

## LINC02688 and PP7080 as novel biomarkers in early diagnosis of gastric cancer

Sadegh Fattahi<sup>a,b,c</sup>, Novin Nikbakhsh<sup>d</sup>, Hassan Taheri<sup>e</sup>, Elham Ghadami<sup>f</sup>,  
 Mohammad Ranaee<sup>g</sup>, Haleh Akhavan-Niaki<sup>b,f,\*</sup>

<sup>a</sup> Student Research Committee, Babol University of Medical Sciences, Babol, Iran

<sup>b</sup> Cellular and Molecular Biology Research Center, Health Research Institute, Babol University of Medical Sciences, Babol, Iran

<sup>c</sup> North Research Center of Pasteur Institute, Amol, Iran

<sup>d</sup> Department of Surgery, Rouhani Hospital Babol University of Medical Sciences, Babol, Iran

<sup>e</sup> Department of Internal Medicine, Rouhani Hospital Babol University of Medical Sciences, Babol, Iran

<sup>f</sup> Department of Genetics, Faculty of Medicine, Babol University of Medical Sciences, Babol, Iran

<sup>g</sup> Department of Pathology, Rouhani Hospital, Babol University of Medical Sciences, Babol, Iran

### ARTICLE INFO

#### Keywords:

Gastric cancer  
 LncRNAs  
 Gene expression  
 Biomarker

### ABSTRACT

Despite considerable progress in gastric cancer screening, prevention, and treatment, it remains a major cause of morbidity and mortality worldwide. Due to late diagnosis of the disease, early potential diagnostic biomarkers are needed. Accumulating evidence indicates that non-coding RNAs have potential applications as diagnostic and prognostic biomarkers in gastric cancer. Herein, we investigated the expression levels of two novel non-coding RNAs, long intergenic non-protein coding RNA 2688 (*LINC02688*) and *LOC25845* (*PP7080*) by real-time PCR for the first time in 47 gastric cancer patients. We found significant downregulation of *LINC02688* and *LOC25845* (*PP7080*) with 3.44 and 2.2-fold decrease, respectively in tumoral tissues in comparison with their adjacent non-tumoral counterparts ( $P < 0.0001$ ). Our data also indicates that more than 96% and 88% of patients showed unchanged or decreased expression of *LINC02688* and *LOC25845* (*PP7080*), respectively. As most gastric cancer patients showed lower expression of these two lncRNAs, no significant association between clinicopathological features of the patients and the level of *LINC02688* and *LOC25845* (*PP7080*) expression could be detected. Furthermore, ROC curve analysis indicated that *LINC02688* and *PP7080* can serve as good predictive biomarkers for distinguishing tumoral tissues from their adjacent non-tumoral counterparts. Taken together, our findings suggested that these two novel tumor suppressor non-coding RNAs may act as novel diagnostic biomarkers for diagnosis of carcinogenesis event even at earlier stages of gastric adenocarcinoma.

### 1. Introduction

Gastric cancer (GC) is the fifth most frequently diagnosed cancer, and remains the third leading cause of cancer-related mortality worldwide, accounting for about 8.2% of all death cases [1]. Based on GLOBOCAN 2018 data more than 1,000,000 new cases and around 783,000 deaths from GC occurred in 2018 [1]. Both genetic and epigenetic factors as well as environmental determinants affect GC development [2–10]. Despite considerable progresses achieved in the treatment of GC, it remains a major clinical challenge due to late stage diagnosis and its poor prognosis [11–13]. Consequently, finding reliable prognostic biomarkers improves early diagnosis with an increased opportunity for

designing strategies for the prevention and treatment of GC. It is now well established that long non-coding RNAs (lncRNAs), which are defined as transcripts with more than 200 nucleotides, have diverse roles in regulating gene expression, and can be used as biomarkers for cancer diagnosis and prognosis as well as potential targets for cancer therapy [14–17]. In a previous pilot study, we unveiled important novel lncRNAs that were down-regulated in GC using RNA-sequencing (RNA-seq) technology by comparing the transcriptome level of tumoral tissues with their normal counterparts (accepted manuscript and unpublished data). Here the expression levels of two novel lncRNAs that were significantly downregulated were evaluated using Taqman real time PCR among gastric cancerous and non-cancerous tissues.

\* Corresponding author. Department of Genetics, Faculty of Medicine, Babol University of Medical Sciences, Babol, Iran.

E-mail address: [halehakhavan@yahoo.com](mailto:halehakhavan@yahoo.com) (H. Akhavan-Niaki).

<https://doi.org/10.1016/j.ncrna.2021.04.002>

Received 28 July 2020; Received in revised form 1 December 2020; Accepted 16 April 2021

Available online 21 April 2021

2468-0540/© 2021 The Authors. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co. Ltd. This is an open access article under the CC

BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 2. Materials and methods

### 2.1. Patients

A total of 94 specimens including 47 tumor and their adjacent non-tumoral tissues were collected from newly diagnosed gastric adenocarcinoma patients who underwent gastrectomy at hospitals in Mazandaran province, Iran during September 2015 to June 2018, and were enrolled into this study. All samples were snap-frozen in the liquid nitrogen after surgery, and stored at  $-80^{\circ}\text{C}$  for further analyzes. Written informed consent was obtained from all patients prior to enrolment in the study. This study was approved by the Ethics Committee of Babol University of Medical Sciences, and all procedures were performed in compliance with Helsinki declaration. Pathologic diagnosis of the tumoral and non-tumoral tissues have been performed by using hematoxylin and eosin staining, and infection status (*H. pylori*, CMV, HHV6, and EBV) was evaluated using PCR and real time PCR method as described previously [18].

### 2.2. RNA extraction

Approximately 10 mg of tumoral tissues and their adjacent non-tumoral counterparts were homogenized using liquid nitrogen. Subsequently, total RNA was extracted using RNX-plus reagent (Cinagen, Iran) according to the manufacturer's instructions. The purity and quantity of total RNA were measured by using NanoDrop 2000 (Thermo Scientific, USA), and 28S/18S ratio (28S and 18S ribosomal RNA bands) was evaluated by electrophoresis on 1.8% agarose gel to monitor the RNA integrity.

### 2.3. Quantitative real-time PCR analysis (qPCR)

Expression levels of two lncRNAs including LINC02688 and LOC25845 (PP7080) were examined by qRT-PCR in 47 pairs of gastric adenocarcinoma tissues and their normal adjacent counterparts. First-strand cDNA was synthesized with SuperScript II reverse transcriptase (Invitrogen, USA) using a gene specific stem-loop primer [19,20]. QRT-PCR was performed in a StepOne Plus real time instrument (Applied Biosystems, USA) using HotStar-Taq DNA Polymerase kit (Qiagen, Germany) according to the manufacturer's instructions. Amplification program consisted of initial denaturation at  $95^{\circ}\text{C}$  for 15 min, followed by 45 cycles of 15 s at  $95^{\circ}\text{C}$ , 1 min at  $60^{\circ}\text{C}$ . The expression levels of target RNAs were normalized to those of *RNU6*. All reactions were performed in triplicate. The sequences of forward and reverse primers along with universal Taqman probe are presented in Table 1. The sequence of Taqman probe was FAM 5' TGGATGTGCTGCGGCGTTTTATCAT 3' BHQ-1, and the sequence of reverse primer was 5' GTATCCAGTGCTGCGACCGT 3'.

Stem-loop sequence is underlined; target portion of mRNA is showed in red Bold.

**Table 1**

Sequences of primers used for evaluation of Non-coding RNAs expression.

Accession Number	Gene Name	Primers 5' → 3'
>NR_004394.1	<i>RNU6-1</i>	Specific forward primer: CTCGTTTCGGCAGCACATATAC RT-PCR primer GTCGTATCCAGTGTCTGCGACCGTATGGATGTGCTGCGGCGTTTTATCATGCAGTGGATACGACGTGTGCATCCTTGC
>NR_024158.1	<i>LOC25845</i>	Specific forward primer: GCCCTCGCCAGGTCITTTGAAC RT-PCR primer GTCGTATCCAGTGTCTGCGACCGTATGGATGTGCTGCGGCGTTTTATCATGCAGTGGATACGACTCCTGACTGGTG
>NR_160890.1	<i>LINC02688</i>	Specific forward primer: GGTCTGGTGTCTCTGACTTCTG RT-PCR primer GTCGTATCCAGTGTCTGCGACCGTATGGATGTGCTGCGGCGTTTTATCATGCAGTGGATACGACCCAGGACTCTC

### 2.4. Statistical analysis

Relative gene expression data was performed by StepOne software v2.3 and the  $2^{-\Delta\Delta\text{Ct}}$  method. Comparisons of results between groups were performed by paired Student's *t*-test. Receiver operating characteristic curve (ROC) analysis was performed using Prism software. A value of  $P < 0.05$  was considered as statistically significant.

## 3. Results

### 3.1. Clinicopathological features

The median age of patients was 67 years (range, 34–85 years), at the time of diagnosis, and also the male-to-female ratio was 3:1. Overall, more than half of the tumors were found at proximal position of the stomach (cardia, fundus, and body), and poorly differentiated tumors were observed in 65% of patients. In general, two-third of tumors were in stage I and II. Family history was found in 12.7% of the patients. Approximately, 90% of patients had at least one infection (EBV or CMV or HHV6 or *H. pylori*).

### 3.2. Expression levels of LINC02688 and LOC25845 (PP7080)

As shown in Fig. 1, LINC02688 and LOC25845 (PP7080) were significantly down-regulated by more than 3.44 and 2.2-fold in tumor samples in comparison with their adjacent non-tumoral tissues, respectively. Scatter plot analysis indicated that LINC02688 was significantly down-regulated in 32 out of 47 (68%) tumoral tissues in comparison with their adjacent non-tumoral counterparts. Also, out of 47 GC patients, 25 (53%) showed statistically significant down-regulation of LOC25845 (PP7080) in tumoral tissues in comparison with their adjacent non-tumoral counterparts (Fig. 2). Interestingly, LINC02688 and LOC25845 (PP7080) expressions were not detected in 2 and 1 tumoral tissue, respectively. There was no significant association between clinicopathological features of the patients and the level of LINC02688 and LOC25845 (PP7080) expression (Fig. 3).

### 3.3. Association of the infection status and LINC02688 and LOC25845 expression level

Association between four studied infections including EBV, CMV, HHV6, and *H. pylori* with the level of LINC02688 and LOC25845 (PP7080) was investigated in GC patients. No significant difference in LINC02688 and LOC25845 (PP7080) expressions level was detected between patients with and without viral infections. However, there was a significant difference for LINC02688 and LOC25845 (PP7080) mRNA levels in *H. pylori* positive GC patients in comparison with *H. pylori* negative patients (Fig. 4) ( $P = 0.0003$ , and  $P = 0.0001$ , respectively).

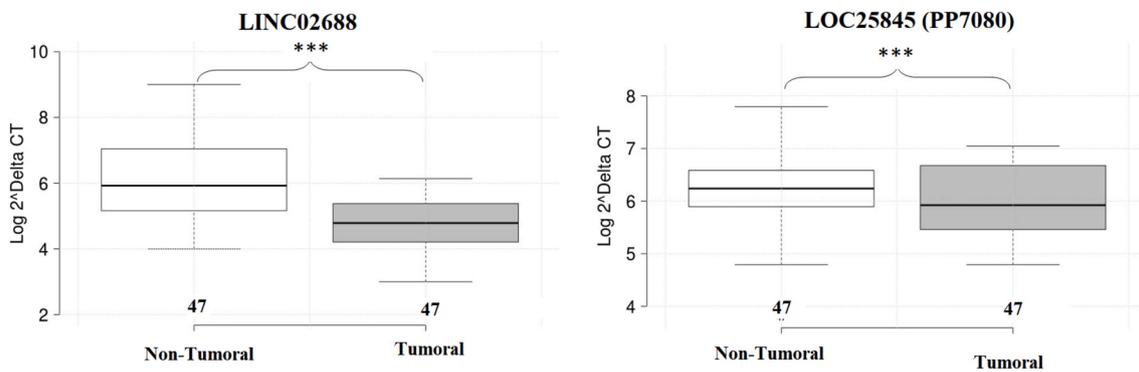


Fig. 1. *LINC02688* and *LOC25845 (PP7080)* expression fold change in 47 patients (tumoral tissues in comparison with non-tumoral tissues). Analyses were performed in triplicate, and data are shown as mean  $\pm$  SD. \*\*\* indicates significant difference  $P < 0.0001$ .

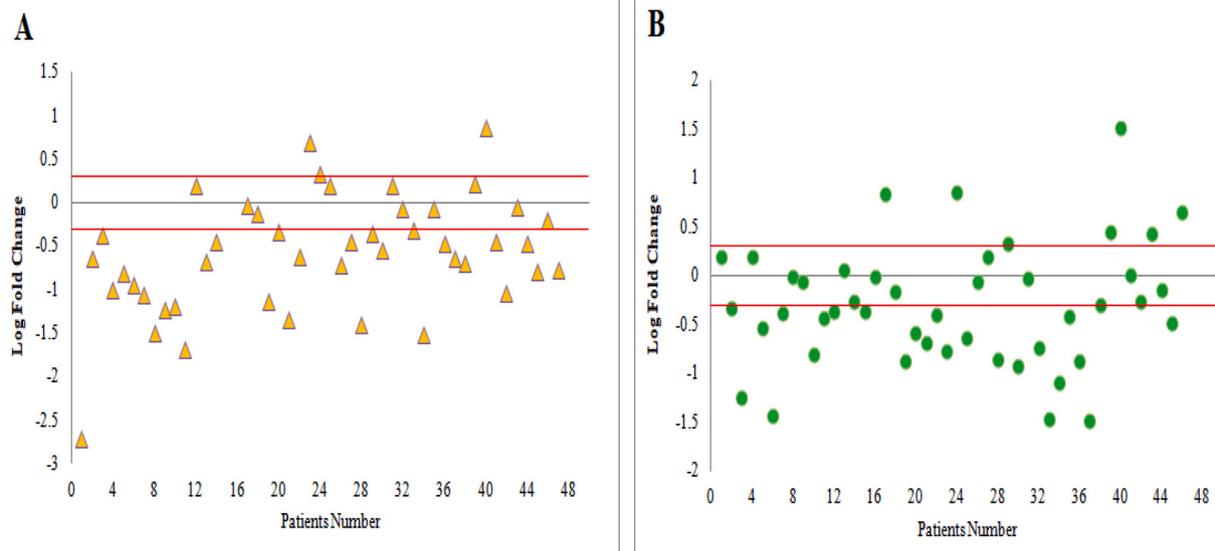


Fig. 2. Scatter plot of lncRNAs expression in tumoral and adjacent non-tumoral tissues. A) *LINC02688*; B) *LOC25845 (PP7080)*. Gene expression variations were considered in 47 gastric adenocarcinoma patients. Each plot represents a person's gene expression change in tumor tissue in comparison with adjacent normal tissue. Horizontal red lines represent cut-off values corresponding to log two-fold changes in expression. The upper and lower parts of the red lines indicate upregulation, and downregulation in the tumoral tissues in comparison with non-tumoral tissues, respectively.

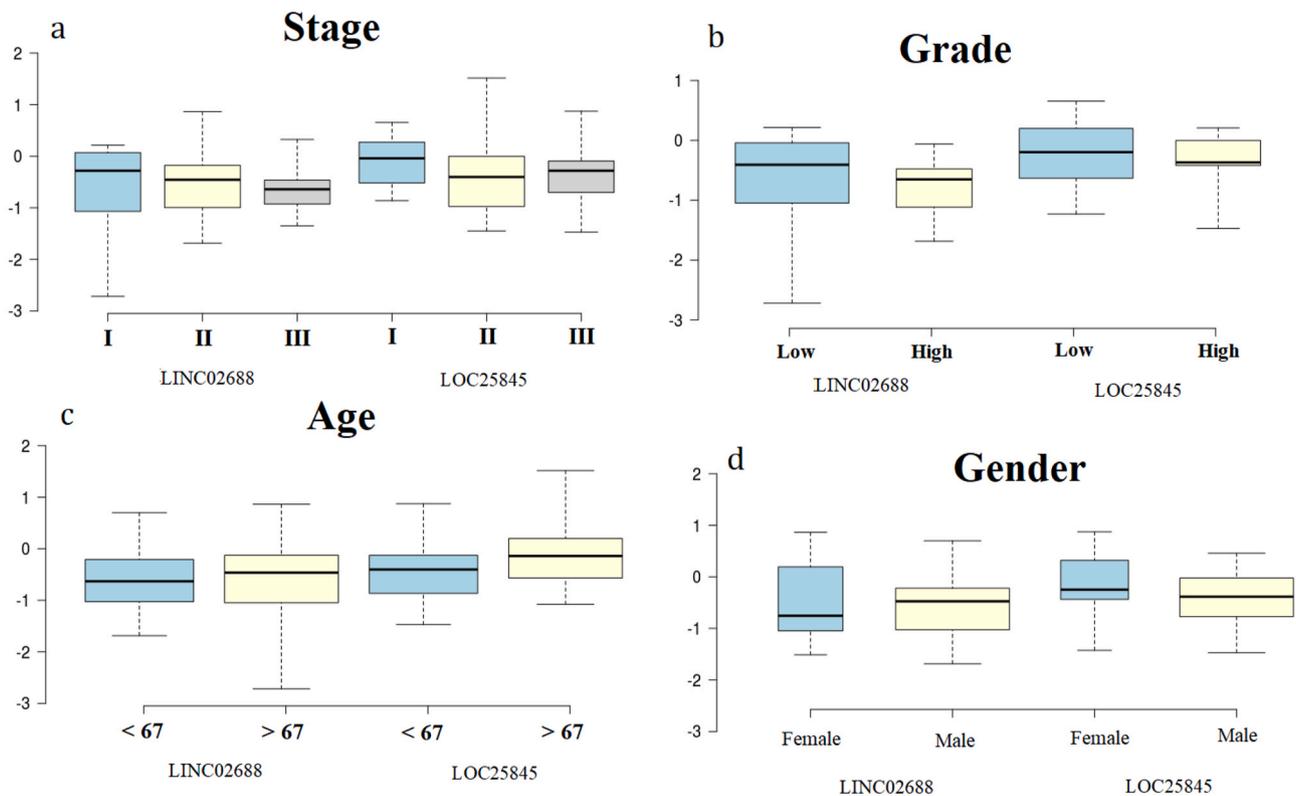
### 3.4. Receiver operating characteristic (ROC) curve analysis

The prognostic power of the candidate lncRNAs was evaluated by calculating the area under the curve (AUC) of receiver operating characteristic (ROC) curve. The AUCs of *LINC0268* and *LOC25845* were 0.63 (sensitivity 71.11% and specificity 51.06%,  $P = 0.02$ ) and 0.68 (sensitivity 80.43% and specificity 53.19%,  $P = 0.002$ ), respectively in tumoral tissues (Fig. 5). Also, based on the AUC values, *LINC0268* and *LOC25845 (PP7080)* had the best performance in differentiation of male from female patients (Fig. 5).

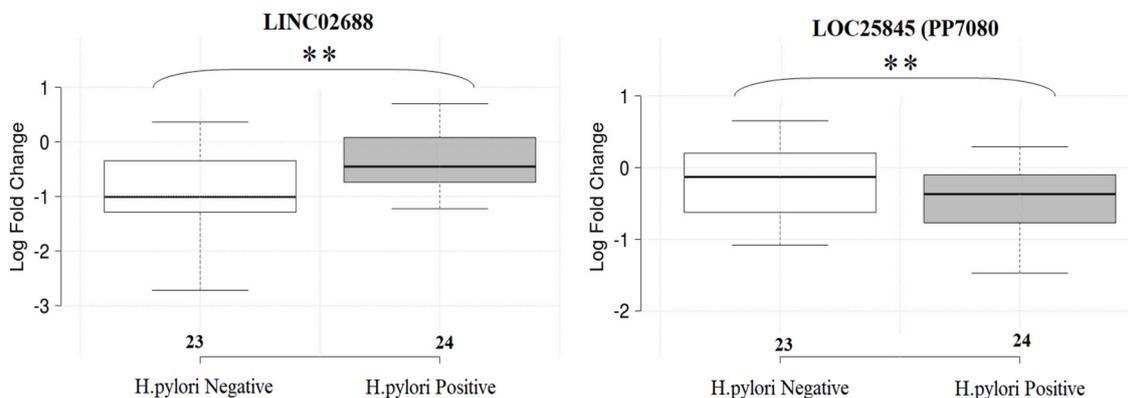
## 4. Discussion

The advent of high-throughput RNAseq witnessed a growing recognition for identifying thousands of unknown lncRNAs whose aberrant expression is closely associated with the initiation and development of cancers. Further findings demonstrated that many novel lncRNAs are dysregulated in GC, and closely related to tumorigenesis, metastasis, and prognosis or diagnosis. For instance, long intergenic non-protein coding RNA 941 (*LINC00941*) was found to be an independent predictor of tumor depth and distant metastasis in GC [21]. Also, long intergenic noncoding RNA 01296 was shown to mediate tumorigenesis through

sponging miR-122 in GC [22]. It was also shown that lncRNA *SUMO1P3* which is a direct target of CCHC-type zinc finger nucleic acid binding protein (CNBP) is an independent predictor for invasion and drug resistance in GC [23]. lncRNA *SNHG3* is another regulatory RNA involved in the progression of GC by regulating mediator complex subunit 18 (*MED18*) gene [24]. In the present study, we evaluated for the first time the expression level of two novel lncRNAs including *LOC25845 (PP7080)* and *LINC02688* in 47 pairs of GC tumoral tissues and their adjacent non-tumoral counterparts. Although the two novel lncRNAs analyzed in the present study were significantly down-regulated in tumor tissues in comparison with their paired adjacent normal gastric tissues, no significant association has been found between their expression level and patient's gender, tumor stage, tumor grade, and lymph node metastasis, highlighting their potential application in diagnosing tumors even at early stages or low dedifferentiation levels in GC. Furthermore, ROC curves analysis was performed in order to evaluate the predictive ability of lncRNAs expression in GC. The *LOC25845 (PP7080)* and *LINC02688* transcript levels had more than 70% specificity in this regard, indicating that these lncRNAs can serve as a potential biomarker in evaluating GC patients. Therefore, for suspicious samples at earlier stages of the disease, assessing the expression level of these lncRNAs could be useful to evaluating the malignancy of



**Fig. 3.** Association between LINC02688 and LOC25845 (PP7080) expression and clinical characteristics of gastric cancer patients. The expression level was represented by box plots using web (<http://ualcan.path.uab.edu/index.html>) for patients characteristics including (a) cancer stages, (b) tumor grade, (c) age, and (d) gender.



**Fig. 4.** Variations of lncRNAs according to *H. pylori* infection. LINC02688 and LOC25845 (PP7080) mRNA expression in *H. pylori*-positive (N = 24) and *H. pylori*-negative (N = 23) gastric cancers

\* indicates significant difference  $P < 0.01$  and \*\* indicates significant difference  $P < 0.001$ .

the collected tissue. In line with our study, microarray data (GSE49355 and GSE62321) of human colon tumor indicated that PP7080 was downregulated in liver metastasis of colon carcinoma [25]. Also, Huang et al. by analyzing TCGA database that included 460 patients with colon adenocarcinoma indicated that the expression of PP7080 was dysregulated in colon adenocarcinoma [26].

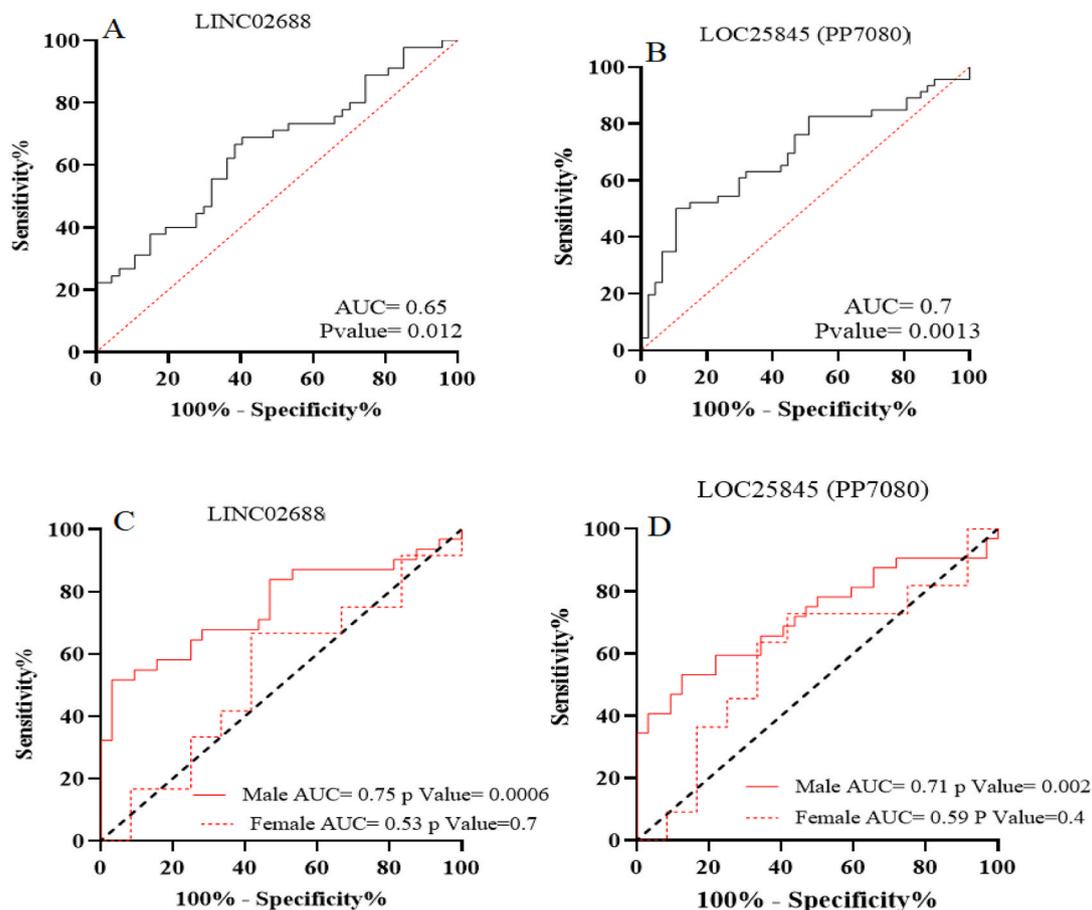
### 5. Conclusion

The present study for the first time revealed that LOC25845 (PP7080) and LINC02688 were downregulated in GC tissues, and their expressions were decreased from early to advanced stages of GC. Further studies

with more clinical samples of different type of cancers, and different ethnic populations are necessary in order to investigate the exact roles of LOC25845 (PP7080) and LINC02688 in development of cancers. These novel lncRNAs could be considered as a potential target for early diagnosis of GC in the future.

### Ethics approval

This study was approved by the Ethics Committee of Babol University of Medical Sciences and all procedures were performed in compliance with Helsinki declaration.



**Fig. 5.** The discriminatory ability of the candidate lncRNAs in gastric cancer patients. ROC curve analysis of the A) LINC02688 and B) *LOC25845* in tumoral tissues in comparison with their adjacent non-tumoral counterparts. ROC curves analysis of the (C) LINC02688 and (D) *LOC25845* (PP7080) in gender groups. AUC: area under the curve; ROC: receiver operating characteristic.

#### CRediT authorship contribution statement

**Sadegh Fattahi:** Methodology, Formal analysis, Investigation, Writing – original draft. **Novin Nikbakshsh:** Resources, Visualization. **Hassan Taheri:** Resources, Visualization. **Elham Ghadami:** Investigation, Resources. **Mohammad Ranaee:** Investigation, Visualization. **Haleh Akhavan-Niaki:** Conceptualization, Writing – review & editing, Supervision, Project administration.

#### Declaration of competing interest

Authors declare no conflict of interest.

#### References

- [1] F. Bray, J. Ferlay, I. Soerjomataram, R.L. Siegel, L.A. Torre, et al., Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries, *Ca - Cancer J. Clin.* 68 (6) (2018) 394–424.
- [2] A.A. Samadani, N. Nikbakshsh, H. Taheri, S. Shafae, S. Fattahi, et al., CDX1/2 and KLF5 expression and epigenetic modulation of sonic hedgehog signaling in gastric adenocarcinoma, *Pathol. Oncol. Res.* 25 (3) (2019) 1215–1222.
- [3] M. Kosari-Monfared, N. Nikbakshsh, S. Fattahi, E. Ghadami, M. Ranaee, et al., CTNBP1 downregulation is associated with tumor grade and viral infections in gastric adenocarcinoma, *J. Cell. Physiol.* 234 (3) (2019) 2895–2904.
- [4] S. Fattahi, M. Kosari-Monfared, E. Ghadami, M. Golpour, P. Khodadadi, et al., Infection-associated epigenetic alterations in gastric cancer: new insight in cancer therapy, *J. Cell. Physiol.* 233 (12) (2018) 9261–9270.
- [5] E. Ghadami, N. Nikbakshsh, S. Fattahi, M. Kosari-Monfared, M. Ranaee, et al., Epigenetic alterations of *CYLD* promoter modulate its expression in gastric adenocarcinoma: a footprint of infections, *J. Cell. Physiol.* 234 (4) (2019) 4115–4124.
- [6] S. Fattahi, M. Pilehchian Langroudi, H. Akhavan-Niaki, Hedgehog signaling pathway: epigenetic regulation and role in disease and cancer development, *J. Cell. Physiol.* 233 (8) (2018) 5726–5735.
- [7] S. Fattahi, M. Golpour, F. Amjadi-Moheb, M. Sharifi-Pasandi, P. Khodadadi, et al., DNA methyltransferases and gastric cancer: insight into targeted therapy, *Epigenomics* 10 (11) (2018) 1477–1497.
- [8] Y.Y. Lee, M.H. Derakhshan, Environmental and lifestyle risk factors of gastric cancer, *Arch. Iran. Med.* 16 (6) (2013) 358–365.
- [9] K. Yamashita, S. Sakuramoto, M. Watanabe, Genomic and epigenetic profiles of gastric cancer: potential diagnostic and therapeutic applications, *Surg. Today* 41 (1) (2011) 24–38.
- [10] H. Takeshima, T. Ushijima, Accumulation of genetic and epigenetic alterations in normal cells and cancer risk, *Npj Precision Oncol.* 3 (1) (2019) 7.
- [11] A.E. Dassen, V.E. Lemmens, L.V. van de Poll-Franse, G.J. Creemers, S. J. Breninkmeijer, et al., Trends in incidence, treatment and survival of gastric adenocarcinoma between 1990 and 2007: a population-based study in The Netherlands, *Eur. J. Canc.* 46 (6) (2010) 1101–1110.
- [12] G. Liu, M. Xu, T. Gao, L. Xu, P. Zeng, et al., Surgical compliance and outcomes in gastric cancer: a population-based cohort study, *J. Canc.* 10 (4) (2019) 779–788.
- [13] J. Zhang, L. Gan, Z. Wu, S. Yan, X. Liu, et al., The influence of marital status on the stage at diagnosis, treatment, and survival of adult patients with gastric cancer: a population-based study, *Oncotarget* 8 (14) (2017) 22385–22405.
- [14] Q.Q. Yang, Y.F. Deng, Long non-coding RNAs as novel biomarkers and therapeutic targets in head and neck cancers, *Int. J. Clin. Exp. Pathol.* 7 (4) (2014) 1286–1292.
- [15] M. Rasool, A. Malik, S. Zahid, M.A. Basit Ashraf, M.H. Qazi, et al., Non-coding RNAs in cancer diagnosis and therapy, *Non-Coding RNA Resear.* 1 (1) (2016) 69–76.
- [16] S. Chandra Gupta, Y. Nandan Tripathi, Potential of long non-coding RNAs in cancer patients: from biomarkers to therapeutic targets, *Int. J. Canc.* 140 (9) (2017) 1955–1967.
- [17] M. Sarfi, M. Abbastabar, E. Khalili, Long noncoding RNAs biomarker-based cancer assessment, *J. Cell. Physiol.* 234 (10) (2019) 16971–16986.
- [18] S. Fattahi, N. Nikbakshsh, H. Taheri, E. Ghadami, M. Kosari-Monfared, et al., Prevalence of multiple infections and the risk of gastric adenocarcinoma development at earlier age, *Diagn. Microbiol. Infect. Dis.* 92 (1) (2018) 62–68.

- [19] S. Fattahi, M. Pilehchian Langroudi, A.A. Samadani, N. Nikbakhsh, M. Asouri, et al., Application of unique sequence index (USI) barcode to gene expression profiling in gastric adenocarcinoma, *J. Cell Commun. Sig.* 11 (1) (2017) 97–104.
- [20] S. Fattahi, G. Amirbozorgi, M. Lotfi, B. Amini Navaei, S. Kavosian, et al., Development of a universal taqman probe for mRNA gene expression analysis, *Iran. J. Sci. Technol. Trans. A-Science* 42 (2) (2018) 363–370.
- [21] H. Liu, N. Wu, Z. Zhang, X. Zhong, H. Zhang, et al., Long non-coding RNA LINC00941 as a potential biomarker promotes the proliferation and metastasis of gastric cancer, *Front. Genet.* 10 (5) (2019).
- [22] Q.-H. Qin, Z.-Q. Yin, Y. Li, B.-G. Wang, M.-F. Zhang, Long intergenic noncoding RNA 01296 aggravates gastric cancer cells progress through miR-122/MMP-9, *Biomed. Pharmacother.* 97 (2018) 450–457.
- [23] Y. Guo, Y. Wang, Y. Ma, G. Chen, P. Yue, et al., Upregulation of lncRNA SUMO1P3 promotes proliferation, invasion and drug resistance in gastric cancer through interacting with the CNBP protein, *RSC Adv.* 10 (10) (2020) 6006–6016.
- [24] Y. Xuan, Y. Wang, Long non-coding RNA SNHG3 promotes progression of gastric cancer by regulating neighboring MED18 gene methylation, *Cell Death Dis.* 10 (10) (2019) 694.
- [25] J. Liu, D. Wang, C. Zhang, Z. Zhang, X. Chen, et al., Identification of liver metastasis-associated genes in human colon carcinoma by mRNA profiling, *Chin. J. Canc. Resear. Chung-Kuo Yen Cheng Yen Chiu* 30 (6) (2018) 633–646.
- [26] W. Huang, Z. Liu, Y. Li, L. Liu, G. Mai, Identification of long noncoding RNAs biomarkers for diagnosis and prognosis in patients with colon adenocarcinoma, *J. Cell. Biochem.* 120 (3) (2019) 4121–4131.