



Draft Genome Sequences of Four Streptomycin-Sensitive *Erwinia amylovora* Strains Isolated from Commercial Apple Orchards in Ohio

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ABSTRACT *Erwinia amylovora* is the causative agent of fire blight, a devastating disease of apples and pears worldwide. Here, we report draft genome sequences of four streptomycin-sensitive strains of *E. amylovora* that were isolated from diseased apple trees in Ohio.

Fire blight, which is caused by *Erwinia amylovora*, is among the most devastating bacterial diseases of apples worldwide and occurs annually in Ohio orchards. Antibiotics, especially streptomycin sulfate, are the most effective strategy to control this disease (1). However, widespread use of streptomycin has led to the emergence of streptomycin-resistant (SmR) *E. amylovora* strains in orchards across the United States (2). We sequenced the genomes of four streptomycin-sensitive (SmS) strains of *E. amylovora* that had been isolated from diseased commercial apple trees in Ohio.

Bacterial isolations from symptomatic shoots were conducted using Crosse-Goodman medium and nutrient broth yeast (NBV) agar as described previously (3). *Erwinia amylovora* strains (Table 1) were screened for SmR using a bioassay test (4). Single colonies were restored from 30% glycerol stocks by streaking on NBV medium, and total genomic DNA was extracted using the Nextera DNA Flex microbial colony extraction protocol (5). Extracted DNA was quantified by spectrophotometry and adjusted to 20 ng/ μ l for library preparation. Sequencing libraries were prepared using the Illumina DNA preparation kit, and the libraries were sequenced on the Illumina iSeq 100 platform with 150-bp paired-end sequencing. Default parameters were used for all software unless otherwise specified. Illumina Local Run Manager software was used to convert and trim the resulting sequences. The quality of sequenced reads was assessed with FastQC v0.11.9 (6). SPAdes v3.14.1 was used to *de novo* assemble the *E. amylovora* genomes and determine genome coverage (7). Genomes were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v5.2 (8–10).

Classification of the assembled genomes was conducted by average nucleotide identity (ANI) analysis using the enveomics collection (11) and LINbase with genome sequence as the identification method (12–16). SmR in *E. amylovora* occurs either from the presence of *strA* and *strB* on plasmids pEA29 or pEA34 or through a mutation in codon 43 of *rpsL* (17, 18). The presence of SmR genes was analyzed by mapping strain reads to *E. amylovora* plasmid pEA34 (GenBank accession number [M96392.1](#)) and *rpsL* (GenBank accession number [NC_013961.1](#)) with the programs BWAv0.17 and IGVv2.10.3 and by conducting BLAST searches for these genes against the assembled genomes (19–21). The four *E. amylovora* strains were nearly identical to the reference strain (*E. amylovora* ATCC 49946 [GenBank accession number [FN666575.1](#)]), with ANI

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values ranging from 99.89% to 99.98% (Table 1). LINbase results confirmed *E. amylovora* as the best match for each sequenced genome. All four Ohio strains contained the *E. amylovora* strain Ea88 ubiquitous plasmid pEA29 (GenBank accession number NC_005706.1) but not *strA*, *strB*, or pEA34, indicating an SmS genotype (17, 18).

The genome sequences and genomic analysis workflow for the SmS strains provide a baseline to screen and monitor for SmR in Ohio apple orchards. Further genomic analysis of *E. amylovora* will increase our understanding of the genetic basis for resistance, allowing us to better address the sustainability of streptomycin use for fire blight management.

Data availability. Data were deposited in NCBI GenBank (BioProject accession number PRJNA756955). The partial genomes were also deposited in LINbase. The BioSample accession number, GenBank accession number, and LINbase number for each *E. amylovora* strain are presented in Table 1.

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