



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Review article

The underpinning biology relating to multiple sclerosis disease modifying treatments during the COVID-19 pandemic



David Baker^a, Sandra Amor^{a,b,*}, Angray S. Kang^{a,c}, Klaus Schmierer^{a,d}, Gavin Giovannoni^{a,d}

^a Blizard Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, E1 2AT; United Kingdom

^b Pathology Department, VUmc, Amsterdam UMC, Amsterdam, The Netherlands

^c Centre for Oral Immunobiology and Regenerative Medicine, Institute of Dentistry, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, United Kingdom

^d Clinical Board:Medicine (Neuroscience), The Royal London Hospital, Barts Health NHS Trust, London, United Kingdom

A B S T R A C T

Background: SARS-CoV-2 viral infection causes COVID-19 that can result in severe acute respiratory distress syndrome (ARDS), which can cause significant mortality, leading to concern that immunosuppressive treatments for multiple sclerosis and other disorders have significant risks for both infection and ARDS.

Objective: To examine the biology that potentially underpins immunity to the SARS-Cov-2 virus and the immunity-induced pathology related to COVID-19 and determine how this impinges on the use of current disease modifying treatments in multiple sclerosis.

Observations: Although information about the mechanisms of immunity are scant, it appears that monocyte/macrophages and then CD8 T cells are important in eliminating the SARS-CoV-2 virus. This may be facilitated via anti-viral antibody responses that may prevent re-infection. However, viral escape and infection of leucocytes to promote lymphopenia, apparent CD8 T cell exhaustion coupled with a cytokine storm and vascular pathology appears to contribute to the damage in ARDS.

Implications: In contrast to ablative haematopoietic stem cell therapy, most multiple-sclerosis-related disease modifying therapies do not particularly target the innate immune system and few have any major long-term impact on CD8 T cells to limit protection against COVID-19. In addition, few block the formation of immature B cells within lymphoid tissue that will provide antibody-mediated protection from (re)infection. However, adjustments to dosing schedules may help de-risk the chance of infection further and reduce the concerns of people with MS being treated during the COVID-19 pandemic.

1. SARS-Cov-2 and COVID-19 a new pandemic

COVID-19 is the pandemic disease caused by severe acute respiratory syndrome (SARS) coronavirus two (SARS-CoV-2) infection (Zhu et al., 2020a; Zhou et al., 2020). About 80% of people infected with SARS-CoV-2 develop a self-limiting illness, 20% require hospitalisation, largely due to cardiovascular issues and about 5% require critical care and potential ventilatory support (Kimball et al., 2020; Day, 2020). The mortality in those requiring ventilatory support is about 40–50% (Weiss and Murdoch 2020; Zhu et al., 2020b). Death from COVID-19 is associated with older age and comorbidities such as cardiovascular disease, smoking, lung disease, obesity and diabetes (Zhu et al., 2020a; Lippi et al., 2020; Richardson et al., 2020). Mortality in young people and those without comorbidities may be related to

excessive viral load (Lui et al 2020a; Chen et al., 2020a). Whilst the typical clinical features requiring self-isolation, and potentially hospitalization are fever, dry cough and shortness of breath related to respiratory tract infection, other symptoms such as headache and gastrointestinal symptoms may go unnoticed or under-appreciated leading to spreading of the virus (Zhu et al., 2020b; Richardson et al., 2020; Huang et al., 2020). People shed infective virus days before symptoms occur and continue to shed virus via the lungs and faeces whilst symptoms develop and resolve, often for more than 7 days after symptom onset (Lauer et al., 2020; Xu et al., 2020a; He et al.2020a).

SARS-CoV-2 is a betacoronavirus closely-related to virus that caused the SARS outbreak in 2002-2004 (Zhou et al.2020). The viral ribonucleic acid (RNA) is bound by the nucleocapsid protein and is encapsulated in a host cell membrane-derived lipid envelope containing

Abbreviations: ACE2, angiotensin converting enzyme two; ARDS, acute respiratory distress syndrome; ASC, antibody secreting cells; CNS, central nervous system; DMT, disease modifying therapies; (HSCT), haematopoietic stem cell therapy; IRT, immune reconstitution therapies; MS, multiple sclerosis; RBD, receptor binding domain; RNA, ribonucleic acid; SARS, Severe acute respiratory syndrome

* Corresponding author at: PhD, Neuropathology, Dept Pathology, Amsterdam UMC Locatie VUmc, ZH 2E 49, De Boelelaan 1117, 1081 HV Amsterdam, The Netherlands.

E-mail addresses: david.baker@qmul.ac.uk (D. Baker), s.amor@amsterdamumc.nl (S. Amor).

<https://doi.org/10.1016/j.msard.2020.102174>

Received 21 April 2020; Received in revised form 29 April 2020; Accepted 30 April 2020

2211-0348/ © 2020 Elsevier B.V. All rights reserved.

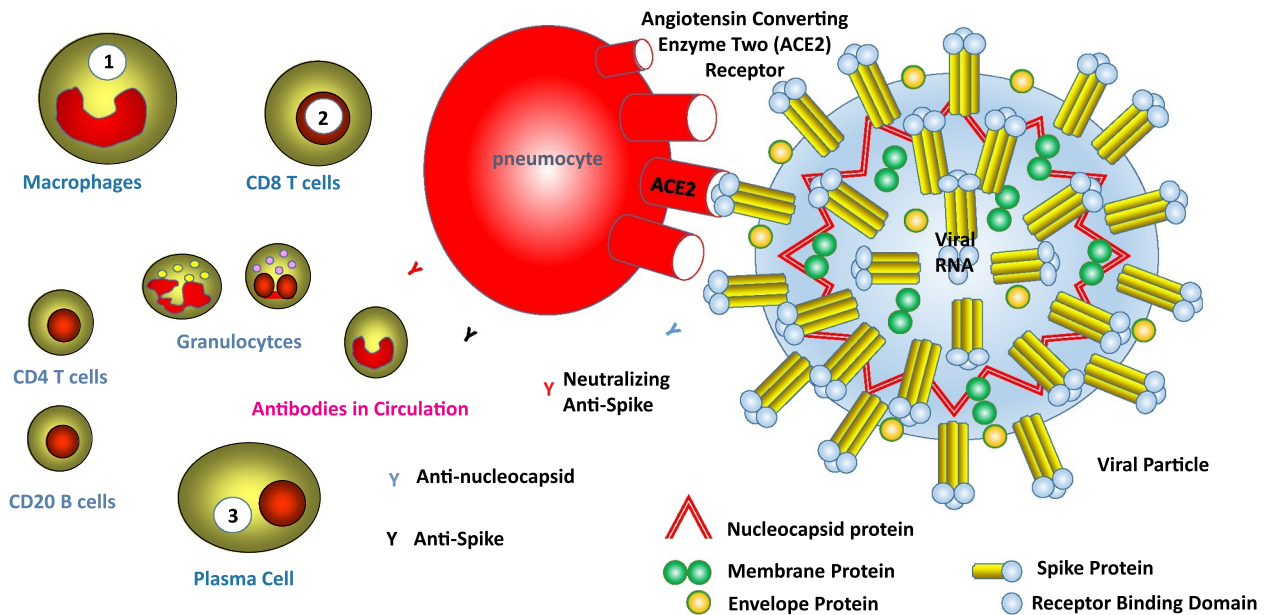


Fig. 1. The protective and destructive immune response against the SARS-CoV-2 virus.

Immune cells target the SARS-CoV-2 virus that initially involves the innate immune response, which is then supplemented with anti-viral cytotoxic T cell responses and neutralizing and binding antibodies.

the viral spike, envelope and membrane proteins (Chen et al., 2020b, Lu et al., 2020). The spike protein contains the receptor binding domain (RBD), which is important for binding to the angiotensin converting enzyme two (ACE2) cell receptor, and thus key to the cellular target, host range and viral infection (Zhou et al., 2020; Ou et al., 2020; Shi et al., 2020. Fig. 1). Viral ACE2 binding is facilitated by host cell, serine proteases such as TMPRSS2 necessary to prime the spike protein (Hoffman et al., 2020; Tai et al., 2020). The ACE2 receptor is expressed on the vasculature and is present in many tissues, such as the kidney, gut, cardiomyocytes and lung epithelia (Hamming et al., 2004; Lukassen et al., 2020). There is very low expression of ACE2 on immune cells, but other co-receptors, including: CD147, proteases and probably lectins, based on similarities with the SARS-CoV virus, may be important in SARS-CoV-2 entry (Letko et al 2020; Yang et al., 2004; Wang et al 2020a; Granberg et al., 2005).

2. Multiple sclerosis in the COVID-19 era

The immune system provides vital defence against viral infections. This has led to concern for people taking immunosuppressive agents, as immune compromised people are particularly vulnerable to infection (Coles et al., 2020; Willis and Robertson 2020; Luna et al., 2020). Infections are more common in people taking DMT and are more frequent with the higher efficacy drugs (Willis and Robertson 2020; Luna et al., 2020). This is consistent with their more potent immunosuppressive activities. Immunosuppressed people have been advised to self-shield and socially distance themselves to avoid infection and will remain a problem, until herd immunity, anti-viral agents or effective vaccines have been developed (Kwok et al., 2020; Stein 2020). Multiple sclerosis (MS) is a major neurological disease that causes disability and can require hospitalisation for uncontrolled disease activity (Compston and Coles 2008). MS is currently managed with immunomodulatory drugs (Pardo and Jones 2017). This has led neurologists to recommend maintaining the *status quo* or curtailing the use of certain disease modifying treatments (DMT) in a pragmatic or non-pragmatic way (Coles et al 2020, Giovannoni et al., 2020;

Brownlee et al., 2020. Table 1). It is understandable that a conservative “*primum non nocere*” (first do not harm) approach was adopted when considering treatments, given the paucity of knowledge surrounding SARS-CoV2 biology when COVID-19 first emerged. However, it is important to recognize the risks of poorly controlled MS may outweigh the perceived risks from COVID-19 (Giovannoni et al., 2020; Brownlee et al., 2020) and an essential goal of MS care must be to limit SARS-CoV-2 infection. Therefore, care must be to prevent disease activation and limit the need for hospitalization that could potentially increase the risk of exposure to SARS-CoV-2. This must be balanced by the requirement of hospitalization for infusion treatments and the level of monitoring that each agent requires, that is particularly arduous with alemtuzumab, but minimal with ocrelizumab and glatiramer acetate (Pardo and Jones 2017).

It is important that such recommendations about treatment are made on a rational basis using knowledge of the mode of actions of the various agents and their ability to impact on the functioning of the components of the immune system. This is important as there is no evidence that immunosuppressed people are at increased risk to coronavirus infections (D'Antiga 2020). Therefore, to understand the risks posed to people with MS using DMT, it is crucial to understand the mechanisms of action, the impact of the treatments on infection-risk, vaccination responses and the mechanisms of pathology and immunity to SARS-CoV-2. Although there are gaps in our knowledge, understanding can be gained from the study of SARS-CoV infection, as well as other coronaviruses and lower respiratory tract infections (Channappanavar et al., 2014; Prompetchara et al., 2020; Rokni et al., 2020, Sarzi-Puttini et al., 2020).

3. Immune response against SARS-CoV-2 virus

Protection against coronaviruses involves both the innate and adaptive immunity, typical for most viral infections (Yen et al., 2006; Prompetchara et al., 2020). However, consistent with SARS, some influenza infections and COVID-19, it appears to be the immune response and destruction of virally-infected cells and lung epithelial tissue that

Table 1
Initial recommendations use of MS-related DMT by some European neurological associations.

Summary of SIN/ABN Guidelines						
At risk category	Class	Trade Name	Safe to start treatment	On treatment	COVID-19 infection	Mode of action
Low	Interferon-beta	Betaferon, Avonex, Rebif, Plegridy	YES	CONTINUE	STOP	Immunomodulatory (not immunosuppressive), pleiotropic immune effects
Low	Glatiramer acetate	Copaxone	YES	CONTINUE	STOP	Immunomodulatory (not immunosuppressive), pleiotropic immune effects
Low	Teriflunomide	Aubagio	YES	CONTINUE	STOP	Dihydro-oxotetrahydro-pyrimidine dehydrogenase inhibitor (reduced de novo pyrimidine synthesis), anti-proliferative
Low	Dimethyl fumarate	Tecfidera	YES	CONTINUE	STOP	Pleiotropic, NRF2 activation, downregulation of NFKβ
Low	Natalizumab	Tysabri	YES	CONTINUE	STOP	Anti-VLA4, selective adhesion molecule inhibitor
Low	S1P modulators	Fingolimod (Gilenya)	YES	CONTINUE	STOP	Selective S1P modulator, prevents egress of lymphocytes from lymph nodes
Intermediate	Anti-CD20	Ocrelizumab (Ocrevus)	NO (YES)	SUSPEND	DELAY	Anti-CD20, B-cell depleter
High*	Cladribine	Mavenclad	NO	SUSPEND	DELAY	Deoxyadenosine (purine) analogue, adenosine deaminase inhibitor, selective T and B cell depletion
High*	Alemtuzumab	Lemtrada	NO	SUSPEND	DELAY	Anti-CD52, non-selective immune depleter
High*	HSCT	-	NO	-	DELAY	Non-selective immune depleter

* risk refers to acquiring infection during the immunodepletion phase. Post immune reconstitution the risk is low.

Composite guidelines generated from recommendations to treat MS from the Society of Italian Neurologists (SIN) and the Association of British Neurologists (Coles et al. 2020).

cause the acute respiratory distress syndromes (ARDS) and the, sometimes fatal, pneumonia (Chen et al., 2020b; Zhang et al., 2020a). It appears that the immune response to SARS-CoV-2 occurs in two phases involving an immune and a tissue, often lung, damaging phase.

3.1. Immune phase

Following infection there is an asymptomatic period of 4–5 days, although some reports indicate this can be up to 3 weeks (Pung et al., 2020; Lauer et al., 2020; Lai et al., 2020), during which time the virus attempts to escape immune surveillance through the inhibition of interferon production and blockade of interferon receptor signalling activity, similar to SARS-CoV (Prompetchara et al., 2020; Chu et al., 2020; O'Brien et al., 2020). There is an early immune response where the innate and then the adaptive immune response eliminates the virus as seen in non-human primates and by inference in humans (Bao et al., 2020; Thevarajan et al., 2020). Given that the majority of infections are asymptomatic (Kimball et al., 2020; Day 2020) indicates that this is a dominant mechanism in most people with COVID-19. *In vitro* data suggest an early innate response, notably from the alveolar macrophages and/or monocytes that may be recruited from the circulation (Yen et al., 2006; Thevarajan et al., 2020). Histological studies of cancerous lungs of people subsequently positive for COVID19, exhibited significant macrophage activity (Cai et al., 2020; Tian et al., 2020a). Thus, macrophages rather than neutrophils appear to be important as an early defence mechanism in SARS and COVID-19 lesions (Prompetchara et al., 2020; Tian et al., 2020a; Cai et al., 2020). This is probably followed by a CD8 cytotoxic T cell response that is generated within days of infection (Channappanavar et al., 2014; Prompetchara et al., 2020; Thevarajan et al., 2020).

SARS-CoV-2 may be eliminated before significant blood antibody titres are generated as seen in non-human primate infections and case reports (Fig.2. Thevarajan et al., 2020; Bao et al., 2020; Soresina et al., 2020). These antibody responses are generated around 12 days (IgM) and 14 days (IgG), although this is earlier in some individuals (Zhoa et al. 2020; Okba et al., 2020; Xiang et al., 2020). Antibodies are predominantly generated against the nucleocapsid and spike proteins (Okba et al., 2020; de Assis et al., 2020). Antibodies against the RBD of the spike protein are clearly neutralizing, are able to prevent infection (Okba et al., 2020; Tai et al., 2020; Tian et al., 2020b). These appear to be protective, as evidenced by the use of convalescent sera to protect against severe COVID-19 (Duan et al., 2020; Pei et al., 2020; Shen et al., 2020). People with X-

linked agammaglobulinemia have been infected and survived COVID-19 (Soresina et al., 2020). This further suggests that B cells and immunoglobulin may not be an obligate immune element required for protection against SARS-CoV-2 infection. Although CD8 T cells are important in viral immunity, antibodies will however, be an essential for the vaccination response to prevent primary infection and reinfection. Most infected subjects will develop an immunoglobulin anti-viral response within 1 month (Zhao et al. 2020; Okba et al., 2020; de Assis et al., 2020). This appears to prevent re-infection as shown in non-human primates (Bao et al., 2020. Fig. 2). However, immunity may not be completely protective since people with COVID-19 can rarely present with SARS-CoV-2 re-activation (Ye et al., 2020; Chen et al., 2020c). However, as the virus may persist in many sites and may not be eliminated at the same rates (Chen et al., 2020d). This may in part explain why viral RNA is detected in faeces when nasopharyngeal swabs become negative (Chen et al., 2020d). There are clearly viral variants (Foster et al., 2020; Yao et al., 2020a) and may be important as vaccines will need to target disease-causing pathogenic variants. This data suggests that immunosuppression of macrophage function and probably CD8 activity may limit anti-viral protection, while blunting or inhibition of antibody formation may limit immunity to reinfection.

3.2. Destructive phase

Although most people appear to tolerate COVID-19 a significant number of people experience respiratory distress (Chen et al., 2020e; Zhu et al., 2020b). It has also been suggested that abnormal coagulation, pulmonary embolism, and endothelial dysfunction are other pathologies of severe COVID-19, which could in part be related to virus and inflammation-induced oxidative stress (Fox et al., 2020; Poor et al., 2020). However, severe disease is associated with peripheral blood neutrophilia and notably lymphopenia (Chen et al., 2020e, Lui et al., 2020b, Wang et al., 2020b), where viral load relates to the severity of lymphopenia (Lui et al., 2020c). The lymphopenia could relate to sequestration of cells into the infected tissues as part of the anti-viral response. Post-mortem histology demonstrates significant mononuclear infiltration into the lung and often, but not always, a paucity of natural killer cells and neutrophils, unless associated with secondary infection (Xu et al., 2020b; Fox et al., 2020; Yao et al., 2020b; Magro et al., 2020; Aurelio Sonzogni et al. 2020). There is a paucity of B cells and perhaps of relevance is that the lymphocytes are predominantly CD4 T cells (Xu et al., 2020b; Yao et al. 2020b). Low

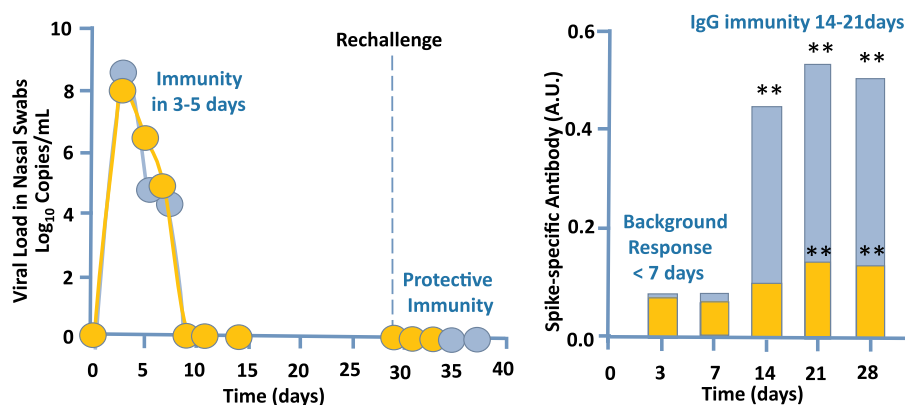


Fig. 2. Removal of the SARS-CoV-2 virus occurs before a significant anti-viral antibody response is generated. Rhesus macaques were infected with coronavirus and the viral titre was assessed using nasal swabs. Animals were re-infected one month later. The results show the responses of two individual (blue and orange) monkeys, as seen in two additional monkeys, relating to viral titre and anti-viral antibody response. A.U. arbitrary units. Adapted from Bao et al. 2020. DoI.org/10.1101/2020.03.13.990226

peripheral blood CD8 T cell numbers are a poor prognostic feature (Du et al., 2020) consistent with a common feature of the COVID-19 lung pathology, where there is a paucity of CD8 T cells (Xu et al. 2020b; Yao et al., 2020b; Zhang et al., 2020b). This may reflect senescence and exhaustion of the anti-viral CD8 response (Zheng et al., 2020; Cossarizza et al., 2020). Whether this contributes to severe disease and fatality remains to be established. However, this would be consistent with age being a major poor prognostic feature (Huang et al., 2020). It appears that T cells can be infected via CD147 (Wang et al., 2020c). In addition, infection and expression of envelope protein and Open Reading Frame protein sequestration that has been shown to have an apoptotic effect at least after SARS-CoV infection (Yang et al., 2005). This may play a role in the lymphopenia and immune suppression of the anti-viral response. There is marked atrophy of lymphoid tissues that may contribute to the lymphopenic state (Chen et al., 2020f; Zhang et al., 2020b, Yao et al., 2020b). Macrophages may also become infected and can take-up the virus due to expression of CD147, lectins and Toll-like receptors known to recognize SARS-CoV pathogen associated molecular pattern recognition elements such as single stranded viral RNA or uptake of viral antibody complexes (Wang et al., 2020a; Yang et al., 2004; Li et al 2013; Iwasaki and Yang, 2020). Macrophage activity may contribute to the lymphopenia (Chen et al., 2020f). Macrophage derived cytokines are produced, which lead to cytokines storms associated with worse prognosis (Chen et al., 2020c; Herold et al., 2020; Wen et al., 2020; Wilk et al., 2020). Therefore, agents such as IL-6 receptor and IL-1 blockers used in rheumatoid arthritis, and the case of IL-6R off-label in neuromyelitis optica, are being used to limit severe COVID-19 (Luo et al., 2020). Plasma cell-supporting cytokines such as TNFSF13 may be associated with recovery (Wen et al., 2020), however, the antibody response may contribute to macrophage hyper-activation. As such, severe disease is associated with the highest titres of antibodies (Liu et al., 2019; Zhao et al., 2020) and antibody-dependent enhancement of disease may occur (Iwasaki and Yang, 2020). There is complement activation, vascular damage and microthrombi that develop, indicative of damage consistent with oxidative stress, IgG3 anti-viral responses and IgG antibody-dependent cellular cytotoxicity by macrophages and in some instance neutrophils (Magro et al., 2020; Zhang et al., 2020b). Interestingly, it has been shown that spike-specific antibody may promote IL-8 and CCL2 production that skews macrophage accumulation towards a destructive phenotype (Liu et al 2019). As such in other lower respiratory tract infections antibodies can sometimes have destructive potential (Kim et al., 1969), therefore immunomodulation during periods of lung damage may offer some benefit.

4. Mechanisms driving multiple sclerosis may be distinct from COVID-19 protection and pathogenesis

Although it is widely considered that CD4, TH17 T cells are the central mediators of MS (Kunkl et al., 2020), all active DMT inhibit

memory B cell activity in a hierarchical fashion that reflects their therapeutic activity (Pardo and Jones 2017, Baker et al. 2017, Baker et al., 2018). This could be secondary to inhibition of T cell function (Sabatino et al., 2019a, 2019b). Targeting memory B cell subsets, and possibly CD4, Th17 T cells, is not likely to prevent SARS-CoV-2 elimination by CD8 T cells and the innate immune responses. This may only be relevant with continuous treatments that maintain peripheral B cells in a nadir state and prevent antibody secreting cell (ASC) formation (Sabatino et al., 2019a; Baker et al., 2020a). However, ASC can be generated by germinal centre cells independent of the CD27+, memory B cell pathway (Baker et al., 2018; Hammarlund et al., 2017; Khodadadi et al., 2019). Novel vaccine responses will be generated from the immature/naïve B cell compartments that regenerate most rapidly following B cell depleting therapies (Baker et al., 2017b, Baker et al., 2019, Baker et al., 2020a). Once formed, anti-viral responses will reside within the long-lived plasma cell pool with lymphoid tissue and bone marrow (Khodadadi et al., 2019; Baker et al., 2018). Plasma cells are relatively quiescent (Khodadadi et al., 2019) and thus avoid the action of agents targeting proliferating cells and they also express low levels of CD52, deoxycytidine kinase and CD20 targeted by high-efficacy, depleting DMT (Baker et al., 2020b; Sabatino et al., 2019a, Baker et al., 2019). Furthermore, they reside predominantly in the bone marrow, a site that may not be effectively targeted by depleting antibodies as cell elimination requires entry of antibodies, complement components and effector accessory cells required for depletion (Baker et al., 2018). Thus once formed, plasma cells may not be particularly well targeted by the current DMT, except haematopoietic stem cell therapy (HSCT) that purges the lymphoid tissues. It will be important to consider how best to deliver a SARS-CoV-2 vaccine in the future (Amanat and Krammer 2020; Chen et al., 2020g). Strategies could be developed for the highly-active agents that accommodate the long-depletion of memory B cells and the more rapid population of naïve cells to allow a vaccination response against SARS-CoV-2 whilst maintaining protection against MS.

5. Low efficacy MS immunomodulators are unlikely to limit anti-viral immunity

The components of the immune response that drive autoimmunity and control infection use overlapping cellular mechanisms. Therefore, removal of significant immune subsets may have the capacity to reduce anti-viral responses in a manner that reflects their immunosuppressive potential. Low treatment-efficacy agents such as glatiramer acetate, beta interferons and teriflunomide (Table 1) are not associated with significant immunosuppression, notable increased risk of viral infections, nor lack of responsiveness to vaccines (Pardo and Jones 2017; Comi et al., 2019a; Wijnands et al., 2018; Olberg et al., 2018; Hauser et al., 2019). Indeed, interferon beta and teriflunomide may have anti-viral activity that could be beneficial (Hensley et al., 2004; Bilger et al). As such beta interferon has been shown to inhibit SARS-

CoV replication and is currently being trialled in COVID-19 (Dahl et al., 2004; Hensley et al., 2004; NCT04350671; NCT04343768). However, these agents have a downside in that they are not that effective in controlling MS disease activity.

6. Moderate efficacy MS immunomodulators carry higher, but modest infection risks

Dimethyl fumarate is modestly immunosuppressive and targets lymphocytes rather than monocytes (Pardo and Jones 2017; Diebold et al., 2018). Immature/transitional B cells are less affected compared to memory B cell targeting (Mehta et al., 2019). Although plasmablasts and plasma cells can be affected by dimethyl fumarate therapy (Mehta et al., 2019), immunoglobulin levels are not unduly reduced (Diebold et al., 2018). Importantly, vaccine responses in people on dimethyl fumarate were no different to those treated with beta interferons (von Hehm et al., 2017). However, in some individuals persistent lymphopenia has been reported (Mehta et al., 2019; Diebold et al., 2018), notably about 20% of people will exhibit CD8 T cell levels below the lower limit of normal (Mehta et al., 2019). Although this is not generally associated with increased infection rates (Boffa et al., 2020), viral infections, including upper respiratory and lung infections occur with the monomethyl fumarate producing compounds (Pardo and Jones 2017; Diebold et al., 2018; Perini et al., 2018; Fernández et al., 2017; Naismith et al., 2019).

Functional lymphopenia occurs with sphingosine-1-phosphate receptor modulators such as fingolimod (Pardo and Jones 2017; Diebold et al., 2018). This appears to modestly elevate efficacy and infection risks (Pardo and Jones 2017; Kalincik et al., 2019). These agents are reported to sequester lymphocytes within lymphoid tissues and exhibit limited activity on the innate immune response (Kowarik et al., 2011; Pardo and Jones 2017; Thomas et al., 2017; Angerer et al., 2018). Fingolimod targets CD4 more than CD8 T cells and notably the naïve and central memory T cell subsets to retain them in lymphoid tissues where anti-viral responses would be generated (Kowarik et al., 2011; Angerer et al., 2018; Hjoorth et al., 2020). It also exhibits a more modest decrease in effector memory CD4 and CD8 T cells that will enter inflamed tissues (Angerer et al., 2018). Infections rates are modest (Diebold et al., 2018), but some bacterial and viral, infections such as herpes and varicella, are marginally more common after fingolimod treatment (Calibresi et al., 2014; Pardo and Jones 2017; Diebold et al. 2019). There may be subtle differences between fingolimod and the other sphingosine-1-phosphate receptor modulators in terms of infections and adverse effects, however it has a relatively long-half-life compared to other agents, which may be relevant if one wants to stop treatment (Subei et al., 2015; Swallow et al., 2020). A small scale trial of fingolimod has been reported for severe COVID-19 (NCT04280588). Sphingosine-1-phosphate is involved with maintaining the germinal centre and B cell niche (Cinamon et al., 2008) and there may be reduced serum immunoglobulin level following fingolimod treatment (Zoehner et al., 2019) as such vaccine responses are slightly reduced compared to the interferons (Olberg et al., 2018; Signoriello et al., 2020) as occurs with natalizumab (Olberg et al., 2018).

7. Natalizumab as the preferred high-efficacy agent

Currently natalizumab is perceived to be the high-efficacy treatment of choice (Coles et al., 2020. Table 1). Natalizumab, unlike depleting highly-active DMT, is potentially more rapidly reversible using plasma exchange and is not likely to inhibit migration of immune cells into lymphoid tissues and prevent novel immune responses, and as such has no or limited influence on vaccine antibody responses

(Vågberg et al., 2012; Kaufman et al., 2014; Olber et al., 2018). The value of the use of natalizumab may also be enhanced because it is perceived to inhibit T cell migration into the central nervous system (CNS) (Yednock et al.1992; Schwab et al., 2015). However, both B cells and importantly monocytes express alpha 4 integrin (CD49d) and thus the antibody directed to CD49d inhibits monocyte binding to vascular cell adhesion molecule one (VCAM-1) (Yednock et al., 1992; Hyduk et al., 2009). Importantly, although natalizumab is used to block migration into the inflamed CNS and gut (Schwab et al., 2015), VCAM-1 is expressed in virally-inflamed lungs (Brodie et al., 1999). Therefore, CD49d is likely involved in mononuclear cell diapedesis into the inflamed lung during SARS-CoV-2 infection (Brodie et al., 1999; Yen et al., 2006). This potential activity is perhaps consistent with increased lung infections in MS following treatment with natalizumab (Polman et al., 2006). Furthermore, that SARS-CoV-2 is neutrotrophic, (Baig et al., 2020; Moriguchi et al., 2020, Helms et al. 2020) suggests that a potential risk of natalizumab treatment is that it blocks viral immunosurveillance of the CNS (Hoepner et al., 2014), however this issue is perhaps limited by the extended interval dosing suggested to limit MS activation and reduce the risk of progressive multifocal leukoencephalopathy (Ryerson et al., 2019; Clerico et al., 2020). Thus, whilst natalizumab use could potentially be a risk factor for severe COVID-19, it is likely to limit monocyte and T cell damage to the lung and avoiding severe complications.

8. High-efficacy depleting agents are not the same and have distinct COVID-19 risks

Based on initial suggestions, immune reconstitution therapies (IRT) were not recommended to be started and ongoing treatment, i.e. additional courses, should be delayed (Table 1. Coles et al., 2020). Autologous HSCT is seen as a high-risk strategy to initiate during the mass-infection stage of COVID-19 pandemic (Table 1) and will probably remain so until herd immunity (Kwok et al., 2020) develops. Myeloablative HSCT removes both the adaptive and notably the innate immune systems and it is already well recognised that loss of the neutrophils, monocytes and other elements of the innate immune system increases the risk of mortality from infection, and until the innate and adaptive immune response reconstitutes people will be at risk for some time (Storek et al., 2008; Ge et al., 2019; Rush et al., 2019). However, once reconstituted the capacity to generate new immune responses occurs as seen following vaccination against childhood infections, to replace the lost immunity due to the HSCT procedure (Brinkmann et al., 2007, Rush et al., 2019). Therefore, there are clear risks from viral infections until the immune system reconstitutes. It is suggested that current licenced IRT, which both deplete T and B cells (Baker et al., 2017b; Baker et al., 2017c) carry similar risk (Coles et al., 2020). However, this does not accommodate the biologies and as such, oral cladribine is dissimilar to alemtuzumab, in terms of its risk for SARS-CoV-2 infection and appears more similar to ocrelizumab in its immunodepletion profile (Table 2).

8.1. Alemtuzumab

This is a CD52-depleting antibody that induces long-lasting and marked (80-90%) depletion of CD4, CD8 T cells and memory B cells (Table 2. Baker et al. 2017; Akgün et al., 2020). Alemtuzumab induces long-term disease remission if treated sufficiently early after symptom onset (Cohen et al., 2012. Havrdova et al., 2017, He et al., 2020b). Two short cycles of treatment give long-term disease remission. Alemtuzumab treatment cycles are generally given at least 12 months apart, but this interval may be extended up to 18 months, which supports the important activity of memory B cells as they, and CD4 T cells, can be

Table 2

High efficacy agents are not the same and oral cladribine is more similar to ocrelizumab than alemtuzumab.

	ALEMTUZUMAB ¹	CLADRIBINE ⁷	OCRELIZUMAB ¹²
Practicality	INFUSION ¹	ORAL TABLETS ⁸	INFUSION
	STEROIDS TO STOP CRS ¹	NO STEROIDS	STEROIDS TO STOP CRS
	HOSPITAL VISITS REQUIRED	HOSPITAL VISITS NOT REQUIRED	HOSPITAL VISIT REQUIRED
	MONITORING FREQUENT ¹	MONITORING MINIMAL ⁸	MONITORING MINIMAL ¹²
	DRUG PERSISTANCE ~1 MONTH ¹	DRUG PERSISTANCE 1 DAY ⁸	DRUG PERSISTANCE 5-6 MONTHS ¹²
Differential Infection Risk	EARLY DELETION MONOCYTE ²⁻⁴	MONOCYTES IN NORMAL RANGE ⁹	MONOCYTES IN NORMAL RANGE ¹³
	NEUTROPHILS IN NORMAL RANGE ^{3,5}	NEUTROPHILS IN NORMAL RANGE (~10%) ⁹	NEUTROPHILS IN NORMAL RANGE ¹²
	CD4 DEPLETED (70-90%) ⁵	CD4 IN NORMAL RANGE (40-50%) ⁹	CD4 IN NORMAL RANGE (~2%) ¹³
	CD8 DEPLETED (70-90%) ⁵	CD8 IN NORMAL RANGE (30-40%) ⁹	CD8 IN NORMAL RANGE (6-8%) ¹³
	NK CELLS IN NORMAL RANGE (40%)	NK CELLS IN NORMAL RANGE (50%) ⁹	NK CELLS IN NORMAL RANGE (<10%) ¹³
Efficacy	IMMATURE B CELL DEPLETED (3-6 MONTHS) ⁵	IMMATURE B CELL DEPLETED (6-9 MONTHS) ¹⁰	IMMATURE B CELLS DEPLETED PERMANENTLY ^{13,14}
	MEMORY B CELLS DEPLETED (> 1 YEAR) ⁵	MEMORY B CELLS DEPLETED (> 1YEAR) ¹⁰	MEMORY B CELLS DELETED PERMANENTLY ^{13,14}
	PLASMA CELLS. LOW CD52 ⁵	PLASMA CELLS LOW DEOXYCYTODINE KINASE ¹¹	PLASMA CELLS, NO CD20 ¹²
	EARLY INFECTION RISK ¹	LIMITED INCREASED RISK ⁸	LIMITED INCREASED RISK ¹²
	(EARLY ANTI-VIRAL REQUIRED)	(MAINLY BACTERIAL, INCREASE HERPES)	(MAINLY BACTERIAL, INCREASE HERPES)
Infection risk	VACCINATION COMPETENT AFTER 6 MONTHS ⁷	?	VACCINATION COMPETENT and BUT BLUNTED ¹⁵

Different characteristics of alemtuzumab, cladribine and ocrelizumab, relevant to efficacy and side-effect potential and their capacity to control MS and exhibit an effective anti-viral immune response. CRS cytokine release syndrome. NK natural killer cell. 1. Lemtrada® 2019. 2. Thomas et al. 2016. 3. Baker et al. 2017d. 4. Gross et al. 2016. 5. Baker et al. 2017b. 6. Baker et al. 2020b. 7. McCarthy et al. 2013. 8. Mavencad® 2018. 9. Baker et al. 2017c. 10. Ceroni et al. 2018. 11. Baker et al. 2019. 12. Ocrevus® 2018; 13. Baker et al. 2020a. 14. Fernandez-Verlascio et al. 2019. 15. Stokmaier et al. 2018.

depleted for at least this time (Tuohy et al., 2015; Havrdova et al., 2017; Akgün et al., 2020). However, alemtuzumab induces transient monocyte depletion and can induce very long-term CD4 and CD8 T cell depletion (Kousin-Ezewu et al., 2014; Thomas et al., 2016; Baker et al., 2017b; Akgün et al., 2020). This influences responses to viral and other infections (Cohen et al., 2012; Wray et al., 2019) and could thus impact on SARS-CoV-2 outcome. Severe lymphopenia increases the risk of infections and pneumonia (Warny et al., 2018). Neutropenia after alemtuzumab can be marked and significant, but is unusual (Baker et al., 2017d). Infection risk is notable following infusion and decreases with time as cellular repopulation occurs (Buonomo et al 2018; Wray et al., 2019). Alemtuzumab has a relatively short half-life and is cleared from the circulation

within about a month (Li et al., 2018). Therefore, surviving cells can repopulate in response to infection and given the relatively low dose and delivery over a single week, allows cells escaping elimination to recover. Transitional/immature B cells rapidly repopulate in the relative absence of T cell regulation, possibly related to limited purging of the bone marrow, and can generate anti-drug responses within a month of treatment in 60-83% of people in the virtual absence of peripheral B and T cell (Baker et al., 2017b; Baker et al., 2020b). Therefore, perhaps it may be possible to generate anti-viral responses. As such childhood vaccine responses persist and novel vaccine responses are not notably inhibited with alemtuzumab within 6 months of treatment (McCarthy et al 2013). Thus with time people with MS are likely to be able to generate a SARS-CoV-2 response and respond to vaccination. Although the treatment protocol means that few infusion visits are required (Cohen et al., 2012; Havrdora et al., 2017), the adverse events, notably the secondary autoimmunities that develop in many people with MS (Tuohy et al., 2015; Havrdora et al., 2017) means that intensive monitoring is required, compared to ocrelizumab that required essentially no inter-infusion monitoring (Pardo and Jones 2017).

8.2. Ocrelizumab

This is a CD20-depleting antibody used to treat relapsing and active primary progressive MS (Hauser et al., 2017; Montalban et al., 2017). This depletes peripheral B cells including memory B cells (Fernandez Velasco et al., 2019). Based on a common mechanism of action (Baker et al., 2017a), there is an unanswered question of whether ocrelizumab will behave like alemtuzumab and cladribine and provide long-term disease inhibition from a short-term treatment cycle (Table 2). Even if it acts as an IRT, based on memory B cell depletion and slow repopulation characteristics (Palanichamy et al., 2014; Baker et al., 2018), it may provide some comfort to suggest that delays of 6-12 months may be feasible without MS disease activity reoccurring. The latter is based on information from off-label and phase I/II studies with rituximab in MS (Bar-Or et al., 2008; Juto et al., 2020) and phase II extension trial data of ocrelizumab (Kappos et al., 2012; Baker et al., 2020a). As such retreatment to maintain remission based on repopulation of CD27+ memory B cell population, after 3-4 cycles it seems that doses, at least with rituximab, can be extended to less than once a year (Novi et al., 2019). Given that ocrelizumab exhibits depletion for a longer duration than rituximab suggests similar or better results can be obtained with rituximab (Baker et al., 2020a). Although ocrelizumab can deplete CD8 T cells, this is only a relatively mild steady state depletion of only 6-8% depletion of CD8 cells and 1-2% of CD4 T cells and has a minor impact on monocytes (Gingele et al., 2018; Baker et al., 2020a). Although infections are generally mild following ocrelizumab treatment (Hauser et al., 2017), some viral infections do occur and can be serious and very rarely life threatening (Hauser et al., 2017; Nicolini et al., 2019). Importantly, this may become a problem with persistent B cell depletion as that which occurs with ocrelizumab (Hauser et al., 2017). In time this can lead to IgM, IgA and IgG hypogammaglobulinemia that will increase infection risk (Tallantrye et al., 2018; Vollmer et al., 2020). However, a delay in repeated cycles may allow immature cells that provide immunity to new infections to partially regenerate, although this process is slow with ocrelizumab (Kappos et al., 2012; Baker et al., 2020a), and improve the vaccination response. Consistent with marked B cell depletion, it is apparent that vaccination responses are blunted when initiated 3 months after infusion however, they are not absent (Stokmaier et al., 2018). As plasma cells do not express CD20, once formed they will not be directly targeted by ocrelizumab (Sabitino et al., 2019a). Ofatumumab is a novel subcutaneous CD20-depleting antibody awaiting licencing following a successful phase III programme (Hauser et al 2019). Ofatumumab dosing shows relatively rapid repopulation of immature B cells compared

Table 3
Our opinion of altered risks of different MS DMT for COVID-19.

At risk category	Rank	Class	Trade Name	Mode of action	Efficacy Class	Class	Safe to start treatment	Advice regarding treatment	In the event of COVID-19 infection?	Immunosuppression?	Response to future SARS-CoV-2 vaccine	Attributes and caveats
Very low	1	Interferon-beta	Betaferon, Avonex, Rebif, Plegridy	Immunomodulatory (not immunosuppressive), pleiotropic immune effects	Moderate	Maintenance immunomodulatory	Yes	Continue	Continue	No	Likely to be intact	Has antiviral properties that may be beneficial in the case of COVID-19
Very low	2	Glatiramer acetate	Copaxone	Immunomodulatory (not immunosuppressive), pleiotropic immune effects	Moderate	Maintenance immunomodulatory	Yes	Continue	Continue	No	Likely to be intact	-
Very low	3	Cladribine / Alemtuzumab / Mitoxantrone / HSCT	see below	Post-immune reconstitution with normal innate and adaptive immunity (lymphocyte count > 1000/mm ³)	High / Very high	IRT	N/A	N/A	N/A	No	Likely to be intact	Some patients who may have mitoxantrone or chemotherapy-induced (HSCT) cardiomyopathy may be at increased risk of severe COVID-19
Very low	4	Teriflunomide	Aubagio	Dihydro-orotate dehydrogenase inhibitor (reduced de novo pyrimidine synthesis), anti-proliferative	Moderate (1st-line) / Moderate to high (2nd-3rd-line)	Maintenance immunomodulatory	Yes	Continue	Continue	Possible (no well-defined immunosuppressive signature)	Likely to be intact	Has antiviral properties that may be beneficial in the case of COVID-19
Low	5	Dimethyl fumarate	Tecfidera	Pleiotropic, NRF2 activation, downregulation of NFKβ	Moderate (2nd-3rd-line) / High (1st-line)	Maintenance immunosuppressive	Probably	Continue / Switch if lymphopenic	Continue	Likely to be intact	Likely to be intact	The risk can only be considered low in patients who do not develop a persistent lymphopenia. Patients with a total lymphocyte count of less than 800/mm ³ should be considered to be at a higher risk of developing complications from COVID19 infection.
Low	6	Natalizumab (EID / extended interval dosing)	Tysabri	Anti-VLA4, selective adhesion molecule inhibitor	Very high	Maintenance immunosuppressive	Yes	Continue	Continue or miss infusion depending on timing	Yes, continuous	Likely to be intact	As COVID-19/SARS-CoV-2 is neurotropic natalizumab will potentially prevent viral clearance from the CNS; this risk is likely to be very low on EID or extended interval dosing. We still have concerns about creating an environment in mucosal surfaces and the gut that may promote prolonged viral shedding; again this risk will be lower with EID.
Low	7	Anti-CD20	Ocrelizumab (Ocrevus), Ofatumumab, Rituximab, Ublituximab	Anti-CD20, B-cell depleter	Very high	Maintenance immunosuppressive	Probably	Risk assessment - continue or suspend dosing	Temporary suspension of dosing depending on timing	Yes, continuous	Blunted, particularly to glycoprotein components of a vaccine	Does drop the both CD4+ and CD8+ T-cell populations by up to 20% and this may interact with other factors to affect antiviral responses. Theoretical risk that ocrelizumab and other anti-CD20 therapies may result in prolonged viral shedding.

(continued on next page)

Table 3 (continued)

Intermediate	Cladribine	Mavenclad	Deoxyadenosine (purine) analogue, adenosine deaminase inhibitor, selective T and B cell depletion	High / Very high (highly-active RMS)	IRT (semi-selective)	Probably	Risk assessment - continue or suspend dosing	Temporary suspension of dosing depending on timing	Yes, intermittent	Possibly blunted.	Only reduces the T-cell compartment by ~50% and has less of an impact on the CD8+ population. Provided total lymphocyte counts are above 500/mm ³ allowing appropriate antiviral responses should be maintained. Theoretical risk that in the immune depletion phase cladribine may result in prolonged viral shedding.
Intermediate	SIP modulators	Fingolimod (Gilenya), Siponimod (Mazent), Ozanimod, Ponesimod	Selective SIP modulator, prevents egress of lymphocytes from lymph nodes	High	Maintenance immunosuppressive	Probably	Continue	Continue or temporary suspension of dosing	Yes, continuous	Blunted	Theoretical risk that SIP modulators may result in prolonged viral shedding. Paradoxically SIP modulators may reduce the severity of COVID-19; fingolimod is currently being trialed.
Intermediate	Natalizumab (SIP / standard interval dosing)	Tysabri	Anti-VLA4, selective adhesion molecule inhibitor	Very high	Maintenance immunosuppressive	Yes	Continue, but consider EID	Continue or miss infusion depending on timing	Yes, continuous	Likely to be intact	As COVID-19/SARS-CoV-2 is neurotropic natalizumab will prevent viral clearance from the CNS. Intermediate risk; higher theoretical risk on SID. I have that natalizumab will create an environment in mucosal surfaces and the gut that may promote prolonged viral shedding.
High*	Mitoxantrone	Novatrone	Immune depleter (topoisomerase inhibitor)	Very high	IRT (non-selective)	No	Suspend dosing	Suspend dosing	Yes, intermittent	Blunted	Theoretical risk that in the immune depletion phase mitoxantrone may result in prolonged viral shedding.
High*	Alemtuzumab	Lemtrada	Anti-CD52, non-selective immune depleter	Very high	IRT (non-selective)	No	Suspend dosing	Suspend dosing	Yes, intermittent	Blunted	Theoretical risk that in the immune depletion phase alemtuzumab may result in prolonged viral shedding.
High*	HSCT	-	Immune depletion and haemopoietic stem cell reconstitution	Very high	IRT (non-selective)	No	Suspend dosing	Suspend dosing	Yes, intermittent	Blunted	Theoretical risk that in the immune depletion phase HSCT may result in prolonged viral shedding.

* risk refers to acquiring an infection during the immunodepletion phase (E.g. rank 11,12,13). Post immune reconstitution the risk is low (rank 3). This opinion was formed 18 April 2020.

with slower repopulation with ocrelizumab (Savelieva et al., 2017; Baker et al., 2020a) and thus it remains to be established if the advantage of home injection and reversibility changes the use of anti-CD20 therapies compared to infusions with rituximab and ocrelizumab (Hauser et al. 2017, Hauser et al., 2019). Likewise, the real-life extended dosing experiment with rituximab and ocrelizumab is ongoing (Table 1). If data captured by registries shows maintained efficacy it is likely that the dosing schedule of ocrelizumab will eventually change on grounds of conveniences, safety and cost-effectiveness, although this will need formal testing (Novi et al. 2019; Baker et al., 2020a).

8.3. Cladribine

This is an oral small molecule that behaves as an IRT that gives long term-term benefit from short treatment cycles (Giovannoni et al., 2010; Giovannoni et al., 2018). This is a B and T cell depletion agent that is eliminated within one day of treatment (Table 2) (Baker et al., 2017c; Baker et al., 2019; Hermann et al., 2019). Treatment induces depletion via apoptosis rather than cell lysis and thus avoids the need for steroids to manage infusion reactions associated with alemtuzumab and ocrelizumab (Cohen et al., 2012; Hauser et al., 2017). Cladribine can induce comparable long-term memory B cell depletion similar to that observed with alemtuzumab, but without the innate cell and the severe lymphopenia associated with alemtuzumab (Ceronie et al., 2018; Ruggieri et al., 2019). Indeed, the T cell depletion is more modest and CD4 cells are depleted by about 40-50% and CD8 T cells are depleted by 30-40% compared to baseline. In comparison, alemtuzumab results in B and T cell depletion of 80-90% (Baker et al. 2017c). As such the T cells generally remain within the lower limit of the normal range as do natural killer cells that show modest depletion (Baker et al., 2017c; Comi et al., 2019b). The CD19+ B cells recover perhaps slower than post-alemtuzumab, as cladribine probably penetrates and acts more in lymphoid tissues, and the dosing schedule of doses being given a month apart targets any rapidly emerging cells (Baker et al., 2017c, Baker et al., 2019). However, B cells probably emerge faster after cladribine compared to ocrelizumab as depleting titres of ocrelizumab remain high for months after infusion (Genovese et al., 2008; Baker et al., 2017c, Baker et al., 2020a). Unfortunately, there is no information available concerning the influence of vaccination responses of oral cladribine. Although there is an increased risk of viral infections (Giovannoni et al., 2010), these are notably less severe than with alemtuzumab (Pardo and Jones 2017) associated with the milder immunosuppression induced by cladribine. Thus, oral cladribine, behaves like a chemical CD19/CD20 depleter with some additional T cell activity and is perhaps functionally closer to CD20-depleting antibodies than CD52 depleting antibody. It has the advantage that treatment is not continuous. During the time of self-isolation and shielding to prevent COVID-19 agents such as cladribine may have some merit as it is a high efficacy IRT that can be administered at home, with minimal post-dosing monitoring requirements (Table 2).

9. Preliminary experience and personal view of treatment

As analysis of the mechanisms of action of the different DMT coupled with emerging knowledge of the anti-viral and pathogenic mechanisms in COVID-19, suggest that initial fears relating to immunosuppression in MS, have yet to be realised, supporting that found in the SARS epidemics (D'Antiga 2020; Giovannoni 2020). The pragmatic approach of examining an individual patient's circumstances, their prognostic profile and level of MS disease activity may help guide treatment approaches (Giovannoni et al., 2020; Giovannoni 2020). Although these are early days in the initial infection wave of COVID-19,

already a number of people with MS have been infected with SAR-CoV-2 with the majority surviving based on early social media and registry data. Although a few people with MS have died they have tended to be older, with more advanced disease and multiple comorbidities. There are now over 360 people with MS and COVID-19 within Italian COVID MS registry with only 5 reported deaths, with only 2 people being treated with DMT and all having comorbidities associated with poor COVID-19 prognosis in the general population. Thus, there does not yet appear evidence that people with MS are at particular risk of severe COVID-19. As such we suggest that risks should be reviewed (Table 3) and advice regarding the risks associated with individual MS-DMT adjusted. Delays in treatment cycles may provide information on the biology of relapsing MS and, if successful may change prescribing habits in the future as there are risk/cost/benefit advantages of reduced dosing frequency. Thus, it will be interesting to determine whether one returns to the current *status quo* after the COVID-19 pandemic wanes or whether extended interval-dosing remains. Likewise, it will be intriguing to determine if real-life data shows that ocrelizumab exhibits IRT-like characteristics whereby long-term benefit can be seen with only short-term treatment cycle as seen with alemtuzumab and oral cladribine. The positive aspect of this unfortunate human experiment created by the SARS-CoV-2 epidemic, is that it will teach us more about the biology of MS and help inform how best to treat this disease and as safely as possible

Disclosures

No company was involved in the decision to write or was involved in the content of this paper. Therefore, disclosures are not considered relevant, however within the past 5 years: DB received consultancy/speaker fees from: Canbex therapeutics, Immunebio, Lundbeck, Merck, Novartis, Sanofi Genzyme. SA has received consultancy from Novartis. SA is section editor of multiple sclerosis and related disorders and associative editor at Clinical and Experimental Immunology. ASK has nothing relevant to declare. KS has received consultancy, speaker fees from: Biogen, Lipomed, Merck, Novartis, Roche, Sanofi-Genzyme and Teva. GG has received received consultancy, speaker fees or research support from: Abbvie, Actelion, Atara, Biogen, Canbex therapeutics, Celgene, MedDay, Merck, Novartis, Roche, Sanofi-Genzyme, Takeda, Teva. GG has received consultancy, speaker fees or research support from: Abbvie, Actelion, Atara, Biogen, Canbex therapeutics, Celgene, MedDay, Merck, Novartis, Roche, Sanofi-Genzyme, Takeda, Teva. Editor of multiple sclerosis and related disorders.

References

- Akgün, K, Blankenburg, J, Marggraf, M, et al., 2020. Event-Driven immunoprofiling predicts return of disease activity in alemtuzumab-treated multiple sclerosis. *Front Immunol.* 11, 56.
- Amanat, F, Kramer, F, 2020. SARS-CoV-2 Vaccines: status report. *Immunity pii: S1074-7613(20)30120-*.
- Angerer, IC, Hecker, M, Koczan, D, et al., 2018. Transcriptome profiling of peripheral blood immune cell populations in multiple sclerosis patients before and during treatment with a sphingosine-1-phosphate receptor modulator. *CNS Neurosci. Ther.* 24, 193–201.
- Baig, AM, Khaleeq, A, Ali, U, Syeda, H, 2020. Evidence of the COVID-19 virus targeting the CNS: tissue distribution, host-virus interaction, and proposed neurotropic mechanisms. *ACS Chem. Neurosci* 11, 995–998.
- Baker, D, Marta, M, Pryce, G, et al., 2017a. Memory B Cells are major targets for effective immunotherapy in relapsing multiple sclerosis. *EBioMedicine* 16, 41–50.
- Baker, D, Herrod, SS, Alvarez-Gonzalez, C, et al., 2017b. Interpreting lymphocyte re-constitution data from the pivotal phase 3 trials of Alemtuzumab. *JAMA Neurol* 74, 961–969.
- Baker, D, Herrod, SS, Alvarez-Gonzalez, C, et al., 2017c. Both cladribine and alemtuzumab may affect MS via B-cell depletion. *Neurol. Neuroimmunol. Neuroinflamm.* 4 (4), e360.

- Baker, D, Giovannoni, G, Schmierer, K, 2017d. Marked neutropenia: Significant but rare in people with multiple sclerosis after alemtuzumab treatment. *Mult. Scler. Relat. Disord* 18, 181–183.
- Baker, D, Pryce, G, Amor, S, et al., 2018. Learning from other autoimmunities to understand targeting of B cells to control multiple sclerosis. *Brain* 141, 2834–2847.
- Baker, D, Pryce, G, Herrod, SS, Schmierer, K, 2019. Potential mechanisms of action related to the efficacy and safety of cladribine. *Mult. Scler. Relat. Disord.* 30, 176–186.
- Baker, D, Pryce, G, James, LK, et al., 2020a. The ocrelizumab phase II extension trial suggests the potential to improve the risk:benefit balance in multiple sclerosis. *bioRxiv*. <https://doi.org/10.1101/2020.01.09.20016774>.
- Baker, D, Ali, L, Saxena, G, Pryce, G, et al., 2020b. The irony of humanization: alemtuzumab, the first, but one of the most immunogenic, humanized monoclonal antibodies. *Front Immunol.* 11, 124.
- Bar-Or, A, Calabresi, PA, Arnold, D, et al., 2008. Rituximab in relapsing-remitting multiple sclerosis: a 72-week, open-label, phase I trial. *Ann. Neurol.* 63, 395–400.
- Bilger, A, Ploshway, J, Ma, S, et al., 2017. Leflunomide/teriflunomide inhibit Epstein-Barr virus (EBV)-induced lymphoproliferative disease and lytic viral replication. *Oncotarget* 8, 44266–44280.
- Bao, L, Deng, W, Gao, H, et al., 2020. Reinfection could not occur in SARS-CoV-2 infected rhesus macaques. *BioRxiv*, 990226. <https://doi.org/10.1101/2020.03.13.990226>. .03.13.
- Boffa, G, Bruschi, N, Cellerino, M, et al., 2020. Fingolimod and dimethyl-fumarate-derived lymphopenia is not associated with short-term treatment response and risk of infections in a real-life ms population. *CNS Drugs* 34, 425–432.
- Brinkman, DM, Jol-van der Zijde, CM, ten Dam, MM, et al., 2007. Resetting the adaptive immune system after autologous stem cell transplantation: lessons from responses to vaccines. *J. Clin. Immunol.* 27, 647–658.
- Brodie, SJ, de la Rosa, C, Howe, JG, et al., 1999. Pediatric AIDS-associated lymphocytic interstitial pneumonia and pulmonary arterio-occlusive disease: role of VCAM-1/VLA-4 adhesion pathway and human herpesviruses. *Am. J. Pathol.* 154, 1453–1464.
- Brownlee, W, Bourdette, D, Broadley, S, et al., 2020. Treating multiple sclerosis and neuromyelitis optica spectrum disorder during the COVID-19 pandemic. *Neurology*. <https://doi.org/10.1212/WNL.0000000000009507>. pii: 10.1212/WNL.0000000000009507[Epub].
- Buonomo, AR, Zappulo, E, Viceconte, G, et al., 2018. Risk of opportunistic infections in patients treated with alemtuzumab for multiple sclerosis. *Expert Opin. Drug. Saf.* 17, 709–717.
- Cai, Y, Hao, Z, Gao, Y, et al., 2020. COVID-19 in the perioperative period of lung resection: a brief report from a single thoracic surgery department in Wuhan. *China J. Thorac. Oncol.* <https://doi.org/10.1016/j.jtho.2020.04.003>. pii: S1556-0864(20)30298-7[Epub].
- Calabresi, PA, Radue, EW, Goodin, D, et al., 2014. *Lancet Neurol.* 2014 Jun;13(6):545-56. Safety and efficacy of fingolimod in patients with relapsing-remitting multiple sclerosis (FREEDOMS II): a double-blind, randomised, placebo-controlled, phase 3 trial. *Lancet Neurol.* 13, 545–556.
- Ceronie, B, Jacobs, BM, Baker, D, et al., 2018. Cladribine treatment of multiple sclerosis is associated with depletion of memory B cells. *J. Neurol.* 265, 1199–1209.
- Channappanavar, R, Fett, C, Zhao, J, et al., 2014. Virus-specific memory CD8 T cells provide substantial protection from lethal severe acute respiratory syndrome coronavirus infection. *J. Virol.* 88, 11034–11044.
- Chen, D, Xu, W, Lei, Z, Huang, Z, et al., 2020c. Recurrence of positive SARS-CoV-2 RNA in COVID-19: A case report. *Int. J. Infect. Dis.* 93, 297–299. <https://doi.org/10.1016/j.ijid.2020.03.003>. Mar 5[Epub ahead of print].
- Chen, G, Wu, D, Guo, W, et al., 2020e. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J. Clin. Invest.* <https://doi.org/10.1172/JCI137244>. pii: 137244[Epub].
- Chen, Y, Feng, X, Diao, B, et al., 2020f. The novel severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) directly decimates human spleens and lymph nodes. *medRxiv*. <http://doi.org/10.1101/2020.03.27.20045427>.
- Chen, W, Lan, Y, Yuan, X, et al., 2020a. Detectable 2019-nCoV viral RNA in blood is a strong indicator for the further clinical severity. *Emerg. Microbes Infect.* 9, 469–473.
- Chen, WH, Strych, U, Hotez, PJ, Bottazzi, ME, 2020g. The SARS-CoV-2 Vaccine Pipeline: an Overview. *Curr. Trop. Med. Rep.* (Mar 3), 1–4.
- Chen, Y, Chen, L, Deng, Q, et al., 2020d. The Presence of SARS-CoV-2 RNA in Feces of COVID-19 Patients. *J. Med. Virol.* <https://doi.org/10.1002/jmv.25825>. [Epub].
- Chen, Y, Liu, Q, Guo, D, 2020b. Emerging coronaviruses: Genome structure, replication, and pathogenesis. *J. Med. Virol.* 92, 418–423.
- Chu, H, Chan, JF, Wang, Y, et al., 2020. Comparative replication and immune activation profiles of SARS-CoV-2 and SARS-CoV in human lungs: an ex vivo study with implications for the pathogenesis of COVID-19. *Clin. Infect. Dis.* <https://doi.org/10.1093/cid/ciaa410>. pii: ciaa410[Epub].
- Cinamon, G, Zachariah, MA, Lam, OM, et al., 2008. Follicular shuttling of marginal zone B cells facilitates antigen transport. *Nat. Immunol.* 9, 54–62.
- Clerico, M, De Mercanti, SF, Signori, A, et al., 2020. Extending the Interval of Natalizumab Dosing: Is Efficacy Preserved? *Neurotherapeutics* 17, 200–207.
- Cohen, JA, Coles, AJ, Arnold, DL, et al., 2012. Alemtuzumab versus interferon beta 1a as first-line treatment for patients with relapsing-remitting multiple sclerosis: a randomised controlled phase 3 trial. *Lancet* 380, 1819–1828.
- Coles, A, Lim M, Giovannoni G, et al. ABN guidance on the use of disease-modifying therapies in multiple sclerosis in response to the threat of a coronavirus epidemic. 2020 Apr (https://cdn.ymaws.com/www.theabn.org/resource/collection/65C334C7-30FA-45DB-93AA-74B3A20293/02.04.20_ABN_Guidance_on_DMTs_for_MS_and_COVID19_VERSION_4_April_2nd.pdf).
- Comi, G, Cook, S, Giovannoni, G, et al., 2019b. Effect of cladribine tablets on lymphocyte reduction and repopulation dynamics in patients with relapsing multiple sclerosis. *Mult. Scler. Relat. Disord.* 29, 168–174.
- Comi, G, Miller, AE, Benamor, M, et al., 2019a. Characterizing lymphocyte counts and infection rates with long-term teriflunomide treatment: Pooled analysis of clinical trials. *Mult. Scler.* <https://doi.org/10.1177/1352458519851981>. [Epub].
- Compston, A, Coles, A, 2008. Multiple sclerosis. *Lancet* 372, 1502–1517.
- Cossarizza, A, De Biasi, S, Guaraldi, G, et al., 2020. Modena COVID-19 working group (moco19)#. Sars-cov-2, the virus that causes COVID-19: cytometry and the new challenge for global health. *Cytometry A* 97, 340–343.
- D'Antiga, L, 2020. Coronaviruses and immunosuppressed patients. The facts during the third epidemic. *Liver Transpl.* <https://doi.org/10.1002/lt.25756>. [Epub].
- Dahl, H, Linde, A, Strannegård, O, 2004. In vitro inhibition of SARS virus replication by human interferons. *Scand. J. Infect. Dis.* 36, 829–831.
- Day, M, 2020. Covid-19: four fifths of cases are asymptomatic, China figures indicate. *BMJ* 369, m1375. <https://doi.org/10.1136/bmj.m1375>.
- de Assis RR, Jain A, Nakajima R, Jasinskas A, et al. Analysis of SARS-CoV-2 Antibodies in COVID-19 convalescent plasma using a coronavirus antigen microarray. *BioRxiv* doi: 10.1101/2020.04.15.043364.
- Diebold, M, Sievers, C, Bantug, G, et al., 2018. Dimethyl fumarate influences innate and adaptive immunity in multiple sclerosis. *J. Autoimmun.* 86, 39–50.
- Diebold, M, Fischer-Barnicol, B, Tsagkas, C, et al., 2019. Hepatitis E virus infections in patients with MS on oral disease-modifying treatment. *Neurol. Neuroimmunol. Neuroinflamm.* 6 (5) pii e594.
- Du, RH, Liang, LR, Yang, CQ, et al., 2020. Predictors of mortality for patients with COVID-19 Pneumonia caused by SARS-CoV-2: a prospective cohort study. *Eur. Respir. J.* <https://doi.org/10.1183/13993003.00524-2020>. pii: 2000524[Epub].
- Duan, K, Liu, B, Li, C, et al., 2020. Effectiveness of convalescent plasma therapy in severe COVID-19 patients. *Proc. Natl. Acad. Sci. U S A.* <https://doi.org/10.1073/pnas.2004168117>. pii: 202004168[Epub].
- Fernández, Ó, Giovannoni, G, Fox, RJ, et al., 2017. Efficacy and safety of delayed-release dimethyl fumarate for relapsing-remitting multiple sclerosis in prior interferon users: an integrated analysis of define and confirm. *Clin. Ther.* 39, 1671–1679.
- Fernandez Velasco, JI, Villarrubia Migallon, N, Monreal, E, et al., 2019. Effects of ocrelizumab treatment in peripheral blood leukocyte subsets of primary progressive multiple sclerosis patients. *P686 25 (S2)*, 341–342.
- Forster, P, Forster, L, Renfrew, C, Forster, M, 2020. Phylogenetic network analysis of SARS-CoV-2 genomes. *Proc. Natl. Acad. Sci. U S A* Apr 8. pii: 202004999.
- Fox, SE, Akmatbekov, A, Harbert, JL, et al., 2020. Pulmonary and cardiac pathology in COVID-19: the first autopsy series from New Orleans. *MedRxiv*. <https://doi.org/10.1101/2020.04.06.20050575>.
- Ge, F, Lin, H, Li, Z, Chang, T, 2019. Efficacy and safety of autologous hematopoietic stem-cell transplantation in multiple sclerosis: a systematic review and meta-analysis. *Neurol. Sci.* 40, 479–487.
- Genovese, MC, Kaine, JL, Lowenstein, MB, et al., 2008. Ocrelizumab, a humanized anti-CD20 monoclonal antibody, in the treatment of patients with rheumatoid arthritis: a phase I/II randomized, blinded, placebo-controlled, dose-ranging study. *Arthritis Rheum.* 58, 2652–2661.
- Giovannoni, G, Comi, G, Cook, S, et al., 2010. A placebo-controlled trial of oral cladribine for relapsing multiple sclerosis. *N. Engl. J. Med.* 362, 416–426.
- Giovannoni, G, Soelberg Sorensen, P, Cook, S, et al., 2018. Safety and efficacy of cladribine tablets in patients with relapsing-remitting multiple sclerosis: results from the randomized extension trial of the CLARITY study. *Mult. Scler.* 24, 1594–1604.
- Giovannoni, G, Hawkes, C, Lechner-Scott, J, et al., 2020. The COVID-19 pandemic and the use of MS disease-modifying therapies. *Mult. Scler. Rel. Disord.* <https://doi.org/10.1016/j.msard.2020.102073>.
- Giovannoni, G, 2020. Anti-CD20 immunosuppressive disease-modifying therapies and COVID-19. *Mult Scler Rel Disord.* <https://doi.org/10.1016/j.msard.2020.102135>. [Epub].
- Gingele, S, Jacobus, TL, Konen, FF, et al., 2018. Ocrelizumab Depletes CD20⁺ T Cells in Multiple Sclerosis Patients. *Cells* 8 (1). <https://doi.org/10.3390/cells8010012>. Dec 28pii: E12.
- Gramberg, T, Hofmann, H, Möller, P, et al., 2005. LSECtin interacts with filovirus glycoproteins and the spike protein of SARS coronavirus. *Virology* 340, 224–236.
- Gross, CC, Ahmetspahic, D, Ruck, T, et al., 2016. Alemtuzumab treatment alters circulating innate immune cells in multiple sclerosis. *Neurol. Neuroimmunol. Neuroinflamm.* 3 (6), e289.
- Hammarlund, E, Thomas, A, Amanna, LJ, et al., 2017. Plasma cell survival in the absence of B cell memory. *Nat. Commun.* 8, 1781.
- Hamming, I, Timens, W, Bulthuis, ML, et al., 2004. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J. Pathol.* 203, 631.
- Hauser, SL, Bar-Or, A, Comi, G, et al., 2017. Ocrelizumab versus interferon beta-1a in relapsing multiple sclerosis. *N Engl. J. Med.* 376, 221–234.
- Hauser, SL, Bar-Or, A, Cohen, J, et al., 2019. Efficacy and safety of ofatumumab versus

- teriflunomide in relapsing multiple sclerosis: results of the phase 3 ASCLEPIOS I and II trials. *Mult. Scler.* 25 (S2), 890–891.
- Havrdova, E, Arnold, DL, Cohen, JA, et al., 2017. Alemtuzumab CARE-MS I 5-year follow-up: durable efficacy in the absence of continuous MS therapy. *Neurology* 89, 1107–1116.
- He, X, Lau, EHY, Wu, P, et al., 2020a. Temporal dynamics in viral shedding and transmissibility of COVID-19. Rapid asymptomatic transmission of COVID-19 during the incubation period demonstrating strong infectivity in a cluster of youngsters aged 16–23 years outside Wuhan and characteristics of young patients with COVID-19: a prospective contact-tracing study. *Nat. Med.* <https://doi.org/10.1038/s41591-020-0869-5>.
- He, A, Merkel, B, Brown, JW, et al., 2020b. Timing of high-efficacy therapy for multiple sclerosis: a retrospective observational cohort study. *Lancet Neurol.* 19, 307–316.
- Helms, J, Kremer, S, Merdji, H, et al., 2020. Neurologic Features in Severe SARS-CoV-2 Infection. *N Engl. J. Med.* <https://doi.org/10.1056/NEJMc2008597>. [Epub].
- Hensley, LE, Fritz, LE, Jahrling, PB, et al., 2004. Interferon-beta 1a and SARS coronavirus replication. *Emerg. Infect. Dis.* 10, 317–319.
- Hermann, R, Karlsson, MO, Novakovic, AM, et al., 2019. The clinical pharmacology of cladribine tablets for the treatment of relapsing multiple sclerosis. *Clin. Pharmacokinet.* 58, 283–297.
- Herold, T, Jurinovic, V, Amreich, C, et al., 2020. Level of IL-6 predicts respiratory failure in hospitalized symptomatic COVID-19 patients. *medRxiv*, 20047381. <https://doi.org/10.1101/2020.04.01.20047381>. 04.01.
- Hjorth, M, Dandu, N, Mellergård, J, 2020. Treatment effects of fingolimod in multiple sclerosis: Selective changes in peripheral blood lymphocyte subsets. *PLoS One* 15 (2), e0228380.
- Hoffmann, M, Kleine-Weber, H, et al., 2020. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* pii: S0092-8674(20)30229-4. [Epub].
- Hoepner, R, Faissner, S, Salmen, A, et al., 2014. Efficacy and side effects of natalizumab therapy in patients with multiple sclerosis. *J. Cent. Nerv. Syst. Dis.* 6, 41–49.
- Huang, L, Zhang, X, Zhang, X, et al., 2020. Rapid asymptomatic transmission of COVID-19 during the incubation period demonstrating strong infectivity in a cluster of youngsters aged 16–23 years outside Wuhan and characteristics of young patients with COVID-19: a prospective contact-tracing study. *J. Infect.* <https://doi.org/10.1016/j.jinf.2020.03.006>. Apr 10pii: S0163-4453(20)30117-1[Epub].
- Hyduk, SJ, Cybulsky, ML., 2009. Role of alpha4beta1 integrins in chemokine-induced monocyte arrest under conditions of shear stress. *Microcirculation* 16, 17–30.
- Iwasaki, A, Yang, Y., 2020. The potential danger of suboptimal antibody responses in COVID-19. *Nat. Rev. Immunol.* <https://doi.org/10.1038/s41577-020-0321-6>.
- Juto, A, Fink, K, Al Nimer, F, Piehl, F, 2020. Interrupting rituximab treatment in relapsing-remitting multiple sclerosis; no evidence of rebound disease activity. *Mult. Scler. Relat. Disord.* 37, 101468.
- Kalincik, T, Kubala Havrdova, E, et al., 2019. Comparison of fingolimod, dimethyl fumarate and teriflunomide for multiple sclerosis. *J. Neurol. Neurosurg. Psychiatry.* 90 (4), 458–468.
- Kappos, L, Li, D, Calabresi, P, O'Connor, P, et al., 2012. Long-term safety and efficacy of ocrelizumab in patients with relapsing-remitting multiple sclerosis: week 144 results of a Phase II, randomised, multicentre trial. *Mult. Scler. J.* 18 (Suppl. 4), 140–141.
- Kaufman, M, Pardo, G, Rossman, H, et al., 2014. Natalizumab treatment shows no clinically meaningful effects on immunization responses in patients with relapsing-remitting multiple sclerosis. *J. Neurol. Sci.* 341, 22–27.
- Khodadadi, L, Cheng, Q, Radbruch, A, Hiepe, F, 2019. The Maintenance of Memory Plasma Cells. *Front Immunol.* 10, 721.
- Kim, HW, Canchola, JG, Brandt, CD, et al., 1969. Respiratory syncytial virus disease in infants despite prior administration of antigenic inactivated vaccine. *Am. J. Epidemiol.* 89 (4), 422–434.
- Kimball, A, Hatfield, KM, Arons, M, et al., 2020. Asymptomatic and Presymptomatic SARS-CoV-2 Infections in Residents of a Long-Term Care Skilled Nursing Facility - King County, Washington, March 2020. *MMWR Morb. Mortal Wkly. Rep.* 69, 377–381.
- Kousin-Ezewu, O, Azzopardi, L, Parker, RA, et al., 2014. Accelerated lymphocyte recovery after alemtuzumab does not predict multiple sclerosis activity. *Neurology* 82, 2158–2164.
- Kowarik, MC, Pellkofer, HL, Cepok, S, et al., 2011. Differential effects of fingolimod (FTY720) on immune cells in the CSF and blood of patients with MS. *Neurology* 76, 1214–1221.
- Kunkl, M, Frasca, S, Amormino, C, et al., 2020. T Helper Cells: The Modulators of Inflammation in Multiple Sclerosis. *Cells* 9 (2). <https://doi.org/10.3390/cells9020482>. pii: E482.
- Kwok, KO, Lai, F, Wei, WI, et al., 2020. Herd immunity - estimating the level required to halt the COVID-19 epidemics in affected countries. *J. Infect Mar* 21. pii: S0163-4453(20)30154-7.
- Lai, CC, Liu, YH, Wang, CY, et al., 2020. Asymptomatic carrier state, acute respiratory disease, and pneumonia due to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2): Facts and myths. *J. Microbiol. Immunol. Infect* pii: S1684-1182(20)30040-2. [Epub].
- Lauer, SA, Grantz, KH, Bi, Q, et al., 2020. The incubation period of coronavirus disease 2019 (COVID-19) from publicly reported confirmed cases: estimation and application. *Ann. Intern. Med.* <https://doi.org/10.7326/M20-0504>. Mar 10[Epub].
- Lemtrada® EU Summary of product characteristics. Nov 2019.
- Letko, M, Marzi, A, Munster, V, 2020. Functional assessment of cell entry and receptor usage for SARS-CoV-2 and other lineage B betacoronaviruses. *Nat. Microbiol* 5, 562–569.
- Li, Y, Chen, M, Cao, H, et al., 2013. Extraordinary GU-rich single-strand RNA identified from SARS coronavirus contributes an excessive innate immune response. *Microbes Infect* 15, 88–95.
- Li, Z, Richards, S, Surks, HK, et al., 2018. Clinical pharmacology of alemtuzumab, an anti-CD52 immunomodulator, in multiple sclerosis. *Clin. Exp. Immunol* 194, 295–314.
- Liu, L, Wei, Q, Lin, Q, et al., 2019. Anti-spike IgG causes severe acute lung injury by skewing macrophage responses during acute SARS-CoV infection. *JCI Insight* 4, e123158.
- Liu, Y, Liao, W, Wan, L, et al., 2020a. Correlation Between Relative Nasopharyngeal Virus RNA Load and Lymphocyte Count Disease Severity in Patients with COVID-19. *Viral Immunol.* <https://doi.org/10.1089/vim.2020.0062>. [Epub].
- Liu, Y, Liao, W, Wan, L, et al., 2020c. Correlation Between Relative Nasopharyngeal Virus RNA Load and Lymphocyte Count Disease Severity in Patients with COVID-19. *Viral Immunol.* <https://doi.org/10.1089/vim.2020.0062>. [Epub].
- Liu, Z, Long, W, Tu, M, et al., 2020b. Lymphocyte subset (CD4+, CD8+) counts reflect the severity of infection and predict the clinical outcomes in patients with COVID-19. *J. Infect* pii: S0163-4453(20)30182-1.
- Lippi, G, Mattiuzzi, C, Sanchis-Gomar, F, et al., 2020. Clinical and demographic characteristics of patients dying from COVID-19 in Italy versus China. *J. Med. Virol.* <https://doi.org/10.1002/jmv.25860>. Apr 10[Epub ahead of print].
- Lu, R, Zhao, X, Li, J, et al., 2020. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. *Lancet* 395 (10224), 565–574.
- Lukassen, S, Chua, RL, Trefzer, T, et al., 2020. SARS-CoV-2 receptor ACE2 and TMPRSS2 are primarily expressed in bronchial transient secretory cells. *EMBO J.* e105114. <https://doi.org/10.15252/embj.20105114>. [Epub].
- Luna, G, Alping, P, Burman, et al., 2020. Infection risks among patients with multiple sclerosis treated with fingolimod, natalizumab, rituximab, and injectable therapies. *JAMA Neurol.* 77, 184–191.
- Luo, P, Liu, Y, Qiu, L, et al., 2020. Tocilizumab treatment in COVID-19: A single center experience. *J. Med. Virol.* <https://doi.org/10.1002/jmv.25801>. [Epub].
- Magro, C, Mulvey, JJ, Berlin, D, et al., 2020. Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: a report of five cases. *Transl. Res.* <https://doi.org/10.1016/j.trsl.2020.04.007>. pii: S1931-5244(20)30070-0[Epub].
- Mavenclad® EU Summary of product characteristics. Jul 2018.
- McCarthy, CL, Tuohy, O, Compston, DA, et al., 2013. Immune competence after alemtuzumab treatment of multiple sclerosis. *Neurology* 81, 872–876.
- Mehta, D, Miller, C, Arnold, DL, et al., 2019. Effect of dimethyl fumarate on lymphocytes in RRMS: Implications for clinical practice. *Neurology* 92, e1724–e1738.
- Montalban, X, Hauser, SL, Kappos, L, et al., 2017. Ocrelizumab versus placebo in primary progressive multiple sclerosis. *Clinical Investigators.* *N Engl. J. Med.* 376, 209–220.
- Moriguchi, T, Harii, N, Goto, J, et al., 2020. A first Case of Meningitis/Encephalitis associated with SARS-Coronavirus-2. *Int. J. Infect. Dis* pii: S1201-9712(20)30195-8.
- Naismith, RT, Wolinsky, JS, Wundes, A, et al., 2019. Diroximel fumarate (DRF) in patients with relapsing-remitting multiple sclerosis: Interim safety and efficacy results from the phase 3 EVOLVE-MS-1 study. *Mult. Scler Nov* 4:1352458519881761 [Epub].
- Nicolini, LA, Canepa, P, Caligiuri, P, et al., 2019. Fulminant hepatitis associated with echovirus 25 during treatment with ocrelizumab for multiple sclerosis. *JAMA Neurol.* 76, 866–867.
- Novi, G, Fabbri, S, Bovis, F, et al., 2019. Tailoring B-cells depleting therapy in MS according to memory B-cells monitoring: a pilot study. *P971. Mult Scler.* 25 (S2), 509–510.
- O'Brien, TR, Thomas, DL, Jackson, SS, et al., 2020. Weak induction of interferon expression by sars-cov-2 supports clinical trials of interferon lambda to treat early COVID-19. *Clin. Infect. Dis.* <https://doi.org/10.1093/cid/ciaa453>. ciaa453[Epub].
- Okba NMA, Müller MA, Li W, et al. Severe acute respiratory syndrome coronavirus 2-specific antibody responses in coronavirus disease 2019 patients. *Emerg Infect Dis.* 2020;26(7). doi: 10.3201/eid2607.200841. [Epub].
- Olberg, HK, Eide, GE, Cox, RJ, et al., 2018. Antibody response to seasonal influenza vaccination in patients with multiple sclerosis receiving immunomodulatory. *Eur. J. Neurol.* 25, 527–534.
- Ocrevus® EU Summary Product Characteristics, Sep 2018.
- Ou, X, Liu, Y, Lei, X, et al., 2020. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. *Nat. Commun.* 11, 1620.
- Palanichamy, A, Jahn, S, Nickles, D, et al., 2014. Rituximab efficiently depletes increased CD20-expressing T cells in multiple sclerosis patients. *J. Immunol.* 193, 580–586.
- Pardo, G, Jones, DE., 2017. The sequence of disease-modifying therapies in relapsing multiple sclerosis: safety and immunologic considerations. *J. Neurol.* 264, 2351–2374.
- Pei, S, Yuan, X, Zhang, Z, et al., 2020. Convalescent plasma to treat covid-19: Chinese strategy and experiences. *MedRxiv.* <https://doi.org/10.1101/2020.04.07.20056440>.
- Perini, P, Rinaldi, F, Puthenparampil, M, et al., 2018. Herpes simplex virus encephalitis temporally associated with dimethyl fumarate-induced lymphopenia in a multiple

- sclerosis patient. *Mult. Scler. Relat. Disord.* 26, 68–70.
- Polman, CH, O'Connor, PW, Havrdova, E, et al., 2006. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl. J. Med.* 354, 899–910.
- Poor, HD, Venetuoilo, CE, Tolbert, T, et al., 2020. critical illness pathophysiology driven by diffuse pulmonary thrombi and pulmonary endothelial dysfunction responsive to thrombolysis. medRxiv, 20057125. <https://doi.org/10.1101/2020.04.17.20057125>. 04.17.
- Promptchara, E, Ketloy, C, Palaga, T, 2020. Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic. *Asian Pac. J. Allergy Immunol.* 38, 1–9.
- Pung, R, Chiew, CJ, Young, BE, et al., 2020. Investigation of three clusters of COVID-19 in Singapore: implications for surveillance and response measures. *Lancet* 395 (10229), 1039–1046.
- Richardson, S, Hirsch, JS, Narasimhan, M, et al., 2020. Presenting Characteristics, Comorbidities, and Outcomes Among 5700 Patients Hospitalized With COVID-19 in the New York City Area. *JAMA*. <https://doi.org/10.1001/jama.2020.6775>. [Epub].
- Rokni, M, Ghasemi, V, Tavakoli, Z, 2020. Immune responses and pathogenesis of SARS-CoV-2 during an outbreak in Iran: Comparison with SARS and MERS. *Rev. Med. Virol.* <https://doi.org/10.1002/rmv.2107>. Apr 8[Epub].
- Ruggieri, M, Gargano, C, Iaffaldano, A, et al., 2019. Changes in lymphocyte subpopulations in highly active multiple sclerosis patients during cladribine treatment. *Eur. J. Neurol.* 26 (Suppl 1), 491.
- Rush, CA, Atkins, HL, Freedman, MS, 2019. Autologous Hematopoietic Stem Cell Transplantation in the Treatment of Multiple Sclerosis. *Cold Spring Harb. Perspect. Med.* 9 (3) pii: a029082.
- Ryerson, LZ, Foley, J, Chang, I, et al., 2019. Risk of natalizumab-associated PML in patients with MS is reduced with extended interval dosing. *Neurology* 93, e1452–e1462.
- Sabatino Jr, JJ, Zamvil, SS, Hauser, SL, 2019a. B-Cell Therapies in Multiple Sclerosis. *Cold Spring Harb. Perspect. Med.* 9 pii: a032037.
- Sabatino Jr, JJ, Wilson, MR, Calabresi, PA, et al., 2019b. Anti-CD20 therapy depletes activated myelin-specific CD8+ T cells in multiple sclerosis. *Proc. Natl. Acad. Sci. U S A* 116, 25800–25807.
- Sarzi-Puttini, P, Giorgi, V, Sirotti, S, et al., 2020. COVID-19, cytokines and immunosuppression: what can we learn from severe acute respiratory syndrome? *Clin. Exp. Rheumatol.* 38, 337–342.
- Savelieva, M, Kahn, J, Bagger, M, et al., 2017. Comparison of the B-cell recovery time following discontinuation of anti-CD20 therapies. *EP1624. Mult. Scler.* 23 (S3), 852–853.
- Schwab, N, Schneider-Hohendorf, T, Wiendl, H, 2015. Therapeutic uses of anti- α 4-integrin (anti-VLA-4) antibodies in multiple sclerosis. *Int. Immunol.* 27 (1), 47–53.
- Shen, C, Wang, Z, Zhao, F, et al., 2020. Treatment of 5 Critically Ill Patients With COVID-19 With Convalescent Plasma. *JAMA*. <https://doi.org/10.1001/jama.2020.4783>. Mar 27[Epub].
- Shi, J, Wen, Z, Zhong, G, et al., 2020. Susceptibility of ferrets, cats, dogs, and other domesticated animals to SARS-coronavirus 2. *Science*. <https://doi.org/10.1126/science.abb7015>. pii: eabb7015[Epub].
- Signoriello, E, Bonavita, S, Sinisi, L, et al., 2020. Is antibody titer useful to verify the immunization after VZV Vaccine in MS patients treated with Fingolimod? A case series. *Mult. Scler. Relat. Disord.* 40, 101963.
- Soresina, A, Moratto, D, Chiarini, M, et al., 2020. Two X-linked agammaglobulinemia patients develop pneumonia as COVID-19 manifestation but recover. *Pediatr. Allergy Immunol.* <https://doi.org/10.1111/pai.13263>. [Epub].
- Stein, RA, 2020. COVID-19 and rationally layered social distancing. *Int. J. Clin. Pract.* e13501. <https://doi.org/10.1111/ijcp.13501>. Mar 14[Epub].
- Stokmaier, D, Winthrop, K, Chognot, C, et al., 2018. Effect of ocrelizumab on vaccine responses in patients with multiple sclerosis (S36.002). *Neurology* 90 (15 Suppl) S36.002.
- Storek, J, Geddes, M, Khan, F, et al., 2008. Reconstitution of the immune system after hematopoietic stem cell transplantation in humans. *Semin Immunopathol.* 30 (4), 425–437.
- Subei, AM, Cohen, JA., 2015. Sphingosine 1-phosphate receptor modulators in multiple sclerosis. *CNS Drugs.* 29 (7), 565–575.
- Swallow, E, Patterson-Lomba, O, Yin, L, et al., 2020. Comparative safety and efficacy of ozanimod versus fingolimod for relapsing multiple sclerosis. *J. Comp. Eff. Res.* 9 (4), 275–285.
- Tallantyre, EC, Whittam, DH, Jolles, S, et al., 2018. Secondary antibody deficiency: a complication of anti-CD20 therapy for neuroinflammation. *J. Neurol.* 265, 1115–1122.
- Tai, W, He, L, Zhang, X, et al., 2020. Characterization of the receptor-binding domain (RBD) of 2019 novel coronavirus: implication for development of RBD protein as a viral attachment inhibitor and vaccine. *Cell Mol. Immunol.* <https://doi.org/10.1038/s41423-020-0400-4>. [Epub].
- Tian, S, Hu, W, Niu, L, et al., 2020a. Pulmonary Pathology of Early-Phase 2019 Novel Coronavirus (COVID-19) Pneumonia in Two Patients With Lung Cancer. *J. Thorac. Oncol.* 28 pii: S1556-0864(20)30132-5.
- Tian, X, Li, C, Huang, A, et al., 2020b. Potent binding of 2019 novel coronavirus spike protein by a SARS coronavirus-specific human monoclonal antibody. *Emerg. Microbes Infect.* 9 (1), 382–385.
- Thevarajan, I, Nguyen, THO, Koutsakos, M, et al., 2020. Breadth of concomitant immune responses prior to patient recovery: a case report of non-severe COVID-19. *Nat. Med.* 26 (4), 453–455.
- Thomas, K, Eisele, J, Rodriguez-Leal, FA, et al., 2016. Acute effects of alemtuzumab infusion in patients with active relapsing-remitting MS. *Neurol. Neuroimmunol. Neuroinflamm.* 3 (3), e228.
- Thomas, K, Sehr, T, Proschmann, U, et al., 2017. Fingolimod additionally acts as immunomodulator focused on the innate immune system beyond its prominent effects on lymphocyte recirculation. *J. Neuroinflammation* 14 (1), 41.
- Tuohy, O, Costelloe, L, Hill-Cawthorne, G, et al., 2015. Alemtuzumab treatment of multiple sclerosis: long-term safety and efficacy. *J. Neurol. Neurosurg. Psychiatry* 86, 208–215.
- Vågberg, M, Kumlin, U, Svenningsson, A, 2012. Humoral immune response to influenza vaccine in natalizumab-treated MS patients. *Neurol. Res.* 34 (7), 730–733.
- Vollmer, B, Vollmer, T, Corboy, J, Alvarez, E, 2020. Evaluation of risk factors in developing lymphopenia and hypogammaglobulinemia in anti-CD20 treated multiple sclerosis patients. *Neurology* S29.002.
- von Hehn, C, Howard, J, Liu, S, et al., 2017. Immune response to vaccines is maintained in patients treated with dimethyl fumarate. *Neurol. Neuroimmunol. Neuroinflamm.* 5 (1), e409. <https://doi.org/10.1212/NXLI.0000000000000409>. eCollection 2018 Jan.
- Wang, D, Hu, B, Hu, C, et al., 2020b. Clinical Characteristics of 138 Hospitalized Patients With 2019 Novel Coronavirus-Infected Pneumonia in Wuhan, China. *JAMA* 323, 1061. <https://doi.org/10.1001/jama.2020.1585>.
- Wang, K, Chen, W, Zhou, YS, et al., 2020a. SARS-CoV-2 invades host cells via a novel route: CD147-spike protein. *BioRxiv*. <https://doi.org/10.1101/2020.03.14.988345>.
- Wang, X, Xu, W, Hu, G, et al., 2020c. SARS-CoV-2 infects T lymphocytes through its spike protein-mediated membrane fusion. *Cell. Mol. Immunol.* <https://doi.org/10.1038/s41423-020-0424-9>. [Epub].
- Warny, M, Helby, J, Nordestgaard, BG, et al., 2018. Lymphopenia and risk of infection and infection-related death in 98,344 individuals from a prospective Danish population-based study. *PLoS Med* 15 (11), e1002685.
- Weiss, P, Murdoch, DR., 2020. Clinical course and mortality risk of severe COVID-19. *Lancet* 395 (10229), 1014–1015 Mar.
- Wen, W, Su, W, Tang, H, et al. immune cell profiling of covid-19 patients in the recovery stage by single-cell sequencing. medRxiv. doi: 10.1101/2020.03.23.20039362.
- Willis, MD, Robertson, NP., 2020 d. Multiple sclerosis and the risk of infection: considerations in the threat of the novel coronavirus, COVID-19/SARS-CoV-2. *J. Neurol.* <https://doi.org/10.1007/s00415-020-09822-3>. [Epub].
- Wilk, AJ, Rustagi, A, Zhou, NQ, et al., 2020. A single cell atlas of the peripheral immune response to severe COVID-10. *MedRxiv* <https://doi.org/10.1101/2020.04.17.20069930>.
- Wijnands, JMA, Zhu, F, Kingwell, E, et al., 2018. Disease-modifying drugs for multiple sclerosis and infection risk: a cohort study. *J. Neurol. Neurosurg. Psychiatry.* 89, 1050–1056.
- Wray, S, Havrdova, E, Snyderman, DR, et al., 2019. Infection risk with alemtuzumab decreases over time: pooled analysis of 6-year data from the CAMMS223, CARE-MS I, and CARE-MS II studies and the CAMMS03409 extension study. *Mult. Scler.* 25, 1605–1617.
- Xiang, F, Wang, X, He, X, et al., 2020. Antibody Detection and Dynamic Characteristics in Patients with COVID-19. *Clin. Infect. Dis.* <https://doi.org/10.1093/cid/ciaa461>. pii: ciaa461[Epub ahead of print].
- Xu, Y, Li, X, Zhu, B, Liang, H, et al., 2020a. Characteristics of pediatric SARS-CoV-2 infection and potential evidence for persistent fecal viral shedding. *Gong S. Nat. Med.* 26, 502–505.
- Xu, Z, Shi, L, Wang, Y, et al., 2020b. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir. Med.* 8, 420–422.
- Yang, ZY, Huang, Y, Ganesh, L, et al., 2004. pH-dependent entry of severe acute respiratory syndrome coronavirus is mediated by the spike glycoprotein and enhanced by dendritic cell transfer through DC-SIGN. *J. Virol.* 78 (11), 5642e5650.
- Yang, Y, Xiong, Z, Zhang, S, et al., 2005. Bcl-xL inhibits T-cell apoptosis induced by expression of SARS coronavirus E protein in the absence of growth factors. *Biochem. J.* 392, 135–143.
- Yao, XH, Li, TY, He, ZC, et al., 2020. A pathological report of three COVID-19 cases by minimally invasive autopsies. *W. Zhonghua Bing Li Xue Za Zhi.* 49 (0), E009.
- Yao, H, Lu, X, Chen, Q, et al., 2020a. Patient-derived mutations impact pathogenicity of SARS-CoV2. medRxiv, 20060160. <https://doi.org/10.1101/2020.04.14.20060160>.
- Ye, G, Pan, Z, Pan, Y, et al., 2020a. Clinical characteristics of severe acute respiratory syndrome coronavirus 2 reactivation. *J. Infect.* <https://doi.org/10.1016/j.jinf.2020.03.001>. pii: S0163-4453(20)30114-6.
- Yednock, TA, Cannon, C, Fritz, LC, et al., 1992. Prevention of experimental autoimmune encephalomyelitis by antibodies against alpha 4 beta 1 integrin. *Nature* 356, 63–66.
- Yen, YT, Liao, F, Hsiao, CH, et al., 2006. Modelling the early events of severe acute respiratory syndrome coronavirus infection in vitro. *BA. J. Virol.* 80, 2684–2693.
- Zhang, Y, Gao, Y, Qiao, L, et al., 2020a. Inflammatory response cells during acute respiratory distress syndrome in patients with coronavirus disease 2019 (COVID-19). *Ann. Intern. Med.* <https://doi.org/10.7326/L20-0227>.
- Zhang, T, Sun, LX, Feng, RE, 2020b. Comparison of clinical and pathological features between severe acute respiratory syndrome and coronavirus disease 2019]. *Zhonghua Jie He He Hu Xi Za Zhi.* 43 (0). <https://doi.org/10.3760/cma.j.cn112147->

- 20200311-00312. Apr 3[Epub ahead of print].
- Zhao, J, Yuan, Q, Wang, H, et al., 2020. Antibody responses to SARS-CoV-2 in patients of novel coronavirus disease 2019. *Clin. Infect. Dis.* 2020. <https://doi.org/10.1093/cid/ciaa344>. pii: ciaa344.[Epub].
- Zheng, M, Gao, Y, Wang, G, et al., 2020. Functional exhaustion of antiviral lymphocytes in COVID-19 patients. *Cell. Mol. Immunol.* <https://doi.org/10.1038/s41423-020-0402-2>.
- Zhou, P, Yang, XL, Wang, XG, et al., 2020. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* 579, 270–273.
- Zhu, J, Ji, P, Pang, J, Zhong, Z, et al., 2020b. Clinical characteristics of 3,062 COVID-19 patients: a meta-analysis. *J. Med. Virol.* <https://doi.org/10.1002/jmv.25884>. Apr [Epub].
- Zhu, N, Zhang, D, Wang, W, et al., 2020a. China Novel Coronavirus investigating and research team. a novel coronavirus from patients with pneumonia in China, 2019. *N Engl. J. Med.* 382, 727–733.
- Zoehner, G, Miclea, A, Salmen, A, et al., 2019. Reduced serum immunoglobulin G concentrations in multiple sclerosis: prevalence and association with disease-modifying therapy and disease course. *Ther. Adv. Neurol. Disord.* 12, 1756286419878340.