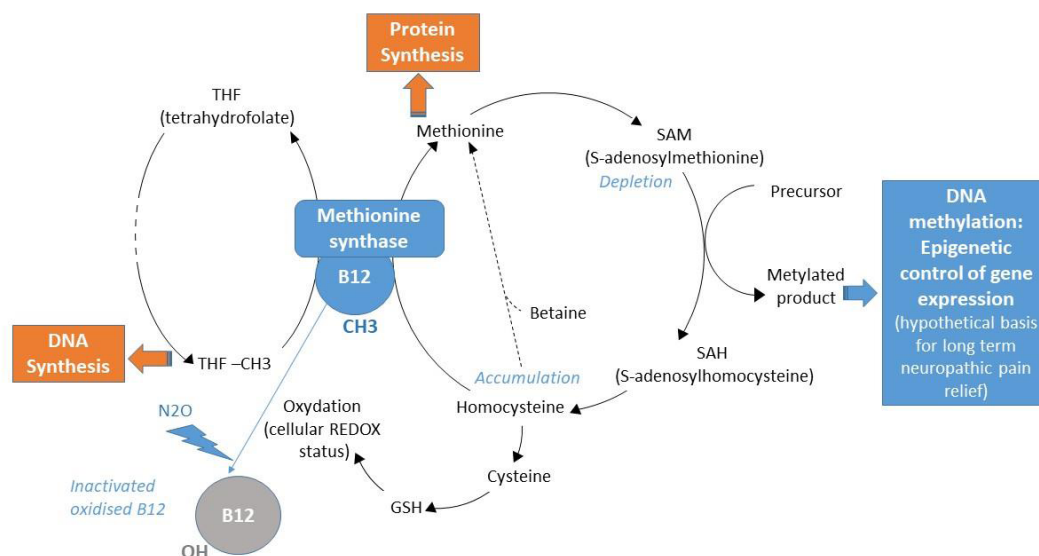


Figure 3: Hypothetical link between oxidation of vitamin B12 by N₂O and epigenetic regulation as a basis for long-lasting effects on pain regulation.

Note: Prevention of methyl transfer from the folate cycle by B12-dependent methionine synthase induces depletion in SAM, the main cofactor for DNA methylation, which may induce long-term effects on neuropathic pain. The hypothesis placing methionine as an integrator of reduction-oxidation status regulating transmethylation between tetrahydrofolate and SAM cycles was deduced from Trivedi et al.⁸⁹ N₂O: Nitrous oxide; SAM: S-adenosylmethionine.

As with many other pain conditions, multimodal therapy is often required for patients with chronic neuropathic pain, and N₂O may have a role as adjunctive therapy, enhancing the analgesic effects of other agents. N₂O mediates activation of the anti-nociceptive descending pathway, together with attenuation of MOR-NMDAR interaction and concurrent potentiation of extrasynaptic GABA_A and glycine receptors, α 2-ARs, noradrenaline release, pro-enkephalin production and DOR- α 2AR interaction.^{68,83} Therefore, pre- or co-medication with N₂O could allow a reduction in the therapeutic doses of other analgesic agents, limiting the occurrence of undesirable adverse events, without compromising therapeutic efficacy. For example, N₂O uncouples the MOR-NMDAR interaction, which would help to re-sensitize MORs and limit opioid tolerance. By activating the α 2-ARs, N₂O might allow for a lower dose of opioid to be used, while the attenuating effects of N₂O on 5-HT₃ signaling might also facilitate a reduced dose of SNRIs.

N₂O might also have a role as an adjunct to anticonvulsant therapy for the treatment of chronic neuropathic pain. Anticonvulsants often need to be used in high doses to achieve analgesic effects in chronic neuropathic pain, placing patients at risk of unwanted adverse events.⁹⁸ Since both N₂O and anticonvulsants potentiate GABAergic nociceptive pathways, using the two in combination might allow a lower dose of anticonvulsant to be used. However, caution should be exercised since the negative effects of N₂O on vitamin B12-dependent mechanisms of erythropoiesis could increase the risk of hematologic adverse events with some anticonvulsants such as carbamazepine.^{99,100}

SUMMARY AND CONCLUSIONS

Chronic pain is a leading health problem, affecting a large number of people worldwide.² Moreover, the pathophysiological processes of chronic pain development are complex,

and reflect the plasticity of the peripheral and central nervous systems.³⁶ Because no single mechanism is responsible for chronic pain, treatment is also complex and usually involves multimodal therapy targeting more than one nociceptive pathway. Chronic pain management requires a thorough assessment of a patient's pain condition as well as any underlying causes.⁶ Current pharmacologic management strategies for chronic pain are often limited in maintaining long-term pain control and are all associated with potential issues, such as adverse events, dependence or tolerance.

The current algorithmic and heuristic analysis was designed to identify a potential interface between the mechanism of chronic pain and the pharmacological effects of N₂O in areas of the brain and spinal cord where chronic pain originates. The findings suggest that there are supportive reasons for N₂O to have beneficial effects in the treatment of chronic neuropathic pain, not only via targeting NMDA receptor. Ongoing research is exploring the clinical potential for N₂O in this setting.

Acknowledgements

We would like to thank Catherine Rees and Mimi Chan, PhD, of Springer Healthcare Communications, for medical writing assistance. We thank Ira Katz, of Air Liquide Santé International, for critical proof reading.

Author contributions

GF conceived the project and proof outline. BB and AM brought their expertise in preclinical neuroscience and pain. CB ensured the medical reliability of the content with her experience of clinical pain treatment. GF, BB, AM, CB equally contributed to the final version of the manuscript. FI and AB performed the systems biology computation and elaborated the mechanistic theories. All authors approved the final version of the manuscript for publication.

Conflicts of interest

BB, AM, CB and GF are employees of Air Liquide Santé International (ALSI). ALSI is currently marketing N₂O in analgesia, as an add-on to general anesthetic agents and for the prevention of procedural pain in a fixed 50% mixture with O₂. ALSI is also conducting a clinical trial (EudraCT #: 2015-004779-64) with N₂O in chronic neuropathic pain. FI and AB are employees of Bio-Modeling Systems, funded



by Air Liquide Santé International for this system biology analysis.

Financial support

This work was supported by Air Liquide Santé International.

Copyright license agreement

The Copyright License Agreement has been signed by all authors before publication.

Plagiarism check

Checked twice by iThenticate.

Peer review

Externally peer reviewed.

Open access statement

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

Additional file

Additional file 1: Computer-Assisted Deductive Integration method.

REFERENCES

- Treede RD, Rief W, Barke A, et al. A classification of chronic pain for ICD-11. *Pain*. 2015;156:1003-1007.
- World Health Organization. World health organization supports global effort to relieve chronic pain. Geneva: 2004.
- DiBonaventura MD, Sadosky A, Concialdi K, et al. The prevalence of probable neuropathic pain in the US: results from a multimodal general-population health survey. *J Pain Res*. 2017;10:2525-2538.
- International Association for the Study of Pain. Epidemiology of neuropathic pain: How common is neuropathic pain, and what is its impact? Global Year Against Neuropathic Pain, 2014-2015. 2014.
- Moulin DE, Clark AJ, Gilron I, et al. Pharmacological management of chronic neuropathic pain - consensus statement and guidelines from the Canadian Pain Society. *Pain Res Manag*. 2007;12:13-21.
- Dworkin RH, O'Connor AB, Backonja M, et al. Pharmacologic management of neuropathic pain: evidence-based recommendations. *Pain*. 2007;132:237-251.
- Gadal F, Bozic C, Pillot-Brochet C, et al. Integrated transcriptome analysis of the cellular mechanisms associated with Ha-ras-dependent malignant transformation of the human breast epithelial MCF7 cell line. *Nucleic Acids Res*. 2003;31:5789-5804.
- Gadal F, Starzec A, Bozic C, et al. Integrative analysis of gene expression patterns predicts specific modulations of defined cell functions by estrogen and tamoxifen in MCF7 breast cancer cells. *J Mol Endocrinol*. 2005;34:61-75.
- Iris F. Psychiatric systems medicine: closer at hand than anticipated but not with the expected portrait. *Pharmacopsychiatry*. 2012;45 Suppl 1:S12-21.
- Iris F, Filiou M, Turck CW. Differential proteomics analyses reveal anxiety-associated molecular and cellular mechanisms in cingulate cortex synapses. *J Neuropsychiatry Clin Neurosci*. 2014;2:25-42.
- Iris F, Gea M, Lampe PH, Querleux B. Heuristic modelling applied to epidermal homeostasis. In: Querleux B, ed. *Computational biophysics of the skin*. New York, NY: Pan Stanford. 2015.
- Iris F, Gea M, Lampe PH, Santamaria P. Production and implementation of predictive biological models. *Med Sci (Paris)*. 2009;25:608-616.
- Nussbaumer M, Asara JM, Teplýtska L, et al. Selective mitochondrial targeting exerts anxiolytic effects in vivo. *Neuropsychopharmacology*. 2016;41:1751-1758.
- Pouillot F, Blois H, Iris F. Genetically engineered virulent phage banks in the detection and control of emergent pathogenic bacteria. *Biosecur Bioterror*. 2010;8:155-169.
- Turck CW, Iris F. Proteome-based pathway modelling of psychiatric disorders. *Pharmacopsychiatry*. 2011;44 Suppl 1:S54-61.
- Iris F, Beopoulos A, Gea M. How scientific literature analysis yields innovative therapeutic hypothesis through integrative iterations. *Curr Opin Pharmacol*. 2018;18:67-70.
- Woessner J. The classification of pain. In: Boswell MV, Cole BE, eds. *Wiener's pain management: A practical guide for clinicians*. 7th ed. Bosa Roca, FL: CRC Press; 2005:35-47.
- Baron R. Mechanisms of disease: neuropathic pain--a clinical perspective. *Nat Clin Pract Neurol*. 2006;2:95-106.
- Woolf CJ, Ma Q. Nociceptors--noxious stimulus detectors. *Neuron*. 2007;55:353-364.
- Woolf CJ. The pathophysiology of peripheral neuropathic pain--abnormal peripheral input and abnormal central processing. *Acta Neurochir Suppl (Wien)*. 1993;58:125-130.
- Ji RR, Samad TA, Jin SX, Schmoll R, Woolf CJ. p38 MAPK activation by NGF in primary sensory neurons after inflammation increases TRPV1 levels and maintains heat hyperalgesia. *Neuron*. 2002;36:57-68.
- Latremoliere A, Woolf CJ. Central sensitization: a generator of pain hypersensitivity by central neural plasticity. *J Pain*. 2009;10:895-926.
- Vardeh D, Naranjo JF. Anatomy and physiology: mechanisms of nociceptive transmission. In: Yong RJ, Nguyen M, Nelson E, Urman RD, eds. *Pain Medicine: An Essential Review*. Cham: Springer International Publishing; 2017:3-5.
- Neumann S, Braz JM, Skinner K, Llewellyn-Smith IJ, Basbaum AI. Innocuous, not noxious, input activates PKCgamma interneurons of the spinal dorsal horn via myelinated afferent fibers. *J Neurosci*. 2008;28:7936-7944.
- Schliessbach J, Maurer K. Pharmacology of pain transmission and modulation. In: Yong RJ, Nguyen M, Nelson E, Urman RD, eds. *Pain medicine: An essential review*. Cham, Switzerland: Springer; 2017:7-9.
- Kohno T, Kumamoto E, Higashi H, Shimoji K, Yoshimura M. Actions of opioids on excitatory and inhibitory transmission in substantia gelatinosa of adult rat spinal cord. *J Physiol*. 1999;518 (Pt 3):803-813.
- Gassner M, Ruscheweyh R, Sandkühler J. Direct excitation of spinal GABAergic interneurons by noradrenaline. *Pain*. 2009;145:204-210.
- Abe K, Kato G, Katafuchi T, Tamae A, Furue H, Yoshimura M. Responses to 5-HT in morphologically identified neurons in the rat substantia gelatinosa in vitro. *Neuroscience*. 2009;159:316-324.
- Wood JN, Boorman JP, Okuse K, Baker MD. Voltage-gated sodium channels and pain pathways. *J Neurobiol*. 2004;61:55-71.
- Mathie A, Veale EL. Two-pore domain potassium channels: potential therapeutic targets for the treatment of pain. *Pflugers Arch*. 2015;467:931-943.
- Chemin J, Patel AJ, Duprat F, Lauritzen I, Lazdunski M, Honoré E. A phospholipid sensor controls mechanogating of the K⁺ channel TREK-1. *EMBO J*. 2005;24:44-53.
- Fink M, Duprat F, Lesage F, et al. Cloning, functional expression and brain localization of a novel unconventional outward rectifier K⁺ channel. *EMBO J*. 1996;15:6854-6862.
- Maingret F, Lauritzen I, Patel AJ, et al. TREK-1 is a heat-activated background K(+) channel. *EMBO J*. 2000;19:2483-2491.
- Patel AJ, Honoré E, Maingret F, et al. A mammalian two pore domain mechano-gated S-like K⁺ channel. *EMBO J*. 1998;17:4283-4290.
- Chen SS, Zhang JM. Progress in sympathetically mediated pathological pain. *J Anesth Perioper Med*. 2015;2:216-225.
- D'Mello R, Dickenson AH. Spinal cord mechanisms of pain. *Br J Anaesth*. 2008;101:8-16.
- De Felipe C, Herrero JF, O'Brien JA, et al. Altered nociception, analgesia and aggression in mice lacking the receptor for substance P. *Nature*. 1998;392:394-397.
- Vinik AI, Mehrabyan A. Diabetic neuropathies. *Med Clin North Am*. 2004;88:947-999, xi.
- Attal N, Cruccu G, Haanpää M, et al. EFNS guidelines on pharmacological treatment of neuropathic pain. *Eur J Neurol*. 2006;13:1153-1169.
- Verdu B, Decosterd I, Buclin T, Stiefel F, Berner A. Antidepressants for the treatment of chronic pain. *Drugs*. 2008;68:2611-2632.
- Arnold LM. Management of psychiatric comorbidity in fibromyalgia. *Curr Psychiatry Rep*. 2006;8:241-245.
- Ohayon MM, Schatzberg AF. Using chronic pain to predict depressive morbidity in the general population. *Arch Gen Psychiatry*. 2003;60:39-47.
- Tan T, Barry P, Reken S, Baker M. Pharmacological management of neuropathic pain in non-specialist settings: summary of NICE guidance. *BMJ*. 2010;340:c1079.
- Finnerup NB, Attal N, Haroutounian S, et al. Pharmacotherapy for neuropathic pain in adults: a systematic review and meta-analysis. *Lancet Neurol*. 2015;14:162-173.
- Griebeler ML, Morey-Vargas OL, Brito JP, et al. Pharmacologic interventions for painful diabetic neuropathy: An umbrella systematic review and comparative effectiveness network meta-analysis. *Ann Intern Med*. 2014;161:639-649.
- Wiffen PJ, Derry S, Bell RF, et al. Gabapentin for chronic neuropathic pain in adults. *Cochrane Database Syst Rev*. 2017;6:CD007938.
- Zutler M, Holty JE. Opioids, sleep, and sleep-disordered breathing. *Curr Pharm Des*. 2011;17:1443-1449.
- Franklin GM, American Academy of Neurology. Opioids for chronic noncancer pain: a position paper of the American Academy of Neurology. *Neurology*. 2014;83:1277-1284.
- Crofford LJ. Adverse effects of chronic opioid therapy for chronic musculoskeletal pain. *Nat Rev Rheumatol*. 2010;6:191-197.
- Chapman CR, Lipschitz DL, Angst MS, et al. Opioid pharmacotherapy for chronic non-cancer pain in the United States: a research guideline for developing an evidence-base. *J Pain*. 2010;11:807-829.



51. Jungquist CR, Flannery M, Perlis ML, Grace JT. Relationship of chronic pain and opioid use with respiratory disturbance during sleep. *Pain Manag Nurs*. 2012;13:70-79.
52. Walker JM, Farney RJ, Rhondeau SM, et al. Chronic opioid use is a risk factor for the development of central sleep apnea and ataxic breathing. *J Clin Sleep Med*. 2007;3:455-461.
53. Manchikanti L, Abdi S, Atluri S, et al. American Society of Interventional Pain Physicians (ASIPP) guidelines for responsible opioid prescribing in chronic non-cancer pain: Part 2--guidance. *Pain Physician*. 2012;15:S67-116.
54. de Vasconcellos K, Sneyd JR. Nitrous oxide: are we still in equipoise? A qualitative review of current controversies. *Br J Anaesth*. 2013;111:877-885.
55. Yamakura T, Harris RA. Effects of gaseous anesthetics nitrous oxide and xenon on ligand-gated ion channels. Comparison with isoflurane and ethanol. *Anesthesiology*. 2000;93:1095-1101.
56. Jevtović-Todorović V, Todorović SM, Mennerick S, et al. Nitrous oxide (laughing gas) is an NMDA antagonist, neuroprotectant and neurotoxin. *Nat Med*. 1998;4:460-463.
57. Ranft A, Kurz J, Becker K, et al. Nitrous oxide (N2O) pre- and post-synaptically attenuates NMDA receptor-mediated neurotransmission in the amygdala. *Neuropharmacology*. 2007;52:716-723.
58. Chan MT, Wan AC, Gin T, Leslie K, Myles PS. Chronic postsurgical pain after nitrous oxide anesthesia. *Pain*. 2011;152:2514-2520.
59. Ohashi Y, Guo T, Orii R, Maze M, Fujinaga M. Brain stem opioidergic and GABAergic neurons mediate the antinociceptive effect of nitrous oxide in Fischer rats. *Anesthesiology*. 2003;99:947-954.
60. Bessière B, Laboureyras E, Chateauraynaud J, Laulin JP, Simonnet G. A single nitrous oxide (N2O) exposure leads to persistent alleviation of neuropathic pain in rats. *J Pain*. 2010;11:13-23.
61. Gruss M, Bushell TJ, Bright DP, Lieb WR, Mathie A, Franks NP. Two-pore-domain K⁺ channels are a novel target for the anesthetic gases xenon, nitrous oxide, and cyclopropane. *Mol Pharmacol*. 2004;65:443-452.
62. Patel AJ, Honoré E, Lesage F, Fink M, Romey G, Lazdunski M. Inhalational anesthetics activate two-pore-domain background K⁺ channels. *Nat Neurosci*. 1999;2:422-426.
63. Orestes P, Bojadzic D, Lee J, et al. Free radical signalling underlies inhibition of Ca_v3.2 T-type calcium channels by nitrous oxide in the pain pathway. *J Physiol*. 2011;589:135-148.
64. Todorovic SM, Jevtovic-Todorovic V, Mennerick S, Perez-Reyes E, Zorumski CF. Ca_v(3.2) channel is a molecular substrate for inhibition of T-type calcium currents in rat sensory neurons by nitrous oxide. *Mol Pharmacol*. 2001;60:603-610.
65. Honoré E. The neuronal background K_{2P} channels: focus on TREK1. *Nat Rev Neurosci*. 2007;8:251-261.
66. Georgiev SK, Kohno T, Ikoma M, Yamakura T, Baba H. Nitrous oxide inhibits glutamatergic transmission in spinal dorsal horn neurons. *Pain*. 2008;134:24-31.
67. de Lera Ruiz M, Kraus RL. Voltage-gated sodium channels: structure, function, pharmacology, and clinical indications. *J Med Chem*. 2015;58:7093-7118.
68. Zhou C, Liu J, Chen XD. General anesthesia mediated by effects on ion channels. *World J Crit Care Med*. 2012;1:80-93.
69. Fan L, Sonoda S, Watanabe M, et al. Effects of nitrous oxide and isoflurane on the L-type calcium current of rabbit ventricular myocytes and their modulation by beta-adrenoceptor stimulation. *Masui*. 2007;56:386-394.
70. Maze M, Fujinaga M. Recent advances in understanding the actions and toxicity of nitrous oxide. *Anaesthesia*. 2000;55:311-314.
71. Sawamura S, Kingery WS, Davies MF, et al. Antinociceptive action of nitrous oxide is mediated by stimulation of noradrenergic neurons in the brainstem and activation of [alpha]2B adrenoceptors. *J Neurosci*. 2000;20:9242-9251.
72. Zorumski CF, Nagele P, Mennerick S, Conway CR. Treatment-resistant major depression: rationale for NMDA receptors as targets and nitrous oxide as therapy. *Front Psychiatry*. 2015;6:172.
73. Duttaroy A, Gomez J, Gan JW, et al. Evaluation of muscarinic agonist-induced analgesia in muscarinic acetylcholine receptor knockout mice. *Mol Pharmacol*. 2002;62:1084-1093.
74. Wess J, Duttaroy A, Gomez J, et al. Muscarinic receptor subtypes mediating central and peripheral antinociception studied with muscarinic receptor knockout mice: a review. *Life Sci*. 2003;72:2047-2054.
75. Mennerick S, Jevtovic-Todorovic V, Todorovic SM, Shen W, Olney JW, Zorumski CF. Effect of nitrous oxide on excitatory and inhibitory synaptic transmission in hippocampal cultures. *J Neurosci*. 1998;18:9716-9726.
76. Zacny JP, Yajnik S, Coalson D, et al. Flumazenil may attenuate some subjective effects of nitrous oxide in humans: a preliminary report. *Pharmacol Biochem Behav*. 1995;51:815-819.
77. Suzuki T, Koyama H, Sugimoto M, Uchida I, Mashimo T. The diverse actions of volatile and gaseous anesthetics on human-cloned 5-hydroxytryptamine₃ receptors expressed in *Xenopus* oocytes. *Anesthesiology*. 2002;96:699-704.
78. De Angelis F, Tata AM. Analgesic effects mediated by muscarinic receptors: mechanisms and pharmacological approaches. *Cent Nerv Syst Agents Med Chem*. 2016;16:218-226.
79. Nabekura J, Xu TL, Rhee JS, Li JS, Akaike N. Alpha2-adrenoceptor-mediated enhancement of glycine response in rat sacral dorsal commissural neurons. *Neuroscience*. 1999;89:29-41.
80. Yalcin I, Charlet A, Cordero-Erausquin M, et al. Nociceptive thresholds are controlled through spinal beta2-subunit-containing nicotinic acetylcholine receptors. *Pain*. 2011;152:2131-2137.
81. Mody I, Pearce RA. Diversity of inhibitory neurotransmission through GABA(A) receptors. *Trends Neurosci*. 2004;27:569-575.
82. Bowery NG, Bettler B, Froestl W, et al. International Union of Pharmacology. XXXIII. Mammalian gamma-aminobutyric acid(B) receptors: structure and function. *Pharmacol Rev*. 2002;54:247-264.
83. Emmanouil DE, Quock RM. Advances in understanding the actions of nitrous oxide. *Anesth Prog*. 2007;54:9-18.
84. Al-Hasani R, Bruchas MR. Molecular mechanisms of opioid receptor-dependent signaling and behavior. *Anesthesiology*. 2011;115:1363-1381.
85. Zhang C, Davies MF, Guo TZ, Maze M. The analgesic action of nitrous oxide is dependent on the release of norepinephrine in the dorsal horn of the spinal cord. *Anesthesiology*. 1999;91:1401-1407.
86. Overland AC, Kitto KF, Chabot-Doré AJ, et al. Protein kinase C mediates the synergistic interaction between agonists acting at alpha2-adrenergic and delta-opioid receptors in spinal cord. *J Neurosci*. 2009;29:13264-13273.
87. Descalzi G, Ikegami D, Ushijima T, Nestler EJ, Zachariou V, Narita M. Epigenetic mechanisms of chronic pain. *Trends Neurosci*. 2015;38:237-246.
88. Liang L, Lutz BM, Bekker A, Tao YX. Epigenetic regulation of chronic pain. *Epigenomics*. 2015;7:235-245.
89. Trivedi MS, Deth R. Redox-based epigenetic status in drug addiction: a potential contributor to gene priming and a mechanistic rationale for metabolic intervention. *Front Neurosci*. 2014;8:444.
90. Nunn JF. Clinical aspects of the interaction between nitrous oxide and vitamin B12. *Br J Anaesth*. 1987;59:3-13.
91. Trivedi MS, Deth RC. Role of a redox-based methylation switch in mRNA life cycle (pre- and post-transcriptional maturation) and protein turnover: implications in neurological disorders. *Front Neurosci*. 2012;6:92.
92. Muñoz I, Urizar I, Casis L, Irazusta J, Subirán N. The epigenetic regulation of the opioid system: new individualized prompt prevention and treatment strategies. *J Cell Biochem*. 2015;116:2419-2426.
93. Herman BH, Vocci F, Bridge P. The effects of NMDA receptor antagonists and nitric oxide synthase inhibitors on opioid tolerance and withdrawal. Medication development issues for opiate addiction. *Neuropsychopharmacology*. 1995;13:269-293.
94. Dickenson AH, Sullivan AF. Evidence for a role of the NMDA receptor in the frequency dependent potentiation of deep rat dorsal horn nociceptive neurones following C fibre stimulation. *Neuropharmacology*. 1987;26:1235-1238.
95. Chabot-Doré AJ, Schuster DJ, Stone LS, Wilcox GL. Analgesic synergy between opioid and alpha2-adrenoceptors. *Br J Pharmacol*. 2015;172:388-402.
96. Ben Boujema M, Laboureyras E, Pype J, Bessière B, Simonnet G. Nitrous oxide persistently alleviates pain hypersensitivity in neuropathic rats: A dose-dependent effect. *Pain Res Manag*. 2015;20:309-315.
97. Turan A, Sarwar S, Atim A, et al. Nitrous oxide for the treatment of chronic low back pain. *Anesth Analg*. 2015;121:1350-1359.
98. Dworkin RH, O'Connor AB, Audette J, et al. Recommendations for the pharmacological management of neuropathic pain: an overview and literature update. *Mayo Clin Proc*. 2010;85:S3-14.
99. Hesdorffer CS, Longo DL. Drug-Induced Megaloblastic Anemia. *N Engl J Med*. 2015;373:1649-1658.
100. Sobotka JL, Alexander B, Cook BL. A review of carbamazepine's hematologic reactions and monitoring recommendations. *DICP*. 1990;24:1214-1219.

Date of submission: July 7, 2020

Date of decision: July 24, 2020

Date of acceptance: July 24, 2020

Date of web publication: Feb 26, 2021