

CircANKRD11 Knockdown Protects HPMECs from Cigarette Smoke Extract-Induced Injury by Regulating miR-145-5p/BRD4 Axis

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Background: Chronic obstructive pulmonary disease (COPD) is a major cause of death because of its high incidence and mortality, which is chiefly resulted from cigarette smoke exposure. A large number of studies show that circular RNA (circRNA) participates in regulating COPD process. This study aims to reveal the role of circRNA ankyrin repeat domain 11 (circANKRD11) in cigarette smoke extract (CSE)-induced cell apoptosis, inflammation, and oxidative stress.

Methods: The expression of circANKRD11, microRNA-145-5p (miR-145-5p) and bromodomain-containing 4 (BRD4) mRNA was detected by quantitative real-time polymerase chain reaction. The expression of apoptosis-related proteins and BRD4 protein was determined by Western blot. Cell apoptosis was detected by flow cytometry and Western blot. Cell inflammation was demonstrated by determining the levels of interleukin-1 β (IL-1 β), IL-6 and tumor necrosis factor- α (TNF- α) through enzyme-linked immunosorbent assay. Oxidative stress was investigated by the reactive oxygen species (ROS) and malondialdehyde (MDA) determination assays as well as superoxide dismutase (SOD) activity assay. The binding relationship between miR-145-5p and circANKRD11 or BRD4 was predicted by circinteractome or MicroT_CDS online database, and identified by dual-luciferase reporter, RNA immunoprecipitation or RNA pull-down assay.

Results: CircANKRD11 and BRD4 expression were increased, whereas miR-145-5p expression was decreased in the lung tissues of smokers with or without COPD and CSE-induced HPMECs compared with the lung tissues of non-smokers as well as untreated HPMECs, respectively. CircANKRD11 silencing ameliorated CSE-induced cell apoptosis, inflammation, and oxidative stress. CircANKRD11 acted as a sponge of miR-145-5p, and regulated CSE-induced cell injury via sponging miR-145-5p. Additionally, miR-145-5p mimics protected against CSE-induced cell injury through targeting BRD4.

Conclusion: CircANKRD11 absence protected HPMECs from CSE-induced injury by regulating BRD4 through associating with miR-145-5p, which demonstrated that circANKRD11 had the potential to act as a diagnosis biomarker for smoker-caused COPD.

Keywords: COPD, circANKRD11, miR-145-5p, BRD4

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Introduction

Chronic obstructive pulmonary disease (COPD) is accompanied by respiratory symptom and is a leading cause of fatality rate globally.¹ Chronic bronchiolitis and emphysema are two main features of COPD and are commonly resulted from virus infection as well as toxic particles or gases.² Among the environmental

factors, cigarette smoking (CS) is deemed to be the most vital one and CS-caused COPD patients account for more than 80% of all COPD suffers yearly.³ CS can indirectly cause endothelial cell injury, and then lead to the upregulation of cell apoptotic rate and oxidative stress.^{4,5} Nevertheless, the underlying mechanisms of CS-induced COPD have not been fully revealed.

Circular RNA is an evolutionarily conserved noncoding RNA and is found in various organisms, featured by high expression, stability and conservatism.^{6,7} Based on these characteristics, circRNA can be employed as a prognostic or diagnostic biomarker for many diseases.⁸ CircRNA is also involved in the incidence and development of pulmonary diseases. For instance, circRNA ankyrin repeat domain (ANKRD) 36 (circANKRD36) absence attenuated lipopolysaccharide (LPS)-triggered cell apoptosis and inflammation via hoisting microRNA-31-3p (miR-31-3p) in lung fibroblasts.⁹ Huang et al indicated that circ_0001953 and circ_0009024 were related to the severity of active tuberculosis.¹⁰ Zhou et al explained circ_0016070 contributed to cell growth, and thereby was correlated with pulmonary arterial hypertension by binding to miR-942.¹¹ Additionally, circ_100395¹² and circ_0078767¹³ were found to inhibit cell proliferative and invasive capacities in lung cancer. Nevertheless, there was little study on the mechanism of smoke-induced COPD mediated by circRNA. Our previous data showed circANKRD11 expression was increased in cigarette smoke extract (CSE)-treated pulmonary microvascular endothelial cells (HPMECs). Whether circANKRD11 regulated smoke-related COPD process was revealed in this study.

MiRNA is a small RNA and consists of about 20 nucleotides, regulating the expression of interest gene via binding to its non-coding region.¹⁴ A plenty of miRNAs are different in expression in the lung specimens between smokers with COPD and smokers without COPD.^{2,15} MiRNA also participates in regulating the pathogenesis of COPD by targeting mRNA. As reported by Diao et al miR-132 mimics upregulated the levels of interleukin-1 β (IL-1 β) and tumor necrosis factor- α (TNF- α) through interacting with suppressor of cytokine signaling 5 (SOCS5) in human monocyte-like cells (THP-1).¹⁶ MiR-543 was lowly expressed in the lung tissue samples of smokers with acute exacerbation COPD, and its overexpression could attenuate CSE-induced upregulation of interferon- γ (INF- γ) and TNF- α via associating with fas apoptotic inhibitory molecule 2 (FAIM2).¹⁷ Bromodomain protein 4 (BRD4) belongs to Bromodomain and Extra-Terminal domain protein family and is an important regulator in gene transcription.¹⁸

Current researchers revealed that miR-145-5p¹⁹ and BRD4 overexpression²⁰ were involved in CSE-triggered cell apoptosis and inflammation in bronchial epithelial cells or HPMECs, suggesting miR-145-5p and BRD4 played vital parts in regulating CSE-induced COPD. Here we found that miR-145-5p possessed the complementary sequences of BRD4 and circANKRD11.

Thus, the impacts of circANKRD11 silencing on CSE-induced cell injury of HPMECs were revealed. The relationships among circANKRD11, miR-145-5p and BRD4 were disclosed. Furthermore, whether circANKRD11 modulated CSE-aroused HPMEC injury by miR-145-5p/BRD4 pathway was unveiled.

Materials and Methods

Sample Collection and the Ethics Committee

In accordance with the Declaration of Helsinki, the lung tissues from non-smokers (N=10), smokers without COPD (N=17) and smokers with COPD (N=21) were collected from pneumonectomy suffers who underwent a surgery of solitary non-small cell lung cancer in the Second Hospital of Hebei Medical University. Obtained tissues were stored at -80°C in a refrigerator. The written informed consent was signed by the subjects before operation. The Ethics Committee of the Second Hospital of Hebei Medical University approved this study. The clinical features of the subjects are listed in Table 1.

CSE Preparation

CSE was prepared based on the method shown by Richter and his colleagues.²¹ In brief, Liqun cigarettes (Tobacco Company; Xi'an, China) were combusted, and the smoke was bubbled through 25 mL Dulbecco's modified Eagle's medium (DMEM; Procell, Wuhan, China) with vacuum pumps (LeyNow, Dongguan, China). Then, the extract was filtered with 0.22 μm filters (Millipore, Bradford, MA, USA). And the obtained solution was referred as 100% CSE and diluted to various concentrations (0%, 0.5%, 1.0%, 2.5% and 5%) with fetal bovine serum (FBS)-free DMEM (Procell).

Cell Culture and CSE Treatment

Human pulmonary microvascular endothelial cells (HPMECs) and human bronchial epithelioid cells were purchased from Otwo Biotech (Shenzhen, China), and cultured in DMEM (Procell) or Roswell Park Memorial Institute-1640 (RPMI-1640; Procell) with 10% FBS (Procell) and

Table 1 Characteristics of Subjects in This Study

	Non-Smokers	Smokers	Smokers with COPD
Number	10	17	21
Age (Years)	61.3±3.1	62.2±4.5	64.5±5.8
Male	10(100%)	17(100%)	21(100%)
Smoking history (pack-years)	0	42.5±3.6**	48.9±4.9*##
BMI (kg/m ²)	26.3±4.5	28.9±4.2	27.7±5.9
FEV ₁ (L)	3.51±0.11	3.02±0.13	2.08±0.12*##
FVC (L)	4.01±0.33	3.75±0.38	3.66±0.15
FEV ₁ /FVC%	81.02±1.2	82.51±1.5	61.71±3.2*##
FEV ₁ (% predicted)	108.1±6.1	96.7±3.8	61.5±4.4*##
Occupational dust exposure, n			
Yes	1	2	2
No	9	15	19
Occupational hazard gas exposure, n			
Yes	2	1	1
No	8	16	20

Notes: *p<0.05 and **p<0.01, different from non-smoker; #p<0.05 and ##p<0.01, different from smokers.

Abbreviations: BMI, body mass index; FVC, forced vital capacity; FEV₁, forced expiratory volume in one second.

1% penicillin/streptomycin (Procell) at 37°C in an incubator with 5% CO₂. For CSE-related experiments, HPMECs were treated with CSE (0%, 0.5%, 1.0%, 2.5% or 5%) and cultured for the defined time (0, 6, 12, 24 or 36 h).

Cell Transfection

The small interfering RNA against circANKRD11 (si-circANKRD11), miR-145-5p mimics (miR-145-5p), miR-145-5p inhibitors (in-miR-145-5p), and control groups (si-NC, miR-NC and in-miR-NC) were provided by GenePharma (Shanghai, China). The overexpression plasmid of BRD4 (BRD4), the wild-type (WT) vectors of circANKRD11 (circANKRD11 WT) and the 3'-untranslated region (3'UTR) of BRD4 (BRD4 3'UTR WT), the mutant (MUT) vectors of circANKRD11 (circANKRD11 MUT) and BRD4 3'UTR (BRD4 3'UTR MUT) and pcDNA 3.1 (+) (pcDNA) were obtained from Genesee Co., Ltd. (Guangzhou, China). Constructed plasmids or synthesized oligonucleotides mixed with TurboFect reagent (Thermo Fisher, Waltham, MA, USA) were incubated with wells according to the instruction of manufacture. The sequences of synthesized oligonucleotides were displayed as below. Si-circANKRD11 5'-GACACAG CAGACGGTTGATGA-3'; miR-145-5p 5'-GUCCAGUUU UCCCAGGAAUCCCU-3'; in-miR-145-5p 5'-AGGGAU UCCUGGAAAACUGGAC-3'; si-NC 5'-CCTCTACCTGT CGCTGAGCTGTAAT-3'; miR-NC 5'-UUUGUACUAC ACAAAAGUACUG-3' and in-miR-NC 5'-CAGUACUU UUGUGUAGUACAAA-3'.

Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR)

Collected lung tissues and HPMECs were split with TransZol (TransGen, Beijing, China), and RNA was isolated with an RNAsimple kit (Tiangen, Beijing, China). Then, cDNA was synthesized with a FastKing RT Kit (Tiangen) or MicroRNA Reverse Transcription Kit (Thermo Fisher). To quantify analysis of circANKRD11, miR-145-5p and BRD4, cDNA was mixed with primers and SuperReal PreMix Color (Tiangen), and reacted on a thermocycler (Thermo Fisher). The value of cycle threshold (Ct) was assessed with the 2^{-ΔΔCt} method with β-actin as a reference. The sequences of sense and antisense primers were circANKRD11 5'-GTGA GGGAGCGAGCCAGCAA-3' and 5'-ACCCTTGGGCA TCGTCCTG-3'; miR-145-5p 5'-ACACTCCAGCTGGG GTCCAGTTTTCCCAGGA-3' and 5'-TGGTGTCTGTG GAGTCG-3'; BRD4 5'-CGTCTCCCTGACATTTCTTT-3' and 5'-GGTAATCCCTGGCAGTAGTT-3'; β-actin 5'-CAC CATTGGCAATGAGCGGTTTC-3' and 5'-AGGTCTTTCG GGATGTCCACGT-3'.

Annexin V-Fluorescein Isothiocyanate (Annexin V-FITC) and Propidium Iodide (PI) Double Staining Assay

To detect the apoptotic rate of HPMECs, Annexin V-FITC/PI detection kit (Solarbio, Beijing, China) was employed. Briefly, HPMECs were collected after digestion with

trypsin (Thermo Fisher). And cells were mixed with Binding Buffer (Solarbio) by vibrating. Then, Annexin V-FITC (Solarbio) and PI (Solarbio) were seriatim incubated with cells in dark. Samples were assessed with a flow cytometry (Thermo Fisher).

Western Blot Analysis

NP-40 buffer (Beyotime, Shanghai, China) was firstly employed to lyse cells, and lysates were mixed with loading buffer (Thermo Fisher). The mixture was then boiled in boiling water. After that, lysates were loaded by 15% bis-tris-acrylamide gel (Thermo Fisher). The protein bands were electrotransferred onto nitrocellulose membranes (Membrane Solutions, Shanghai, China), which were then immersed in 8% defatted milk (Solarbio). Then, the membranes were incubated with primary antibodies against B-cell lymphoma-2 (Bcl-2) (1:1000; Abcam, Cambridge, UK), BCL2-associated x protein (Bax) (1:5000; Abcam), Cleaved caspase 3 (c-caspase 3; 1:5000; Abcam), pro-caspase 3 (p-caspase 3; 1:2000; Abcam), BRD4 (1:800; Abcam) and β -actin (1:1000; Abcam), respectively. Afterwards, secondary antibody against IgG H&L (Abcam; 1:8000) was utilized to incubate the membranes. Finally, the protein bands were visualized by dropwise adding evoECL Plus (Beyotime) onto the membranes. β -actin was employed as a control.

Enzyme-Linked Immunosorbent Assay (ELISA)

The levels of IL-1 β , IL-6 and TNF- α were quantified by ELISA kits (Beyotime). In brief, cell supernatant was collected up by centrifuging at 300 g for 6 min. Cell supernatant was diluted with dilution buffer (Beyotime) and added into reaction wells. Two hours later, reaction wells were washed, and incubated with biotinylated antibodies (Beyotime) for 1 h. After that, streptavidin-marked horseradish peroxidase (Beyotime) and 3,3',5,5'-Tetramethylbenzidine (TMB) were severally added into the wells and incubated in dark for 20 min. The stop solution (Beyotime) was added, and samples were assessed using a microplate reader (Thermo Fisher) with the wavelength at 450 nm.

Reactive Oxygen Species (ROS) Detection Assay

ROS level was detected by Cellular ROS Assay kit (Abcam). Briefly, HPMECs or CSE-treated HPMECs were cultured in 6-well plates and transfected with si-circANKRD11, si-NC,

in-miR-145-5p, in-miR-NC, miR-145-5p, miR-NC, BRD4 or pcDNA according the different purposes. Forty-eight hours later, cells were collected and seeded in clear bottom 96-well plates. After 24 h, dichlorodihydrofluorescein diacetate (DCFH-DA) was utilized to stain cells in dark. The plates were measured by a fluorescence plate reader (Molecular Device, Sunnyvale, CA, USA) at excitation wavelength/emission wavelength=485 nm/535 nm.

Malondialdehyde (MDA) Determination Assay

MDA level was quantified by lipid peroxidation MDA assay kit (Abcam). In brief, cultured cells were harvested and washed using cold phosphate buffer solution (PBS; Solarbio). Then, cells were homogenized in Lysis Solution (Abcam), and insoluble material was removed by centrifuging. After that, thiobarbituric acid (TBA) was added into the wells and incubated at 95°C for 1 h. Supernatant was assessed by measuring the wavelength at 532 nm using a microplate reader (Thermo Fisher).

Superoxide Dismutase (SOD) Activity Detection

SOD activity was detected by SOD activity assay kit (Abcam). In brief, grown HPMECs were collected and lysed with Tris/HCl (Millipore). Supernatant was harvested by centrifuging, and then placed on ice. Samples were mixed with Trifluoroacetic acid sodium salt (WST), Working Solution (Abcam) and Enzyme Working Solution (Abcam). SOD activity was determined by analyzing the output of wavelength at 450 nm with a microplate reader (Thermo Fisher).

Dual-Luciferase Reporter Assay

The binding sequence between miR-145-5p and circANKRD11 or BRD4 was severally predicted by circinteractome (https://circinteractome.nia.nih.gov/api/v2/mirnasearch?circular_rna_query=hsa_circ_0040929&mirna_query=&submit=miRNA+Target+Search) and MicroT_CDS online databases (http://diana.imis.athena-innovation.gr/DianaTools/index.php?r=microT_CDS/results&keywords=hsa-miR-145-5p%20%20%20ENSG00000141867&genes=ENSG00000141867%20&mirnas=hsa-miR-145-5p%20&descr=&threshold=0.1&page=1). Built luciferase reporter plasmids were mixed with miR-145-5p or NC, and transfected into HPMECs with TurboFect reagent (Thermo Fisher). At the second day after transfection,

cells were collected and firefly luciferase activity was quantified by Dual-Lucy Assay Kit (Solarbio) with Renilla luciferase activity as a control.

RNA Immunoprecipitation (RIP) Assay

RIP assay was conducted using a Magna RIP kit (Millipore). In brief, HPMECs were lysed with RIP buffer (Millipore) containing RNase inhibitor (Millipore). Then, lysates were incubated with antibody against argonaute2 (Anti-AGO2; 1:50; Abcam) and immunoglobulin G (Anti-IgG; 1:100; Abcam), respectively. After 4 h, magnetic protein A/G beads (Millipore) were incubated with lysates for 24 h. Then, proteinase k (Millipore) was added to digest proteins, and the contents of circANKRD11 and miR-145-5p were determined by qRT-PCR.

RNA Pull-Down Assay

HPMECs were seeded in 12-well plates and treated with biotinylated wild-type miR-145-5p (Bio-miR-145-5p WT; Sangon, Shanghai, China), biotinylated mutant miR-145-5p (Bio-miR-145-5p MUT; Sangon) and biotinylated NC (Bio-NC; Sangon), respectively. Cells were cultured for another 48 h and lysed with RIP buffer (Millipore). Then, streptavidin-coupled beads (Invitrogen, Carlsbad, CA, USA) were incubated with the lysates. The magnetic beads were eluted, and the RNA complex pulled down was assessed by qRT-PCR.

Statistical Analysis

SPSS 21.0 software (IBM, Somers, NY, USA) was used to analyze the data derived from the three independent duplicate tests. Significant differences were compared with two-tailed Student's *t*-tests between the two groups and with

one-way analysis of variance (ANOVA) or Kruskal–Wallis test among three or more groups. The statistical significance was deemed when *P* value <0.05.

Results

CircANKRD11 Expression Was Increased in the Tissues of COPD Patients and CSE-Induced HPMECs

CircANKRD11 expression was firstly detected in the lung tissues of smokers without or with COPD. And qRT-PCR data showed circANKRD11 level was upregulated in smokers without COPD compared with non-smokers, and was higher in smokers with COPD than those without COPD (Figure 1A). This finding suggested that circANKRD11 expression might be correlated with the concentration of cigarette. To prove the guess, circANKRD11 expression was detected in CSE-induced HPMECs. Results presented that circANKRD11 expression was upregulated in a dose- and time-dependent manner after treatment of CSE in HPMECs (Figure 1B and C). HPMECs were treated with 2.5% CSE for 24 h in subsequent study. These results demonstrated that circANKRD11 might participate in regulating the process of smoke-caused COPD.

CircANKRD11 Knockdown Attenuated CSE-Induced Cell Apoptosis, Inflammation, and Oxidative Stress in Pulmonary Endothelial Cells

In order to reveal that whether circANKRD11 was enrolled in CSE-triggered lung injury, the effects between CSE and

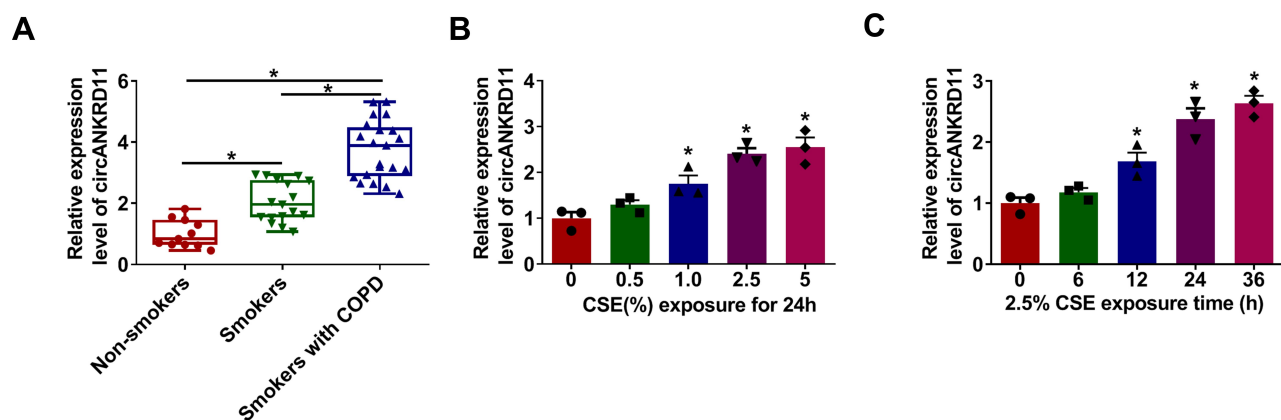


Figure 1 CircANKRD11 was overexpressed in the tissues of COPD patients and CSE-induced HPMECs. (A) CircANKRD11 expression was detected by qRT-PCR in the lung tissues of non-smokers (N=10), smokers without COPD (N=17) and smokers with COPD (N=21). (B) CircANKRD11 expression was determined by qRT-PCR in HPMECs after treatment of CSE (0%, 0.5%, 1.0%, 2.5% and 5%) for 24 h. (C) 2.5% CSE was exposed into HPMECs for 0, 6, 12, 24 and 36 h, respectively, and circANKRD11 expression was determined by qRT-PCR. **P*<0.05.

circANKRD11 silencing on cell apoptosis, inflammation, and oxidative stress were firstly explored in HPMECs. Results initially presented that CSE treatment increased circANKRD11 expression, whereas this effect was reversed after circANKRD11 silencing (Figure 2A). Additionally, CSE exposure induced cell apoptosis, while circANKRD11 knockdown hindered this impact (Figure 2B). To further demonstrate the effects between CSE and circANKRD11 silencing on the apoptosis of HPMECs, the levels of apoptosis-related proteins (Bcl-2, Bax, c-caspase 3 and p-caspase 3) were detected. Western blot analysis exhibited that CSE treatment decreased Bcl-2 protein expression, and increased the protein level of Bax and the value of c-caspase 3/p-caspase 3; however, these effects were abolished by si-circANKRD11 (Figure 2C). Additionally, the production of IL-1 β , IL-6 and TNF- α was promoted after treatment of CSE in HPMECs, but circANKRD11 knockdown impaired these effects (Figure 2D–F). Furthermore, CSE treatment upregulated the levels of ROS and MDA, and repressed SOD activity, which were reversed after circANKRD11 silencing in HPMECs (Figure 2G–I). Collectively, the above data demonstrated that circANKRD11 knockdown could ameliorate CSE-induced HPMEC injury.

Next, to determine whether the above observations could be repeatable in other human pulmonary endothelial cells, we studied the effects of circANKRD11 knockdown on inflammation and oxidative stress in CSE-treated HBE cells. Results initially presented the high efficiency of si-circANKRD11 in reducing circANKRD11 expression (Figure S1A). Additionally, results showed that the production of IL-1 β , IL-6, TNF- α , ROS and MDA was promoted, and SOD activity was repressed after treatment of CSE in HBE cells, but circANKRD11 knockdown impaired these effects (Figure S1B–G). The above evidences demonstrated that circANKRD11 knockdown attenuated CSE-induced cell inflammation and oxidative stress in HBE cells. Based on the similar results in HPMECs and HBE cells, further experiments were performed in HPMECs.

CircANKRD11 Acted as a Sponge of miR-145-5p

In order to reveal the mechanism of circANKRD11 in CSE-induced cell injury, the miRNA associated with circANKRD11 was continued to be explored. Circinteractome online database showed that miR-

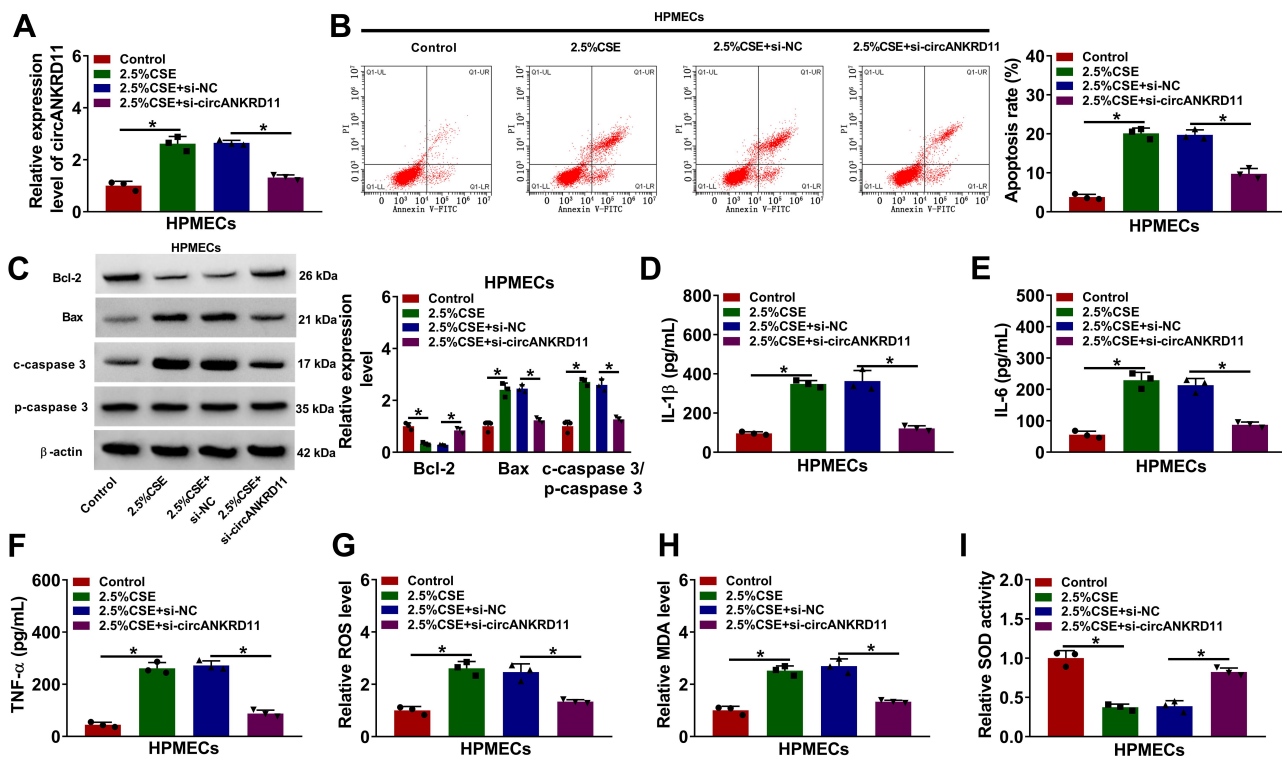


Figure 2 CircANKRD11 absence protected HPMECs from CSE-induced injury. (A–I) HPMECs were treated with 0% CSE (Control), 2.5% CSE, 2.5% CSE+si-NC and 2.5% CSE+si-circANKRD11, respectively. (A) CircANKRD11 expression was determined by qRT-PCR. (B) Cell apoptosis was detected by Annexin V-FITC and PI double staining assay. (C) The protein expression of Bcl-2, Bax, c-caspase 3 and p-caspase 3 was determined by Western blot analysis. (D–F) The levels of IL-1 β , IL-6 and TNF- α were detected by ELISA. (G) The level of ROS was detected by ROS detection assay. (H) MDA level was detected by MDA determination assay. (I) SOD activity was detected by SOD activity detection assay. * P <0.05.

145-5p possessed the binding sites of circANKRD11 (Figure 3A). To identify the binding relationship, the efficiency of synthesized miR-145-5p and in-miR-145-5p in affecting miR-145-5p expression was firstly detected. The results from qRT-PCR presented that miR-145-5p expression was apparently upregulated by miR-145-5p mimics, and downregulated by miR-145-5p inhibitors (Figure 3B). Subsequently, dual-luciferase reporter assay showed the relative luciferase activity was dramatically repressed in the co-transfection group of circANKRD11 WT and miR-145-5p, but not in circANKRD11 MUT and miR-145-5p co-transfection group (Figure 3C). RIP assay displayed that circANKRD11 and miR-145-5p were significantly enriched in Anti-AGO2 group in comparison with Anti-IgG group in HPMECs (Figure 3D). Meanwhile,

RNA pull-down assay exhibited that circANKRD11 content was notably higher in Bio-miR-145-5p WT group than in Bio-miR-145-5p MUT group in HPMECs (Figure 3E). Additionally, qRT-PCR data presented that miR-145-5p expression was lower in the lung tissues of smokers without COPD than in non-smokers, and was greatly lower in the lung tissues of smokers with COPD than in smokers without COPD (Figure 3F). MiR-145-5p expression was also decreased after treatment of CSE in HPMECs in comparison with untreated HPMECs (Figure 3G). Furthermore, miR-145-5p inhibitors also attenuated circANKRD11 silencing-mediated promotion impact on miR-145-5p expression in HPMECs (Figure 3H). Thus, the above data showed circANKRD11 was directly associated with miR-145-5p in HPMECs.

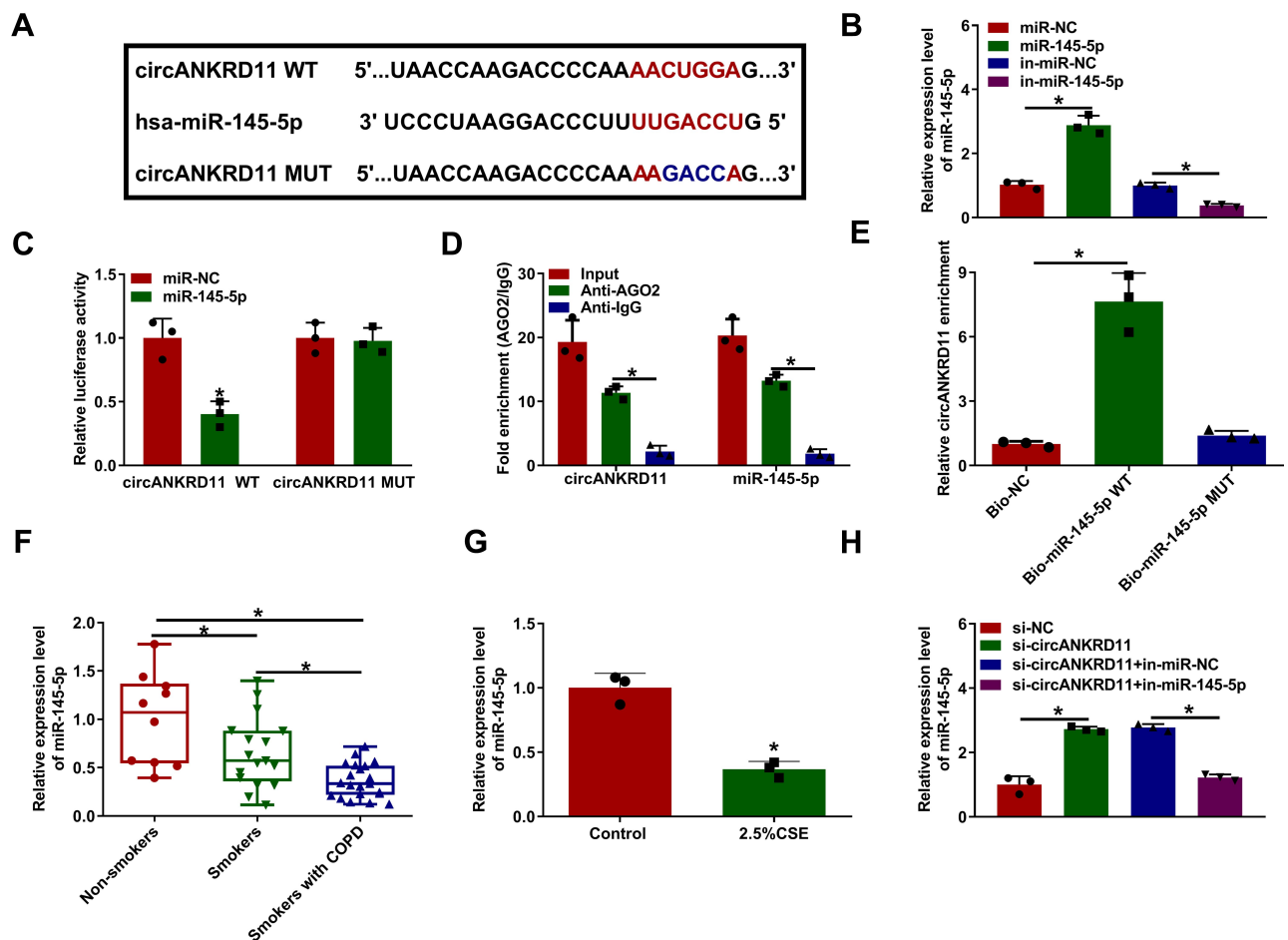


Figure 3 CircANKRD11 directly bound to miR-145-5p in HPMECs. (A) Circinteractome online database was employed to predict the binding sites between circANKRD11 and miR-145-5p. (B) MiR-145-5p expression was detected by qRT-PCR in HPMECs transfected with miR-NC, miR-145-5p, in-miR-NC or in-miR-145-5p. (C–E) Dual-luciferase reporter, RIP and RNA pull-down assays were employed to demonstrate the binding relationship between circANKRD11 and miR-145-5p in HPMECs. (F) MiR-145-5p expression was detected by qRT-PCR in the lung tissues of non-smokers (N=10), smokers without COPD (N=17) and smokers with COPD (N=21). (G) MiR-145-5p expression was determined by qRT-PCR in 2.5% CSE-treated HPMECs and untreated HPMECs. (H) The impacts between circANKRD11 silencing and miR-145-5p inhibitors on miR-145-5p expression were determined by qRT-PCR in HPMECs. * $P < 0.05$.

CircANKRD11 Regulated CSE-Induced Cell Apoptosis, Inflammation, and Oxidative Stress by Sponging miR-145-5p

Given the binding relationship between circANKRD11 and miR-145-5p, whether miR-145-5p participated in circANKRD11-mediated effects on CSE-caused cell injury was revealed in this part. Results showed miR-145-5p inhibitors reversed circANKRD11 silencing-mediated repressive impact on cell apoptosis in CSE-induced HPMECs (Figure 4A). Meanwhile, circANKRD11 knockdown upregulated Bcl-2 protein expression, and downregulated Bax protein expression and the value of c-caspase 3/p-caspase 3; however, these impacts were impaired by miR-145-5p inhibitors (Figure 4B). The repressive impacts of circANKRD11 knockdown on the production of IL-1 β , IL-6 and TNF- α were also attenuated after transfection of in-miR-145-5p (Figure 4C-E). Furthermore, circANKRD11 absence downregulated the levels of ROS and MDA, and promoted SOD activity, whereas these effects were abolished by miR-145-5p inhibitors (Figure

4F-H). Thus, these evidences demonstrated that circANKRD11 modulated CSE-induced HPMEC injury by binding to miR-149-5p.

CircANKRD11 Regulated BRD4 Expression by Interacting with miR-145-5p

The target gene of miR-145-5p was continued to be sought. MicroT_CDS online database presented that BRD4 3'UTR contained the binding sites of miR-145-5p (Figure 5A). Subsequently, dual-luciferase reporter assay displayed that the relative luciferase activity was apparently inhibited in BRD4 3'UTR WT and miR-145-5p co-transfection group, but not in BRD4 3'UTR MUT and miR-145-5p co-transfection group (Figure 5B). The results from qRT-PCR displayed high mRNA and protein expression of BRD4 in the lung tissues of smokers with COPD compared with non-smokers, and higher mRNA and protein expression of BRD4 in the lung tissues of

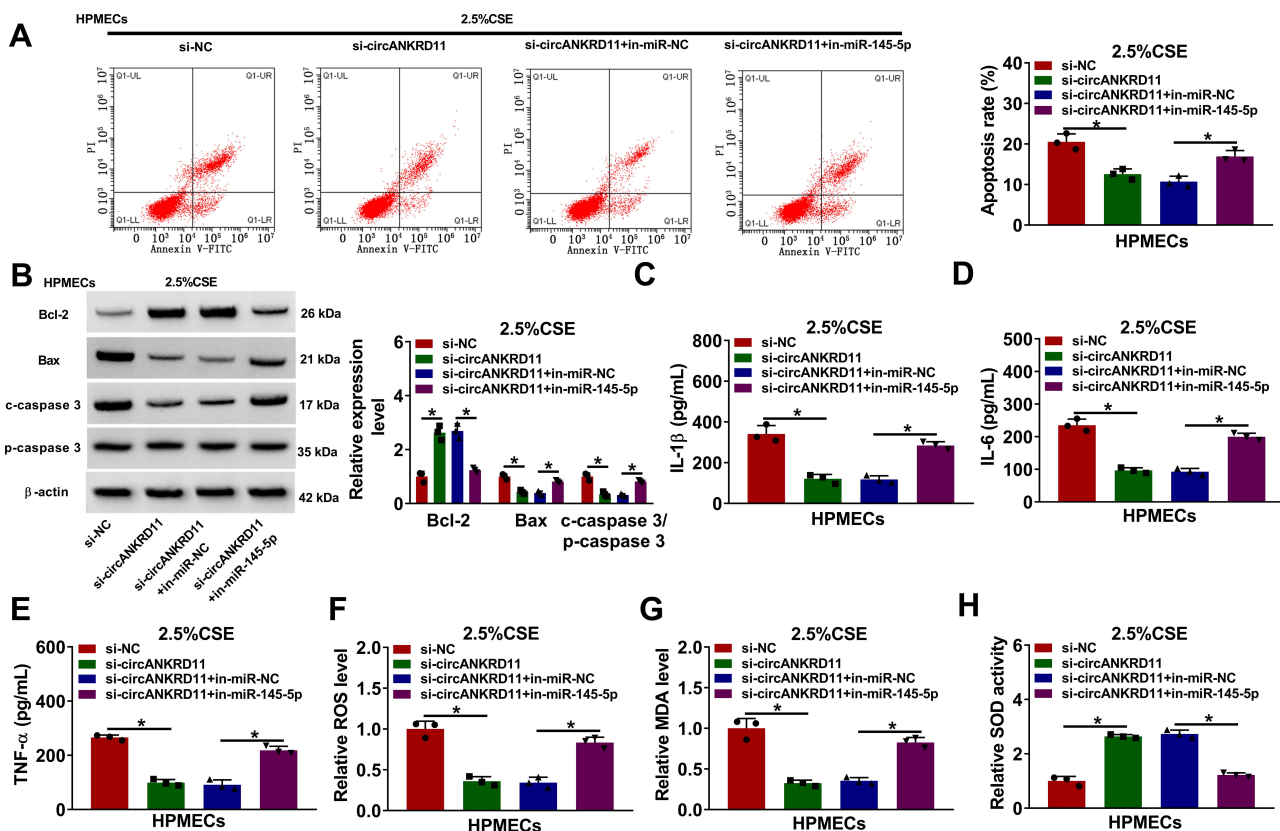


Figure 4 CircANKRD11 knockdown repressed cell apoptosis, inflammation, and oxidative stress by interacting with miR-145-5p in CSE-induced HPMECs. (A–H) 2.5% CSE-induced HPMECs were severally transfected with si-NC, si-circANKRD11, si-circANKRD11+in-miR-NC and si-circANKRD11+in-miR-145-5p. (A) Annexin V-FITC and PI double staining assay was employed to detect cell apoptosis. (B) The protein levels of Bcl-2, Bax, c-caspase 3 and p-caspase 3 were determined by Western blot analysis. (C–E) The levels of IL-1 β , IL-6 and TNF- α were detected by ELISA. (F) The level of ROS was detected by ROS detection assay. (G) MDA level was detected by MDA determination assay. (H) SOD activity was detected by SOD activity detection assay. * P <0.05.

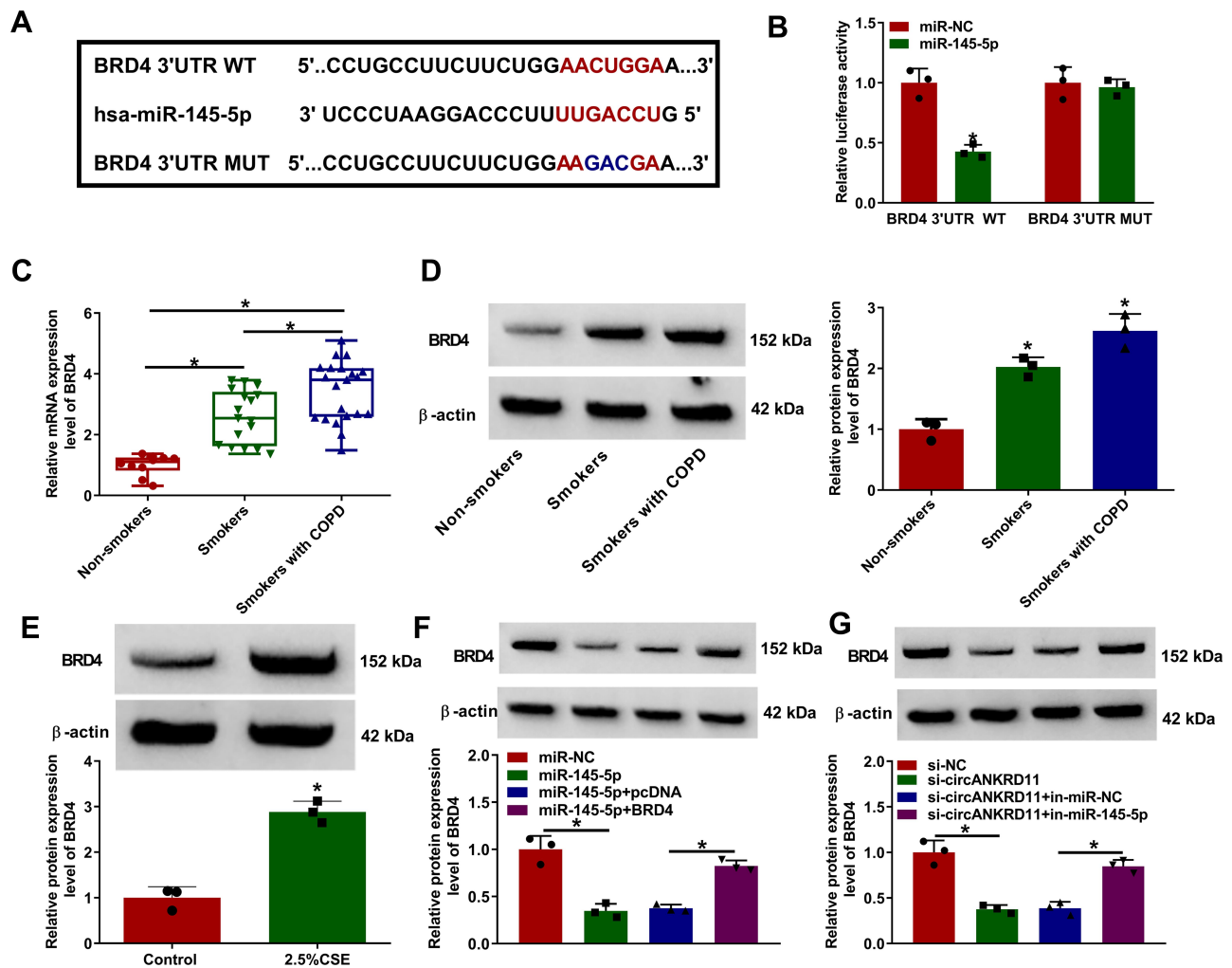


Figure 5 CircANKRD11 modulated BRD4 via binding to miR-145-5p. (A) The binding sites between miR-145-5p and BRD4 were predicted by microT_CDS online database. (B) Luciferase activities were detected by dual-luciferase reporter assay in HPMECs. (C and D) The mRNA and protein expression of BRD4 were determined by qRT-PCR and Western blot, respectively, in the lung tissues of non-smokers (N=10), smokers without COPD (N=17) and smokers with COPD (N=21). (E) BRD4 protein expression was detected by Western blot analysis in HPMECs treated with 0% CSE or 2.5% CSE. (F) The effects between miR-145-5p mimics and BRD4 overexpression on the protein expression of BRD4 were determined by Western blot analysis in HPMECs. (G) The impacts between circANKRD11 silencing and miR-145-5p inhibitors on BRD4 protein expression were revealed by Western blot analysis in HPMECs. * $P < 0.05$.

smokers with COPD as compared to smokers without COPD (Figure 5C and D). Additionally, BRD4 protein expression was also increased in 2.5% CSE-induced HPMECs when compared with untreated HPMECs (Figure 5E). The repressive impact of miR-145-5p mimics on BRD4 protein expression was also restored by BRD4 overexpression (Figure 5F). These results demonstrated BRD4 was a target gene of miR-145-5p. Furthermore, Western blot analysis presented that circANKRD11 knockdown decreased BRD4 protein expression, whereas miR-145-5p inhibitors reversed this impact (Figure 5G), suggesting that circANKRD11 regulated BRD4 expression by associating with miR-145-5p.

MiR-145-5p Mimics Suppressed Cell Apoptosis, Inflammation, and Oxidative Stress by Targeting BRD4 in CSE-Induced HPMECs

MiR-145-5p had been proved to bind to BRD4, whether it regulated CSE-aroused HPMEC injury by binding to BRD4 was further disclosed. To answer question, the effects between miR-145-5p mimics and BRD4 overexpression on cell apoptosis, inflammation, and oxidative stress were displayed. Results showed miR-145-5p mimics repressed cell apoptosis, but BRD4 overexpression attenuated this effect (Figure 6A). MiR-145-5p mimics also upregulated Bcl-2

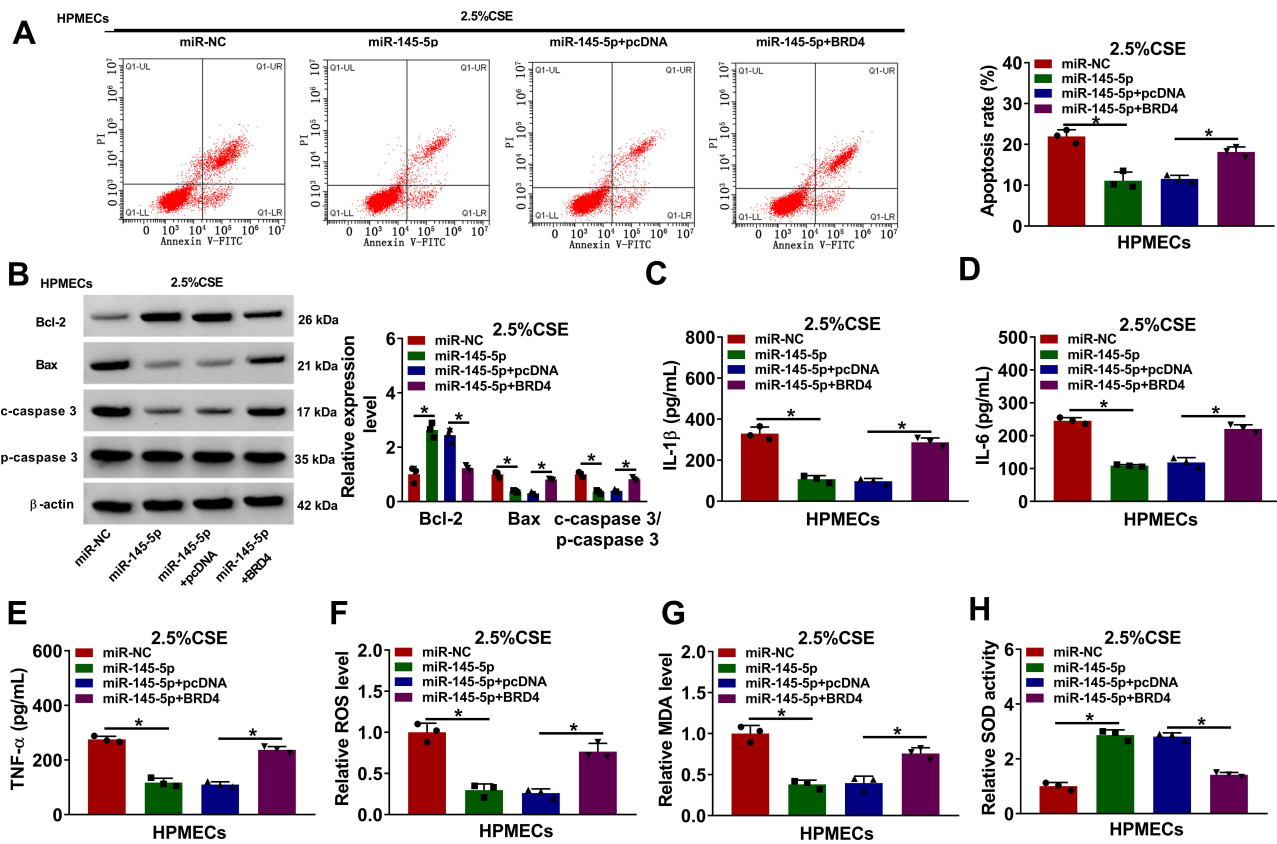


Figure 6 MiR-145-5p mimics protected against CSE-induced HPMEC injury through interacting with BRD4. (A–H) HPMECs were transfected with miR-NC, miR-145-5p, miR-145-5p+pcDNA and miR-145-5p+BRD4, respectively, under 2.5% CSE treatment in HPMECs. (A) Cell apoptosis was detected by Annexin V-FITC and PI double staining assay. (B) The protein expression of Bcl-2, Bax, c-caspase 3 and p-caspase 3 was determined by Western blot analysis. (C–E) The levels of IL-1 β , IL-6 and TNF- α were detected by ELISA. (F) The level of ROS was detected by ROS detection assay. (G) MDA level was detected by MDA determination assay. (H) SOD activity was detected by SOD activity detection assay. * P <0.05.

protein expression, and downregulated the protein level of Bax and the value of c-caspase 3/p-caspase 3; however, these effects were reversed after transfection of BRD4 (Figure 6B). Additionally, the production of IL-1 β , IL-6 and TNF- α was inhibited by miR-145-5p mimics, which was impaired by ectopic BRD4 expression (Figure 6C–E). Furthermore, the levels of ROS and MDA were downregulated, and SOD activity was promoted after transfection of miR-145-5p, whereas these effects were reversed after BRD4 overexpression (Figure 6F–H). These results demonstrated miR-145-5p could modulate CSE-induced HPMEC injury via targeting BRD4.

Discussion

The features of circRNAs in structure and expression make them serve as diagnostic biomarkers or therapeutic targets for diseases.²² In COPD, circRNAs have been seldom investigated previously. Current reports demonstrated that circ_0001859 could be employed as a diagnostic and prognostic biomarker based on its dysregulation in CS-treated lung

tissues of mice and peripheral blood of patients with COPD as well as the analysis of receiver operating characteristics (ROC) curve.²³ Duan et al analyzed the expression profiles of circRNAs and mRNAs, and demonstrated that circRNAs played vital roles in COPD process via regulating the immune balance and inflammatory responses.²⁴ In mechanism, Ma and his colleagues explained that circ_0061052 implicated in epithelial–mesenchymal transition of CSE-induced airway remodeling by controlling forkhead box C1/snail pathway through binding to miR-515-5p.²⁵ Circ_0006872 was shown to facilitate CSE-triggered cell injury by mediating cell apoptosis, inflammatory response and oxidative stress via regulating miR-145-5p/nuclear factor kappaB pathway.²⁶ In this paper, a novel circRNA, circANKRD11, was revealed that its silencing protected HPMECs from CSE-induced injury by miR-145-5p/BRD4 axis.

CS can induce cell apoptosis in various types of cells, such as fibroblasts and vascular endothelial cells.^{27,28} CS is the best-defined stimulant that causes COPD and leads to nonreversible pulmonary injury.²⁹ In this research, circANKRD11 was

augmented in the lung tissue samples of smokers without COPD, and was greatly augmented in smokers with COPD. Another result presented that circANKRD11 expression was dose- and time-dependently upregulated in CSE-treated HPMECs. These evidences suggested that circANKRD11 might be enrolled in the regulation of smoke-related COPD process. Subsequently, loss-of function experiments displayed that CSE treatment induced the upregulation of apoptotic rate and inflammatory factors and promoted oxidative stress, but these effects were attenuated by reduced circANKRD11, implying circANKRD11 knockdown could restrain CSE-induced cell injury.

CircRNAs commonly served as competing endogenous RNAs to sponge miRNAs.³⁰ So, the miRNA interacted with circANKRD11 was predicted. Our data presented miR-145-5p bound to circANKRD11, which was proved by qRT-PCR, dual-luciferase reporter, RIP and RNA pull-down assays. Current data presented that miR-145-5p had various roles in the process of diseases. For example, miR-145-5p could repress cell growth of glioma³¹ and hepatocellular carcinoma,³² attenuated hypertrophic scar,³³ regulated sensitivity of prolactinoma to bromocriptine³⁴ and suppressed airway remodeling in CSE-triggered COPD.³⁵ From our results, miR-145-5p was lowly expressed in smokers with or without COPD and CSE-induced HPMECs compared with non-smokers and untreated HPMECs, respectively, and miR-145-5p inhibitors restored circANKRD11 silencing-mediated effects on CSE-induced cell injury, meaning miR-145-5p could inhibit cell apoptosis, inflammation and oxidative stress, which was further proved by the previous data.^{19,26} Besides, these results explained that circANKRD11 regulated CSE-induced cell injury by sponging miR-145-5p.

Next, the target gene of miR-145-5p was continued to be sought. BRD4 was identified as an interest gene of miR-145-5p through dual-luciferase reporter assay. BRD4 has been revealed to repress the production of IL-1 β -caused inflammatory factors in human airway epithelial cells.³⁶ Liu et al showed BRD4 downregulation could protect mice from LPS-induced COPD by blocking NF- κ B pathway.³⁷ Of note, Castellucci et al indicated that IL-10 could repress LPS-triggered expression of TNF- α and C-X-C motif chemokine ligand 8 (CXCL8) by damaging BRD4-dokjing sites.³⁸ Additionally, BRD4 was also found to regulate CSE-triggered cell proliferation, apoptosis and inflammatory response in BEAS-2B cells.³⁹ In this paper, data also presented that BRD4 was augmented in the lung tissues of smokers with or without COPD as well as CSE-treated HPMECs, and that BRD4 induced cell apoptosis as well as

promoted inflammation and oxidative stress, which were in line with the study of Song et al.²⁰ In the meantime, the repressive impacts of BRD4 overexpression on miR-145-5p-mediated action implied that miR-145-5p regulated CSE-induced cell injury by targeting BRD4. Further, rescue experiments displayed that circANKRD11 regulated BRD4 expression by binding to miR-145-5p.

All in all, circANKRD11 knockdown mitigated CSE-induced cell apoptosis, inflammatory response and oxidative stress in HPMECs. CircANKRD11 was proved to bind to miR-145-5p, which was further found to target BRD4. Additionally, circANKRD11 knockdown attenuated CSE-induced cell injury by binding to miR-145-5p. And miR-145-5p mimics ameliorated CSE-caused cell damage via interacting with BRD4. Furthermore, circANKRD11 was presented to regulate BRD4 through associating with miR-145-5p. Collectively, circANKRD11 regulated CSE-induced HPMEC injury by controlling BRD4 through sponging miR-145-5p (as shown in Figure 7). This study not only demonstrates that circANKRD11 has the potential to serve as a diagnostic biomarker for smoke-related COPD, but also provides a theoretical basis for studying circRNA-directed therapy for smoke-caused COPD.

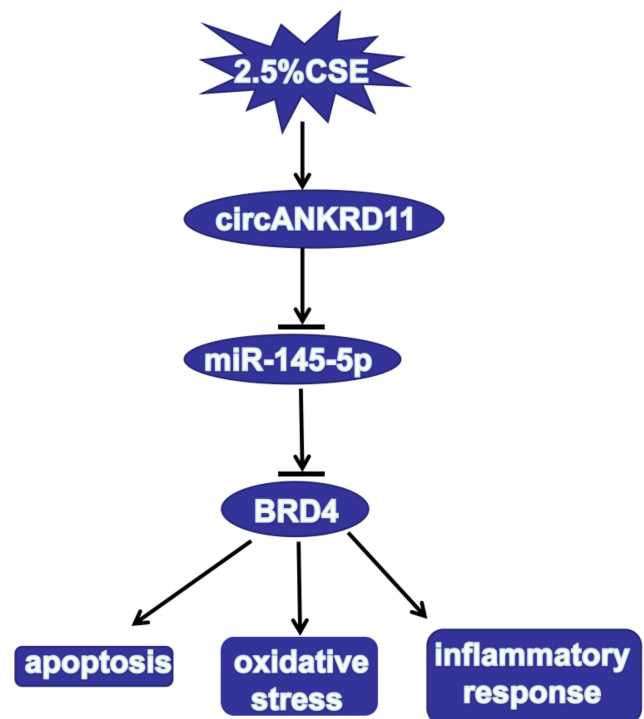


Figure 7 The illustration of circANKRD11 in regulating CSE-induced cell injury of HPMECs. CircANKRD11 regulated CSE-induced cell apoptosis, oxidative stress and inflammatory reaction by controlling BRD4 expression by sponging miR-145-5p.

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Disclosure

The authors declare that they have no financial conflicts of interest.

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