

A Glimpse of Inflammation and Anti-Inflammation Therapy in Diabetic Kidney Disease

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Diabetic kidney disease (DKD) is a common complication of diabetes mellitus and a major cause of end-stage kidney disease (ESKD). The pathogenesis of DKD is very complex and not completely understood. Recently, accumulated evidence from in vitro and in vivo studies has demonstrated that inflammation plays an important role in the pathogenesis and the development of DKD. It has been well known that a variety of pro-inflammatory cytokines and related signaling pathways are involved in the procession of DKD. Additionally, some anti-hyperglycemic agents and mineralocorticoid receptor antagonists (MRAs) that are effective in alleviating the progression of DKD have antiinflammatory properties, which might have beneficial effects on delaying the progression of DKD. However, there is currently a lack of systematic overviews. In this review, we focus on the novel pro-inflammatory signaling pathways in the development of DKD, including the nuclear factor kappa B (NF-ĸB) signaling pathway, toll-like receptors (TLRs) and myeloid differentiation primary response 88 (TLRs/MyD88) signaling pathway, adenosine 5'monophosphate-activated protein kinase (AMPK) signaling pathways, inflammasome activation, mitochondrial DNA (mtDNA) release as well as hypoxia-inducible factor-1(HIF-1) signaling pathway. We also discuss the related anti-inflammation mechanisms of metformin, finerenone, sodium-dependent glucose transporters 2 (SGLT2) inhibitors, Dipeptidyl peptidase-4 (DPP-4) inhibitors, Glucagon-like peptide-1 (GLP-1) receptor agonist and traditional Chinese medicines (TCM).

Keywords: diabetic kidney disease, inflammation, signaling pathway, anti-Inflammation, anti-hyperglycemic, mineralocorticoid receptor antagonists, traditional Chinese medicine

INTRODUCTION

Diabetic kidney disease (DKD), which is also known as diabetic nephropathy before, is the most common microvascular complication of diabetes mellitus (Thomas et al., 2015; Doshi and Friedman, 2017; Sun et al., 2022). The number of DKD patients has increased year by year, which has led to a huge burden on global public health. Recent studies show that 21.8%–40% of diabetic patients may progress to DKD, which is the main cause of end-stage renal disease (ESRD) (Zhang J. et al., 2020; Mottl et al., 2022). In addition, nearly 38.8% of ESRD in 2018 was attributable to DKD in America (Johansen et al., 2021). In China and India, the proportion of ESRD in DKD patients was about 1/5

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Inflammation and Diabetic Kidney Disease

(23%) and 1/3 (31.2%), respectively (Hussain et al., 2019; Zhang L. et al., 2019). On the other hand, treatment of DKD mainly includes lifestyle interventions, such as exercise, weight loss, smoking cessation and sodium restriction, and medication to control hyperglycemia as well as blood pressure (Doshi and Friedman, 2017; Anders et al., 2018). However, current treatments cannot completely delay the progression to ESRD. Therefore, it is still necessary to further explore the pathogenesis of DKD and find new therapeutic targets.

Previous studies demonstrate that many factors are implicated in the progression of DKD, such as metabolic abnormalities and hemodynamic changes caused by hyperglycemia and insulin resistance, etc (Forbes and Thorburn, 2018). It has been wellknown that overproduction of reactive oxygen species (ROS), abnormal autophagy, senescence are also involved in the development of DKD (Ren et al., 2020; Sun et al., 2022). Recently, accumulated evidence suggested that inflammation plays a key role in the pathophysiology of DKD. For example, oxidative stress (Huang W. et al., 2020), abnormal autophagy (Han et al., 2021), and senescence (Xiong and Zhou, 2019) could lead to the abnormal immune-inflammation in DKD. Furthermore, several clinical trials found that non-steroidal selective mineralocorticoid receptor antagonists (MRA) could delay the progression of DKD by inhibiting inflammation (Donath, 2014; Komada and Muruve, 2019). These data strongly suggest that inflammation plays an important role in the development of DKD.

Inflammation is a biologically conserved immune defense mechanism (Cronkite and Strutt, 2018). In human, inflammation plays a dual role in various disease. Early inflammatory responses may promote tissue repair. However, severe inflammation would lead to secondary injury resulting in tissue damage or fibrosis (Nathan and Ding, 2010; Rathinam and Chan, 2018). In addition, increasing evidence indicates that in chronic non-infectious diseases, such as diabetes and its complications including DKD and atherosclerosis, persistent systemic or local inflammation plays a critical role in the progression of disease (Pichler et al., 2017; Tang and Yiu, 2020; Engelen et al., 2022; Rohm et al., 2022). A large number of infiltrated macrophages were observed in the renal biopsy specimens in patients with DKD, and the expression of inflammatory cytokines such as Interleukin 6 (IL-6), Interleukin 1 beta (IL-1β), and tumor necrosis factor alpha (TNF- α) were increased (Navarro-Gonzalez et al., 2011; Araujo et al., 2020). In recent years, the microarray analysis has also showed that the expression of pro-inflammatory genes and fibrosis-related genes were significantly increased in patients with DKD or animal models, which was accompanied with increased levels of inflammatory cytokines in peripheral blood specimens (Wilson et al., 2019; Levin et al., 2020; Sur et al., 2021). These data further indicate that there is a notable association between inflammation and the development of DKD.

In this review, we discuss the role of some novel and key proinflammatory signaling pathways in the progression of DKD, such as the nuclear factor kappa B (NF- κ B) signaling pathway, toll-like receptors (TLRs) and myeloid differentiation primary response 88 (TLRs/MyD88) signaling pathway, adenosine 5'- monophosphate-activated protein kinase (AMPK) signaling pathways, inflammasome activation, mitochondrial DNA (mtDNA) release and hypoxia-inducible factor-1(HIF) signaling pathway. We also introduce the role of antihyperglycemic drugs, non-steroidal selective MRA and traditional Chinese medicine in anti-inflammation during the treatment of DKD.

HYPERGLYCEMIA-INDUCED INFLAMMATION IN DKD

Hyperglycemia and insulin resistance are the most common pathological characteristics in patients with DKD and can lead to systemic low-grade inflammation (Saraheimo et al., 2003; Leehey, 2020). Hyperglycemia can lead to overproduction of ROS and mitochondrial DNA release, which further cause inflammation in renal parenchymal cells by different signaling pathways (Sun et al., 2022).

NF-\kappaB Signaling

The transcription factor NF-KB is a key mediator of inflammation and NF-kB remains inactive while binding to the proteins of the IkappaB (IκB) family (mainly IκBα) (Sun, 2011, 2017; Zhang Q. et al., 2017). There are two pathways for the activation of NF-κB: the canonical pathway and the non-canonical pathway (Zhang S. et al., 2017). Under the stimulation of various immune receptors, such as pattern recognition receptors (PRR), TNF receptors (TNFR), T cell receptors (TCR) and B-cell receptors, intracellular $I\kappa B\alpha$ is degraded and the canonical NF- κB pathway is activated (Sun, 2017). The non-canonical NF- κB activation is primarily associated with ligands of a subset of TNFR superfamily members, such as the lymphotoxin beta receptor (LTBR), B-cell activating factor receptor (BAFFR), Cluster of differentiation 40 (CD40), and Receptor activator of nuclear factor κ B (RANK) (Sun, 2017; Hou et al., 2018). NF-κB mediates the activation of downstream signaling pathways including inflammasome activation, HIF signaling pathways and AMPK signaling pathways (Rius et al., 2008; Afonina et al., 2017). In addition, the activation of NF-kB signaling pathway inhibits the anti-inflammatory pathway, Sirtuin 1 (SIRT1) signaling pathway, which is closely related to the development of non-infectious chronic diseases such as atherosclerosis and DKD (Stein et al., 2010; Kauppinen et al., 2013; Sun et al., 2021). Overall, the NF-kB signaling pathway plays a key role in the inflammatory response.

It has been observed that activated NF- κ B signaling pathway is associated with inflammation and fibrosis in DKD (Mezzano et al., 2004; Schmid et al., 2006; Liu P. et al., 2014). Foresto-Neto et al. found that the activation of NF- κ B signaling pathway increased the expression of the inflammatory cytokines IL-6, which preceded the kidney damage in the db/db mice (Foresto-Neto et al., 2020). Microarray analysis of renal biopsies in patients with DKD also suggested that a number of NF- κ B targets were significantly upregulated (Schmid et al., 2006). These data suggest the key role of abnormal activation of NF- κ B in the progression of DKD. A recent study found that delivery of Smad7 siRNA to the kidney through ultrasound microbubbles could reduce the production of IL-1 β and monocyte chemoattractant protein 1 (MCP1) in the renal tissue of db/db mice by inhibiting NF- κ B signaling pathway (Ka et al., 2012). Sun et al. demonstrated that dephosphorylated P65 NF- κ B could attenuate renal inflammation-related damage in STZ-induced diabetic mice by downregulating the expression levels of TNF- α , IL-1 β , and cyclooxygenase 2 (COX-2). (Zhang M. et al., 2019). These data further confirm that the NF- κ B signaling pathway plays a critical role in the development of DKD and targeting NF- κ B signaling pathway might be a potential therapeutic strategy for DKD.

TLRs-MyD88 Signaling Pathway

Toll-like receptors (TLRs) is an evolutionarily ancient family of pattern recognition receptors and an important component of innate immunity (Tang and Yiu, 2020). Activation of TLRs in response to pathogen-associated molecular patterns (PAMPs) or danger-associated molecular patterns (DAMPs) could lead to inflammation in injured tissue (Fitzgerald and Kagan, 2020; Tang and Yiu, 2020). Myeloid differentiation factor 88 (MyD88) is a critical adaptor protein in innate immunity and central hub in inflammatory response (Xu et al., 2000; Deguine and Barton, 2014), which could respond to the activation of TLRs. The activation of TLRs-MyD88 signaling pathway might initiate the translocation of NF-KB into the nucleus, thus promoting the transcription of pro-inflammatory factors and chemokines, such as IL-1β, IL-6, MCP-1 etc. (Warner and Nunez, 2013; Fitzgerald and Kagan, 2020). These studies suggest that TLRs-MyD88 signaling pathway plays a key role in the inflammatory response.

It has been demonstrated that the high glucose (HG) could activate TLRs-MyD88 signaling pathway in the kidneys of DKD, which is the initiator of renal interstitial fibrosis (Lin and Tang, 2014; Liu and Zen, 2021). Liu et al. found that the mRNA and protein levels of TLR4 and MyD88 increased in renal tubular epithelial cells treated with HG, which was accompanied with the activation of NF-KB and increased expression of chemokine MCP-1 (Liu R. et al., 2014). Zhang et al. found that pharmacological inhibition of MyD88 by LM8, a new smallmolecule inhibitor of MyD88, could alleviate the progression of renal fibrosis by inhibiting the activation of NF-kB and reducing the expression of TNF-α and IL-1β in both streptozotocin (STZ)induced diabetic mice and db/db mice (Zhang et al., 2022). These data indicate that targeting TLRs-MyD88 might be a potential therapeutic option to reduce renal fibrosis in DKD. In addition, there are a series of Chinese herbal medicines to reduce renal fibrosis by inhibiting TLRs-MyD88 signaling-mediated inflammation (Lu et al., 2019; Arigela et al., 2021; Guo et al., 2021).

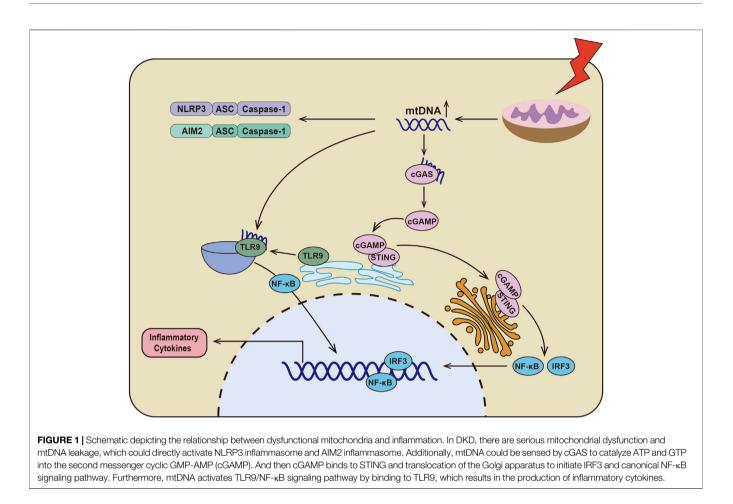
Inflammasome Activation

Inflammasome was first proposed by Tschopp et al., in 2002, which could modulate inflammation by the activation of caspases (Martinon et al., 2002; Schroder and Tschopp, 2010; Guo et al., 2015). There are canonical inflammasomes and non-canonical inflammasomes. The former are caspase-1-dependent

inflammasomes and the non-canonical inflammasomes depend on caspase-11 in mouse and caspase-4 or caspase-5 (the homologue of caspase-11) in humans (Stowe et al., 2015; Van Opdenbosch and Lamkanfi, 2019). It is well known that the activation of the inflammasome is associated with a series of pattern recognition receptors (PRRs) in the membrane and cytoplasm, which include nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs), Toll-like receptors (TLRs), C-type lectin receptors (CLRs), Rig-I-like receptors (RLRs), and absent in melanoma 2 (AIM2)-like receptors (ALRs) (Schroder and Tschopp, 2010; Strowig et al., 2012; Evavold and Kagan, 2019). This series of PRRs could rapidly respond to danger signals such as PAMPs or DAMPs to activate inflammasomes (Guo et al., 2015).

Accumulated data suggests that the inflammasome contributes to the progression of DKD (Strowig et al., 2012; Masood et al., 2015). Upregulated expression of IL-1 β and Interleukin 18 (IL-18) was observed in the peripheral blood or renal tissue of DKD patients (Uzu et al., 2011; Lei et al., 2019). In addition, the activation of the NLRP3 inflammasome was not only present in the renal infiltrating macrophages (Shahzad et al., 2015; Zhang J. et al., 2021), but also in podocytes (Xiong et al., 2020; Wu et al., 2021), tubular epithelial cells (Han et al., 2019; Xie et al., 2019) and mesangial cells (Yi et al., 2017; Tung et al., 2018). Inhibition or knockout of NLRP3 has also been shown to be effective in delaying the progression of DKD. It has been reported that the NLRP3-specific small molecule inhibitor MCC950 could alleviate renal damage and fibrosis in db/db mice (Wang et al., 2017; Zhang W. et al., 2019; Ding et al., 2021) and knockdown of NLRP3 has a significant reno-protective effect in STZ-induced diabetic mice (Wu et al., 2018). In addition the NLRP3 inflammasome, NLRP1 and NLRC4 to inflammasome were also activated in the kidneys of DKD mice and associated with increased urinary albumin excretion and renal damage (Yuan et al., 2016; Soares et al., 2018). Furthermore, Luan et al. demonstrated that NLRC5 gene deficiency also reduced the inflammation and renal damage and delayed the progression of renal fibrosis in STZ-induced diabetic mice (Luan et al., 2018).

Other novel inflammasomes such as NLRP2, NLRP6, NLRP10, NLRP12, and AIM2 are also found to be involved in the regulation of inflammation and their expressions are upregulated in various kidney diseases (Yuan et al., 2016; Luan et al., 2018; Wu et al., 2018). It has been shown that the above inflammasomes participate in the pathogenesis of non-diabetic kidney diseases (Lech et al., 2010; Komada et al., 2018; Valino-Rivas et al., 2020). Valiño-Rivas et al. found that NLRP6 deficiency aggravated ischemia-reperfusion (I/R) induced acute kidney injury (Valino-Rivas et al., 2020). The AIM2 inflammasome is a sensor for endogenous Double Stranded DNA (dsDNA) that mediates canonical and non-canonical inflammasome activation. The expression level of AIM2 inflammasome is also associated with the progression of hepatitis B-related glomerulonephritis and lupus nephritis (Zhang et al., 2013; Zhen et al., 2014; Komada et al., 2018). These data suggested that the inflammasomes might also have potential role in the pathogenesis of chronic kidney disease



(CKD). However, there is currently a lack of studies on investigating the effect of inflammasomes above in DKD, thus it needs to be further explored in future.

AMPK Signaling Pathways

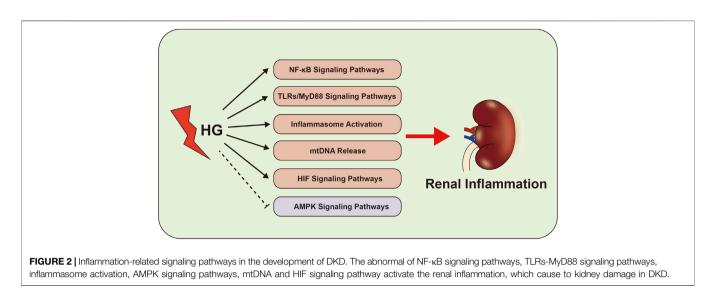
AMP-activated protein kinase (AMPK) is a serine/threonine kinase that is involved in regulating cellular metabolism by activating energy-producing pathways (Cordero et al., 2018; Lyons and Roche, 2018; Larabi et al., 2020) (Wang et al., 2018), In addition, AMPK plays a key role in regulating autophagy (Li and Chen, 2019) and other biological functions. Recent studies have also suggested that AMPK is involved in inflammation, particularly the activation of NLRP3 inflammasome (Youm et al., 2015; Xian et al., 2021). It is wellknown that the AMPK dysfunction is associated with a variety of diseases including diabetes mellitus (Chen et al., 2021) and cancers (Wang et al., 2016).

Interestingly, AMPK signaling pathway was related to the renal inflammation (Garcia and Shaw, 2017). Previous study by our group showed that DsbA-L could inhibit the activation of NLRP3 by activating AMPK phosphorylation in the kidney of DKD mice, while the expression levels of IL-18 and IL-1 β were increased in that of DKD mice treated with compound C (an AMPK inhibitor) (Yang et al., 2021a). AMPK also inhibits the expression of inducible nitric oxide synthase (iNOS), thereby

inhibiting the activation of NLRP3 inflammasome (Pilon et al., 2004; Liao et al., 2021). These studies confirmed the renoprotective and anti-inflammatory effects of AMPK signaling pathway in DKD. In addition, AMPK activation induced mitophagy, which could clear damaged mitochondria in cells and reduce the production of ROS and activation of NLRP3 inflammasome (Seabright et al., 2020; Yang et al., 2021b; Drake et al., 2021). What's more, a variety of drugs for the treatments of DKD such as metformin, could also activate phosphorylation of AMPK to inhibit renal inflammation (Zhang X.-X. et al., 2020). Therefore, understanding mechanisms of AMPK signaling pathway would be critical to improving the strategy of DKD prevention and treatment.

Mitochondrial DNA Release

When mitochondria are dysfunctional, the mitochondrial DNA (mtDNA) translocates to cytoplasm and initiates the inflammation through various signaling pathways (**Figure 1**) (Zhang et al., 2010; Fang et al., 2016; Riley and Tait, 2020). First, mtDNA, as an endogenous DAMPs, could directly activate the inflammasomes (Nakahira et al., 2011), which conclude NLRP3 (Shimada et al., 2012; Zhong et al., 2018) and AIM2 (Bae et al., 2019; Xu et al., 2021). In addition, mtDNA could initiate the cGAS-STING signaling pathway. The mtDNA is translocated to the cytoplasm and associated with the DNA



receptor cyclic GMP-AMP Synthase (cGAS), causing the activation of cGAS, which catalyzes ATP and GTP into the second messenger, cyclic GMP-AMP (cGAMP) (Wu et al., 2013). Stimulator of interferon genes (STING) was localized in the endoplasmic reticulum and activated by cGAMP. STING then translocated to the Golgi apparatus to initiate interferon regulatory factor 3 (IRF3) and canonical NF-kB signaling pathway (Ishikawa and Barber, 2008; Hopfner and Hornung, 2020). Recently, it has also been found that the hypomethylated mtDNA is the ligands of TLR9 for regulating inflammation (Goulopoulou et al., 2012; Rodriguez-Nuevo et al., 2018). When cells are under stress, TLR9 is translocated to endolysosome from endoplasmic reticulum (Latz et al., 2004; Leifer et al., 2004; Fang et al., 2016), where hypomethylated mtDNA binds to TLR9 to activate TLR9/NF-KB and TLR9/MyD88 signaling pathways, finally resulting in increased production of TNF-α, IL-6, IL-1β, and MCP-1 (Zhang et al., 2014; Wei et al., 2015; Rodriguez-Nuevo et al., 2018). These data indicat that mtDNA release after mitochondria damage plays a key role in the inflammatory response.

As we know that proximal tubular cells of the kidney are rich in mitochondria and mitochondrial dysfunction and mtDNA release have been recognized in various kidney disease (Forbes and Thorburn, 2018; Ahmad et al., 2021). A large amount of mtDNA was detected in the blood and urine of patients with DKD, which is also related to the severity of interstitial fibrosis (Malik et al., 2009; Huang Y. et al., 2020; Jin et al., 2021). The level of mtDNA was increased in primary mesangial cells treated with high glucose, which is accompanied with enhanced TLR9 activation (Czajka et al., 2015). Chung et al. confirmed that STING gene deficiency significantly reduced renal fibrosis and the expression of TNF- α , IL-1 β , IL-6 and MCP1 (Chung et al., 2019). There is no doubt that the activation of the mtDNA-cGAS-STING signaling pathway is related to renal fibrosis in DKD. Meanwhile, Myakala et al. also found that sacubitril/valsartan could regulate mitochondrial function and alleviate mtDNA release to inhibit mtDNA-cGAS-STING signaling pathway, ultimately delaying the progression of DKD. This data

indicated that targeting mtDNA may be a powerful solution for the treatment of DKD. In addition, it has been reported that mitophagy inhibits the activation of NLRP3 inflammasome and the secretion of inflammatory cytokines such as IL-1 β in DKD (Han et al., 2021). These data suggest that mitochondrial homeostasis disruption and mtDNA release are important initiator in renal inflammation, and it might be a potential therapeutic target for the treatment of DKD.

HIF Signaling Pathways

Hypoxia-inducible factor (HIF) is conserved in biological evolution and is an important mediator of inflammation (Palazon et al., 2014; McGettrick and O'Neill, 2020). The structure of HIF contains α and β subunits, which include three a subunits (HIF-1a, HIF-2a, HIF-3a) and three β subunits (HIF-1 β , HIF-2 β , HIF- 3 β). It has been reported that HIF-1 α and HIF-2 α are related to the inflammation (Keith et al., 2011). A large amount of evidence shows that the activation of HIF-1α under hypoxia regulates the transcription level of NF-κB (Palazon et al., 2014). Additionally, the fatty acid oxidation (FAO)-mediated NLRP3 inflammasome is activated when the HIF-2a is deficient in macrophage (Li et al., 2021). Of note, inflammation also results in the activation of HIF signaling pathway (Watts and Walmsley, 2019; McGettrick and O'Neill, 2020). Zhang et al. found that SARS-CoV-2 infection induced nuclear translocation of NF-KB to upregulate the mRNA level of HIF-1a via IL18/IL18R1 (Zhang L. et al., 2021). In addition, endogenous DAMPs could upregulate expression of HIF-1a and induced the production of IL-1β (Tannahill et al., 2013). It could be seen that the HIF signaling pathway and inflammation are interdependent in multiple dimensions.

Recent studies have shown that HIF signaling pathway is closely related to the development of renal interstitial fibrosis in DKD (Li et al., 2019; Jiang et al., 2020). The activation of HIF signaling indicates that the kidney is intolerant to hypoxia, which contributes to the initial adaptive response to hypoxia and tissue repair (Isoe et al., 2010; Garcia-Pastor et al., 2019). Our team has found that HIF-1 α activation exerts a reno-protective effect by regulating mitochondrial dynamics through HO-1 in the early stage of DKD in STZ-induced diabetic mice (Jiang et al., 2020). Yu et al. also found that HIF-1a/Parkin/PINK1-mediated mitophagy improved mitochondrial function in renal tubular epithelial cells, whereas inhibition of HIF-1a by YC-1 (a specific inhibitor of HIF-1a) promoted inflammation and increased the production of IL-1 β and IL-18 (Yu et al., 2021). These data indicate the reno-protective effect of HIF in the early stage of DKD. However, HIF signaling pathway could also promote chronic inflammation and interstitial fibrosis in CKD including the development of DKD (Wang et al., 2014; Catrina and Zheng, 2021). Co-treatment of mesangial cells with HG and HIF-1a siRNA reduce the production of inflammation and fibrosis factors such as endothelin-1, TGF-β1, CTGF and VEGF (Shao et al., 2016). It has been reported that high doses of MK-8617 (hypoxia-inducible factor-prolyl hydroxylase inhibitor) can promote the tubulointerstitial fibrosis by activating HIF-1a/KLF5/TGF-B1 axis (Li et al., 2019). These data indicate that persistent activation of HIF signaling pathway is an important mediator of the progression of renal interstitial fibrosis and is closely related to the inflammation of DKD (Figure 2).

In addition to be involved in the pathogenesis of DKD, inflammation has recently been shown to play a role in diabetic cardiomyopathy (Quagliariello et al., 2020). Under hyperglycemia condition, it has been demonstrated that the heart would be more susceptible to drug damage or ischemiareperfusion injury because of hyperglycemia-induced inflammation (Peng et al., 2015; Quagliariello et al., 2020). Quagliariello et al. found that hyperglycemia could activate the NLRP3 inflammasome in cardiomyocytes and enhance the cardiotoxicity of ipilimumab (an anti-cancer drug for breast cancer) (Quagliariello et al., 2020). Additionally, diabetic cardiomyopathy might result in low myocardial reserve (Moir et al., 2006). Treatment of kaempferol to hyperglycemia-induced myocardial damage might reduce ROS production and the expression of TNF-α by inhibiting the NF-κB signaling pathway (Chen et al., 2018). These data suggest that HG leads to cardiac damage and decreases myocardial reserve through the inflammatory signaling pathways.

ANTI-INFLAMMATION TREATMENT IN DKD

The treatments of DKD mainly include management of hyperglycemia, hypertension and hyperlipidemia. Actually, these drugs especially the nonsteroidal selective mineralocorticoid receptor antagonist (MRA) finerenone and some Chinese traditional medicine could alleviate the development of DKD by regulating the inflammation.

Metformin

Metformin has been used for treatment of diabetes mellitus since 1950s, and is now the most widely used anti-hyperglycemic drug (Flory and Lipska, 2019). There's a lot of evidence that metformin has not only anti-hyperglycemic properties, but also renoprotective effect. A cohort study showed that treatment of 10,426 patients with type 2 diabetes and chronic kidney disease at stage 3B with metformin significantly reduced the risk of all-cause mortality and ESRD events (Kwon et al., 2020). Similar result was observed in metformin treated DKD animal models, which also showed improved renal function and alleviated renal tubulointerstitial fibrosis (Zhang Q. et al., 2017). Recently, studies have shown that the reno-protective effect of metformin might relate to the mechanisms of anti-inflammation. Among them, AMPK signaling pathway might be the most important anti-inflammatory mechanism of metformin. Recently, Ma et al. found that metformin inhibited v-ATPase activity by inducing the binding of PEN2 to the ATP6AP1 subunit, and then activated lysosomal AMPK (Ma et al., 2022). The work of Han et al. suggested that metformin maintained mitochondrial homeostasis through AMPK-mediated mitophagy and inhibited the activation of NLRP3 inflammasome in DKD (Han et al., 2021). In addition to AMPK signaling pathway, metformin could also regulate renal inflammation by HIF signaling pathway (Takiyama et al., 2011), intestinal flora (Induri et al., 2022), and autophagy (Bharath et al., 2020).

Finerenone

Nonsteroidal selective MRA is a novel drug with cardio-renal protection. Compared with traditional MRAs such as spironolactone and eplerenone, nonsteroidal selective MRA has significantly improved efficacy and safety. Finerenone (also known as BAY 94-8862) is currently approved by FDA for treatment of DKD. A series of clinical studies confirmed the reno-protective effect of finerenone in DKD. The phase III FIDELIO-DKD trial has shown that finerenone significantly delayed renal failure and eGFR decline. In addition to finerenone, several nonsteroidal MRAs are currently in development (Patel et al., 2021), such as AZD9977 (phase I) (Whittaker et al., 2020), apararenone (phase II) (Wada et al., 2021) and esaxerenone (phase III) (Ito et al., 2020), which might also have reno-protective effect in DKD by regulating inflammation.

It has been well known that overactivation of MR is a key event for chronic inflammation by increasing the recruitment of neutrophil, macrophage and Th1 & Th17 cells and upregulating the expression of pro-inflammatory factors and fibrotic-related factors including TGF-a, endothelin 1, PAI-1, CTGF (Young and Rickard, 2015). There is sufficient evidence to suggest that treatment with MRA including finerenone might alleviate the progression of chronic kidney disease and eliminate the renal inflammation by reducing the expression of proinflammatory cytokines, such as MCP-1, TNF-a and Matrix metalloproteinase-12 (MMP-12) (Han et al., 2006; Huang et al., 2014). Martínez et al. found that finerenone regulated the activation of NF-kB signaling pathway through neutrophil gelatinase-associated lipocalin (NGAL) and inhibited the inflammation in cardiac remodeling after myocardial infarction (Martinez-Martinez et al., 2017). Bhuiyan et al. also found that esaxerenone, another nonsteroidal selective, significantly reduced the expression of MCP-1 and inflammatory cell infiltration in DKD by inhibiting ROS (Bhuiyan et al., 2019). These studies suggest that nonsteroidal selective MRA can alleviate the progression of DKD through targeting inflammatory factors.

SGLT2 Inhibitors

Sodium-glucose cotransporter 2 inhibitor (SGLT2i) is a novel anti-hyperglycemic drug by inhibiting reabsorption of glucose in the kidney (Ferrannini, 2017). Several clinical trials have confirmed that SGLT2i have excellent cardio-renal benefit and low risk of hypoglycemia (Perkovic et al., 2019). Interestingly, the SGLT2i could reduce the tissue low-grade inflammation (Bonnet and Scheen, 2018; Packer, 2020). A retrospective study showed that compared to glimepiride, SGLT2i canagliflozin significantly reduced the expression levels of inflammatory markers and fibrosis markers, such as tumor necrosis factor receptor 1 (TNFR1), matrix metalloproteinase-7 (MMP7), Fibronectin (FN) (Heerspink et al., 2019) and IL-6. The anti-inflammatory mechanism of SGLT2i might relate to HIF-1a in DKD (Bessho et al., 2019). In addition, SGLT2i also can regulate mitochondrial energy metabolism through reducing mitochondrial Ca²⁺ concentration and activating AMPK signaling pathway (Hawley et al., 2016; Baartscheer et al., 2017). Recent study demonstrates that Canagliflozin exerts the anti-inflammatory effect by activating AMPK-Akt-eNOS pathway (Heerspink et al., 2019). Interestingly, it has also been reported that SGLT2i could inhibit inflammasome activation. Kim et al. found that compared to sulfonamide, SGLT2i significantly inhibited the activation of NLRP3 inflammasome and reduced the production of IL-1 β (Kim et al., 2020). Birnbaum et al. also found that SGLT2i could inhibit the activation of NLRP3 inflammasome and reduce the mRNA expression of IL-1β, IL-6, TNF-α in BTBR ob/ob mice (Birnbaum et al., 2018). In conclusion, the reno-protective effect of SGLT2i in DKD might be related to anti-inflammation.

DPP-4 Inhibitor and GLP-1 Receptor Agonist

Glucagon-like peptide 1 (GLP-1) is an incretin hormone that stimulates insulin secretion in response to food intake (Baggio and Drucker, 2007). GLP-1 binds to the GLP-1 receptor in the pancreas and then regulates blood sugar by promoting insulin secretion. The half-life of natural GLP-1 is very short (1-2 min) because it is rapidly degraded by the ubiquitous proteolytic enzyme DPP-4 (Bell et al., 1983; Hansen et al., 1999). Recently, dipeptidyl peptidase-4 inhibitor (DDP-4i) and GLP-1 receptor agonists (GLP1RA) have been widely used in antihyperglycemic medications by interfering with GLP-1 expression (McGuire et al., 2019; Shaman et al., 2022). Recent studies demonstrated that DDP-4i and GLP-1RA could not only control blood sugar by regulating the secretion of insulin, but also exert anti-inflammatory and reno-protective effects in DKD (Gangadharan et al., 2016; Coppolino et al., 2018; Shaman et al., 2022). Many studies have suggested that DDP-4i could reduce the secretion of MCP-1 from renal parenchymal cells and inhibit the chemotaxis and activation of mono-macrophages (Ishibashi et al., 2011; Shah et al., 2011; Coppolino et al., 2018). DDP-4i could also inhibit the activity of NF-κB signaling pathway in renal tissue and reduce the expression of TNF-a in STZ-induced diabetic mice (Kodera et al., 2014; Gangadharan et al., 2016). Additionally, supplementation of exendin-4 (a GLP-1 analog) could reduce the secretion of proinflammatory factors in the kidney of diabetic mouse, including TNF-α, IL-1β, MCP-1, Intercellular Adhesion Molecule 1 (ICAM-1), etc. and alleviate the progression of DKD (Hendarto et al., 2012; Sancar-Bas et al., 2015). Furthermore, Hasan et al. found that in GLP-1R-deficient mice with 5/6 nephrectomy, linagliptin (a DDP-4 inhibitor) also significantly inhibited the expression of TGF-β, Collagen I, and Phospho-Mothers against decapentaplegic homolog 3 (pSMAD3) and reduced renal fibrosis (Hasan et al., 2019). These studies suggest that DPP-4i and GLP-1RA might have a reno-protective role in DKD through anti-inflammation, which is independent of the anti-hyperglycemia effect. The role of GLP-1 and DDP-4i on anti-inflammation have been confirmed, as well. Zobel el al. found liraglutide treatment (1.8 mg/day) could reduce the gene expression of TNF- α , IL-1 β in the peripheral blood mononuclear cells (PBMC) of patients with type 2 diabetes (Zobel et al., 2021). The study of Tremblay et al. also indicated that the treatment with sitagliptin significantly reduced the plasma levels of C-reactive protein (CRP), IL-6, IL-18, secreted phospholipase-A2 (sPLA2), soluble intercellular adhesion molecule-1 and E-selectin (Tremblay et al., 2014).

Traditional Chinese Medicine

Traditional Chinese medicines (TCM) such as Astragalus, Cordyceps sinensis, Tripterygium wilfordii, Fructus arctii, Panax notoginseng, Berberine, etc. are also powerful weapons against DKD (Wen et al., 2017; Zhong et al., 2019). A systematic review and meta-analysis including 29 trials suggested that TCM significantly reduced urinary protein excretion rate and urinary protein (Xiao et al., 2013). Furthermore, Chinese herbal medicines combined with ACEI or ARB have a better effect in reducing urinary protein in patients with DKD compared to ACEI or ARB therapy alone? (Xiao et al., 2013). Some studies indicate that the mechanism of TCM in DKD might be related to antiinflammation. The study by Zhong et al. suggested that arctigenin, an extract of burdock, reduced proteinuria in diabetic mice through inhibiting NF-KB-mediated inflammation by binding to protein phosphatase 2 A (PP2A) in podocytes exposed to HG ambiance (Zhong et al., 2019). The Tripterygium wilfordii Hook. f (GTW) could inhibit the activation of NF-KB signaling pathway and reduce inflammatory cell infiltration as well as the mRNA expression of IL-1 β and TNF- α in STZ-induced diabetic mice combined with unilateral nephrectomy (Wu et al., 2017). Guo et al. found that maackiain could reduce renal inflammation and the expression of MCP-1 and TNF-a by inhibiting TLRs-MyD88 signaling pathway in STZ-induced diabetic rat (Guo et al., 2021). In addition, the antiinflammatory effect of TCM is also related to reducing ROS, regulating autophagy and inhibiting epithelial-mesenchymal transition (Li et al., 2017; Zhang et al., 2021a; Zhang et al., 2021b). All of these researches demonstrate that TCM might be a potential weapon for the treatment of DKD by anti-inflammation.

In addition to TCM, nutraceuticals derived from Chinese herbal medicines could prevent DKD, for example, boswellic acid, curcumin and quercetin (Lu et al., 2015; Lu et al., 2017; Asgharpour and Alirezaei, 2021). A systematic review and Metaanalysis revealed that curcumin supplementation significantly improved blood sugar, lipid and blood pressure control and decreased the serum creatinine in DKD patients (Jie et al., 2021). Lu et al. also found that curcumin could delay fibrosis progression in DKD mice by inhibiting the initiation of NLRP3 inflammasome and reduce the expression of IL-1 β in renal tissue (Lu et al., 2017). Additionally, the present studies have revealed the protective effect of quercetin, a natural AMPK activator, in STZ-induced diabetic rat (Zhang J. et al., 2021). Lu et al. also found that treatment of quercetin could alleviate the progression of renal fibrosis in STZ-induced diabetic rat (Lu et al., 2015).

However, TCM has limitations such as low bioavailability, which greatly limits its broad application in the treatment of DKD (Liu et al., 2013; Zhang C. et al., 2019). The study of Rosso et al. suggested that hyaluronic acid (HA-NPs) and sub-micron particles (a CD44-targeted vector) were effective means to deliver bio-actives to targeted tissue in a specific and controlled manner (Rosso et al., 2013). This suggests that delivering TCM to the kidney might be a potential strategy to improve the bioavailability. Importantly, the expression of CD44 is low in the normal kidney and would specifically increase during renal damage (Zhao et al., 2019). The study of Hu et al. confirmed that CD44-targeted hyaluronic acid-curcumin prodrug could reduce the ROS in renal tubular epithelial cell. Moreover, Wang et al. have found the bioavailability and therapeutic effect of rhein, an active ingredient of TCM, were significantly improved in STZ-induced diabetic mice through kidney-targeted rhein-loaded liponanoparticles (Wang et al., 2019). This study further confirmed the feasibility of kidney-targeted therapies. Thus, it might be effective to transport TCM to the kidney through hyaluronic vector, liponanoparticle, etc. However, this conjecture needs be verified through further experiments.

Mesenchymal Stem Cells and Extracellular Vesicles

Mesenchymal Stem Cells (MSCs) and extracellular vesicles (EVs) are current research hotspots and pre-clinical agents for the treatment of DKD (Tang et al., 2019; Savio-Silva et al., 2020; Shen et al., 2020). Previous studies suggested that systemic injection of MSCs improved glycemic control but showing no renal protection (Savio-Silva et al., 2020). Recent evidence suggests that proteinuria is decreased after inducing renal homing of MSC by ultrasound-targeted micro-bubble destruction (UTMD) (Marquez-Curtis and Janowska-Wieczorek, 2013). Xiang et al. found that intravenous injection of human umbilical cordderived MSCs could reduce the expression of inflammatory factors such as IL-6, IL-1 β and TNF- α in the kidney and blood, and alleviate the renal interstitial fibrosis in the STZ-induced diabetic rat (Xiang et al., 2020). These studies suggest that MSCs treatment might be beneficial to reduce the renal inflammation and alleviate the progression of DKD.

The EVs include exosomes, microvesicles and apoptotic bodies. It has been found that the EVs play an important role in the progression of DKD (Tang et al., 2019). Under the stimulation of exosomes secreted from HG-treated macrophages, the mRNA and protein levels of TNF- α , IL-1 β and MCP-1 in renal mesangial cells increased. Furthermore, intravenous injection of the exosomes into C57BL/6 mice

resulted in the increased expression of TNF- α , pro-IL-1 β and IL-1 β and renal interstitial fibrosis (Zhu et al., 2019). This data suggested that EVs secreted from damaged macrophage or renal cells could promote renal inflammation and interstitial fibrosis in DKD. Additionally, Jiang et al. have found that intravenous injection of the exosomes secreted by human urine-derived stem cells could inhibit the apoptosis of podocytes and renal tubular epithelial cells, and reduce urinary microalbumin excretion in STZ-induced diabetic rat (Jiang et al., 2016). This suggests that EVs derived from MSCs may be excellent biological agents to alleviate the progression of interstitial fibrosis in DKD.

DISCUSSION

Inflammation plays an important role in the progression of DKD (Perez-Morales et al., 2019). In this review, we have discussed and summarized the role of NF- κ B signaling pathways, AMPK TLRs-MyD88 signaling pathways, signaling pathway, inflammasome activation, mtDNA release and HIFsignaling pathway in the inflammation-induced renal damage of DKD. To date, drugs directly targeting inflammation are limited in treatment of patients with DKD, but the commonly used antihyperglycemic drugs such as metformin, SGLT2i, DPP-4i, GLP-1AR and TCM also exhibit excellent anti-inflammatory effect. Interestingly, finerenone (a reno-protective nonsteroidal selective MRA) also exhibits strong anti-inflammatory efficacy to protect the kidney and heart in patients with DKD. This data suggested that attention should be paid to inflammation and anti-Inflammation therapy in DKD in future research.

In addition, MSCs and EVs have shown the curative effect and anti-inflammatory effect in the treatment of DKD animal models (Tang et al., 2019). However, both MSCs and EVs are mainly used for animal experiments currently and their efficacy and safety are uncertain in human body.

AUTHOR CONTRIBUTIONS

CBL and MY wrote the first draft of the manuscript. SLL, JFY, and CRL provided consultations on the preparation of the work. During the revision, LL and HFL participated in editing and revising the manuscript. LS contributed to manuscript revision and approved the submitted version.

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REFERENCES

- Afonina, I. S., Zhong, Z., Karin, M., and Beyaert, R. (2017). Limiting Inflammation-The Negative Regulation of NF-κB and the NLRP3 Inflammasome. *Nat. Immunol.* 18, 861–869. doi:10.1038/ni.3772
- Ahmad, A. A., Draves, S. O., and Rosca, M. (2021). Mitochondria in Diabetic Kidney Disease. *Cells* 10, 2945. doi:10.3390/cells10112945
- Anders, H.-J., Huber, T. B., Isermann, B., and Schiffer, M. (2018). CKD in Diabetes: Diabetic Kidney Disease versus Nondiabetic Kidney Disease. *Nat. Rev. Nephrol.* 14, 361–377. doi:10.1038/s41581-018-0001-y
- Araújo, L. S., Torquato, B. G. S., da Silva, C. A., dos Reis Monteiro, M. L. G., dos Santos Martins, A. L. M., da Silva, M. V., et al. (2020). Renal Expression of Cytokines and Chemokines in Diabetic Nephropathy. *BMC Nephrol.* 21, 308. doi:10.1186/s12882-020-01960-0
- Arigela, C. S., Nelli, G., Gan, S. H., Sirajudeen, K. N. S., Krishnan, K., Abdul Rahman, N., et al. (2021). Bitter Gourd Honey Ameliorates Hepatic and Renal Diabetic Complications on Type 2 Diabetes Rat Models by Antioxidant, Antiinflammatory, and Anti-apoptotic Mechanisms. *Foods* 10, 2872. doi:10.3390/ foods10112872
- Asgharpour, M., and Alirezaei, A. (2021). Herbal Antioxidants in Dialysis Patients: A Review of Potential Mechanisms and Medical Implications. *Ren. Fail.* 43, 351–361. doi:10.1080/0886022X.2021.1880939
- Baartscheer, A., Schumacher, C. A., Wüst, R. C. I., Fiolet, J. W. T., Stienen, G. J. M., Coronel, R., et al. (2017). Empagliflozin Decreases Myocardial Cytoplasmic Na+ through Inhibition of the Cardiac Na+/H+ Exchanger in Rats and Rabbits. *Diabetologia* 60, 568–573. doi:10.1007/s00125-016-4134-x
- Bae, J. H., Jo, S. I., Kim, S. J., Lee, J. M., Jeong, J. H., Kang, J. S., et al. (2019). Circulating Cell-free mtDNA Contributes to AIM2 Inflammasome-Mediated Chronic Inflammation in Patients with Type 2 Diabetes. *Cells* 8, 328. doi:10. 3390/cells8040328
- Baggio, L. L., and Drucker, D. J. (2007). Biology of Incretins: GLP-1 and GIP. Gastroenterology 132, 2131–2157. doi:10.1053/j.gastro.2007.03.054
- Bell, G. I., Sanchez-Pescador, R., Laybourn, P. J., and Najarian, R. C. (1983). Exon Duplication and Divergence in the Human Preproglucagon Gene. *Nature* 304, 368–371. doi:10.1038/304368a0
- Bessho, R., Takiyama, Y., Takiyama, T., Kitsunai, H., Takeda, Y., Sakagami, H., et al. (2019). Hypoxia-inducible Factor-1α Is the Therapeutic Target of the SGLT2 Inhibitor for Diabetic Nephropathy. *Sci. Rep.* 9, 14754. doi:10.1038/ s41598-019-51343-1
- Bharath, L. P., Agrawal, M., McCambridge, G., Nicholas, D. A., Hasturk, H., Liu, J., et al. (2020). Metformin Enhances Autophagy and Normalizes Mitochondrial Function to Alleviate Aging-Associated Inflammation. *Cell Metab.* 32, 44–55. doi:10.1016/j.cmet.2020.04.015
- Bhuiyan, A. S., Rafiq, K., Kobara, H., Masaki, T., Nakano, D., and Nishiyama, A. (2019). Effect of a Novel Nonsteroidal Selective Mineralocorticoid Receptor Antagonist, Esaxerenone (CS-3150), on Blood Pressure and Renal Injury in High Salt-Treated Type 2 Diabetic Mice. *Hypertens. Res.* 42, 892–902. doi:10. 1038/s41440-019-0211-0
- Birnbaum, Y., Bajaj, M., Yang, H.-C., and Ye, Y. (2018). Combined SGLT2 and DPP4 Inhibition Reduces the Activation of the Nlrp3/ASC Inflammasome and Attenuates the Development of Diabetic Nephropathy in Mice with Type 2 Diabetes. Cardiovasc Drugs Ther. 32, 135–145. doi:10.1007/s10557-018-6778-x
- Bonnet, F., and Scheen, A. J. (2018). Effects of SGLT2 Inhibitors on Systemic and Tissue Low-Grade Inflammation: The Potential Contribution to Diabetes Complications and Cardiovascular Disease. *Diabetes & Metabolism* 44, 457–464. doi:10.1016/j.diabet.2018.09.005
- Catrina, S.-B., and Zheng, X. (2021). Hypoxia and Hypoxia-Inducible Factors in Diabetes and its Complications. *Diabetologia* 64, 709–716. doi:10.1007/s00125-021-05380-z
- Chen, M., Huang, N., Liu, J., Huang, J., Shi, J., and Jin, F. (2021). AMPK: A Bridge between Diabetes Mellitus and Alzheimer's Disease. *Behav. Brain Res.* 400, 113043. doi:10.1016/j.bbr.2020.113043
- Chen, X., Qian, J., Wang, L., Li, J., Zhao, Y., Han, J., et al. (2018). Kaempferol Attenuates Hyperglycemia-Induced Cardiac Injuries by Inhibiting Inflammatory Responses and Oxidative Stress. *Endocrine* 60, 83–94. doi:10. 1007/s12020-018-1525-4

- Chung, K. W., Dhillon, P., Huang, S., Sheng, X., Shrestha, R., Qiu, C., et al. (2019). Mitochondrial Damage and Activation of the STING Pathway Lead to Renal Inflammation and Fibrosis. *Cell Metab.* 30, 784–799. doi:10.1016/j.cmet.2019. 08.003
- Coppolino, G., Leporini, C., Rivoli, L., Ursini, F., di Paola, E. D., Cernaro, V., et al. (2018). Exploring the Effects of DPP-4 Inhibitors on the Kidney from the Bench to Clinical Trials. *Pharmacol. Res.* 129, 274–294. doi:10.1016/j.phrs.2017.12.001
- Cordero, M. D., Williams, M. R., and Ryffel, B. (2018). AMP-activated Protein Kinase Regulation of the NLRP3 Inflammasome during Aging. *Trends Endocrinol. Metabolism* 29, 8–17. doi:10.1016/j.tem.2017.10.009
- Cronkite, D. A., and Strutt, T. M. (2018). The Regulation of Inflammation by Innate and Adaptive Lymphocytes. J. Immunol. Res. 2018, 1–14. doi:10.1155/ 2018/1467538
- Czajka, A., Ajaz, S., Gnudi, L., Parsade, C. K., Jones, P., Reid, F., et al. (2015). Altered Mitochondrial Function, Mitochondrial DNA and Reduced Metabolic Flexibility in Patients with Diabetic Nephropathy. *EBioMedicine* 2, 499–512. doi:10.1016/j.ebiom.2015.04.002
- Deguine, J., and Barton, G. M. (2014). MyD88: A Central Player in Innate Immune Signaling. *F1000Prime Rep.* 6, 97. doi:10.12703/P6-97
- Ding, H., Li, J., Li, Y., Yang, M., Nie, S., Zhou, M., et al. (2021). MicroRNA-10 Negatively Regulates Inflammation in Diabetic Kidney via Targeting Activation of the NLRP3 Inflammasome. *Mol. Ther.* 29, 2308–2320. doi:10.1016/j.ymthe. 2021.03.012
- Donath, M. Y. (2014). Targeting Inflammation in the Treatment of Type 2 Diabetes: Time to Start. Nat. Rev. Drug Discov. 13, 465–476. doi:10.1038/ nrd4275
- Doshi, S. M., and Friedman, A. N. (2017). Diagnosis and Management of Type 2 Diabetic Kidney Disease. *Clin. J. Am. Soc. Nephrol.* 12, 1366–1373. doi:10.2215/ CJN.11111016
- Drake, J. C., Wilson, R. J., Laker, R. C., Guan, Y., Spaulding, H. R., Nichenko, A. S., et al. (2021). Mitochondria-localized AMPK Responds to Local Energetics and Contributes to Exercise and Energetic Stress-Induced Mitophagy. *Proc. Natl. Acad. Sci. U.S.A.* 118. doi:10.1073/pnas.2025932118
- Engelen, S. E., Robinson, A. J. B., Zurke, Y.-X., and Monaco, C. (2022). Therapeutic Strategies Targeting Inflammation and Immunity in Atherosclerosis: How to Proceed? *Nat. Rev. Cardiol.*, 1–21. doi:10.1038/s41569-021-00668-4
- Evavold, C. L., and Kagan, J. C. (2019). Inflammasomes: Threat-Assessment Organelles of the Innate Immune System. *Immunity* 51, 609–624. doi:10. 1016/j.immuni.2019.08.005
- Fang, C., Wei, X., and Wei, Y. (2016). Mitochondrial DNA in the Regulation of Innate Immune Responses. *Protein Cell* 7, 11–16. doi:10.1007/s13238-015-0222-9
- Ferrannini, E. (2017). Sodium-Glucose Co-transporters and Their Inhibition: Clinical Physiology. Cell Metab. 26, 27–38. doi:10.1016/j.cmet.2017.04.011
- Fitzgerald, K. A., and Kagan, J. C. (2020). Toll-like Receptors and the Control of Immunity. *Cell* 180, 1044–1066. doi:10.1016/j.cell.2020.02.041
- Flory, J., and Lipska, K. (2019). Metformin in 2019. Jama 321, 1926–1927. doi:10. 1001/jama.2019.3805
- Forbes, J. M., and Thorburn, D. R. (2018). Mitochondrial Dysfunction in Diabetic Kidney Disease. Nat. Rev. Nephrol. 14, 291–312. doi:10.1038/nrneph.2018.9
- Foresto-Neto, O., Albino, A. H., Arias, S. C. A., Faustino, V. D., Zambom, F. F. F., Cenedeze, M. A., et al. (2020). NF-κB System Is Chronically Activated and Promotes Glomerular Injury in Experimental Type 1 Diabetic Kidney Disease. *Front. Physiol.* 11, 84. doi:10.3389/fphys.2020.00084
- Gangadharan Komala, M., Gross, S., Zaky, A., Pollock, C., and Panchapakesan, U. (2016). Saxagliptin Reduces Renal Tubulointerstitial Inflammation, Hypertrophy and Fibrosis in Diabetes. *Nephrology* 21, 423–431. doi:10.1111/ nep.12618
- Garcia, D., and Shaw, R. J. (2017). AMPK: Mechanisms of Cellular Energy Sensing and Restoration of Metabolic Balance. *Mol. Cell* 66, 789–800. doi:10.1016/j. molcel.2017.05.032
- García-Pastor, C., Benito-Martínez, S., Moreno-Manzano, V., Fernández-Martínez, A. B., and Lucio-Cazaña, F. J. (2019). Mechanism and Consequences of the Impaired Hif-1a Response to Hypoxia in Human Proximal Tubular HK-2 Cells Exposed to High Glucose. *Sci. Rep.* 9, 15868. doi:10.1038/s41598-019-52310-6
- Goulopoulou, S., Matsumoto, T., Bomfim, G. F., and Webb, R. C. (2012). Toll-likeReceptor 9 Activation: A Novel Mechanism Linking Placenta-Derived

Mitochondrial DNA and Vascular Dysfunction in Pre-eclampsia. *Clin. Sci.* (*Lond*) 123, 429–435. doi:10.1042/CS20120130

- Guo, H., Callaway, J. B., and Ting, J. P.-Y. (2015). Inflammasomes: Mechanism of Action, Role in Disease, and Therapeutics. *Nat. Med.* 21, 677–687. doi:10.1038/ nm.3893
- Guo, J., Li, J., Wei, H., and Liang, Z. (2021). Maackiain Protects the Kidneys of Type 2 Diabetic Rats via Modulating the Nrf2/HO-1 and TLR4/NF-κB/Caspase-3 Pathways. *Dddt* 15, 4339–4358. doi:10.2147/DDDT.S326975
- Han, S.-Y., Kim, C.-H., Kim, H.-S., Jee, Y.-H., Song, H.-K., Lee, M.-H., et al. (2006). Spironolactone Prevents Diabetic Nephropathy through an Anti-inflammatory Mechanism in Type 2 Diabetic Rats. J. Am. Soc. Nephrol. 17, 1362–1372. doi:10. 1681/ASN.2005111196
- Han, Y.-c., Tang, S.-q., Liu, Y.-t., Li, A.-m., Zhan, M., Yang, M., et al. (2021). AMPK Agonist Alleviate Renal Tubulointerstitial Fibrosis via Activating Mitophagy in High Fat and Streptozotocin Induced Diabetic Mice. *Cell Death Dis.* 12, 925. doi:10.1038/s41419-021-04184-8
- Han, Y., Xu, X., Tang, C., Gao, P., Chen, X., Xiong, X., et al. (20192018). Corrigendum to 'Reactive Oxygen Species Promote Tubular Injury in Diabetic Nephropathy: The Role of the Mitochondrial Ros-Txnip-Nlrp3 Biological axis. *Redox Biol.* 16, 32–46. doi:10.1016/j.redox.2019.101216
- Hansen, L., Deacon, C. F., Ørskov, C., and Holst, J. J. (1999). Glucagon-Like Peptide-1-(7-36)Amide Is Transformed to Glucagon-like Peptide-1-(9-36) Amide by Dipeptidyl Peptidase IV in the Capillaries Supplying the L Cells of the Porcine Intestine1. *Endocrinology* 140, 5356–5363. doi:10.1210/endo.140. 11.7143
- Hasan, A. A., von Websky, K., Reichetzeder, C., Tsuprykov, O., Gaballa, M. M. S., Guo, J., et al. (2019). Mechanisms of GLP-1 Receptor-independent Renoprotective Effects of the Dipeptidyl Peptidase Type 4 Inhibitor Linagliptin in GLP-1 Receptor Knockout Mice with 5/6 Nephrectomy. *Kidney Int.* 95, 1373–1388. doi:10.1016/j.kint.2019.01.010
- Hawley, S. A., Ford, R. J., Smith, B. K., Gowans, G. J., Mancini, S. J., Pitt, R. D., et al. (2016). The na+/Glucose Cotransporter Inhibitor Canagliflozin Activates AMPK by Inhibiting Mitochondrial Function and Increasing Cellular AMP Levels. *Diabetes* 65, 2784–2794. doi:10.2337/db16-0058
- Heerspink, H. J. L., Perco, P., Mulder, S., Leierer, J., Hansen, M. K., Heinzel, A., et al. (2019). Canagliflozin Reduces Inflammation and Fibrosis Biomarkers: A Potential Mechanism of Action for Beneficial Effects of SGLT2 Inhibitors in Diabetic Kidney Disease. *Diabetologia* 62, 1154–1166. doi:10.1007/s00125-019-4859-4
- Hendarto, H., Inoguchi, T., Maeda, Y., Ikeda, N., Zheng, J., Takei, R., et al. (2012). GLP-1 Analog Liraglutide Protects against Oxidative Stress and Albuminuria in Streptozotocin-Induced Diabetic Rats via Protein Kinase A-Mediated Inhibition of Renal NAD(P)H Oxidases. *Metabolism* 61, 1422–1434. doi:10. 1016/j.metabol.2012.03.002
- Hopfner, K.-P., and Hornung, V. (2020). Molecular Mechanisms and Cellular Functions of cGAS-STING Signalling. *Nat. Rev. Mol. Cell Biol.* 21, 501–521. doi:10.1038/s41580-020-0244-x
- Hou, Y., Liang, H., Rao, E., Zheng, W., Huang, X., Deng, L., et al. (2018). Non-canonical NF-κB Antagonizes STING Sensor-Mediated DNA Sensing in Radiotherapy. *Immunity* 49, 490–503. doi:10.1016/j.immuni. 2018.07.008
- Huang, L. L., Nikolic-Paterson, D. J., Han, Y., Ozols, E., Ma, F. Y., Young, M. J., et al. (2014). Myeloid Mineralocorticoid Receptor Activation Contributes to Progressive Kidney Disease. J. Am. Soc. Nephrol. 25, 2231–2240. doi:10.1681/ ASN.2012111094
- Huang, W., Man, Y., Gao, C., Zhou, L., Gu, J., Xu, H., et al. (2020). Short-Chain Fatty Acids Ameliorate Diabetic Nephropathy via GPR43-Mediated Inhibition of Oxidative Stress and NF-κB Signaling. Oxidative Med. Cell. Longev. 2020, 1–21. doi:10.1155/2020/4074832
- Huang, Y., Chi, J., Wei, F., Zhou, Y., Cao, Y., and Wang, Y. (2020). Mitochondrial DNA: A New Predictor of Diabetic Kidney Disease. *Int. J. Endocrinol.* 2020, 1–7. doi:10.1155/2020/3650937
- Hussain, S., Habib, A., and Najmi, A. K. (2019). Limited Knowledge of Chronic Kidney Disease Among Type 2 Diabetes Mellitus Patients in India. *Ijerph* 16, 1443. doi:10.3390/ijerph16081443
- Induri, S. N. R., Kansara, P., Thomas, S. C., Xu, F., Saxena, D., and Li, X. (2022). The Gut Microbiome, Metformin, and Aging. Annu. Rev. Pharmacol. Toxicol. 62, 85–108. doi:10.1146/annurev-pharmtox-051920-093829

- Ishibashi, Y., Nishino, Y., Matsui, T., Takeuchi, M., and Yamagishi, S.-i. (2011). Glucagon-like Peptide-1 Suppresses Advanced Glycation End Product-Induced Monocyte Chemoattractant Protein-1 Expression in Mesangial Cells by Reducing Advanced Glycation End Product Receptor Level. *Metabolism* 60, 1271–1277. doi:10.1016/j.metabol.2011.01.010
- Ishikawa, H., and Barber, G. N. (2008). STING Is an Endoplasmic Reticulum Adaptor that Facilitates Innate Immune Signalling. *Nature* 455, 674–678. doi:10.1038/nature07317
- Isoe, T., Makino, Y., Mizumoto, K., Sakagami, H., Fujita, Y., Honjo, J., et al. (2010). High Glucose Activates HIF-1-Mediated Signal Transduction in Glomerular Mesangial Cells through a Carbohydrate Response Element Binding Protein. *Kidney Int.* 78, 48–59. doi:10.1038/ki.2010.99
- Ito, S., Kashihara, N., Shikata, K., Nangaku, M., Wada, T., Okuda, Y., et al. (2020). Esaxerenone (CS-3150) in Patients with Type 2 Diabetes and Microalbuminuria (ESAX-DN). *Clin. J. Am. Soc. Nephrol.* 15, 1715–1727. doi:10.2215/CJN.06870520
- Jiang, N., Zhao, H., Han, Y., Li, L., Xiong, S., Zeng, L., et al. (2020). HIF-1α Ameliorates Tubular Injury in Diabetic Nephropathy via HO-1-Mediated Control of Mitochondrial Dynamics. *Cell Prolif.* 53, e12909. doi:10.1111/cpr. 12909
- Jiang, Z.-z., Liu, Y.-m., Niu, X., Yin, J.-y., Hu, B., Guo, S.-c., et al. (2016). Exosomes Secreted by Human Urine-Derived Stem Cells Could Prevent Kidney Complications from Type I Diabetes in Rats. Stem Cell Res. Ther. 7, 24. doi:10.1186/s13287-016-0287-2
- Jie, Z., Chao, M., Jun, A., Wei, S., and LiFeng, M. (2021). Effect of Curcumin on Diabetic Kidney Disease: A Systematic Review and Meta-Analysis of Randomized, Double-Blind, Placebo-Controlled Clinical Trials. *Evidence-Based Complementary Altern. Med.* 2021, 1–14. doi:10.1155/2021/6109406
- Jin, L., Yu, B., Armando, I., and Han, F. (2021). Mitochondrial DNA-Mediated Inflammation in Acute Kidney Injury and Chronic Kidney Disease. Oxidative Med. Cell. Longev. 2021, 1–12. doi:10.1155/2021/9985603
- Johansen, K. L., Chertow, G. M., Foley, R. N., Gilbertson, D. T., Herzog, C. A., Ishani, A., et al. (2021). US Renal Data System 2020 Annual Data Report: Epidemiology of Kidney Disease in the united states. *Am. J. Kidney Dis.* 77, A7–A8. doi:10.1053/j.ajkd.2021.01.002
- Ka, S. M., Yeh, Y. C., Huang, X. R., Chao, T. K., Hung, Y. J., Yu, C. P., et al. (2012). Kidney-targeting Smad7 Gene Transfer Inhibits Renal TGF-β/MAD Homologue (SMAD) and Nuclear Factor κB (NF-κB) Signalling Pathways, and Improves Diabetic Nephropathy in Mice. *Diabetologia* 55, 509–519. doi:10. 1007/s00125-011-2364-5
- Kauppinen, A., Suuronen, T., Ojala, J., Kaarniranta, K., and Salminen, A. (2013). Antagonistic Crosstalk between NF-κB and SIRT1 in the Regulation of Inflammation and Metabolic Disorders. *Cell. Signal.* 25, 1939–1948. doi:10. 1016/j.cellsig.2013.06.007
- Keith, B., Johnson, R. S., and Simon, M. C. (2011). HIF1α and HIF2α: Sibling Rivalry in Hypoxic Tumour Growth and Progression. *Nat. Rev. Cancer* 12, 9–22. doi:10.1038/nrc3183
- Kim, S. R., Lee, S.-G., Kim, S. H., Kim, J. H., Choi, E., Cho, W., et al. (2020). SGLT2 Inhibition Modulates NLRP3 Inflammasome Activity via Ketones and Insulin in Diabetes with Cardiovascular Disease. *Nat. Commun.* 11, 2127. doi:10.1038/ s41467-020-15983-6
- Kodera, R., Shikata, K., Takatsuka, T., Oda, K., Miyamoto, S., Kajitani, N., et al. (2014). Dipeptidyl Peptidase-4 Inhibitor Ameliorates Early Renal Injury through its Anti-inflammatory Action in a Rat Model of Type 1 Diabetes. *Biochem. Biophysical Res. Commun.* 443, 828–833. doi:10.1016/j.bbrc.2013. 12.049
- Komada, T., Chung, H., Lau, A., Platnich, J. M., Beck, P. L., Benediktsson, H., et al. (2018). Macrophage Uptake of Necrotic Cell DNA Activates the AIM2 Inflammasome to Regulate a Proinflammatory Phenotype in CKD. J. Am. Soc. Nephrol. 29, 1165–1181. doi:10.1681/ASN.2017080863
- Komada, T., and Muruve, D. A. (2019). The Role of Inflammasomes in Kidney Disease. Nat. Rev. Nephrol. 15, 501–520. doi:10.1038/s41581-019-0158-z
- Kwon, S., Kim, Y. C., Park, J. Y., Lee, J., An, J. N., Kim, C. T., et al. (2020). The Long-Term Effects of Metformin on Patients with Type 2 Diabetic Kidney Disease. *Diabetes Care* 43, 948–955. doi:10.2337/dc19-0936
- Larabi, A., Barnich, N., and Nguyen, H. T. T. (2020). New Insights into the Interplay between Autophagy, Gut Microbiota and Inflammatory Responses in IBD. Autophagy 16, 38–51. doi:10.1080/15548627.2019.1635384

- Latz, E., Schoenemeyer, A., Visintin, A., Fitzgerald, K. A., Monks, B. G., Knetter, C. F., et al. (2004). TLR9 Signals after Translocating from the ER to CpG DNA in the Lysosome. *Nat. Immunol.* 5, 190–198. doi:10.1038/ni1028
- Lech, M., Avila-Ferrufino, A., Skuginna, V., Susanti, H. E., and Anders, H.-J. (2010). Quantitative Expression of RIG-like Helicase, NOD-like Receptor and Inflammasome-Related mRNAs in Humans and Mice. *Int. Immunol.* 22, 717–728. doi:10.1093/intimm/dxq058
- Leehey, D. J. (2020). Targeting Inflammation in Diabetic Kidney Disease: Is There a Role for Pentoxifylline? *Kidney360* 1, 292–299. doi:10.34067/KID.0001252019
- Lei, Y., Devarapu, S. K., Motrapu, M., Cohen, C. D., Lindenmeyer, M. T., Moll, S., et al. (2019). Interleukin-1β Inhibition for Chronic Kidney Disease in Obese Mice with Type 2 Diabetes. *Front. Immunol.* 10, 1223. doi:10.3389/fimmu.2019. 01223
- Leifer, C. A., Kennedy, M. N., Mazzoni, A., Lee, C., Kruhlak, M. J., and Segal, D. M. (2004). TLR9 Is Localized in the Endoplasmic Reticulum Prior to Stimulation. *J. Immunol.* 173, 1179–1183. doi:10.4049/jimmunol.173.2.1179
- Levin, A., Reznichenko, A., Witasp, A., Liu, P., Greasley, P. J., Sorrentino, A., et al. (2020). Novel Insights into the Disease Transcriptome of Human Diabetic Glomeruli and Tubulointerstitium. *Nephrol. Dial. Transpl.* 35, 2059–2072. doi:10.1093/ndt/gfaa121
- Li, X.-y., Wang, S.-s., Han, Z., Han, F., Chang, Y.-p., Yang, Y., et al. (2017). Triptolide Restores Autophagy to Alleviate Diabetic Renal Fibrosis through the miR-141-3p/PTEN/Akt/mTOR Pathway. *Mol. Ther. - Nucleic Acids* 9, 48–56. doi:10.1016/j.omtn.2017.08.011
- Li, X., Zhang, X., Xia, J., Zhang, L., Chen, B., Lian, G., et al. (2021). Macrophage HIF-2α Suppresses NLRP3 Inflammasome Activation and Alleviates Insulin Resistance. *Cell Rep.* 36, 109607. doi:10.1016/j.celrep.2021.109607
- Li, Y., and Chen, Y. (2019). AMPK and Autophagy. Adv. Exp. Med. Biol. 1206, 85–108. doi:10.1007/978-981-15-0602-4_4
- Li, Z. L., Lv, L. L., Wang, B., Tang, T. T., Feng, Y., Cao, J. Y., et al. (2019). The Profibrotic Effects of MK-8617 on Tubulointerstitial Fibrosis Mediated by the KLF5 Regulating Pathway. *FASEB J.* 33, 12630–12643. doi:10.1096/fj. 201901087RR
- Liao, K., Lv, D.-Y., Yu, H.-L., Chen, H., and Luo, S.-X. (2021). iNOS Regulates Activation of the NLRP3 Inflammasome through the sGC/cGMP/PKG/TACE/ TNF-α axis in Response to Cigarette Smoke Resulting in Aortic Endothelial Pyroptosis and Vascular Dysfunction. *Int. Immunopharmacol.* 101, 108334. doi:10.1016/j.intimp.2021.108334
- Lin, M., and Tang, S. C. W. (2014). Toll-like Receptors: Sensing and Reacting to Diabetic Injury in the Kidney. *Nephrol. Dial. Transplant.* 29, 746–754. doi:10. 1093/ndt/gft446
- Liu, J.-Y., Lee, K.-F., Sze, C.-W., Tong, Y., Tang, S. C.-W., Ng, T.-B., et al. (2013). Intestinal Absorption and Bioavailability of Traditional Chinese Medicines: A Review of Recent Experimental Progress and Implication for Quality Control. J. Pharm. Pharmacol. 65, 621–633. doi:10.1111/j.2042-7158.2012.01608.x
- Liu, M., and Zen, K. (2021). Toll-Like Receptors Regulate the Development and Progression of Renal Diseases. *Kidney Dis.* 7, 14–23. doi:10.1159/000511947
- Liu, P., Li, F. A., Qiu, M., and He, L. (2014). Expression and Cellular Distribution of TLR4, MyD88, and NF-κB in Diabetic Renal Tubulointerstitial Fibrosis, *In Vitro* and *In Vivo. Diabetes Res. Clin. Pract.* 105, 206–216. doi:10.1016/j.diabres. 2014.04.020
- Liu, R., Zhong, Y., Li, X., Chen, H., Jim, B., Zhou, M.-M., et al. (2014). Role of Transcription Factor Acetylation in Diabetic Kidney Disease. *Diabetes* 63, 2440–2453. doi:10.2337/db13-1810
- Lu, M., Yin, N., Liu, W., Cui, X., Chen, S., and Wang, E. (2017). Curcumin Ameliorates Diabetic Nephropathy by Suppressing NLRP3 Inflammasome Signaling. *BioMed Res. Int.* 2017, 1–10. doi:10.1155/2017/1516985
- Lu, Q., Ji, X.-J., Zhou, Y.-X., Yao, X.-Q., Liu, Y.-Q., Zhang, F., et al. (2015). Quercetin Inhibits the mTORC1/p70S6K Signaling-Mediated Renal Tubular Epithelial-Mesenchymal Transition and Renal Fibrosis in Diabetic Nephropathy. *Pharmacol. Res.* 99, 237–247. doi:10.1016/j.phrs.2015.06.006
- Lu, S., Zhang, H., Wei, X., Huang, X., Chen, L., Jiang, L., et al. (2019). 2-dodecyl-6methoxycyclohexa-2,5-diene-1,4-dione Isolated from Averrhoa Carambola L.
 Root Ameliorates Diabetic Nephropathy by Inhibiting the TLR4/MyD88/NFκB Pathway. *Dmso* Vol. 12, 1355–1363. doi:10.2147/DMSO.S209436
- Luan, P., Zhuang, J., Zou, J., Li, H., Shuai, P., Xu, X., et al. (2018). NLRC5 Deficiency Ameliorates Diabetic Nephropathy through Alleviating Inflammation. *FASEB J.* 32, 1070–1084. doi:10.1096/fj.201700511RR

- Lyons, C., and Roche, H. (2018). Nutritional Modulation of AMPK-Impact upon Metabolic-Inflammation. *Ijms* 19, 3092. doi:10.3390/ijms19103092
- Ma, T., Tian, X., Zhang, B., Li, M., Wang, Y., Yang, C., et al. (2022). Low-dose Metformin Targets the Lysosomal AMPK Pathway through PEN2. *Nature* 603, 159–165. doi:10.1038/s41586-022-04431-8
- Malik, A. N., Shahni, R., and Iqbal, M. M. (2009). Increased Peripheral Blood Mitochondrial DNA in Type 2 Diabetic Patients with Nephropathy. *Diabetes Res. Clin. Pract.* 86, e22–e24. doi:10.1016/j.diabres.2009.07.002
- Marquez-Curtis, L. A., and Janowska-Wieczorek, A. (2013). Enhancing the Migration Ability of Mesenchymal Stromal Cells by Targeting the SDF-1/ CXCR4 axis. *BioMed Res. Int.* 2013, 1–15. doi:10.1155/2013/561098
- Martínez-Martínez, E., Buonafine, M., Boukhalfa, I., Ibarrola, J., Fernández-Celis, A., Kolkhof, P., et al. (2017). Aldosterone Target NGAL (Neutrophil Gelatinase-Associated Lipocalin) Is Involved in Cardiac Remodeling after Myocardial Infarction through NFκB Pathway. *Hypertension* 70, 1148–1156. doi:10.1161/ HYPERTENSIONAHA.117.09791
- Martinon, F., Burns, K., and Tschopp, J. (2002). The Inflammasome. *Mol. Cell* 10, 417–426. doi:10.1016/s1097-2765(02)00599-3
- Masood, H., Che, R., and Zhang, A. (2015). Inflammasomes in the Pathophysiology of Kidney Diseases. *Kidney Dis.* 1, 187–193. doi:10.1159/000438843
- McGettrick, A. F., and O'Neill, L. A. J. (2020). The Role of HIF in Immunity and Inflammation. *Cell Metab.* 32, 524–536. doi:10.1016/j.cmet.2020.08.002
- McGuire, D. K., Alexander, J. H., Johansen, O. E., Perkovic, V., Rosenstock, J., Cooper, M. E., et al. (2019). Linagliptin Effects on Heart Failure and Related Outcomes in Individuals with Type 2 Diabetes Mellitus at High Cardiovascular and Renal Risk in CARMELINA. *Circulation* 139, 351–361. doi:10.1161/ CIRCULATIONAHA.118.038352
- Mezzano, S., Aros, C., Droguett, A., Burgos, M. E., Ardiles, L., Flores, C., et al. (2004). NF- B Activation and Overexpression of Regulated Genes in Human Diabetic Nephropathy. *Nephrol. Dial. Transplant.* 19, 2505–2512. doi:10.1093/ ndt/gfh207
- Moir, S., Hanekom, L., Fang, Z. Y., Haluska, B., Wong, C., et al. (2006). Relationship between Myocardial Perfusion and Dysfunction in Diabetic Cardiomyopathy: A Study of Quantitative Contrast Echocardiography and Strain Rate Imaging. *Heart* 92, 1414–1419. doi:10.1136/hrt.2005. 079350
- Mottl, A. K., Alicic, R., Argyropoulos, C., Brosius, F. C., Mauer, M., Molitch, M., et al. (2022). KDOQI US Commentary on the KDIGO 2020 Clinical Practice Guideline for Diabetes Management in CKD. Am. J. Kidney Dis. 79, 457–479. doi:10.1053/j.ajkd.2021.09.010
- Nakahira, K., Haspel, J. A., Rathinam, V. A. K., Lee, S.-J., Dolinay, T., Lam, H. C., et al. (2011). Autophagy Proteins Regulate Innate Immune Responses by Inhibiting the Release of Mitochondrial DNA Mediated by the NALP3 Inflammasome. *Nat. Immunol.* 12, 222–230. doi:10.1038/ni.1980
- Nathan, C., and Ding, A. (2010). Nonresolving Inflammation. Cell 140, 871–882. doi:10.1016/j.cell.2010.02.029
- Navarro-González, J. F., Mora-Fernández, C., de Fuentes, M. M., and García-Pérez, J. (2011). Inflammatory Molecules and Pathways in the Pathogenesis of Diabetic Nephropathy. *Nat. Rev. Nephrol.* 7, 327–340. doi:10.1038/nrneph. 2011.51
- Packer, M. (2020). Role of Impaired Nutrient and Oxygen Deprivation Signaling and Deficient Autophagic Flux in Diabetic CKD Development: Implications for Understanding the Effects of Sodium-Glucose Cotransporter 2-Inhibitors. J. Am. Soc. Nephrol. 31, 907–919. doi:10. 1681/ASN.2020010010
- Palazon, A., Goldrath, A. W., Nizet, V., and Johnson, R. S. (2014). HIF Transcription Factors, Inflammation, and Immunity. *Immunity* 41, 518–528. doi:10.1016/j.immuni.2014.09.008
- Patel, V., Joharapurkar, A., and Jain, M. (2021). Role of Mineralocorticoid Receptor Antagonists in Kidney Diseases. *Drug Dev. Res.* 82, 341–363. doi:10.1002/ddr. 21760
- Peng, J., Li, X., Zhang, D., Chen, J.-K., Su, Y., Smith, S. B., et al. (2015). Hyperglycemia, P53, and Mitochondrial Pathway of Apoptosis Are Involved in the Susceptibility of Diabetic Models to Ischemic Acute Kidney Injury. *Kidney Int.* 87, 137–150. doi:10.1038/ki.2014.226
- Pérez-Morales, R. E., del Pino, M. D., Valdivielso, J. M., Ortiz, A., Mora-Fernández, C., and Navarro-González, J. F. (2019). Inflammation in Diabetic Kidney Disease. Nephron 143, 12–16. doi:10.1159/000493278

- Perkovic, V., Jardine, M. J., Neal, B., Bompoint, S., Heerspink, H. J. L., Charytan, D. M., et al. (2019). Canagliflozin and Renal Outcomes in Type 2 Diabetes and Nephropathy. N. Engl. J. Med. 380, 2295–2306. doi:10.1056/NEJMoa1811744
- Pichler, R., Afkarian, M., Dieter, B. P., and Tuttle, K. R. (2017). Immunity and Inflammation in Diabetic Kidney Disease: Translating Mechanisms to Biomarkers and Treatment Targets. Am. J. Physiology-Renal Physiology 312, F716–F731. doi:10.1152/ajprenal.00314.2016
- Pilon, G., Dallaire, P., and Marette, A. (2004). Inhibition of Inducible Nitric-Oxide Synthase by Activators of AMP-Activated Protein Kinase. J. Biol. Chem. 279, 20767–20774. doi:10.1074/jbc.M401390200
- Quagliariello, V., De Laurentiis, M., Cocco, S., Rea, G., Bonelli, A., Caronna, A., et al. (2020). NLRP3 as Putative Marker of Ipilimumab-Induced Cardiotoxicity in the Presence of Hyperglycemia in Estrogen-Responsive and Triple-Negative Breast Cancer Cells. *Ijms* 21, 7802. doi:10.3390/ijms21207802
- Rathinam, V. A. K., and Chan, F. K.-M. (2018). Inflammasome, Inflammation, and Tissue Homeostasis. *Trends Mol. Med.* 24, 304–318. doi:10.1016/j.molmed. 2018.01.004
- Riley, J. S., and Tait, S. W. (2020). Mitochondrial DNA in Inflammation and Immunity. *EMBO Rep.* 21, e49799. doi:10.15252/embr.201949799
- Rius, J., Guma, M., Schachtrup, C., Akassoglou, K., Zinkernagel, A. S., Nizet, V., et al. (2008). NF-κB Links Innate Immunity to the Hypoxic Response through Transcriptional Regulation of HIF-1α. *Nature* 453, 807–811. doi:10.1038/ nature06905
- Rodríguez-Nuevo, A., Díaz-Ramos, A., Noguera, E., Díaz-Sáez, F., Duran, X., Muñoz, J. P., et al. (2018). Mitochondrial DNA and TLR9 Drive Muscle Inflammation upon Opa1 Deficiency. *Embo J.* 37. doi:10.15252/embj. 201796553
- Rohm, T. V., Meier, D. T., Olefsky, J. M., and Donath, M. Y. (2022). Inflammation in Obesity, Diabetes, and Related Disorders. *Immunity* 55, 31–55. doi:10.1016/j. immuni.2021.12.013
- Rosso, F., Quagliariello, V., Tortora, C., Di Lazzaro, A., Barbarisi, A., and Iaffaioli, R. V. (2013). Cross-linked Hyaluronic Acid Sub-micron Particles: *In Vitro* and *In Vivo* Biodistribution Study in Cancer Xenograft Model. *J. Mater Sci. Mater Med.* 24, 1473–1481. doi:10.1007/s10856-013-4895-4
- Sancar-Bas, S., Gezginci-Oktayoglu, S., and Bolkent, S. (2015). Exendin-4 Attenuates Renal Tubular Injury by Decreasing Oxidative Stress and Inflammation in Streptozotocin-Induced Diabetic Mice. *Growth factors*. 33, 419–429. doi:10.3109/08977194.2015.1125349
- Saraheimo, M., Teppo, A.-M., Forsblom, C., Fagerudd, J., and Groop, P.-H. (2003). Diabetic Nephropathy Is Associated with Low-Grade Inflammation in Type 1 Diabetic Patients. *Diabetologia* 46, 1402–1407. doi:10.1007/s00125-003-1194-5
- Sávio-Silva, C., Beyerstedt, S., Soinski-Sousa, P. E., Casaro, E. B., Balby-Rocha, M. T. A., Simplício-Filho, A., et al. (2020). Mesenchymal Stem Cell Therapy for Diabetic Kidney Disease: A Review of the Studies Using Syngeneic, Autologous, Allogeneic, and Xenogeneic Cells. *Stem Cells Int.* 2020, 1–28. doi:10.1155/2020/ 8833725
- Schmid, H., Boucherot, A., Yasuda, Y., Henger, A., Brunner, B., Eichinger, F., et al. (2006). Modular Activation of Nuclear Factor-κB Transcriptional Programs in Human Diabetic Nephropathy. *Diabetes* 55, 2993–3003. doi:10.2337/db06-0477
- Schroder, K., and Tschopp, J. (2010). The Inflammasomes. Cell 140, 821-832. doi:10.1016/j.cell.2010.01.040
- Seabright, A. P., Fine, N. H. F., Barlow, J. P., Lord, S. O., Musa, I., Gray, A., et al. (2020). AMPK Activation Induces Mitophagy and Promotes Mitochondrial Fission while Activating TBK1 in a PINK1-Parkin Independent Manner. FASEB J. 34, 6284–6301. doi:10.1096/fj.201903051R
- Shah, Z., Kampfrath, T., Deiuliis, J. A., Zhong, J., Pineda, C., Ying, Z., et al. (2011). Long-term Dipeptidyl-Peptidase 4 Inhibition Reduces Atherosclerosis and Inflammation via Effects on Monocyte Recruitment and Chemotaxis. *Circulation* 124, 2338–2349. doi:10.1161/CIRCULATIONAHA.111.041418
- Shahzad, K., Bock, F., Dong, W., Wang, H., Kopf, S., Kohli, S., et al. (2015). Nlrp3inflammasome Activation in Non-myeloid-derived Cells Aggravates Diabetic Nephropathy. *Kidney Int.* 87, 74–84. doi:10.1038/ki.2014.271
- Shaman, A. M., Bain, S. C., Bakris, G. L., Buse, J. B., Idorn, T., Mahaffey, K. W., et al. (2022). Effect of the Glucagon-like Peptide-1 Receptor Agonists Semaglutide and Liraglutide on Kidney Outcomes in Patients with Type 2 Diabetes: Pooled Analysis of SUSTAIN 6 and LEADER. *Circulation* 145, 575–585. doi:10.1161/ CIRCULATIONAHA.121.055459

- Shao, Y., Lv, C., Wu, C., Zhou, Y., and Wang, Q. (2016). Mir-217 Promotes Inflammation and Fibrosis in High Glucose Cultured Rat Glomerular Mesangial Cells via Sirt1/HIF-1α Signaling Pathway. *Diabetes Metab. Res. Rev.* 32, 534–543. doi:10.1002/dmrr.2788
- Shen, A.-R., Zhong, X., Tang, T.-T., Wang, C., Jing, J., Liu, B.-C., et al. (2020). Integrin, Exosome and Kidney Disease. *Front. Physiol.* 11, 627800. doi:10.3389/ fphys.2020.627800
- Shimada, K., Crother, T. R., Karlin, J., Dagvadorj, J., Chiba, N., Chen, S., et al. (2012). Oxidized Mitochondrial DNA Activates the NLRP3 Inflammasome during Apoptosis. *Immunity* 36, 401–414. doi:10.1016/j.immuni.2012.01.009
- Soares, J. L. S., Fernandes, F. P., Patente, T. A., Monteiro, M. B., Parisi, M. C., Giannella-Neto, D., et al. (2018). Gain-of-function Variants in NLRP1 Protect against the Development of Diabetic Kidney Disease: NLRP1 Inflammasome Role in Metabolic Stress Sensing? *Clin. Immunol.* 187, 46–49. doi:10.1016/j. clim.2017.10.003
- Stein, S., Lohmann, C., Schäfer, N., Hofmann, J., Rohrer, L., Besler, C., et al. (2010). SIRT1 Decreases Lox-1-Mediated Foam Cell Formation in Atherogenesis. *Eur. Heart J.* 31, 2301–2309. doi:10.1093/eurheartj/ehq107
- Stowe, I., Lee, B., and Kayagaki, N. (2015). Caspase-11: Arming the Guards against Bacterial Infection. *Immunol. Rev.* 265, 75–84. doi:10.1111/imr.12292
- Strowig, T., Henao-Mejia, J., Elinav, E., and Flavell, R. (2012). Inflammasomes in Health and Disease. *Nature* 481, 278–286. doi:10.1038/nature10759
- Sun, H.-J., Xiong, S.-P., Cao, X., Cao, L., Zhu, M.-Y., Wu, Z.-Y., et al. (2021). Polysulfide-mediated Sulfhydration of SIRT1 Prevents Diabetic Nephropathy by Suppressing Phosphorylation and Acetylation of P65 NF-κB and STAT3. *Redox Biol.* 38, 101813. doi:10.1016/j.redox.2020.101813
- Sun, H., Saeedi, P., Karuranga, S., Pinkepank, M., Ogurtsova, K., Duncan, B. B., et al. (2022). IDF Diabetes Atlas: Global, Regional and Country-Level Diabetes Prevalence Estimates for 2021 and Projections for 2045. *Diabetes Res. Clin. Pract.* 183, 109119. doi:10.1016/j.diabres.2021.109119
- Sun, S.-C. (2011). Non-canonical NF-κB Signaling Pathway. Cell Res. 21, 71–85. doi:10.1038/cr.2010.177
- Sun, S.-C. (2017). The Non-canonical NF-κB Pathway in Immunity and Inflammation. Nat. Rev. Immunol. 17, 545–558. doi:10.1038/nri.2017.52
- Sur, S., Nguyen, M., Boada, P., Sigdel, T. K., Sollinger, H., and Sarwal, M. M. (2021). FcER1: A Novel Molecule Implicated in the Progression of Human Diabetic Kidney Disease. *Front. Immunol.* 12, 769972. doi:10.3389/fimmu. 2021.769972
- Takiyama, Y., Harumi, T., Watanabe, J., Fujita, Y., Honjo, J., Shimizu, N., et al. (2011). Tubular Injury in a Rat Model of Type 2 Diabetes Is Prevented by Metformin. *Diabetes* 60, 981–992. doi:10.2337/db10-0655
- Tang, S. C. W., and Yiu, W. H. (2020). Innate Immunity in Diabetic Kidney Disease. Nat. Rev. Nephrol. 16, 206–222. doi:10.1038/s41581-019-0234-4
- Tang, T.-T., Lv, L.-L., Lan, H.-Y., and Liu, B.-C. (2019). Extracellular Vesicles: Opportunities and Challenges for the Treatment of Renal Diseases. *Front. Physiol.* 10, 226. doi:10.3389/fphys.2019.00226
- Tannahill, G. M., Curtis, A. M., Adamik, J., Palsson-McDermott, E. M., McGettrick, A. F., Goel, G., et al. (2013). Succinate Is an Inflammatory Signal that Induces IL-1 β through HIF-1 α . *Nature* 496, 238–242. doi:10. 1038/nature11986
- Thomas, M. C., Brownlee, M., Susztak, K., Sharma, K., Jandeleit-Dahm, K. A. M., Zoungas, S., et al. (2015). Diabetic Kidney Disease. *Nat. Rev. Dis. Prim.* 1, 15018. doi:10.1038/nrdp.2015.18
- Tremblay, A. J., Lamarche, B., Deacon, C. F., Weisnagel, S. J., and Couture, P. (2014). Effects of Sitagliptin Therapy on Markers of Low-Grade Inflammation and Cell Adhesion Molecules in Patients with Type 2 Diabetes. *Metabolism* 63, 1141–1148. doi:10.1016/j.metabol.2014.06.004
- Tung, C.-W., Hsu, Y.-C., Shih, Y.-H., Chang, P.-J., and Lin, C.-L. (2018). Glomerular Mesangial Cell and Podocyte Injuries in Diabetic Nephropathy. *Nephrology* 23 (Suppl. 4), 32–37. doi:10.1111/nep.13451
- Uzu, T., Yokoyama, H., Itoh, H., Koya, D., Nakagawa, A., Nishizawa, M., et al. (2011). Elevated Serum Levels of Interleukin-18 in Patients with Overt Diabetic Nephropathy: Effects of Miglitol. *Clin. Exp. Nephrol.* 15, 58–63. doi:10.1007/ s10157-010-0343-7
- Valiño-Rivas, L., Cuarental, L., Nuñez, G., Sanz, A. B., Ortiz, A., and Sanchez-Niño, M. D. (2020). Loss of NLRP6 Expression Increases the Severity of Acute Kidney Injury. *Nephrol. Dial. Transpl.* 35, 587–598. doi:10.1093/ndt/ gfz169

- Van Opdenbosch, N., and Lamkanfi, M. (2019). Caspases in Cell Death, Inflammation, and Disease. *Immunity* 50, 1352–1364. doi:10.1016/j.immuni. 2019.05.020
- Wada, T., Inagaki, M., Yoshinari, T., Terata, R., Totsuka, N., Gotou, M., et al. (2021). Apararenone in Patients with Diabetic Nephropathy: Results of a Randomized, Double-Blind, Placebo-Controlled Phase 2 Dose-Response Study and Open-Label Extension Study. *Clin. Exp. Nephrol.* 25, 120–130. doi:10.1007/s10157-020-01963-z
- Wang, G., Li, Q., Chen, D., Wu, B., Wu, Y., Tong, W., et al. (2019). Kidney-targeted Rhein-Loaded Liponanoparticles for Diabetic Nephropathy Therapy via Size Control and Enhancement of Renal Cellular Uptake. *Theranostics* 9, 6191–6208. doi:10.7150/thno.37538
- Wang, Q., Liu, S., Zhai, A., Zhang, B., and Tian, G. (2018). AMPK-mediated Regulation of Lipid Metabolism by Phosphorylation. *Biol. Pharm. Bull.* 41, 985–993. doi:10.1248/bpb.b17-00724
- Wang, S., Li, Y., Fan, J., Zhang, X., Luan, J., Bian, Q., et al. (2017). Interleukin-22 Ameliorated Renal Injury and Fibrosis in Diabetic Nephropathy through Inhibition of NLRP3 Inflammasome Activation. *Cell Death Dis.* 8, e2937. doi:10.1038/cddis.2017.292
- Wang, Z., Wang, N., Liu, P., and Xie, X. (2016). AMPK and Cancer. Exp. Suppl. 107, 203–226. doi:10.1007/978-3-319-43589-3_9
- Wang, Z., Zhu, Q., Li, P.-L., Dhaduk, R., Zhang, F., Gehr, T. W., et al. (2014). Silencing of Hypoxia-Inducible Factor-1α Gene Attenuates Chronic Ischemic Renal Injury in Two-Kidney, One-Clip Rats. Am. J. Physiology-Renal Physiology 306, F1236–F1242. doi:10.1152/ajprenal.00673.2013
- Warner, N., and Núñez, G. (2013). MyD88: A Critical Adaptor Protein in Innate Immunity Signal Transduction. J. I. 190, 3–4. doi:10.4049/jimmunol.1203103
- Watts, E. R., and Walmsley, S. R. (2019). Inflammation and Hypoxia: HIF and PHD Isoform Selectivity. *Trends Mol. Med.* 25, 33–46. doi:10.1016/j.molmed. 2018.10.006
- Wei, X., Shao, B., He, Z., Ye, T., Luo, M., Sang, Y., et al. (2015). Cationic Nanocarriers Induce Cell Necrosis through Impairment of Na+/K+-ATPase and Cause Subsequent Inflammatory Response. *Cell Res.* 25, 237–253. doi:10. 1038/cr.2015.9
- Wen, Y., Yan, M., Zhang, B., and Li, P. (2017). Chinese Medicine for Diabetic Kidney Disease in China. Nephrology 22 (Suppl. 4), 50–55. doi:10.1111/nep. 13149
- Whittaker, A., Kragh, Å. M., Hartleib-Geschwindner, J., Albayaty, M., Backlund, A., Greasley, P. J., et al. (2020). Safety, Tolerability, and Pharmacokinetics of the Mineralocorticoid Receptor Modulator AZD9977 in Healthy Men: A Phase I Multiple Ascending Dose Study. *Clin. Transl. Sci.* 13, 275–283. doi:10.1111/cts. 12705
- Wilson, P. C., Wu, H., Kirita, Y., Uchimura, K., Ledru, N., Rennke, H. G., et al. (2019). The Single-Cell Transcriptomic Landscape of Early Human Diabetic Nephropathy. Proc. Natl. Acad. Sci. U.S.A. 116, 19619–19625. doi:10.1073/pnas. 1908706116
- Wu, J., Sun, L., Chen, X., Du, F., Shi, H., Chen, C., et al. (2013). Cyclic GMP-AMP Is an Endogenous Second Messenger in Innate Immune Signaling by Cytosolic DNA. *Science* 339, 826–830. doi:10.1126/science.1229963
- Wu, M., Han, W., Song, S., Du, Y., Liu, C., Chen, N., et al. (2018). NLRP3 Deficiency Ameliorates Renal Inflammation and Fibrosis in Diabetic Mice. *Mol. Cell. Endocrinol.* 478, 115–125. doi:10.1016/j.mce.2018.08.002
- Wu, M., Yang, Z., Zhang, C., Shi, Y., Han, W., Song, S., et al. (2021). Inhibition of NLRP3 Inflammasome Ameliorates Podocyte Damage by Suppressing Lipid Accumulation in Diabetic Nephropathy. *Metabolism* 118, 154748. doi:10.1016/ j.metabol.2021.154748
- Wu, W., Yang, J.-J., Yang, H.-M., Huang, M.-M., Fang, Q.-J., Shi, G., et al. (2017).
 Multi-glycoside of Tripterygium Wilfordii Hook. F. Attenuates Glomerulosclerosis in a Rat Model of Diabetic Nephropathy by Exerting Anti-microinflammatory Effects without Affecting Hyperglycemia. *Int. J. Mol. Med.* 40, 721–730. doi:10.3892/ijmm.2017.3068
- Xian, H., Liu, Y., Rundberg Nilsson, A., Gatchalian, R., Crother, T. R., Tourtellotte,
 W. G., et al. (2021). Metformin Inhibition of Mitochondrial ATP and DNA
 Synthesis Abrogates NLRP3 Inflammasome Activation and Pulmonary
 Inflammation. *Immunity* 54, 1463–1477. doi:10.1016/j.immuni.2021.05.004
- Xiao, Y., Liu, Y., Yu, K., Zhou, L., Bi, J., Cheng, J., et al. (2013). The Effect of Chinese Herbal Medicine on Albuminuria Levels in Patients with Diabetic

Nephropathy: A Systematic Review and Meta-Analysis. Evidence-Based Complementary Altern. Med. 2013, 1–11. doi:10.1155/2013/937549

- Xie, C., Wu, W., Tang, A., Luo, N., and Tan, Y. (2019). IncRNA GAS5/miR-452-5p Reduces Oxidative Stress and Pyroptosis of High-Glucose-Stimulated Renal Tubular Cells. Dmso 12, 2609–2617. doi:10.2147/DMSO.S228654
- Xiong, W., Meng, X.-F., and Zhang, C. (2020). Inflammasome Activation in Podocytes: A New Mechanism of Glomerular Diseases. *Inflamm. Res.* 69, 731–743. doi:10.1007/s00011-020-01354-w
- Xiong, Y., and Zhou, L. (20192019). The Signaling of Cellular Senescence in Diabetic Nephropathy. Oxidative Med. Cell. Longev. 2019, 1–16. doi:10.1155/ 2019/7495629
- Xu, L., Zhou, J., Che, J., Wang, H., Yang, W., Zhou, W., et al. (2021). Mitochondrial DNA Enables AIM2 Inflammasome Activation and Hepatocyte Pyroptosis in Nonalcoholic Fatty Liver Disease. Am. J. Physiology-Gastrointestinal Liver Physiology 320, G1034–G1044. doi:10.1152/ajpgi.00431.2020
- Xu, Y., Tao, X., Shen, B., Horng, T., Medzhitov, R., Manley, J. L., et al. (2000). Structural Basis for Signal Transduction by the Toll/interleukin-1 Receptor Domains. *Nature* 408, 111–115. doi:10.1038/35040600
- Yang, M., Luo, S., Jiang, N., Wang, X., Han, Y., Zhao, H., et al. (2021a). DsbA-L Ameliorates Renal Injury through the AMPK/NLRP3 Inflammasome Signaling Pathway in Diabetic Nephropathy. *Front. Physiol.* 12, 659751. doi:10.3389/ fphys.2021.659751
- Yang, M., Wang, X., Han, Y., Li, C., Wei, L., Yang, J., et al. (2021b). Targeting the NLRP3 Inflammasome in Diabetic Nephropathy. *Cmc* 28, 8810–8824. doi:10. 2174/0929867328666210705153109
- Yi, H., Peng, R., Zhang, L.-y., Sun, Y., Peng, H.-m., Liu, H.-d., et al. (2017). LincRNA-Gm4419 Knockdown Ameliorates NF-κB/NLRP3 Inflammasome-Mediated Inflammation in Diabetic Nephropathy. *Cell Death Dis.* 8, e2583. doi:10.1038/cddis.2016.451
- Youm, Y.-H., Nguyen, K. Y., Grant, R. W., Goldberg, E. L., Bodogai, M., Kim, D., et al. (2015). The Ketone Metabolite β-hydroxybutyrate Blocks NLRP3 Inflammasome-Mediated Inflammatory Disease. *Nat. Med.* 21, 263–269. doi:10.1038/nm.3804
- Young, M. J., and Rickard, A. J. (2015). Mineralocorticoid Receptors in the Heart: Lessons from Cell-Selective Transgenic Animals. J. Endocrinol. 224, R1–R13. doi:10.1530/JOE-14-0471
- Yu, L., Wang, Y., Guo, Y. H., Wang, L., Yang, Z., Zhai, Z. H., et al. (2021). HIF-1α Alleviates High-Glucose-Induced Renal Tubular Cell Injury by Promoting Parkin/PINK1-Mediated Mitophagy. *Front. Med.* 8, 803874. doi:10.3389/ fmed.2021.803874
- Yuan, F., Kolb, R., Pandey, G., Li, W., Sun, L., Liu, F., et al. (2016). Involvement of the NLRC4-Inflammasome in Diabetic Nephropathy. *PLoS One* 11, e0164135. doi:10.1371/journal.pone.0164135
- Zhang, C., Zhu, X., Li, L., Ma, T., Shi, M., Yang, Y., et al. (2019). A Small Molecule Inhibitor MCC950 Ameliorates Kidney Injury in Diabetic Nephropathy by Inhibiting NLRP3 Inflammasome Activation. *Dmso* 12, 1297–1309. doi:10. 2147/DMSO.S199802
- Zhang, J., Huang, L., Shi, X., Yang, L., Hua, F., Ma, J., et al. (2020). Metformin Protects against Myocardial Ischemia-Reperfusion Injury and Cell Pyroptosis via AMPK/NLRP3 Inflammasome Pathway. *Aging* 12, 24270-24287. doi:10. 18632/aging.202143
- Zhang, J.-Z., Liu, Z., Liu, J., Ren, J.-X., and Sun, T.-S. (2014). Mitochondrial DNA Induces Inflammation and Increases TLR9/NF-κB Expression in Lung Tissue. *Int. J. Mol. Med.* 33, 817–824. doi:10.3892/ijmm.2014.1650
- Zhang, J., Yang, X., Zhang, X., Lu, D., and Guo, R. (2021). Electro-Acupuncture Protects Diabetic Nephropathy-Induced Inflammation through Suppression of NLRP3 Inflammasome in Renal Macrophage Isolation. *Emiddt* 21, 2075–2083. doi:10.2174/1871530321666210118161721
- Zhang, L., Zhao, M.-H., Zuo, L., Wang, Y., Yu, F., Zhang, H., et al. (2019). China Kidney Disease Network (CK-NET) 2015 Annual Data Report. *Kidney Int. Suppl.* 9, e1-e81. doi:10.1016/j.kisu.2018.11.001
- Zhang, L., Li, M., Wang, Z., Sun, P., Wei, S., Zhang, C., et al. (2021). Cardiovascular Risk after SARS-CoV-2 Infection Is Mediated by IL18/IL18R1/HIF-1 Signaling Pathway axis. *Front. Immunol.* 12, 780804. doi:10.3389/fimmu.2021.780804
- Zhang, M., Chen, Y., Yang, M. j., Fan, X. R., Xie, H., Zhang, L., et al. (2019). Celastrol Attenuates Renal Injury in Diabetic Rats via MAPK/NF-κB Pathway. *Phytotherapy Res.* 33, 1191–1198. doi:10.1002/ptr.6314

- Zhang, Q.-y., Xu, S.-j., Qian, J.-c., Yang, L.-b., Chen, P.-q., Wang, Y., et al. (2022). Pharmacological Inhibition of MyD88 Suppresses Inflammation in Tubular Epithelial Cells and Prevents Diabetic Nephropathy in Experimental Mice. Acta Pharmacol. Sin. 43, 354–366. doi:10.1038/s41401-021-00766-6
- Zhang, Q., Lenardo, M. J., and Baltimore, D. (2017). 30 Years of NF-κB: A Blossoming of Relevance to Human Pathobiology. *Cell* 168, 37–57. doi:10.1016/ j.cell.2016.12.012
- Zhang, Q., Liu, X., Sullivan, M. A., Shi, C., and Deng, B. (2021a). Protective Effect of Yi Shen Pai Du Formula against Diabetic Kidney Injury via Inhibition of Oxidative Stress, Inflammation, and Epithelial-To-Mesenchymal Transition in Db/db Mice. Oxidative Med. Cell. Longev. 2021, 1–15. doi:10.1155/2021/7958021
- Zhang, Q., Raoof, M., Chen, Y., Sumi, Y., Sursal, T., Junger, W., et al. (2010). Circulating Mitochondrial DAMPs Cause Inflammatory Responses to Injury. *Nature* 464, 104–107. doi:10.1038/nature08780
- Zhang, Q., Song, W., Zhao, B., Xie, J., Sun, Q., Shi, X., et al. (2021b). Quercetin Attenuates Diabetic Peripheral Neuropathy by Correcting Mitochondrial Abnormality via Activation of AMPK/PGC-1a Pathway In Vivo and In Vitro. Front. Neurosci. 15, 636172. doi:10.3389/fnins.2021.636172
- Zhang, S., Xu, H., Yu, X., Wu, Y., and Sui, D. (2017). Metformin Ameliorates Diabetic Nephropathy in a Rat Model of Low-Dose Streptozotocin-Induced Diabetes. *Exp. Ther. Med.* 14, 383–390. doi:10.3892/etm.2017.4475
- Zhang, W., Cai, Y., Xu, W., Yin, Z., Gao, X., and Xiong, S. (2013). AIM2 Facilitates the Apoptotic DNA-Induced Systemic Lupus Erythematosus via Arbitrating Macrophage Functional Maturation. J. Clin. Immunol. 33, 925–937. doi:10. 1007/s10875-013-9881-6
- Zhang, W., Huai, Y., Miao, Z., Qian, A., and Wang, Y. (2019). Systems Pharmacology for Investigation of the Mechanisms of Action of Traditional Chinese Medicine in Drug Discovery. *Front. Pharmacol.* 10, 743. doi:10.3389/ fphar.2019.00743
- Zhang, X.-X., Kong, J., and Yun, K. (2020). Prevalence of Diabetic Nephropathy Among Patients with Type 2 Diabetes Mellitus in China: A Meta-Analysis of Observational Studies. J. Diabetes Res. 2020, 1–11. doi:10.1155/2020/2315607
- Zhao, X., Chen, X., Chima, A., Zhang, Y., George, J., Cobbs, A., et al. (2019). Albumin Induces CD44 Expression in Glomerular Parietal Epithelial Cells by Activating Extracellular Signal-regulated Kinase 1/2 Pathway. J. Cell. Physiology 234, 7224–7235. doi:10.1002/jcp.27477

- Zhen, J., Zhang, L., Pan, J., Ma, S., Yu, X., Li, X., et al. (2014). AIM2 Mediates Inflammation-Associated Renal Damage in Hepatitis B Virus-Associated Glomerulonephritis by Regulating Caspase-1, IL-1β, and IL-18. *Mediat. Inflamm.* 2014, 1–9. doi:10.1155/2014/190860
- Zhong, Y., Lee, K., Deng, Y., Ma, Y., Chen, Y., Li, X., et al. (2019). Arctigenin Attenuates Diabetic Kidney Disease through the Activation of PP2A in Podocytes. *Nat. Commun.* 10, 4523. doi:10.1038/s41467-019-12433-w
- Zhong, Z., Liang, S., Sanchez-Lopez, E., He, F., Shalapour, S., Lin, X.-j., et al. (2018). New Mitochondrial DNA Synthesis Enables NLRP3 Inflammasome Activation. *Nature* 560, 198–203. doi:10.1038/s41586-018-0372-z
- Zhu, Q. J., Zhu, M., Xu, X. X., Meng, X. M., and Wu, Y. G. (2019). Exosomes from High Glucose-Treated Macrophages Activate Glomerular Mesangial Cells via TGF-β1/Smad3 Pathway *In Vivo* and *In Vitro. FASEB J.* 33, 9279–9290. doi:10. 1096/fj.201802427RRR
- Zobel, E. H., Ripa, R. S., von Scholten, B. J., Rotbain Curovic, V., Kjaer, A., Hansen, T. W., et al. (2021). Effect of Liraglutide on Expression of Inflammatory Genes in Type 2 Diabetes. *Sci. Rep.* 11, 18522. doi:10. 1038/s41598-021-97967-0

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