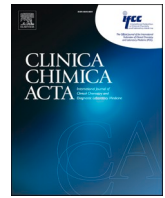




Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Answer to the letter of Lippi & Plebani entitled “Not all SARS-CoV-2 IgG and neutralizing antibody assays are created equal”

ARTICLE INFO

Keywords

SARS-CoV-2
Neutralizing antibody
anti-S-RBD
BNT162b2

Dear Editor,

We really appreciate the interest of Lippi and Plebani [1] for our recent paper, which offers us the opportunity to provide some additional clues. The primary aim of our study was to collect data on the long-term kinetics of binding and functional anti-SARS-CoV-2 antibodies in a cohort of BNT162b2-vaccinated health care workers (HCW) over an observation period of 6 months post-vaccination. The main findings were the not unexpected decline of binding anti-RBD antibodies and the lack of a consistent and time-independent correlation between anti-RBD IgG levels and neutralizing bioactivity, which settled at high level from T2 (50 days post-1st dose administration) thereafter with a non significant declining trend between T2 and T3 (6 months post-1st dose administration). Lippi and Plebani pointed out that a systematic lack of accuracy in surrogate virus neutralization assay (sVNT) platforms compared to conventional ones should prevent us from concluding that the absolute anti-RBD level is not a reliable proxy of neutralizing bioactivity in BNT162b2-vaccinated individuals.

We certainly agree that sVNT platforms should not replace live virus neutralization assays as the gold standard for identifying and titering neutralizing antibodies to SARS-CoV-2 wild-type and variant strains, as we also admitted as a limitation of our study. However, we hint that the accuracy of these tests and their correlations relies on the clinical setting to which they are applied. Indeed, despite the existence of a WHO International Standard for both binding and neutralizing assays [2], it is not yet clear how the clinical setting (COVID-19 vaccinated vs. experienced patients) and the emergence of variants of concern could undermine the harmonization process between these different assay platforms under real-world conditions [3,4]. sVNT platforms, by investigating the wild-type RBD-induced neutralizing activity only, are likely to underestimate the neutralizing potency of serum from COVID-19 experienced individuals, especially when VOC are involved [5,6]. On the other hand, as repeatedly shown by comparative analyses in vaccinated cohorts [7–9], the diagnostic reliability of sVNT seems to be retained when assessing vaccine-induced immune-responses, provided that the mRNA sequence of the vaccine RBD matches that of the recombinant RBD in the binding and competition assays. The cross-sectional studies cited by Lippi and Plebani [5,6] quantified humoral immunogenicity markers in

two cohorts of convalescent COVID-19 patients while the cohort studied by Bayart et al. [10] is as well affected by a large number (31%) of individuals with a positive serological scrutiny for SARS-CoV-2 infection before vaccination. In fact, the primary aim of the study by Meyer et al. [5] was the diagnostic value and comparability of different immune assay platforms for a diagnosis of SARS-CoV-2 infection rather than the long-term monitoring of the immune response to a spike-based vaccine. Notably, the very same study of Sholukh et al. [6] showed that the correlations between the sVNT, on the one side, and 4 live virus/pseudovirus neutralization assays, on the other, are much higher ($r = 0.73–0.8$) when the percentage of neutralization (ND_{50} in the original manuscript) is considered instead of the ND_{50} titer reported by Lippi and Plebani. Further on, all cited studies did not assess whether patients were infected by the Wuhan original strain or a variant strain, which, for the above reasons, could affect inter-assays harmonization. As to the study by Bayart et al. [10], which showed a much faster decline in neutralizing antibody titers using a live VNT than we assessed with our sVNT, available literature has so far provided contrasting results about long-term decay of circulating antibodies after mRNA vaccination, with some studies providing evidence for persistent and sustained neutralizing bioactivity of serum up to 6 months from vaccination [11,12]. It's our opinion that such discrepancies are resulting from multiple factors, including but not limited to inter-assays inconsistencies, such as previous exposure to cross-reacting common cold coronaviruses, age, history of previous COVID-19, sex, and other comorbidities in vaccinated cohorts.

We would also like to remark that in our study we did not disclaim the existence of a correlation between anti-RBD IgG levels and neutralizing bioactivity in vaccinated individuals nor that the high neutralizing activity we assessed at 6 months is necessarily protective, since the 1 month-apart swab-based monitoring protocol we use might have missed asymptomatic carriers. Actually, we observed an increasing strength of correlations from T2 and T3 despite a declining titer of anti-RBD IgG, likely as a result of affinity maturation of IgG by somatic hypermutation of spike/RBD-specific B cells [13–15]. This temporal trend has already been tracked after SARS-CoV-2 vaccination [12] and implies that an absolute conversion factor between anti-RBD IgG levels

<https://doi.org/10.1016/j.cca.2021.12.021>

Received 21 December 2021; Accepted 21 December 2021

Available online 25 December 2021

0009-8981/© 2021 Elsevier B.V. All rights reserved.

and neutralizing titers would be unreliable. As opposite, the studies cited by Lippi and Plebani [16,17] were not designed to provide data on anti-RBD IgG and neutralizing activity and their correlation on the long-term, when divergent kinetics may become apparent. Should a neutralizing antibodies absolute threshold of protection ever become available (hopefully using the WHO International Standard), implementation of the anti-RBD IgG level as a time-independent standalone correlate of protection may confuse risk stratification [14].

We are grateful to Lippi and Plebani for their appropriate remarks, which gave us the opportunity to better explain and contextualize our findings, but, for the above reasons, we believe the arguments and studies they brought about would not impair our findings. On the contrary, we believe that sVNT are user-friendly, high-throughput, standardized [18] immune-assays that may represent invaluable tools when implemented in the appropriate clinical and epidemiological context.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- [1] G. Lippi, M. Plebani, Not all SARS-CoV-2 IgG and neutralizing antibody assays are created equal.
- [2] P.A. Kristiansen, M. Page, V. Bernasconi, G. Mattiuzzo, P. Dull, K. Makar, S. Plotkin, I. Knezevic, WHO International Standard for anti-SARS-CoV-2 immunoglobulin, *Lancet* 397 (10282) (2021) 1347–1348.
- [3] M. Infantino, M. Pieri, M. Nuccetelli, V. Grossi, B. Lari, F. Tomassetti, G. Calugi, S. Pancani, M. Benucci, P. Casprini, M. Manfredi, S. Bernardini, The WHO International Standard for COVID-19 serological tests: towards harmonization of anti-spike assays, *Int. Immunopharmacol.* 100 (2021) 108095, <https://doi.org/10.1016/j.intimp.2021.108095>.
- [4] D. Giavarina, M. Carta, Improvements and limits of anti SARS-CoV-2 antibodies assays by WHO (NIBSC 20/136) standardization, *Diagnosis (Berl.)* (2021), <https://doi.org/10.1515/dx-2021-0126>. Epub ahead of print. PMID: 34851563.
- [5] B. Meyer, J. Reimerink, G. Torriani, F. Brouwer, G.-J. Godeke, S. Yerly, M. Hoogerwerf, N. Vuilleumier, L. Kaiser, I. Eckerle, C. Reusken, Validation and clinical evaluation of a SARS-CoV-2 surrogate virus neutralisation test (sVNT), *Emerg. Microbes. Infect.* 9 (1) (2020) 2394–2403.
- [6] A.M. Sholukh, A. Fiore-Gartland, E.S. Ford, M.D. Miner, Y.J. Hou, L.V. Tse, H. Kaiser, H. Zhu, J. Lu, B. Madarampalli, A. Park, F.A. Lempp, R. St. Germain, E. L. Bossard, J.J. Kee, K. Diem, A.B. Stuart, P.B. Rupert, C. Brock, M. Buerger, M. K. Doll, A.K. Randhawa, L. Stamatatos, R.K. Strong, C. McLaughlin, M.-L. Huang, K. R. Jerome, R.S. Baric, D. Montefiori, L. Corey, A.M. Caliendo, Evaluation of Cell-Based and Surrogate SARS-CoV-2 Neutralization Assays, *J. Clin. Microbiol.* 59 (10) (2021), <https://doi.org/10.1128/JCM.00527-21>.
- [7] F. Neumann, R. Rose, J. Römpke, O. Grobe, T. Lorentz, H. Fickenscher, A. Krumbholz, Development of SARS-CoV-2 Specific IgG and Virus-Neutralizing Antibodies after Infection with Variants of Concern or Vaccination, *Vaccines (Basel)* 9 (7) (2021) 700, <https://doi.org/10.3390/vaccines9070700>.
- [8] J. Favresse, C. Gillot, L. Di Chiaro, C. Eucher, M. Elsen, S. Van Eeckhoudt, C. David, L. Morimont, J.-M. Dogné, J. Douxfils, Neutralizing Antibodies in COVID-19 Patients and Vaccine Recipients after Two Doses of BNT162b2, *Viruses* 13 (7) (2021) 1364, <https://doi.org/10.3390/v13071364>.
- [9] K.-H. Chan, K.-Y. Leung, R.-R. Zhang, D. Liu, Y. Fan, H. Chen, K.-Y. Yuen, I.-N. Hung, Performance of a Surrogate SARS-CoV-2-Neutralizing Antibody Assay in Natural Infection and Vaccination Samples, *Diagnostics (Basel)* 11 (10) (2021) 1757, <https://doi.org/10.3390/diagnostics11101757>.
- [10] J.-L. Bayart, J. Douxfils, C. Gillot, C. David, F. Mullier, M. Elsen, C. Eucher, S. Van Eeckhoudt, T. Roy, V. Gerin, G. Wieers, C. Laurent, M. Closset, J.-M. Dogné, J. Favresse, Waning of IgG, Total and Neutralizing Antibodies 6 Months Post-Vaccination with BNT162b2 in Healthcare Workers, *Vaccines (Basel)* 9 (10) (2021) 1092, <https://doi.org/10.3390/vaccines9101092>.
- [11] N. Doria-Rose, M.S. Suthar, M. Makowski, S. O'Connell, A.B. McDermott, B. Flach, J.E. Ledgerwood, J.R. Mascola, B.S. Graham, B.C. Lin, S. O'Dell, S.D. Schmidt, A. T. Widge, V.-V. Edara, E.J. Anderson, L. Lai, K. Floyd, N.G. Roupael, V. Zarnitsyna, P.C. Roberts, M. Makhene, W. Buchanan, C.J. Luke, J.H. Beigel, L. A. Jackson, K.M. Neuzil, H. Bennett, B. Leav, J. Albert, P. Kunwar, mRNA-1273 Study Group. Antibody Persistence through 6 Months after the Second Dose of mRNA-1273 Vaccine for Covid-19, *N. Engl. J. Med.* 384 (23) (2021) 2259–2261.
- [12] E.G. Levin, Y. Lustig, C. Cohen, R. Fluss, V. Indenbaum, S. Amit, R. Doolman, K. Asraf, E. Mendelson, A. Ziv, C. Rubin, L. Freedman, Y. Kreiss, G. Regev-Yochay, Waning Immune Humoral Response to BNT162b2 Covid-19 Vaccine over 6 Months, *N. Engl. J. Med.* 385 (24) (2021) e84, <https://doi.org/10.1056/NEJMoa2114583>.
- [13] F. Struck, P. Schreiner, E. Staschik, et al., Vaccination versus infection with SARS-CoV-2: Establishment of a high avidity IgG response versus incomplete avidity maturation, *J. Med. Virol.* 93 (2021) 6765–6777.
- [14] K.P. Bliden, T. Liu, D. Sreedhar, et al., Evolution of Anti-SARS-CoV-2 IgG Antibody and IgG Avidity Post Pfizer and Moderna mRNA Vaccinations, *medRxiv* (2021), <https://doi.org/10.1101/2021.06.28.21259338>.
- [15] R.R. Goel, M.M. Painter, S.A. Apostolidis, D. Mathew, W. Meng, A.M. Rosenfeld, K. A. Lundgreen, A. Reynaldi, D.S. Khoury, A. Pattekar, S. Gouma, L. Kuri-Cervantes, P. Hicks, S. Dysinger, A. Hicks, H. Sharma, S. Herring, S. Korte, A.E. Baxter, D. A. Oldridge, J.R. Giles, M.E. Weirick, C.M. McAllister, M. Awofolaju, N. Tanenbaum, E.M. Drapeau, J. Dougherty, S. Long, K. D'Andrea, J.T. Hamilton, M. McLaughlin, J.C. Williams, S. Adamski, O. Kuthuru, I. Frank, M.R. Betts, L. A. Vella, A. Grifoni, D. Weiskopf, A. Sette, S.E. Hensley, M.P. Davenport, P. Bates, E.T. Luning Prak, A.R. Greenplate, E.J. Wherry, mRNA vaccines induce durable immune memory to SARS-CoV-2 and variants of concern, *Science* 374 (6572) (2021), <https://doi.org/10.1126/science.abm0829>.
- [16] C. Irsara, A.E. Egger, W. Prokop, et al., Clinical validation of the Siemens quantitative SARS-CoV-2 spike IgG assay (sCOVG) reveals improved sensitivity and a good correlation with virus neutralization titers, *Clin. Chem. Lab Med.* 59 (2021) 1453–1462.
- [17] A. Padoan, C. Cosma, F. Bonfante, F.D. Rocca, F. Barbaro, C. Santarossa, L. Dall'Olmio, M. Pagliari, A. Bortolami, A. Cattelan, V. Cianci, D. Basso, M. Plebani, SARS-CoV-2 neutralizing antibodies after one or two doses of Comirnaty (BNT162b2, BioNTech/Pfizer): Kinetics and comparison with chemiluminescent assays, *Clin. Chim. Acta* 523 (2021) 446–453.
- [18] F. Zhu, T. Althaus, C.W. Tan, A. Costantini, W.N. Chia, N. Van Vinh Chau, L.e. Van Tan, G. Mattiuzzo, N.J. Rose, E. Voiglio, L.-F. Wang, WHO international standard for SARS-CoV-2 antibodies to determine markers of protection, *Lancet Microbe.* (2021), [https://doi.org/10.1016/S2666-5247\(21\)00307-4](https://doi.org/10.1016/S2666-5247(21)00307-4).

Giacomo Malipiero, Danilo Villalta*

Immunologia e Allergologia, Ospedale Santa Maria degli Angeli, Pordenone, Italy

* Corresponding author at: Immunologia e Allergologia, Ospedale S. Maria degli Angeli, Pordenone, Via Montereale 24, 33170 Pordenone, Italy.

E-mail address: danilo.villalta@asfo.sanita.fvg.it (D. Villalta).