



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



## Research paper

# Validation of a combined ELISA to detect IgG, IgA and IgM antibody responses to SARS-CoV-2 in mild or moderate non-hospitalised patients

A.M. Cook<sup>a</sup>, S.E. Faustini<sup>b</sup>, L.J. Williams<sup>a,\*</sup>, A.F. Cunningham<sup>c</sup>, M.T. Drayson<sup>b</sup>, A. M. Shields<sup>b,d</sup>, D. Kay<sup>a</sup>, L. Taylor<sup>e</sup>, T. Plant<sup>b</sup>, A. Huissoon<sup>c,d</sup>, G. Wallis<sup>a</sup>, S. Beck<sup>d</sup>, S.E. Jossi<sup>c</sup>, M. Perez-Toledo<sup>c</sup>, M.L. Newby<sup>f</sup>, J.D. Allen<sup>f</sup>, M. Crispin<sup>f</sup>, S. Harding<sup>a</sup>, A.G. Richter<sup>b,d</sup>

<sup>a</sup> The Binding Site Group Ltd, 8 Calthorpe Road, Birmingham B15 1QT, UK

<sup>b</sup> Clinical Immunology Service, University of Birmingham College of Medical and Dental Sciences, Birmingham B15 2TT, UK

<sup>c</sup> Institute of Immunology and Immunotherapy, University of Birmingham, Birmingham B15 2TT, UK

<sup>d</sup> University Hospitals Birmingham, NHS Foundation Trust, Birmingham B15 2GW, UK

<sup>e</sup> The Royal Wolverhampton NHS Trust, Wolverhampton Road, Wolverhampton, West Midlands WV10 0QP, UK

<sup>f</sup> School of Biological Sciences, University of Southampton, Southampton SO17 1BJ, UK

## ARTICLE INFO

## Keywords:

COVID-19  
Dried blood spot  
Trimeric spike protein  
Mild disease  
Low prevalence  
Serological

## ABSTRACT

**Background:** Frequently SARS-CoV-2 results in mild or moderate disease with potentially lower concentrations of antibodies compared to those that are hospitalised. Here, we validated an ELISA using SARS-CoV-2 trimeric spike glycoprotein, with targeted detection of IgG, IgA and IgM (IgGAM) using serum and dried blood spots (DBS) from adults with mild or moderate disease.

**Methods:** Targeting the SARS-CoV-2 trimeric spike, a combined anti-IgG, IgA and IgM serology ELISA assay was developed using 62 PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples,  $\geq 14$  days post symptom onset and 624 COVID-19 negative samples. The assay was validated using 73 PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples,  $\geq 14$  days post symptom onset and 359 COVID-19 negative serum samples with an additional 81 DBSs. The assay was further validated in 226 PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples,  $\geq 14$  days post symptom onset and 426 COVID-19 negative clinical samples.

**Results:** A sensitivity and specificity of 98.6% (95% CI, 92.6–100.0), 98.3% (95% CI, 96.4–99.4), respectively, was observed following validation of the SARS-CoV-2 ELISA. No cross-reactivities with endemic coronaviruses or other human viruses were observed, and no change in results were recorded for interfering substances. The assay was stable at temperature extremes and components were stable for 15 days once opened. A matrix comparison showed DBS to correlate with serum results. Clinical validation of the assay reported a sensitivity of 94.7% (95% CI, 90.9–97.2%) and a specificity of 98.4% (95% CI, 96.6–99.3%).

**Conclusions:** The human anti-IgGAM SARS-CoV-2 ELISA provides accurate and sensitive detection of SARS-CoV-2 antibodies in non-hospitalised adults with mild or moderate disease. The use of dried blood spots makes the assay accessible to the wider community.

## 1. Introduction

The COVID-19 pandemic resulting from severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection was declared an international public health emergency in early 2020 (Organization WH, 2020a). Unlike previous coronavirus infection outbreaks, such as SARS-CoV and Middle East Respiratory Syndrome, the lack of rapid, overt symptomatic presentation in 80% of patients (Organization WH, 2020b;

Organization WH, 2020c), has meant seroconversion rather than molecular testing has become an established method to understand SARS-CoV-2 epidemiology.

Asymptomatic or even mild infection cases represent a challenge for in vitro SARS-CoV-2 diagnostic assays, both in terms of understanding the required sensitivity and specificity (Deeks et al., 2020), and for obtaining samples from molecular tested positive patients. In addition, as seroconversion to different immunoglobulin types is not a synchronous process, testing of the major humoral immune system components

\* Corresponding author at: The Binding Site Group, 8 Calthorpe Road, Birmingham B15 1QT, UK.

E-mail address: [Leigh.Williams@bindingsite.com](mailto:Leigh.Williams@bindingsite.com) (L.J. Williams).

<https://doi.org/10.1016/j.jim.2021.113046>

Received 17 November 2020; Received in revised form 18 March 2021; Accepted 20 March 2021

Available online 26 March 2021

0022-1759/© 2021 Published by Elsevier B.V.

## Abbreviations

DBS	Dried Blood Spots
ELISA	Enzyme Linked Immunosorbent Assay
PCR	Polymerase Chain Reaction
SARS-CoV-2	Severe Acute Respiratory Syndrome Coronavirus 2

may be preferable to limiting assessment to individual isotypes such as IgG, IgA & IgM (IgGAM).

Here we describe the validation of an ELISA, utilising recombinant SARS-CoV-2 trimeric spike glycoprotein (Wrapp et al., 2020; Watanabe et al., 2020), with targeted detection of IgG, IgA and IgM antibodies in non-hospitalised adults with mild or moderate disease.

## 2. Materials & methods

### 2.1. Samples

Serum and Dried Blood Spot (DBS) samples were collected from consenting non-hospitalised polymerase chain reaction (PCR) positive adults, presenting with mild or moderate symptoms of COVID-19 disease,  $\geq 14$  days post symptom onset, and from individuals COVID-19 negative (Organization WH, 2020c; Prevention CfDCA, 2020; Shah et al., 2020). Samples collected prior to the pandemic (pre December 2019) were deemed COVID-19 negative. Serum samples were stored at  $-20^{\circ}\text{C}$  and thawed before analysis.

Residual anonymised samples were used in this study from several sources. For the specificity and sensitivity analysis residual samples were from commercial sources (BioIVT, approved by Western Institutional Review Board - IRB20161665; TCS Bioscience approved by The Diagnostics Investigational Review Board- IRB112015-01.1; anonymised normal samples provided by Plasma Services Group were collected with patient consent via an FDA registered blood establishment (registration number 3005275238)) and further residual samples collected in accordance with REC 2002/201; South Birmingham Research Ethics Service. For dried blood spot comparison, residual anonymised samples were collected from University of Birmingham (REC 2002/201; South Birmingham Research Ethics Service). For cross reactivity analysis, samples were provided by the Amsterdam University Medical Center of the University of Amsterdam, the Netherlands (approved by the Medical Ethics Committee; MEC 07/182MEC 07/182). Samples were used in accordance with the world medical association declaration of Helsinki Ethical principles for Medical Research involving human subjects.

### 2.2. ELISA

A stabilised trimeric spike protein preparation with 95% purity (Watanabe et al., 2020), assessed by silver stain, was coated onto ELISA plates (Nunc High Bind, Thermo Fisher Scientific, Loughborough, UK) at  $1.0\ \mu\text{g}/\text{mL}$  in PBS for 24 h. Plates were blocked using 2% BSA blocking reagent (Sigma, Gillingham, UK) prepared in PBS-0.1% Tween 20 and incubated for 1 h at room temperature. Subsequently, plates were washed three times with PBS-1% Tween +0.08% Proclin300 and air dried before packing in airtight in aluminium-coated packs with drying agent (IDS, Gateshead, UK).

High, low, negative and cut-off calibrator controls were generated using COVID-19 negative pooled normal human and specific human monoclonal IgG1, IgA and IgM anti-SARS-CoV-2 S glycoprotein (CR3022, Absolute Antibodies, Upper Heyford, UK) (Huo et al., 2020).  $100\ \mu\text{L}$  of either the controls (neat dilution), calibrator (neat dilution) or serum samples (diluted 1/40) were added to the coated plate and incubated for 30 min at room temperature. The plate was then washed

three times with PBS-0.1% Tween and purified horseradish peroxidase (HRP)-labelled polyclonal rabbit antibody to human IgG, IgA, IgM (DAKO, USA) ( $100\ \mu\text{L}$ ) was added to each well and then incubated for a further 30 min at room temperature (RT). The plate was washed a further three times and substrate solution containing 3,3',5,5'-tetramethylbenzidine ( $100\ \mu\text{L}$ , Surmodics Inc., Eden Prairie, MN, USA) was added to each well and then incubated at RT in the dark for 10 min. The reaction was stopped by the addition of phosphoric acid ( $100\ \mu\text{L}$ ) and the plate read at 450 nm using either Biotek Synergy HTX microplate reader or an automated liquid handler (Dynex Technologies, USA).

### 2.3. Pilot study: sensitivity, specificity and cut-off co-efficient

COVID-19 negative samples ( $n = 624$ , median 63 years of age, range 18–98) and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples,  $\geq 14$  days post-symptom onset ( $n = 62$ , median 44 years of age, range 19–63) were analysed using manual and automated platforms using the human anti-IgGAM SARS-CoV-2 ELISA. The cut-off coefficient was determined, and the clinical study powered.

## 3. Validation of the manufacturing process

### 3.1. Sensitivity and specificity of the ELISA

Sensitivity and specificity of the human anti-IgGAM SARS-CoV-2 ELISA was assessed using COVID-19 negative serum samples ( $n = 359$ ), and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples ( $n = 73$ ),  $\geq 14$  days post symptom onset, manually.

### 3.2. Precision of the ELISA

Precision was measured in accordance with the Clinical and Laboratory Standards Institute guidelines EP12-A2 (Wayne, 2008). Forty COVID-19 negative serum samples replicates were spiked ( $< 1\%$ ) with specific human monoclonal IgG1 anti-SARS-CoV-2 S glycoprotein (CR3022), at C5, C50 and C95. The C50 level, defined as a response of 1.0, and at the C5- C95 interval, defined as being bounded by the response of 0.8–1.2 ( $\pm 20\%$  of C50 response). The acceptable ranges for testing each of these three levels was defined as  $\pm 5\%$  of the target response. Acceptance criteria was defined as, C5:  $\geq 36$  negatives out of 40 replicates; C50: 14–26 positives out of 40 replicates; C95:  $\geq 36$  positives out of 40 replicates. The coefficient of variation (% CV) was additionally calculated for each cut-off.

### 3.3. Interference with the ELISA

Interference analysis was performed by spiking haemoglobin (2000 mg/L), bilirubin (200 mg/L), triglyceride (3000 mg/L) into quintuplicate serum samples that were either antibody positive or antibody negative, (normal human serum spiked with/without CR3022). Controls were generated by spiking samples with equivalent volumes of saline. The assay was deemed to have passed the interference assessment if there was no change in the reportable value following the addition of the interfering agent.

## 4. Cross reactivity of the ELISA with other respiratory diseases

Cross-reactivity of the assay was assessed in patients with known respiratory diseases; endemic coronaviruses ( $n = 11$ ), influenza and parainfluenza ( $n = 14$ ), enteroviruses ( $n = 5$ ), Epstein Bar virus ( $n = 4$ ), adenovirus ( $n = 1$ ) or respiratory syncytial virus ( $n = 4$ ). Positive cross-reactivity was identified if the cut-off index value was  $\geq 1$ .

### 4.1. Stability of the ELISA

Extremes of temperature stability was determined in triplicate

following 24 h storage at either 37 °C or – 20 °C. Open vial stability was assessed over 15 days with 5 day interval sampling. Briefly, ELISAs were opened in parallel and components re-capped and stored at 4 °C.

#### 4.2. Dried blood spot versus serum samples

Paired DBS and serum samples (49 PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples and 32 COVID-19 negative samples) collected at the same time, were assayed using the human anti-IgGAM SARS-CoV-2 ELISA. Capillary blood samples (50 µL per spot) were collected onto 226 grade forensic collection cards (Ahlstrom Munksjo, Germany). Individual 12 mm diameter pre-perforated DBS spots were isolated using a sterile pipette tip and placed into a universal tube at a ratio of one spot to 250 µL 0.05% PBS Tween-20 (Sigma). Tubes were briefly vortexed and incubated overnight at room temperature. DBS eluate was subsequently harvested into a microtube and centrifuged at 10,600 ×g for 10 min at room temperature and stored at 4 °C until used (Morley et al., 2020). DBS samples were applied to the assay at a 1 in 4 dilution, resulting in a 1 in 40 overall dilution. Results were analysed using Bland Altman, Passing-Bablok and Cohen's kappa statistic.

#### 4.3. Clinical study

COVID-19 negative (n = 426) and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples (n = 226), ≥14 days post symptom onset, were assayed using the human anti-IgGAM SARS-CoV-2 ELISA on the automated platform.

#### 4.4. Statistics

All graphs and statistical analysis were generated using Graph Pad Prism statistical software version 5.04 or Analyse-it® for Microsoft Excel. Assay validation sensitivity, specificity and precision (95% CI and % CVs) were calculated using Analyse-it. The matrix comparison was analysed using Bland-Altman, Passing Bablok and Cohen's kappa statistics (Analyse-it). To achieve an expectant 98%, minimum 93%, sensitivity in the clinical study, a 90% power with 5% alpha error was determined from the pilot study. A p < 0.05 was considered statistically significant.

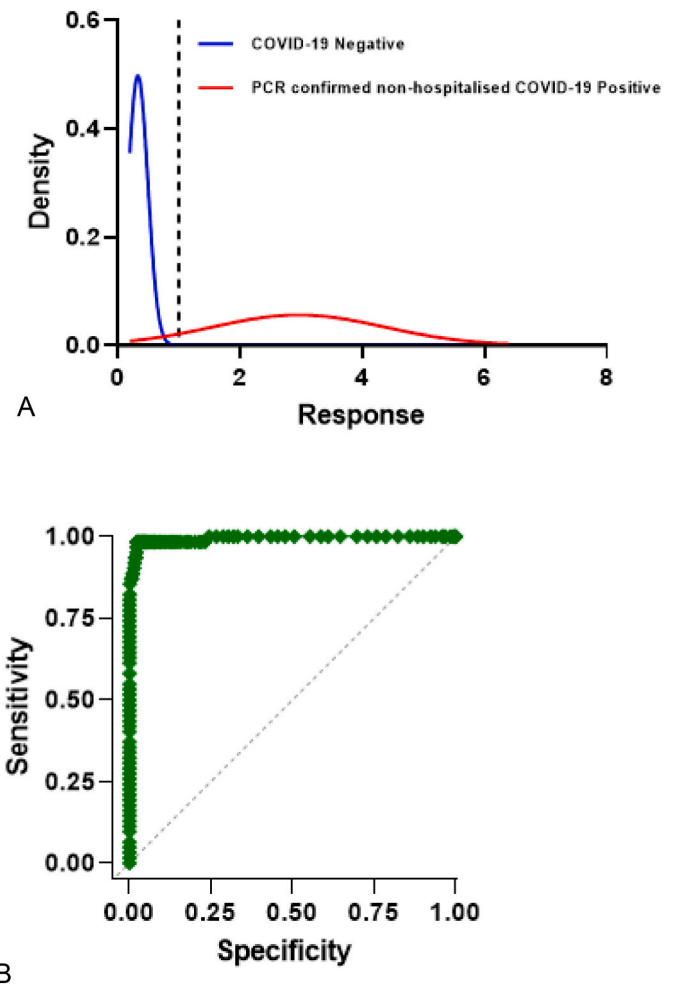
### 5. Results

#### 5.1. Pilot study to determine sensitivity, specificity and cut-off co-efficient of the ELISA

The pilot study with 624 COVID-19 negative and 62 PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples, which were at least 14 days from symptom onset were analysed on the human anti-IgGAM SARS-CoV-2 ELISA. The ELISA output result was reported as a ratio to the cut-off calibrator (run in parallel) and multiplied by the cut-off co-efficient. The cut-off coefficient was determined from the pilot study, results were reported as the sample to plate ratio (samples optical density at 450 nm /cut off calibrator optical density at 450 nm). From

the sample to plate ratio results, sensitivity and specificity was calculated and the cut-off calibrator co-efficient determined. This would maintain batch-to-batch consistency and was defined here as 1.15. A sensitivity of 98.5% (95% CI 91.8–100%) and specificity of 97.6% (95% CI 96.1–98.7%) was achieved, (Table 1 & Fig. 1).

From the pilot study a minimum of 167 PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples, ≥14 days post symptom onset would be required for the clinical study to determine whether the ELISA test delivered the expected sensitivity of 98% (minimum of 93%)



**Fig. 1.** The distribution of responses and sensitivity and specificity of the human anti-IgGAM SARS-CoV-2 ELISA.

The cumulative standard normal distribution of optical densities of (A) COVID-19 negative samples (n = 624, blue) and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples, ≥14 days post symptom onset (n = 62, red). (B) ROC curve of sensitivity (98.5%, 95% CI 91.4–99.7%) and specificity (97.6%, 95% CI 96.1–98.5%).

**Table 1**

Sensitivity and specificity of the pilot, validation and clinical study using the human anti-IgGAM SARS-CoV-2 ELISA. Assay assessed using COVID-19 negative samples and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples, ≥14 days post symptom onset.

Pilot, validation & clinical studies: specificity & sensitivity						
Study	Category	Number	Positive	Negative	Specificity (95% CI)	Sensitivity (95% CI)
Pilot Study (manual & automated)	COVID-19 negative	624	15	609	97.6 (96.1–98.5)	98.5% (91.4–99.7)
	PCR confirmed, non-hospitalised COVID-19 positive	62	61	1		
Validation Study (manual)	COVID-19 negative	359	6	353	98.3 (96.4–99.4)	98.6 (92.6–100.0)
	PCR confirmed, non-hospitalised COVID-19 positive	73	72	1		
Clinical Study (automated)	COVID-19 negative	426	7	419	98.4 (96.6–99.3)	94.7 (90.9–97.2)
	PCR confirmed, non-hospitalised COVID-19 positive	226	214	12		

with an alpha error of 5% and power of 90%.

### 5.2. Validation of the manufacturing process

Sensitivity and specificity were assessed in COVID-19 negative samples ( $n = 359$ ) and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples,  $\geq 14$  days post symptom onset ( $n = 73$ ). COVID-19 negative samples were identified as negative for SARS-CoV-2 antibodies when assay results were less than 1 and PCR confirmed COVID-19 positive samples were identified as positive when the assay results were greater than 1, using the human anti-IgGAM SARS-CoV-2 ELISA (Fig. 2). This resulted in an assay sensitivity of 98.6% (95% CI 92.6–100.0%) and specificity of 98.3% (95% CI 96.4–99.4%), (Table 1).

Assay precision was assessed in 40 replicates by following the CLSI EP12-A2 guidelines to determine whether the C5 to C95 response range (range of responses at which imprecision around the C50 occurs) falls within 20% of C50. The precision of the assay was demonstrated at  $\pm 20\%$  of C50 as C5 gave a mean of 0.77 (40 negatives out of 40 replicates, 4.56% CV), C50 gave a mean of 1.02 (25 positives out of 40 replicates, 5.07% CV) and C95 gave a mean of 1.13 (40 positives out of 40 replicates, 3.97% CV), (Fig. 3).

Generally, no change in overall result outcome were observed following potential analytical interference from increased haemoglobin, bilirubin or triglyceride concentrations, (Fig. 4 A). Two samples containing elevated bilirubin did result in increased result outputs, however, only 1 sample resulted in a change of response. Assessing cross-reactivity with other human viruses and coronaviruses, 39 samples tested negative covering 8 disease categories, (Fig. 4 B). Kit components were stable once opened for 15 days and for one day at extremes of temperature (Fig. 4 C & D).

### 5.3. Dried blood spot versus serum samples

Responses were measured in 81 matched serum and DBS samples, with unknown PCR status. A strong correlation was observed between the two matrices when assessed using the ELISA (Passing Bablok  $r = 0.959$ ,  $y = 0.16-0.91x$ ), with a mean positive bias towards the DBS compared to serum (Bland Altman 0.05, 95% CI -0.773 to 0.679), (Fig. 5). An almost perfect agreement, 0.83 (Cohen's kappa) was observed between the two sampling matrices.

### 5.4. Clinical study

The assay was validated in clinical samples from an independent

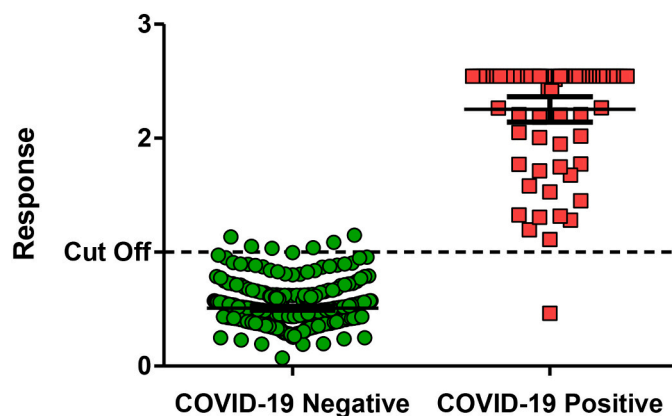


Fig. 2. IgGAM response in COVID-19 negative ( $n = 359$ ) and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples,  $\geq 14$  days post symptom onset ( $n = 73$ ).

A sensitivity and specificity of 98.6% (95% CI 92.6–100%) and 98.3% (95% CI 96.4–99.4%), respectively, was reported using an assay cut-off of 1.0.

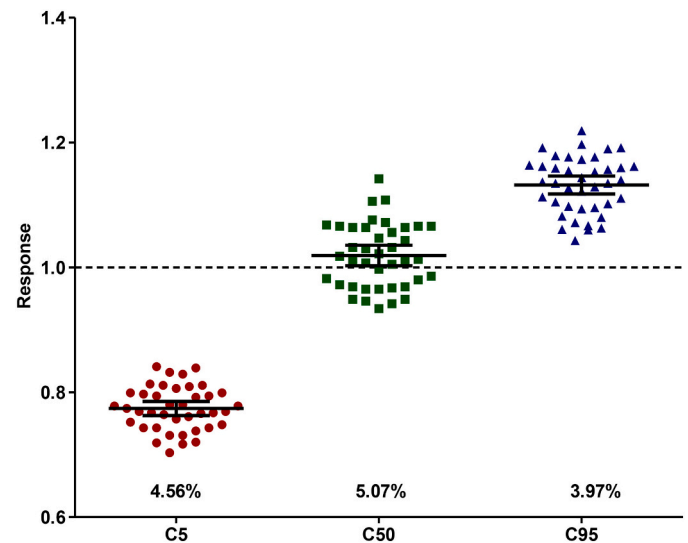


Fig. 3. Precision of the human anti-IgGAM SARS-CoV-2 ELISA.

Precision was measured using clinical and lab standards guidelines EP12-A2 in 40 replicates and assessed at clinical cut-offs C5, C50 and C95. The mean and coefficient of variation (%) was calculated for each clinical cut-off.

group of COVID-19 negative samples ( $n = 426$ ) and PCR-confirmed non-hospitalised, mild or moderate COVID-19 samples,  $\geq 14$  days post symptom onset ( $n = 226$ ). The sensitivity of the anti-IgGAM SARS-CoV-2 ELISA in this cohort was 94.7% (95% CI, 90.9–97.2%) and specificity 98.4% (95% CI, 96.6–99.3%), (Fig. 6 & Table 1).

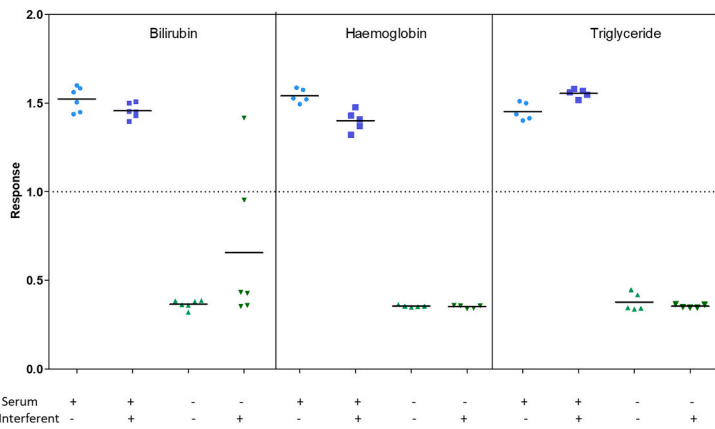
## 6. Discussion

Seroepidemiological studies are essential in understanding population-level exposure to SARS-CoV-2, determining correlates of protection from re-infection and guiding public health policy. The accuracy of such studies is determined by the performance characteristics of the immunoassay used to determine serostatus. Using samples derived from non-hospitalised adults with mild or moderate COVID-19 disease we developed a highly sensitive (98.6%) and specific (98.3%) anti-IgGAM SARS-CoV-2 ELISA. The assay showed no cross reactivity with endemic coronaviruses or other respiratory diseases, even though 30% of common colds are caused by alpha and beta coronaviruses (Paules et al., 2020).

By validating this assay using samples from non-hospitalised adults, we demonstrate the human anti-IgGAM SARS-CoV-2 ELISA to be a reliable assay for future seroepidemiological studies of the general population, the majority of whom will not require hospitalisation following infection with SARS-CoV-2. Current testing in the community has been problematic due to the inadequate sensitivities and specificities of current point of care tests in non-hospitalised patients with mild or moderate disease (Deeks et al., 2020).

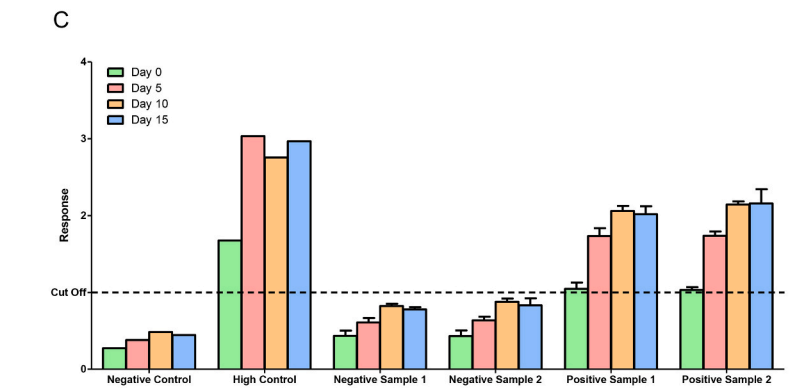
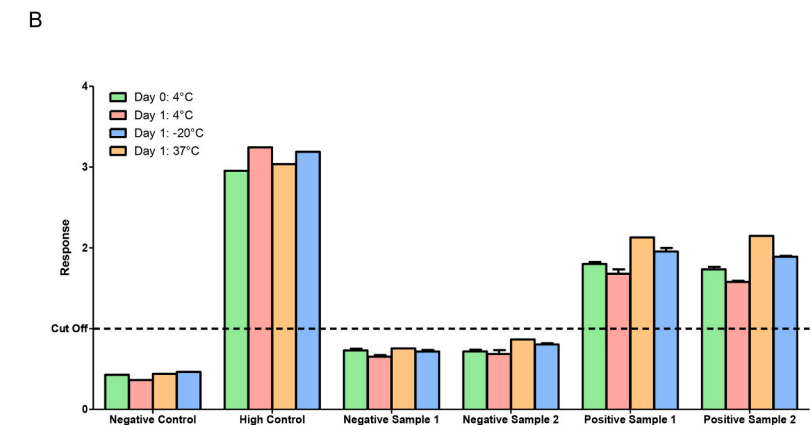
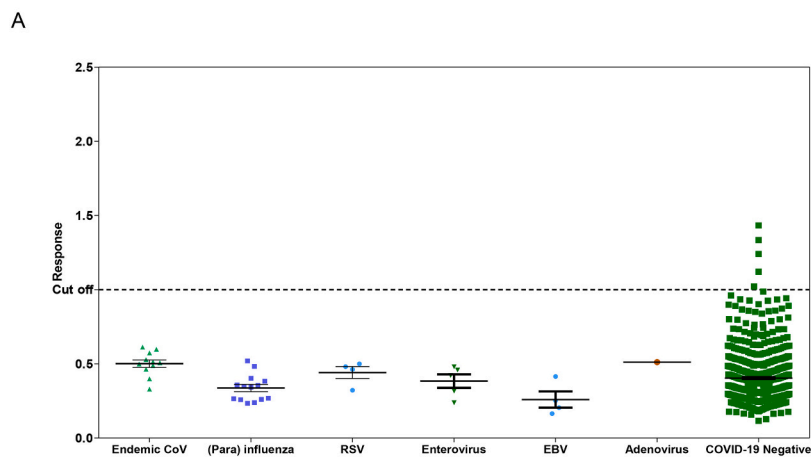
The assay targets nearly the entirety of the trimeric spike protein, and thereby offers a greater range of epitopes, as reports suggest neutralising antibodies to SARS-CoV-2 are not only targeted towards the receptor binding domain but also elicit high affinity and cross-reactivity to other sites across the spike (Wrapp et al., 2020; Brouwer et al., 2020; Rogers et al., 2020; Chi et al., 2020; Liu et al., 2020). Faustini et al. compared the receptor binding domain, S1 domain and nucleocapsid and although these targets were excellent at detecting antibodies in severe COVID-19 patients, they were not as effective when antibody concentrations were more limited (Faustini et al., 2020). Serological assays have been developed to detect the antibody response present in severe disease (hospitalised patients). This assay was therefore developed and tested in non-hospitalised adults, at least 14 days post-symptom onset, with mild or moderate COVID-19, where the antibody





**Fig. 4.** Stability and cross reactivity of antibody positive or antibody negative serum samples on the anti-IgGAM SARS CoV-2 ELISA.

The assay was assessed for (A) interfering substances, bilirubin, haemoglobin and triglyceride (n = 5 replicates per test) in antibody positive or antibody negative serum samples; and (B) evaluated for cross reactivity with human viruses and other endemic coronaviruses (n = 39). Additionally, the kit components were tested for (C) stability at extreme temperatures (-20 °C, 4 °C and 37 °C) (n = 3) and (D) stability once opened (n = 4).

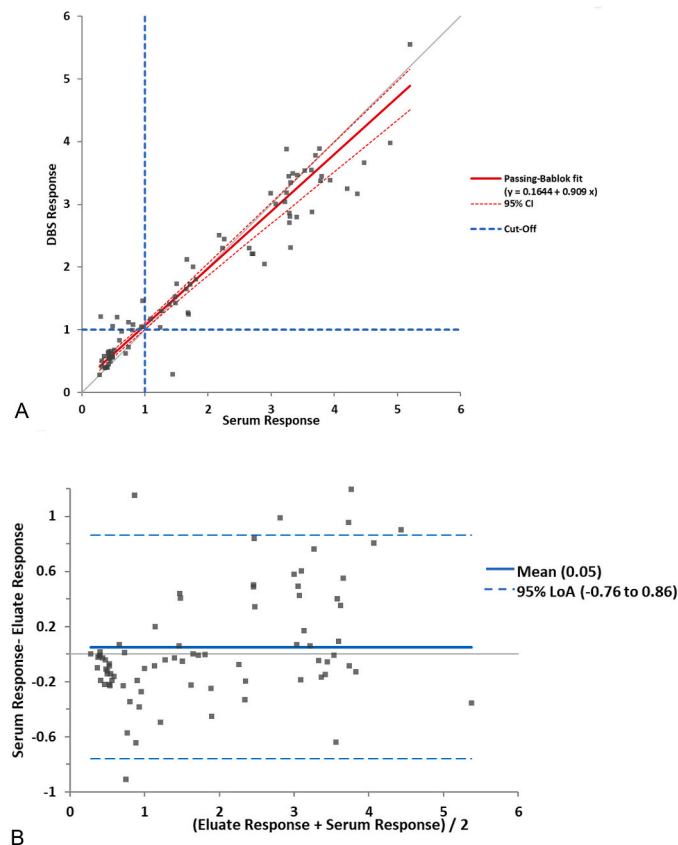


concentrations are much lower than in hospitalised patients with severe disease (Deeks et al., 2020; Faustini et al., 2020; Sun et al., 2020).

The assay measures anti-IgG, IgA and IgM as reports suggest an asynchronous serological response. In adults, Seow et al. showed that the kinetics of IgGAM only occurred synchronously in 50% patients, with 9.7% developing either IgG, A and M first, 9.7% developing IgM and IgG together, 3% developing IgA and IgM together, and 3% developing IgA and IgG together (Seow et al., 2020). Valdivia et al. reported that 3.5% asymptomatic patients presented with IgM while negative for IgG (Valdivia et al., 2021). Long et al. showed that seroconversion for IgG and IgM occurred synchronously or simultaneously (Long et al., 2020). Additionally, the longevity of the antibody responses is still under investigation. Studies by Iyer et al. showed that adult patients were IgA and IgM seronegative by 51 and 47 days, respectively, while IgG persisted to the end of their evaluation period (75 days) (Iyer et al., 2020). Gudbjartsson, reported IgG persisting without reduction up to 4 months after diagnosis (Gudbjartsson et al., 2020). A potential limitation associated with measuring the combined IgGAM result to that of IgG alone is that the IgA and IgM antibody response will reduce as the immune response prevails, therefore measurement of IgG alone could be indicative of the longer term response.

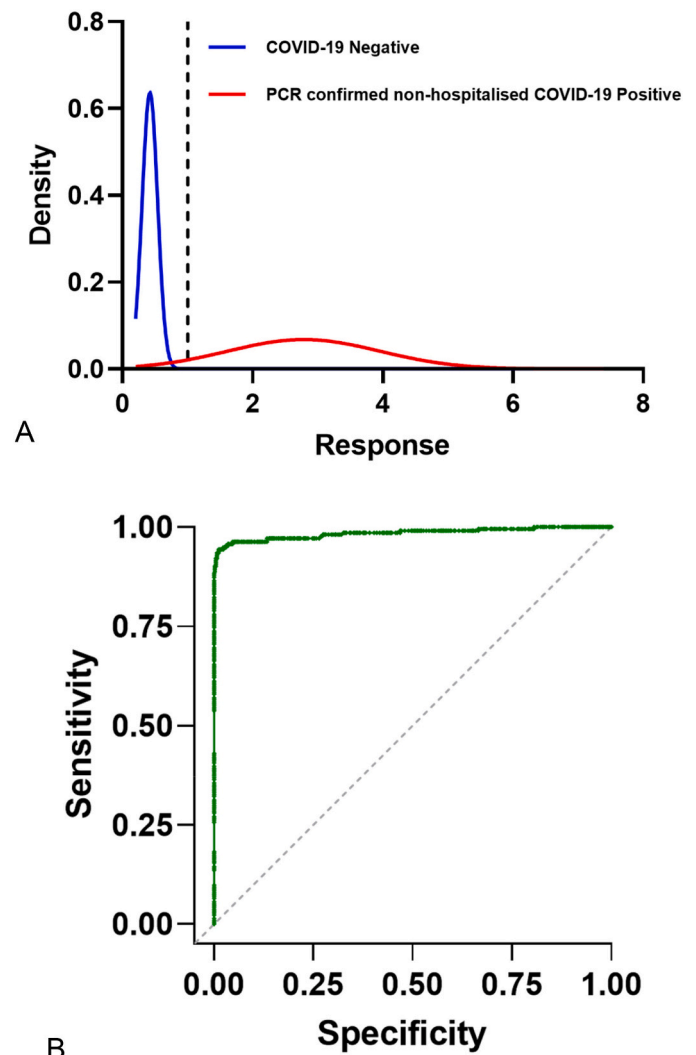
This assay benefits by being able to assess SARS-CoV-2 IgGAM antibodies in both mild and moderate disease, while also being compatible with remote sampling. Here, antibodies obtained from DBSs were shown to correlate well with serum results. DBSs are readily used in analytical

assays for viruses such as those for human immunodeficiency virus (HIV) and hepatitis B and C (Freeman et al., 2018; Ross et al., 2013; Cassol et al., 1992) They are also frequently used for other applications such as screening of neonates, metabolic profiling, pharmacokinetic, forensic and environmental assays (Freeman et al., 2018; Gupta and Mahajan, 2018; Bjorkesten et al., 2017). The major advantages with using DBS's include: the requirement for a smaller volume of blood, reduced risk of potential bacterial contamination, non-invasive blood collection, easy delivery-to- test centre, low cost and stability for prolonged periods of time with minimal deterioration of analytes (Morley et al., 2020; Freeman et al., 2018; Gupta and Mahajan, 2018). The additional advantage in the current COVID-19 pandemic include, ensuring patients do not have to attend hospitals and sites with greater risk of infection. Rickman et al. reported in a major London Teaching Hospital 15% of in-patient COVID-19 cases developed as a result of varied transmission routes within the hospital (Rickman et al., 2020) and Meredith et al. also reported strong and plausible epidemiological links following the identification of 35 clusters of identical viruses



**Fig. 5.** Comparison of the anti-IgGAM SARS-CoV-2 ELISA response in DBS and serum samples.

The response was measured in 81 DBS and serum matched samples. A comparison between the two collection methods were assessed using (A) Passing-Bablok analysis ( $r = 0.959$ ,  $y = 0.16 + 0.91x$ ) and (B) Bland-Altman analysis (central line demonstrates mean difference and broken lines shows the limits of agreement,  $-0.76$  to  $0.86$ ).



**Fig. 6.** Clinical study.

The cumulative standard normal distribution of (A) COVID-19 negative samples ( $n = 426$ , blue) and PCR confirmed, mild or moderate COVID-19 positive samples,  $\geq 14$  days post symptom onset samples ( $n = 226$ , red). (B) ROC curve of sensitivity (94.7%, 95% CI 90.9–97.2%) and specificity (98.4%, 95% CI 96.6–99.3%).

associated among patients in their hospital (Meredith et al., 2020). Therefore, when using DBSs, the test becomes accessible to all, including those that are vulnerable and those beyond developed countries.

In conclusion, utilising the trimeric spike protein and measuring IgG, IgA and IgM simultaneously, the human anti-IgGAM SARS-CoV-2 ELISA assay provides accurate and sensitive detection of antibody responses to SARS-CoV-2 in adults with mild or moderate disease. The use of dried blood spots makes this assay more accessible to the wider community.

## Declarations

LJW, AMC, GW, SH & DK are members of The Binding Site Group Ltd.

## Acknowledgments

We thank Lia van der Hoek for seasonal coronaviruses samples used in the cross reactivity study from the Amsterdam Cohort Studies (ACS) on HIV infection and AIDS. The ACS is a collaboration among the Public Health Service of Amsterdam, the Amsterdam University Medical Center of the University of Amsterdam, the Sanquin Blood Supply Foundation, the Medical Center Jan van Goyen and the HIV Focus Center of the DC-Clinics. It is part of the Netherlands HIV Monitoring Foundation and financially supported by the Center for Infectious Disease Control of the Netherlands National Institute for Public Health and the Environment. The authors thank all ACS participants for their contribution, as well as the ACS study nurses, data managers and lab technicians.

## References

- Bjorksten, J., Enroth, S., Shen, Q., Wik, L., Hougaard, D.M., Cohen, A.S., et al., 2017. Stability of proteins in dried blood spot biobanks. *Mol. Cell. Proteomics* 16 (7), 1286–1296.
- Brouwer, P.J.M., Caniels, T.G., van der Straten, K., Snitselaar, J.L., Aldon, Y., Bangaru, S., et al., 2020. Potent neutralizing antibodies from COVID-19 patients define multiple targets of vulnerability. *Science* 369 (6504), 643–650.
- Cassol, S., Salas, T., Gill, M.J., Montpetit, M., Rudnik, J., Sy, C.T., et al., 1992. Stability of dried blood spot specimens for detection of human immunodeficiency virus DNA by polymerase chain reaction. *J. Clin. Microbiol.* 30 (12), 3039–3042.
- Chi, X., Yan, R., Zhang, J., Zhang, G., Zhang, Y., Hao, M., et al., 2020. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2. *Science* 369 (6504), 650–655.
- Deeks, J.G., Dinnes, J., Takwoingi, Y., Davenport, C., Spijker, R., Taylor-Phillips, S., et al., 2020. Antibody tests for identification of current and past infection with SARS-CoV-2. *Cochrane Database Syst. Rev.* 6, CD013652.
- Faustini, S.E., Jossi, S.E., Perez-Toledo, M., Shields, A., Allen, J.D., Watanabe, Y., et al., 2020 Jun 18. Detection of antibodies to the SARS-CoV-2 spike glycoprotein in both serum and saliva enhances detection of infection. *medRxiv*. <https://doi.org/10.1101/2020.06.16.20133025>.
- Freeman, J.D., Rosman, L.M., Ratcliff, J.D., Strickland, P.T., Graham, D.R., Silbergeld, E. K., 2018. State of the science in dried blood spots. *Clin. Chem.* 64 (4), 656–679.
- Gudbjartsson, D.F., Norddahl, G.L., Melsted, P., Gunnarsdottir, K., Holm, H., Eythorsson, E., et al., 2020. Humoral immune response to SARS-CoV-2 in Iceland. *N. Engl. J. Med.* 382, 2302–2315.
- Gupta, K., Mahajan, R., 2018. Applications and diagnostic potential of dried blood spots. *Int. J. Appl. Basic Med. Res.* 8 (1), 1–2.
- Huo, J., Zhao, Y., Ren, J., Zhou, D., Duyvesteyn, H.M.E., Ginn, H.M., et al., 2020. Neutralization of SARS-CoV-2 by destruction of the prefusion spike. *Cell Host Microbe* 28 (3), 445–454 (e6).
- Iyer, A.S., Jones, F.K., Nodoushani, A., Kelly, M., Becker, M., Slater, D., et al., 2020. Dynamics and significance of the antibody response to SARS-CoV-2 infection. *medRxiv* 5 (52).
- Liu, L., Wang, P., Nair, M.S., Yu, J., Rapp, M., Wang, Q., et al., 2020. Potent neutralizing antibodies against multiple epitopes on SARS-CoV-2 spike. *Nature* 584 (7821), 450–456.
- Long, Q.X., Liu, B.Z., Deng, H.J., Wu, G.C., Deng, K., Chen, Y.K., et al., 2020. Antibody responses to SARS-CoV-2 in patients with COVID-19. *Nat. Med.* 26 (6), 845–848.
- Meredith, L.W., Hamilton, W.L., Warne, B., Houldcroft, C.J., Hosmillo, M., Jahun, A.S., et al., 2020. Rapid implementation of SARS-CoV-2 sequencing to investigate cases of health-care associated COVID-19: a prospective genomic surveillance study. *Lancet Infect. Dis.* 20 (11), 1263–1272.
- Morley, G.L., Taylor, S., Jossi, S., Perez-Toledo, M., Faustini, S.E., Marcial-Juarez, E., et al., 2020. Sensitive detection of SARS-CoV-2-specific antibodies in dried blood spot samples. *Emerg. Infect. Dis.* 26 (12).
- Organization WH, 2020a. WHO Director-General's Opening Remarks at the Media Briefing on COVID-19-11 March 2020. Available from: <https://www.who.int/dg/speeches/detail/who-director-general-s-opening-remarks-at-the-media-briefing-on-covid-19-11-march-2020>.
- Organization WH, 2020b. Coronavirus Disease 2019 (COVID-19) Situation Report-41.
- Organization WH, 2020c. Clinical Management of COVID-19-Interim Guidance.
- Paules, C.I., Marston, H.D., Fauci, A.S., 2020. Coronavirus infections-more than just the common cold. *JAMA.* 323 (8), 707–708.
- Prevention CfDca, 2020. Interim Clinical Guidance for Management of Patients With confirmed Coronavirus Disease (COVID-19). Available from: <https://www.cdc.gov/coronavirus/2019-ncov/hcp/clinical-guidance-management-patients.html>.
- Rickman, H.M., Rampling, T., Shaw, K., Martinez-Garcia, G., Hail, L., Coen, P., et al., 15 February 2021. Nosocomial transmission of COVID-19: a retrospective study of 66 hospital-acquired cases in a London teaching hospital. *Clin. Infect. Dis.* 72 (4), 690–693.
- Rogers, T.F., Zhao, F., Huang, D., Beutler, N., Burns, A., He, W.T., et al., 2020. Isolation of potent SARS-CoV-2 neutralizing antibodies and protection from disease in a small animal model. *Science* 369 (6506), 956–963.
- Ross, S., Stambouli, O., Gruner, N., Marcus, U., Cai, W., Zhang, W., et al., 2013. Detection of infections with hepatitis B virus, hepatitis C virus, and human immunodeficiency virus by analyses of dried blood spots-performance characteristics of the ARCHITECT system and two commercial assays for nucleic acid amplification. *Virology* 45, 72.
- Seow, J., Graham, C., Merrick, B., Acors, S., Pickering, S., Steel, K.J.A., et al., December 2020. Longitudinal observation and decline of neutralizing antibody responses in the three months following SARS-CoV-2 infection in humans. *Nat. Microbiol.* 5, 1598–1607.
- Shah, S.J., Barish, P.N., Prasad, P.A., Kistler, A., Neff, N., Kamm, J., et al., 2020. Clinical features, diagnostics, and outcomes of patients presenting with acute respiratory illness: a retrospective cohort study of patients with and without COVID-19. *EClinicalMedicine* 100518.
- Sun, B., Feng, Y., Mo, X., Zheng, P., Wang, Q., Li, P., et al., 2020. Kinetics of SARS-CoV-2 specific IgM and IgG responses in COVID-19 patients. *Emerg. Microbes Infect.* 9 (1), 940–948.
- Valdivia, A., Torres, I., Huntley, D., Alcaraz, M.J., Albert, E., 2021. Solano de la Asuncion C, et al. Caveats in interpreting SARS-CoV-2 IgM positive/IgG negative antibody profile in asymptomatic health care workers. *J. Med. Virol.* 93, 634–636.
- Watanabe, Y., Allen, J.D., Wrapp, D., McLellan, J.S., Crispin, M., 2020. Site-specific glycan analysis of the SARS-CoV-2 spike. *Science* 369 (6501), 330–333.
- Wayne, P.A., 2008. CLSI. User Protocol for Evaluation of Qualitative Test Performance; Approved Guideline, Second ed.
- Wrapp, D., Wang, N., Corbett, K.S., Goldsmith, J.A., Hsieh, C.L., Abiona, O., et al., 2020. Cryo-EM structure of the 2019-nCoV spike in the prefusion conformation. *Science* 367 (6483), 1260–1263.