

OPEN

Isorhamnetin Alleviates Steatosis and Fibrosis in Mice with Nonalcoholic Steatohepatitis

Munkhzul Ganbold¹, Yohei Owada², Yusuke Ozawa², Yasuhiro Shimamoto³, Farhana Ferdousi⁴, Kenichi Tomimaga³, Yun-Wen Zheng², Nobuhiro Ohkohchi² & Hiroko Isoda^{1,3,4,5*}

Nonalcoholic steatohepatitis (NASH) is the most severe and progressive form of nonalcoholic fatty liver disease (NAFLD), which can lead to life-threatening conditions, however, there is still no approved drug for the treatment of NASH. In this study we used human-like NASH mouse model and treated orally with isorhamnetin at a dose of 50 mg/kg to analyze the effect of isorhamnetin on the progression of NASH. NASH-induced mice represented severe steatosis with inflammation, and fibrosis in liver accompanied with high level of liver injury markers in serum. Isorhamnetin treatment reduced intrahepatic lipid accumulation and TG content by inhibiting *de novo* lipogenic pathway in NASH-induced mice. Consistent with this, isorhamnetin-treated NASH mice showed improved liver injury markers, reduced collagen deposition as well as decreased gene expression of fibrogenic markers. Taken together, here we showed for the first time that synthesized isorhamnetin alleviates pathologic features of NASH and thus can potentially contribute to NASH drug development.

Nonalcoholic fatty liver disease (NAFLD) is one of the leading causes of chronic liver disease in the worldwide, with its global prevalence of 25% among adult population¹. NASH is characterized by hepatic lipid accumulation exceeding 5% of liver weight in the absence of heavy alcohol consumption associated with the histological features of hepatocyte damage, lobular inflammation, and fibrosis. The most commonly used hypothesis on its etiology is called “Two-hit theory” in which the “first hit” usually arising from metabolic complications of obesity, insulin resistance, diabetes mellitus, and metabolic syndrome. Oxidative stress, immune system, inflammation, gut and adipose tissue-derived factors, and genetic background, grouped as the “Second hit”, are shown to play an important role in the progression of NASH from NAFLD by inducing liver injury and fibrosis. Instead of a single risk factor in natural course of NASH, simultaneous persistence of more than one metabolic disorder may play pivotal role by causing subsequent pathologic features. However, the relevant signaling pathways and molecular mechanisms responsible for its progression from NAFLD to NASH remain unexplained. This gap in our knowledge considerably limits drug development against NASH. Other than lifestyle modification through diet and exercise, there are currently no other approved treatments for NASH^{2,3}.

It is worth to mention that a suitable animal model is crucial in the research progress of drug discovery. Several diet- and drug-induced mouse models have been used for their morphologic and histopathologic similarities to human NASH. Methionine and choline deficient (MCD) diet-induced NASH mice model is a commonly used aggressive NASH model, however, this model seems to develop neither peripheral insulin resistance nor obesity^{4,5}. Similarly, HFD-induced NAFLD rodent model could not develop hepatic fibrosis, the most important histological predictor for NASH progression in human. Nonetheless, neither MCD diet-induced NASH mice model nor HFD-induced NAFLD rodent model could represent full NASH hallmarks if the real pathologic context of human NASH needs to be considered. Kubota *et al.* and Tsuchida *et al.* proposed a combination of diet and chemical inducers for murine NASH models, however, it requires 8 and 12 weeks until NASH resembling features

¹School of Integrative and Global Majors (SIGMA), University of Tsukuba, Tsukuba, Ibaraki, 305-8572, Japan.

²Department of Gastrointestinal and Hepato-Biliary-Pancreatic Surgery, Faculty of Medicine, University of Tsukuba, Tsukuba, Ibaraki, 305-8575, Japan. ³Interdisciplinary Research Center for Catalytic Chemistry, National Institute of Advanced Industrial Science and Technology (AIST), Tsukuba, Ibaraki, 305-8565, Japan. ⁴Alliance for Research on the Mediterranean and North Africa (ARENA), University of Tsukuba, Tsukuba, Ibaraki, 305-8577, Japan. ⁵Faculty of Life and Environment Science, University of Tsukuba, Tsukuba, 305-8572, Japan. *email: isoda.hiroko.ga@u.tsukuba.ac.jp

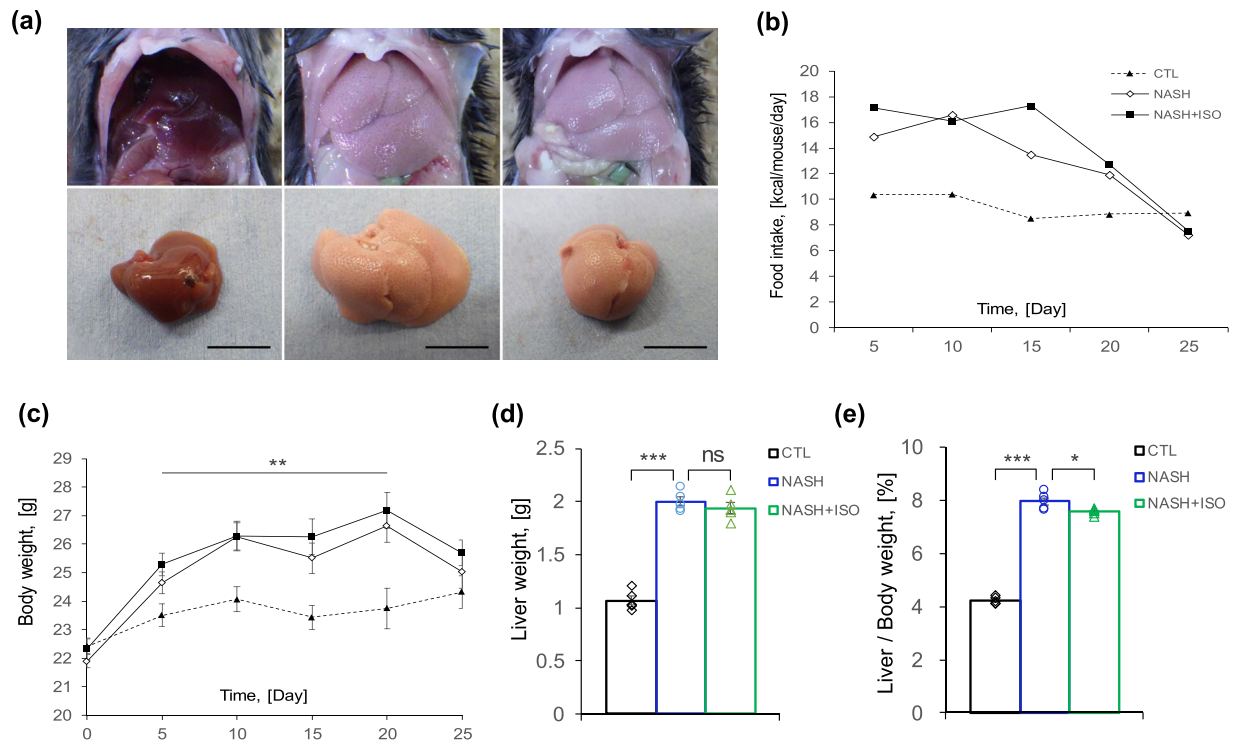


Figure 1. Biometrics of NASH-induced mice treated with isorhamnetin. (a) Representative liver aspect of experimental groups: control (CTL), NASH-induced (NASH), and NASH-induced with isorhamnetin treatment (NASH + ISO) group (scale bar = 1 cm). (b) Average food intake in calories of experimental groups presented as kcal per mouse per day (n = 8/group). (c) Body weight change (n = 8/group). (d) Effect of isorhamnetin on liver weight (n = 5/group, NASH vs. NASH + ISO: p = 0.389) and (e) liver on body weight ratio (n = 5/group, NASH vs. NASH + ISO: p = 0.025). Data are shown as mean ± SEM with significance *p < 0.05, **p < 0.01, ***p < 0.001 and ns = non-significant.

occur^{6,7}. Interestingly, Owada *et al.* used triple combination of diet and chemical inducers to develop human-like NASH in murine within 4 weeks⁸. Thus, in this study, we used the NASH model developed by Owada *et al.* which represents all pathologic features present in human NASH⁸.

Natural flavonoids have been shown to have bioactive effects against metabolic diseases^{9–11}. Isorhamnetin is a natural flavonoid found in several plants and plant-derived foods as well as an immediate metabolite of quercetin. It has been shown to inhibit proliferation of breast and lung cancer cells^{12,13}, protect HepG2 cells against oxidative stress¹⁴, attenuate inflammatory bowel disease¹⁵, and repress adipogenesis in 3T3-L1 cells¹⁶. Research studies including investigations in our laboratory showed its antioxidant, anti-obesity, and antifibrotic effects in rodents treated with isorhamnetin or plant extracts rich in isorhamnetin^{17–19}. Its aglycone parent – quercetin was already demonstrated to possess anti-fibrotic and hepatoprotective activity^{20,21}, however, reports on bioavailability of quercetin showed that a majority of the absorbed latter is found in its methylated form – isorhamnetin which is maintained in plasma longer than quercetin^{22,23}. It strongly implies a potential role of isorhamnetin as a mediator of the beneficial effect of quercetin. Structurally, previous comparative analysis revealed that aglycone flavonoids exert more biological activity compared to their glycones²⁴.

Those collected evidence led us to hypothesize that isorhamnetin may have efficient biological activity against hallmark features of NASH. To this end, we induced NASH in mice and treated them with isorhamnetin by oral gavage and evaluated its biological efficacy in the NASH model.

Results

Isorhamnetin reduced liver weight without affecting body weight in NASH-induced mice. Morphologically, livers from both NASH-induced (NASH) and NASH treated with isorhamnetin (NASH + ISO) groups were significantly enlarged and pale in color indicating fatty liver aspect compared to those of control (CTL) group, which were reddish-brown in color and small-sized (Fig. 1a). Compared to CTL group, the other two groups had increased food intake in calories and weight gain. However, no significant difference was observed in daily calorie intake and change in body weight between NASH and NASH + ISO groups (Fig. 1b,c) although both groups slightly lost body weight and food intake by the end of experiment which is specific to this model. Liver weight of NASH and NASH + ISO groups were indifferent (Fig. 1d). In contrast, liver on body weight ratio, which is extensively used as a clinical parameter to assess liver enlargement, was significantly reduced in NASH + ISO mice compared to NASH group (Fig. 1e), suggesting that isorhamnetin decreased liver weight before affecting body weight.

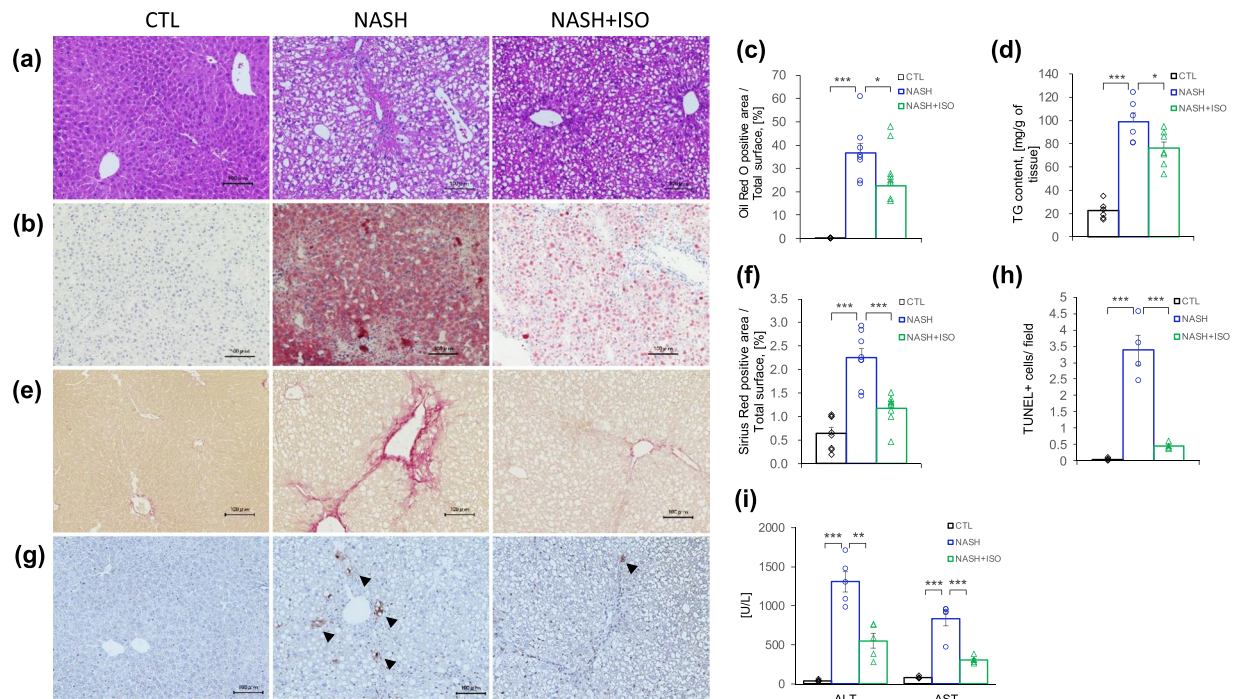


Figure 2. Effect of isorhamnetin on histopathology of NASH-induced mouse liver. Representative microscopic images of HE-stained (a), and oil red O-stained (b) liver sections (scale bar = 0.1 mm). (c) Quantification of oil red O positive area (n = 8/group, NASH vs. NASH + ISO group: $p = 0.017$). (d) Quantification of triglycerides in liver (mg/g of tissue, n = 6/group in CTL and NASH, n = 7 in NASH + ISO group, NASH vs. NASH + ISO: $p = 0.029$). (e) Representative microscopic images of Sirius red-stained liver sections (scale bar = 0.1 mm). (f) Quantification of Sirius red positive area of liver section (n = 8/group). (g) Representative microscopic images of apoptotic cells detected by TUNEL staining in liver sections (scale bar = 0.1 mm). Black arrow show TUNEL positive cells. Hematoxylin counterstaining. (h) Number of TUNEL positive cells per x20 field (n = 4/group). (i) Serum level of ALT and AST measured by automated biochemical analyzer (n = 5/group, ALT value of NASH vs. NASH + ISO: $p = 0.0017$) Data are shown as mean \pm SEM with significance * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$.

Isorhamnetin ameliorated liver histopathology of mice with NASH. Histopathological evaluation remains the standard method to diagnose and evaluate NASH. Therefore, liver steatosis was evaluated with Hematoxylin and Eosin (HE) and oil red O staining (Fig. 2a,b). Compared to the CTL group, NASH-induced liver exhibited severe accumulation of lipids exceeding 37% of oil red O positive area, whereas isorhamnetin treatment mitigated it until 22% (Fig. 2e). Concordant with the ameliorated steatosis, the triglycerides (TG) content in liver was also significantly reduced in NASH + ISO group compared to NASH group (76 mg/g and 99 mg/g of liver weight, respectively, $p = 0.029$) (Fig. 2f). This finding correlates with the reduced liver to body weight ratio observed in NASH + ISO mice.

One of the NASH features that distinguish it from benign steatosis is hepatic fibrosis characterized by excessive deposition of extracellular matrix (ECM). To address this hallmark, we evaluated collagen deposited area stained with Sirius red-stained (Figs 2c, and S1). NASH-induced liver showed greater collagen deposition surrounding portal vein than in those of CTL group. Interestingly, NASH-induced mice, when treated with isorhamnetin, had diminished collagen. The morphometric analysis further confirmed that isorhamnetin treatment reduced deposition of collagen from 2.25 to 1.17% of total surface ($p < 0.001$ vs. NASH) (Fig. 2g), indicating that isorhamnetin treatment efficiently improved fibrotic condition of NASH-induced liver.

Consistent damage in the liver leads to hepatocyte death by apoptosis and necrosis when the liver fails to respond adequately to this pathologic condition²⁵, and increasing number of apoptotic cells contribute to exacerbating the progression of fibrosis²⁶. Serum levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST), enzymes indicating liver injury state, were increased significantly in NASH-induced mice compared to CTL ($p < 0.001$ for both enzymes). However, NASH + ISO group had significantly decreased ALT and AST enzyme levels when compared with NASH group ($p < 0.01$ and $p < 0.001$ vs. NASH, respectively), suggesting that NASH-induced liver was less injured when treated with isorhamnetin (Fig. 2i). Moreover, highest number of apoptotic cells was found in NASH-induced mice compared to other two groups revealed by TUNEL assay (Fig. 2d,h). In addition, isorhamnetin treatment on hepatocyte and hepatic stellate cell lines did not affect the cell viability after 24 h (Fig. S2,a,b). Lipid accumulation induced by oleic and palmitic acid in HepG2 cells was not affected by isorhamnetin treatment (Fig. S2,c). These findings collectively suggest that isorhamnetin treatment alleviated histopathologic condition of NASH-induced liver by reducing steatosis, fibrosis, and number of apoptotic cells.

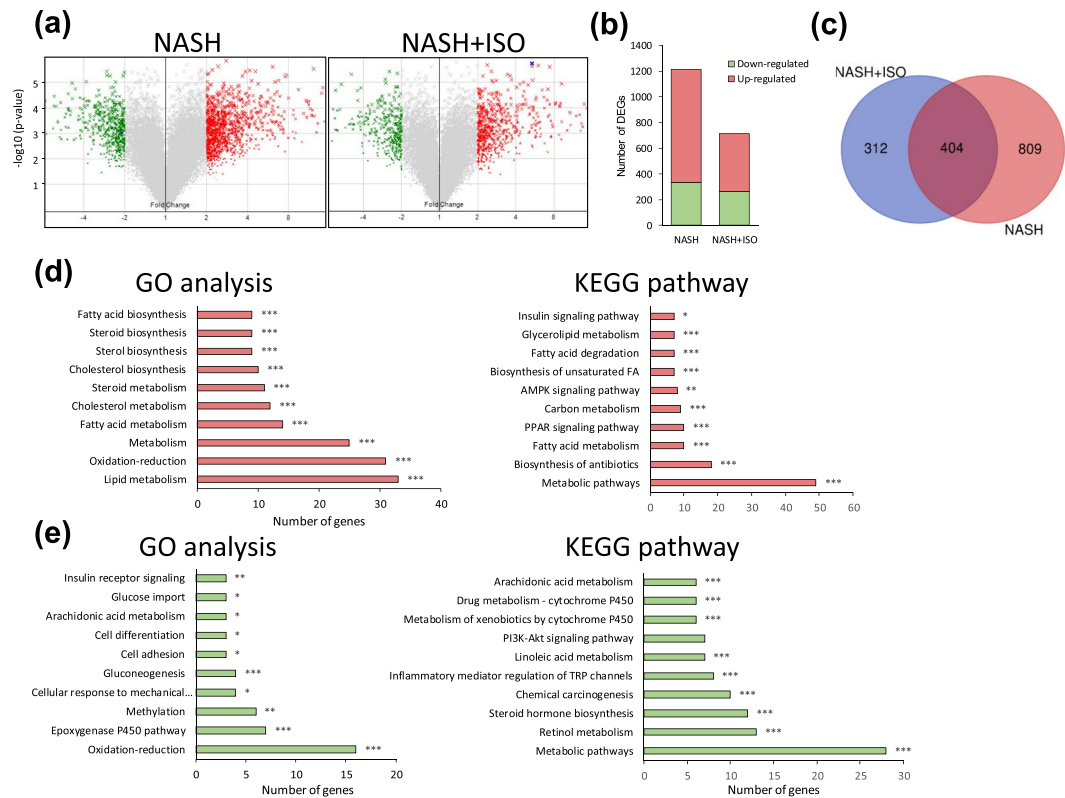


Figure 3. Hepatic gene expression profile in NASH and NASH + ISO group compared to CTL group. **(a)** Volcano plot of DEGs with fold-change over ± 2.0 (green: down-regulated, and red: up-regulated genes) between NASH vs. CTL, and NASH + ISO vs. CTL. **(b)** Quantitation of number of DEGs. **(c)** Venn diagram showing the overlap of DEGs between NASH vs. CTL, and NASH + ISO vs. CTL. GO process analysis and KEGG pathway for the up-regulated 257 genes **(d)** and down-regulated 147 genes **(e)** from overlapping 404 DEGs with p-value as * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

Gene expression profile and related pathways following NASH-induction and changes after isorhamnetin treatment. To clarify the expression pattern of genes in the liver following the induction of NASH and its changes regulated by isorhamnetin treatment, we have performed a microarray gene expression analysis. Among the 45078 probe sets, the number of genes with fold change greater than ± 2.0 and p value threshold of less than 0.05 compared to CTL were considered as differentially expressed genes (DEGs) as shown in volcano plot (Fig. 3a). The number of upregulated DEGs in NASH was remarkably reduced in NASH + ISO (Fig. 3b). The 404 overlapping genes were found between NASH and NASH + ISO (Fig. 3c). In gene ontology (GO) analysis, the upregulated genes resulting from the overlapped genes were greatly associated with lipid metabolism (GO:0006629), oxidation reduction process (GO:0055114), metabolic (GO:0008152) especially fatty acid (GO:0006631) and cholesterol (GO:0008203) metabolic processes (Fig. 3d), while downregulated genes were related to oxidation-reduction (GO:0055114), epoxygenase P450 pathway (GO:0019373), and methylation process (GO:0032259) (Fig. 3e). Along with the GO process, the upregulated genes were enriched in the metabolic pathways (mmu01100), biosynthesis of antibiotics (mmu01130), fatty acid metabolism (mmu01212), and PPAR signaling pathway (mmu03320) in Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis (Fig. 3d). In addition, metabolic pathways, retinol metabolism (mmu00830), and steroid hormone biosynthesis (mmu00140) pathways enriched by downregulated genes were top-ranked in KEGG pathway (Fig. 3e). These data show that isorhamnetin treatment reduced the number of altered gene expressions due to the induction of NASH.

Changes in lipid metabolic process and inhibition of *de novo* lipogenesis. The substantial numbers of genes were upregulated in lipid metabolism with the development of NASH as revealed by the GO analysis (Fig. S3). Thus, we analyzed the genes (58 genes in total) identified by heat map comparing NASH vs. CTL and NASH + ISO vs. CTL (Fig. 4a). Interestingly, the reduced level of expression for 42 genes was found in NASH + ISO compared to NASH. Next, we sought to distinguish genes by pathway axis which is involved in lipid metabolic process. As expected, the essential gene expressions in fatty acid metabolism, steroid biosynthesis, and PPAR signaling pathway were invariably decreased in NASH + ISO, while the median change of gene expression level in fatty acid degradation was not different between groups although a slight decrease in genes associated with fatty acid degradation was observed in NASH + ISO group (Fig. 4b and Supplementary Dataset). Moreover, *de novo* lipogenesis (DNL) is known to contribute nearly 30% of lipid accumulation in liver^{27,28}. Thus, we evaluated the individual genes identified as the key regulators in DNL pathway such as Sterol regulatory element binding protein 1 (SREBP1c), fatty acid synthase (FAS), and acetyl-Coenzyme A carboxylase alpha (ACCI)^{27,29}. We found

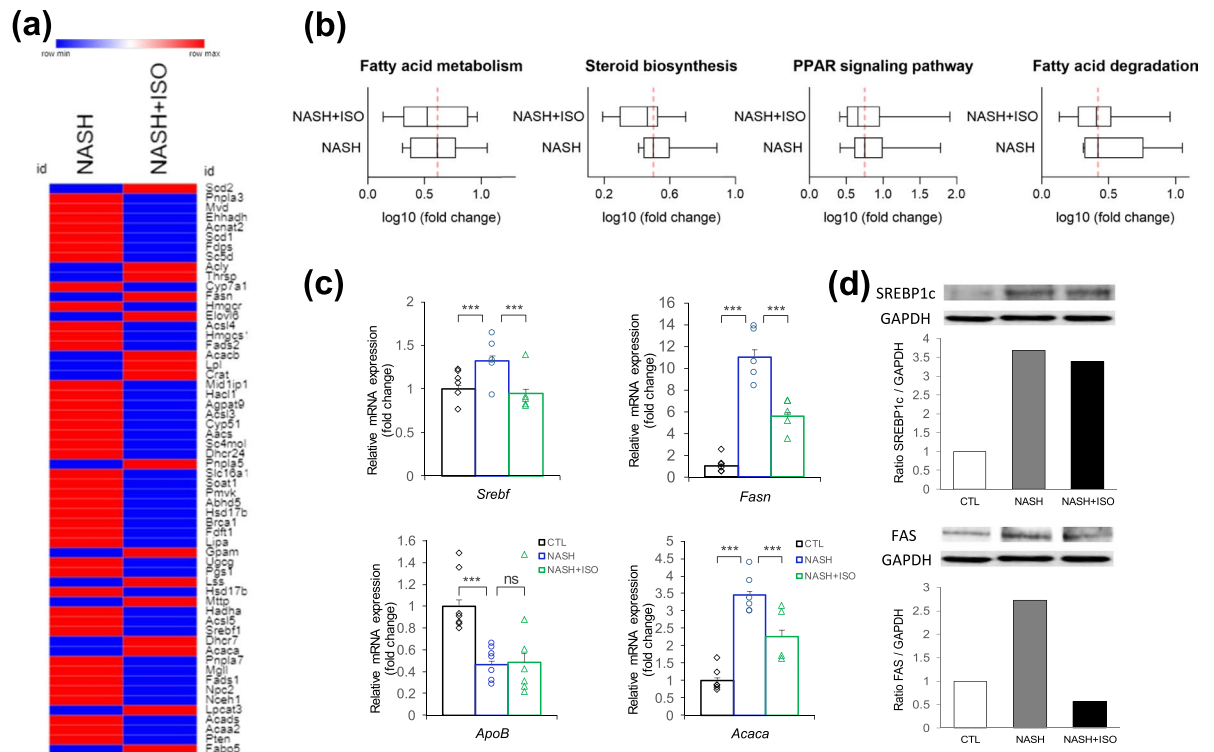


Figure 4. Changes in lipid metabolic process. **(a)** Heat map showing DEGs involved in lipid metabolic pathways between NASH and NASH + ISO compared to CTL group. **(b)** Box plot with median averaging genes involved in fatty acid metabolism (13 genes averaged), steroid biosynthesis (9 genes averaged), PPAR signaling pathway (10 genes averaged), and fatty acid degradation (7 genes averaged). Red dash line shows the median of NASH group **(c)** Relative mRNA expression level of de novo lipogenic genes ($n = 4-6/\text{group}$) by qPCR analysis. Data are shown as mean \pm SEM with significance $***p < 0.001$. **(d)** Immunoblot analysis for SREBP1c and Fatty acid synthase (FAS) in liver tissue and quantification of band intensities normalized to GAPDH.

that mRNA expression of SREBP1c, FAS, consistent with the corresponding protein level (Fig. 4d), and ACC1 was significantly upregulated ($p < 0.001$) in NASH-induced liver compared to CTL group, while SREBP1c-mediated DNL pathway was considerably inhibited ($p < 0.001$ for all genes) in NASH + ISO group when compared with NASH group (Fig. 4c). Decreased level of apolipoprotein B (*ApoB*), a rate-determining protein of lipid export in liver, is observed in NASH. The *ApoB* gene expression level was significantly reduced in NASH group compared to CTL but the level in NASH + ISO was similar to that of NASH group (Fig. 4c). These findings collectively show that the substantially altered lipid metabolic process in NASH was efficiently improved, and DNL was in part inhibited by the isorhamnetin treatment.

Changes in oxidation reduction process and activation of HSCs. Oxidative stress is well known to be one of the main drivers in the development of fibrosis and cell death by causing cell damage and liver injury^{30,31}. In response to hepatic insults and increased secretion of pro-inflammatory cytokines such as transforming growth factor beta ($\text{TGF}\beta$), HSCs become activated from the quiescent state which leads to excessive deposition of extracellular matrix (ECM) and eventually scarring. Expression level of genes implicated in oxidation reduction process was greatly elevated in NASH (Fig. S3). When all upregulated redox gene expressions (57 genes in total) occurred in NASH were compared with NASH + ISO in heat map, important proportion of genes (41 genes) expression was decreased in NASH + ISO (Fig. 5a). To yield a clearer understanding of the effect of isorhamnetin on redox-mediated processes, we further analyzed genes implicated in HSCs activation, proliferation, Nrf2-mediated oxidative stress and apoptosis. The mean expression level of genes involved in HSC activation such as transforming growth factor beta receptor 1 (*Tgfb1*) and several types of collagen was significantly greater in NASH, and decreased to near CTL level in NASH + ISO, indicating the activation of HSCs was decreased after isorhamnetin treatment (Fig. 5b and Supplementary Dataset). Among 41 genes averaged in proliferation process such as several genes encoding cell division cycle-associated proteins (*Cdca2*, *Cdca3*, *Cdca8*), globally upregulated genes were overrepresented in NASH, while downregulated in NASH + ISO. Nrf2-mediated oxidative stress markers including 38 genes were slightly ameliorated in NASH + ISO compared to NASH. Similarly, genes included in regulation of apoptosis encoding such as TNF-receptor superfamily proteins (*Tnf*, *Tnfrsf11b*, *Tnfrsf12a*, *Tnfrsf21*), Fas cell surface death receptor (*Fas*), and caspases (*Casp2*, *Casp6*, *Casp7*) were higher in NASH compared to NASH + ISO. $\text{TGF}\beta$ is known to mediate hepatic fibrosis by activating HSCs which result in increased ECM production³². Thus, we measured individual mRNA expression level of $\text{TGF}\beta$, and collagen type I. The results indicated that the NASH + ISO group showed consistently decreased mRNA levels compared to

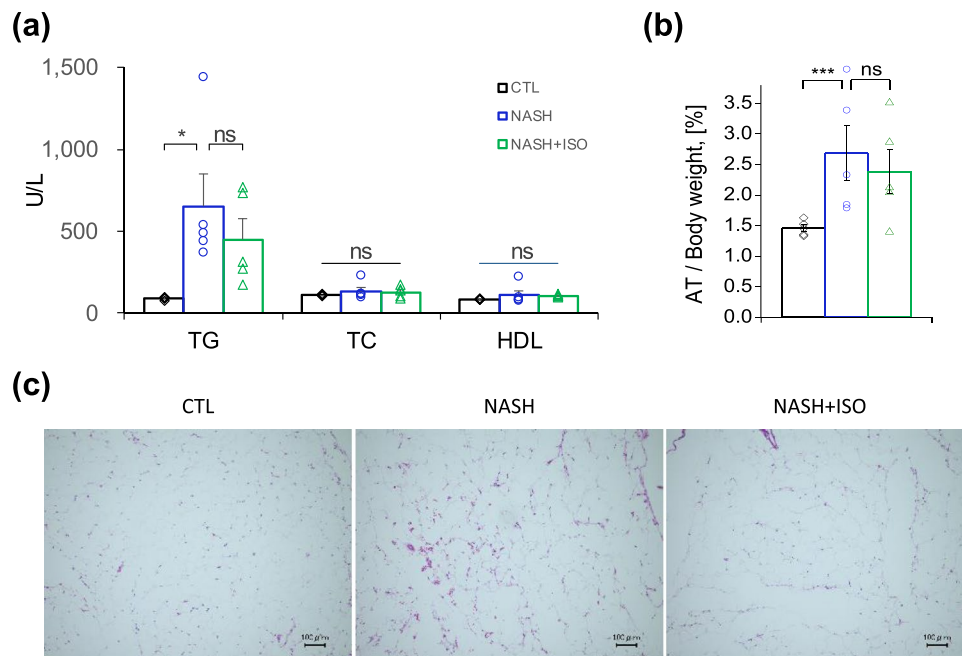


Figure 6. Effect of isorhamnetin on adipose tissue and lipid profile in serum. **(a)** Serum level of TG, TC, and HDL measured by automated biochemical analyzer ($n = 5/\text{group}$, CTL vs. NASH: $p = 0.0215$). **(b)** Adipose tissue on body weight ratio ($n = 7/\text{group}$). Data are shown as mean \pm SEM with significance * $p < 0.05$, *** $p < 0.001$ and ns = non-significant. **(c)** Representative microscopic images of HE-stained adipose tissue sections (scale bar = 0.1 mm).

Discussion

Researchers still have been seeking an effective drug although the underlying molecular mechanism responsible for etiology of NASH is not completely elucidated yet. Several studies showed curative and preventive effects of natural flavonoids against the liver diseases^{20,33}. A recent rodent NASH model that we used in our study can be induced within four weeks and was able to show the set of main hallmarks present in human NASH, including liver steatosis, injury and apoptosis, and fibrosis. This advantage allowed us to study the effect of isorhamnetin not only in liver itself but also in systemic level. In this study, we demonstrated that oral administration of isorhamnetin could consistently alleviate pathologic features of NASH when compared to non-treated NASH-induced mice with the intention of finding a beneficial dietary supplement to drug development.

Liver steatosis, inflammation, and fibrosis are interrelated and are likely to aggravate each other by positive feedback leading to progression of NASH³⁴. Lipid accumulation in the liver precedes the onset of NASH arising from three main sources: around 60% from fat-rich diet, 10–20% from lipolysis of adipose tissue, and 20–30% from hepatic DNL^{27,35}. In current clinical practice, diet source and lipolysis of adipose tissue can be controlled up to certain extent by managing diet regime combined with pharmacological strategies. In NAFLD patients, DNL pathway in liver is constantly activated because of insulin resistance and contributes to exacerbation of hepatic steatosis^{36–38}. Main regulator of DNL pathway is insulin-induced SREBP1c which triggers its downstream gene expressions required for fatty acid synthesis. SREBP1c is activated following the cleavage in Golgi and matured in endoplasmic reticulum. Overactivation of SREBP1c induces endoplasmic reticulum stress which associates with liver steatosis due to increased oxidative stress³⁹. Hepatic lipid content was reduced by isorhamnetin treatment from 37% to 23% which brings NASH-induced liver back to mild steatosis category (5–33% as mild, and 34–66% as moderate) according to steatosis grade of NAS score⁴⁰. We found that isorhamnetin treatment of NASH-induced mice could attenuate DNL pathway by regulating lipogenic key transcription factor, SREBP1c, which, in turn, downregulated lipogenic enzymes. This result correlates with the histological findings of reduced hepatic TG content and intrahepatic lipid accumulation in isorhamnetin-treated NASH mice compared to untreated NASH mice. It is seemingly possible that the inhibition of DNL pathway may be resulted in the alleviation of liver steatosis by 20–30% because the mRNA expression level of ApoB was not differed by the isorhamnetin treatment. ApoB is a protein synthesized in liver and is required for the formation of very-low-density lipoprotein (VLDL), which exports TG from liver and delivers to body tissues, including adipose tissue. Previous studies demonstrated that hepatic ApoB production is decreased in NASH patients resulting in diminished secretion of VLDL and increased hepatic steatosis⁴¹.

Yu Zhang *et al.* showed that four weeks of isorhamnetin treatment could reduce the body weight significantly in diet-induced NAFLD mice and leptin-deficient obese mice model⁴². Moreover, previous metabolic studies have also observed the difference in body weight among groups after at least four weeks of treatment with isorhamnetin^{42,43}. Thus, in our study, two weeks of treatment was more likely premature to observe a detectable difference in body weight and liver weight between NASH and NASH + ISO although the liver weight was slightly reduced in NASH + ISO group. On the other hand, at systemic level isorhamnetin was also reported to improve

glucose metabolism and insulin sensitivity, and protect against lipid peroxidation in diet-induced obese mice and streptozotocin-induced diabetic rats⁴⁴. We found that the liver injury markers were greatly decreased following the isorhamnetin treatment.

The mechanism that underlies hepatic fibrogenesis is primarily initiated by the activation of HSCs in response to hepatic insults. TGF β , a potent fibrogenic cytokine, plays a pivotal role in the activation of HSC by triggering its effector molecules to stimulate fibrogenic gene expressions and to induce collagen production. The ensembles of genes implicated in HSC activation, proliferation, and apoptosis were downregulated in NASH + ISO accompanied by the downregulation of genes involved in Nrf-mediated oxidative stress. In addition, mRNA expression of TGF β and collagen by isorhamnetin were reduced suggesting that isorhamnetin may also exert its biological activity on HSCs which are the main sources of TGF β secretion and TGF β -mediated production of collagen in ECM. Yang *et al.* reported that isorhamnetin isolated from *oenanthe javanica* exerted anti-fibrotic effect in mice liver with CCl₄-induced fibrosis by preventing the activation of TGF β -induced smad2/3 pathway¹⁹. In our study, we did not exclude possible inflammatory insults from adipose tissue and hepatic steatosis-related intra-hepatic deregulation of gene expression since the ‘second hits’ possibly act as positive feedback to exaggerate ‘first hits’. In this study, we showed that isorhamnetin could prevent the activation of TGF β -mediated fibrogenesis in NASH-induced mice. Additionally, the release of apoptotic bodies derived from injury-induced parenchymal cell apoptosis, activation of immune cells due to systemic inflammation, signaling from Kupffer cells, and lipid peroxidation are considered as fibrogenic factors leading to HSCs activation⁴⁵. Chronic fibrotic state and hepatic cell death by apoptosis are positively correlated with the severity of NASH^{26,46}. We have found that gene expressions related to apoptosis and the number of apoptotic cells in liver were greatly reduced in the treated group. These results suggest that the isorhamnetin treatment may reverse in longer-term fibrosis and liver injury in NASH by mitigating systemic inflammation as well as by preventing HSCs activation.

Obesity, insulin resistance, and type 2 diabetes are all considered as risk factors for the development of NAFLD and NASH, which are primarily characterized by an ectopic accumulation of lipid in liver. Adipose tissue, especially visceral one, is known to be responsible for elevated lipolysis and systemic inflammation due to insulin resistance resulting in hepatic lipid accumulation and inflammation³⁴. Although the lipid profile measured in non-fasting serum demonstrated insignificant difference between treated and non-treated NASH-induced mice, adipose tissue of NASH-induced mice was more inflamed, as evidenced by the increased number of macrophage infiltration, while adipocytes of NASH + ISO were greatly ameliorated. Of note, similar studies that used flavonoids to treat HFD-induced metabolic disorders in rodents also noted indifference of lipid profile in serum²¹ but found the net amelioration of systemic inflammation accompanied with reduced adipose tissue and body weight after longer duration of treatment in diet-induced mice or *db/db* mice^{18,42}.

Altogether, our results demonstrated that isorhamnetin elicits a beneficial effect on hallmarks of NASH by improving steatosis, injury, and fibrosis in a human-like NASH-induced mouse model. This hepatoprotective effect of isorhamnetin was correlated to the inhibition of DNL and fibrogenic gene expressions; the alleviation of liver TG content; and the diminution of hepatic collagen deposition accompanied with the reduced number of apoptotic hepatocytes. Thus, isorhamnetin can be a novel candidate for the consideration of additional compound in NASH drug development.

Material and Methods

Animals. Six-week-old male C57BL/6J mice (Charles River Laboratories JAPAN Inc., Kanagawa, Japan) were maintained at room temperature in a 12 h light/dark cycle. After one week of acclimatization with standard chow diet and tap water *ad libitum*, all animals were randomly assigned (Day 0) into three experimental groups (n = 8/group): Control (CTL), NASH, and NASH treated with isorhamnetin (NASH + ISO). All animal procedures were approved by the Animal Study Committee of University of Tsukuba (No.17–312) and were handled according to the Guiding Principles for the Care and Use of Animals in the Field of Physiological Sciences approved by The Physiological Society of Japan.

Chemicals. Isorhamnetin was synthesized from commercially available quercetin (Fujifilm Wako Pure Chemical Corp., Tokyo, Japan) according to the protocol⁴⁷ and used for *in vivo* experiment. Commercially available selective agonist for LXR α and LXR β (T0901317) (Cayman Chemical, Ann Arbor, MI, USA); and carbon tetrachloride (CCl₄) (Wako Pure Chemical Industries Ltd., Osaka, Japan) were used.

Experimental design and procedure. NASH was induced according to the protocol⁸. Briefly, the CTL group mice were fed with laboratory chow diet and received vehicles. NASH and NASH + ISO groups were subjected to high fat diet (60% cal from fat, D12492, Research Diets Inc., New Brunswick, NJ, USA) from Day 0–24. Rodents received intraperitoneal injections of CCl₄ at 0.1 ml/kg of body weight 4 times (Day 14, 17, 21, and 24), and T0901317 at 2.5 ml/kg of body weight 5 times (Day 20–24) to induce NASH. Corn oil and dimethyl sulfoxide (DMSO) were used as a vehicle for CCl₄ and T0901317 injections respectively. NASH + ISO group was treated with daily oral administration of isorhamnetin at 50 mg/kg of body weight (vehicle for CTL and NASH group) for last two weeks (Day 11–24). Body weight and food intake were measured every day. At the end of experiment (Day 25) blood was collected from retro-orbital sinus using capillary tube after mice are slightly anesthetized with isoflurane inhalation. Serum was separated by centrifugation at 3000 rpm for 10 min and stored at –20 °C until biochemical analysis. Liver and epididymal fat were isolated immediately after the exsanguination. Median lobe of liver was stocked in either liquid nitrogen or cryopreserves for further analysis. The remaining liver was fixed in 10% neutral buffered formalin for histological analysis.

Blood biochemical analysis. The TC, TG, HDL, ALT, and AST levels in serum were measured using FUJI DRI-CHEM 7000 automated chemistry analyzer (Fujifilm Corp. Tokyo, Japan).

Histological analysis. Tissues embedded in paraffin were sliced into sections and stained with H&E, and Sirius red using the standard protocol for histopathology analysis of liver. Cryopreserved liver tissues were sliced and used for oil red O staining with hematoxylin counterstaining. Stained slides were observed under a BZ-9000 BioRevo digital microscope (Keyence Corp., Osaka, Japan), and were analyzed using ImageJ.

Apoptosis TUNEL assay. Apoptotic cells *in situ* were detected in paraffin-embedded liver sections using DeadEnd Colorimetric TUNEL System (Promega Corp., Madison, WI, USA) according to the manufacturer's instruction. Sections were counter stained with hematoxylin. Stained slides were observed under a BZ-9000 BioRevo digital microscope (Keyence Corp., Osaka, Japan), and were analyzed using ImageJ.

Gene expression analysis. The total RNA of liver tissues was extracted using ISOGEN reagent (Nippon Gene Co., Ltd. Toyama, Japan) according to the manufacturer's protocol. First-strand cDNA was amplified from the total RNA (100 ng) using SuperScript III Reverse Transcriptase kit (Invitrogen, Carlsbad, CA, USA). The quantification of total RNA and cDNA was measured on NanoDrop 2000 spectrophotometer (Thermo Scientific, Wilmington, DE, USA). Real-time quantitative PCR of target gene expression was assayed by TaqMan predesigned primers (Applied Biosystems, Foster City, CA, USA) and TaqMan Gene Expression Master Mix (Life Technologies, Carlsbad, USA) using the 7500 Fast Real-Time PCR System (Applied Biosystems, Foster City, CA, USA). The following primers were bought from Applied Biosystems: sterol regulatory element binding protein 1 (*Srebfl*) (Mm00550338_m1), fatty acid synthase (*Fasn*) (Mm00662319_m1), acetyl-Coenzyme A carboxylase alpha (*Acaca*) (Mm01304257_m1), transforming growth factor beta 1 (*Tgfb1*) (Mm01178820_m1), collagen type 1 alpha 1 (*Col1a1*) (Mm00801666_g1), apolipoprotein B (*Apob*) (Mm01545150_m1), and glyceraldehyde-3-phosphate dehydrogenase (*Gapdh*) (Mm99999915_g1). The $2^{-\Delta\Delta C_t}$ method was applied to calculate the relative mRNA expression levels using *Gapdh* as a housekeeping endogenous control.

Total RNA extraction and microarray analysis of liver. The total RNA of liver tissues was extracted using ISOGEN reagent (Nippon Gene Co., Ltd. Toyama, Japan) according to the manufacturer's protocol. The quality and quantity of total RNA was measured on NanoDrop 2000 spectrophotometer (Thermo Scientific, Wilmington, DE, USA), and 100 ng of RNA input was used for microarray analysis. The gene expression profiling was performed using GeneChip 3' IVT PLUS Reagent Kit (Affymetrix Inc., Santa Clara, CA, USA) with Affymetrix® 3' IVT Array Strips for GeneAtlas® System (GeneChip® MG-430 PM) according to the manufacturer's user guide. Raw intensity values were obtained using GeneAtlas™ Imaging Station and were normalized using Expression Console Software provided by the Affymetrix following robust multichip average (RMA) algorithm (<http://www.affymetrix.com>). Subsequently, the raw data set was transferred to the Transcriptome Analysis Console (TAC) v4.0 (ThermoFisher inc.). A threshold with fold-change ≥ 2.0 and $p < 0.05$ compared to CTL were considered as DEGs. Further analyses were conducted using Functional annotation tool of The Database for Annotation, Visualization and Integrated Discovery (DAVID) v6.8 online bioinformatics database to identify enriched Gene Ontology (GO) and KEGG pathways. Transcriptome dataset is available at NCBI Gene Expression Omnibus database (www.ncbi.nlm.nih.gov/geo) with accession number GSE137635. Heat maps were visualized with DEGs in each biological process identified by GO using Morpheus online tool (<https://software.broadinstitute.org/morpheus>). Box plots analyses were visualized using GraphPad Prism 8 which show the average of genes implicated in each process (grouping of genes by biological process as mentioned in Cazanave *et al.*⁴⁸).

Western blotting. Total protein was extracted from liver tissue (50 mg) using radio immunoprecipitation assay (RIPA) buffer (Sigma-Aldrich, St. Louis, USA) containing protease inhibitor cocktail (P8340, Sigma-Aldrich, St. Louis, MO, USA), and quantified with 2-D Quant kit (GE Healthcare, Chicago, USA) following the manufacturer's instruction. Protein samples (20 μ g) were resolved in 10% SDS-PAGE and transferred to nitrocellulose membrane. After blocking in Odyssey blocking buffer (LI-COR, NE, USA) for 2 hours, membranes were blotted with primary antibody for overnight at 4°C and incubated with appropriate Alexa Fluor® conjugated secondary antibodies. Imaging and quantification of signal intensity were detected using the Odyssey Fc Imaging System (LI-COR, NE, USA). Primary antibodies: rabbit anti- α SMA (ab5694), rabbit anti-FAS (ab22759), rabbit anti-SREBP1 (ab28481), mouse anti-GAPDH (ab8245), and secondary antibodies: Alexa Fluor® 488-conjugated donkey anti-rabbit (ab150073), Alexa Fluor® 594-conjugated donkey anti-mouse (ab150108) were purchased from Abcam. Uncropped images of western blotting gel are available in the Supplementary Files.

Quantification of liver triglyceride. Total TG content in the liver was quantified using commercially available colorimetric kit (Cayman Chemical, Ann Arbor, MI, USA).

Statistical analysis. All data are expressed as the mean \pm SEM, unless otherwise mentioned. Normality test was performed to confirm the normal distribution. A one-way analysis of variance (ANOVA) followed by Tukey's posthoc test was applied to assess the statistical significance of difference among the treatment groups. Statistical outliers were excluded from data analysis. A value of $p < 0.05$ was considered significant for all results. Statistical analyses were performed using IBM SPSS Statistics version 24.0.

Data availability

All data generated or analyzed during this study are included in this published article and its Supplementary Files. Microarray data are deposited in the Gene Expression Omnibus (GEO) under Accession Number: GSE137635.

Received: 8 January 2019; Accepted: 21 October 2019;

Published online: 07 November 2019

References

1. Younossi, Z. M. *et al.* Global epidemiology of nonalcoholic fatty liver disease—Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* **64**, 73–84 (2016).
2. Serfaty, L. Management of patients with non-alcoholic steatohepatitis (NASH) in real life. *Liver Int.* **38**, 52–55 (2018).
3. Sanyal, A. J. & Miller, V. Regulatory Science and Drug Approval for Alcoholic and Nonalcoholic Steatohepatitis. *Gastroenterology* **150**, 1723–1727 (2016).
4. Leclercq, I. A., Lebrun, V. A., Stärkel, P. & Horsmans, Y. J. Intrahepatic insulin resistance in a murine model of steatohepatitis: Effect of PPAR γ agonist pioglitazone. *Lab. Investig.* **87**, 56–65 (2007).
5. Stephenson, K. *et al.* Updates on dietary models of nonalcoholic fatty liver disease: Current studies and insights. *Gene Expr.* **18**, 5–17 (2018).
6. Kubota, N. *et al.* A high-fat diet and multiple administration of carbon tetrachloride induces liver injury and pathological features associated with non-alcoholic steatohepatitis in mice. *Clin. Exp. Pharmacol. Physiol.* **40**, 422–430 (2013).
7. Tsuchida, T. *et al.* A simple diet- and chemical-induced murine NASH model with rapid progression of steatohepatitis, fibrosis and liver cancer. *J. Hepatol.* **69**, 385–395 (2018).
8. Owada, Y. *et al.* Novel non-alcoholic steatohepatitis model with histopathological and insulin-resistant features. *Pathol. Int.* **68**, 12–22 (2018).
9. Hossain, M. K. *et al.* Molecular mechanisms of the anti-obesity and anti-diabetic properties of flavonoids. *Int. J. Mol. Sci.* **17** (2016).
10. Vinayagam, R. & Xu, B. Antidiabetic properties of dietary flavonoids: A cellular mechanism review. *Nutr. Metab.* **12** (2015).
11. Procházková, D., Boušová, I. & Wilhelmová, N. Antioxidant and prooxidant properties of flavonoids. *Fitoterapia* **82**, 513–523 (2011).
12. Hu, S. *et al.* Isorhamnetin inhibits cell proliferation and induces apoptosis in breast cancer via Akt and mitogen-activated protein kinase signaling pathways. *Mol. Med. Rep.* **12**, 6745–6751 (2015).
13. Lee, H. J. *et al.* Mitochondria-cytochrome C-caspase-9 cascade mediates isorhamnetin-induced apoptosis. *Cancer Lett.* **270**, 342–353 (2008).
14. Yang, J. H. *et al.* Isorhamnetin protects against oxidative stress by activating Nrf2 and inducing the expression of its target genes. *Toxicol. Appl. Pharmacol.* **274**, 293–301 (2014).
15. Dou, W. *et al.* Plant flavonol isorhamnetin attenuates chemically induced inflammatory bowel disease via a PXR-dependent pathway. *J. Nutr. Biochem.* **25**, 923–933 (2014).
16. Lee, J. *et al.* Isorhamnetin represses adipogenesis in 3T3-L1 cells. *Obes. (Silver Spring)* **17**, 226–232 (2009).
17. Zar Kalai, F. *et al.* Antiobesity effects of an edible halophyte *Nitraria retusa* Forssk in 3T3-L1 preadipocyte differentiation and in C57B6/J mice fed a high fat diet-induced obesity. *Evidence-based Complement. Altern. Med.* **2013**, 368658 (2013).
18. Zar Kalai, F., Han, J., Ksouri, R., Abdely, C. & Isoda, H. Oral administration of *Nitraria retusa* ethanolic extract enhances hepatic lipid metabolism in db/db mice model ‘BKS.Cg-Dock7m+/+ Leprdb/J’ through the modulation of lipogenesis-lipolysis balance. *Food Chem. Toxicol.* **72**, 247–256 (2014).
19. Yang, J. H. *et al.* Isorhamnetin attenuates liver fibrosis by inhibiting TGF- β /Smad signaling and relieving oxidative stress. *Eur. J. Pharmacol.* **783**, 92–102 (2016).
20. Wu, L. *et al.* Quercetin prevents hepatic fibrosis by inhibiting hepatic stellate cell activation and reducing autophagy via the TGF- β 1/Smads and PI3K/Akt pathways. *Sci. Rep.* **7**, 9289 (2017).
21. Marcolin, É. *et al.* Quercetin Decreases Liver Damage in Mice with Non-Alcoholic Steatohepatitis. *Basic Clin. Pharmacol. Toxicol.* **112**, 385–391 (2013).
22. Zhang, L. *et al.* Quercetin aglycone is bioavailable in murine pancreas and pancreatic xenografts. *J. Agric. Food Chem.*, <https://doi.org/10.1021/jf101192k> (2010).
23. Li, G. *et al.* Pharmacokinetic properties of isorhamnetin, kaempferol and quercetin after oral gavage of total flavones of *Hippophae rhamnoides* L. in rats using a UPLC-MS method. *Fitoterapia* **83**, 182–191 (2012).
24. Perez-Vizcaino, F., Duarte, J. & Santos-Buelga, C. The flavonoid paradox: Conjugation and deconjugation as key steps for the biological activity of flavonoids. *J. Sci. Food Agric.* **92**, 1822–1825 (2012).
25. Yang, L. *et al.* TGF- β signaling in hepatocytes participates in steatohepatitis through regulation of cell death and lipid metabolism. *Hepatology* **59**, 483–495 (2014).
26. Feldstein, A. E. *et al.* Hepatocyte apoptosis and Fas expression are prominent features of human nonalcoholic steatohepatitis. *Gastroenterology* **125**, 437–443 (2003).
27. Donnelly, K. L. *et al.* Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. *J. Clin. Invest.* **115**, 1343–1351 (2005).
28. Postic, C. & Girard, J. Contribution of de novo fatty acid synthesis to hepatic steatosis and insulin resistance: lessons from genetically engineered mice. *J. Clin. Invest.* **118**, 829–838 (2008).
29. Horton, J. D., Goldstein, J. L. & Brown, M. S. SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. *J. Clin. Invest.* **109**, 1125–1131 (2002).
30. Richter, K., Konzack, A., Pihlajaniemi, T., Heljasvaara, R. & Kietzmann, T. Redox-fibrosis: Impact of TGF β 1 on ROS generators, mediators and functional consequences. *Redox Biol.* **6**, 344–352 (2015).
31. Arauz, J., Ramos-Tovar, E. & Muriel, P. Redox state and methods to evaluate oxidative stress in liver damage: From bench to bedside. *Ann. Hepatol.* **15**, 160–173 (2016).
32. Bian, Z. & Ma, X. Liver fibrogenesis in non-alcoholic steatohepatitis. *Front. Physiol.* **3 JUL**, 1–7 (2012).
33. Hernández-Aquino, E. & Muriel, P. Beneficial effects of naringenin in liver diseases: Molecular mechanisms. *World J. Gastroenterol.* **24**, 1679–1707 (2018).
34. Chen, Z., Yu, R., Xiong, Y., Du, F. & Zhu, S. A vicious circle between insulin resistance and inflammation in nonalcoholic fatty liver disease. *Lipids Health Dis.* **16**, 1–9 (2017).
35. Reccia, I. *et al.* Non-alcoholic Fatty Liver Disease: A sign of systemic disease. *Metabolism.* **72**, 94–108 (2017).
36. Diraison, F., Moulin, P. & Beylot, M. Contribution of hepatic de novo lipogenesis and reesterification of plasma non esterified fatty acids to plasma triglyceride synthesis during non-alcoholic fatty liver disease. *Diabetes Metab.* **29**, 478–485 (2003).
37. Ameer, F., Scanduzzi, L., Hasnain, S., Kalbacher, H. & Zaidi, N. De novo lipogenesis in health and disease. *Metabolism* **63**, 895–902 (2014).
38. Lambert, J. E., Roman, M. A. R., Browning, J. D. & Parks, E. J. Increased De Novo Lipogenesis Is a Distinct Characteristic of Individuals With Nonalcoholic Fatty Liver Disease. *Gastroenterology* **146**, 726–735 (2014).
39. Ferré, P. & Fougère, F. Hepatic steatosis: A role for de novo lipogenesis and the transcription factor SREBP-1c. *Diabetes, Obes. Metab.*, <https://doi.org/10.1111/j.1463-1326.2010.01275.x> (2010).
40. Brunt, E. M. *et al.* The NAS and the Histopathologic Diagnosis of NAFLD: Distinct Clinicopathologic Meanings. *Hepatology* **53**, 810–820 (2011).
41. Charlton, M., Sreekumar, R., Rasmussen, D., Lindor, K. & Nair, K. S. Apolipoprotein synthesis in nonalcoholic steatohepatitis. *Hepatology* **35**, 898–904 (2002).
42. Zhang, Y. *et al.* Dietary component isorhamnetin is a PPAR γ antagonist and ameliorates metabolic disorders induced by diet or leptin deficiency. *Sci. Rep.* **6**, 19288 (2016).

43. Rodríguez-Rodríguez, C. *et al.* The effect of isorhamnetin glycosides extracted from *Opuntia ficus-indica* in a mouse model of diet induced obesity. *Food Funct.* **6**, 805–815 (2015).
44. Yokozawa, T., Kim, H. Y., Cho, E. J., Choi, J. S. & Chung, H. Y. Antioxidant effects of isorhamnetin 3,7-di-O- β -D-glucopyranoside isolated from mustard leaf (*Brassica juncea*) in rats with streptozotocin-induced diabetes. *J. Agric. Food Chem.* **50**, 5490–5495 (2002).
45. Lee, U. E. & Friedman, S. L. Mechanisms of Hepatic Fibrogenesis. *Best Pract. Res. Clin. Gastroenterol.* **25**, 195–206 (2011).
46. Chakraborty, J. B., Oakley, F. & Walsh, M. J. Mechanisms and Biomarkers of Apoptosis in Liver Disease and Fibrosis. *Int. J. Hepatol.* **2012**, 1–10 (2012).
47. Kato, K., Ninomiya, M., Tanaka, K. & Koketsu, M. Effects of Functional Groups and Sugar Composition of Quercetin Derivatives on Their Radical Scavenging Properties. *J. Nat. Prod.* **79**, 1808–1814 (2016).
48. Cazanova, S. *et al.* The Transcriptomic Signature of Disease Development and Progression of Nonalcoholic Fatty Liver Disease. *Sci. Rep.* **7**, 1–13 (2017).

Acknowledgements

This work was partially supported by the JST-JICA - Japan Science and Technology Agency (JST) and the Japan International Cooperation Agency (JICA) (SATREPS), and JST-Center of Innovation (COI) project; and MJĒED - Mongolian-Japan Engineering Education Development Project. Authors also thank Ako Takahashi for her kind help during *in vivo* test.

Author contributions

M.G., N.O., H.I.: conceptualization and methodology; F.F. and Y.Z.W.: validation; M.G.: formal analysis, data curation, writing – original draft preparation; M.G., O.Yo., O.Yu., F.F.: investigation; M.G. and F.F.: visualization; T.K. and S.Y.: resources (the synthesis of isorhamnetin); Y.W.Z., N.O., H.I.: writing – review and editing, supervision, project administration; N.O. and H.I.: funding acquisition. All authors reviewed the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41598-019-52736-y>.

Correspondence and requests for materials should be addressed to H.I.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2019