



Perioperative “remote” acute lung injury: recent update

Zhaosheng Jin, Ka Chun Suen, Daqing Ma[✉]

Anaesthetics, Pain Medicine and intensive Care, Department of Surgery and Cancer, Faculty of Medicine, Imperial College London, Chelsea & Westminster Hospital, London SW10 9NH, UK.

Abstract

Perioperative acute lung injury (ALI) is a syndrome characterised by hypoxia and chest radiograph changes. It is a serious post-operative complication, associated with considerable mortality and morbidity. In addition to mechanical ventilation, remote organ insult could also trigger systemic responses which induce ALI. Currently, there are limited treatment options available beyond conservative respiratory support. However, increasing understanding of the pathophysiology of ALI and the biochemical pathways involved will aid the development of novel treatments and help to improve patient outcome as well as to reduce cost to the health service. In this review we will discuss the epidemiology of peri-operative ALI; the cellular and molecular mechanisms involved on the pathological process; the clinical considerations in preventing and managing perioperative ALI and the potential future treatment options.

Keywords: acute lung injury, intraoperative care/adverse effects, postoperative complications, inflammation, anaesthetics, general, fluid therapy

Introduction

The term acute lung injury (ALI) was first introduced in 1994 by the American–European Consensus Conference Committee^[1]. It is defined as acute onset hypoxia with $\text{PaO}_2/\text{FiO}_2$ between 200–300; presence of bilateral infiltrates on the chest radiograph; in absence of pulmonary hypertension or other cardiac pathologies. The term was coined to identify cases which are not severe enough to fall into the criteria of Acute Respiratory Distress Syndrome (ARDS), for the ease of identification and further research. Since then, ALI has attracted significant attention in both clinical and laboratory research. Interestingly, general abdominal surgery carries similar ALI risk as general thoracic surgeries, and animal studies of remote trauma and non-pulmonary transplant demonstrates features of ALI,

suggesting that peri-operative ALI is at least partially attributable to remote injury. In this article, we will discuss the epidemiology of perioperative ALI, as well as the biochemical mechanisms and clinical considerations of perioperative ALI due to remote injury.

Epidemiology

In the general surgical population, the incidence of ARDS is reported to be 0.2%^[2]. Indeed, it is thought that patients who undergo elective minor orthopaedic and pelvic surgeries are at minimal risk of developing perioperative ALI^[3–4]. Thoracic and abdominal surgeries are associated with higher risks. In patients without significant risk factors, the average ALI incidence in thoracic and abdominal surgery is 1.3–

[✉] Corresponding author: Daqing Ma, MD, PhD, Anaesthetics, Pain Medicine and Intensive Care, Department of Surgery and Cancer, Faculty of Medicine, Imperial College London Chelsea and Westminster Hospital, London, UK. Tel/Fax: (0044) 020 3315 8495/(0044) 020 3315 5109, Email: d.ma@imperial.ac.uk.

Received 28 April 2016, Revised 01 June 2016, Accepted 16 July 2016, Epub 10 September 2016

CLC number: R614, Document code: A

The authors reported no conflict of interests.

This is an open access article under the Creative Commons Attribution (CC BY 4.0) license, which permits others to distribute, remix, adapt and build upon this work, for commercial use, provided the original work is properly cited.

4%^[5-7], with 1.2%-2.3% incidence of respiratory failure^[8-9]. Interestingly, despite the surgical manipulation of the lung, the incidence of perioperative ALI in lobectomy and pneumonectomy is only 2.8%-5%. In cardiac and aortic surgery, the incidence of perioperative ALI is also around 5%; however, this can increase to as high as 28%-35% in high risk procedures^[10-11]. Esophagectomy is associated with a high incidence of ALI and reported to be between 16%-41%^[12-14]. There is not enough published data to create a comprehensive list of surgeries and associated ALI risks, but recommendations for a surgery related risk stratification system have been proposed by Kor *et al.*^[11].

The pre-morbid state of the patient plays a significant role of the development of perioperative ALI. Two cohort studies involving a total of 7,126 patients with pre-operative risk factors for developing ALI reported an incidence of 6.8%-7.5%^[11,15]. A number of studies have looked into pre-operative patient parameters and constructed ALI prediction models; however, their application in clinical practice has not been reported. Emergency surgery is the most consistently reported predictor of ALI; other frequently reported predictors of ALI include age, pre-operative renal failure, chronic obstructive pulmonary disease (COPD) and pneumonia, hypoalbuminaemia and alcohol consumption. These four models employed different markers of respiratory distress (desaturation, tachypnoea, dysapnoea and oxygen requirement), all of which are statistically significant predictors from the literature that are listed in **Table 1**^[3,11,15-18].

Development of perioperative ALI is associated with significantly worse outcomes (See **Fig. 1** for summary

of ALI clinical outcomes). Patients with ALI are twice as likely to be admitted to intensive care unit (ITU) and require an average of an 8 day stay (which is 4-8 times longer than those without ALI). This includes a relative risk ratio of 2.5 for mechanical ventilation compared to normal ventilation, in which patients spend on average 6 days on mechanical ventilation. This is 6 times longer than the duration of non ALI patients. This prolongs the duration of the hospital stay to an average of 15-20 days, up to 2.5 times longer compared to patients without ALI^[7,15]. The prolonged ITU and hospital stay is not without its own risks. The total in-patient mortality rate is between 22 to 24%, which is 5-10 times higher than comparative population without ALI. However, most of this is accounted for by ITU mortality of 20%^[15,19-20]. Even when the patient recovers from the episode and is discharged, their long term prognosis is still significantly worse than those without ALI. A two year follow up study reported that survivors of perioperative ALI required on average of 2 episodes of readmission and 6 days of hospital stay per year while another study reported 30 day mortality to be up to 30% and 90 day mortality to be 55%^[16,21].

Apart from the significant mortality and loss of quality of life from the extended ITU and hospital stay experienced by patients, perioperative ALI is also associated with a significant cost to the health care system. In a study based on American health care expenditure in 2013, it has been estimated that initial hospital management of ALI costs approximately \$100,000, with another \$35,000 spent on two years of follow up treatment^[21].

In summary, perioperative ALI can be a common

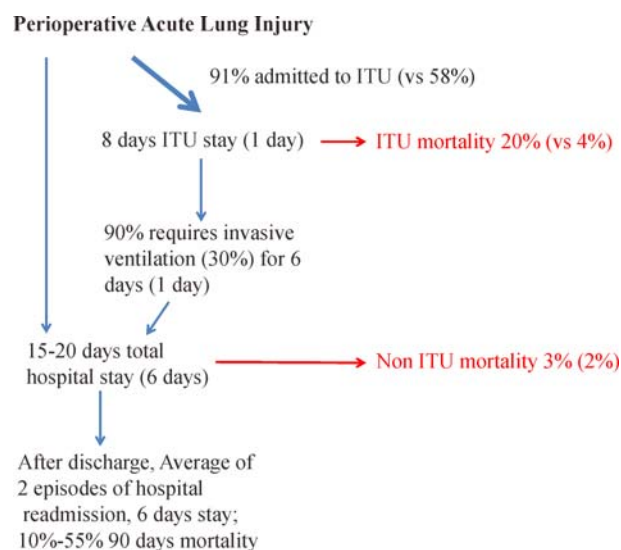


Fig. 1 "Roadmap" of postoperative acute lung injury (ALI). Illustrating the higher rate of invasive ventilation, ITU admission; longer length of ITU and hospital stay; re-admission; as well as the much higher ITU and 90 day mortality, non-ALI cohort data presented in brackets for comparison ^[7,15-16,19-21].

Table 1 Summary of tested parameters from 6 acute lung injury (ALI) predictive models

	Tested and reported statistical significance* (<i>n</i> = 6)	Tested and reported no statistical significance* (<i>n</i> = 6)
General		
Gender	1	1
Age	2	-
Functionally not independent	1	-
Weight loss	1	-
Alcohol	2	-
Smoking	1	2
Admission not from home	1	-
Obesity	1	-
Past medical history, pulmonary		
COPD	2	-
preoperative pneumonia	2	-
preoperative dysapnoea	1	-
preoperative desaturation	1	1
preoperative tachypnoea	2	-
Aspiration	1	-
FiO ₂	1	-
Others		
ASA grade	1	1
preoperative sepsis	1	1
preoperative renal failure	2	-
Cirrhosis	1	-
Albumin	2	-
Shock	1	-
preoperative anaemia	1	-
Advanced cancer	1	-
Perioperative factors		
Emergency surgery	5	-
Surgery duration	1	-
Fluid infusion	1	-
Blood Transfusion	1	-

Note: *: number of studies which tested the parameter and demonstrated statistical significance. **: number of studies which tested the parameter and reported that there is no statistical significance [3,11,15-18]. COPD: chronic obstructive pulmonary disease.

complication in certain patient groups, and is associated a significantly poor outcome and high treatment cost. Currently, most cases of perioperative ALIs are managed conservatively and, therefore, further investigation into its pathophysiology and treatment is very necessary.

Molecular mechanism of acute lung injury

From the literature reviewed, most of the human studies and an animal model of ALI reported similar presentations of clinical, histological and biochemical

changes (**Fig. 2**), despite the varied aetiology of lung injury in the studies. Here, we will attempt to summarise the biochemical pathways found to be involved with the pathogenesis of acute lung injury.

Inflammation

Histologically, ALI is typically associated with increased neutrophil infiltration, increased vascular permeability, and increased tissue oedema, all of which are characteristic features of an inflammatory process. Both human studies and animal models of ALI invariably reported increased production of systemic

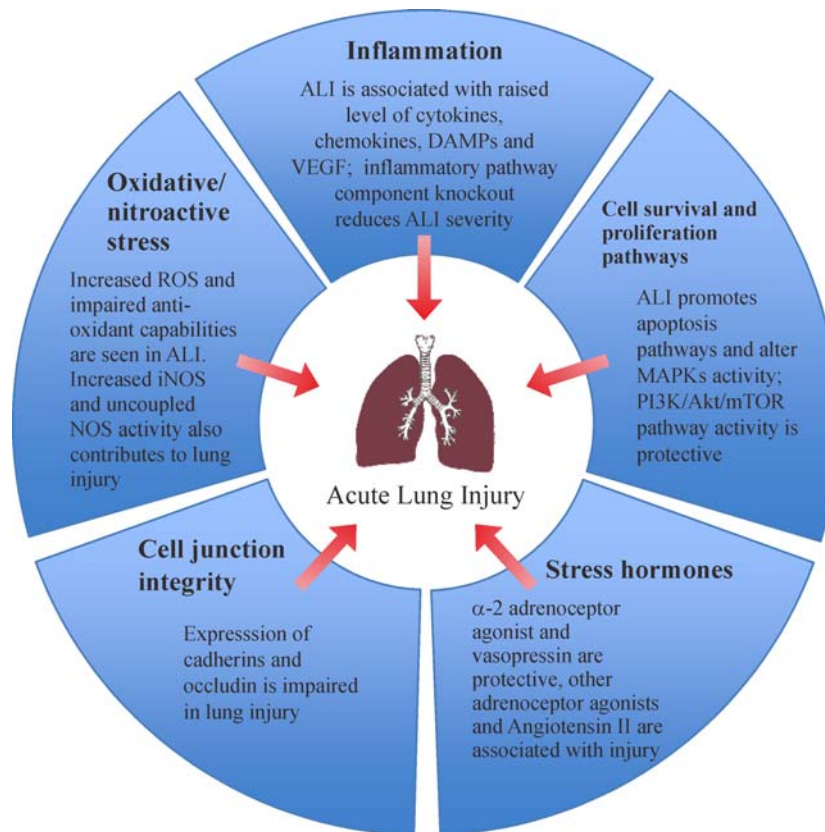


Fig. 2 Summary of molecular mechanism of acute lung injury (ALI). ALI is characterized by inflammation, increased oxidative and nitroactive stress, impaired cell junction integrity, release of stress hormones, and altered cell survival and proliferation pathways. ACE: angiotensin converting enzyme, DAMPs: damage-associated molecular pattern molecules; iNOS: inducible nitric oxide synthase; MCP1: monochemotactic protein 1; MAPK: mitogen-activated protein kinases; MIP1: macrophage inflammatory protein 1; ROS: reactive oxygen species; VEGF: vascular endothelial derived growth factor.

and local cytokines (IL-1 β , IL-6, IL-8, IL-10, and TNF- α), chemokines (CXCL1, CINC1, MIP1, and MCP1) and other immune mediators. This occurs without the presence of endotoxin in remote organ injury due to trauma, ischaemia and transplant^[22-25]. The inflammatory mediators are thought to play a significant role in the pathogenesis of ALI, as eliminating various parts of the inflammatory cascade alleviates the severity of lung injury^[26-30].

Tissue injury, whether of the lung or remote organ, leads directly to cell injury and necrosis. In this process, TNF- α and IL-1 β are released^[31]. This has several effects. For example, TNF- α increases vascular permeability, which results in an increased recruitment of neutrophils and macrophages. It can inactivate I κ B thorough phosphorylation, thereby lifting the inhibition of NF- κ B, which further upregulates the expression of IL-1 β ^[32-33]. Increased levels of TNF- α and IL-1 β recruit more macrophages and neutrophils, as well as promoting their survival. IL-1 β is also able to upregulate the production of acute phase proteins, such as CRP and complement, as well as promote the expression of other cytokines, chemokines and adhesion molecules^[34].

Among the cytokines upregulated by TNF- α and IL-1 β are IL-6 and IL-8. IL-6, which is produced mainly by endothelial cells, modulates the immune response by altering the expression of neutrophil, macrophage and T cell chemokines (including CXCL1, MIP1, MCP1 and CCL5, all of which are reported to be associated with the pathogenesis of ALI), and also upregulates adhesion molecules such as selectin ICAM-1 and VCAM-1. IL-8, which is a chemoattractant produced by macrophages, induces neutrophil chemotaxis, and upregulates the phagocytic function of neutrophils^[35].

Interestingly, although elevation in IL-10 is reported by a number of studies in association with ALI, it is actually an anti-inflammatory cytokine, which inhibits TNF- α , IL-1 β , and NF- κ B, as well as causing de-adherence of macrophages^[36]. It is likely that its role in ALI is more regulatory in nature. This is difficult to prove, however, as most studies to date demonstrate an increased level of IL-10 with ALI mimetics and a reduced level of IL-10 with ALI treatment^[37].

Toll like receptors are pattern recognition receptors, which are usually known for their affinity to bacterial endotoxins. However, studies have shown that knockout

of toll like receptor 4 (TLR4), as well as its adaptor protein MyD88 is associated with reduction in ALI severity in apparently aseptic conditions^[26-28]. More recent studies have alluded to the possibility that as well as bacterial molecular patterns, TLR4 is also able to identify endogenous ligands associated with tissue injury^[38]. TLR-4/MyD88 complex acts through downstream molecule IKK, which phosphorylates and inactivates I κ B, thereby lifting inhibition on NF- κ B^[38-39].

Vascular endothelial growth factor (VEGF) is a group of growth factors which promotes angiogenesis and has some chemotactic functions. It has been demonstrated that VEGF expression is upregulated by various inflammatory mediators described above, including IL-1 β , IL-6 and TNF- α ^[40-41]. It has been reported that administration of VEGF increases lung vasculature permeability, and VEGF inhibition alleviates lung injury^[42-44]; however, reports of VEGF attenuating lung injury also exist^[45-46]. It is possible that the effect of VEGF depends on the timing in relation to the injury.

Oxidative and nitrosative stress

Studies have shown that remote ALI is associated with increased oxidative stress markers, and treatments which reverse ALI are associated with reduced level of oxidative stress, suggesting that oxidative stress may be involved in the pathogenesis of ALI^[47]. More directly, anti-oxidant ammonium pyrrolidinedithiocarbamate has been reported to reduce ALI after liver transplant in rats^[48]. Most of the endogenous reactive oxygen species (ROS) are produced by NADPH oxidase which is neutralised by a number of anti-oxidant enzymes such as superoxide dismutase, glutathione S transferase, catalase and heme-oxygenase. When the anti-oxidant capability of the cell is exhausted, oxidative damage induces the activation of Bcl-2, which triggers mitochondrial breakdown and apoptosis^[49]. In the context of ALI, an increase in ROS production, as well as impaired anti-oxidant capabilities are seen with remote organ ischaemia, haemorrhage, hypoxia and burns/blast trauma^[50-53].

Recently, studies have also investigated the role of nitric oxide (NO) and nitric oxide synthase (NOS) in ALI. NOS is a group of enzymes which catalyses the generation of NO from arginine; there are three isoforms of NOS in human, including neuronal isoform nNOS, endothelial isoform eNOS and inducible isoform iNOS. Whereas eNOS is constitutively expressed and is generally thought to play a protective role, iNOS expression is upregulated by cytokines such as IL-1 β and TNF- α , and is thought to contribute to tissue injury^[54]. This isoform specific effect has been demonstrated by Sedoris^[55]. While NO can act as an anti-

oxidant, excessive oxidative stress can cause accumulation of peroxynitrite, a reactive nitrogen species that results in injury through oxidation or nitration^[56]. A number of animal models of Lipopolysaccharide (LPS) that induces ALI have reported an increased level of iNOS, while treatment of ALI is associated with reduced level of iNOS^[57-59]. Similar findings have also been seen in ALI secondary to high volume ventilation, ischaemia/reperfusion injury and chemical injury^[60-62]. The role of eNOS is less clear, while eNOS knockout is associated with reduced lung injury after LPS exposure, reduced eNOS activity worsens lung injury secondary to bowel and brain ischaemic/reperfusion injury^[62-64]. This suggests that the role of NO is pathology specific and the effect is likely concentration dependent.

Asymmetric dimethylarginine (ADMA) is a structural analogue of L-arginine, which competitively binds to all isoforms of NOS. The binding causes the uncoupling of NOS, which diminishes NO production and increases ROS production^[65]. The process is usually kept in check by dimethylargininase (DDAH) which hydrolyses ADMA^[66]. Reduced DDAH activity and excessive ADMA have been reported in association with ALI, while increased DDAH activity reduces the extent of ALI^[62,64,67].

Cell survival and proliferation pathways

A number of animal models of ALI secondary to remote organ injury and other conditions have reported an altered activity of the PI3K/Akt/mTOR pathway, a series of signalling molecules which plays a vital role in cell proliferation. General anaesthetic agents exert their cytoprotective effects at least in part by upregulating this pathway. PI3K and mTOR are reported to inhibit the expression of NF- κ B; mTOR could regulate downstream molecules like HIF1 α ^[68], which upregulates the expression of antioxidative enzymes, promotes cell survival, and encourages angiogenesis; however, its role can be dual depending on elevated level as it has been reported with both improvement and deterioration of lung injury^[68-69]. One of the downstream effects of HIF-1 α is to inhibit HMGB1 release from the nucleus. HMGB1 is a chromatin protein, that acts as a damage-associated molecular pattern. It can interact with TLR4/MyD88, which upregulates NF- κ B and MAPK; it can also interact with RAGE. HMGB1 is implicated in brain trauma induced acute lung injury; it is reported that RAGE knockdown is protective against ischaemic-reperfusion, and high soluble RAGE is associated with prolonged ventilation and lung transplant failure^[70].

A number of studies have also demonstrated the role of apoptotic pathways in ALI. BAX and BAK are pro-

apoptotic members of the Bcl-2 family. This is usually kept in check through binding to other proteins or through sequestering of proteins needed for activation^[71]. ALI models are associated with increased levels of BAX and BAK, as well as reduced level of the pro-survival Bcl-2^[68,72-73]. One possible mechanism of this is oxidative stress causing the activation of BAX and BAK^[49]. Activated BAX and BAK are converted from monomers to oligomers, which form pores on the mitochondrial membranes. This facilitates translocations of cytochrome c, which activates the caspase cascade and causes apoptosis^[71]. Increased caspase activity is also seen in patients with ALI, while ALI treatments reduce caspase activities^[72-74]. However, the activation of the apoptotic pathway is likely to be a result of existing cellular insult, not the direct cause of ALI.

Changes in MAPKs are also seen in ALI models. MAPKs are a group of serine/threonine kinases which integrate stress signals and phosphorylate downstream signals to promote cell survival or cell death. There are three conventional signalling pathways, p38 MAPK, ERK1/2 and JNK. p38 MAPK has a number of pro-survival and pro-apoptotic functions; JNK has a variety of functions, including induction of cell apoptosis in response to various cellular stresses, including ischaemia/reperfusion and inflammatory cytokines such as TNF- α ; whereas ERK1/2 are thought to have mainly pro-survival functions^[75]. In addition, it is thought that MAPKs act as downstream signal of TLR to increase the expression of IL-1, IL-6 and IL-8^[76]. Increased level of phosphorylated ERK and JNK are seen in ALI models^[77-80]. It is possible that while increased JNK is related to lung injury, ERK1/2 is a protective response against the injury. However, it is not possible to confirm without more studies.

Cell junction integrity

One of the hallmarks of ALI is increased vascular permeability, which leads to oedema, protein extravasation and neutrophil infiltration. A number of molecules have been implicated in the breakdown of intercellular stability. Occludin is a polypeptide vital for the formation of tight junction, and reported to be down-regulated by remote organ ischemia reperfusion injury^[81-82]. Cadherins are polypeptides which form adherence junctions and interacts with intracellular actin. It was demonstrated that the expression of cadherin and endothelial barrier function are damaged by direct lung injury^[83-84].

Stress hormones

Perioperative state, as well as any preoperative

pathologies are frequently associated with increased physiological stress. The relationship between stress hormones and severity of lung injury is not well studied; however, recent studies are beginning to demonstrate their important in ALI pathogenesis.

Adrenaline is the hormone responsible for sympathetic activation, which is associated with a number of systemic disease states, including burn injury, sepsis and ARDS^[85-86]. It has been reported that administration of adrenaline worsens lung histology and associated inflammatory response in ALI, whereas β -adrenoceptor antagonist alleviates ALI^[22,87-88]. Dexmedetomidine is an α -2 adrenoceptor agonist, and has been found to reduce lung injury secondary to remote ischaemia/reperfusion injury, chest trauma as well as surgical pneumoperitoneum^[89-91]. The benefit is partially reversed by α -2 adrenoceptor antagonist atipamezole, which suggests that dexmedetomidine possesses protection at least in part *via* α -2 adrenoceptor against ALI^[92]. However, the benefit of α -2 antagonist in sepsis induced ALI suggests that the role of α -2 adrenoceptor is pathology specific^[88].

Another stress related hormone vasopressin has been reported to play a protective role in ALI. Administration of vasopressin reduces pulmonary oedema and airway secretion, and improves alveolar fluid clearance^[93-95]. In addition, a study of ARDS patients reported that terlipressin administration was associated with significantly better oxygenation^[96].

The renin-angiotensin system has also been linked to ALI. Angiotensin II is a peptide hormone, converted from angiotensin I by ACE. It can be further modified into angiotensin 1-7 by ACE2. It has been reported that inhibition of ACE or antagonism of angiotensin II is associated with significantly less lung injury and oedema; as well as reduced cytokine and chemokine expression^[77,97]. ACE2 and angiotensin 1-7, however, have been reported to be protective against ALI secondary to LPS, bleomycin administration and acid inhalation^[42,77,98]. The possible mechanisms include reversing vascular permeability caused by VEGF and down-regulating the pro-apoptotic mediators^[42,72,99].

BNP is a 32 amino acid peptide secreted mainly by cardiomyocytes. It is synthesized from preproBNP, which is converted to proBNP, then enzymatically cleaved into BNP and the inactive fragment NT-proBNP. In addition to modulating vascular tone and sodium homeostasis, BNP has been found to dampen inflammatory reaction and promote survival of cardiomyocytes^[100]. Although limited, studies into the role of BNP in ALI show promising results. It has been reported that in patients, ALI is associated with significantly increased level of systemic BNP, which may

represent up-regulated compensatory response^[101-102]. In animal models of ALI, administration of recombinant human BNP has been shown to reduce the severity of lung injury, as well as the associated inflammatory response and oxidative stress^[103-104]. However, more studies are needed to further validate both the diagnostic and therapeutic value of BNP in humans.

ALI secondary to remote organ injury and transplant

While undesirable, surgical manipulation invariably leads to tissue injury, and it has been observed that even in pathologies where lungs are not directly damaged, acute lung injury can ensue from remote organ injury. Perhaps the most well described aetiology of remote organ injury that induces ALI is traumatic brain injury. In TBI cases, acute lung injury is among the most common non-neuronal organ dysfunction, with an incidence of 9%^[70,105]. Severe trauma in general is also associated with a high risk of ALI^[106]. In addition, high incidence of ALI has also been reported in liver transplant and renal transplant^[107-109]. More indirectly, ALI could be reliably reproduced in animals through organ transplant and remote organ ischaemia^[24,110]. The severity of ALI correlates to the length of remote organ insult^[48,111]. These findings all support the role of lung-remote organs crosstalk in perioperative acute lung injury.

ALI causing remote organ injury has been shown to cause systemic inflammation. Animal studies of transplant related ALI consistently reported increased serum level of cytokines such as IL-1 β and TNF- α ; these cytokines are also found to be increased lung tissue^[28,53,112]. Studies have also shown that disabling part of the inflammatory pathway such as TLR knock-out, NF- κ B inhibition and preventing leucocyte adhesion can reduce the extent of lung injury and the level of cytokines in the lungs^[28,113]. This suggests that cytokines released from remote organ injury could spread to the lungs *via* the blood supply, where it activates pro-inflammatory pathways in the lungs and leads to ALI. In addition to circulating cytokines, remote organ injury can also cause the release of proinflammatory damage associated molecular pattern such as HMGB1 into the circulation, which can also activate the proinflammatory pathways in the lungs^[70].

In addition to inflammation, a number of studies have reported increased oxidative stress in animal models of ALI secondary to remote injury. Limb trauma reduces SOD activity and Glutathione, while increases the levels of hydrogen peroxide and MDA. This pattern is seen systemically in the serum as well as lung tissue. Similar pattern is also seen with transplant^[23,50] and

haemorrhagic shock^[104]. In addition, there is some evidence that oxidative stress is alleviated through over-expression of SOD and through glutathione administration^[114-115]. This suggests that oxidative stress may directly contribute towards the pathogenesis of ALI.

While not well studied, dexmedetomidine has demonstrated protective effect in myocardial and renal ischaemia/reperfusion injury^[89,92]. This suggests that abnormality in sympathetic activation may play a role in ALI secondary to remote organ injury.

Interestingly, despite the wealth of animal studies showing that intestinal ischaemic-reperfusion injury causes acute lung injury, there is no literature of similar condition in humans. Indeed, the incidence of perioperative ALI is not well presented in the literature outside the topic of thoracic surgery, and may pose an area for future study.

Clinical considerations

Protective ventilation

It is now widely accepted that protective ventilation, a combination of low tidal volume, use of PEEP and recruitment manoeuvre is significantly associated with incidence of perioperative ALI. The PROtective Ventilation group (PROVE) has organised a number of larger scale randomised control trials (RCT). The IMPROVE trial, published in 2013, reported that the incidence of perioperative ALI in abdominal surgery is 0.5% when patients receive protective ventilation compared with 3% of those who received conventional ventilation^[116]. Similar results were also found in a meta-analysis which looked into thoracic and neurosurgery^[117]. However, protective ventilation does not seem to modify the mortality rate^[118-120].

Studies have also looked into the benefits of individual components of protective ventilation. The PROVHILO trial looked into the benefit of PEEP and recruitment manoeuvre with fixed tidal volume. It was found that high PEEP and alveolar recruitment itself did not lead to improvement in oxygenation. On the contrary, PEEP was associated with higher incidence of intra-operative hypotension^[121]. This also echoes the findings in non-surgical patients^[122]. When low tidal volume ventilation on its own, the results were also somewhat conflicting. While two meta-analyses of 4700 cases reported significantly lower rate of ALI with low tidal volume alone, they did not standardise the protocol for PEEP and recruitment manoeuvre. When controlled for PEEP and recruitment manoeuvre, low tidal volume did not reduce the rate of ALI^[6,122-123]. However, studies report that use of protective ventilation does not seem to modify the overall mortality rate^[124-125].

One lung ventilation is a special ventilation technique used in thoracic surgery, which is associated with significantly higher rate of perioperative ALI. This could also be reduced by the use of protective ventilation, low tidal volume of 6-8 mL/kg with PEEP and recruitment manoeuvre was associated with significantly lower rate of ALI^[126-127]. Other parameters of ventilator setting may also affect the incidence of ALI. A study by Hu *et al.* reported that pressure controlled volume guaranteed ventilation that resulted in better maintenance of lung compliance and blunts increase in inflammatory markers^[128].

Anaesthetic agents

In recent years, a number of published studies looked into the effect of anaesthetic agents on the progression of ALI, with some interesting results.

Sevoflurane, isoflurane and propofol have all been demonstrated to exhibit anti-inflammatory and cytoprotective effects in animal models of ALI, and the benefit is shown consistently in experiments involving LPS exposure, transplant and remote organ injury models, and ventilator induced lung injury models^[37,79,81].

Sevoflurane is used in the majority of anaesthetic agent studies. While there are very little available data on the effect of sevoflurane administration on the mortality rate or recovery time in ALI models, an *in-vivo* animal model consistently demonstrated that administration of sevoflurane in ALI model is associated with reduced histological change, reduced wet: dry ratio and improved ventilation parameters (higher pO₂ and lower pCO₂)^[51,129-130]. Sevoflurane administration is also associated with significantly lower neutrophil infiltration in the pulmonary tissue of the ALI models^[131]. In addition to the histological and blood gas parameter changes, sevoflurane administration is also associated with lower levels of proinflammatory cytokines, most notably Il-1a, Il-6, TNF- α , and lower level of chemokines^[80,132]. Sevoflurane administration is also associated with significantly lower NF-kb expression^[130].

Sevoflurane administration has also been shown to reduce the activity of cyclo-oxygenase, lipo-oxygenase and cytosolic phospholipase A₂ activities, thereby reducing the production of leukotriene and thromboxane levels^[133-134]. Sevoflurane administration is also associated with reduced expression of TLR4^[135].

Similar findings are also seen with isoflurane, with promising *in-vivo* survival data. In an experimental model of AKI using zymosan, a fungal surface glucan, isoflurane administration has been demonstrated to increase the survival rate 3-5 folds^[73,136]. This is associated with less histological damage, less protein

exudate and pulmonary oedema, and reduction in proinflammatory cytokines similar to that of sevoflurane. The studies also looked into pathways related to cell survival, and found that isoflurane administration is associated with significantly reduced caspase activities, downregulation of NF-kb through reduced expression and upregulated i-kb expression, and affects a number of apoptosis related mediators including BAX and Bcl-2. These are likely to account for the cytoprotective effect of isoflurane^[73,81,136-137].

Both isoflurane and sevoflurane are thought to play a role in maintaining the integrity of tight junction between airway epithelial cells. Breakdown of the tight junction with increased permeability is noted in both ventilation-induced lung injury and LPS models of ALI. Both volatile agents are noted to upregulate cell junction proteins zona occludens 1 and occludin expression, with normalisation of epithelial permeability^[81-82].

There are several explanations to the anti-inflammatory and cytoprotective effects of volatile agent. It was found that in human and rat cell lines, trifluorinated carbon molecule significantly reduces the expression of inflammatory cytokines Il-1, Il-6, and Il-8 and chemokines MIP-1 and CINC-1, which is associated with reduced neutrophil chemotaxis. In addition, trifluorinated carbon also seems to downregulate the caspase activity^[74]. Fortis *et al.* found that GABA administration also significantly reduces the expression of cytokines and chemokines, and this is negated by the co-administration of picrotoxin, a GABA receptor antagonist^[37]. Further study into the protective mechanism of inhalational agents is needed as this could lead to the development of better ALI treatments.

Propofol has demonstrated protective effect in ALI models. Zhao *et al.* demonstrated that propofol administration is associated with 2-fold increase in survival in a LPS model of ALI. Findings of reduced histological damage, pulmonary oedema and reduced pro-inflammatory cytokine profile were noted, similar to that of sevoflurane and isoflurane administration. Propofol has also been noted to have anti-oxidative properties, administration is associated with increased SOD and Nrf-2 activities, which reduces tissue hydrogen peroxide and MDA^[50,138].

Comparative studies of sevoflurane and propofol found that sevoflurane administration was associated with less neutrophil infiltration and lower cytokine expression^[131,139]. In terms of human study, there are four studies with a total of 130 participants, comparing the outcome of propofol and sevoflurane in perioperative ALI; overall, there were no significant differences in the incidence of ARDS and the reported biochemical

difference is conflicting between the studies^[25,140-142].

Xenon is a novel general anaesthetic agent, which has previously demonstrated neuroprotective effects. It also has anti-inflammatory and anti-apoptotic properties in ALI secondary to remote renal injury^[68].

In summary, there is now a growing body of evidence that general anaesthetic agents have significant protective effect against ALI, and have a role to play in the prevention and treatment of ALI. Given that most operations possess a high risk of developing perioperative ALI are done under general anaesthesia, the only clinical relevance would be the choice of anaesthetics, however, there is currently no conclusive human study to prove either is superior, although animal studies points towards inhalational agents.

Fluid administration and transfusion

In most operations, intravenous fluid is routinely administered, with blood component transfusion sometimes in case of high blood loss. However, a number of observation studies have reported that during the perioperative period, high volume of fluid administration is associated with significantly higher incidence of ALI^[143-145]. It has been demonstrated in RCTs of perioperative ALI cases that conservative fluid management is associated with better oxygenation and shorter intubation time^[146-147]. In a large RCT of medical and surgical patients with ALI, it was found that conservative fluid administration is associated with significantly better oxygenation, lung compliance, and 60 day survival^[148].

Nevertheless, it gets more complex in trauma. In animal studies, haemorrhage is associated with significantly decreased oxygenation, increased pulmonary vascular permeability and cell infiltration^[104,149]. However, in trauma patients, fluid administration is still linked with incidence of ALI. Two observational studies with a total of more than 2300 patients identified that the rate of ALI/ARDS is significantly higher in patients administered with larger volume of IV fluid^[20,150]. However, it is not known if this is an association or a causation, as larger fluid administration could be associated with severity of the trauma.

Like fluid administration, blood product transfusion has also been associated with the development of ALI^[20,151]. It is noted that the incidence of ALI associated with perioperative transfusion is significantly higher than the incidence in the general population^[152], and that the incidence of perioperative transfusion associated acute lung injury (TRALI) is significantly higher with larger volume of transfusion^[4]. A number of specific blood components are suggested as the cause of transfusion-related ALI, including erythrocyte derived

micro-particles and serum antibodies and platelet released VEGF^[43,153-154]. However, the exact mechanism is likely to be complex and multi-factorial.

Conclusion and way forward

In summary, perioperative ALI is a complex pathology which involves the activation of inflammatory pathways, increased endovascular permeability, increased oxidative stress and change in stress hormones. It occurs as a result of the interaction between surgical and anaesthetic factors, and patient's pre-operative condition. This is best described by the multi-causal model such as the one proposed by Middleburg *et al.*^[155].

In certain patient cohorts, perioperative ALI could be a frequent and devastating complication, associated with long periods of invasive monitoring and treatment, longer stay in hospital, more long term complications and increased mortality. This is also associated with significant cost to the health system.

In addition to protective ventilation, use of inhalational anaesthetic agent and conservative fluid administration, a number of potential prophylaxis and treatment for perioperative ALI have been identified and investigated in recent years. In animal studies, suppression of various parts of inflammatory cascade consistently reduced the severity of lung injury. One method to suppress inflammation in humans is with corticosteroids. Indeed, administration of corticosteroid is associated with significantly milder histological and mechanical lung changes, and significantly lower cytokine level compared to the control group^[156-157]. It is, however, worth noting that the role of corticosteroids in paediatric patients with is less clear, with conflicting data regarding its benefits^[158].

Neutrophil elastase is a proteinase secreted by neutrophils and macrophages during inflammation, and knockout studies suggest that it plays a role in leucocyte recruitment, inflammatory mediator release and phagocytosis^[159]. In patients with ARDS secondary to sepsis, selective neutrophil elastase inhibitor sivelestat has been reported to improve oxygenation, reduce lung injury and shorten the length of ICT stay^[160-161]. In the context of postoperative ALI without sepsis, while it improves inflammatory mediator levels, the benefit on prognosis is unclear^[162-164].

Other treatments which have shown potential benefit in human studies include therapeutic ventilation hypercapnia and terlipressin administration which have been shown to improve oxygenation and reduce lung injury^[96,165].

A wide range of interventions have shown benefit in animal studies, some using substances which are already licenced for use in humans. Hypertonic saline has shown benefits in animal studies, studies reported significantly lower lung injury and oedema associated with hypertonic saline administration, which is also associated with lower rate of mortality^[166-167]. TNF- α inhibition has shown promising results in animal model of ALI, which may warrant further human studies with existing anti-TNF therapy^[168-169].

As discussed above, manipulation of the renin-angiotensin-aldosterone system has shown promise as treatments for ALI. ACE inhibition, ARB blockade and upregulation of ACE II have all been shown to reduce the severity of ALI in animal models^[42,77,98]. Mineralocorticoid antagonist spironolactone administration has also been associated with reduced ALI in an ischaemic-reperfusion model^[170]. Similarly, α -2 adrenoceptor agonist dexmedetomidine and vasopressin have also shown therapeutic benefits in animal models^[92,94,96] whereas studies suggests that use of adrenaline in ALI should be avoided^[87].

Despite the limited conservative management options available for perioperative ALI, more potential treatment modalities are emerging which have shown promising results. However, larger scale human studies are needed to validate those findings, and potentially contribute towards a much lower mortality and morbidity rate associated with perioperative ALI.

References

- [1] Bernard GR, Artigas A, Brigham KL, et al. The American-European Consensus Conference on ARDS. Definitions, mechanisms, relevant outcomes, and clinical trial coordination[J]. *Am J Respir Crit Care Med*, 1994, 149(3 Pt 1): 818–824.
- [2] Blum JM, Stentz MJ, Dechert R, et al. Preoperative and intraoperative predictors of postoperative acute respiratory distress syndrome in a general surgical population[J]. *Anesthesiology*, 2013, 118(1): 19–29.
- [3] Licker M, Schweizer A, Ellenberger C, et al. Perioperative medical management of patients with COPD[J]. *Int J Chron Obstruct Pulmon Dis*, 2007, 2(4): 493–515.
- [4] Clifford L, Jia Q, Subramanian A, et al. Characterizing the epidemiology of postoperative transfusion-related acute lung injury[J]. *Anesthesiology*, 2015, 122(1): 12–20.
- [5] Hemmes SN, Serpa Neto A, Schultz MJ. Intraoperative ventilatory strategies to prevent postoperative pulmonary complications: a meta-analysis[J]. *Curr Opin Anaesthesiol*, 2013, 26(2): 126–133.
- [6] Gu WJ, Wang F, Liu JC. Effect of lung-protective ventilation with lower tidal volumes on clinical outcomes among patients undergoing surgery: a meta-analysis of randomized controlled trials[J]. *CMAJ*, 2015, 187(3): E101–E109.
- [7] Serpa Neto A, Hemmes SN, Barbas CS, et al., and the PROVE Network investigators. Incidence of mortality and morbidity related to postoperative lung injury in patients who have undergone abdominal or thoracic surgery: a systematic review and meta-analysis[J]. *Lancet Respir Med*, 2014, 2(12): 1007–1015.
- [8] Winkler EA, Yue JK, Birk H, et al. Perioperative morbidity and mortality after lumbar trauma in the elderly[J]. *Neurosurg Focus*, 2015, 39(4): E2.
- [9] McAlister FA, Bertsch K, Man J, et al. Incidence of and risk factors for pulmonary complications after nonthoracic surgery[J]. *Am J Respir Crit Care Med*, 2005, 171(5): 514–517.
- [10] Shi S, Chen C, Zhao D, et al. The role of plasma gelsolin in cardiopulmonary bypass induced acute lung injury in infants and young children: a pilot study[J]. *BMC Anesthesiol*, 2014, 14: 67.
- [11] Kor DJ, Lingineni RK, Gajic O, et al. Predicting risk of postoperative lung injury in high-risk surgical patients: a multicenter cohort study[J]. *Anesthesiology*, 2014, 120(5): 1168–1181.
- [12] Dancer RCA, Parekh D, Calfee CS, et al. American Thoracic Society International Conference: Smoking and the risk of acute lung injury in patients undergoing oesophagectomy[C]. *American Thoracic Society*; 2014, A1153-A.
- [13] Wu WC, Wang Y, Wang X, et al. Clinical analysis of acute lung injury after esophagectomy[J]. *J Cancer Res Ther*, 2014, 10(Suppl): 314–318.
- [14] Perkins GD, Gates S, Park D, et al., and the BALTI-Prevention Collaborators. The beta agonist lung injury trial prevention. A randomized controlled trial[J]. *Am J Respir Crit Care Med*, 2014, 189(6): 674–683.
- [15] Gajic O, Dabbagh O, Park PK, et al., and the U.S. Critical Illness and Injury Trials Group: Lung Injury Prevention Study Investigators (USCIITG-LIPS). Early identification of patients at risk of acute lung injury: evaluation of lung injury prediction score in a multicenter cohort study[J]. *Am J Respir Crit Care Med*, 2011, 183(4): 462–470.
- [16] Canet J, Gallart L, Gomar C, et al., and the ARISCAT Group. Prediction of postoperative pulmonary complications in a population-based surgical cohort[J]. *Anesthesiology*, 2010, 113(6): 1338–1350.
- [17] Feltracco P, Carollo C, Barbieri S, et al. Early respiratory complications after liver transplantation[J]. *World J Gastroenterol*, 2013, 19(48): 9271–9281.
- [18] Arozullah AM, Daley J, Henderson WG, et al., and the The National Veterans Administration Surgical Quality Improvement Program. Multifactorial risk index for predicting postoperative respiratory failure in men after major noncardiac surgery[J]. *Ann Surg*, 2000, 232(2): 242–253.

- [19] Tandon S, Batchelor A, Bullock R, et al. Peri-operative risk factors for acute lung injury after elective oesophagectomy[J]. *Br J Anaesth*, 2001, 86(5): 633–638.
- [20] Zielinski MD, Jenkins D, Cotton BA, et al., and the AAST Open Abdomen Study Group. Adult respiratory distress syndrome risk factors for injured patients undergoing damage-control laparotomy: AAST multicenter post hoc analysis[J]. *J Trauma Acute Care Surg*, 2014, 77(6): 886–891.
- [21] Ruhl AP, Lord RK, Panek JA, et al. Health care resource use and costs of two-year survivors of acute lung injury. An observational cohort study[J]. *Ann Am Thorac Soc*, 2015, 12(3): 392–401.
- [22] Xiang L, Lu S, Mittwede PN, et al. b(2)-Adrenoreceptor blockade improves early posttrauma hyperglycemia and pulmonary injury in obese rats[J]. *Am J Physiol Heart Circ Physiol*, 2014, 307(4): H621–H627.
- [23] Chi X, Guo N, Yao W, et al. Induction of heme oxygenase-1 by hemin protects lung against orthotopic autologous liver transplantation-induced acute lung injury in rats[J]. *J Transl Med*, 2016, 14(1): 35.
- [24] Fan Z, Yao J, Li Y, et al. Anti-inflammatory and antioxidant effects of curcumin on acute lung injury in a rodent model of intestinal ischemia reperfusion by inhibiting the pathway of NF-Kb[J]. *Int J Clin Exp Pathol*, 2015, 8(4): 3451–3459.
- [25] Potočnik I, Novak Janković V, Šostarič M, et al. Anti-inflammatory effect of sevoflurane in open lung surgery with one-lung ventilation[J]. *Croat Med J*, 2014, 55(6): 628–637.
- [26] Altemeier WA, Liles WC, Villagra-Garcia A, et al. Ischemia-reperfusion lung injury is attenuated in MyD88-deficient mice [J]. *PLoS One*, 2013, 8(10): e77123.
- [27] Chun CD, Liles WC, Frevert CW, et al. Mechanical ventilation modulates Toll-like receptor-3-induced lung inflammation via a MyD88-dependent, TLR4-independent pathway: a controlled animal study[J]. *BMC Pulm Med*, 2010, 10: 57.
- [28] Chi X, Yao W, Zhang A, et al. Downregulation of Lung Toll-Like Receptor 4 Could Effectively Attenuate Liver Transplantation-Induced Pulmonary Damage at the Early Stage of Reperfusion[J]. *Mediators Inflamm*, 2015, 2015: 383907.
- [29] You Z, Feng D, Xu H, et al. Nuclear factor-kappa B mediates one-lung ventilation-induced acute lung injury in rabbits[J]. *J Invest Surg*, 2012, 25(2): 78–85.
- [30] Guzel A, Kanter M, Guzel A, et al. Anti-inflammatory and antioxidant effects of infliximab on acute lung injury in a rat model of intestinal ischemia/reperfusion[J]. *J Mol Histol*, 2012, 43(3): 361–369.
- [31] Hempel SL, Monick MM, Hunninghake GW. Effect of hypoxia on release of IL-1 and TNF by human alveolar macrophages[J]. *Am J Respir Cell Mol Biol*, 1996, 14(2): 170–176.
- [32] Lawrence T. The nuclear factor NF-kappaB pathway in inflammation[J]. *Cold Spring Harb Perspect Biol*, 2009, 1(6): a001651.
- [33] Miyamoto S, Maki M, Schmitt MJ, et al. Tumor necrosis factor alpha-induced phosphorylation of I kappa B alpha is a signal for its degradation but not dissociation from NF-kappa B[J]. *Proc Natl Acad Sci U S A*, 1994, 91(26): 12740–12744.
- [34] Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family[J]. *Annu Rev Immunol*, 2009, 27: 519–550.
- [35] Scheller J, Chalaris A, Schmidt-Arras D, et al. The pro- and anti-inflammatory properties of the cytokine interleukin-6[J]. *Biochim Biophys Acta*, 2011, 1813(5): 878–888.
- [36] Pierson W, Liston A. A new role for interleukin-10 in immune regulation[J]. *Immunol Cell Biol*, 2010, 88(8): 769–770.
- [37] Fortis S, Spieth PM, Lu WY, et al. Effects of anesthetic regimes on inflammatory responses in a rat model of acute lung injury[J]. *Intensive Care Med*, 2012, 38(9): 1548–1555.
- [38] Tsan MF, Gao B. Endogenous ligands of Toll-like receptors [J]. *J Leukoc Biol*, 2004, 76(3): 514–519.
- [39] Ray M, Yu S, Sharda DR, et al. Inhibition of TLR4-induced IκB kinase activity by the RON receptor tyrosine kinase and its ligand, macrophage-stimulating protein[J]. *J Immunol*, 2010, 185(12): 7309–7316.
- [40] Maloney JP, Gao L. Proinflammatory Cytokines Increase Vascular Endothelial Growth Factor Expression in Alveolar Epithelial Cells[J]. *Mediators Inflamm*. 2015;2015:387842.
- [41] Nagineni CN, Kommineni VK, William A, et al. Regulation of VEGF expression in human retinal cells by cytokines: implications for the role of inflammation in age-related macular degeneration[J]. *J Cell Physiol*, 2012, 227(1): 116–126.
- [42] Yu X, Lin Q, Qin X, et al. ACE2 Antagonizes VEGFa to Reduce Vascular Permeability During Acute Lung Injury[J]. *Cell Physiol Biochem*, 2016, 38(3): 1055–1062.
- [43] Maloney JP, Ambruso DR, Voelkel NF, et al. Platelet Vascular Endothelial Growth Factor is a Potential Mediator of Transfusion-Related Acute Lung Injury[J]. *J Pulm Respir Med*, 2014, 4: 4.
- [44] Sato T, Paquet-Fifield S, Harris NC, et al. VEGF-D promotes pulmonary oedema in hyperoxic acute lung injury[J]. *J Pathol*, 2016, 239(2): 152–161.
- [45] Yang Y, Chen QH, Liu AR, et al. Synergism of MSC-secreted HGF and VEGF in stabilising endothelial barrier function upon lipopolysaccharide stimulation via the Rac1 pathway[J]. *Stem Cell Res Ther*, 2015, 6: 250.
- [46] Song J, Lu H, Zheng X, et al. Effects of Vascular Endothelial Growth Factor in Recovery Phase of Acute Lung Injury in Mice[J]. *Lung*, 2015, 193(6): 1029–1036.
- [47] Liu CH, Zhang WD, Wang JJ, et al. Senegenin Ameliorate Acute Lung Injury Through Reduction of Oxidative Stress and Inhibition of Inflammation in Cecal Ligation and Puncture-Induced Sepsis Rats[J]. *Inflammation*, 2016, 39(2): 900–906.
- [48] Jiang A, Liu C, Liu F, et al. Liver cold preservation induce lung surfactant changes and acute lung injury in rat liver transplantation[J]. *World J Gastroenterol*, 2012, 18(4): 323–

- 330.
- [49] Maroto R, Perez-Polo JR. BCL-2-related protein expression in apoptosis: oxidative stress versus serum deprivation in PC12 cells[J]. *J Neurochem*, 1997, 69(2): 514–523.
- [50] Yao W, Luo G, Zhu G, et al. Propofol activation of the Nrf2 pathway is associated with amelioration of acute lung injury in a rat liver transplantation model[J]. *Oxid Med Cell Longev*, 2014, 2014: 258567.
- [51] Luo C, Yuan D, Zhao W, et al. Sevoflurane ameliorates intestinal ischemia-reperfusion-induced lung injury by inhibiting the synergistic action between mast cell activation and oxidative stress[J]. *Mol Med Rep*, 2015, 12(1): 1082–1090.
- [52] Ning J, Mo L, Yi B, et al. Therapeutic Whole-body Hypothermia Protects Remote Lung, Liver, and Kidney Injuries after Blast Limb Trauma in Rats[J]. *Anesthesiology*, 2016, 124(6): 1360–1371.
- [53] Ning J, Mo L, Zhao H, et al. Transient regional hypothermia applied to a traumatic limb attenuates distant lung injury following blast limb trauma[J]. *Crit Care Med*, 2014, 42(1): e68–e78.
- [54] Bhat NR, Zhang P, Bhat AN. Cytokine induction of inducible nitric oxide synthase in an oligodendrocyte cell line: role of p38 mitogen-activated protein kinase activation[J]. *J Neurochem*, 1999, 72(2): 472–478.
- [55] Sedoris KC, Ovechkin AV, Gozal E, et al. Differential effects of nitric oxide synthesis on pulmonary vascular function during lung ischemia-reperfusion injury[J]. *Arch Physiol Biochem*, 2009, 115(1): 34–46.
- [56] Bloodsworth A, O'Donnell VB, Freeman BA. Nitric oxide regulation of free radical- and enzyme-mediated lipid and lipoprotein oxidation[J]. *Arterioscler Thromb Vasc Biol*, 2000, 20(7): 1707–1715.
- [57] Li WC, Zou ZJ, Zhou MG, et al. Effects of simvastatin on the expression of inducible NOS in acute lung injury in septic rats [J]. *Int J Clin Exp Pathol*, 2015, 8(11): 15106–15111.
- [58] Grailer JJ, Haggadone MD, Sarma JV, et al. Induction of M2 regulatory macrophages through the β 2-adrenergic receptor with protection during endotoxemia and acute lung injury[J]. *J Innate Immun*, 2014, 6(5): 607–618.
- [59] Gross CM, Rafikov R, Kumar S, et al. Endothelial nitric oxide synthase deficient mice are protected from lipopolysaccharide induced acute lung injury[J]. *PLoS One*, 2015, 10(3): e0119918.
- [60] Gao S, Guan S, Li H, et al. Ameliorating effects of low tidal volume ventilation with associated hypercapnia on pneumoperitoneum-induced lung injury by inhibition of Toll-like receptor 4[J]. *Int J Clin Exp Med*, 2015, 8(2): 1814–1823.
- [61] Ma L, Chen X, Wang R, et al. 3,5,4'-Tri-O-acetylresveratrol decreases seawater inhalation-induced acute lung injury by interfering with the NF- κ B and i-NOS pathways[J]. *Int J Mol Med*, 2016, 37(1): 165–172.
- [62] Zhu Q, He G, Wang J, et al. Protective effects of fenofibrate against acute lung injury induced by intestinal ischemia/reperfusion in mice[J]. *Sci Rep*, 2016, 6: 22044.
- [63] Breithaupt-Faloppa AC, Fantozzi ET, de Assis Ramos MM, et al. Protective effect of estradiol on acute lung inflammation induced by an intestinal ischemic insult is dependent on nitric oxide[J]. *Shock*, 2013, 40(3): 203–209.
- [64] Wu YH, Zhang X, Wang DH. Role of asymmetric dimethylarginine in acute lung injury induced by cerebral ischemia/reperfusion injury in rats[J]. *Nan Fang Yi Ke Da Xue Xue Bao*, 2011, 31(8): 1289–1294.
- [65] Sharma S, Smith A, Kumar S, et al. Mechanisms of nitric oxide synthase uncoupling in endotoxin-induced acute lung injury: role of asymmetric dimethylarginine[J]. *Vascul Pharmacol*, 2010, 52(5-6): 182–190.
- [66] Teerlink T, Luo Z, Palm F, et al. Cellular ADMA: regulation and action[J]. *Pharmacol Res*, 2009, 60(6): 448–460.
- [67] Aggarwal S, Gross CM, Kumar S, et al. Dimethylarginine dimethylaminohydrolase II overexpression attenuates LPS-mediated lung leak in acute lung injury[J]. *Am J Respir Cell Mol Biol*, 2014, 50(3): 614–625.
- [68] Zhao H, Huang H, Ologunde R, et al. Xenon Treatment Protects against Remote Lung Injury after Kidney Transplantation in Rats[J]. *Anesthesiology*, 2015, 122(6): 1312–1326.
- [69] Liu Z, Zhang B, Wang XB, et al. Hypertonicity contributes to seawater aspiration-induced lung injury: Role of hypoxia-inducible factor 1 α [J]. *Exp Lung Res*, 2015, 41(6): 301–315.
- [70] Weber DJ, Allette YM, Wilkes DS, et al. The HMGB1-RAGE Inflammatory Pathway: Implications for Brain Injury-Induced Pulmonary Dysfunction[J]. *Antioxid Redox Signal*, 2015, 23(17): 1316–1328.
- [71] Westphal D, Kluck RM, Dewson G. Building blocks of the apoptotic pore: how Bax and Bak are activated and oligomerize during apoptosis[J]. *Cell Death Differ*, 2014, 21(2): 196–205.
- [72] Ji Y, Gao F, Sun B, et al. Angiotensin-Converting Enzyme 2 Inhibits Apoptosis of Pulmonary Endothelial Cells During Acute Lung Injury Through Suppressing SMAD2 Phosphorylation[J]. *Cell Physiol Biochem*, 2015, 35(6): 2203–2212.
- [73] Li JT, Wang H, Li W, Wang LF, Hou LC, Mu JL, et al. Anesthetic isoflurane posttreatment attenuates experimental lung injury by inhibiting inflammation and apoptosis[J]. *Mediators Inflamm*. 2013, 2013: 108928.
- [74] Urner M, Limbach LK, Herrmann IK, et al. Fluorinated groups mediate the immunomodulatory effects of volatile anesthetics in acute cell injury[J]. *Am J Respir Cell Mol Biol*, 2011, 45(3): 617–624.
- [75] Wada T, Penninger JM. Mitogen-activated protein kinases in apoptosis regulation[J]. *Oncogene*, 2004, 23(16): 2838–2849.
- [76] Peroval MY, Boyd AC, Young JR, et al. A critical role for MAPK signalling pathways in the transcriptional regulation of toll like receptors[J]. *PLoS One*, 2013, 8(2): e51243.
- [77] Li Y, Cao Y, Zeng Z, et al. Angiotensin-converting enzyme 2/angiotensin-(1-7)/Mas axis prevents lipopolysaccharide-induced apoptosis of pulmonary microvascular endothelial

- cells by inhibiting JNK/NF- κ B pathways[J]. *Sci Rep*, 2015, 5: 8209.
- [78] Li Y, Zeng Z, Li Y, et al. Angiotensin-converting enzyme inhibition attenuates lipopolysaccharide-induced lung injury by regulating the balance between angiotensin-converting enzyme and angiotensin-converting enzyme 2 and inhibiting mitogen-activated protein kinase activation[J]. *Shock*, 2015, 43(4): 395–404.
- [79] Kim SH, Li M, Pyeon TH, et al. The volatile anesthetic sevoflurane attenuates ventilator-induced lung injury through inhibition of ERK1/2 and Akt signal transduction[J]. *Korean J Anesthesiol*, 2015, 68(1): 62–69.
- [80] Steurer M, Schl pfer M, Steurer M, et al. The volatile anaesthetic sevoflurane attenuates lipopolysaccharide-induced injury in alveolar macrophages[J]. *Clin Exp Immunol*, 2009, 155(2): 224–230.
- [81] Englert JA, Macias AA, Amador-Munoz D, et al. Isoflurane Ameliorates Acute Lung Injury by Preserving Epithelial Tight Junction Integrity[J]. *Anesthesiology*, 2015, 123(2): 377–388.
- [82] Chai J, Long B, Liu X, et al. Effects of sevoflurane on tight junction protein expression and PKC- α translocation after pulmonary ischemia-reperfusion injury[J]. *Exp Mol Med*, 2015, 47: e167.
- [83] Dong WW, Liu YJ, Lv Z, Mao YF, Wang YW, Zhu XY, et al. Lung endothelial barrier protection by resveratrol involves inhibition of HMGB1 release and HMGB1-induced mitochondrial oxidative damage via an Nrf2-dependent mechanism[J]. *Free Radic Biol Med*. 2015;88(Pt B):404–16.
- [84] Meliton A, Meng F, Tian Y, et al. Role of Krev Interaction Trapped-1 in Prostacyclin-Induced Protection against Lung Vascular Permeability Induced by Excessive Mechanical Forces and Thrombin Receptor Activating Peptide 6[J]. *Am J Respir Cell Mol Biol*, 2015, 53(6): 834–843.
- [85] Ballard-Croft C, Maass DL, Sikes P, et al. Activation of stress-responsive pathways by the sympathetic nervous system in burn trauma[J]. *Shock*, 2002, 18(1): 38–45.
- [86] Annane D, Trabold F, Sharshar T, et al. Inappropriate sympathetic activation at onset of septic shock: a spectral analysis approach[J]. *Am J Respir Crit Care Med*, 1999, 160(2): 458–465.
- [87] Ota S, Yazawa T, Tojo K, et al. Adrenaline aggravates lung injury caused by liver ischemia-reperfusion and high-tidal-volume ventilation in rats[J]. *J Intensive Care*, 2016, 4: 8.
- [88] Ji MH, Zhu XL, Liu FF, et al. Alpha 2A-adrenoreceptor blockade improves sepsis-induced acute lung injury accompanied with depressed high mobility group box-1 levels in rats [J]. *Cytokine*, 2012, 60(3): 639–645.
- [89] Kip G,  elik A, Bilge M, et al. Dexmedetomidine protects from post-myocardial ischaemia reperfusion lung damage in diabetic rats[J]. *Libyan J Med*, 2015, 10: 27828.
- [90] Geze S, Cekic B, Imamođlu M, et al. Use of dexmedetomidine to prevent pulmonary injury after pneumoperitoneum in ventilated rats[J]. *Surg Laparosc Endosc Percutan Tech*, 2012, 22(5): 447–453.
- [91] Wu X, Song X, Li N, et al. Protective effects of dexmedetomidine on blunt chest trauma-induced pulmonary contusion in rats[J]. *J Trauma Acute Care Surg*, 2013, 74(2): 524–530.
- [92] Gu J, Chen J, Xia P, et al. Dexmedetomidine attenuates remote lung injury induced by renal ischemia-reperfusion in mice[J]. *Acta Anaesthesiol Scand*, 2011, 55(10): 1272–1278.
- [93] Wang X, Ma S, Liu Y, et al. Effects and mechanism analysis of combined infusion by levosimendan and vasopressin on acute lung injury in rats septic shock[J]. *Cell Biochem Biophys*, 2014, 70(3): 1639–1645.
- [94] Westphal M, Rehberg S, Maybauer MO, et al. Cardiopulmonary effects of low-dose arginine vasopressin in ovine acute lung injury[J]. *Crit Care Med*, 2011, 39(2): 357–363.
- [95] Deng W, Wang D. Effect of arginine vasopressin on alveolar fluid clearance in rats with acute lung injury[J]. *Nan Fang Yi Ke Da Xue Xue Bao*, 2015, 35(11): 1602–1605.
- [96] Hua F, Wang X, Zhu L. Terlipressin decreases vascular endothelial growth factor expression and improves oxygenation in patients with acute respiratory distress syndrome and shock[J]. *J Emerg Med*, 2013, 44(2): 434–439.
- [97] Deng W, Deng Y, Deng J, et al. Losartan attenuated lipopolysaccharide-induced lung injury by suppression of lectin-like oxidized low-density lipoprotein receptor-1[J]. *Int J Clin Exp Pathol*, 2015, 8(12): 15670–15676.
- [98] Zambelli V, Bellani G, Borsa R, et al. Angiotensin-(1-7) improves oxygenation, while reducing cellular infiltrate and fibrosis in experimental Acute Respiratory Distress Syndrome [J]. *Intensive Care Med Exp*, 2015, 3(1): 44.
- [99] Bao H, Gao F, Xie G, et al. Angiotensin-Converting Enzyme 2 Inhibits Apoptosis of Pulmonary Endothelial Cells During Acute Lung Injury Through Suppressing MiR-4262[J]. *Cell Physiol Biochem*, 2015, 37(2): 759–767.
- [100] Rosenblatt-Velin N, Badoux S, Liaudet L. Pharmacological Therapy in the Heart as an Alternative to Cellular Therapy: A Place for the Brain Natriuretic Peptide[J]? *Stem Cells Int*. 2016;2016:5961342.
- [101] Sun YZ, Gao YL, Yu QX, et al. Assessment of acute lung injury/acute respiratory distress syndrome using B-type brain natriuretic peptide[J]. *J Int Med Res*, 2015, 43(6): 802–808.
- [102] Reel B, Oishi PE, Hsu JH, et al. Early elevations in B-type natriuretic peptide levels are associated with poor clinical outcomes in pediatric acute lung injury[J]. *Pediatr Pulmonol*, 2009, 44(11): 1118–1124.
- [103] Cao X, Xia HY, Zhang T, et al. Protective effect of lyophilized recombinant human brain natriuretic peptide on renal ischemia/reperfusion injury in mice[J]. *Genet Mol Res*, 2015, 14(4): 13300–13311.
- [104] Song Z, Zhao X, Liu M, et al. Recombinant human brain natriuretic peptide attenuates trauma/haemorrhagic shock-induced acute lung injury through inhibiting oxidative stress and the NF- κ B-dependent inflammatory/MMP-9 pathway[J].

- Int J Exp Pathol*, 2015, 96(6): 406–413.
- [105] Aisiku IP, Yamal JM, Doshi P, et al. The incidence of ARDS and associated mortality in severe TBI using the Berlin definition[J]. *J Trauma Acute Care Surg*, 2016, 80(2): 308–312.
- [106] Cordts Filho RdeM, Parreira JG, Perlingeiro JA, et al. Pelvic fractures as a marker of injury severity in trauma patients[J]. *Rev Col Bras Cir*, 2011, 38(5): 310–316.
- [107] Pirat A, Ozgur S, Torgay A, et al. Risk factors for postoperative respiratory complications in adult liver transplant recipients[J]. *Transplant Proc*, 2004, 36(1): 218–220.
- [108] Canet E, Osman D, Lambert J, et al. Acute respiratory failure in kidney transplant recipients: a multicenter study[J]. *Crit Care*, 2011, 15(2): R91.
- [109] Zhao W, Ge X, Sun K, et al. Acute respiratory distress syndrome after orthotopic liver transplantation[J]. *J Crit Care*, 2016, 31(1): 163–167.
- [110] Iida T, Takagi T, Katada K, et al. Rapamycin Improves Mortality Following Intestinal Ischemia-Reperfusion via the Inhibition of Remote Lung Inflammation in Mice[J]. *Digestion*, 2015, 92(4): 211–219.
- [111] Jiang A, Liu C, Song Y, et al. NF- κ B induced the donor liver cold preservation related acute lung injury in rat liver transplantation model[J]. *PLoS One*, 2011, 6(9): e24960.
- [112] Zhao H, Ning J, Lemaire A, et al. Necroptosis and parthanatos are involved in remote lung injury after receiving ischemic renal allografts in rats[J]. *Kidney Int*, 2015, 87(4): 738–748.
- [113] Hashimoto K, Kim H, Oishi H, et al. Annexin V homodimer protects against ischemia reperfusion-induced acute lung injury in lung transplantation[J]. *J Thorac Cardiovasc Surg*, 2016, 151(3): 861–868.
- [114] Aggarwal S, Dimitropoulou C, Lu Q, et al. Glutathione supplementation attenuates lipopolysaccharide-induced mitochondrial dysfunction and apoptosis in a mouse model of acute lung injury[J]. *Front Physiol*, 2012, 3: 161.
- [115] Hassett P, Curley GF, Contreras M, et al. Overexpression of pulmonary extracellular superoxide dismutase attenuates endotoxin-induced acute lung injury[J]. *Intensive Care Med*, 2011, 37(10): 1680–1687.
- [116] Futier E, Constantin JM, Paugam-Burtz C, et al., and the IMPROVE Study Group. A trial of intraoperative low-tidal-volume ventilation in abdominal surgery[J]. *N Engl J Med*, 2013, 369(5): 428–437.
- [117] Sutherasan Y, Vargas M, Pelosi P. Protective mechanical ventilation in the non-injured lung: review and meta-analysis [J]. *Crit Care*, 2014, 18(2): 211.
- [118] Severgnini P, Selmo G, Lanza C, et al. Protective mechanical ventilation during general anesthesia for open abdominal surgery improves postoperative pulmonary function[J]. *Anesthesiology*, 2013, 118(6): 1307–1321.
- [119] Park SJ, Kim BG, Oh AH, et al. Effects of intraoperative protective lung ventilation on postoperative pulmonary complications in patients with laparoscopic surgery: prospective, randomized and controlled trial[J]. *Surg Endosc*, 2016.
- [120] Ge Y, Yuan L, Jiang X, et al. Effect of lung protection mechanical ventilation on respiratory function in the elderly undergoing spinal fusion[J]. *Zhong Nan Da Xue Xue Bao Yi Xue Ban*, 2013, 38(1): 81–85.
- [121] Hemmes SN, Gama de Abreu M, Pelosi P, et al., and the PROVE Network Investigators for the Clinical Trial Network of the European Society of Anaesthesiology. High versus low positive end-expiratory pressure during general anaesthesia for open abdominal surgery (PROVHILO trial): a multicentre randomised controlled trial[J]. *Lancet*, 2014, 384(9942): 495–503.
- [122] Santa Cruz R, Rojas JJ, Nervi R, et al. High versus low positive end-expiratory pressure (PEEP) levels for mechanically ventilated adult patients with acute lung injury and acute respiratory distress syndrome[J]. *Cochrane Database Syst Rev*, 2013, 6(6): CD009098.
- [123] Yang D, Grant MC, Stone A, et al. A meta-analysis of intraoperative ventilation strategies to prevent pulmonary complications: is low tidal volume alone sufficient to protect healthy lungs[J]? *Ann Surg*, 2016, 263(5):881-887.
- [124] Guay J, Ochroch EA. Intraoperative use of low volume ventilation to decrease postoperative mortality, mechanical ventilation, lengths of stay and lung injury in patients without acute lung injury[J]. *Cochrane Database Syst Rev*, 2015, 12(12): CD011151.
- [125] Serpa Neto A, Cardoso SO, Manetta JA, et al. Association between use of lung-protective ventilation with lower tidal volumes and clinical outcomes among patients without acute respiratory distress syndrome: a meta-analysis[J]. *JAMA*, 2012, 308(16): 1651–1659.
- [126] Yang M, Ahn HJ, Kim K, et al. Does a protective ventilation strategy reduce the risk of pulmonary complications after lung cancer surgery?: a randomized controlled trial[J]. *Chest*, 2011, 139(3): 530–537.
- [127] Choi YS, Bae MK, Kim SH, et al. Effects of Alveolar Recruitment and Positive End-Expiratory Pressure on Oxygenation during One-Lung Ventilation in the Supine Position[J]. *Yonsei Med J*, 2015, 56(5): 1421–1427.
- [128] Hu X, Shen H, Li X, et al. [Effects of volume-controlled ventilation and pressure-controlled volume- guaranteed mode during one-lung ventilation on circulation, pulmonary function and lung injury][J]. *Zhonghua Yi Xue Za Zhi*, 2014, 94(13): 1006–1009.
- [129] Casanova J, Garutti I, Simon C, et al. The effects of anesthetic preconditioning with sevoflurane in an experimental lung autotransplant model in pigs[J]. *Anesth Analg*, 2011, 113(4): 742–748.
- [130] Otsuki T, Ishikawa M, Hori Y, et al. Volatile anesthetic sevoflurane ameliorates endotoxin-induced acute lung injury via microRNA modulation in rats[J]. *Biomed Rep*, 2015, 3(3): 408–412.
- [131] Ferrando C, Aguilar G, Piqueras L, et al. Sevoflurane, but not

- propofol, reduces the lung inflammatory response and improves oxygenation in an acute respiratory distress syndrome model: a randomised laboratory study[J]. *Eur J Anaesthesiol*, 2013, 30(8): 455–463.
- [132] Voigtsberger S, Lachmann RA, Leutert AC, et al. Sevoflurane ameliorates gas exchange and attenuates lung damage in experimental lipopolysaccharide-induced lung injury[J]. *Anesthesiology*, 2009, 111(6): 1238–1248.
- [133] Liu R, Yang Y, Li Y, et al. Effects of sevoflurane on pulmonary cytosolic phospholipase A₂ and clara cell secretory protein expressions in rabbits with one-lung ventilation-induced lung injury[J]. *Nan Fang Yi Ke Da Xue Xue Bao*, 2013, 33(4): 469–473.
- [134] Liu R, Luo J, Li J, et al. Protective mechanisms of sevoflurane against one-lung ventilation-induced acute lung injury: role of cyclooxygenase-2 and 5-lipoxygenase pathways[J]. *Nan Fang Yi Ke Da Xue Xue Bao*, 2013, 33(5): 625–630.
- [135] Sun XJ, Li XQ, Wang XL, et al. Sevoflurane inhibits nuclear factor- κ B activation in lipopolysaccharide-induced acute inflammatory lung injury via toll-like receptor 4 signaling[J]. *PLoS One*, 2015, 10(4): e0122752.
- [136] Wang H, Fan J, Li NL, et al. A subanesthetic dose of isoflurane during postconditioning ameliorates zymosan-induced neutrophil inflammation lung injury and mortality in mice[J]. *Mediators Inflamm*. 2013, 2013: 479628.
- [137] Faller S, Strosing KM, Ryter SW, et al. The volatile anesthetic isoflurane prevents ventilator-induced lung injury via phosphoinositide 3-kinase/Akt signaling in mice[J]. *Anesth Analg*, 2012, 114(4): 747–756.
- [138] Zhao LL, Hu GC, Zhu SS, et al. Propofol pretreatment attenuates lipopolysaccharide-induced acute lung injury in rats by activating the phosphoinositide-3-kinase/Akt pathway[J]. *Braz J Med Biol Res*, 2014, 47(12): 1062–1067.
- [139] Wakabayashi S, Yamaguchi K, Kumakura S, et al. Effects of anesthesia with sevoflurane and propofol on the cytokine/chemokine production at the airway epithelium during esophagectomy[J]. *Int J Mol Med*, 2014, 34(1): 137–144.
- [140] Erturk E, Topaloglu S, Dohman D, et al. The comparison of the effects of sevoflurane inhalation anesthesia and intravenous propofol anesthesia on oxidative stress in one lung ventilation[J]. *Biomed Res Int*. 2014, 2014: 360936.
- [141] Jin Y, Zhao X, Li H, et al. Effects of sevoflurane and propofol on the inflammatory response and pulmonary function of perioperative patients with one-lung ventilation[J]. *Exp Ther Med*, 2013, 6(3): 781–785.
- [142] Feng H, Wang GM, Qiao Y, et al. Effects of sevoflurane preconditioning on lung injury during one lung ventilation[J]. *Int J Clin Exp Med*, 2015, 8(8): 13634–13638.
- [143] Evans RG, Naidu B. Does a conservative fluid management strategy in the perioperative management of lung resection patients reduce the risk of acute lung injury[J]? *Interact Cardiovasc Thorac Surg*, 2012, 15(3): 498–504.
- [144] Chau EH, Slinger P. Perioperative fluid management for pulmonary resection surgery and esophagectomy[J]. *Semin Cardiothorac Vasc Anesth*, 2014, 18(1): 36–44.
- [145] Licker M, Diaper J, Villiger Y, et al. Impact of intraoperative lung-protective interventions in patients undergoing lung cancer surgery[J]. *Crit Care*, 2009, 13(2): R41.
- [146] Stewart RM, Park PK, Hunt JP, et al., and the National Institutes of Health/National Heart, Lung, and Blood Institute Acute Respiratory Distress Syndrome Clinical Trials Network. Less is more: improved outcomes in surgical patients with conservative fluid administration and central venous catheter monitoring[J]. *J Am Coll Surg*, 2009, 208(5): 725–735., discussion 735–737.
- [147] Zhang J, Chen CQ, Lei XZ, et al. Goal-directed fluid optimization based on stroke volume variation and cardiac index during one-lung ventilation in patients undergoing thoracoscopy lobectomy operations: a pilot study[J]. *Clinics (Sao Paulo)*, 2013, 68(7): 1065–1070.
- [148] Wiedemann HP, Wheeler AP, Bernard GR, et al., and the National Heart, Lung, and Blood Institute Acute Respiratory Distress Syndrome (ARDS) Clinical Trials Network. Comparison of two fluid-management strategies in acute lung injury [J]. *N Engl J Med*, 2006, 354(24): 2564–2575.
- [149] Rani M, Zhang Q, Oppeltz RF, et al. Gamma delta T cells regulate inflammatory cell infiltration of the lung after trauma-hemorrhage[J]. *Shock*, 2015, 43(6): 589–597.
- [150] Kasotakis G, Sideris A, Yang Y, et al., and the Inflammation and Host Response to Injury Investigators. Aggressive early crystalloid resuscitation adversely affects outcomes in adult blunt trauma patients: an analysis of the Glue Grant database [J]. *J Trauma Acute Care Surg*, 2013, 74(5): 1215–1221., discussion 1221–1222.
- [151] Gajic O, Dara SI, Mendez JL, et al. Ventilator-associated lung injury in patients without acute lung injury at the onset of mechanical ventilation[J]. *Crit Care Med*, 2004, 32(9): 1817–1824.
- [152] Morita Y, Pretto EA Jr. Increased incidence of transfusion-related acute lung injury during orthotopic liver transplantation: a short report[J]. *Transplant Proc*, 2014, 46(10): 3593–3597.
- [153] Kanai R, Iijima T, Hashimoto S, et al. Impact of immunoreactive substances contained in apheresis platelet concentrate on postoperative respiratory function in surgical patients receiving platelet transfusion: a prospective cohort study[J]. *Transfus Med*, 2013, 23(5): 344–350.
- [154] Jy W, Ricci M, Shariatmadar S, et al. Microparticles in stored red blood cells as potential mediators of transfusion complications[J]. *Transfusion*, 2011, 51(4): 886–893.
- [155] Middelburg RA, van Stein D, Briët E, et al. The role of donor antibodies in the pathogenesis of transfusion-related acute lung injury: a systematic review[J]. *Transfusion*, 2008, 48(10): 2167–2176.
- [156] Ju NY, Gao H, Huang W, et al. Therapeutic effect of inhaled budesonide (Pulmicort® Turbuhaler) on the inflammatory

- response to one-lung ventilation[J]. *Anaesthesia*, 2014, 69(1): 14–23.
- [157] Theroux MC, Fisher AO, Rodriguez ME, et al. Prophylactic methylprednisolone to reduce inflammation and improve outcomes from one lung ventilation in children: a randomized clinical trial[J]. *Paediatr Anaesth*, 2015, 25(6): 587–594.
- [158] Yehya N, Servaes S, Thomas NJ, et al. Corticosteroid exposure in pediatric acute respiratory distress syndrome[J]. *Intensive Care Med*, 2015, 41(9): 1658–1666.
- [159] Young RE, Thompson RD, Larbi KY, et al. Neutrophil elastase (NE)-deficient mice demonstrate a nonredundant role for NE in neutrophil migration, generation of proinflammatory mediators, and phagocytosis in response to zymosan particles in vivo[J]. *J Immunol*, 2004, 172(7): 4493–4502.
- [160] Hayakawa M, Katabami K, Wada T, et al. Sivelestat (selective neutrophil elastase inhibitor) improves the mortality rate of sepsis associated with both acute respiratory distress syndrome and disseminated intravascular coagulation patients[J]. *Shock*, 2010, 33(1): 14–18.
- [161] Tsuboko Y, Takeda S, Mii S, et al. Clinical evaluation of sivelestat for acute lung injury/acute respiratory distress syndrome following surgery for abdominal sepsis[J]. *Drug Des Devel Ther*, 2012, 6: 273–278.
- [162] Inoue N, Oka N, Kitamura T, et al. Neutrophil elastase inhibitor sivelestat attenuates perioperative inflammatory response in pediatric heart surgery with cardiopulmonary bypass[J]. *Int Heart J*, 2013, 54(3): 149–153.
- [163] Lee SK, Son BS, Hwang JJ, et al. The use of neutrophil elastase inhibitor in the treatment of acute lung injury after pneumonectomy[J]. *J Cardiothorac Surg*, 2013, 8: 69.
- [164] Eguchi T, Yoshida K, Kondo R, et al. Sivelestat prevents cytoskeletal rearrangements in neutrophils resulting from lung re-expansion following one-lung ventilation during thoracic surgery[J]. *Inflammation*, 2013, 36(6): 1479–1484.
- [165] Gao W, Liu DD, Li D, et al. Effect of Therapeutic Hypercapnia on Inflammatory Responses to One-lung Ventilation in Lobectomy Patients[J]. *Anesthesiology*, 2015, 122(6): 1235–1252.
- [166] Bihari S, Dixon DL, Lawrence MD, et al. Induced hypernatraemia is protective in acute lung injury[J]. *Respir Physiol Neurobiol*, 2016, 227: 56–67.
- [167] Petroni RC, Biselli PJ, de Lima TM, et al. Hypertonic Saline (NaCl 7.5%) Reduces LPS-Induced Acute Lung Injury in Rats [J]. *Inflammation*, 2015, 38(6): 2026–2035.
- [168] Weifeng Y, Li L, Yujie H, et al. Inhibition of Acute Lung Injury by TNFR-Fc through Regulation of an Inflammation-Oxidative Stress Pathway[J]. *PLoS One*, 2016, 11(3): e0151672.
- [169] Yu Y, Gao M, Li H, et al. Pulmonary artery perfusion with anti-tumor necrosis factor alpha antibody reduces cardiopulmonary bypass-induced inflammatory lung injury in a rabbit model[J]. *PLoS One*, 2013, 8(12): e83236.
- [170] Barut F, Ozacmak VH, Turan I, et al. Reduction of Acute Lung Injury by Administration of Spironolactone After Intestinal Ischemia and Reperfusion in Rats[J]. *Clin Invest Med*, 2016, 39(1): E15–E24.