

● REVIEW

Basics on the use of acid-sensing ion channels' inhibitors as therapeutics

Jamileh Dibas¹, Houssam Al-Saad², Adnan Dibas^{2,*}

¹ Faculty of Pharmacy, Applied University, Amman, Jordan

² North Texas Eye Research Institute, Department of Pharmacology and Neuroscience, University of North Texas Health Science Center at Fort Worth, Fort Worth, TX, USA

Funding: This work was supported by the BrightFocus Foundation and intramural grant from North Texas Health Science Center at Fort Worth (to AD).

Abstract

Since the discovery of acid-sensing ion channels in 1997, their importance in the health of neurons and other non-neuronal cells has gained significant importance. Acid-sensing ion channels play important roles in mediating pain sensation during diseases such as stroke, inflammation, arthritis, cancer, and recently migraine. More interestingly, acid-sensing ion channels may explain the sex differences in pain between males and females. Also, the ability of acid-sensing ion channel blockers to exert neuroprotective effects in a number of neurodegenerative diseases has added a new dimension to their therapeutic value. The current failure rate of ~45% of new drugs (due to toxicity issues) and saving of up to 7 years in the life span of drug approval makes drug repurposing a high priority. If acid-sensing ion channels' blockers undergo what is known as "drug repurposing", there is a great potential to bring them as medications with known safety profiles to new patient populations. However, the route of administration remains a big challenge due to their poor penetration of the blood brain and retinal barriers. In this review, the promise of using acid-sensing ion channel blockers as neuroprotective drugs is discussed.

Key Words: optic nerve; glaucoma; neurodegeneration; neuroprotection; acid; sensing; ion channel; calpain

Introduction

Under normal physiological conditions, extracellular pH (pH_o) is maintained around ~7.3 while intracellular pH (pH_i) is ~7. However, under pathological conditions such as an ischemic stroke, seizure, and inflammation, pH is greatly shifted towards acidification, a mechanism mediated in part by anaerobic glycolysis and the accumulation of lactic acid and protons. Neurons are mostly at risk of uncontrolled acidification leading to their death and permanent neurological impairment. As a defensive mechanism, cells including neurons express a family of proteins known as the acid-sensing ion channels (ASICs) that are sensitive to changes in extracellular pH reduction.

ASICs, a family of cation-selective, voltage insensitive, and proton-sensing channels belongs to a proton-gated family of the degenerin-epithelial channel family of cation channels, which are responsible for Na^+ influx (Waldmann et al., 1997). In the human genome, there are four ASIC genes that have been assigned the symbols of *Accn1* (ASIC2), *Accn2* (ASIC1), *Accn3* (ASIC3), and *Accn4* (ASIC4) (Lin et al., 2015). A novel *Accn5* encodes a related channel, BLNAC/BASIC that is however different from ASICs (Lin et al., 2015). ASIC1 is the most abundant ASIC subunit in the mammalian central nervous system (Chen et al., 1998). Similar to ASIC1a, ASIC3 has equal sensitivity to H^+ however; it is mainly expressed in peripheral nervous system (Waldmann et al., 1997).

ASICs form trimeric ion channels composed of either identical (homotrimeric) or different (heterotrimeric) subunits. ASICs are permeable to monovalent cations ($\text{Na}^+ > \text{K}^+$) and protons (Waldmann et al., 1997), and some ASICs (ASIC1a homomeric channels and ASIC1a/2b heteromeric channels) are also permeable to divalent cations such as calcium, suggesting

that they could play a particularly important role in intracellular signaling as well as membrane excitability (Stankowska et al., 2018). Interestingly, while both ASIC1a and ASIC3 are active at neutral pH (7.1–7.3), ASIC2a is only active at low pH (~4.5) (Hesselager et al., 2004).

ASIC1 Blockers are Potential Therapeutics

ASICs have been associated with many neurodegenerative disorders and recent observations suggest that ASICs are major contributors to axonopathy in such pathologies. The inhibition of ASICs is neuroprotective in stroke, Huntington's disease (Walker, 2007), multiple sclerosis (Vergo et al., 2011), and Parkinson's disease (Arias et al., 2008). List of diseases where ASIC blockers exerted neuroprotective effects is shown in **Figure 1A**. Using optic nerve crush in rats (Stankowska et al., 2018) and ischemia/reperfusion (retinal stroke model) in mice (Dibas et al., 2018), we demonstrated that blockers of ASIC1 such as amiloride (nonselective) and psalmotoxin-1 (selective ASIC1a blocker), exerted neuroprotective effects on retinal ganglion cells. In both models, an upregulation of ASIC1 and ASIC2 was observed (ASIC1 significantly increased in optic nerve extracts and ASIC1 and ASIC2 increased in retinal ganglion cells in ischemia/reperfusion model). Not surprisingly, acidification increased intracellular Ca^{2+} ($[\text{Ca}^{2+}]_i$) in isolated primary rat retinal ganglion cells, an effect attenuated by psalmotoxin-1 (Stankowska et al., 2018). We have shown and others that ASIC upregulation and activation can induce extracellular Ca^{2+} influx and increase in intracellular Ca^{2+} (Stankowska et al., 2018). ASIC1a-induced influx of extracellular Ca^{2+} mediating ischemic neuronal death, can be prevented by either reducing extracellular Ca^{2+} or ASIC1a inhibitors (psalmotoxin-1 and

*Correspondence to:

Adnan Dibas, PhD,
adnan.dibas@unthsc.edu.

doi: 10.4103/1673-5374.245466

Received: June 28, 2018

Accepted: October 17, 2018

amiloride) (Xiong et al., 2004; Yermolaieva et al., 2004; Gao et al., 2005). Elevated intracellular Ca^{2+} is a known activator of the calcium-dependent calpains. Activation of calpain 1 *via* autolysis plays a key role in mediating neuronal apoptosis as it cleaves essential proteins in neurons such as fodrins. Myelin basic proteins and axonal neurofilament protein are also known substrates for calpain and their loss greatly affects structure of neurons (Li and Banik, 1995; Das et al., 2013). Calpains are involved in retinal neurodegeneration in a number of injury models (Wu et al., 2004; Oka et al., 2006; McKernan et al., 2007; Huang et al., 2010).

In optic nerve crush in rats and ischemia/reperfusion (retinal stroke model) in mice, calpain-1 activation was evident as increased fodrin cleavage was detected. Heat shock protein (HSP)70, a known neuroprotective protein and a substrate for calpain, was degraded following retinal ischemic stroke (Dibas et al., 2018). HSP70 induction or overexpression has been shown to be neuroprotective in retinal ganglion cells in an intraocular pressure-model and during optic nerve crush (Ueda et al., 1998; Park et al., 2001a). We have shown that ischemia/reperfusion induced-upregulation of ASIC1 correlated with degradation of HSP70 and the appearance of a lower band of 55 kDa. While psalmotoxin-1 pretreatment did prevent fodrin cleavage, a well-known calpain substrate, unfortunately it failed to block HSP70-induced cleavage. This inability to affect HSP70 may explain the partial rescue of retinal ganglion cells by psalmotoxin-1. HSP70 protein is considered a master switch in neuroprotection. Its mechanism of protection involves different pathways. One mechanism involves enhancement of autophagy as HSP70 stabilized lysosomes by binding to endolysosomal phospholipid, bis(monoacylglycero)phosphate, and acted as a chaperone recycling damaged proteins to lysosomes for elimination (Kirkegaard et al., 2010). HSP70 also blocked maturation of apoptosome by binding to key elements preventing the recruiting of caspases and their activation (Beere et al., 2000). Furthermore, Matsumori et al. (2005) have shown that HSP70 attenuated caspase-unrelated cellular death pathways as well. Also, HSP70 blocked c-Jun N-terminal kinase-dependent and p38 mitogen-activated protein kinase signaling pathways (Gabai et al., 1997; Park et al., 2001b). HSP70 suffered oxidization by free radicals-induced carbonylation of its Arg469 amino-acid leading to its degradation by calpains. HSP70 depletion causes lysosomal destabilization and autophagy failure that lead ultimately to neurodegeneration (Zhu et al., 2002; Oikawa et al., 2009; Sahara and Yamashima, 2010; Furukawa and Koriyama, 2016). Our data is consistent with that reported by Nakajima et al 2006, who have shown that monkey retinas subjected to hypoxia suffered from neuronal degeneration *via* calpain activation and degradation of fodrin and HSP70 (Nakajima et al., 2006).

Finally, at least three clinical trials (NCT01802489, NCT01879527 and NCT01910259) are currently underway or finished assessing blocking ASICs using amiloride (nonselective blocker) for the treatment of optic neuritis. The results from first clinical trial (NCT01902489) was recently published and although it appears that amiloride did not demonstrate any neuroprotective effects based on the scanning laser polarimetry (which measures the thickness of the peripapillary retinal fiber layer), visual evoked potentials measures were enhanced in the amiloride group. However, newer trials have been suggested

with changed window of opportunities (McKee et al., 2017). The results from NCT01879527 and NCT01910259 clinical trials have not been released yet although both supposedly ended.

The Anti-Nociceptive Effects of ASIC Blockers

Peripheral ASICs are involved in cutaneous pain. Acidification of human skin was accompanied by cutaneous pain (Steen et al., 1995), an effect that was blocked by amiloride, a nonselective ASIC blocker (Ugawa et al., 2002; Jones et al., 2004). Amiloride also exerted analgesic effects following topical application in pain models of inflammatory and postoperative pain in rodents (Kuduk et al., 2009). Interestingly, ASIC inhibition may be helpful for migraine patients as amiloride reduced headache symptoms in a human clinical trial (Holland et al., 2012). Elevated ASIC proteins have been also detected in human degenerated intervertebral discs compared to healthy tissues suggesting a role for ASICs in inflammation and osteoarthritis-induced joint pain (Cuesta et al., 2014). Not surprisingly, intra-articular administrations of anthopleura elegantissima toxin 2 (APETx2), an ASIC3 blocker, reduced pain-related behavior and inflammation-induced cartilage damage in animals (Izumi et al., 2012). Interestingly, the known non-steroid anti-inflammatory drugs (ibuprofen, aspirin and diclofenac) directly inhibited the activity of both ASIC1a and ASIC3 channels with ASIC2a being partially blocked by diclofenac (0.5 mM) (Voilley et al., 2001).

The Neuroprotective Properties of Psalmotoxin-1 and Related Peptides

Various ASIC inhibitors have also been isolated from venoms. Psalmotoxin-1, a toxin isolated from the venom of tarantula spider, is specific to ASIC1a homomeric and ASIC1a/2b heteromeric channels has exerted neuroprotective effects in several injury models including; ischemic stroke (Xiong et al., 2004), porcine models of cerebral ischemia (Yang et al., 2011), ischemic stroke in conscious hypertensive rats (McCarthy et al., 2015), optic nerve crush in rats (Stankowska et al., 2018), and retinal stroke model in mice (Dibas et al., 2018). Hi1a is a recently discovered ASIC1a blocker with almost double the size of PcTx1 (psalmotoxin-1; 75 amino acids) (Chassagnon et al., 2017), is more potent at inhibiting ASIC1a than PcTx1 with an estimated half maximal inhibitory concentration (IC50) of ~400 pM. Chassagnon et al., 2017 have shown that intracerebroventricular injection of Hi1a (2 ng/kg) exerted significant neuroprotective effects as late as 8 hours post-injury. Also, novel peptides known as mambalgins 1, 2, and 3 isolated from the venom of mamba snakes are also potent inhibitors of ASIC1a as homotrimer or heterotrimers. Mambalgin-1 inhibited currents from ASIC1a and ASIC1b homomers (ASIC1a and ASIC1b are isoforms of ASIC1) as well as from heteromers of ASIC1a-ASIC2a, ASIC1a-ASIC2b or ASIC1a-ASIC1b (ASIC2a and ASIC2b are isoforms of ASIC2) (Diochot et al., 2012; Mourier et al., 2016). APETx2, on the other hand, inhibits ASIC3 containing channels (Chagot et al., 2005). However, to date, there are no known selective ASIC2a/2b inhibitors.

Does ASICs' Differential Gene Expression Explain Female Over Sensitivity

Overwhelming evidence is suggesting sex-based disparities

between males and females with a clear majority of chronic patients being females (Mogil, 2012). Women with knee osteoarthritis have greater pain than men (Sluka et al., 2012). Women also demonstrate higher pain sensitivity and prevalence of chronic visceral pain conditions such as fibromyalgia, migraine, and functional gastrointestinal disorders (Unruh, 1996). The exact mechanisms for why women constitute a large majority of chronic patients are not known. While psychological, hormonal, and genetic differences have been suggested to explain reasons making women more sensitive to pain than men and why women endure more pain post operationally and experience different outcomes to therapeutical interventions, ASICs's differential expression between both genders may play a role.

Studies have shown that female bladders are more sensitive to acidic injuries than male bladders. ASIC2 (which is only activated at acidic pH of ~4.5), is expressed at almost double the levels in female muscle bladder compared to male mouse bladders (Kobayashi et al., 2009), which could possibly be a contributor to the increased susceptibility in females. Male mice injected with 2-guanidine-4-methylguanidine (an ASIC3 agonist), in their paws licked their 2-guanidine-4-methylguanidine-injected paw ~34% less than female mice suggesting a clear difference in pain sensitivity between genders (Lzurietia Munoz et al., 2018). However, there are no current studies on ASICs's expression in different tissues in males and females although this could help explain the substantial sex differences in clinical and experimental pain outcomes for both genders. A list of diseases where ASIC blockers may exert sex-based anti-nociceptive effects is shown in Figure 1B.

Future Directions and Limitations

Although ASICs's inhibitors are known for their pain-attenuation or control as well diuretics (in the case of amiloride), their promising neuroprotective effects exerted by ASIC1a selective blockers are opening a new exciting field in the fight against neurodegenerative diseases. The introduction of ASIC1a targeted agents (e.g., psalmotoxin-1, H1a, and mambalgins) has raised the question whether alternate clinical trial designs and window trials, are better suited to evaluate such new drugs. However, there are limitations due to lack of delivery of such inhibitors without invasive techniques due to the inability of penetrating the blood-brain barrier and the retinal blood barrier. Blood-brain barrier is impermeable to almost 98% of small molecules (> 600 Daltons) and impermeable to large molecules. Current procedures of delivering drugs to brain such as intracerebroventricular, intracerebral, and intrathecal injections, require

specialized experience and not only carry high risk of infections but also cannot be repeatedly used in patients. Similarly, intravitreal injection although successful in delivering drugs to the retina, has the risk of side effects and limitation on the number of times to be used in the human eyes. However, intranasal administration provides a non-invasive route for central nervous system drug delivery that successfully bypassed the blood-brain barrier. There are 102 completed clinical trials that assessed the intranasal administration delivery of numerous drugs with promising results (clinicaltrials.gov). There is a great difference in brain deposits of psalmotoxin-1 when injected intravenously vs. intranasal administration. Following intravenously administration of radiolabeled psalmotoxin-1, data clearly indicated that psalmotoxin-1 is unlikely to cross the blood-brain barrier in any appreciable amount. To the contrary, using intranasal administration of 5 µg psalmotoxin-1, ~1.3 ng (~0.03%) was detected in the olfactory bulb and many brain compartments (Er, 2017). McCarthy et al. (2015) reported that intracerebroventricular administration of psalmotoxin-1 at 1 ng/kg (~25 pg for a 25 g mouse) 2 hours post stroke exerted neuroprotective effects in mice. More interestingly, Pignataro et al. (2007) administered psalmotoxin-1 (intranasal administration) at 25 ng/mouse, also provided neuroprotection when given within the first 4 hours post brain ischemic stroke. Finally, it remains of high importance to evaluate any role for ASICs in gender-based pain sensation and the potential of development of masculinity/feminine-dependent therapy.

Author contributions: Article writing: JD, HAS and AD.

Conflicts of interest: None declared.

Financial support: This work was supported by the BrightFocus Foundation and intramural grant from University of North Texas Health Science Center at Fort Worth (to AD).

Copyright license agreement: The Copyright License Agreement has been signed by all authors before publication.

Plagiarism check: Checked twice by iThenticate.

Peer review: Externally peer reviewed.

Open access statement: This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

Open peer reviewer: Steven Levy, MD Stem Cells, USA.

Additional file: Open peer review report 1.

References

Arias RL, Sung ML, Vasylyev D, Zhang MY, Albinson K, Kubek K, Kagan N, Beyer C, Lin Q, Dwyer JM, Zaleska MM, Bowlby MR, Dunlop J, Monaghan M (2008) Amiloride is neuroprotective in an MPTP model of Parkinson's disease. *Neurobiol Dis* 31:334-341.

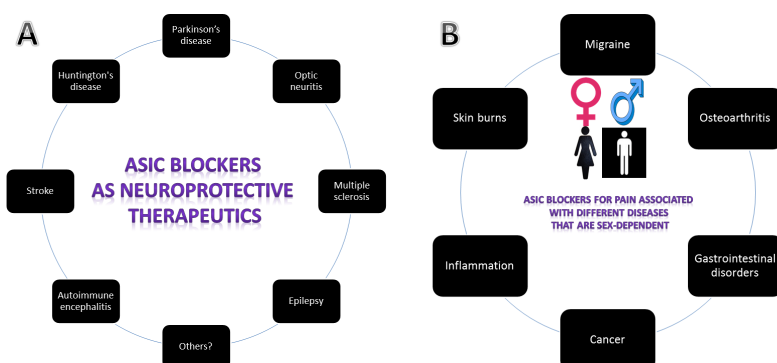


Figure 1 The promising role of ASIC blockers as therapeutics.

(A) List of diseases where ASIC blockers exerted neuroprotective effects. (B) List of diseases where ASIC blockers may exert sex-based anti-nociceptive effects. ASICs: Acid-sensing ion channel.

- Beere HM, Wolf BB, Cain K, Mosser DD, Mahboubi A, Kuwana T, Taylor P, Morimoto RI, Cohen GM, Green DR (2000) Heat-shock protein 70 inhibits apoptosis by preventing recruitment of procaspase-9 to the Apaf-1 apoptosome. *Nat Cell Biol* 2:469-475.
- Chagot B, Escoubas P, Diochot S, Bernard C, Lazdunski M, Darbon H (2005) Solution structure of APETx2, a specific peptide inhibitor of ASIC3 proton-gated channels. *Protein Sci* 14:2003-2010.
- Chassagnon IR, McCarthy CA, Chin YK, Pineda SS, Keramidis A, Mobli M, Pham V, De Silva TM, Lynch JW, Widdop RE, Rash LD, King GF (2017) Potent neuroprotection after stroke afforded by a double-knot spider-venom peptide that inhibits acid-sensing ion channel 1a. *Proc Natl Acad Sci U S A* 114:3750-3755.
- Chen CC, England S, Akopian AN, Wood JN (1998) A sensory neuron-specific, proton-gated ion channel. *Proc Natl Acad Sci U S A* 95:10240-10245.
- Cuesta A, Del Valle ME, Garcia-Suarez O, Vina E, Cabo R, Vazquez G, Cobo JL, Murcia A, Alvarez-Vega M, Garcia-Cosamalon J, Vega JA (2014) Acid-sensing ion channels in healthy and degenerated human intervertebral disc. *Connect Tissue Res* 55:197-204.
- Das A, Guyton MK, Smith A, Wallace Gt, McDowell ML, Matzelle DD, Ray SK, Banik NL (2013) Calpain inhibitor attenuated optic nerve damage in acute optic neuritis in rats. *J Neurochem* 124:133-146.
- Dibas A, Millar C, Al-Farra A, Yorio T (2018) Neuroprotective effects of psalmotoxin-1, an acid-sensing ion channel (ASIC) inhibitor, in ischemia reperfusion in mouse eyes. *Curr Eye Res* 43:921-933.
- Diochot S, Baron A, Salinas M, Douguet D, Scarzello S, Dabert-Gay AS, Debaule D, Friend V, Alloui A, Lazdunski M, Lingueglia E (2012) Black mamba venom peptides target acid-sensing ion channels to abolish pain. *Nature* 490:552-555.
- Er SY (2017) Biotechnological applications of spider venom peptides. Brisbane, Australia: The University of Queensland.
- Furukawa A, Koriyama Y (2016) A role of heat shock protein 70 in photoreceptor cell death: potential as a novel therapeutic target in retinal degeneration. *CNS Neurosci Ther* 22:7-14.
- Gabai VL, Meriin AB, Mosser DD, Caron AW, Rits S, Shifrin VI, Sherman MY (1997) Hsp70 prevents activation of stress kinases. A novel pathway of cellular thermotolerance. *J Biol Chem* 272:18033-18037.
- Gao J, Duan B, Wang DG, Deng XH, Zhang GY, Xu L, Xu TL (2005) Coupling between NMDA receptor and acid-sensing ion channel contributes to ischemic neuronal death. *Neuron* 48:635-646.
- Hesselager M, Timmermann DB, Ahning PK (2004) pH Dependency and desensitization kinetics of heterologously expressed combinations of acid-sensing ion channel subunits. *J Biol Chem* 279:11006-11015.
- Holland PR, Akerman S, Andreou AP, Karsan N, Wemmie JA, Goadsby PJ (2012) Acid-sensing ion channel 1: a novel therapeutic target for migraine with aura. *Ann Neurol* 72:559-563.
- Huang W, Fileta J, Rawe I, Qu J, Grosskreutz CL (2010) Calpain activation in experimental glaucoma. *Invest Ophthalmol Vis Sci* 51:3049-3054.
- Izumi M, Ikeuchi M, Ji Q, Tani T (2012) Local ASIC3 modulates pain and disease progression in a rat model of osteoarthritis. *J Biomed Sci* 19:77.
- Izurrieta Munoz H, Gonzales EB, Sumien N (2018) Effects of creatine supplementation on nociception in young male and female mice. *Pharmacol Rep* 70:316-321.
- Jones NG, Slater R, Cadiou H, McNaughton P, McMahon SB (2004) Acid-induced pain and its modulation in humans. *J Neurosci* 24:10974-10979.
- Kirkegaard T, Roth AG, Petersen NH, Mahalka AK, Olsen OD, Moilanen I, Zylitz A, Knudsen J, Sandhoff K, Arenz C, Kinnunen PK, Nylandsted J, Jaattela M (2010) Hsp70 stabilizes lysosomes and reverts Niemann-Pick disease-associated lysosomal pathology. *Nature* 463:549-553.
- Kobayashi H, Yoshiyama M, Zakoji H, Takeda M, Araki I (2009) Sex differences in the expression profile of acid-sensing ion channels in the mouse urinary bladder: a possible involvement in irritative bladder symptoms. *BJU Int* 104:1746-1751.
- Kuduk SD, Di Marco CN, Chang RK, Dipardo RM, Cook SP, Cato MJ, Jovanovska A, Urban MO, Leitl M, Spencer RH, Kane SA, Bilodeau MT, Hartman GD, Bock MG (2009) Amiloride derived inhibitors of acid-sensing ion channel-3 (ASIC3). *Bioorg Med Chem Lett* 19:2514-2518.
- Li Z, Banik NL (1995) The localization of mcalpain in myelin: immunocytochemical evidence in different areas of rat brain and nerves. *Brain Res* 697:112-121.
- Lin SH, Sun WH, Chen CC (2015) Genetic exploration of the role of acid-sensing ion channels. *Neuropharmacology* 94:99-118.
- Matsumori Y, Hong SM, Aoyama K, Fan Y, Kayama T, Sheldon RA, Vexler ZS, Ferriero DM, Weinstein PR, Liu J (2005) Hsp70 overexpression sequesters AIF and reduces neonatal hypoxic/ischemic brain injury. *J Cereb Blood Flow Metab* 25:899-910.
- McCarthy CA, Rash LD, Chassagnon IR, King GF, Widdop RE (2015) PcTx1 affords neuroprotection in a conscious model of stroke in hypertensive rats via selective inhibition of ASIC1a. *Neuropharmacology* 99:650-657.
- McKee JB, Cottrill CL, Elston J, Epps S, Evangelou N, Gerry S, Kennard C, Kong Y, Koelewyn A, Kueker W, Leite MI, Palace J, Craner M (2017) Amiloride does not protect retinal nerve fibre layer thickness in optic neuritis in a phase 2 randomised controlled trial. *Mult Scler* doi:10.1177/1352458517742979.
- McKernan DP, Guerin MB, O'Brien CJ, Cotter TG (2007) A key role for calpains in retinal ganglion cell death. *Invest Ophthalmol Vis Sci* 48:5420-5430.
- Mogil JS (2012) Sex differences in pain and pain inhibition: multiple explanations of a controversial phenomenon. *Nat Rev Neurosci* 13:859-866.
- Mourier G, Salinas M, Kessler P, Stura EA, Leblanc M, Tepshi L, Besson T, Diochot S, Baron A, Douguet D, Lingueglia E, Servent D (2016) Mambalgin-1 pain-relieving peptide, stepwise solid-phase synthesis, crystal structure, and functional domain for acid-sensing ion channel 1a inhibition. *J Biol Chem* 291:2616-2629.
- Nakajima E, David LL, Bystrom C, Shearer TR, Azuma M (2006) Calpain-specific proteolysis in primate retina: Contribution of calpains in cell death. *Invest Ophthalmol Vis Sci* 47:5469-5475.
- Oikawa S, Yamada T, Minohata T, Kobayashi H, Furukawa A, Tada-Oikawa S, Hiraku Y, Murata M, Kikuchi M, Yamashima T (2009) Proteomic identification of carbonylated proteins in the monkey hippocampus after ischemia-reperfusion. *Free Radic Biol Med* 46:1472-1477.
- Oka T, Tamada Y, Nakajima E, Shearer TR, Azuma M (2006) Presence of calpain-induced proteolysis in retinal degeneration and dysfunction in a rat model of acute ocular hypertension. *J Neurosci Res* 83:1342-1351.
- Park HS, Lee JS, Huh SH, Seo JS, Choi EJ (2001a) Hsp72 functions as a natural inhibitory protein of c-Jun N-terminal kinase. *EMBO J* 20:446-456.
- Park KH, Cozier F, Ong OC, Caprioli J (2001b) Induction of heat shock protein 72 protects retinal ganglion cells in a rat glaucoma model. *Invest Ophthalmol Vis Sci* 42:1522-1530.
- Pignataro G, Simon RP, Xiong ZG (2007) Prolonged activation of ASIC1a and the time window for neuroprotection in cerebral ischaemia. *Brain* 130:151-158.
- Sahara S, Yamashima T (2010) Calpain-mediated Hsp70.1 cleavage in hippocampal CA1 neuronal death. *Biochem Biophys Res Commun* 393:806-811.
- Sluka KA, Berkley KJ, O'Connor MI, Nicoletta DP, Enoka RM, Boyan BD, Hart DA, Resnick E, Kwok CK, Tosi LL, Coutts RD, Kohrt WM (2012) Neural and psychosocial contributions to sex differences in knee osteoarthritic pain. *Biol Sex Differ* 3:26.
- Stankowska DL, Mueller BH, 2nd, Oku H, Ikeda T, Dibas A (2018) Neuroprotective effects of inhibitors of acid-sensing ion channels (ASICs) in optic nerve crush model in rodents. *Curr Eye Res* 43:84-95.
- Steen KH, Steen AE, Reeh PW (1995) A dominant role of acid pH in inflammatory excitation and sensitization of nociceptors in rat skin, in vitro. *J Neurosci* 15:3982-3989.
- Ueda J, Sawaguchi S, Hanyu T, Yaoeda K, Fukuchi T, Abe H, Ozawa H (1998) Experimental glaucoma model in the rat induced by laser trabecular photocoagulation after an intracameral injection of India ink. *Jpn J Ophthalmol* 42:337-344.
- Ugawa S, Ueda T, Ishida Y, Nishigaki M, Shibata Y, Shimada S (2002) Amiloride-blockable acid-sensing ion channels are leading acid sensors expressed in human nociceptors. *J Clin Invest* 110:1185-1190.
- Unruh AM (1996) Gender variations in clinical pain experience. *Pain* 65:123-167.
- Vergo S, Craner MJ, Etzspenberger R, Attfield K, Friese MA, Newcombe J, Esiri M, Fugger L (2011) Acid-sensing ion channel 1 is involved in both axonal injury and demyelination in multiple sclerosis and its animal model. *Brain* 134:571-584.
- Voilley N, de Weille J, Mamet J, Lazdunski M (2001) Nonsteroid anti-inflammatory drugs inhibit both the activity and the inflammation-induced expression of acid-sensing ion channels in nociceptors. *J Neurosci* 21:8026-8033.
- Waldmann R, Champigny G, Bassilana F, Heurteaux C, Lazdunski M (1997) A proton-gated cation channel involved in acid-sensing. *Nature* 386:173-177.
- Walker FO (2007) Huntington's disease. *Lancet* 369:218-228.
- Wu HY, Tomizawa K, Oda Y, Wei FY, Lu YF, Matsushita M, Li ST, Moriwaiki A, Matsui H (2004) Critical role of calpain-mediated cleavage of calcineurin in excitotoxic neurodegeneration. *J Biol Chem* 279:4929-4940.
- Xiong ZG, Zhu XM, Chu XP, Minami M, Hey J, Wei WL, MacDonald JF, Wemmie JA, Price MP, Welsh MJ, Simon RP (2004) Neuroprotection in ischemia: blocking calcium-permeable acid-sensing ion channels. *Cell* 118:687-698.
- Yang ZJ, Ni X, Carter EL, Kibler K, Martin LJ, Koehler RC (2011) Neuroprotective effect of acid-sensing ion channel inhibitor psalmotoxin-1 after hypoxia-ischemia in newborn piglet striatum. *Neurobiol Dis* 43:446-454.
- Yermolaieva O, Leonard AS, Schnizler MK, Abboud FM, Welsh MJ (2004) Extracellular acidosis increases neuronal cell calcium by activating acid-sensing ion channel 1a. *Proc Natl Acad Sci U S A* 101:6752-6757.
- Zhu Y, Ohlemiller KK, McMahan BK, Gidday JM (2002) Mouse models of retinal ischemic tolerance. *Invest Ophthalmol Vis Sci* 43:1903-1911.