RESEARCH ARTICLE

Variation in Neotropical river otter (*Lontra longicaudis*) diet: Effects of an invasive prey species

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Abstract

Predation is one of the main barriers that exotic species may face in newly colonized areas and may help stop or control the potential negative impacts of invasive species in the environment. We evaluated if the consumption of an invasive prey (armored catfish: *Pterygoplichtys* sp.) affects the dietary niche breadth and trophic level of a native predator (Neotropical river otter: *Lontra longicaudis*) in northern Guatemala. We examined otter scats from three rivers: two where the invasive armored catfish occurred and one without the invasive fish. Samples were collected two and seven years after the first report of the catfish in the area. We performed gross scat analysis and stable isotope analyses of nitrogen and carbon of fecal matter. Where the invasive armored catfish occurred, it was the main prey item for *L. longicaudis*. Particularly in the river outside of protected areas seven years after the first report of the catfish, where it accounted for 49% of the otter diet. Concordance was found between the two techniques to estimate dietary niche breadth and trophic level. The dietary niche breath of otters was narrower seven years after the invasion in comparison to two years after the invasion in both invaded rivers, but the extent of the reduction was lesser inside the protected area. Finally, the trophic level of otters also showed a reduction related to the occurrence of the armored catfish in their diet.

Introduction

Predators may change their diet after an exotic prey species becomes established and abundant in the predator's range [1–4]. Inclusion of such a species in a predator's diet can lead to a shift in the predator's dietary niche, which may become wider or narrower, depending on the intensity of use of the new resource and changes in the use of alternative native prey [5]. Furthermore, the type of prey that a predator eats defines its trophic level (e.g., primary consumer, secondary consumer). Both niche breadth and trophic levels can be evaluated using gross scat analysis and stable isotopes analyses.

An important group of invasive species in freshwater communities are the armored catfishes of the South American family Loricariidae, a diverse group of fishes with 928 valid

species and eight subfamilies, including the genus *Pterygoplichthys*, commonly known as the suckermouth armored catfish (hereafter ACF; [6]). These catfish are very popular in the aquarium trade, easily domesticated, exhibit parental care [7], possess physiological tolerance to adverse conditions [8–12], have wide distribution ranges [13], and possess high reproductive and growth rates [14,15]. They feed on detritus, an abundant resource, especially in human-modified areas, and therefore have a low fractional trophic level (FTL) [13]. These traits contribute to their invasiveness, as they fulfill the six life-history variables associated with species that successfully establish invasive populations [16]. The presence of ACF as an invasive species has been documented for at least 21 countries in five continents [17]. In 2005, an established population of *Pterygoplichthys pardalis* was found in Laguna Frontera at the mouth of the Usumacinta River, Tabasco, Mexico [18]. Two years later, *P. pardalis* was reported in Guatemala in the headwaters of the San Pedro River, a tributary of the Usumacinta River (Juarez-Sanchez and J. F. Moreira, in prep.). The species identification, however, has not been confirmed because *P. pardalis* can be misidentified and confused with other species of *Pterygoplichthys* given that identification is based on ventral color patterns and hybridization with *P. disjunctivus* has been reported elsewhere [19–21].

The ACF has been reported to have positive effects by generating nutrient hotspots, making nutrients available for producers in nutrient-depleted areas [22]. However, the amount of nutrients released by the ACF does not compensate for its grazing pressure [23]. Other negative impacts of ACF have been documented in places where they have established invasive populations. These impacts include asphyxiating native predators in Puerto Rico [24]; preying on native fish eggs and first-feeding fry in Thailand [25]; competing for forage with native species, reducing biofilm from the substrate, and changing the proportions of dissolved nutrients in the Philippines and Mexico [23,26,27]; harassing manatees [28–30]; and possibly promoting erosion with their nesting burrows in Florida and Mexico [7,31]. These impacts could occur anywhere ACF establish an invasive population. Invasive ACF are preyed upon by native piscivorous predators such as common snook (*Centropomus undecimalis*) and the Neotropical cormorant (*Phalacrocorax brasilianus*) [32,33], although their effects on these and other native predators have not been evaluated.

Otters (Lutrinae) are mid-sized carnivores that are top predators in freshwater wetlands and riverine systems because of their high energetic demand and trophic position [34,35]. The Neotropical river otter (*Lontra longicaudis*; hereafter NRO) is a semi-aquatic mustelid that preys primarily on benthic slow-moving fish [36], but also feeds on crustaceans, mollusks, reptiles, and mammals (Table 1). This species is distributed from northern Mexico to northern Argentina, coexisting with different community assemblages of prey species, and adapting its foraging behavior according to the local community [37]. Where ACF are native, they coexist with the NRO and constitute one of the most important prey items in its diet [37–40]. However, the role of ACF as a prey item for NRO in areas where ACF has been introduced is unknown and may be reshaping the foraging ecology of the NRO in those areas.

The main objective of this study was to determine if invasive armored catfish affected the diet of Neotropical river otters. Given that NRO feed on ACF in areas where native populations overlap [38,46,47], we hypothesized that NRO will change their diet to include ACF in rivers where invasive populations of ACF occur. We predicted that where ACF are present, they will become the main prey of NRO and reduce the niche breadth of NRO. If ACF become the main prey of the NRO, we also predicted a lower trophic level for the NRO in areas where ACF are present due to the low trophic level of the ACF.

Table 1. Food items reported as present in diets of Neotropical river otters across their geographic range.

Locality	Primary item	Other items	Citation
Oaxaca, México.	crustaceans (53.0%)	fish (33.1%), insects (9.8%) and amphibians (4.0%)	[41]
México state, México.	fish (92.4%)	invertebrates (3.5%), amphibians (2.9%) and plant matter (1.8%)	[42]
Alto Cauca, Colombia.	fish (76.7%)	insects (12.7%), reptiles (0.7%), and others (9.9%)	[39]
Salta, Argentina.	fish (53%)	insects (24%), crustaceans (16%), amphibians (7%), and reptiles, mammals and mollusks (<0.1%)	[37]
Rio de Janeiro, Brazil.	fish (86%)	crustaceans (71%), amphibians (10%), mammals (3%), birds (0.6%), reptiles (0.2%) and others (0.7%)	[43]
Rio Grande do Sul, Brazil.	Fish (Loricariidae 41.1%, Cichlidae 21%, Pimelodidae 12.6%, Characidae 6.5%)	other fish (12.5%), Megaloptera (3.6%), mammals (1.2%), insects (0.4%), Decapoda (0.1%), birds (0.3%), snakes (0.3%) and plant matter (0.4%)	[38]
Rio Grande do Sul, Brazil.	fish (82.6%)	crustaceans (20.6%), birds (4.5%), mammals and snakes (3.7%), Coleoptera (1.2%), amphibians (0.8%) and mollusks (0.4%)	[44]
Rio Grande do Sul, Brazil	fish	mammals, amphibians, birds, snakes, insects, crustaceans mollusks and eggs.	[45]

Percent values are frequency of occurrence and do not add to 100%.

Materials and methods

This research and its associated fieldwork were conducted under permit from the Consejo Nacional de Areas Protegidas (CONAP), Presidencia de la Republica, Guatemala C.A. No. 047/2015

Study area

The study area is located at northern Guatemala in the district of Petén (between 15.50° N and 17.50° N and 88.50° W and 91.25° W) and includes the Usumacinta and Mopan basins (Fig 1). Precipitation ranges from 1,200 to 4,000 mm/year on a gradient decreasing northward (INSIVU-MEH, 2016). Major habitat types in the study area consist of subtropical moist forest in the north, subtropical very moist forest in the south, and tropical very moist forest in the southeast [48]. The entire study area consists of lowland forest, with elevations ranging from 0 to 1000 masl.

In northern Guatemala, rivers flow into the Gulf of Mexico or into the Caribbean Sea watersheds (Fig 1). Thus, bodies of fresh water are isolated by large expanses of land in the headwaters, and large distances between river mouths along the coast. The Mopan River flows northwards from southern Petén and then east in central Belize into the Caribbean Sea. The Usumacinta River runs northwest into the Gulf of Mexico. Samples were collected from the Mopan River and two tributaries of the Usumacinta River: The San Pedro River and the Pasion River. In Guatemala, the San Pedro River flows along the southern border of Laguna del Tigre National Park with a well preserved riparian forest but it is been threatened by manmade fire use to open land for cattle ranching. The Pasion and the Mopan rivers run through private lands and some small protected areas that are under different land uses, mainly cattle ranching operations and African palm plantations.

In Guatemalan territory the Usumacinta basin has at least 61 fish species distributed in 25 families. The two main families in Usumacinta basin are Cichlidae with 18 species and Poeciliidae with 10 species [6,13,49,50]. To our knowledge, no peer-reviewed document has been published that describes fishes of the Mopan River within the borders of Guatemala. Thus, information about the fish assemblage in this river is based on information from the estuarine area in Belize. Therefore, the number of fish species that we are considering as present and
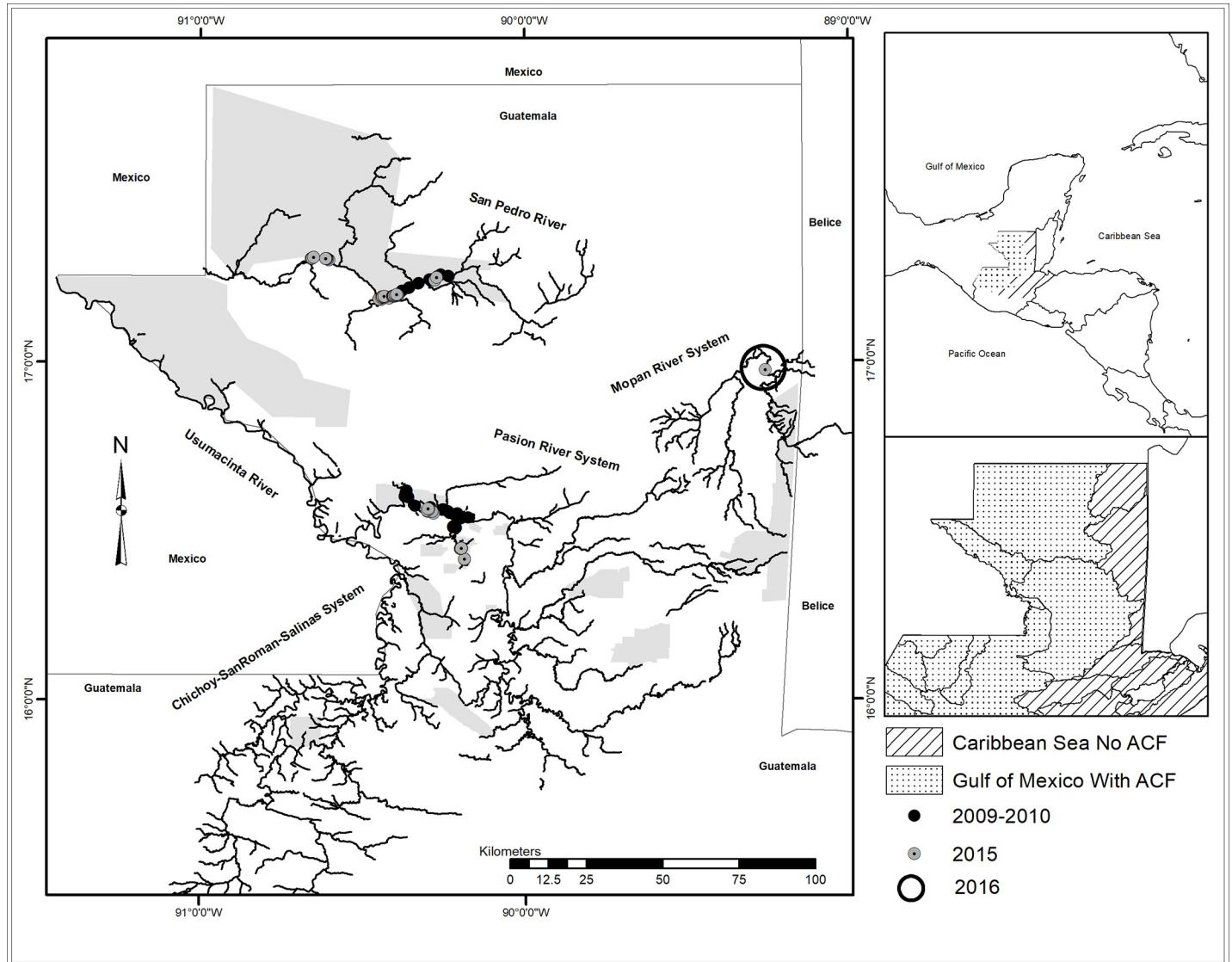
Fig 1. Study area for collection of Neotropical river otter scats in northern Guatemala. Grey circles represent samples collected in 2009–2010; black solid circles represent samples collected in 2015; black hollow circle represents the area where samples were collected in 2016. The dashed area represents the Usumacinta basin divided in sub-basins, where the armored catfish has been reported (ACF). The striped area represents the Caribbean runoff where no ACF has been reported. Grey areas represent protected areas.

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potential prey for otters in the river headwaters within Guatemalan territory may be inflated. In Mopan River, there are at least 103 fish species distributed in 32 families, including the invasive tilapia (*Oreochromis aureus*). The main families are Cichlidae with 14 species and Poeciliidae with 16 species [6,13,51]. Exotic tilapia is widespread across all Guatemala due to multiple introductions, both accidental and deliberate from aquaculture or governmental fisheries restocking. The Asian grass carp (*Ctenopharyngodon idella*) and the ACF have been found in the Usumacinta basin, but the origins of these invasions are not clear.

Scat collection

Samples were collected during three periods: May 2009 –April 2010, May–July 2015, and June 2016. The search for otter scats was conducted from a small boat moving at slow speeds (< 5

km/h) close to the shoreline, with scats and latrines typically found on protruding structures (e.g. rocks or fallen trees). This search was conducted along both shorelines of the river in opposite directions. All scats were collected, placed in paper and/or plastic bags with silica gel, and stored in a dry environment. Otter scats were identified by their appearance, as no other species present in the study area have similar scats (located on protruding sites along the river shore, low fecal matter and high content of fish or crab remains) [52]. If a scat was found but its identification was doubtful, it was collected and included in the analysis only if otter hair from grooming was found on it. Each scat was assigned a unique code and the geographic coordinates of its location were recorded using a handheld GPS unit (GARMIN Astro 320, Garmin Ltd. Kansas City, USA).

We sampled the Usumacinta basin during 2009–2010 using continuous searches along the rivers, including 38.5 km of the San Pedro River (starting from Paso Caballos village and moving west) and 89.1 km of the Pasion River (starting from Sayaxche town and heading west). We sampled the Usumacinta and Mopan basins in 2015 by organizing the search for scats into segments of 10 km, with segments separated by at least 10 km. In the Usumacinta basin, we sampled along 40 km in San Pedro river and 50 km in Pasion river. Surveys began in Paso Caballos for the San Pedro and in Sayaxche for the Pasion. The Mopan River was sampled along 10 km in 2015 near La Polvora military base. In June 2016, local fisherman collected samples in the Mopan River near La Polvora military base, no exact georeference was collected per sample (Fig 1).

Scat sample preparation and analysis

Scats were dried and stored with silica gel for at least 6 months. From each dry scat, the fecal matter was collected. We define as fecal matter the dust that remains after removing all the hard remains (i.e. hair, shells, scales or any skeletal pieces) from the scat. This fecal matter was homogenized using a porcelain mortar and pestle, stored in glass vials and sent to the Light Stable Isotope Mass Spectrometry Laboratory in the Department of Geological Sciences at the University of Florida for stable isotope analysis (SIA) of $\delta^{15}N$ and $\delta^{13}C$. Samples were analyzed using a Thermo Electron DeltaV Advantage isotope ratio mass spectrometer coupled with a ConFlo II interface linked to a Carlo Erba NA 1500 CNHS Elemental Analyzer. All carbon isotopic results are expressed in standard delta notation relative to VPDB. All nitrogen isotopic results are expressed in standard delta notation relative to AIR. Hard remains (i.e., scales, skeleton pieces) were separated and identified to the lowest possible taxonomic level. A list of potential prey species for otters was made, consisting of all the fish species reported in the study area that have a reported maximum total length $\geq$ 100 mm (S1 Table). Size selection was based on the assumption that otters prefer to feed on fish within the 100–150 mm size range [53]. Prey remains that could be identified were fish scales, otoliths or vertebrae; crustacean shells; and mammal hairs. A scale guide was constructed for 68 of the 80 scaled fish species that are found in the sampled river basins and that were consider potential prey of the NRO [54]. Scales were obtained from museum specimens at the Florida Museum of Natural History (FLMNH) and El Colegio de la Frontera Sur in México (ECOSUR). Scales from these fish species were cleaned with water and alcohol, placed on glass slides with nail polish, and sealed with a coverslip to make semi-permanent slides. For 10 catfish species that do not have scales, the identification was based on fin spines, using reference material from the zooarchaeological collection at FLMNH. Hairs found in the scats were identified using a hair-identification guide [55] and reference material from the mammal collection of the Museo de Historia Natural (MUSNAT) at the Universidad de San Carlos de Guatemala (USAC). Otter hair (product of grooming) was saved and pressed between glass slides and coverslips for future analysis.

Data analyses

For data analyses, the sampling units were the rivers (San Pedro, La Pasion, Mopan) with year as factor (2009–2010 and 2015–2016). The year effect represents 2 and 7 years after the advent of the ACF invasion. Comparisons were made over time (i.e., same river, different year) only using data from Pasion and San Pedro rivers where the ACF are present; we additionally looked at differences across river basins in the same sampling years (i.e., different river, same year), combining 2015–2016 records as one year and including the Mopan River where ACF do not occur.

The importance of each prey species can be biased by abundant and conspicuous hard remains that are identifiable for some species, even if those species are consumed in low numbers, due to differential digestibility of prey items. This overestimation of some species can then lead to an underestimate of overall diet diversity. On the other hand, when using SIA of predators, one can measure diet diversity breadth and comparative trophic levels but with no taxonomic information about the prey. For this reason, we used both techniques, expecting to find concordance between them.

Gross scat analysis (GSA)

Accumulation curves were constructed using program EstimateS (Colwell 2013, Connecticut, USA) where the expected number of prey species found in a given number of scats is obtained by

$$\tau(h) = S_{abs} - \sum \alpha_{jh} S_j$$

$$\alpha_{jh} = (H - h)!(H - j)!/(H - h - j)!H!$$

were $\tau(h)$ is the estimated number of species for h number of scats; S_{obs} is the number of species actually observed; S_j is the number of prey species found in j scats; α_{jh} is a combinatorial coefficient; H is the total number of scats; h is the number of possible combination of scats that add up to j scats; and j is the number of scats per moment or segment of the curve [56].

The importance of different food items, including the ACF, in the NRO diet was assessed through GSA, using the percentage of occurrence. Percentage of occurrence was estimated for a prey item by dividing the number of scats with item i by the total number of reported items. To compare the NRO niche breadth between basins, with different prey communities, Levin's index was used:

$$B = 1/\sum p_j^2$$

where p is the proportion of food items from category i [57]. The Levin's niche-breadth index can be standardized using:

$$B_a = B - 1/n - 1$$

where B_a is the standardized Levin's niche-breadth index, B is Levin's niche-breadth index, and n is the number of recorded species. Levin's index ranges from 1 to n and from 0 to 1 in its standardized version. In both cases, its minimum value is reached when all reported prey belongs to only one species (specialist predator) and is at its maximum when all the species are consumed in the same proportion (generalist predator). It has been suggested that values of B_a > 0.6 represent a generalist and values of B_a < 0.4 a specialist [58,59]. To estimate confidence intervals the samples (scats) were randomly selected with replacement (bootstrap), then the index was re-estimated with the resulting set of samples. This procedure was repeated 1000 times and the confidence intervals calculated.

The NRO's fractional trophic level, which represents the trophic distance of a consumer species from producers, in each basin was estimated using Pauly and Palomares's (2005) formula:

$$FTL_i = 1 + \sum_j (FTL_j DC_{ij})$$

where FTL_i is the fractional trophic level of the consumer, +1 is a constant increment for the FTL of a consumer, FTL_j is the fractional trophic level of the prey j, and DC_{ij} is the proportion of contribution of prey j to the diet of consumer i. Prey FTL_j values were obtained from Fish-Base database[13] for fish and as the average FTL reported for a similar species for crabs [60]. The DC_{ij} was based on proportion of occurrence values by river-year combination in the otter scats. To estimate confidence intervals a bootstrap procedure was developed as explained above.

Stable isotope analysis (SIA)

Stable isotope analysis (SIA) measures the proportion of heavy to light stable isotopes in a sample [61,62]; its values are expressed in delta notation (δ) or per mil (‰) and estimated with this equation:

$$\delta = ((R_{sample}/R_{standard}) - 1) * 1000$$

where R = heavy isotope / light isotope obtained with a mass spectrometer.

Isotopic values of a predator are generally higher than those of its prey due to a process called fractionation, wherein the molecules with the lighter isotopes, given their lighter overall weight, react faster and can be metabolized and excreted faster than the heavier ones. This process results in the predator being enriched with a higher proportion of heavier isotopes than its prey [62,63]. The mean value of this fractionation across taxa is 3.4‰ (1 SD = 1‰) for $\delta^{15}N$ and 0.4‰ (1 SD = 1.3‰) for $\delta^{13}C$ [63]. These values are the expected increment of the isotopic value when molecules are assimilated from prey tissue to predator tissue (from lower to higher trophic levels;[64]).

Isotopic values of different tissues, such as bone, blood, hair or muscle, have been used to evaluate the diets of a wide range of species [65–72]. Normally, tissue samples are obtained from dead or captured specimens but these invasive techniques are sometimes difficult or impossible to use, especially for secretive, rare or endangered species. However, controlled experiments have shown that SIA based on feces is sensitive to changes in the diet over periods of 3 hours for insectivorous bats [73] and, thus, represent the isotopic values of the latest meals of the individual that produced the scat[74]. In carnivores and omnivores, SIA based on scats can be used to estimate the main type of prey and nutrient flow, using $\delta^{15}N$ to infer the range of trophic positions or FTLs at which a predator eats, and $\delta^{13}C$ to determine the type of producers that supported the specific trophic chain [63,75–77]. Further, the variance of isotopic values of a population may represent the niche width (or breadth) of a consumer [78].

Taking $\delta^{15}N$ and $\delta^{13}C$ values from individual scats as samples from each river, we evaluated the data for normality using histograms, qq-plots and a Shapiro-Wilk normality test; all values followed a normal distribution. To evaluate differences between variances in $\delta^{13}C$ and $\delta^{15}N$ as a niche breadth metric, a Levene's homoscedasticity test was used. To test for differences in mean $\delta^{15}N$ values between rivers and years we use a two-factor ANOVA after a log transformation of the data to correct for lack of homoscedasticity; a post-hoc paired t-test with Bonferroni adjusted p-values was used to evaluate where the differences occurred. All the statistical procedures except for the species accumulation curves were performed using the program R [79] and its packages lawstat [80], dplyr [81] and ggplots2 [82].

Results

Field collection of scats yielded 286 samples identified as coming from the NRO. After eliminating scats that had some type of contamination (e.g., wood, mud, or termite nest), 177 samples of fecal matter were sent for isotopic analysis (Table 2). We identified 35 scaled fish species, including three nonnative fish species (*Oreochromis aureus*, *Ctenopharyngodon idella* and *Pterygoplichtys* sp.) from otter scats. In addition, remains of unidentified insects, one unidentified reptile, one unidentified mammal, and one species of crabs and one species of crayfish were recovered from the scats (Table 3).

Niche breadth

Pterygoplichtys sp. was the main identifiable prey item in all samples from the Usumacinta basin. Occurrence of ACF in scat samples was highest (49%) in samples collected from Pasion River 7 years after the first report of the catfish, an increase from 9.9% in 2010 (Table 3). ACF occurrence also increased in the San Pedro River, but less than in the Pasion River. *Oreochromis aureus* was an important item (percentage of occurrence > 5%) for otters in the Pasion and San Pedro rivers in 2015 and the Mopan River in 2016 (Table 3).

Based on species accumulation curves, the expected number of prey species was marginally lower in 2015 than in 2010 in Pasion River samples (Fig 2A); no difference was seen for San Pedro River samples (Fig 2B). When all three rivers were compared based on data from 2015–2016, otters from the San Pedro River were expected to have more prey species, those from the Pasion River fewer species, and those from the Mopan River were expected to have a middle number of prey species. Confidence intervals around expected numbers were wide and overlapped, especially between curves from the Mopan River and the other two rivers (Fig 2C). Further, the assumption that all samples used to construct the accumulation curves were independent may have been violated because some of the scats were collected from the same latrine.

Niche breadth (Levin's index, B_a) of the Neotropical river otter was lower 7 years after the ACF invasion when compared to 2 years after the invasion in the San Pedro River ($B_a = 0.53$ in 2009 vs 0.29 in 2015). A similar situation was found in Pasion River ($B_a = 0.47$ in 2010 vs 0.18 in 2015). NRO niche breadth varied among the three rivers in 2015, with similar values in San Pedro River and Mopan River and lower values in Pasion River ($B_a = 0.29$, 0.28 and 0.18 respectively; Table 4).

Table 2. Scats of Neotropical river otters collected in northern Guatemala.

River	No. of scats collected (year)	No. of scats without contamination (year)
Usumacinta	1 (2015)	0
	1 Total	**0 Total**
San Pedro	36 (2009)	20 (2009)
	117 (2015)	55 (2015)
	153 Total	**75 Total**
La Pasion	52 (2010)	36 (2010)
	40 (2015)	34 (2015)
	92 Total	**70 Total**
Mopan	1 (2015)	1 (2015)
	39 (2016)	31 (2016)
	40 Total	**32 Total**

Only scats without contamination were used for fecal matter isotope analyses

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Table 3. Number of records (No.), percentage of scats with each prey species (% S) and percentage of records of each prey species per total of records (% R) found in otter scats collected from the Mopan, Pasion and San Pedro rivers, northern Guatemala.

		Mopan 2015–2016			Pasion 2010			Pasion 2015			San Pedro 2009			San Pedro 2015		
		No.	% S	% R	No.	% S	% R	No.	% S	% R	No.	% S	% R	No.	% S	% R
Belonidae																
	Strongylura hubbsi										3	8.3	2.2			
	Strongylura marina	1	2.5	1.1												
Carangidae																
	Caranx latus	1	2.5	1.1												
Centropomidae																
	Centropomus ensiferus	1	2.5	1.1												
Characidae																
	Astianax fasciatus										9	25.0	6.5	1	0.9	0.3
Chichlidae		,														
	Chuco intermedius	6	15.4	6.6										1	0.9	0.3
	Cincelichthys bocourti	1	2.5	1.1				1	2.5	1.4				1	0.9	0.3
	Cincelichthys pearsei													1	0.9	0.3
	Cribroheros robertsoni	1	2.5	1.1	13	25.0	9.2	4	10.0	5.5	22	61.1	15.8	12	10.3	4.2
	Kihmchithys ufermammi										3	8.3	2.2	1	0.9	0.3
	Maskaheros argenteus				1	1.9	0.7									
	Mayaheros urophtalmus	2	5.0	2.2	5	9.6	3.5	5	12.5	6.8	5	13.9	3.6	17	14.5	5.9
	Oreochromis aureus	8	20.0	8.8	5	9.6	3.5	6	15.0	8.2	4	11.1	2.9	21	17.9	7.3
	Parachromis friedrichsthalii	2	5.0	2.2	9	17.3	6.3	5	12.5	6.8	1	2.8	0.7	4	3.4	1.4
	Petenia splendida				1	1.9	0.7							3	2.6	1.0
	Rheoheros lentiginosus				1	1.9	0.7	1	2.5	1.4						
	Rocio octofasciata				2	3.8	1.4	1	2.5	1.4				3	2.6	1.0
	Thorichthys affinis				2	3.8	1.4							1	0.9	0.3
	Thorichthys aureus	6	15.0	6.6												
	Thorichthys meeki	5	12.5	5.5	11	21.2	7.7	2	5.0	2.7	10	27.8	7.2	25	21.4	8.7
	Thorichthys pasionis				10	19.2	7.0	1	2.5	1.4	10	27.8	7.2	8	6.8	2.8
	Trichromis salvini													2	1.7	0.7
	Vieja bifasciata				3	5.8	2.1	6	15.0	8.2	1	2.8	0.7	21	17.9	7.3
	Vieja melanurus				3	5.8	2.1	1	2.5	1.4				8	6.8	2.8
Cyprinidae																
	Ctenopharyngodon idella				7	13.5	4.9							5	4.3	1.7
Eleotridae																
	Dormitator maculatus	1	2.5	1.1												
Gerreidae																
	Eugerres mexicanus													4	3.4	1.4
Hemiranphidae																
	Hyporhamphus mexicanus										9	25.0	6.5	2	1.7	0.7
Lepisosteidae																
	Aractosteus tropicus										1	2.8	0.7			
Loricariidae																
	Pterygoplichthys spp				14	26.9	9.9	36	90.0	49.3	23	63.9	16.5	75	64.1	26.0
Megalopidae																
	Megalops atlanticus													1	0.9	0.3
Mugilidae																
	Mugil cephalus				1	1.9	0.7									
Poeciliidae																
	Belonesox belizanus				5	9.6	3.5				1	2.8	0.7	12	10.3	4.2
	Poecilia mexicana	1	2.5	1.1	3	5.8	2.1	1	2.5	1.4	17	47.2	12.2	24	20.5	8.3
	Poecilia petenensis				11	21.2	7.7	1	2.5	1.4	17	47.2	12.2	26	22.2	9.0

(Continued)

Table 3. (Continued)

		Mopan 2015–2016			Pasion 2010			Pasion 2015			San Pedro 2009			San Pedro 2015		
		No.	% S	% R	No.	% S	% R	No.	% S	% R	No.	% S	% R	No.	% S	% R
Ariidae, Heptapteridae, Ictaluridae																
	Catfish	8	20.0	8.8	3	5.8	2.1	1	2.5	1.4	3	8.3	2.2	4	3.4	1.4
Pseudothelphusidae																
	Potamocarcinus sp	32	80.0	35.2	30	57.7	21.1							3	2.6	1.0
Palaemonidae																
	Macrobrachium sp													1	0.9	0.3
Unknown Insects		3	7.5	3.3												
Unknown Reptile		3	7.5	3.3				1	2.5	1.4						
Unknown Mammal		1	2.5	1.1												
Unknown		8	20.0	8.8	2	3.8	1.4							2	1.7	0.7
	Totals	91	227.5	100.0	142	273.1	100.0	73	182.5	100.0	139	386.1	100.0	289	247.0	100.0
No. of sp / No. of scats		19/40			22/52			16/40			17/36			29/117		

The precision (one standard deviation of standards) of the δ^{15}N and δ^{13}C reads was 0.097 and 0.080 respectively, n = 34.

https://doi.org/10.1371/journal.pone.0217727.t003

Isotope values ranged from 5.89 to 16.39 for δ^{15}N and -38.31 to -20.61 for δ^{13}C (Fig 3) and did not depart from a normal distribution so no transformations were needed. Variance of δ^{15}N signatures from fecal samples differed among groups (Levene's test for homoscedasticity; W = 2.54, p = 0.042; Fig 4A). Based on pairwise comparisons, variance of δ^{15}N signatures from the Pasion River did not differ significantly between years (σ^2 = 2.45 in 2010 and σ^2 = 1.80 in 2015; W = 0.78, p = 0.37; Fig 4A). In contrast, variance of δ^{15}N differed significantly between years in samples from San Pedro River (σ^2 = 4.83 in 2009 and σ^2 = 1.73 in 2015; W = 6.68, $p < 0.01$; Fig 4A). The δ^{13}C variances also differed among groups (W = 3.23, $p < 0.01$; Fig 4B), with pairwise contrasts indicating that δ^{13}C variances increased across years for Pasion River (σ^2 = 3.65 in 2010 and σ^2 = 6.49 in 2015; W = 3.83, p = 0.05; Fig 4B) and San Pedro River (σ^2 = 2.04 in 2009 and σ^2 = 7.09 in 2015; W = 6.75, p = 0.01; Fig 4B).

Trophic level

Calculations of FTL values excluded information from *Maskaheros argenteus* (found in one sample from the Pasion River), insects, reptiles, crayfish and unknown species because no data on the FTL of those prey items were available. The highest FTL values for NRO came from the Mopan River in 2016, Pasion River in 2010, and San Pedro River in 2009, with lower values from the Pasion and San Pedro rivers in 2015 (Table 5).

Values of δ^{15}N from NRO samples were highest in the Mopan River in 2015 (based on only one specimen), followed by mean values from the Pasion River in 2010 and the San Pedro River in 2009 (Fig 5). Lowest values came from the Pasion and San Pedro rivers in 2015 (Fig 5). Values of δ^{15}N from sites in the Usumacinta basin differed across years (ANOVA, $F_{1,141}$ = 67.98; $p < 0.001$) and across rivers (ANOVA, $F_{1,141}$ = 15.53; $p < 0.001$) with no interaction between the two factors (ANOVA, $F_{1,141}$ = 2.76; p = 0.10). Higher values were found from scats collected during the early sampling years in the Pasion and San Pedro rivers, two years after the first report of the ACF (post-hoc pairwise t-test with Bonferroni adjusted p-values: Pasion 2010 vs. 2015 t = 5.37, df = 68, $p < 0.001$; San Pedro 2009 vs. 2015, t = 5.31, df = 24.122,
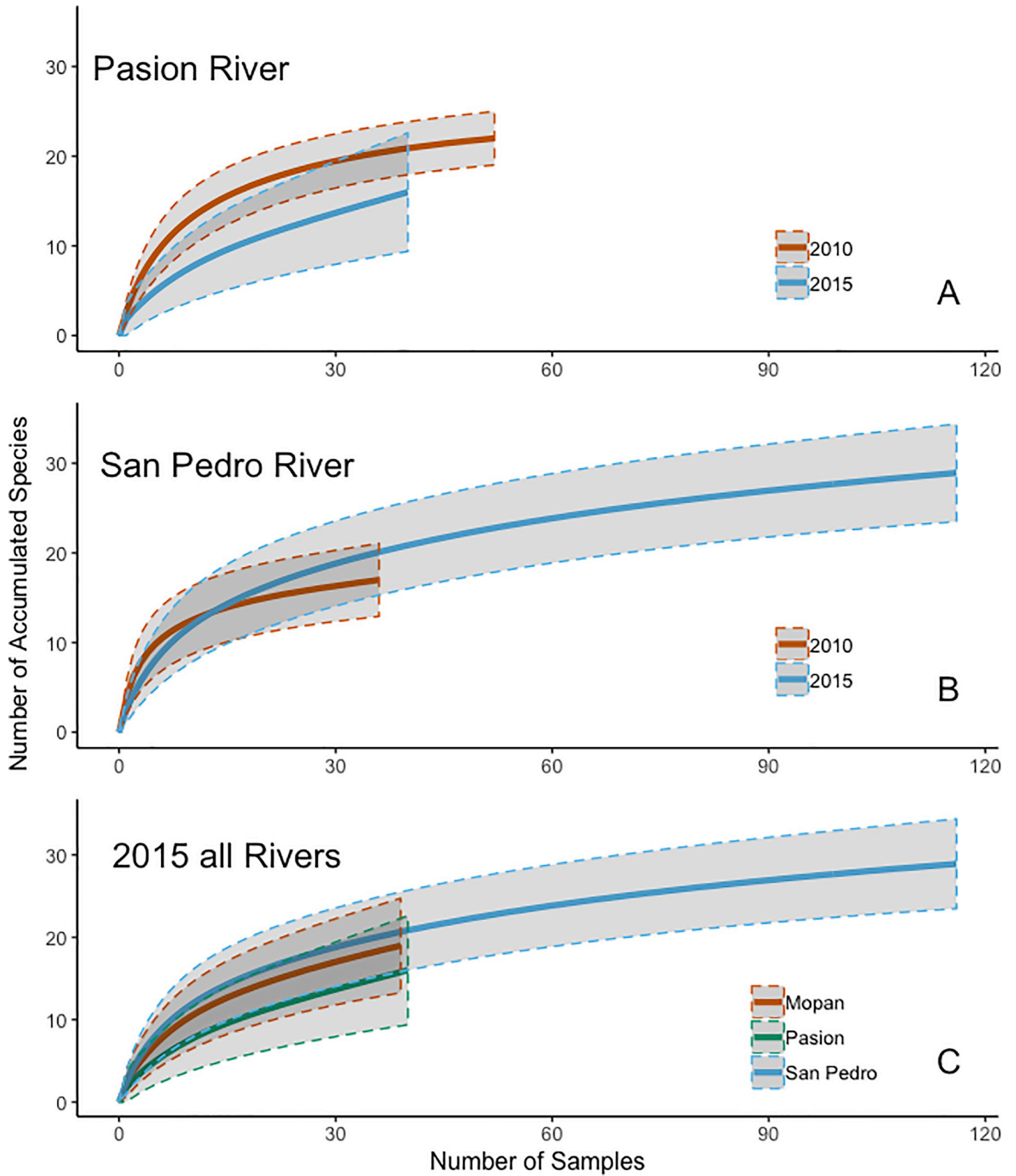
Fig 2. Species accumulation curves for prey species found in scats of Neotropical river otter in the **(A)** Pasion River, Guatemala 2010 and 2015; **(B)** San Pedro River, Guatemala, in 2009 and 2015; and **(C)** Mopan River 2016, Pasion River 2015 and San Pedro River 2015.

$p < 0.001$). Mean values of NRO δ^{15}N did not differ between the Pasion and San Pedro rivers from same sampling years (Pasion vs. San Pedro 2010–2009 t = -0.40, df = 54, p = 1.0; Pasion vs. San Pedro 20015, t = 2.42, df = 87 p = 0.23).

Discussion

Concordance between the gross scat analysis and stable isotope analysis values strongly supports the idea that an increase in consumption of the armored catfish reduced the dietary niche breadth of the neotropical river otter and trophic level at which the otter feeds in northern Guatemala. As predicted, ACF became the main prey species for the NRO in invaded rivers and, consequently, NRO δ^{15}N variances and mean values decreased over time in both invaded rivers (with a weaker decline in Pasion River). The same pattern was observed in the standardized niche breadth index (B_a). Further, the wider niche breadth (B_a values) in the San Pedro River may be related to its higher environmental integrity (located adjacent to a national park) that could help sustain the richness of native NRO prey or reduce the invasiveness of the ACF. This conclusion is supported by the species accumulation curves. Invasive species are predicted to have better chances of establishment in native assemblages that are depleted or disrupted and more likely to have long-term success in systems highly altered by human activity [83]. The increase in δ^{13}C variation over time suggests that the NRO diet has included a prey that consumes different producer types or a prey that consumes producers in a different proportion, likely because of the ability of ACF to exploit a different range/proportion of plant resources than natives from the same trophic guild [84]. Furthermore, the decrease in FTL across rivers (Mopan River showing similar values to San Pedro River and higher than Pasion River) combined with lower mean values of δ^{15}N provide evidence of a reduction in the NRO trophic level associated with ACF presence.

The range of prey types exploited by NRO changed after the invasion of ACF, with the lowest dietary niche breadth found in Pasion River seven years after the invasion. The dietary NRO niche breadth decreased in Pasion and San Pedro rivers, even though the number of prey species consumed by NRO was highest in the San Pedro River in 2015. This result is concordant with the idea that predators can use a wide range of resources but still concentrate on a subset of those resources [85]. It also supports the statement that NRO prey mostly on slow-moving and territorial prey species [36]; the main prey species for NRO in this study included Loricariidae, Cichlidae, large Poeciliidae, and crabs (Table 3).

Results based on GSA and δ^{15}N variances were similar for both indexes, with narrower niche breadth 7 years after initiation of the ACF invasion compared to 2 years after the

Table 4. Neotropical river otter niche breadth (Levin's index, B_a) in the study area.

River	year	Ba	2.5% quantile	97.5% quantile
Mopan	2016	0.29	0.36	0.23
San Pedro	2009	0.53	0.60	0.50
San Pedro	2015	0.29	0.33	0.25
Pasion	2010	0.47	0.59	0.39
Pasion	2015	0.18	0.25	0.11

Quantiles estimated using 1,000 bootstrap randomizations.
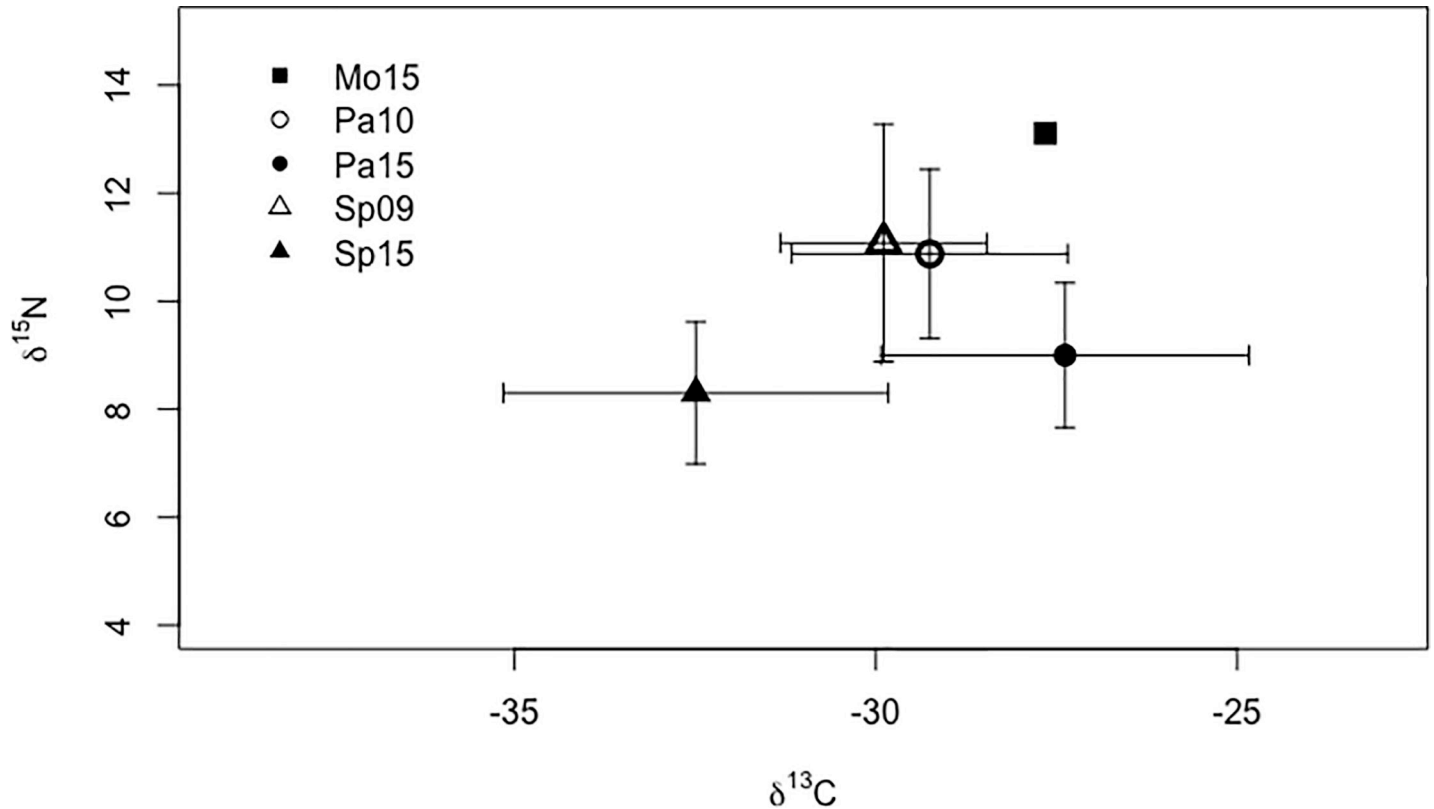
Fig 3. Isotopic values of δ^{15}N and δ^{13}C from Neotropical river otter scats collected from the study area. Error bars are one sd. Mo15 = samples from Mopan River 2015 (n = 1); Pa10, Pa15 = samples from Pasion River 2010 and 2015 (n = 36 in 2010 and 34 in 2015); Sp09, Sp15 = samples from San Pedro River 2009 and 2015 (n = 20 in 2010 and 55 in 2015).

invasion. The narrower dietary niche breadth found in the Pasion River in all situations and with both indexes in relation to the San Pedro River may be a result of Pasion River prey community been already depleted before the arrival of the ACF, and that the Laguna del Tigre National Park may provide some type of protection to the San Pedro River. A similar result was seen in a Bahamas mangrove system for grey snapper (*Lutjanus griseus*) with a reduced niche breadth (based on SIA) found in disturbed areas [5]. Therefore, it is possible that the higher values of NRO niche breadth in San Pedro River in relation to Pasion River are related
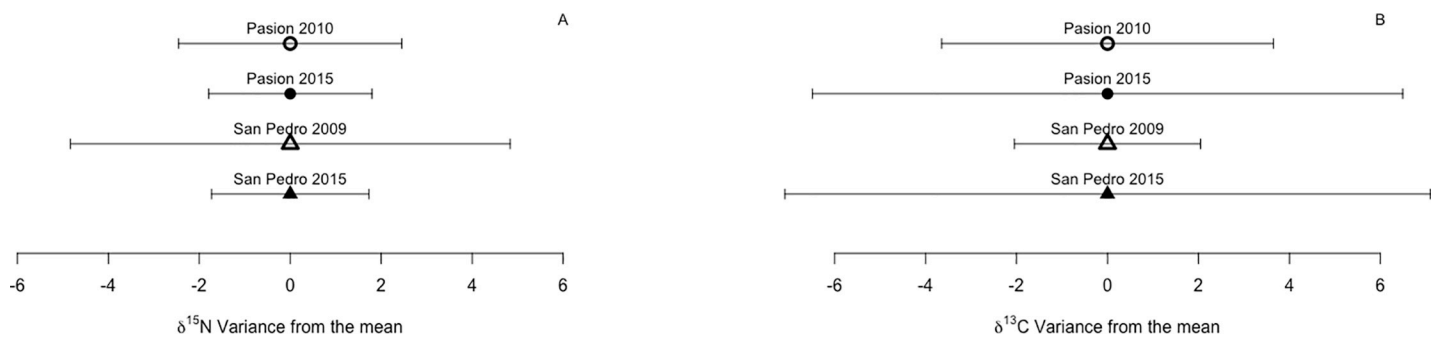
Fig 4. Variance from the mean of isotopes values in fecal samples from Neotropical river otters for (A) δ^{15}N and (B) δ^{13}C. The mean is set to 0 to help visualize the magnitude of the variances. Pasion River n = 36 in 2010 and 34 in 2015; San Pedro River n = 20 in 2010 and 55 in 2015.

Table 5. Neotropical river otter fractional trophic level (FTL) in the study area.

River	year	FTL	2.5% quantile	97.5% quantile
Mopan	2016	3.73	3.83	3.65
San Pedro	2009	3.71	3.79	3.64
San Pedro	2015	3.47	3.53	3.41
Pasion	2010	3.70	3.80	3.62
Pasion	2015	3.48	3.62	3.37

Quantiles estimation using 1,000 bootstrap randomizations.

to differences in the resilience of the two rivers due to differences in habitat conservation. Disturbances may facilitate the ACF or depress populations of native fish. For example, in the Guadalquivir marshes of southwestern Spain, the Eurasian otter (*Lutra lutra*) included high levels of an invasive species (75%; North American red swamp crayfish, *Procambarus clarkii*) in its diet within 10 years of the invasion. In the same area, various waterbirds similarly consumed this invasive species at higher rates in disturbed locations than in natural marshes [1].

In contrast to results from $\delta^{15}N$, variances of $\delta^{13}C$ in fecal samples were greater seven years after the ACF invasion compared to two years after the first sighting. Values of $\delta^{13}C$ represent the plant source of a food chain and a wider variance may indicate that primary consumers exploit a greater range of producers. Loricariidae may exploit a diverse variety of basal sources or a portion that the natives does not exploit, which may help explain the increase in the variance of $\delta^{13}C$ in NRO scats, given the increased presence of ACF in the NRO diet [86].

Native predators may act to reduce invasive species numbers [87,88], and such predation could be one of the main biological drivers by which streams resist the invasion of exotic species [89]. Further, predators from different taxa often adapt to and benefit from the consumption of invasive species [3,4,90]. In this context, NRO may act as a buffer to hold ACF populations at low levels and minimize their potential negative effects on the system. The question that arises from this situation, as in other systems where an invasive species becomes the main prey of a native predator [1], is whether the consumption of ACF by NRO and other native predators can facilitate the predators [3,4,91]. Greater predator populations might increase depredation on native prey that are threatened by overexploitation or habitat loss [92]. This effect is a valid concern in our study area, where cichlids, a group of fish that is highly appreciated by the local artisanal fisheries [93] were exploited as a group without much change when the consumption of ACF increased (Table 3). Also, concern for the increase of negative interactions between native predator and humans becomes relevant when wild predators establish dense populations in or near human-dominated areas [94,95] more research is needed to evaluate this situation.

Both GSA and $\delta^{15}N$ values indicated a reduction in the trophic level at which otters feed in rivers where ACF are present in northern Guatemala. Based on GSA, there were reductions in the FTL of NRO of approximately 0.22 FTL in the Pasion River and 0.24 FTL in the San Pedro River. These reductions may not represent much ecological difference. GSA may, however, under-estimate the consumption of some species and over-estimate the consumption of others either because of differences in digestibility of prey or because we measured presence of prey remains rather than consumed biomass, regardless of the amount of remains (not all remains were identifiable; e.g., spines). In contrast to GSA, SIA may give a more accurate result. Differences in mean $\delta^{15}N$ were as great as 1.88‰ for Pasion River and 2.78‰ for San Pedro River. If we use the widely accepted 3.4‰ enrichment ($\Delta^{15}N$) per FTL, these differences in mean $\delta^{15}N$ may represent changes of 0.5 to 0.8 FTLs in the Pasion and San Pedro rivers, respectively. The
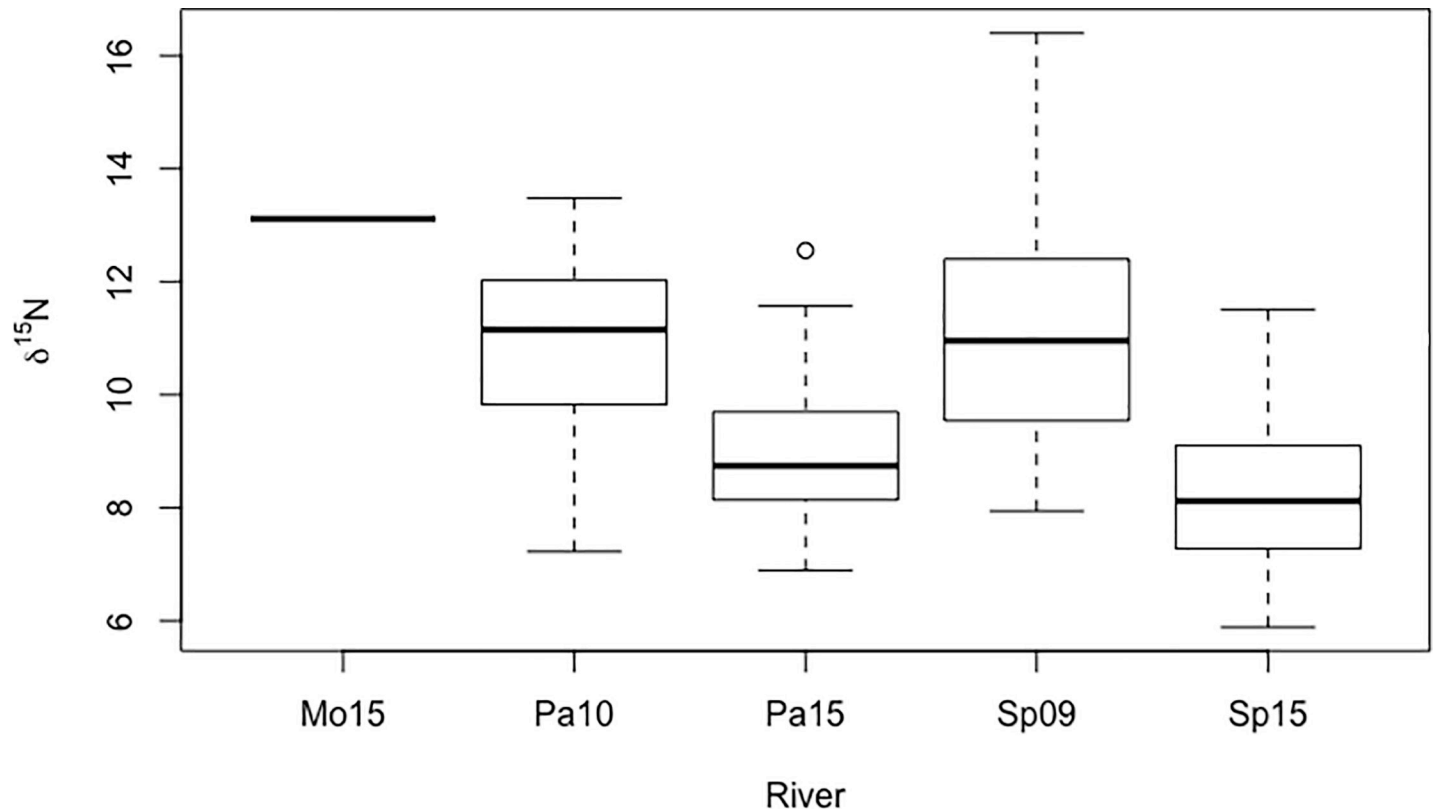
Fig 5. Boxplots for δ¹⁵N in fecal samples from Neotropical river otters in Guatemala. Mo16 = samples from Mopan River 2016 (n = 1); Pa10, Pa15 = samples from Pasion River 2010 and 2015 (n = 36 in 2010 and n = 34 in 2015); Sp09, Sp15 = samples from San Pedro River 2009 and 2015 (n = 20 in 2009 and n = 55 in 2015).

3.4‰ Δ^{15}N value has, however, been criticized. Models and empirical data have shown that this enrichment factor can underestimate FTL of marine predators [96]. In any case, the observed mean δ^{15}N values for NRO in both the Pasion and San Pedro rivers apparently represent a decrease in trophic level.

A reduction in the trophic level at which otters feed can have diverse effects on the riverine ecosystem. These effects may be difficult to anticipate and can compete with or interact with each other. It could mean predator release for other prey species that would benefit from reduced predation pressure [97,98]. On the other hand, consumption of the invasive species may benefit the predator, eventually leading to higher predator densities that could increase pressure on other native species. A model evaluating this situation suggests that predation on native prey by a native predator whose numbers have been enhanced by consumption of an invasive species can be more harmful than direct competition between native and invasive species [99]. Empirical data using SIA for golden eagles (*Aquila chrysaetos*) suggests that these eagles colonized the California Channel Islands after the introduction of feral pigs (*Sus scrofa*) [92]. Nonetheless, eagles still preyed on endemic meso-carnivores, including a fox (*Urocyon littoralis*) and skunk (*Spilogale gracilis amphiala*), pushing the fox towards extinction [92].

Another potential effect that needs to be evaluated is the reduction of trophic levels in the system by moving energy more directly from primary consumers to top predators. This results can occur by eliminating food-web links in the mid-trophic levels through competition or predation facilitated by a numerical response of predators in response to the high abundance of the invasive organisms [1]. A similar situation was found in the United Kingdom, where

researchers compared the fish assemblage in a pond with a low-trophic-level invasive cyprinid (Pseudorasbora parva) composing > 99% of fish present to that in another pond without the cyprinid. They reported a reduction in the δ15N values of piscivorous fish and a mean reduction in the δ15N of the complete fish community [100]. Further studies are needed to investigate the effect of different types of land management, as well as factors that indicate the ecological integrity of communities, on the ability of communities to resist or facilitate the invasion of exotic species and their interactions with native predators.

Supporting information

S1 Table.
(DOCX)

S1 Dataset.
(CSV)

S2 Dataset.
(CSV)

Acknowledgments

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Conceptualization: Diego Juarez-Sanchez.

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Formal analysis: Diego Juarez-Sanchez.

Investigation: Diego Juarez-Sanchez.

Methodology: Diego Juarez-Sanchez.

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Resources: Diego Juarez-Sanchez, John G. Blake, Eric C. Hellgren.

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Validation: Diego Juarez-Sanchez, John G. Blake, Eric C. Hellgren.

Visualization: Diego Juarez-Sanchez.

Writing – original draft: Diego Juarez-Sanchez.

Writing – review & editing: John G. Blake, Eric C. Hellgren.

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