



REVIEW

Promising neuroprotective strategies for traumatic spinal cord injury with a focus on the differential effects among anatomical levels of injury [version 1; referees: 2 approved]

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Abstract

Traumatic spinal cord injury (SCI) is a devastating condition of motor, sensory, and autonomic dysfunction. The significant cost associated with the management and lifetime care of patients with SCI also presents a major economic burden. For these reasons, there is a need to develop and translate strategies that can improve outcomes following SCI. Given the challenges in achieving regeneration of the injured spinal cord, neuroprotection has been at the forefront of clinical translation. Yet, despite many preclinical advances, there has been limited translation into the clinic apart from methylprednisolone (which remains controversial), hypertensive therapy to maintain spinal cord perfusion, and early decompressive surgery. While there are several factors related to the limited translational success, including the clinical and mechanistic heterogeneity of human SCI, the misalignment between animal models of SCI and clinical reality continues to be an important factor. Whereas most clinical cases are at the cervical level, only a small fraction of preclinical research is conducted in cervical models of SCI. Therefore, this review highlights the most promising neuroprotective and neural reparative therapeutic strategies undergoing clinical assessment, including riluzole, hypothermia, granulocyte colony-stimulating factor, glibenclamide, minocycline, Cethrin (VX-210), and anti-Nogo-A antibody, and emphasizes their efficacy in relation to the anatomical level of injury. Our hope is that more basic research will be conducted in clinically relevant cervical SCI models in order to expedite the transition of important laboratory discoveries into meaningful treatment options for patients with SCI.

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Introduction

Traumatic spinal cord injury (SCI), which is caused by external mechanical impact, results in impairment of motor, sensory, and autonomic function at and below the level of injury. Mechanical laceration, contusion, and compression result in cell death, which is further propagated by secondary injury mechanisms which include ischemia, sodium- and calcium-mediated cell injury, glutamatergic excitotoxicity, hemorrhage, and inflammation. The secondary injury amplifies the primary damage and promotes cystic degeneration and glial scar formation, thereby preventing functional recovery. Therefore, targeting secondary injury is a promising therapeutic intervention.

Despite several efficacious preclinical studies for SCI, there have been challenges in achieving successful translation into the clinic. While the disconnect between bench and bedside is not limited to SCI, it is important to recognize the underlying factors and identify solutions. Clinical heterogeneity, complexity of the disease, and the limited regenerative capacity of the spinal cord are among the key causes for poor translation and have been broadly discussed in the literature¹. Yet significantly less emphasis has been placed on the need to apply clinically relevant models of cervical SCI². Given that over 50% of human SCI cases occur at the cervical level³ and the majority of preclinical work involves thoracic injuries (Table 1), translation will require a greater understanding of injury-level subpopulation differences in pathophysiology and therapeutic benefits.

Table 1. Experimental evidence for the efficacy of promising neuroprotective therapies.

Neuroprotective/neural reparative therapy	Injury model, species	Reference
Riluzole	Contusion, T7–T10, rat	69
	Compression, T8, rat	70
	Compression, T6, rat	71
	Contusion, T10, rat	72
	Compression, T11, rat	73
	Contusion, T8, rat	74
	Unilateral contusion, C7, rat	75
	Hemisection, C2, rat	35
	Compression, C7, rat	76
	Unilateral contusion, C7, rat	77
	Compression, C7, rat	78
	Compression, C7, rat	79
	Transection, S2, rat	34
	Hypothermia	Contusion, T8, rat
Compression, T8, rat		81
Compression, T8, rat		82
Compression, T11, rat		83
Contusion, T9, rat		84
Contusion, T10, rat		85
Unilateral contusion, C7, rat		75
Contusion, C5, rat		86

Neuroprotective/neural reparative therapy	Injury model, species	Reference
Glibenclamide	Contusion, T8, rat	87
	Unilateral contusion, T9, mouse	88
	Unilateral contusion, C7, rat	75
	Unilateral contusion, C7, rat	89
	Contusion, C7, rat	90
	Unilateral contusion, C4, rat	42
	Unilateral contusion, C7, rat	77
	Unilateral contusion, C7, rat	91
Granulocyte colony-stimulating factor	Contusion, T10, rat	92
	Compression, T9, rat	93
	Contusion, T9, rat	94
	Hemisection, T10, mouse	95
	Contusion, T8, rat	96
	Contusion, T9, rat	97
	Contusion, T8, rat	44
	Compression, T8, mouse	98
	Compression, T7, mouse	99
	Transection, T8, mouse	100
	Contusion, T8, rat	101
Minocycline	Compression, T8, rat	102
	Contusion, T7, rat	103
	Contusion, T9, rat	104
	Contusion, T9, mouse	105
	Contusion, T9, rat	106
	Contusion, T9, rat	107
	Hemisection, T13, rat	108
	Contusion, T10, rat	109
	Contusion, T9, rat	110
	Contusion, T10, rat	111
	Contusion, T9, rat	112
	Dorsal transection, C7, rat	113
	Unilateral contusion, C5, rat	114
	Compression, T3, mouse	115
	Cethrin (VX-210)	Contusion, T8, mouse
Dorsal hemisection, T7, mouse		117
Dorsal transection, T3, rat		118
Contusion, T9, rat		119
Anti-Nogo-A antibody	Hemisection, T10, rat	120
	Dorsolateral hemisection, T8, rat	121
	T-shape transection, T9, rat	122
	Partial hemisection, T8, monkey	123
	T-shape transection, T8, rat	124
	T-shape transection, T8, rat	125
	T-shape transection, T8, rat	126
	Dorsal hemisection, T8, rat	52
	Partial dorsal transection, T6, rat	53
	Partial hemisection, C7, monkey	54
	Hemisection, C7, monkey	55

The table summarizes the model, anatomical level of spinal cord injury, and the species used to evaluate the effectiveness and mechanisms of action of the neuroprotective therapies undergoing clinical trials. Although this list is not exhaustive, it highlights that thoracic models of spinal cord injury are most commonly applied at the preclinical level. All injury models are bilateral if not stated otherwise.

Differences between the cervical and thoracic cord anatomy, physiology, and immune response may affect the outcome of neuroprotective treatments (Figure 1). Anatomically, the cervical spine has small vertebrae and increased mobility, which make it more susceptible to injury compared to the thoracic region. The cervical spinal cord also has a larger diameter, a greater blood supply, and larger gray and white matter areas⁴. Relatedly, the cervical gray matter vasculature has less pericyte coverage than the thoracic cord, resulting in a blood spinal cord barrier predisposed to increased permeability^{5,6}. Also, some of the most frequent conditions of the spine, such as central cord syndrome⁷ and degenerative cervical myelopathy⁸, affect primarily the cervical region. Of note, while the cervical spinal cord is especially vulnerable to injury and hemorrhage, it may also be more accessible to systemically administered therapeutics. Adding to this complexity, there is emerging evidence demonstrating level-dependent variations in the immune response^{9,10}. For example, interestingly, higher-level injuries may be less prone to chronic autoimmunity^{11,12}. Therefore, as SCI pathophysiology may differ between anatomical levels of injury, there is growing awareness that treatments should be tailored to the patient's injury. Here, we review the most promising neuroprotective approaches, emphasizing their effect differences based on the level of injury (Table 2).

Neuroprotective strategies in current care

Early surgical decompression

The popularized phrase coined by the senior author, "Time is spine", highlights the preclinical^{13,14} and clinical^{15,16} success of early surgical decompression, which aims to realign the spinal column

and relieve bony or ligamentous spinal cord compression. Decompression of intradural pressure, by durotomy alone or durotomy combined with duraplasty, has also been evaluated in experimental SCI^{17,18}. Yet mixed results warrant further research on the efficacy and standardization of these practices. In contrast, early extradural surgical decompression has been shown to reduce tissue damage and improve outcomes following SCI. Even with some concern regarding perioperative hemodynamic changes affecting cord perfusion, most spine surgeons have been, and continue to be, in favor of decompressing the acutely injured spinal cord^{19,20}. As a result, early decompression remains recommended in clinical management guidelines by the American Association of Neurological Surgeons (AANS) and the Congress of Neurological Surgeons²¹. Similarly, the recent AOSpine guideline also recommends decompression within 24 hours of SCI²². While there may be differential efficacy between injury-level subpopulations²³, current evidence is limited by substantial clinical heterogeneity, loss to follow-up, unclear adjustment for baseline factors, and a lack of statistical power. For these reasons, more work is needed to develop customizable treatment regimens and prioritized surgical access to the most benefiting patient subtypes.

Support of mean arterial pressure

Hypotension, hypoxemia, pulmonary dysfunction, and cardiovascular instability are common within the first 7 to 10 days of SCI²⁴. Hemodynamic instability not only limits the opportunity for early surgical intervention but also increases spinal cord ischemia and therefore secondary damage. For this reason, the current AANS and Congress of Neurological Surgeons guideline recommends continuous hemodynamic monitoring, interventions

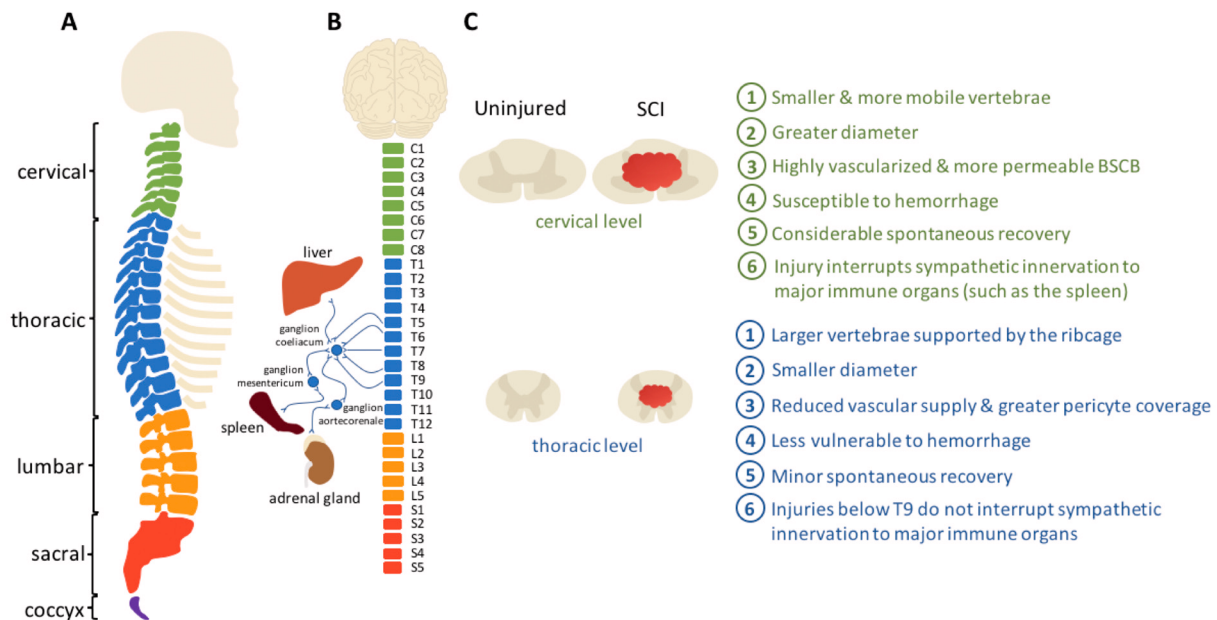


Figure 1. There are several key differences between cervical and thoracic spinal cord injury. (A) The cervical vertebrae are smaller and more mobile than their thoracic counterparts, which are further supported by the rib cage. (B) The cervical spinal cord also has a larger diameter, and injuries at the cervical level interrupt the sympathetic innervation to major immune organs. (C) Moreover, the greater vascularity of the cervical cord increases susceptibility to hemorrhage following trauma. Lastly, injuries at the cervical level allow for considerably more spontaneous recovery compared with injuries at the thoracic level¹²⁸. BSCB, blood spinal cord barrier; SCI, spinal cord injury.

Table 2. Neuroprotective strategies currently in clinical trials.

Neuroprotective/ neural-reparative drug	ClinicalTrials.gov identifier	Status	Enrollment (number of patients, level of injury)		Results	Mechanism of action	Reference
			Thoracic	Cervical			
Riluzole	NCT00876889	Completed	8 (T1–T11)	28 (C4–C8)	Motor score improvement in patients with cervical SCI, particularly with incomplete injuries. No significant effect in patients with thoracic SCI.	Limit excitotoxicity	36
	NCT01597518	Recruiting	0	Est. enrollment 351 (C4–C8)			37
Therapeutic hypothermia	N/A	Completed	0	14 (C4–C7)	Trend toward improvement of motor scores compared with historical controls in the same institution.	Reduce excitotoxicity, inflammation, and vasogenic edema	40
	NCT02991690	Recruiting	0	Est. 120 (C1–C8)		Limit hemorrhage	ClinicalTrials.gov
Glibenclamide	NCT02524379	Recruiting	0	Est. 10 (C2–C8)		Limit hemorrhage	ClinicalTrials.gov
	N/A	Completed	0	28 (C2–C6)	Motor score improvement at 3 months of treatment compared with MPSS historical controls.	Promote neurogenesis and angiogenesis, and reduce inflammation	46
Granulocyte colony- stimulating factor	N/A	Completed	0	17 (C2–C6)	Motor score improvement from 1 week to 1 year after SCI compared with placebo in an open non-randomized trial.		47
	NCT00559494	Completed	17 (T1–T12)	25 (C1–C8)	Motor score improvement in patients with cervical SCI, particularly for motor incomplete. No benefit for patients with thoracic SCI.	Reduce inflammation	49
Minocycline	NCT01828203	Recruiting	0	Est. 248 (C0–C8)			ClinicalTrials.gov
	NCT00500812	Completed	32 (T2–T12)	16 (C4–T1)	Significant motor score improvement in patients with cervical injury. No effect in patients with thoracic SCI.	Inhibit axonal dieback and reduce inflammation	50
Anti-Nogo-A antibody	NCT02669849	Recruiting	0	Est. 150 (C5–C6)			ClinicalTrials.gov
	NCT00406016	Completed	52 patients with injury between C5 and T12; no data available about injury level classification		No adverse effects.	Promote neurite sprouting	127

The table lists the discussed neuroprotective strategies for spinal cord injury (SCI) undergoing clinical evaluation. The status of trials and enrollment information, including level of injury and results, are summarized. This demonstrates that clinical trials are predominately focused on cervical SCI.

Est, estimated; N/A, not applicable; MPSS, methylprednisolone sodium succinate.

correcting hypotension (such as by vasopressor administration), and maintenance of mean arterial blood pressure (MAP) between 85 and 90 mmHg for the first 7 days following cervical injury²⁵. While these recommendations are largely based on a small group of uncontrolled and underpowered studies²⁶, a recent retrospective assessment largely confirmed the published guidelines as well as the neuroprotective potential of vasopressor administration²⁷. As these results need further prospective validation, it will be important to stratify patient populations and identify potential treatment effect differences based on the anatomical level of injury. It is also important to note that MAP support principally aims to maintain appropriate spinal cord perfusion pressure (SCPP), determined by the difference between MAP and intraspinal pressure (ISP). However, as ISP may increase independently of MAP, maintenance of a low ISP or a high SCPP (or both) is gaining increasing attention as an important practice in the acute clinical management of SCI^{28,29}. While initial studies have shown encouraging results about the predictive value of low ISP or high SCPP in neurological recovery, larger multicenter studies are needed to validate these preliminary data²⁹.

Methylprednisolone sodium succinate

Methylprednisolone sodium succinate (MPSS) is a synthetic corticosteroid with potent anti-inflammatory effects and neuroprotective potential in acute traumatic SCI. Concerns about increased risk for infections following MPSS treatment have kept the drug at the forefront of continuous controversy. While it remains the only treatment option for acute SCI, debate regarding optimal dose, time of administration, efficacy, and adverse effects has dominated the field for decades and has dichotomized clinicians around the world. For this reason, there have been three National Acute Spinal Cord Injury Studies (NASCIS) to evaluate the clinical safety and efficacy of varying MPSS dose and timing. Moreover, NASCIS results have been retrospectively analyzed on numerous occasions to derive meaningful conclusions. One of the most recent publications on the topic concluded that MPSS does not increase the risk of infections and confers significant short-term effects when given within the first 8 hours of injury³⁰. Importantly, patients with cervical SCI and reduced baseline injury severity seem to benefit most from this treatment³¹. Given the particularly debilitating nature of cervical injuries, these improvements have tremendous impact on patients' quality of life. Thus, the most recent AOSpine guideline currently recommends a 24-hour treatment of intravenous MPSS when initiated within the first 8 hours of SCI, independently of injury level²².

Promising neuroprotective and neural reparative therapies in clinical trials

Riluzole

Secondary injury involves ionic dysregulation and excitotoxicity. As cell membranes become highly permeable to sodium ions, there is increased calcium influx. Subsequently, high sodium and calcium ion concentrations in neurons trigger the secretion of glutamate from nerve terminals. Increased synaptic glutamate leads to prolonged excitability in the postsynaptic neurons, driving eventual neuronal edema and death.

Riluzole is a benzothiazole (molecular weight of 234.2 Da) which inhibits voltage-gated sodium channels and glutamate release, thereby mitigating excitotoxicity. Riluzole has been reported to slow the progression of amyotrophic lateral sclerosis (ALS)—a progressive motor neuron disease—and currently is the only US Food and Drug Administration (FDA)-approved drug for ALS. In addition, riluzole has been shown to have neuroprotective potential in animal models of Parkinson's disease³² and Huntington's disease³³ and currently is being used in clinical studies of mild Alzheimer's disease (ClinicalTrials.gov identifier NCT01703117). Importantly, riluzole was also shown to suppress spasticity³⁴, a frequent co-morbidity in patients with SCI, and to promote neural preservation in rats with high cervical spinal hemisection injury³⁵.

Capitalizing on the preclinical success of riluzole, a phase I/IIA clinical trial was launched in April 2010 to assess the safety and pharmacokinetics of riluzole in patients with acute traumatic SCI (ClinicalTrials.gov identifier NCT00876889). In this trial, 36 patients with SCI (28 cervical and eight thoracic SCI) received riluzole (50 mg) orally every 12 hours for 28 doses. A control group consisting of 36 patients with SCI—matched for neurological impairment, gender, and age—received the standard of care but no riluzole. Patients who received riluzole showed statistically significant improvement compared with the control group ($P = 0.021$). In particular, patients with incomplete cervical injury—American Spinal Injury Association (ASIA) Impairment Scale B—showed the highest improvement in the International Standards for Neurological Classification of Spinal Cord Injury (ISNCSCI) motor score ($P = 0.037$). In addition to being efficacious, riluzole was shown to be safe for this patient cohort³⁶. Interestingly, there was no difference within the thoracic injury group, as patient numbers were small, patients had more severe injuries, and the ISNCSCI motor scoring is less sensitive to thoracic recovery³⁷.

Based on these results, a phase IIB/III was launched in 2013 to evaluate the efficacy and safety of riluzole in patients with cervical traumatic SCI, entitled "Riluzole in Acute SCI Study" (RISCIS) (estimated enrollment: 351 patients, ClinicalTrials.gov identifier NCT01597518). In this multicenter, randomized, placebo-controlled, double-blinded trial, riluzole (100 mg, twice daily) is administered orally to patients within 24 hours from injury, followed by two 50 mg daily doses for 14 days after SCI. A capsule identical in shape and size to riluzole is administered to patients in the control group. The primary outcome of the study is improvement in ISNCSCI motor scores at 180 days after injury. The study is estimated to be completed by 2021³⁷.

Therapeutic hypothermia

In response to trauma, increased metabolic rate can lead to excitotoxicity and cell death. Local or systemic cooling following insult has been shown to reduce the metabolic demand, thereby limiting cell death. Moreover, therapeutic hypothermia has been shown to reduce inflammatory cell infiltration, myeloperoxidase activity, and vasogenic edema and stabilize the blood-brain barrier³⁸. Despite these benefits, systemic hypothermia may have some serious side effects, including bradycardia, respiratory infections, and deep vein

thrombosis. While local cooling of the spinal cord circumvents many of these concomitant issues, randomized controlled trials are still needed to prove the efficacy of local hypothermia in neurological recovery after SCI. However, in one study, acute (within 8 hours of injury) local hypothermia was shown to improve recovery among cervical and thoracic populations (n = 12 out of 14 cervical SCI, n = 4 out of 6 thoracic SCI) when compared with historical controls³⁹. Similarly, a pilot study of systemic hypothermia in patients with cervical complete SCI (n = 14) demonstrated fewer adverse effects and a trend toward improved recovery compared with age- and injury level-matched historical controls when systemic hypothermia was induced within 9 hours of trauma⁴⁰. Furthermore, a follow-up randomized controlled trial assessing the efficacy of intravascularly delivered systemic hypothermia in acute cervical SCI commenced in May 2017 (estimated enrollment: 120 patients, ClinicalTrials.gov identifier NCT02991690). Although previous multicenter randomized clinical trials found hypothermia to be ineffective in adults with traumatic brain injury⁴¹, expectations for SCI remain hopeful.

Glibenclamide (Glyburide, DiaBeta)

Capillary fragmentation following SCI contributes to hemorrhage. This process is initiated in the capillary-rich gray matter of the injury epicenter and expands rostro-caudally, leading to progressive tissue necrosis, cavitation, and neurological dysfunction. In a rat model of unilateral cervical SCI, Simard *et al.* found that sulfonylurea receptor 1 (SUR1)-regulated Ca^{2+} -activated $[\text{ATP}]_i$ -sensitive non-specific cation ($\text{NC}_{\text{Ca-ATP}}$) channels of the capillary endothelium in the spinal cord are key to capillary fragmentation following SCI⁴². By blocking $\text{NC}_{\text{Ca-ATP}}$ channels with the FDA-approved anti-diabetic drug glibenclamide (Glyburide), Simard *et al.* observed decreased lesion volumes and significant white matter preservation coupled with improved neurobehavioral outcomes⁴². Recently, a phase I/II clinical trial was initiated to assess the safety and neuroprotective effectiveness of Glyburide (DiaBeta) in patients with acute traumatic cervical SCI (estimated enrollment: 10 patients, ClinicalTrials.gov identifier NCT02524379), with an estimated completion date in early 2020.

Granulocyte colony-stimulating factor

Initially overlooked for its potential in the central nervous system (CNS), granulocyte colony-stimulating factor (G-CSF) has shown positive preclinical results for SCI (Table 1). In response to ischemia and CNS injury, G-CSF and its receptor (CD114; G-CSFR) are upregulated in neurons and endogenous stem cells, initiating a compensatory neuroprotective mechanism. By binding to its cognate receptor, G-CSF counteracts programmed cell death in mature neurons, induces neurogenesis, and promotes neuronal differentiation of adult neural stem cells⁴³. Moreover, angiogenesis⁴⁴ and reduced inflammation⁴⁵ have been attributed to the protective actions of G-CSF. Kamiya *et al.* administered G-CSF for 5 consecutive days after cervical SCI and assessed ASIA motor scores 3 months later⁴⁶. The improvements were significant compared with historical controls of patients with cervical SCI receiving high-dose MPSS⁴⁶. In a study by Inada *et al.*, patients with cervical SCI who received G-CSF demonstrated improved recovery compared with a non-treated group⁴⁷. However, the treatment was administered in an open-label and non-randomized fashion⁴⁷. Interestingly, in a

study by Saberi *et al.*, in which G-CSF was administered in patients with chronic SCI, significant motor and sensory recovery was demonstrated, particularly in patients with incomplete cervical SCI⁴⁸. Despite these promising effects, a true double-blinded randomized control clinical trial for G-CSF has yet to be developed.

Minocycline

Inflammatory cytokines produced by resident microglia and astrocytes following trauma attract peripheral immune cells to the spinal cord. Neutrophils and monocytes are the first blood-derived cells to enter the injured parenchyma. While these cells are crucial in cleaning up the cellular debris, they produce inflammatory cytokines, such as tumor necrosis factor-alpha and interferon-gamma, as well as toxic by-products that exacerbate damage.

Minocycline is a tetracycline antibiotic with neuroprotective and anti-inflammatory properties. A single-center, placebo-controlled, double-blinded phase I/II clinical trial was initiated in 2004 to evaluate the efficacy and safety of intravenous minocycline within 12 hours of injury for 7 days. The study, which was completed in 2010 (27 patients received minocycline and 25 received placebo), showed a trend toward improved motor scores in incomplete cervical SCI cases in the absence of any serious adverse effects ($P = 0.05$) but no improvement in thoracic SCI⁴⁹ (ClinicalTrials.gov identifier NCT00559494). Based on these results, a phase III clinical trial, titled “Minocycline in Acute Spinal Cord Injury (MASC)”, was initiated in 2013 and is expected to finish by 2018 (estimated enrollment: 248 patients, ClinicalTrials.gov identifier NCT01828203). Interestingly, a clinical trial evaluating the efficacy of minocycline in reducing neuropathic pain has been successfully completed, but the results have yet to be published (ClinicalTrials.gov identifier NCT01869907). Given that neuropathic pain is a common and debilitating co-morbidity in patients with SCI, the study results will be of significant interest to the field.

Cethrin (VX-210)

The injured spinal cord niche contains growth-inhibitory molecules, such as myelin debris and chondroitin sulfate proteoglycans, that lead to neuron growth cone collapse, thereby inhibiting regeneration. These molecules bind to respective receptors on regenerating neurons, where they initiate a phosphorylation cascade. At the converging point of this cascade are Rho GTPases, a family of intracellular enzymes that regulate cytoskeletal mechanisms and cellular mobility. Cethrin (VX-210) is a recombinant deactivator of RhoA (a member of the Rho family) with dura and cell membrane penetrance. An open-label uncontrolled phase I/IIa clinical trial showed significant neurological improvement in patients with SCI who received Cethrin (48 patients, ClinicalTrials.gov identifier NCT00500812). Benefits were particularly enhanced in patients with cervical SCI compared with their thoracic counterparts⁵⁰, a finding that incentivized the initiation of larger controlled double-blinded clinical trials for patients with cervical SCI. This phase I/II clinical trial will evaluate the safety and efficacy of two doses of VX-210 (formerly known as Cethrin) compared with placebo (a fibrin sealant) when applied extradurally at the site of injury, acutely after cervical SCI (estimated enrollment: 150 patients, ClinicalTrials.gov identifier NCT02669849).

Anti-Nogo-A antibody (ATI-355)

Anti-Nogo-A antibody is a monoclonal antibody against Nogo-A, a protein inhibitor of neurite growth found on adult CNS myelin. Widely assessed at the thoracic level in rodents^{51–53}, in addition to several studies in primate models of cervical SCI^{54,55}, anti-Nogo-A antibody has been shown to promote axonal sprouting and improve functional recovery following injury. A non-randomized, open-label phase I clinical trial of humanized anti-Nogo-A antibody (ATI-355; Novartis Pharmaceuticals) was initiated to assess the feasibility, tolerability, and safety of either repeated intrathecal bolus injections of ATI-355 or continuous intrathecal delivery in acute SCI (4–14 days after injury). In total, 52 cervical and thoracic patients with traumas between C5 and T12 level were recruited in the study, and results are pending dissemination (ClinicalTrials.gov identifier NCT00406016). A phase IIb trial, led by Armin Curt, is expected to begin in Europe shortly.

Emerging neuroprotective approaches

Intravenous immunoglobulin G

Intravenous immunoglobulin G (IVIG) consists of serum immunoglobulin G (IgG) pooled from thousands of healthy donors. Independent laboratory studies in cervical and thoracic models of SCI have shown that IVIG improves recovery by targeting the detrimental inflammatory response in the spinal cord after trauma^{56–58}. While the efficacy of IVIG for SCI has not been assessed in clinical trials, the exciting preclinical results coupled with IVIG's long-term clinical use for the treatment of autoimmune and immunodeficiency conditions make it a promising candidate for SCI clinical trials.

Cell therapies

With cell transplantation as an attractive treatment approach for SCI, a diverse range of cells has been evaluated in preclinical studies, resulting in a plethora of potential mechanisms. In short, transplanted cells have been used for immune modulation, trophic support, scaffolding, re-myelination, and cell replacement⁵⁹. Yet, predominantly applied in the subacute and chronic phases of injury, only a few cell transplantation strategies are thought to have neuroprotective potential for the acutely injured spinal cord.

Mesenchymal stem/stromal cells. Mesenchymal stem/stromal cells (MSCs) are multipotent mesodermal progenitors defined by their *in vitro* adhesion to plastic and their cell surface antigen profile⁶⁰. Readily accessible from various adult tissues such as bone marrow, cartilage, and fat, MSCs are among the most commonly studied cells in regenerative medicine. This popularity has led to significant heterogeneity in MSC isolation, cultivation, and purification procedures, further resulting in mixed therapeutic efficacy among preclinical studies and the increasing number of clinical studies⁶¹. In SCI, MSCs have been reported to dampen inflammation, modulate the immune response, and secrete neuroprotective factors⁵⁹. A 2013 systematic meta-analysis of preclinical studies, involving intrathecal, intraparenchymal, and intravenous infusion of MSCs in various models of cervical and thoracic SCI, determined that the cells, overall, result in improved functional recovery after injury⁶². While this is encouraging, a lot remains to be understood about the identity and function of MSCs. Furthermore, additional mechanis-

tic studies are needed to effectively tailor their therapeutic application for SCI and identify differences in efficacy between anatomical levels of injury.

Final thoughts

Spinal cord level-dependent differences in vertebral structure, anatomy, and peripheral immune organ innervation may affect SCI pathophysiology. Though largely overlooked in preclinical studies, post hoc subgroup analysis from seminal large-scale clinical trials has been indicative of varying treatment efficacy between different anatomical levels of SCI^{50,63}. As a result, the stratification of patients into injury-level subpopulations is being increasingly adopted in trial design, and the aforementioned RISCIS trial is a leading example³⁷.

Notwithstanding its strengths, this approach has considerable challenges. Firstly, given the clinical heterogeneity of SCI pathophysiology, even within the same level of injury, the recruitment of adequate patient numbers to reach statistical power may prove substantially difficult, especially for acute and infrequent injuries. Although multicenter trials may circumvent this issue, they require tremendous coordination, collaboration, and resources. However, a recently published assessment of patient recruitment for acute SCI trials determined that such multicenter Canadian trials are feasible with careful *a priori* planning and registry support⁶⁴. Secondly, stratified analysis is susceptible to confounding effects. For instance, treatment efficacy may be directly affected by patient age or injury causation rather than the anatomical level of injury³. In addition, stratification according to injury level alone is unlikely to account for the significant variability in injury severity, presentation, and patient characteristics. Finally, a greater understanding of the impact of trauma-related mechanisms on the impact and outcomes of SCI is also required.

Therefore, it is increasingly recognized that optimal patient recovery will stem from a combinatorial treatment regimen of integrated pharmacological and rehabilitation-based strategies that will be personalized to their SCI signature (age, medical history, level, completeness, and mechanism of injury). For this reason, translational laboratory studies need to compare neuroprotective efficacy, as well as combinatorial approaches, between different anatomical levels of injury and severity. Moreover, advances in imaging and biochemical biomarkers are needed to help tailor trials within a heterogeneous SCI patient population, narrowing the inclusion window and increasing study power. These approaches can be further applied to better assess treatment efficacy, specifically beyond basic neurological recovery. Apart from outcomes, treatment protocols should ensure sufficient drug delivery to target sites, especially for systemically administered neuroprotective agents with CNS-specific effects⁶⁵. Moreover, coordinated efforts should be made by leading SCI care units to standardize early medical management^{66,67} and monitoring practices⁶⁸ in order to maximize the efficacy of randomized controlled trials. Lastly, with the increasing incidence of traumatic SCI in the elderly³, it is important that more emphasis be placed on optimizing such practices to address the specific needs of this growing demographic.

In conclusion, neuroprotection has the potential to improve recovery of motor, sensory, and autonomic function following SCI. Significant strides in our understanding of SCI pathophysiology, patient presentation, and biomarkers will further align preclinical research with clinical reality, yielding translatable solutions that can benefit patients.

Competing interests

The authors declare that they have no competing interests.

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References



- Blesch A, Tuszynski MH: **Spinal cord injury: plasticity, regeneration and the challenge of translational drug development.** *Trends Neurosci.* 2009; **32**(1): 41–7.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Kwon BK, Hillyer J, Tetzlaff W: **Translational research in spinal cord injury: a survey of opinion from the SCI community.** *J Neurotrauma.* 2010; **27**(1): 21–33.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Singh A, Tetreault L, Kalsi-Ryan S, *et al.*: **Global prevalence and incidence of traumatic spinal cord injury.** *Clin Epidemiol.* 2014; **6**: 309–31.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Kameyama T, Hashizume Y, Sobue G: **Morphologic features of the normal human cadaveric spinal cord.** *Spine (Phila Pa 1976).* 1996; **21**(11): 1285–90.
[PubMed Abstract](#) | [Publisher Full Text](#)
- F** Li Y, Lucas-Osma AM, Black S, *et al.*: **Pericytes impair capillary blood flow and motor function after chronic spinal cord injury.** *Nat Med.* 2017; **23**(6): 733–41.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- Winkler EA, Sengillo JD, Bell RD, *et al.*: **Blood-spinal cord barrier pericyte reductions contribute to increased capillary permeability.** *J Cereb Blood Flow Metab.* 2012; **32**(10): 1841–52.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- McKinley W, Santos K, Meade M, *et al.*: **Incidence and outcomes of spinal cord injury clinical syndromes.** *J Spinal Cord Med.* 2007; **30**(3): 215–24.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Nouri A, Tetreault L, Singh A, *et al.*: **Degenerative Cervical Myelopathy: Epidemiology, Genetics, and Pathogenesis.** *Spine (Phila Pa 1976).* 2015; **40**(12): E675–93.
[PubMed Abstract](#) | [Publisher Full Text](#)
- F** Brommer B, Engel O, Kopp MA, *et al.*: **Spinal cord injury-induced immune deficiency syndrome enhances infection susceptibility dependent on lesion level.** *Brain.* 2016; **139**(Pt 3): 692–707.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Lucin KM, Sanders VM, Jones TB, *et al.*: **Impaired antibody synthesis after spinal cord injury is level dependent and is due to sympathetic nervous system dysregulation.** *Exp Neurol.* 2007; **207**(1): 75–84.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Ibarra A, Jiménez A, Cortes C, *et al.*: **Influence of the intensity, level and phase of spinal cord injury on the proliferation of T cells and T-cell-dependent antibody reactions in rats.** *Spinal Cord.* 2007; **45**(5): 380–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Uldreaj A, Tzekou A, Mothe AJ, *et al.*: **Characterization of the Antibody Response after Cervical Spinal Cord Injury.** *J Neurotrauma.* 2017; **34**(6): 1209–26.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Batchelor PE, Wills TE, Skeers P, *et al.*: **Meta-analysis of pre-clinical studies of early decompression in acute spinal cord injury: a battle of time and pressure.** *PLoS One.* 2013; **8**(8): e72659.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Fehlings MG, Perrin RG: **The timing of surgical intervention in the treatment of spinal cord injury: a systematic review of recent clinical evidence.** *Spine (Phila Pa 1976).* 2006; **31**(11 Suppl): S28–35; discussion S36.
[PubMed Abstract](#) | [Publisher Full Text](#)
- F** Fehlings MG, Vaccaro A, Wilson JR, *et al.*: **Early versus delayed decompression for traumatic cervical spinal cord injury: results of the Surgical Timing in Acute Spinal Cord Injury Study (STASCIS).** *PLoS One.* 2012; **7**(2): e32037.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Furlan JC, Noonan V, Cadotte DW, *et al.*: **Timing of decompressive surgery of spinal cord after traumatic spinal cord injury: an evidence-based examination of pre-clinical and clinical studies.** *J Neurotrauma.* 2011; **28**(8): 1371–99.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Jalan D, Saini N, Zaidi M, *et al.*: **Effects of early surgical decompression on functional and histological outcomes after severe experimental thoracic spinal cord injury.** *J Neurosurg Spine.* 2017; **26**(1): 62–75.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Smith JS, Anderson R, Pham T, *et al.*: **Role of early surgical decompression of the intradural space after cervical spinal cord injury in an animal model.** *J Bone Joint Surg Am.* 2010; **92**(5): 1206–14.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Fehlings MG, Rabin D, Sears W, *et al.*: **Current practice in the timing of surgical intervention in spinal cord injury.** *Spine (Phila Pa 1976).* 2010; **35**(21 Suppl): S166–73.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Wilson JR, Tetreault LA, Kwon BK, *et al.*: **Timing of Decompression in Patients With Acute Spinal Cord Injury: A Systematic Review.** *Global Spine Journal.* 2017; **7**(3 Suppl): 95S–115.
[Publisher Full Text](#)
- Resnick DK: **Updated Guidelines for the Management of Acute Cervical Spine and Spinal Cord Injury.** *Neurosurgery.* 2013; **72** Suppl 2: 1.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Fehlings MG, Wilson JR, Tetreault LA, *et al.*: **A Clinical Practice Guideline for the Management of Patients With Acute Spinal Cord Injury: Recommendations on the Use of Methylprednisolone Sodium Succinate.** *Global Spine Journal.* 2017; **7**(3 Suppl): 203S–11S.
[Publisher Full Text](#)
- Samuel AM, Bohl DD, Basques BA, *et al.*: **Analysis of Delays to Surgery for Cervical Spinal Cord Injuries.** *Spine (Phila Pa 1976).* 2015; **40**(13): 992–1000.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Levi L, Wolf A, Belzberg H: **Hemodynamic parameters in patients with acute cervical cord trauma: description, intervention, and prediction of outcome.** *Neurosurgery.* 1993; **33**(6): 1007–16; discussion 1016–7.
[PubMed Abstract](#)
- Ryken TC, Hurlbert RJ, Hadley MN, *et al.*: **The acute cardiopulmonary management of patients with cervical spinal cord injuries.** *Neurosurgery.* 2013; **72** Suppl 2: 84–92.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Vale FL, Burns J, Jackson AB, *et al.*: **Combined medical and surgical treatment after acute spinal cord injury: results of a prospective pilot study to assess the merits of aggressive medical resuscitation and blood pressure management.** *J Neurosurg.* 1997; **87**(2): 239–46.
[PubMed Abstract](#) | [Publisher Full Text](#)
- F** Hawryluk G, Whetstone W, Saigal R, *et al.*: **Mean Arterial Blood Pressure Correlates with Neurological Recovery after Human Spinal Cord Injury: Analysis of High Frequency Physiologic Data.** *J Neurotrauma.* 2015; **32**(24): 1958–67.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- Chen K, Marsh BC, Cowan M, *et al.*: **Sequential therapy of anti-Nogo-A antibody treatment and treadmill training leads to cumulative improvements after spinal cord injury in rats.** *Exp Neurol.* 2017; **292**: 135–44.
[PubMed Abstract](#) | [Publisher Full Text](#)
- F** Saadoun S, Chen S, Papadopoulos MC: **Intraspinal pressure and spinal cord perfusion pressure predict neurological outcome after traumatic spinal cord injury.** *J Neurol Neurosurg Psychiatry.* 2017; **88**(5): 452–3.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
- F** Evaniw N, Belley-Côté EP, Fallah N, *et al.*: **Methylprednisolone for the Treatment of Patients with Acute Spinal Cord Injuries: A Systematic Review and Meta-Analysis.** *J Neurotrauma.* 2016; **33**(5): 468–81.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
- F** Evaniw N, Noonan VK, Fallah N, *et al.*: **Methylprednisolone for the Treatment of Patients with Acute Spinal Cord Injuries: A Propensity Score-Matched Cohort Study from a Canadian Multi-Center Spinal Cord Injury Registry.** *J Neurotrauma.* 2015; **32**(21): 1674–83.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)

32. Fernandez-Espejo E: **Pathogenesis of Parkinson's disease: prospects of neuroprotective and restorative therapies.** *Mol Neurobiol.* 2004; **29**(1): 15–30. [PubMed Abstract](#) | [Publisher Full Text](#)
33. Bonelli RM, Wenning GK, Kapfhammer HP: **Huntington's disease: present treatments and future therapeutic modalities.** *Int Clin Psychopharmacol.* 2004; **19**(2): 51–62. [PubMed Abstract](#) | [Publisher Full Text](#)
34. Kitzman PH: **Effectiveness of riluzole in suppressing spasticity in the spinal cord injured rat.** *Neurosci Lett.* 2009; **455**(2): 150–3. [PubMed Abstract](#) | [Publisher Full Text](#)
35. Satkunendrarajah K, Nassiri F, Karadimas SK, et al.: **Riluzole promotes motor and respiratory recovery associated with enhanced neuronal survival and function following high cervical spinal hemisection.** *Exp Neurol.* 2016; **276**: 59–71. [PubMed Abstract](#) | [Publisher Full Text](#)
36. Grossman RG, Fehlings MG, Frankowski RF, et al.: **A prospective, multicenter, phase I matched-comparison group trial of safety, pharmacokinetics, and preliminary efficacy of riluzole in patients with traumatic spinal cord injury.** *J Neurotrauma.* 2014; **31**(3): 239–55. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
37. Fehlings MG, Nakashima H, Nagoshi N, et al.: **Rationale, design and critical end points for the Riluzole in Acute Spinal Cord Injury Study (RISCIS): a randomized, double-blinded, placebo-controlled parallel multi-center trial.** *Spinal Cord.* 2016; **54**(1): 8–15. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
38. Martirosyan NL, Patel AA, Carotenuto A, et al.: **The role of therapeutic hypothermia in the management of acute spinal cord injury.** *Clin Neurol Neurosurg.* 2017; **154**: 79–88. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
39. Hansebout RR, Hansebout CR: **Local cooling for traumatic spinal cord injury: outcomes in 20 patients and review of the literature.** *J Neurosurg Spine.* 2014; **20**(5): 550–61. [PubMed Abstract](#) | [Publisher Full Text](#)
40. Levi AD, Casella G, Green BA, et al.: **Clinical outcomes using modest intravascular hypothermia after acute cervical spinal cord injury.** *Neurosurgery.* 2010; **66**(4): 670–7. [PubMed Abstract](#) | [Publisher Full Text](#)
41. Clifton GL, Miller ER, Choi SC, et al.: **Lack of effect of induction of hypothermia after acute brain injury.** *N Engl J Med.* 2001; **344**(8): 556–63. [PubMed Abstract](#) | [Publisher Full Text](#)
42. Simard JM, Tsybalyuk O, Ivanov A, et al.: **Endothelial sulfonylurea receptor 1-regulated NC_{Ca-ATP} channels mediate progressive hemorrhagic necrosis following spinal cord injury.** *J Clin Invest.* 2007; **117**(8): 2105–13. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
43. Schneider A, Krüger C, Steigleder T, et al.: **The hematopoietic factor G-CSF is a neuronal ligand that counteracts programmed cell death and drives neurogenesis.** *J Clin Invest.* 2005; **115**(8): 2083–98. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
44. Kawabe J, Koda M, Hashimoto M, et al.: **Neuroprotective effects of granulocyte colony-stimulating factor and relationship to promotion of angiogenesis after spinal cord injury in rats: laboratory investigation.** *J Neurosurg Spine.* 2011; **15**(4): 414–21. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
45. Hartung T: **Anti-inflammatory effects of granulocyte colony-stimulating factor.** *Curr Opin Hematol.* 1998; **5**(3): 221–5. [PubMed Abstract](#)
46. Kamiya K, Koda M, Furuya T, et al.: **Neuroprotective therapy with granulocyte colony-stimulating factor in acute spinal cord injury: a comparison with high-dose methylprednisolone as a historical control.** *Eur Spine J.* 2015; **24**(5): 963–7. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
47. Inada T, Takahashi H, Yamazaki M, et al.: **Multicenter prospective nonrandomized controlled clinical trial to prove neurotherapeutic effects of granulocyte colony-stimulating factor for acute spinal cord injury: analyses of follow-up cases after at least 1 year.** *Spine (Phila Pa 1976).* 2014; **39**(3): 213–9. [PubMed Abstract](#) | [Publisher Full Text](#)
48. Saberi H, Derakhshanrad N, Yekaninejad MS: **Comparison of neurological and functional outcomes after administration of granulocyte-colony-stimulating factor in motor-complete versus motor-incomplete postrehabilitated, chronic spinal cord injuries: a phase I/II study.** *Cell Transplant.* 2014; **23** Suppl 1: S19–23. [PubMed Abstract](#) | [Publisher Full Text](#)
49. Casha S, Zygun D, McGowan MD, et al.: **Results of a phase II placebo-controlled randomized trial of minocycline in acute spinal cord injury.** *Brain.* 2012; **135**(Pt 4): 1224–36. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
50. Fehlings MG, Theodore N, Harrop J, et al.: **A phase I/IIa clinical trial of a recombinant Rho protein antagonist in acute spinal cord injury.** *J Neurotrauma.* 2011; **28**(5): 787–96. [PubMed Abstract](#) | [Publisher Full Text](#)
51. Bregman BS, Kunkel-Bagden E, Schnell L, et al.: **Recovery from spinal cord injury mediated by antibodies to neurite growth inhibitors.** *Nature.* 1995; **378**(6556): 498–501. [PubMed Abstract](#) | [Publisher Full Text](#)
52. Merkle D, Metz GA, Raineteau O, et al.: **Locomotor recovery in spinal cord-injured rats treated with an antibody neutralizing the myelin-associated neurite growth inhibitor Nogo-A.** *J Neurosci.* 2001; **21**(10): 3665–73. [PubMed Abstract](#)
53. Schnell L, Schwab ME: **Axonal regeneration in the rat spinal cord produced by an antibody against myelin-associated neurite growth inhibitors.** *Nature.* 1990; **343**(6255): 269–72. [PubMed Abstract](#) | [Publisher Full Text](#)
54. Freund P, Schmidlin E, Wannier T, et al.: **Nogo-A-specific antibody treatment enhances sprouting and functional recovery after cervical lesion in adult primates.** *Nat Med.* 2006; **12**(7): 790–2. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
55. Freund P, Wannier T, Schmidlin E, et al.: **Anti-Nogo-A antibody treatment enhances sprouting of corticospinal axons rostral to a unilateral cervical spinal cord lesion in adult macaque monkey.** *J Comp Neurol.* 2007; **502**(4): 644–59. [PubMed Abstract](#) | [Publisher Full Text](#)
56. Brennan FH, Kurniawan ND, Vukovic J, et al.: **IVIg attenuates complement and improves spinal cord injury outcomes in mice.** *Ann Clin Transl Neurol.* 2016; **3**(7): 495–511. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
57. Gok B, Sciuuba DM, Okutan O, et al.: **Immunomodulation of acute experimental spinal cord injury with human immunoglobulin G.** *J Clin Neurosci.* 2009; **16**(4): 549–53. [PubMed Abstract](#) | [Publisher Full Text](#)
58. Nguyen DH, Cho N, Satkunendrarajah K, et al.: **Immunoglobulin G (IgG) attenuates neuroinflammation and improves neurobehavioral recovery after cervical spinal cord injury.** *J Neuroinflammation.* 2012; **9**: 224. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
59. Badner A, Siddiqui AM, Fehlings MG: **Spinal cord injuries: how could cell therapy help?** *Expert Opin Biol Ther.* 2017; **17**(5): 529–41. [PubMed Abstract](#) | [Publisher Full Text](#)
60. Dominici M, Le Blanc K, Mueller I, et al.: **Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement.** *Cytotherapy.* 2006; **8**(4): 315–7. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
61. Squillaro T, Peluso G, Galderisi U: **Clinical Trials With Mesenchymal Stem Cells: An Update.** *Cell Transplant.* 2016; **25**(5): 829–48. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
62. Oliveri RS, Bello S, Biering-Sørensen F: **Mesenchymal stem cells improve locomotor recovery in traumatic spinal cord injury: systematic review with meta-analyses of rat models.** *Neurobiol Dis.* 2014; **62**: 338–53. [PubMed Abstract](#) | [Publisher Full Text](#)
63. Rahimi-Movaghar V: **Clinical trials for the treatment of spinal cord injury: cervical and lumbar enlargements versus thoracic area.** *Brain.* 2009; **132**(Pt 7): e115; author reply e116. [PubMed Abstract](#) | [Publisher Full Text](#)
64. Thibault-Halman G, Rivers CS, Bailey CS, et al.: **Predicting Recruitment Feasibility for Acute Spinal Cord Injury Clinical Trials in Canada Using National Registry Data.** *J Neurotrauma.* 2017; **34**(3): 599–606. [PubMed Abstract](#) | [Publisher Full Text](#)
65. Phang I, Zoumprouli A, Papadopoulos MC, et al.: **Microdialysis to Optimize Cord Perfusion and Drug Delivery in Spinal Cord Injury.** *Ann Neurol.* 2016; **80**(4): 522–31. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
66. Fehlings MG, Cadotte DW, Fehlings LN: **A series of systematic reviews on the treatment of acute spinal cord injury: a foundation for best medical practice.** *J Neurotrauma.* 2011; **28**(8): 1329–33. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
67. Franssen BL, Hosman AJ, van Middendorp JJ, et al.: **Pre-hospital and acute management of traumatic spinal cord injury in the Netherlands: survey results urge the need for standardisation.** *Spinal Cord.* 2016; **54**(1): 34–8. [PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
68. Saadoun S, Papadopoulos MC: **Spinal cord injury: is monitoring from the injury site the future?** *Crit Care.* 2016; **20**(1): 308. [PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
69. Ates O, Cayli SR, Gurses I, et al.: **Comparative neuroprotective effect of sodium channel blockers after experimental spinal cord injury.** *J Clin Neurosci.* 2007; **14**(7): 658–65. [PubMed Abstract](#) | [Publisher Full Text](#)
70. Hachem LD, Mothe AJ, Tator CH: **Evaluation of the effects of riluzole on adult spinal cord-derived neural stem/progenitor cells in vitro and in vivo.** *Int J Dev Neurosci.* 2015; **47**(Pt B): 140–6. [PubMed Abstract](#) | [Publisher Full Text](#)
71. Hama A, Sagen J: **Antinociceptive effect of riluzole in rats with neuropathic spinal cord injury pain.** *J Neurotrauma.* 2011; **28**(1): 127–34. [PubMed Abstract](#) | [Publisher Full Text](#)
72. Mu X, Azbill RD, Springer JE: **Riluzole improves measures of oxidative stress**

- following traumatic spinal cord injury. *Brain Res.* 2000; **870**(1–2): 66–72.
[PubMed Abstract](#) | [Publisher Full Text](#)
73. Stutzmann JM, Pratt J, Boraud T, *et al.*: The effect of riluzole on post-traumatic spinal cord injury in the rat. *Neuroreport.* 1996; **7**(2): 387–92.
[PubMed Abstract](#) | [Publisher Full Text](#)
74. **F** Vasconcelos NL, Gomes ED, Oliveira EP, *et al.*: Combining neuroprotective agents: effect of riluzole and magnesium in a rat model of thoracic spinal cord injury. *Spine J.* 2016; **16**(8): 1015–24.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
75. **F** Hosier H, Peterson D, Tsybalyuk O, *et al.*: A Direct Comparison of Three Clinically Relevant Treatments in a Rat Model of Cervical Spinal Cord Injury. *J Neurotrauma.* 2015; **32**(21): 1633–44.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
76. Schwartz G, Fehlings MG: Evaluation of the neuroprotective effects of sodium channel blockers after spinal cord injury: improved behavioral and neuroanatomical recovery with riluzole. *J Neurosurg.* 2001; **94**(2 Suppl): 245–56.
[PubMed Abstract](#) | [Publisher Full Text](#)
77. Simard JM, Tsybalyuk O, Keledjian K, *et al.*: Comparative effects of glibenclamide and riluzole in a rat model of severe cervical spinal cord injury. *Exp Neurol.* 2012; **233**(1): 566–74.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
78. Wu Y, Satkumdrarajah K, Teng Y, *et al.*: Evaluation of the sodium-glutamate blocker riluzole in a preclinical model of cervical spinal cord injury. *Evid Based Spine Care J.* 2010; **1**(2): 71–2.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
79. Wu Y, Satkumdrarajah K, Teng Y, *et al.*: Delayed post-injury administration of riluzole is neuroprotective in a preclinical rodent model of cervical spinal cord injury. *J Neurotrauma.* 2013; **30**(6): 441–52.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
80. Batchelor PE, Kerr NF, Gatt AM, *et al.*: Hypothermia prior to decompression: buying time for treatment of acute spinal cord injury. *J Neurotrauma.* 2010; **27**(8): 1357–68.
[PubMed Abstract](#) | [Publisher Full Text](#)
81. Faroque M, Hillered L, Holtz A, *et al.*: Effects of moderate hypothermia on extracellular lactic acid and amino acids after severe compression injury of rat spinal cord. *J Neurotrauma.* 1997; **14**(1): 63–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
82. Grulova I, Slovinska L, Nagyova M, *et al.*: The effect of hypothermia on sensory-motor function and tissue sparing after spinal cord injury. *Spine J.* 2013; **13**(12): 1881–91.
[PubMed Abstract](#) | [Publisher Full Text](#)
83. Morino T, Ogata T, Takeba J, *et al.*: Microglia inhibition is a target of mild hypothermic treatment after the spinal cord injury. *Spinal Cord.* 2008; **46**(6): 425–31.
[PubMed Abstract](#) | [Publisher Full Text](#)
84. Seo JY, Kim YH, Kim JW, *et al.*: Effects of Therapeutic Hypothermia on Apoptosis and Autophagy After Spinal Cord Injury in Rats. *Spine (Phila Pa 1976).* 2015; **40**(12): 883–90.
[PubMed Abstract](#) | [Publisher Full Text](#)
85. Yu CG, Jimenez O, Maricillo AE, *et al.*: Beneficial effects of modest systemic hypothermia on locomotor function and histopathological damage following contusion-induced spinal cord injury in rats. *J Neurosurg.* 2000; **93**(1 Suppl): 85–93.
[PubMed Abstract](#) | [Publisher Full Text](#)
86. **F** Lo TP Jr, Cho KS, Garg MS, *et al.*: Systemic hypothermia improves histological and functional outcome after cervical spinal cord contusion in rats. *J Comp Neurol.* 2009; **514**(5): 433–48.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
87. Redondo-Castro E, Hernández J, Mahy N, *et al.*: Phagocytic microglial phenotype induced by glibenclamide improves functional recovery but worsens hyperalgesia after spinal cord injury in adult rats. *Eur J Neurosci.* 2013; **38**(12): 3786–98.
[PubMed Abstract](#) | [Publisher Full Text](#)
88. Simard JM, Woo SK, Norenberg MD, *et al.*: Brief suppression of *Abcc8* prevents autodestruction of spinal cord after trauma. *Sci Transl Med.* 2010; **2**(28): 28ra29.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
89. Popovich PG, Lemeshow S, Gensel JC, *et al.*: Independent evaluation of the effects of glibenclamide on reducing progressive hemorrhagic necrosis after cervical spinal cord injury. *Exp Neurol.* 2012; **233**(2): 615–22.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
90. Simard JM, Popovich PG, Tsybalyuk O, *et al.*: Spinal cord injury with unilateral versus bilateral primary hemorrhage—effects of glibenclamide. *Exp Neurol.* 2012; **233**(2): 829–35.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
91. Simard JM, Popovich PG, Tsybalyuk O, *et al.*: MRI evidence that glibenclamide reduces acute lesion expansion in a rat model of spinal cord injury. *Spinal Cord.* 2013; **51**(11): 823–7.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
92. Chen WF, Chen CH, Chen NF, *et al.*: Neuroprotective Effects of Direct Intrathecal Administration of Granulocyte Colony-Stimulating Factor in Rats with Spinal Cord Injury. *CNS Neurosci Ther.* 2015; **21**(9): 698–707.
[PubMed Abstract](#) | [Publisher Full Text](#)
93. Chung J, Kim MH, Yoon YJ, *et al.*: Effects of granulocyte colony-stimulating factor and granulocyte-macrophage colony-stimulating factor on glial scar formation after spinal cord injury in rats. *J Neurosurg Spine.* 2014; **21**(6): 966–73.
[PubMed Abstract](#) | [Publisher Full Text](#)
94. Dittgen T, Pitzer C, Plass C, *et al.*: Granulocyte-colony stimulating factor (G-CSF) improves motor recovery in the rat impactor model for spinal cord injury. *PLoS One.* 2012; **7**(1): e29880.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
95. Guo Y, Liu S, Zhang X, *et al.*: G-CSF promotes autophagy and reduces neural tissue damage after spinal cord injury in mice. *Lab Invest.* 2015; **95**(12): 1439–49.
[PubMed Abstract](#) | [Publisher Full Text](#)
96. Kadota R, Koda M, Kawabe J, *et al.*: Granulocyte colony-stimulating factor (G-CSF) protects oligodendrocyte and promotes hindlimb functional recovery after spinal cord injury in rats. *PLoS One.* 2012; **7**(11): e50391.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
97. Kato K, Koda M, Takahashi H, *et al.*: Granulocyte colony-stimulating factor attenuates spinal cord injury-induced mechanical allodynia in adult rats. *J Neurol Sci.* 2015; **355**(1–2): 79–83.
[PubMed Abstract](#) | [Publisher Full Text](#)
98. Koda M, Nishio Y, Kamada T, *et al.*: Granulocyte colony-stimulating factor (G-CSF) mobilizes bone marrow-derived cells into injured spinal cord and promotes functional recovery after compression-induced spinal cord injury in mice. *Brain Res.* 2007; **1149**: 223–31.
[PubMed Abstract](#) | [Publisher Full Text](#)
99. Nishio Y, Koda M, Kamada T, *et al.*: Granulocyte colony-stimulating factor attenuates neuronal death and promotes functional recovery after spinal cord injury in mice. *J Neuropathol Exp Neurol.* 2007; **66**(8): 724–31.
[PubMed Abstract](#) | [Publisher Full Text](#)
100. Pitzer C, Klusmann S, Krüger C, *et al.*: The hematopoietic factor granulocyte-colony stimulating factor improves outcome in experimental spinal cord injury. *J Neurochem.* 2010; **113**(4): 930–42.
[PubMed Abstract](#) | [Publisher Full Text](#)
101. Sanli AM, Serbes G, Caliskan M, *et al.*: Effect of granulocyte-colony stimulating factor on spinal cord tissue after experimental contusion injury. *J Clin Neurosci.* 2010; **17**(12): 1548–52.
[PubMed Abstract](#) | [Publisher Full Text](#)
102. Urdziková L, Jendelová P, Glogarová K, *et al.*: Transplantation of bone marrow stem cells as well as mobilization by granulocyte-colony stimulating factor promotes recovery after spinal cord injury in rats. *J Neurotrauma.* 2006; **23**(9): 1379–91.
[PubMed Abstract](#) | [Publisher Full Text](#)
103. Ahmad M, Zakaria A, Almutairi KM: Effectiveness of minocycline and FK506 alone and in combination on enhanced behavioral and biochemical recovery from spinal cord injury in rats. *Pharmacol Biochem Behav.* 2016; **145**: 45–54.
[PubMed Abstract](#) | [Publisher Full Text](#)
104. Aras M, Altas M, Motor S, *et al.*: Protective effects of minocycline on experimental spinal cord injury in rats. *Injury.* 2015; **46**(8): 1471–4.
[PubMed Abstract](#) | [Publisher Full Text](#)
105. Arnold SA, Hagg T: Anti-inflammatory treatments during the chronic phase of spinal cord injury improve locomotor function in adult mice. *J Neurotrauma.* 2011; **28**(9): 1995–2002.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
106. **F** Festoff BW, Ameenuddin S, Arnold PM, *et al.*: Minocycline neuroprotects, reduces microgliosis, and inhibits caspase protease expression early after spinal cord injury. *J Neurochem.* 2006; **97**(5): 1314–26.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
107. Lee SM, Yune TY, Kim SJ, *et al.*: Minocycline reduces cell death and improves functional recovery after traumatic spinal cord injury in the rat. *J Neurotrauma.* 2003; **20**(10): 1017–27.
[PubMed Abstract](#) | [Publisher Full Text](#)
108. Marchand F, Tsantoulas C, Singh D, *et al.*: Effects of Etanercept and Minocycline in a rat model of spinal cord injury. *Eur J Pain.* 2009; **13**(7): 673–81.
[PubMed Abstract](#) | [Publisher Full Text](#)
109. Sonmez E, Kabatas S, Ozen O, *et al.*: Minocycline treatment inhibits lipid peroxidation, preserves spinal cord ultrastructure, and improves functional outcome after traumatic spinal cord injury in the rat. *Spine (Phila Pa 1976).* 2013; **38**(15): 1253–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
110. **F** Tan AM, Zhao P, Waxman SG, *et al.*: Early microglial inhibition preemptively mitigates chronic pain development after experimental spinal cord injury. *J Rehabil Res Dev.* 2009; **46**(1): 123–33.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
111. Teng YD, Choi H, Onario RC, *et al.*: Minocycline inhibits contusion-triggered mitochondrial cytochrome c release and mitigates functional deficits after spinal cord injury. *Proc Natl Acad Sci U S A.* 2004; **101**(9): 3071–6.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
112. Yune TY, Lee JY, Jung GY, *et al.*: Minocycline alleviates death of oligodendrocytes by inhibiting pro-neuro growth factor production in microglia after spinal cord injury. *J Neurosci.* 2007; **27**(29): 7751–61.
[PubMed Abstract](#) | [Publisher Full Text](#)
113. Stirling DP, Khodarahmi K, Liu J, *et al.*: Minocycline treatment reduces delayed oligodendrocyte death, attenuates axonal dieback, and improves functional

- outcome after spinal cord injury.** *J Neurosci.* 2004; **24**(9): 2182–90.
[PubMed Abstract](#) | [Publisher Full Text](#)
114. **F** Wang Z, Nong J, Shultz RB, *et al.*: **Local delivery of minocycline from metal ion-assisted self-assembled complexes promotes neuroprotection and functional recovery after spinal cord injury.** *Biomaterials.* 2017; **112**: 62–71.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
115. Wells JE, Hurlbert RJ, Fehlings MG, *et al.*: **Neuroprotection by minocycline facilitates significant recovery from spinal cord injury in mice.** *Brain.* 2003; **126**(Pt 7): 1628–37.
[PubMed Abstract](#) | [Publisher Full Text](#)
116. Boato F, Hendrix S, Huelsenbeck SC, *et al.*: **C3 peptide enhances recovery from spinal cord injury by improved regenerative growth of descending fiber tracts.** *J Cell Sci.* 2010; **123**(Pt 10): 1652–62.
[PubMed Abstract](#) | [Publisher Full Text](#)
117. Dergham P, Ellezam B, Essagian C, *et al.*: **Rho signaling pathway targeted to promote spinal cord repair.** *J Neurosci.* 2002; **22**(15): 6570–7.
[PubMed Abstract](#)
118. Fournier AE, Takizawa BT, Strittmatter SM: **Rho kinase inhibition enhances axonal regeneration in the injured CNS.** *J Neurosci.* 2003; **23**(4): 1416–23.
[PubMed Abstract](#)
119. Sung JK, Miao L, Calvert JW, *et al.*: **A possible role of RhoA/Rho-kinase in experimental spinal cord injury in rat.** *Brain Res.* 2003; **959**(1): 29–38.
[PubMed Abstract](#) | [Publisher Full Text](#)
120. Atalay B, Bavbek M, Cekinmez M, *et al.*: **Antibodies neutralizing Nogo-A increase pan-cadherin expression and motor recovery following spinal cord injury in rats.** *Spinal Cord.* 2007; **45**(12): 780–6.
[PubMed Abstract](#) | [Publisher Full Text](#)
121. Brösamle C, Huber AB, Fiedler M, *et al.*: **Regeneration of lesioned corticospinal tract fibers in the adult rat induced by a recombinant, humanized IN-1 antibody fragment.** *J Neurosci.* 2000; **20**(21): 8061–8.
[PubMed Abstract](#)
122. Chen S, Smielewski P, Czosnyka M, *et al.*: **Continuous Monitoring and Visualization of Optimum Spinal Cord Perfusion Pressure in Patients with Acute Cord Injury.** *J Neurotrauma.* 2017; **34**(21): 2941–2949.
[PubMed Abstract](#) | [Publisher Full Text](#)
123. Fouad K, Klusman I, Schwab ME: **Regenerating corticospinal fibers in the Marmoset (*Callitrix jacchus*) after spinal cord lesion and treatment with the anti-Nogo-A antibody IN-1.** *Eur J Neurosci.* 2004; **20**(9): 2479–82.
[PubMed Abstract](#) | [Publisher Full Text](#)
124. Gonzenbach RR, Zoerner B, Schnell L, *et al.*: **Delayed anti-nogo-a antibody application after spinal cord injury shows progressive loss of responsiveness.** *J Neurotrauma.* 2012; **29**(3): 567–78.
[PubMed Abstract](#) | [Publisher Full Text](#)
125. **F** Liebscher T, Schnell L, Schnell D, *et al.*: **Nogo-A antibody improves regeneration and locomotion of spinal cord-injured rats.** *Ann Neurol.* 2005; **58**(5): 706–19.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
126. Maier IC, Ichiyama RM, Courtine G, *et al.*: **Differential effects of anti-Nogo-A antibody treatment and treadmill training in rats with incomplete spinal cord injury.** *Brain.* 2009; **132**(Pt 6): 1426–40.
[PubMed Abstract](#) | [Publisher Full Text](#)
127. Zörner B, Schwab ME: **Anti-Nogo on the go: from animal models to a clinical trial.** *Ann N Y Acad Sci.* 2010; **1198** Suppl 1: E22–34.
[PubMed Abstract](#) | [Publisher Full Text](#)
128. Fawcett JW, Curt A, Steeves JD, *et al.*: **Guidelines for the conduct of clinical trials for spinal cord injury as developed by the ICCP panel: spontaneous recovery after spinal cord injury and statistical power needed for therapeutic clinical trials.** *Spinal Cord.* 2007; **45**(3): 190–205.
[PubMed Abstract](#) | [Publisher Full Text](#)

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Version 1

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