

SCIENTIFIC REPORTS



OPEN

Effects of multiple environmental variables on tundra ecosystem respiration in maritime Antarctica

Tao Bao¹, Renbin Zhu¹, Xianglan Li², Wenjuan Ye¹ & Xiao Cheng²

Summertime ecosystem respiration (ER) rates through seven sites were measured at an upland tundra on Fildes Peninsula in maritime Antarctica to investigate effects of topographic gradient, vegetation types and climatic factors on tundra ER rates. Overall the highest ER rates occurred at the tundra top, followed at the middle slope, and the lowest rates at the lower slope. The daily highest ER rates occurred at noon whereas the lowest at 6 am. There was a significant positive correlation ($P < 0.05$) between tundra ER and 0–10 cm soil temperature, but a significant negative correlation ($P < 0.01$) between ER and soil moisture. A high Q_{10} value of 2.69 was obtained when all the data were combined, indicating soil temperature sensitivity of tundra ER. The mean ER at the tundra sites with moss coverage ($72.2 \pm 4.4 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$) was significantly higher ($P < 0.01$) than that at the sites with lichen coverage ($46.8 \pm 8.7 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$). The tundra sites without snow coverage experienced significant CO_2 release, whereas the emission through ER was very low at the tundra sites with snow coverage. Our results indicated that topographic gradient, soil temperature, soil moisture, vegetation types and snow coverage might affect tundra ER in maritime Antarctica.

Carbon dioxide (CO_2) is key active greenhouse gas (GHG) contributing to global warming¹. Increasing atmospheric concentration of CO_2 has stimulated research on its emission from both terrestrial and aquatic environments¹. Ecosystem respiration (ER), as a key component of global carbon cycle, is one of important sources for atmospheric CO_2 . At present, the CO_2 emissions through ecosystem respiration have been extensively investigated from temperate, subtropical and tropical terrestrial ecosystems, and they are affected by the geographical differences at the global scale^{1–3}. Polar Regions would be the most vulnerable to climate change in the world, and numerous researches about effects of climate warming on CO_2 fluxes have occurred in Arctic tundra ecosystems^{4–7}. The studies about CO_2 emissions from Antarctic terrestrial ecosystem mainly concentrated on the McMurdo Dry Valleys of continental Antarctica^{8–10}. However, only few data about CO_2 emissions are available from maritime Antarctic tundra according to the limited references^{11–13}.

To better understand the relationships of CO_2 emissions to environmental parameters, ER rates and net CO_2 fluxes have been measured at a number of sites of boreal or Arctic region and some sites of Antarctic region for the past three decades. Soil physicochemical properties, soil microbial community structure, and vegetation coverage are important factors affecting the ecosystem carbon balance and ER in Arctic and sub-Arctic areas^{4,5}. Higher CO_2 emissions from the tundra are predicted to become a positive feedback to climate warming as a result of increased soil organic matter decomposition and consequent higher nutrient availability⁶. Soil moisture directly affects vegetation distribution and functioning, and controls microbial decomposition influencing soil respiration, and thus it has both direct and indirect effects on CO_2 exchange in tundra ecosystems¹⁴. Vegetation composition and productivity have a direct effect on tundra ER¹⁰. Significantly weak and spatially variable CO_2 emissions were found in the McMurdo Dry Valleys, which are located in continental Antarctica, due to low soil organic carbon contents, and they were driven mainly by soil temperature and moisture, and their interaction with biological elements^{8–10,15}. However, the results from Arctic regions or the McMurdo Dry Valleys of continental Antarctica might not be applied to maritime Antarctic tundra due to differences in vegetation communities, soil nutrients and climatic conditions. The available works are few, and more data about tundra CO_2 emissions

¹Institute of Polar Environment & Anhui Key Laboratory of Polar Environment and Global Change, School of Earth and Space Sciences, University of Science and Technology of China, Hefei, 230026, China. ²College of Global Change and Earth System Science, Beijing Normal University, Beijing, 100875, China. Correspondence and requests for materials should be addressed to R.Z. (email: zhurb@ustc.edu.cn)

are needed in maritime Antarctica^{11–13}. Therefore it is very necessary to conduct the research about CO₂ emission from the tundra in maritime Antarctica.

In maritime Antarctica, the response of tundra ecosystem is sensitive to climate warming, accompanied by the regional changes in precipitation^{16,17}. Warming climate might cause several consequences in the tundra ecosystem, especially in soil formation and the spatial extent of expanding vegetation¹². The most common moss vegetation (*Bryum Pseudotriquetrum* and *Bryum muelenbeckii*) and native vascular plant species (*Colobanthus quitensis* and *Deschampsia antarctica* Desv) have recently been expanding in maritime Antarctic tundra^{16,18–20}. These changes would affect tundra ER rates, and net CO₂ fluxes since tundra vegetation composition and soil physical properties are related to soil organic carbon content, potential mineralization and microorganism activity^{4,21,22}. Therefore, the measurements of ER rates are essential to understand the C cycling dynamic in the maritime Antarctic tundra, and their feedbacks to the regional warming climate¹². Summertime CH₄, N₂O and CO₂ fluxes, and ER rates have also been *in situ* observed from various tundra ecosystems in maritime Antarctica^{13,19,23–25}. The increasing temperatures might decrease tundra CO₂ sink accompanied by the increase in soil respiration^{13,19}. Tundra soil and vegetation are closely linked to the global carbon cycling dynamics, especially CO₂ exchange in maritime Antarctica^{11–13}. More data about tundra ER are required to improve our knowledge about CO₂ exchange.

During the austral summer of 2014/2015, the ER rates at seven sites were measured in an upland tundra on Fildes Peninsula, maritime Antarctica. The objectives of this study were (1) to study summertime and daily variability in tundra ER; (2) to investigate effects of topographic gradient on tundra ER; (3) to examine effects of soil temperature, soil moisture, snow coverage and vegetation types on tundra ER. This is an attempt to increase the tundra ER data set in order to reasonably estimate carbon budget, and to find out environmental variables affecting ER in maritime Antarctica.

Results

Climate and tundra soil characteristics. During the austral summer of 2014/2015, the mean air temperature (AT) showed a small increase from Dec 20, 2014 to Feb 18, 2015, and the daily minimum AT was often below 0°C while the maximum AT was generally above 3°C (Fig. 1). The total precipitation was 114 mm, and total sunlight time (ST) was 117 h. The air temperature, ground temperature (GT) and 10 cm soil temperature (ST₁₀) showed a similar change at the sites GW1, GW2 and GW3, and they increased gradually from December, 2014 to January, 2015, but soil temperatures decreased with soil depths (Fig. 2). The mean AT, GT and ST₁₀ were 3.4 ± 0.6°C, 5.9 ± 1.0°C and 1.5 ± 0.3°C, respectively.

Physicochemical properties of the tundra soils were summarized in Table 1. The mean soil moisture (SM) decreased from the lower slope (59.0%) to the top of tundra (30.4%). The TOC contents and C/N ratios were almost equal to each other in all the soil samples, ranging from 6.7 to 7.1% and from 9.6 to 10.3, respectively, and the tundra soils were neutral with the pH of 6.8–7.3. However, soil TN, NH₄⁺-N, NO₃⁻-N and TS contents were highly variable from the lower slope to the top of tundra due to environmental heterogeneity. The mean soil TN and TS contents ranged from 0.23 to 0.32 mg g⁻¹ and from 0.07 to 0.15 mg g⁻¹, respectively. The mean soil NH₄⁺-N and NO₃⁻-N concentrations increased from the lower slope to the top of the tundra.

Summertime tundra ER along the topographic gradient. During the observation period, tundra ER rates showed a similar fluctuation at the tundra top site GW1, the middle slope site GW2, and the lower slope site GW3. All the three sites experienced a significant release of CO₂ (Fig. 2). The ER rates ranged from 55.4 to 127.2 mg CO₂ m⁻² h⁻¹ at GW1, from 38.5 to 125.7 mg CO₂ m⁻² h⁻¹ at GW2, and from 34.8 to 114.0 mg CO₂ m⁻² h⁻¹ at GW3, respectively. The ER rates showed a significant difference (P < 0.05) between the sites GW1, GW2 and GW3. Overall the highest mean ER rate occurred at GW1 (mean 91.2 ± 10.6 mg CO₂ m⁻² h⁻¹), followed at GW2 (mean 76.9 ± 11.4 mg CO₂ m⁻² h⁻¹), and the lowest at GW3 (mean 67.3 ± 9.5 mg CO₂ m⁻² h⁻¹) (Fig. 2d).

Tundra ER showed a similar variation with air temperature, ground temperature and 10 cm soil temperature during the observation period (Fig. 2c). Overall, tundra ER rates were significantly affected by soil temperature (F = 3.67, P < 0.05), soil moisture (F = 6.36, P < 0.01) and their interaction (F = 3.49, P < 0.05) (Table 2). In addition, there was a significant exponential correlation (R² = 0.37, P < 0.05) between tundra ER and 0–10 cm mean soil temperature. Tundra ER showed a significant negative correlation (R² = 0.60, P < 0.01) with soil moisture (Fig. 3a). Tundra ER can be more accurately predicted using a bivariate linear model that incorporates both soil temperature and moisture as independent variables (Table 3). A high Q₁₀ value of 2.69 was obtained, indicating soil temperature sensitivity of summertime ER in maritime Antarctic tundra. The Q₁₀ showed a negative correlation (R² = 0.83, P < 0.01) with tundra soil temperatures, but a significant positive correlation (R² = 0.81, P < 0.01) with soil moisture based upon a linear regression model (Fig. 3b). No significant correlation (P > 0.05) was found between tundra ER and soil chemical properties (TOC, NH₄⁺-N, NO₃⁻-N, C: N) at all the observation sites although the differences in the emission rates might reflect the ecological and environmental heterogeneity at the different tundra sites.

Diel tundra ER. The ER showed a similar diurnal pattern at the top site GW1 and the slope site GW2 on Feb 2, 2015. The highest emission rates both occurred at noon (10 am to 14 pm), and the lowest at 6 am (Fig. 4a). The daily mean ER rate at GW1 (49.9 mg CO₂ m⁻² h⁻¹) was significantly higher than that at GW2 (32.8 mg CO₂ m⁻² h⁻¹). On Feb 13, the ER rates also showed a similar diurnal pattern at GW1 and GW2, with the lowest ER at 6 am. However, the abnormal low ER rates occurred at 2:00 pm, corresponding to significantly enhanced soil moisture (Fig. 4b). The sudden precipitation took place at that moment, thus high soil water content could suppress soil CO₂ release. The CO₂ emission rates ranged from 51.1 to 95.0 mg CO₂ m⁻² h⁻¹ at GW1 and from 40.0 to 69.0 mg CO₂ m⁻² h⁻¹ at GW2, and mean emission rate (72.0 ± 7.2 mg CO₂ m⁻² h⁻¹) at GW1 was also significantly higher

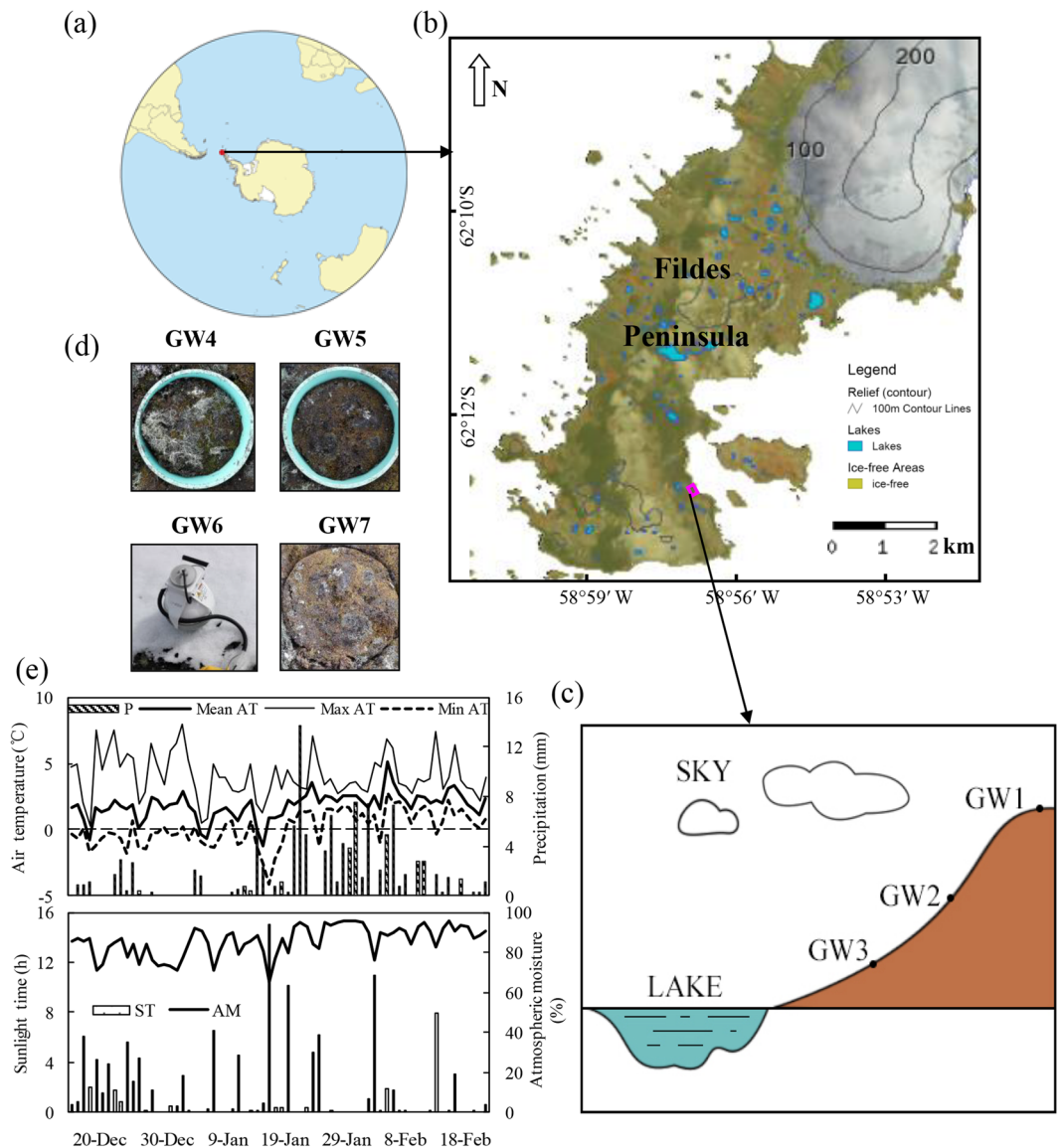


Figure 1. (a) The dot indicates location of the investigation area in maritime Antarctica. (b) Location of the study sites in Fildes Peninsula. (c) The position of the investigation sites GW1, GW2 and GW3 in upland tundra of Fildes Peninsula; (d) The background of the sites GW4, GW5, GW6 and GW7; (e) The climate conditions during tundra ER measurements. Note: The map was drawn using CorelDRAW 2017 (<http://www.corel.com/cn/>) and Microsoft Excel 2016 (<https://products.office.com/zh-cn/excel>) software.

than that at GW2 ($49.0 \pm 4.9 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$). Calculated from 6 individual measurements over 24 h in upland tundra, ER rates between 6:00 and 10:00 were close to the mean diurnal tundra ER.

The daily ER rates showed a similar trend to ground temperatures, and two ER peaks coincided with those of ground temperatures, indicating that high temperature increased tundra ER. Tundra ER showed the lowest values in the morning corresponding to the lowest ground temperature (Fig. 4). The soil temperatures show a large fluctuation at GW1 and GW2, whereas the fluctuation of soil moisture was small, and almost stable. Overall soil temperature was higher, but soil moisture was lower at GW1 than at GW2.

Tundra ER under different vegetation coverage. The tundra ER rates at lichen-covered (GW4) and moss-covered (GW5) tundra sites ranged from 23.8 to $64.9 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$, and from 63.4 to $84.0 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$, respectively. The mean ER at GW5 (three subsites, $n = 9$, $72.2 \pm 4.4 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$) was significantly higher ($P < 0.01$) than that at GW4 (three subsites, $n = 9$, $46.8 \pm 8.7 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$), indicating that moss-covered tundra soil could release more CO_2 than lichen-covered tundra soil (Fig. 5a). For the two sites, ER rates were significantly affected by soil moisture ($F = 10.37$, $P < 0.01$), vegetation types ($F = 22.70$, $P < 0.001$) and their interaction ($F = 6.06$, $P < 0.01$) (Table 2), but no significant change was affected by soil temperature (Table 2), although ER rates increased with soil temperature (Fig. S1).

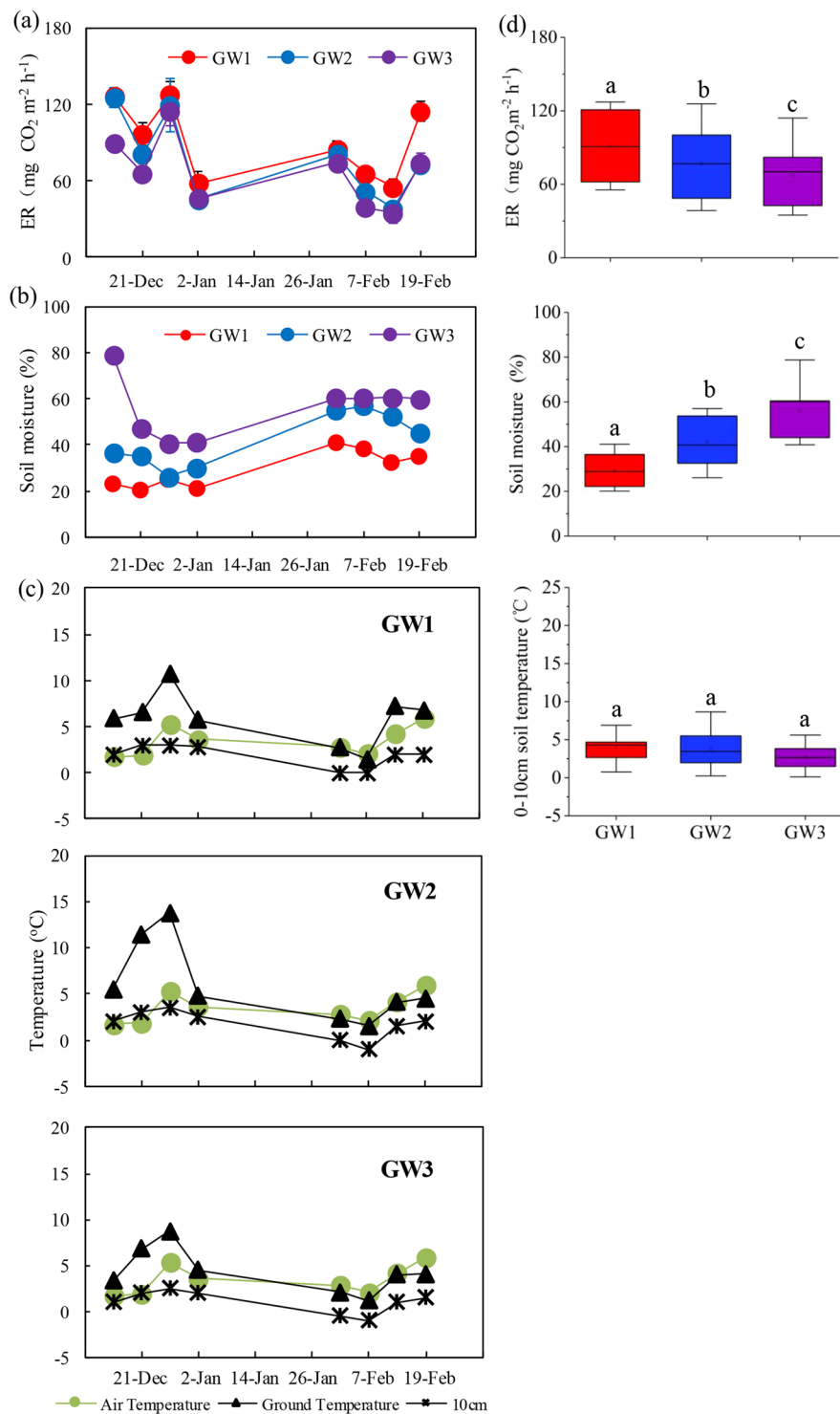


Figure 2. The summertime ER ($\text{mg CO}_2 \text{ m}^{-2} \text{ h}^{-1} \pm \text{SE}$, $n = 3$) along the topographic gradient at an upland tundra. Panel (a) ER, (b) soil moisture, (c) air and soil temperature, and (d) The comparisons of the ER, soil moisture and soil temperature between the sites GW1, GW2 and GW3. Note: GW1, GW2 and GW3 were located at the tundra top, the middle slope and at the lower tundra slope. Error bar indicates standard error of the means ($n = 3$). Boxes enclose the interquartile range, whiskers show the full range. The different lowercase letters indicate statistically significant differences between the means (Fisher's LSD, $P < 0.05$).

Tundra ER under snow coverage. The ER rates showed a large difference between tundra sites with snow coverage (GW6) and without snow coverage (GW7) (Fig. 5b). The site GW7 experienced a significant CO_2 release with the mean emission rate of $67.2 \pm 11.1 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$ (three subsites, $n = 9$), whereas the ER rates at the site

Tundra Sites	Soil Properties							Characteristic plant species
	SM (%)	pH	TOC (%)	NH ₄ ⁺ -N (μg g ⁻¹)	NO ₃ ⁻ -N (μg g ⁻¹)	C/N	S (mg g ⁻¹)	
GW1	30.4	7.3	6.86	0.62	0.33	9.7	0.15	<i>Bryum pseudotriquetrum</i> , <i>Bryum muelenbeckii</i> , <i>Sanionia uncinata</i> , <i>Warnstorfia sarmentosa</i> , <i>Usnea sp.</i>
GW2	45.7	7.1	6.96	0.14	0.17	9.6	0.13	
GW3	59	6.8	6.66	0.06	0.19	10	0.08	
GW4	42.8	6.9	6.67	0.24	0.19	10.3	0.07	
GW5	44.9	7	6.98	0.16	0.16	9.6	0.12	
GW6	54.6	7.2	6.96	0.22	0.18	9.7	0.12	
GW7	47	7.2	7.12	0.24	0.22	9.6	0.14	

Table 1. Soil properties and plant species at the sites for the upland tundra in maritime Antarctica. Note: SM, TOC and C/N indicated soil moisture, total organic carbon and the ratios of soil organic carbon and nitrogen, respectively.

	F	P
GW1–3 Combined (n = 24)		
DA	12.06	<0.001 ^c
ST	3.67	<0.05 ^a
SM	6.36	<0.01 ^b
ST*SM	3.49	<0.05 ^a
GW4–5 Combined (n = 18)		
DA	17.84	<0.001 ^c
ST	2.99	0.112
SM	10.37	<0.01 ^b
VT	22.70	<0.001 ^c
ST*SM	0.25	0.627
ST*VT	2.23	0.147
SM*VT	6.06	<0.01 ^b
ST*SM*VT	2.62	0.114
GW6–7 Combined (n = 18)		
DA	4.26	<0.05 ^a
ST	21.77	<0.001 ^c
SM	5.49	<0.05 ^a
SC	28.10	<0.001 ^c
ST*SM	5.21	<0.05 ^a
ST*SC	24.50	<0.001 ^c
SM*SC	15.16	<0.001 ^c
ST*SM*SC	7.09	<0.01 ^b
GW1–7 Combined (n = 60)		
ST	9.03	<0.01 ^b
SM	6.63	<0.01 ^b
VT	7.35	<0.01 ^b
SC	22.96	<0.001 ^c
ST*SM	5.56	<0.01 ^b
ST*SM*VT	1.70	0.193
ST*SM*SC	4.67	<0.05 ^a
ST*SM*VT*SC	1.03	0.316

Table 2. Tests of significance of date (DA), soil temperature (ST), soil moisture (SM), vegetation types (VT), snow coverage (SC) and their interactions on the tundra ER using multivariate ANOVA (F and P values). Note: Superscripts a, b and c indicate significant effects at P < 0.05, 0.01 and 0.001, respectively.

GW6 was almost extremely low (three subsites, n = 9). The ER rates at the tundra sites without snow coverage was significantly higher (P < 0.01) than those at the sites with snow coverage, indicating that accumulated snow could greatly limit ecosystem respiration or prevent CO₂ emissions from the tundra ecosystem. Overall, snow coverage had a significant effect (F = 28.10, P < 0.001) on tundra ER rates, and the interaction among soil temperature, soil moisture and snow coverage on ER rates (F = 7.09, P < 0.01) was also observed during the observation period (Table 2).

Sites	Equation		
	$F_{ER} = aW + bT + c$	R^2	n
GW1	$F_{ER} = 47.44 - 0.42 W + 8.11 T$	0.46	8
GW2	$F_{ER} = 78.88 - 0.45 W + 4.46 T$	0.49	8
GW3	$F_{ER} = 25.46 - 1.05 W + 12.59 T$	0.42	8
GW1–3 Combined	$F_{ER} = 67.60 - 0.24 W + 6.11 T$	0.54	24

Table 3. Correlations of summertime soil respiration rates and soil moisture, 0–10 cm mean soil temperature at the tundra sites of the upland tundra. Note: Only the equations and R^2 values significant at $p < 0.05$ are shown. F_{ER} : Soil CO_2 respiration rates; W : Soil moisture; T : 0–10 cm mean soil temperature.

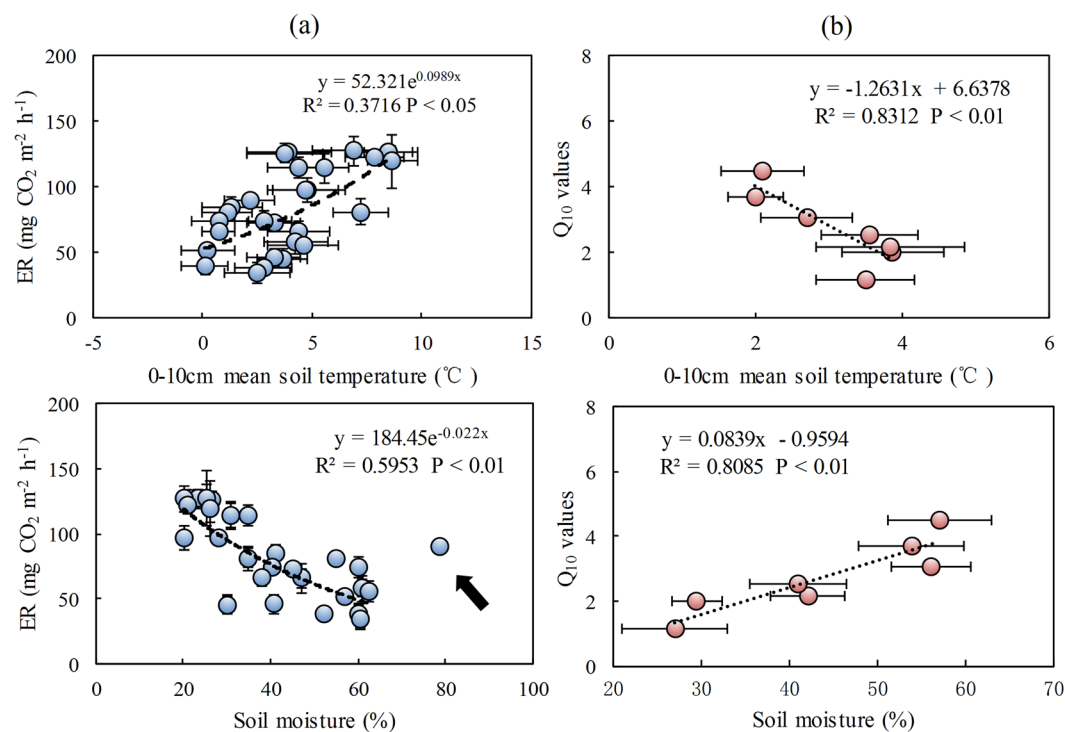


Figure 3. Panel (a) Relationship between ER and 0–10 cm mean soil temperature, soil moisture at the tundra sites. (b) Correlations between tundra Q_{10} values and soil temperatures, soil moisture. The arrow indicates outlier data, which are excluded from the correlation. Note: the figures for Q_{10} fitting at each site were given in Supplementary Material.

Discussion

Previous climate model simulations have shown that both annual and seasonal responses of terrestrial carbon cycle to climate change will change in the 21st century, but with large uncertainty at high latitudes (Antarctic and Arctic)^{26,27}. This study is therefore valuable for the research of terrestrial carbon cycle modelling and the comparisons of mean CO_2 emission rates within different Antarctic regions could approximately reflect the differences of ER. In our study area, the mean CO_2 emission ($46.8\text{--}91.2\text{ mg CO}_2\text{ m}^{-2}\text{ h}^{-1}$) at upland tundra sites was larger than those measured for soils in the Taylor Valley ($15.8\text{--}23.8\text{ mg CO}_2\text{ m}^{-2}\text{ h}^{-1}$)¹⁵, at Rothera Point of Marguerite Bay ($23.8\text{--}30.1\text{ mg CO}_2\text{ m}^{-2}\text{ h}^{-1}$), and at Anchorage Island ($25.3\text{--}33.3\text{ mg CO}_2\text{ m}^{-2}\text{ h}^{-1}$)¹², and close to those ($26.4\text{--}124.1\text{ mg CO}_2\text{ m}^{-2}\text{ h}^{-1}$) measured on lakeshore soils in Garwood Valley, Antarctica⁸. In addition, our results were comparable to those measured at a seal colony ($70.8 \pm 7.6\text{ mg CO}_2\text{ m}^{-2}\text{ h}^{-1}$) and the tundra sites adjacent to a penguin colony¹³. However, the mean ER rates in the study area were almost one tenth of those measured at penguin colony tundra ($201.3 \pm 31.4\text{ mg CO}_2\text{ m}^{-2}\text{ h}^{-1}$)^{13,19}. Higher soil organic carbon level at penguin colony than our study area might provide the favorable environment for microbial activity, resulting in enhanced soil respiration^{13,19}. Hopkins *et al.*²⁸ found that the composition and mineralization of lacustrine detritus and soil organic matter affected soil respiration in Antarctic Garwood Valley soils. However, soil chemical properties were similar to each other at our sites, and no significant correlation ($P > 0.05$) was found between tundra ER and soil TOC, other chemical properties at all the observation sites in our study area.

Overall the highest ER rates occurred at the tundra top, followed at the middle slope, and the lowest emission rate at the lower slope (Fig. 2), indicating that topographic gradient might have an important effect on spatial

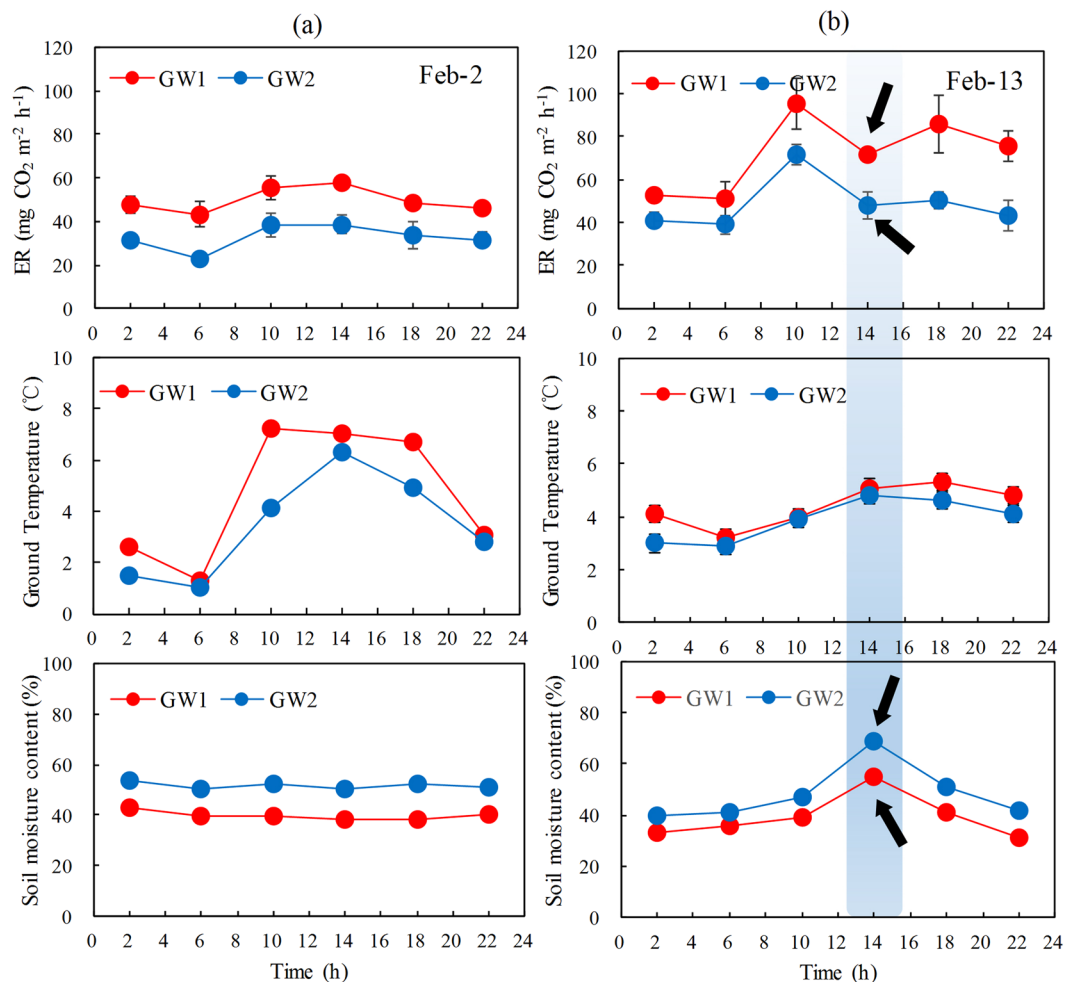


Figure 4. Diurnal variations of tundra ER, soil temperature and soil moisture at the tundra sites GW1 and GW2.

variability in tundra ER. Topography is the primary determinant of soil moisture patterns across arctic landscapes and plays a major role in determining the distribution of vegetation types and plant community succession²⁹. The variation in plant biomass, rates of primary productivity, soil organic matter (SOM) quantity and quality, nutrient status and soil chemistry are all tightly linked to topography³⁰. In total, topography results in spatial variations in environmental variables through establishing elevation gradients, and in turn influences the rates of tundra carbon transformation processes. In our study area, soil temperature increased with the elevation from GW3 to GW1, but soil moisture significantly decreased with the elevation. Our results indicated that topography gradient plays critical roles through their influences on soil moisture, soil temperature and their interaction that in turn affect ER and C biogeochemical cycling in the tundra ecosystems of maritime Antarctica.

On the other hand, corresponding topographical gradients of soil moisture and temperature tend to occur at the landscape and larger spatial scales across the Arctic³¹. In maritime Antarctica, tundra ecosystems generally developed on the elevated, hilly and uneven upland along the coast. Faster rates of organic matter mineralization and decomposition (i.e. CO₂ emissions) occur in inter-hummock areas^{13,30}. In this study, higher soil moisture and lower temperature occurred at the slope site GW3 which might limit soil organic matter decomposition by microorganisms, resulting in soil organic matter accumulation in low-lying areas³⁰. Therefore topography is a key factor for the evaluation of tundra ER at the tundra landscape and larger spatial scales in maritime Antarctica.

The temporal variation range of ER (34.9 to 127.2 mg CO₂ m⁻² h⁻¹) was considerable as tundra ER varied with soil temperatures (Fig. 2). The ER rates showed a similar diurnal pattern at the top site GW1 and the slope site GW2, corresponding to ground temperature (Fig. 4). It was demonstrated that there was a diurnal cycle both of tundra photosynthesis and respiration in the Antarctic Peninsula¹², and our results corroborated previous research. Overall the ER rates showed a significant positive correlation ($P < 0.05$) with soil temperatures in maritime Antarctic tundra (Fig. 3a). The activities of soil or plant enzymes and soil microorganisms were gradually enhanced with the increase in soil temperatures, and then led to increased ecosystem respiration rates, whereas the low temperature would restrict the respiration of the vegetation and soil microorganisms, thus reduced tundra ER rates^{9,12}. Similarly, the ER generally showed a strong positive correlation with soil temperature in Arctic tundra, and climate warming might decrease CO₂ sink through increase in tundra ER^{9,32}. In McMurdo Dry Valley of east Antarctica, several studies also showed a similar correlation between ER and soil temperatures^{10,33}. Overall

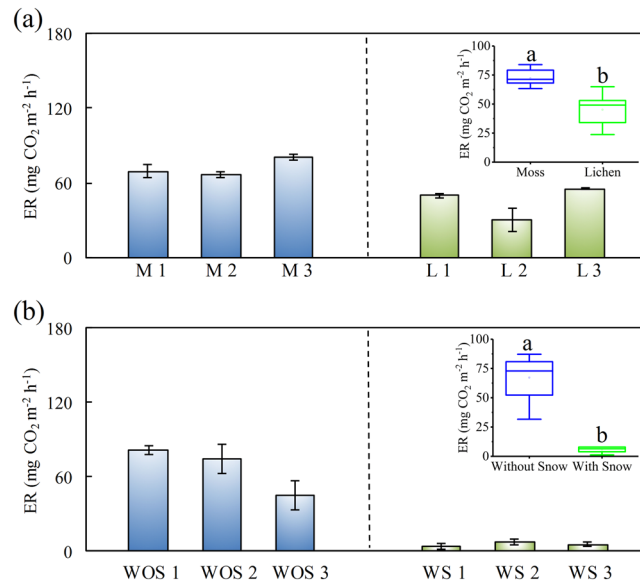


Figure 5. Effects of (a) vegetation types and (b) snow coverage on ER rates at the upland tundra sites. M 1–3 and L 1–3 indicated the sub-sites with moss coverage and with lichen coverage, respectively. WOS 1–3 and WS 1–3 indicated the sub-sites without snow coverage and with snow coverage, respectively. Boxes enclose the interquartile range, whiskers show the full range. The different lowercase letters indicate statistically significant differences between the means (Fisher's LSD, $P < 0.05$).

ER indicated greater temperature sensitivity in Polar Regions than other global regions. The faster increase in temperatures have occurred in maritime Antarctica compared with other global regions in the past few decades¹⁷. The positive correlation of ER with soil temperature might reflect the positive feedback of tundra ecosystem respiration to current warming climate in maritime Antarctica.

A high Q_{10} value ($Q_{10} = 2.69$) showed soil temperature sensitivity of summertime ER in maritime Antarctic tundra. The Q_{10} values for soil respiration ranged from 2.00 to 3.00 on Antarctic Peninsula, and from 2.00 to 4.40 in the Garwood Valley, east Antarctica²⁸. Raich & Schlesinger² reported a Q_{10} range of 1.30–3.30 for different biomes of the world. Our observed Q_{10} values were within the high results reported in the references. Prior research suggested that Q_{10} could vary according to the range of temperatures used for the calculation, and highlighted that it could be subjected to inter-annual and seasonal variations³⁴. In this study, the Q_{10} values showed a negative correlation with tundra soil temperatures (Fig. 3b). Generally the Q_{10} values for ER decreases with the increase in soil temperature^{2,34}. The Q_{10} dependence on temperature might be related with the response of soil microorganisms or soil enzymatic activities to different temperature ranges^{2,34,35}. Significant exponential correlations between tundra ER and soil temperatures (Fig. 3a) indicated that climate warming might decrease tundra CO₂ sink through increase in soil respiration, thus affect tundra CO₂ exchange in maritime Antarctica.

A significant negative correlation ($P < 0.01$) between ER and soil moisture (Fig. 3a) indicated that soil moisture might have an important effect on ER rate. Similarly, a weak negative correlation between soil respiration and soil moisture was found in the McMurdo Dry Valleys, east Antarctica^{10,33}. Soil moisture directly affected tundra vegetation distribution and functioning, and played major roles in structuring soil microbial communities by constraining their physiology and distribution and the chemical substrates used by soil microorganisms²⁸. High soil water content might prevent atmospheric oxygen from diffusing into the soil, and decreased the ecosystem respiration⁴. On the contrary, the drying of near-surface soil layers and consequent increased diffusion of soil gases might allow more rapid efflux of CO₂ emissions to the atmosphere³⁶. In this study, the diel variation of ER was also constrained by soil moisture, and the abnormal low ER occurred at 2:00 pm (Fig. 4b), corresponding to significantly enhanced soil moisture due to the heavy precipitation. Similarly, the diel variation in ER was largely controlled by soil moisture in a desert shrub ecosystem³⁵. Therefore the increase in soil moisture might have a significant impact on C and N mineralization^{10,33}, and further inhibited CO₂ exchanges between tundra ecosystem and atmosphere in maritime Antarctica.

The multivariate regression analysis indicated that about 70% of the variation in ecosystem respiration rate was jointly affected by soil temperature and soil moisture⁴. Generally it was difficult to independently differentiate the effects of soil temperature and soil moisture under field condition due to the hydrothermal interaction^{34,35}. When the soil temperature increased to 10–20 °C, effects of soil moisture on ER rate gradually increased, whereas soil moisture had little effect on ecosystem respiration when the soil temperature was lower than 5 °C^{37,38}. In our study area, the soil temperatures varied from –4 to 10 °C in the summer. However, soil moisture still had an important effect on tundra ER in maritime Antarctica under such low temperatures (Fig. 3a), and a positive correlation ($P < 0.05$) occurred between Q_{10} and soil moisture (Fig. 3b).

The mean ER at the sites with moss coverage was significantly higher than that at the sites with lichen coverage (Fig. 5a). Generally the mosses grow faster than the lichens, and thus have stronger plant respiration. Previous

studies have also demonstrated that CO₂ emission from terrestrial ecosystems was influenced by the types and distribution of tundra vegetation at King George Island and on the Antarctic Peninsula^{11,12}. Therefore vegetation type might be another important factor affecting tundra ER in maritime Antarctica. On the global scale, the ER varied with various vegetation types, and the mean ER rate ranged from 60 mg CO₂ m⁻² h⁻¹ in the tundra and cryolithozone to 1260 mg CO₂ m⁻² h⁻¹ in the tropical rainforest^{2,19,35}. Different vegetation communities could give rise to differences in CO₂ emissions through their influence on soil microbial processes^{31,36}. The distribution characteristics of tundra vegetation species and the closely related microbial communities are greatly affected by topographical moisture gradients. Lichens (*Usnea sp.*) preferentially occupy higher tundra soil positions and could dominate plant species in relatively tough and exposed environments. Mosses (*Bryum muelenbeckii* and *Bryum pseudotriquetrum*) tend to be found on lower slope positions with high moisture²⁹. Wet mosses are characterised by higher biomass and respiration and lower recycling of materials than dry mosses on Signy Island as reported by Davis³⁹. Therefore, the role that tundra plants and the related multiple environmental variables play in altering carbon flux cannot be ignored in maritime Antarctica.

In addition, the mean ER at the sites without snow coverage was significantly higher than that at the sites with snow coverage (Fig. 5b), indicating that snow coverage might decrease tundra ER. Snow coverage affects directly or indirectly litter decomposition and carbon release through the effects on soil temperature, water availability, soil fertility, soil microbial activity, and growing-season length^{40,41}. Several important researches in the low- and high-arctic tundra ecosystems have shown that ER still occurs at the sites under snow cover, and might provide an important contribution to the annual carbon balance, although snow cover could significantly reduce growing-season ER^{7,40,42}. In this study, the interactive effects of soil moisture, soil temperature and snow coverage on ER rates were found (Table 2) and the tundra sites with snow coverage experienced a weak, even extremely low CO₂ release. The accumulated snow might act as an insulating layer, and inhibit the gas exchange between the underlying tundra soil and the atmosphere^{7,41}. Moreover snow melting water increased soil moisture, and might further reduce ER rates⁵. The distribution and coverage of accumulated snow might be important factors affecting the spatial variation of tundra ER in maritime Antarctica.

In general, the synergetic effects among soil moisture, soil temperature, vegetation types and snow coverage on ER rates indicate the complexity of multiple-factors in terms of effects on tundra CO₂ emissions, inferring that more well-designed field experiments are needed to understand ecosystem processes and the factors affecting ER. In addition, tundra ER rates were obtained from the limited sites during short summertime due to local severe climatic conditions, inconvenient traffic and the manpower absence. Effects of multiple-factors on tundra CO₂ emissions need long-term multi-site observations to further evaluate the contribution of tundra ER to the annual carbon balance in maritime Antarctica.

Methods

Study area. The research area is situated on Fildes Peninsula (61°51′–62°15′S, 57°30′–59°00′W; an area of 30 km²) in the southwestern area of King George Island (Fig. 1). Communities of mosses and lichens represent the vegetation on this peninsula⁴³. An upland tundra (S62°12′59″, W58°57′52″), which is 500 m away from Great Wall Station, was selected as the study area for the measurements of ecosystem respiration. The upland tundra has the elevation of about 40 m a.s.l. The sampling grounds were covered completely by mosses with the dominant species of *Bryum pseudotriquetrum* and *Bryum muelenbeckii* or lichens (*Usnea sp.*), and the tundra surface was divided into two distinct areas covered with the mosses and the lichens. The depth of tundra vegetation layer is about 5–10 cm, and under the vegetation cover is an organic clay layer of about 10–15 cm¹⁹. A more detailed description about the study area was given by Zhu *et al.*¹⁹.

Investigation sites and experimental designment. During the summer of 2014/2015, seven sites (GW1–GW7) for the measurements of tundra ER were established along a gentle slope within the upland tundra as shown in Fig. 1: (i) Three observation sites GW1, GW2 and GW3 were equipped with three collars each to test effects of topographic gradient and environmental variables on tundra ER. The sites GW1, GW2 and GW3 were located at the tundra top (about 40 m a.s.l.), the middle slope (about 20 m a.s.l.) and the lower slope (about 10 m a.s.l.), respectively. GW3 was near a lake with the lowest altitude. These three tundra sites GW1–3 were all covered with mosses (*Bryum muelenbeckii* and *Bryum pseudotriquetrum*). The vegetation coverage was about 95% and their height was about 2–3 cm. There were almost no evident differences in the dominant moss species, their height and the phytomass across the sites GW1–3; (ii) The sites GW4 and GW5 were set up in the areas covered with lichens and mosses, respectively, and three sub-sites were selected for each site to study effects of tundra vegetation types on ER. These subsites were almost located at the middle slope (between 15–20 m a.s.l.) on the upland tundra; (iii) The sites GW6 and GW7 (three sub-sites for each site) were established to study effects of accumulated snow coverage on tundra ER. The ground at the site GW6 was covered with 5–6 cm depth accumulated snow, whereas the site GW7 was covered with no snow as the control. These tundra sub-sites were equipped with two collars each, and they were located at the same altitude (about 15 m a.s.l.) with moss vegetation.

In situ ER measurement. ER rates from tundra sites were determined using a CO₂ flux system (LI-COR Biosciences, Lincoln, NE, USA) with an infrared gas analyzer (model LI-8100A), a 20 cm diameter opaque survey chamber (model LI-8100-103) and a laptop to collect data⁴⁴. The opaque survey chamber was manually mounted on the PVC collar in each plot for the measurement of ER. This system operated at the same time during measurements and CO₂ concentrations were quantified through optical absorption spectroscopy. The system had been calibrated at the factory using precision gases (CO₂ gas standards) at controlled temperatures. Polyvinyl chloride (PVC) collars (21.3 cm outer diameter, 20.3 cm inner diameter and 12.5 cm high) were inserted into the soil in the center of each plot at each site, reaching a standard depth of 10 cm. This was done at least 24 h prior to the ER measurements to minimize the effects of the disturbance on CO₂ emissions. In the snow-covered tundra area, the

depth of accumulated snow was only 5–6 cm, and the insertion of 10 cm collar into tundra soil through snow layer was enough to provide the insulation for the measurements of CO₂ fluxes. During the measuring period, ER rates at each site were based on repetitive measurements over 1.5 min, during which time measurements were made of CO₂ concentrations inside the chamber at 3 s intervals⁴⁴. The three PVC collars were kept at each site during the whole observation period, and the order of measurements was varied to ensure that the measuring time did not bias the results, resulting in three replicate measurements per site. During the period from Dec 16, 2014 to Feb 2, 2015, the ER rates were measured eight times at the sites GW1, GW2 and GW3. The ER rates at GW4 and GW5 were measured three times on Dec 26, 2014, Jan 12 and Feb 1, 2016. The rates at GW6 and GW7 were also measured three times on Dec 22 and Dec 26, 2014 and Jan 12, 2015. In addition, the diurnal ER measurements were conducted from 2:00 am to 22:00 pm at GW1 and GW2 on Feb 2 and Feb 13, 2015. The measuring time interval was 4 h during the daytime. Normally, ER values are always positive representing CO₂ emission from the tundra ecosystem to the atmosphere.

Environmental variables and general soil properties. Soil temperature and soil moisture measurements were conducted at the same time as ER at each site. The monitoring systems consisted of soil temperature probes (Campbell L107E thermocouple, accuracy of ±0.2 °C) and soil moisture probes (CS656 water content reflectometer, accuracy of ±2.5%). These probes were placed at 10 cm depths where PVC soil collars were inserted to determine the ER. All probes were connected to a Campbell Scientific CR 1000 data logger, recording data at every 1 second interval during 1.5 min. Meteorological data, e.g. air temperature (AT), daily sunlight time (ST), total precipitation (TP), and ground temperature (GT) were obtained at the weather station of Great Wall Station.

Soil samples were collected in the plots after ER measurements completed in the summer of 2014/2015. The soils were sampled in a hermetically sealed bag and stored at 4 °C until analyzed. Soil moisture was determined after oven-drying field-moist soil samples to a constant mass at 105 °C (24 hours). Soil samples were homogenized manually, and each 10 g sub-sample (fresh weight) was extracted with 100 ml 1 M KCl solution for one hour and then filtered, and soil NH₄⁺-N and NO₃⁻-N concentrations were determined colorimetrically based on Berthelot's reaction and ion chromatography^{13,45}. TOC content was measured by Scheibler's volumetric method²⁰, and TN was quantified on an automatic elemental analyzer (Elementar Vario EL, Hanau, Germany). Soil pH was measured after a 1:3 (soil weight: extractant-volume) dilution of soil with deionized water.

Data analysis. All statistical analyses were performed using SPSS 20.0 (<http://www.spss.com.cn/>) and Microsoft Excel 2016 (<https://products.office.com/zh-cn/excel>) for Windows 10. The univariate exponential function model was used to analyze the relationship between ER and soil temperature according to our previous studies in maritime Antarctica¹³ and the related references^{46,47}:

$$ER = \beta_0 e^{\beta_1 T} \quad (1)$$

where ER is ecosystem respiration (mg CO₂ m⁻² h⁻¹), T is mean soil temperature (°C), and β₀ and β₁ are constants fitted with the least squares technique⁴³. Temperature coefficient (Q₁₀ value), an index of temperature dependence⁴⁸, has been widely used to describe the sensitivity of soil respiration to temperature in different types of terrestrial ecosystems as reviewed by Tuomi *et al.*⁴⁶. The Q₁₀ value was calculated as:

$$Q_{10} = e^{10\beta_1} \quad (2)$$

The Q₁₀ for the whole measurement period (Dec 2014 to Feb 2015) was computed on the basis of the daily averages of respiration rate and soil temperature¹³. The Q₁₀ and mean soil temperature, soil moisture during the measurement period were used to examine the relationships between Q₁₀ and soil temperature, soil moisture. Exponential regression analyses were used to explore the relationships between ER and soil temperature, soil moisture for all the sites. The single-factor and interaction effects of soil moisture, soil temperature, vegetation types and snow coverage on ER rates were detected using multi-way analysis of variance (ANOVA). Standard error (SE) was used to estimate the uncertainty of the mean of individual fluxes. All the data for ER rates were expressed as mean ± SE. The statistically significant differences between ER means at each type of sites were tested using Fisher's Least Significant Difference (LSD, P < 0.05) tests. A stepwise multiple regression analysis was used to analyze the multivariate relationships among ER at the sites GW1–3 and soil temperature, soil moisture and other environmental variables.

References

1. IPCC. IPCC Climate Change 2013: The Physical Science Basis. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA (2013).
2. Raich, J. W. & Schlesinger, W. H. The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate. *Tellus B* **44**, 81–99 (1992).
3. Jungkunst, H. F. & Fiedler, S. Latitudinal differentiated water table control of CO₂, CH₄ and N₂O fluxes from hydromorphic soils: feedbacks to climate change. *Glob. Change Biol.* **13**, 2668–2683 (2007).
4. Davidson, E. A. & Janssens, I. A. Temperature sensitivity of soil carbon decomposition and feedbacks to climate change. *Nature* **440**, 165–173 (2006).
5. Oberbauer, S. F. *et al.* Tundra CO₂ fluxes in response to experimental warming across latitudinal and moisture gradients. *Ecol. Monogr.* **77**, 221–238 (2007).
6. Schuur, E. A. *et al.* The effect of permafrost thaw on old carbon release and net carbon exchange from tundra. *Nature* **459**, 556–559 (2009).
7. Semenchuk, P. R., Christiansen, C. T., Grogan, P., Elberling, B. & Cooper, E. J. Long-term experimentally deepened snow decreases growing-season respiration in a low- and high-arctic tundra ecosystem. *J. Geophys. Res.: Biogeosci.* **121**, 1236–1248 (2016).
8. Gregorich, E. G. *et al.* Emissions of CO₂, CH₄ and N₂O from lakeshore soils in an Antarctic dry valleys. *Soil Biol. Biochem.* **38**, 3120–3129 (2006).

9. Barrett, J. E., Virginia, R. A., Parson, A. N. & Wall, D. H. Soil carbon turnover in the McMurdo dry valleys, Antarctica. *Soil Biol. Biochem.* **38**, 3065–3082 (2006).
10. Ball, B. A. & Virginia, R. A. Controls on diel soil CO₂ flux across moisture gradients in a polar desert. *Antarct. Sci.* **27**, 527–534 (2015).
11. Mendonça, E., La Scala, N., Panosso, A. R., Simas, F. N. & Schaefer, C. E. Spatial variability models of CO₂ emissions from soils colonized by grass (*Deschampsia antarctica*) and moss (*Sanionia uncinata*) in Admiralty Bay, King George Island. *Antarct. Sci.* **23**, 27–33 (2011).
12. Cannone, N., Binelli, G., Worland, M. R., Convey, P. & Guglielmin, M. CO₂ fluxes among different vegetation types during the growing season in Marguerite Bay (Antarctic Peninsula). *Geoderma* **189**, 595–605 (2012).
13. Zhu, R. B., Bao, T., Wang, Q., Xu, H. & Liu, Y. S. Summertime CO₂ fluxes and ecosystem respiration from marine animal colony tundra in maritime Antarctica. *Atmos. Environ.* **98**, 190–201 (2014).
14. Dagg, J. & Lafleur, P. Vegetation community, foliar nitrogen, and temperature effects on tundra CO₂ exchange across a soil moisture gradient. *Arct. Antarct. Alp. Res.* **43**, 189–197 (2011).
15. Burkins, M. B., Virginia, R. A. & Wall, D. H. Organic carbon cycling in Taylor Valley, Antarctica: quantifying soil reservoir and soil respiration. *Glob. Change Biol.* **7**, 113–125 (2001).
16. Convey, P. & Smith, R. I. L. Response of terrestrial Antarctic ecosystems to climate change. *Plant Ecol.* **182**, 1–10 (2006).
17. Convey, P. *et al.* Antarctic climate change and the environment. *Antarct. Sci.* **21**, 541–563 (2009).
18. Vera, M. L. Colonization and demographic structure of *Deschampsia antarctica* and *Colobanthus quitensis* along an altitudinal gradient on Livingston Island, South Shetland Islands, Antarctica. *Polar Res.* **30**, 7146 (2011).
19. Zhu, R. B., Ma, D. W. & Xu, H. Summertime N₂O, CH₄ and CO₂ exchanges from a tundra marsh and an upland tundra in maritime Antarctica. *Atmos. Environ.* **83**, 269–281 (2014).
20. Cannone, N., Guglielmin, M., Convey, P., Worland, M. R. & Longo, S. F. Vascular plant changes in extreme environments: effects of multiple drivers. *Climatic change* **134**, 651–665 (2016).
21. Park, J. H. & Day, T. A. Temperature response of CO₂ exchange and dissolved organic carbon release in a maritime Antarctic tundra ecosystem. *Polar Biol.* **30**, 1535–1544 (2007).
22. Carvalho, J. V. S. *et al.* Impact of expected global warming on C mineralization in maritime Antarctic soils: results of laboratory experiments. *Antarct. Sci.* **22**, 485–493 (2010).
23. Huiskes, A. H. L., Boschker, H. T. S., Lud, D. & Moerdijk-Poortvliet, T. C. W. Stable isotope ratios as a tool for assessing changes in carbon and nutrient sources in Antarctic terrestrial ecosystems. *Plant Ecol.* **182**, 79–86 (2006).
24. Vieira, F. C. B. *et al.* *In situ* methane and nitrous oxide fluxes in soil from a transect in Hennequin Point, King George Island, Antarctica. *Chemosphere* **90**, 497–504 (2013).
25. Bao, T. *et al.* Potential effects of ultraviolet radiation reduction on tundra nitrous oxide and methane fluxes in maritime Antarctica. *Sci. Rep.* **8**, 3716 (2018).
26. Liu, Y. *et al.* Changes in interannual climate sensitivities of terrestrial carbon fluxes during the 21st century predicted by CMIP5 Earth System Models. *J. Geophys. Res.: Biogeosci.* **121**, 903–918 (2016).
27. Liu, Y., Piao, S., Lian, X., Ciais, P. & Smith, W. K. Seasonal Responses of Terrestrial Carbon Cycle to Climate Variations in CMIP5 Models: Evaluation and Projection. *J. Climate* **30**, 6481–6503 (2017).
28. Hopkins, D. W. *et al.* Carbon, nitrogen and temperature controls on microbial activity in soils from an Antarctic dry valley. *Soil Biol. Biochem.* **38**, 3130–3140 (2006).
29. Walker, D. A. Hierarchical subdivision of arctic tundra based on vegetation response to climate, parent material, and topography. *Glob. Change Biol.* **6**, 19–34 (2000).
30. Paré, M. C. & Bedard-Haughn, A. Landscape-scale N mineralization and greenhouse gas emissions in Canadian Cryosols. *Geoderma* **189**, 469–479 (2012).
31. Stewart, K. J., Coxson, D. & Siciliano, S. D. Small-scale spatial patterns in N₂-fixation and nutrient availability in an arctic hummock-hollow ecosystem. *Soil Biol. Biochem.* **43**, 133–140 (2011).
32. Fouché, J., Keller, C., Allard, M. & Ambrosi, J. P. Diurnal evolution of the temperature sensitivity of CO₂ efflux in permafrost soils under control and warm conditions. *Sci. Total Environ.* **581**, 161–173 (2017).
33. Shanhan, F. L., Almond, P. C., Clogh, T. J. & Smith, C. M. S. Abiotic processes dominate CO₂ fluxes in Antarctic soils. *Soil Biol. Biochem.* **53**, 99–111 (2012).
34. Xu, M. & Qi, Y. Soil surface CO₂ efflux and its spatial and temporal variations in a young pond erosa pine plantation in northern California. *Glob. Change Biol.* **7**, 667–677 (2001).
35. Wang, B. *et al.* Soil moisture modifies the response of soil respiration to temperature in a desert shrub ecosystem. *Biogeosci.* **11**, 259–268 (2014).
36. Brummell, M. E., Farrell, R. E. & Siciliano, S. D. Greenhouse gas soil production and surface fluxes at a high arctic polar oasis. *Soil Biol. Biochem.* **52**, 1–12 (2012).
37. Zak, D. R., Holmes, W. E. & MacDonald, N. W. Soil temperature, matric potential, and the kinetics of microbial respiration and nitrogen mineralization. *Soil Sci. Society America J.* **63**, 575–584 (1999).
38. Sponseller, R. A. Precipitation pulses and soil CO₂ flux in a Sonoran desert ecosystem. *Glob. Change Biol.* **13**, 426–436 (2007).
39. Davis, R. C. Structure and function of two Antarctic terrestrial moss communities. *Ecol. Monogr.* **51**, 125–143 (1981).
40. Schimel, J. P., Bilbrough, C. & Welker, J. M. Increased snow depth affects microbial activity and nitrogen mineralization in two Arctic tundra communities. *Soil Biol. Biochem.* **36**, 217–227 (2004).
41. Baptist, F., Yoccoz, N. G. & Choler, P. Direct and indirect control by snow cover over decomposition in alpine tundra along a snowmelt gradient. *Plant soil* **328**, 397–410 (2010).
42. Morgner, E., Elberling, B., Strebel, D. & Cooper, E. J. The importance of winter in annual ecosystem respiration in the High Arctic: effects of snow depth in two vegetation types. *Polar Res.* **29**, 58–74 (2010).
43. Bockheim, J. G. & Hall, K. J. Permafrost, active-layer dynamics and periglacial environments of continental Antarctica: Periglacial and Permafrost Research in the Southern Hemisphere. *Soil Sci. Society America J.* **98**, 82–90 (2002).
44. Thomazini, A. *et al.* Geospatial variability of soil CO₂-C exchange in the main terrestrial ecosystems of Keller Peninsula, Maritime Antarctica. *Sci. Total Environ.* **562**, 802–811 (2016).
45. Bao, T., Zhu, R., Bai, B. & Xu, H. Potential methane production rates and its carbon isotopic composition from ormithogenic tundra soils in coastal Antarctic. *Adv. Polar Sci.* **27**, 21–30 (2016).
46. Tuomi, M., Vanhala, P., Karhu, K., Fritze, H. & Liski, J. Heterotrophic soil respiration- comparison of different models describing its temperature dependence. *Ecol. Model.* **211**, 182–190 (2008).
47. Gershenson, A., Bader, N. E. & Cheng, W. Effects of substrate availability on the temperature sensitivity of soil organic matter decomposition. *Glob. Change Biol.* **15**, 176–183 (2009).
48. Tjoelker, M. G., Oleksyn, J. & Reich, P. B. Modelling respiration of vegetation: evidence for a general temperature-dependent Q₁₀. *Glob. Change Biol.* **7**, 223–230 (2001).

Acknowledgements

The current work was supported by the National Natural Science Foundation of China (Grant Nos 41776190 and 41576181). We thank the editors and reviewers for their constructive comments to improve this paper.

Author Contributions

R.B.Z., X.L.L. and T.B. conceptualized and designed the study, X.L.L. and X.C. provided the test instrument, T.B. collected the samples from the Antarctica, T.B. and W.J.Y. analyzed the samples, R.B.Z. and T.B. calculated and evaluated the data statistically. Finally, R.B.Z. and T.B. interpreted the results, designed and wrote the manuscript. All authors reviewed and discussed the manuscript.

Additional Information

Supplementary information accompanies this paper at <https://doi.org/10.1038/s41598-018-30263-6>.

Competing Interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018