

Review

¹³C-NMR Spectral Data of Alkaloids Isolated from *Psychotria* Species (Rubiaceae)

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Abstract: The genus *Psychotria* (Rubiaceae) comprises more than 2000 species, mainly found in tropical and subtropical forests. Several studies have been conducted concerning their chemical compositions, showing that this genus is a potential source of alkaloids. At least 70 indole alkaloids have been identified from this genus so far. This review aimed to compile ¹³C-NMR data of alkaloids isolated from the genus *Psychotria* as well as describe the main spectral features of different skeletons.

Keywords: Rubiaceae; *Psychotria*; ¹³C-NMR spectral data

1. Introduction

In phytochemistry and related areas, structural elucidation techniques play a key role because precise knowledge of the chemistry of plants requires unequivocal structural characterization of its metabolites to obtain information related to the taxonomy of plant groups. Moreover, correct identification of biologically active compounds is important, both to understand their possible mechanisms of action and propose chemical modifications aimed at enhancing their activity.

The characterization of natural products requires, apart from patience and dedication, knowledge about spectroscopic techniques (interpretation of these data) and the biosynthesis of different types of metabolites. Comparison with literature data is another important auxiliary tool that aids the structural characterization of a given compound. In this context, finding a material that provides as much information as possible about the spectral data of metabolites isolated from a genus (such as *Psychotria*) may enable saving time.

The genus *Psychotria* (Rubiaceae) comprises more than 2000 species, which occur mostly in tropical and subtropical regions [1], with many of these species being employed in folk medicine to treat several diseases [2,3]. The biological potential of the chemical constituents of the species of this genus has possibly motivated several studies regarding the chemical composition of such species. Most of these have focused on investigating alkaloid fractions obtained by acid-base extraction, probably owing to the biological importance of this type of metabolite. Such efforts have led to the isolation

and/or identification of various alkaloids, primarily indole-type. Some of them exhibit some biological properties such as analgesic [4,5], antioxidant [6], antiparasitic [7], and cytotoxic [8,9] activities. This review aimed to compile ^{13}C -NMR spectral data of alkaloids isolated from *Psychotria* species as well as to discuss the main spectral features observed for the different types of skeletons.

2. Discussion

2.1. ^{13}C -NMR Chemical Shifts of Monoterpene Indole Alkaloids Isolated from *Psychotria* Species

Monoterpene indole alkaloids (MIAs) comprise a wide group of secondary metabolites, found mainly in the Apocynaceae, Loganiaceae, and Rubiaceae families [10]. Their biosynthesis involves a reaction between tryptamine (derived from tryptophan) and the iridoid secologanin, catalyzed by strictosidine synthase [11]. This initial step leads to the formation of strictosidine (**1**, Table 1, Figure 1), the key precursor of other MIAs.

Strictosidine (**1**) was isolated from *P. elata* [12] and *P. nuda* (data not reported), and presents an *ortho*-substituted ring system (as do most of the MIAs isolated from this genus), characterized by the presence of four methine carbon signals at δ_{C} 118.8 (CH-9), 120.1 (CH-10), 122.7 (CH-11), and 112.0 (CH-12), and two quaternary carbon signals at δ_{C} 127.9 (C-8) and 137.9 (C-13). The signals of two quaternary carbons at δ_{C} 133.2 (C-2) and 107.7 (C-7), along with a methine carbon at δ_{C} 52.4 (CH-3) and two methylene carbons at δ_{C} 42.9 (CH₂-5) and 21.0 (CH₂-6) complete the tetrahydro- β -carboline system. The secologanin moiety is confirmed by the presence of signals resonating at δ_{C} 170.6 (C-22), δ_{C} 109.9 (C-16), and 156.1 (C-17), indicative of an α,β -unsaturated carboxyl group, a terminal vinyl at δ_{C} 135.7 (CH-19) and 119.5 (CH₂-18), besides signals at δ_{C} 97.5 (CH-21), 45.6 (CH-20), 35.9 (CH₂-14), and 32.5 (CH-15). The anomeric carbon signals of the glucose unit are observed at δ_{C} 100.3 (CH-1'), with four mono-oxygenated methines in the interval from δ_{C} 78.6 to 71.7 and one oxygenated methylene at δ_{C} 62.9 [13].

Strictosidine (**1**) may function as a precursor of other biosynthetic pathways, leading to different skeletons and consequently changes in spectral properties. Carbonylation at C-5 (δ_{C} = 176.5 ppm), as observed for 5 α -carboxystrictosidine (**4**), for example, promotes a chemical shift displacement of CH₂-6 ($\Delta\delta$ = 4.2 ppm, β effect) when compared with **1**, as can be seen in Table 2. A similar pattern was observed for methylation of N-4 on correantoside (**7**) isolated from *P. correa* [14], where $\Delta\delta$ variations (β effect) of 5.4 and 3.5 ppm are observed for CH-3 and CH₂-5, respectively. For 10-hydroxycorreantoside (**8**), it is possible to observe the electronic influence of a hydroxyl by the inductive effect at the *ipso* carbon (C-10) and an increase in the electron densities at the *ortho*(CH-9 and CH-11) and *para* (C-13) positions by the mesomeric effect. On the basis of this mesomeric effect, the signals corresponding to carbon atoms at the *ortho*, CH-9 (δ_{C} 118.8 (**1**) and 104.4 (**8**), $\Delta\delta_{\text{C}}$ = -14.8 ppm) and CH-11 (δ_{C} 122.7 (**1**) and 114.2 (**8**), $\Delta\delta_{\text{C}}$ = -8.5 ppm) and *para* positions, C-13 (δ_{C} 137.9 (**1**) and 131.4 (**8**), $\Delta\delta_{\text{C}}$ = -6.5 ppm), are displaced upfield.

Other metabolic pathways of this class of alkaloid revealed cyclization reactions involving N-1 (compounds **5** and **6**) or N-4 (compounds **7**–**14**) with C-22, or N-1 with C-18 and N-4 with C-22, as particularly observed for stachyoside (**30**) isolated from *P. stachyoides* [15] (Figure 2). Strictosamide (**5**), isolated from four different species, [16–19] is an example of lactam formation between N-4 and C-22. By examining Table 2, it is possible to notice, apart from the absence of a methoxyl group (carbomethoxy function) signal at δ_{C} 52.4, a slight difference in the chemical shift of C-22 (δ_{C} 167.1 ppm), when compared with compound **1** (δ_{C} 170.6 ppm), as well as a $\Delta\delta_{\text{C}}$ variation of 6.9 ppm for C-17. In contrast, correantoside (**7**) exemplified the first possibility involving cyclization between N-1 and C-22. It is possible, in this case, to observe the variation in the chemical shifts of the *ortho*CH-12 ($\Delta\delta$ = 4.0 ppm) and *para* CH-10 ($\Delta\delta$ = 4.1 ppm) atoms, promoted by the inductive and mesomeric effects of the carboxyl group at C-22. These effects were also observed for compounds **8** ($\Delta\delta_{\text{C}}$ = 4.8 (CH-12) ppm), **13** ($\Delta\delta_{\text{C}}$ = 4.4 (CH-12) and 4.1 (CH-10) ppm), **14** ($\Delta\delta_{\text{C}}$ = 4.4 (CH-12) and 4.5 (CH-10) ppm), **18** ($\Delta\delta_{\text{C}}$ = 7.8 (CH-12) and 6.2 (CH-10) ppm), and **19** ($\Delta\delta_{\text{C}}$ = 7.3 (CH-12) and

5.4 (CH-10) ppm), showing that the downfield displacements of the CH-12 and CH-10 signals may be used to suggest that *N*-1 is attached to C-22.

There are some examples of alkaloids isolated from this genus, whose biosynthesis involves hydrolysis of a glycoside moiety such as (*E/Z*)-vallesiachotamines, **23** and **24**, isolated from *P. bahiensis* [17], and 10-hydroxy-*iso*-depeaninol (**27**) and *N*-oxide-10-hydroxyantirrhine (**29**) isolated from *P. prunifolia* [20]. These types of skeletons may be suggested by analysis of the region of the ¹³C spectrum that is typical of sugars, revealing the absence of the typical signal of the anomeric carbon around δ_C 100.0, apart from additional signals of the oxy-carbons characteristic of this unit.

Kerber et al. reported the isolation of a new MIA from *P. brachyceras* leaves [21], named brachycerine (**33**), which showed a new alkaloid skeleton. Its biosynthesis involved the coupling of tryptamine to a 1-*epi*-loganin derivative. Psychollatine (**34**), a new MIA from *P. umbellata* [22], presented a terpenoid derivative from geniposide. Both alkaloids as well as compounds **21**, **22**, and **35** revealed an important characteristic in their ¹³C spectra: the absence of typical signals of a terminal vinyl group ($\sim\delta_C$ 119 ppm). In contrast, bahienosides A (**38**) and B (**37**), isolated from *P. bahiensis* [17], showed duplicate terminal vinyl group signals relative to two secologanin moieties. Figure 2 shows typical carbon assignments, which may indicate some different structural possibilities in comparison with those values observed for strictosidine (**1**).

Table 1. Monoterpene indole alkaloids from *Psychotria* species.

Compounds	Species	References	¹³ C-NMR Data
Strictosidine (1)	<i>P. elata</i> <i>P. acuminata</i>	[12]	[13]
Strictosidinic acid (2)	<i>P. barbiflora</i> <i>P. myriantha</i>	[1,23–25]	[25]
Palicoside (3)	<i>P. racemosa</i>	[12]	[26]
5 α -Carboxystrictosidine (4)	<i>P. acuminata</i> <i>P. bahiensis</i> <i>P. bahiensis</i>	[17,23]	[27]
Strictosamide (5)	<i>P. nuda</i> <i>P. prunifolia</i> <i>P. suterella</i>	[16–19]	[18]
<i>N</i> , β -D-Glucopyranosilvincosamide (6)	<i>P. leiocarpa</i>	[28]	[28]
Correantose (7)	<i>P. correae</i>	[14]	[14]
10-Hydroxycorreantose (8)	<i>P. correae</i>	[14]	[14]
Correantine B (9)	<i>P. correae</i>	[14]	[14]
20- <i>epi</i> -Correantine B (10)	<i>P. correae</i>	[14]	[14]
Correantine A (11)	<i>P. correae</i>	[14]	[14]
Correantine C (12)	<i>P. correae</i>	[14]	[14]
<i>N</i> -Desmethyl-correantose (13)	<i>P. stachyoides</i>	[29]	[29]
<i>Nor</i> -Methyl-23-oxo-correantose (14)	<i>P. stachyoides</i>	[15]	[15]
14-Oxoprunifoleine (15)	<i>P. prunifolia</i>	[18,20]	[18]
17-Vinyl-19-oxa-2-azonia-12-azapentacyclo [14.3.1.0 ^{2,14} .0 ^{5,13} .0 ^{6,11}]icosa-2(14),3,5(13),6(11),7,9-hexaene (16)	<i>P. prunifolia</i>	[18]	[18]
Naucleatine (17)	<i>P. suterella</i>	[19]	[30]
Correantosine E (18)	<i>P. stachyoides</i>	[31]	[31]
Correantosine F (19)	<i>P. stachyoides</i>	[31]	[31]
Lagamboside (20)	<i>P. acuminata</i>	[23]	[23]
<i>N</i> ⁴ -[1-((<i>R</i>)-2-Hydroxypropyl)]-psychollatine (21)	<i>P. umbellata</i>	[32]	[32]
<i>N</i> ⁴ -[1-((<i>S</i>)-2-Hydroxypropyl)]-psychollatine (22)	<i>P. umbellata</i>	[32]	[32]
(<i>E/Z</i>)-Vallesiachotamine (23 + 24)	<i>P. bahiensis</i> <i>P. laciniata</i>	[17,33]	[34]
Isodolichantose (25)	<i>P. correae</i>	[14]	[14]
Angustine (26)	<i>P. bahiensis</i> <i>P. laciniata</i>	[17,33]	[35]
10-Hydroxy- <i>iso</i> -depeaninol (27)	<i>P. prunifolia</i>	[20]	[20]
10-Hydroxy-antirrhine (28)	<i>P. prunifolia</i>	[20]	[20]
<i>N</i> -Oxide-10-hydroxyantirrhine (29)	<i>P. prunifolia</i>	[20]	[20]
Stachyoside (30)	<i>P. stachyoides</i>	[15]	[15]
Lyaloside (31)	<i>P. laciniata</i> <i>P. suterella</i>	[19,36]	[37]
Myrianthosine (32)	<i>P. myriantha</i>	[25]	[25]

Table 1. Cont.

Compounds	Species	References	¹³ C-NMR Data
Brachycerine (33)	<i>P. brachyceras</i>	[21]	[21]
Psychollatine (34)	<i>P. umbellata</i>	[5,22,38]	[22]
3,4-Dehydro-18,19-β-epoxy-psychollatine (35)	<i>P. umbellata</i>	[32]	[32]
Desoxycordifoline (36)	<i>P. acuminata</i>	[23]	[39]
Bahienoside B (37)	<i>P. acuminata</i>	[17,23]	[17]
Bahienoside A (38)	<i>P. bahiensis</i>	[17]	[17]

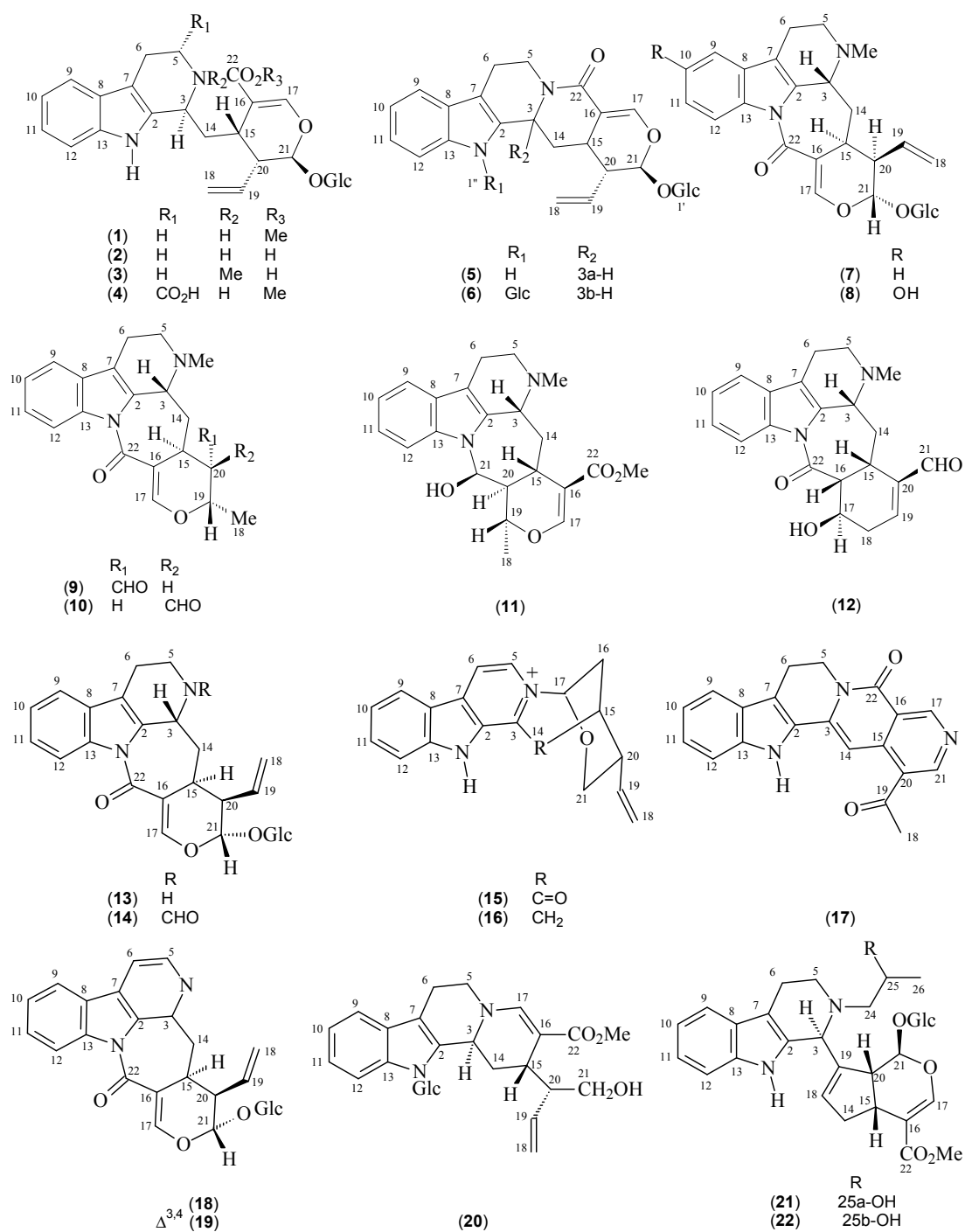


Figure 1. Cont.

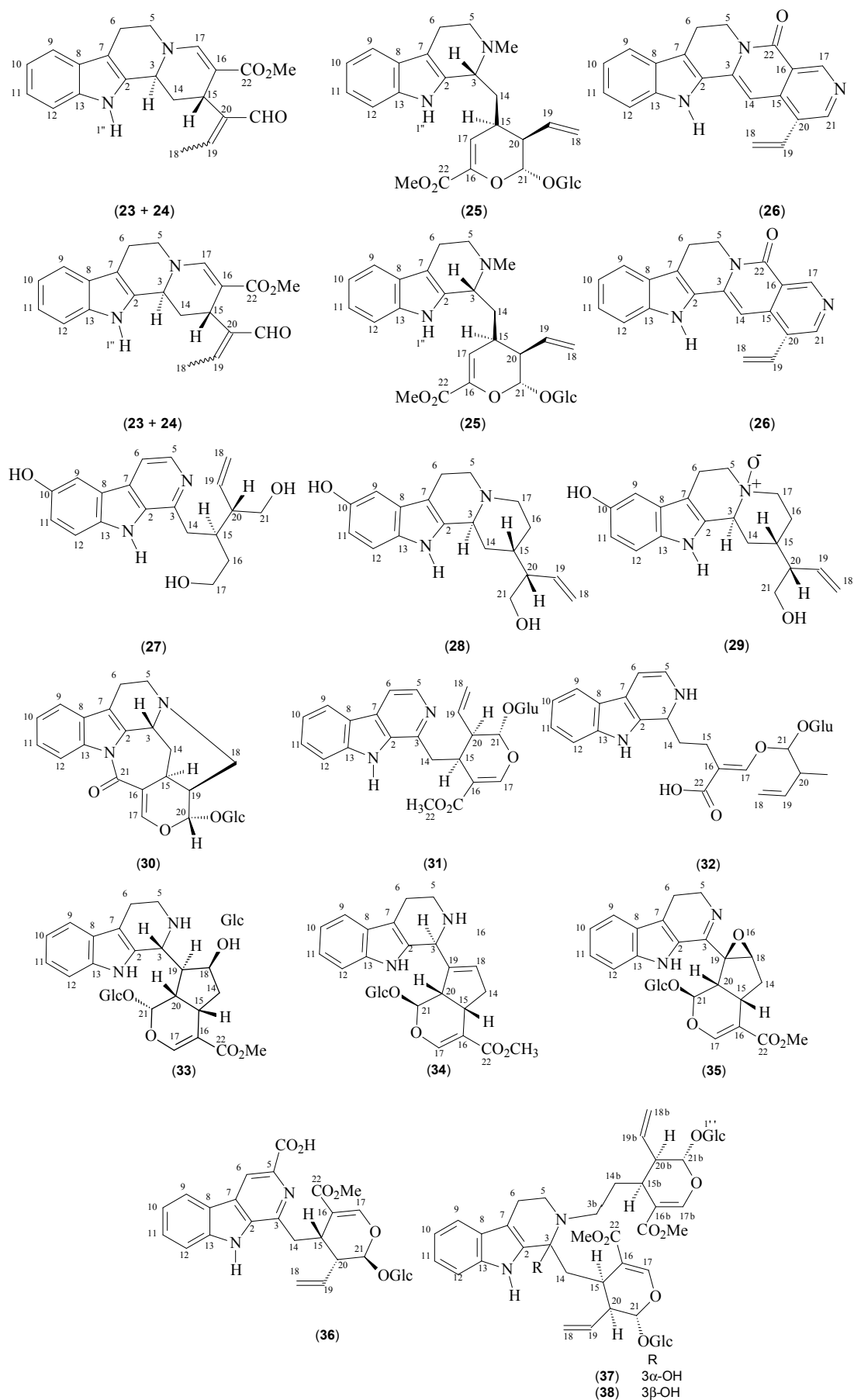


Figure 1. Structures of monoterpene indole alkaloids from *Psychotria* species.

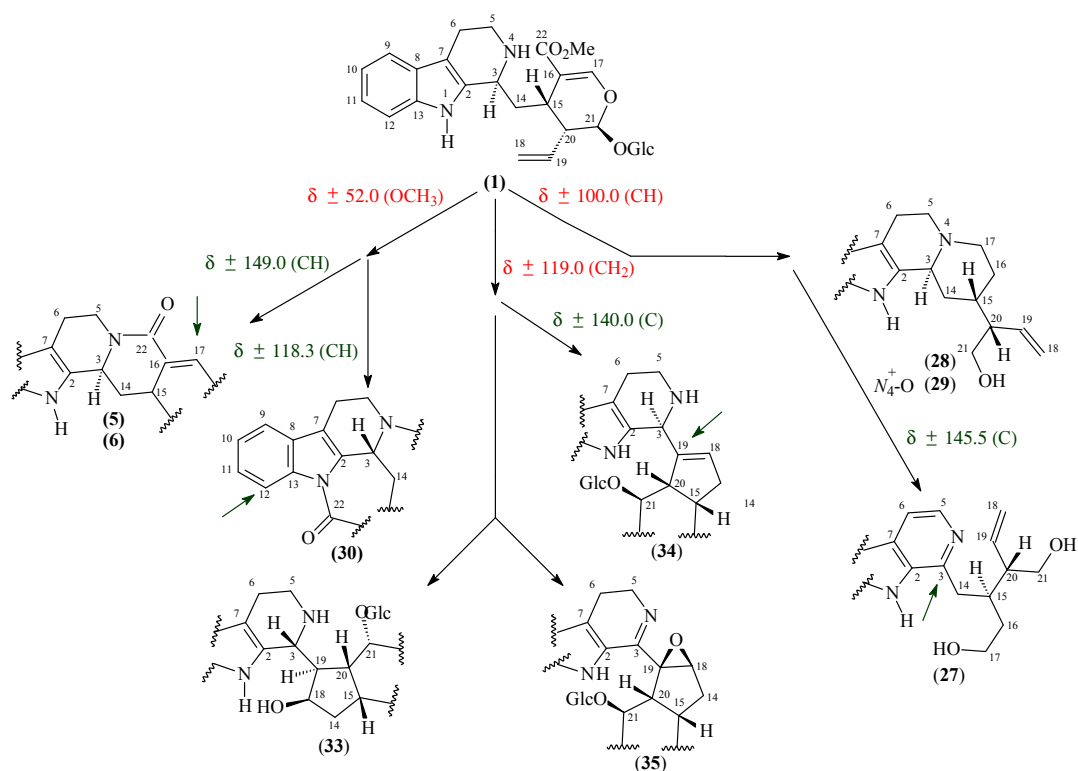


Figure 2. Structural signaling based on specific signals compared with values for strictosidine (1): absence of a given signal in red and presence in green.

Table 2. ¹³C-NMR data of MIAs from *Psychotria* species.

Carbons	Compounds/ δ_C (ppm)									
	1 ^I	2 ^{III}	3 ^{III}	4 ^I	5 ^I	6 ^I	7 ^I	8 ^I	9 ^{II}	10 ^{II}
C										
2	133.2	132.3	134.7	133.2	134.8	136.1	134.3	133.8	132.9	133.0
7	107.7	106.0	105.2	109.0	110.3	111.5	115.7	115.3	114.8	114.8
8	127.9	126.1	126.6	128.0	128.7	129.5	130.4	131.3	129.1	129.1
13	137.9	135.8	135.8	138.4	137.8	137.7	137.3	131.4	136.0	136.0
22	170.6	170.0	168.4	170.9	167.1	166.3	168.2	167.8	166.2	166.2
16	109.9	113.4	112.5	109.9	109.2	109.1	112.2	112.0	108.6	109.6
CH										
3	52.4	49.6	56.1	53.2	55.1	54.5	57.8	58.2	56.4	56.7
5	-	-	-	60.1	-	-	-	-	-	-
9	118.8	117.8	117.4	118.8	118.7	119.3	119.2	104.4	118.1	118.0
10	120.1	118.7	118.1	120.1	120.2	121.3	124.2	155.1	123.2	123.2
11	122.7	121.2	120.3	122.6	122.6	122.9	125.5	114.2	124.6	124.6
12	112.0	111.5	110.8	112.1	112.3	114.8	116.0	116.8	115.4	115.2
15	32.5	31.8	30.6	32.4	24.9	27.9	35.7	35.6	29.7	29.2
17	156.1	150.0	151.8	156.1	149.2	149.2	155.7	155.5	158.0	156.4
19	135.7	135.6	135.6	135.2	134.4	133.4	135.1	135.0	70.2	69.4
20	45.6	44.3	44.0	45.7	44.7	44.1	45.4	45.4	51.8	53.9
21	97.5	95.1	95.9	97.6	98.1	97.5	97.4	97.3	-	-
CH₂										
5	42.9	40.0	45.2	60.1	44.8	41.6	46.4	46.7	45.5	45.5
6	21.0	19.2	15.9	25.2	22.1	22.3	18.8	18.8	17.6	17.7
14	35.9	33.7	35.3	35.6	27.3	35.6	34.4	34.1	39.1	35.3
18	119.5	117.8	117.8	119.6	120.6	120.7	119.2	119.3	-	-

Table 2. Cont.

Glucose										
1'	-	-	100.7	100.8	-	-	-	100.1	100.1	87.6
2'	-	-	74.9	74.9	-	-	-	74.8	74.7	72.4
3'	-	-	78.7	78.2	-	-	-	78.0	77.9	79.4
4'	-	-	71.8	71.7	-	-	-	71.7	71.7	71.8
5'	-	-	78.2	78.7	-	-	-	78.0	78.6	81.2
6'	-	-	63.1	63.0	-	-	-	62.9	62.9	63.0
CHO	-	-	-	163.9	-	-	-	-	-	-
CO ₂ Me	51.1	-	-	-	-	-	-	-	-	51.2
Carbons	Compounds/ δ_C (ppm)									
	21 ^I	22 ^I	23 ^{III}	24 ^{III}	25 ^I	26 ^{III}	27 ^I	28 ^I	29 ^I	
C										
2	134.0	133.4	133.1	133.6	134.0	126.8	136.9	130.5	131.0	
3	-	-	-	-	-	136.9	145.5	-	-	
7	108.4	108.4	106.6	107.4	106.5	114.8	130.6	106.0	105.7	
8	138.6	138.1	126.2	127.0	128.1	125.5	123.1	128.6	128.3	
10	-	-	-	-	-	-	152.6	151.8	152.0	
13	128.4	128.0	136.1	136.8	137.8	138.5	137.5	133.1	133.6	
15	-	-	-	-	-	139.0	-	-	-	
16	112.2	112.0	93.2	93.4	112.0	119.8	-	-	-	
19	141.0	142.1	-	-	-	-	-	-	-	
20	-	-	146.1	143.9	-	127.8	-	-	-	
22	169.7	169.0	166.9	167.6	169.8	161.1	-	-	-	
CH										
3	61.7	59.3	48.6	47.9	58.8	-	-	57.0	71.6	
5	-	-	-	-	-	-	135.7	-	-	
6	-	-	-	-	-	-	114.6	-	-	
9	118.6	118.0	117.4	118.4	118.7	119.9	106.6	103.2	103.3	
10	119.5	120.0	118.3	119.2	119.9	119.9	-	-	-	
11	121.9	122.0	120.7	121.6	122.3	124.6	120.4	112.9	113.2	
12	112.0	112.0	110.8	111.8	111.8	112.0	113.7	112.8	113.0	
14	-	-	-	-	-	93.8	-	-	-	
15	33.0	35.3	27.4	30.5	30.5	-	36.4	31.1	30.6	
17	153.3	153.0	147.2	148.5	154.0	149.7	-	-	-	
18	132.6	131.0	-	-	-	-	-	-	-	
19	-	-	152.0	146.3	135.8	130.2	138.1	138.7	138.2	
20	49.0	48.4	-	-	45.5	-	51.0	50.8	52.3	
21	95.6	97.0	-	-	97.8	147.7	-	-	-	
25	65.1	66.3	-	-	-	-	-	-	-	
CH₂										
5	49.0	49.6	49.8	50.7	47.9	40.4	-	52.4	69.0	
6	21.4	19.7	21.3	22.2	17.9	19.2	-	18.1	20.6	
14	39.4	39.5	32.9	32.9	34.5	-	37.0	31.6	28.5	
17	-	-	-	-	-	-	61.4	48.0	59.1	
18	-	-	-	-	119.8	119.8	118.7	118.5	118.5	
21	-	-	-	-	-	-	64.4	64.0	63.8	
24	62.4	61.6	-	-	-	-	-	-	-	
CH₃										
18	-	-	14.3	13.8	-	-	-	-	-	
26	20.7	21.2	-	-	-	-	-	-	-	
MeN-Glucose										
1'	100.1	100.1	-	-	100.5	-	-	-	-	
2'	74.6	74.8	-	-	74.7	-	-	-	-	

Table 2. Cont.

3'	78.0	78.0	-	-	78.6	-	-	-	-
4'	78.2	71.6	-	-	71.6	-	-	-	-
5'	76.2	78.5	-	-	78.0	-	-	-	-
6'	62.7	62.5	-	-	62.9	-	-	-	-
CHO	-	-	195.5	191.5	-	-	-	-	-
CO ₂ Me	51.6	51.7	49.7	50.8	51.9	-	-	-	-
	Compounds/δ_C (ppm)								
Carbons	30^I	31	32^{III}	33^I	34^I	35^I	36^I	37^I	38^I
C									
2	137.1	140.3	134.8	130.7	131.1	128.8	135.6	135.0	138.0
3	-	143.8	-	-	-	158.9	142.9	-	-
5	-	-	-	-	-	-	135.6	-	-
7	118.4	121.0	121.0	108.3	107.9	118.8	128.4	107.3	106.6
8	129.2	126.9	121.5	127.7	127.6	139.9	121.7	128.4	128.0
13	139.1	134.6	140.2	112.3	138.1	126.1	141.6	137.8	138.0
15									
19	-	-	-	-	140.0	67.3	-	-	-
16	114.8	109.9	112.0	111.8	112.2	110.2	108.7	112.1	111.5
21	169.0	-	-	-	-	-	-	-	-
22	-	166.6	170.0	169.1	169.1	168.9	171.3	169.7	170.0
22b	-	-	-	-	-	-	-	169.5	169.4
CH									
3	51.7	-	48.5	54.7	53.7	-	-	58.8	59.6
5	-	137.3	137.0	-	-	-	-	-	-
6	-	112.6	118.0	-	-	-	114.2	-	-
9	119.7	121.4	126.6	118.9	119.0	121.2	121.4	120.6	118.7
10	125.2	119.0	118.9	120.2	120.3	121.1	119.9	119.7	120.0
11	126.8	127.6	127.5	123.2	123.6	126.2	128.4	122.0	122.5
12	118.3	111.8	112.5	112.3	112.3	113.6	111.6	112.0	112.0
15	32.9	30.1	-	35.5	37.5	31.7	34.5	31.5	31.6
17	148.7	151.6	151.0	153.5	153.4	153.1	153.2	154.0	154.7
18	-	-	-	74.3	138.5	62.5	-	-	-
19	53.3	134.0	134.5	49.0	-	-	133.8	136.2	136.1
20	95.5	42.9	45.5	41.9	49.0	43.8	44.4	45.5	45.4
21	-	95.9	95.4	99.0	99.4	95.2	96.1	98.2	97.9
15b	-	-	-	-	-	-	-	30.3	30.5
17b	-	-	-	-	-	-	-	153.2	153.5
19b	-	-	-	-	-	-	-	135.7	135.5
20b	-	-	-	-	-	-	-	44.8	44.8
21b	-	-	-	-	-	-	-	98.5	98.3
CH₂									
5	48.0	-	-	41.8	42.1	48.2	-	44.8	44.8
6	21.2	-	-	24.4	20.5	20.1	-	17.6	17.4
14	43.7	32.1	45.6	43.5	40.5	34.7	34.0	36.9	36.7
15	-	-	30.0	-	-	-	-	-	-
17	-	-	-	-	-	-	-	-	-
18	72.4	118.6	118.9	-	-	-	117.6	119.8	119.8
3b	-	-	-	-	-	-	-	52.0	51.9
14b	-	-	-	-	-	-	-	28.0	27.4
18b	-	-	-	-	-	-	-	120.1	120.1
CH₃									
Me	-	-	10.4	-	-	-	-	-	-
Glucose									
1'	100.3	98.79	98.6	100.6	101.5	99.4	99.0	100.4	100.4
2'	74.9	73.1	73.0	74.0	71.0	74.7	73.2	74.6	74.8 ^c
3'	78.5	77.3	69.9	71.1	78.6	78.1	76.6	78.0	78.6 ^a
4'	71.9	71.10	77.3	78.3	74.6	72.1	70.4	71.6	71.7 ^d
5'	78.5	77.8	76.8	77.7	77.6	78.8	76.6	78.4	78.2 ^b
6'	63.0	61.2	61.0	62.1	61.8	63.3	61.8	62.9	62.9 ^e

Table 2. Cont.

1''	-	-	-	-	-	-	-	100.3	100.4
2''	-	-	-	-	-	-	-	74.8	74.6 ^c
3''	-	-	-	-	-	-	-	78.1	78.4 ^a
4''	-	-	-	-	-	-	-	71.6	71.6 ^d
5''	-	-	-	-	-	-	-	78.3	78.0 ^b
6''	-	-	-	-	-	-	-	62.8	62.8 ^e
CO ₂ Me	-	50.7	-	51.8	51.9	51.9	50.6	52.1	52.1

^I CD₃OD, ^{II} CDCl₃, ^{III} DMSO-*d*₆, letters (a–e) indicate signals that may be interchanged.

2.2. ¹³C-NMR Chemical Shifts of Pyrrolidinoindoline Alkaloids Isolated from *Psychotria* Species

Some studies have also reported that the isolation of pyrrolidinoindoline alkaloids seems to be specific to the *Psychotria* species (Table 3). As shown in Figure 3, their chemical structures present the condensation of some *N*-methyltryptamine units with different connection patterns, mainly involving C-3a-C3'a, C-3'a-C-7, and *N*-C-3'a bonds or containing *N*-methyltryptamine units linked to a bisquinoline part. The compound (+)-chimonanthine (40) was isolated from several *Psychotria* species [40–42] and is an example of a dimer that presents a C-3a-C-3'a-type linkage between its two units. Its ¹³C-NMR spectrum exhibited 11 carbon-signal equivalents for both units. The signals at δ_C 52.4 (CH₂-2) and 84.6 (C-8a) are typical of carbons bearing one and two nitrogen atoms, respectively. The signals at δ_C 33.2 and 63.6 were attributed to C-3 and C-3a, respectively, whereas the signal at δ_C 33.8 is consistent with a methyl carbon attached to a nitrogen atom. The *ortho*-substituted aromatic rings are characterized by signals at δ_C 124.9 (CH-4/CH-4'), 128.3 (C-4a/C-4a'), 122.3 (CH-5), 119.8 (CH-5'), 129.9 (CH-6/CH-6'), 110.5 (CH-7/CH-7'), and 150.5 (C-7a/C-7a') [40] (Table 4).

Since some compounds with more than two units present a chimonanthine portion in their structures, the monitoring of C-3a and C-7 (main binding sites) and their neighborhood may be a good alternative, in order to determine the positions of the other monomeric units. Hodgkinsine (52) occurs frequently in the genus [41–46] and presents a third unit with a C-3''a-C-7' linkage. In this case, besides replacement of a methine aromatic carbon by a quaternary carbon (C-7'), observing the up field displacements of C-6' and C-4' ($\Delta\delta$ around 3.0 ppm) is possible probably because of the presence of a group that increases the electron densities of these positions (comparison with compound 40). Takayama et al. (2004), however, reported the isolation of psychopentamine (60) from *P. rostrata*², which showed a new type of linkage between C-3'''a and C-5'' [2].

The chemical study of *P. calocarpa* leaves [43] led to the isolation of a new alkaloid named psychotriazine (45), which presents a tryptamine unit linked to a pyrroloindole unit by an *N*-C3'a linkage. This type of junction was also observed for psychohenin (46) and compound 48 isolated from *P. henryi* [47,48] and may be indicated by the presence of a quaternary carbon (C-3'a) that resonates at δ_C 79.4, 77.8, and 76.7 ppm, in the three compounds, respectively. In contrast, psychotrimine (53), isolated from *P. rostrate* [2] shows, besides the *N*-C-3'a bond, an *N*-C-7' linkage indicated by the signal of a quaternary aromatic carbon C-7' at δ_C 121.5 ppm.

Alkaloids with more complex structures, containing from four to seven units, such as quadrigemines A–C (55–57), psychotridine (61), oleidine (64), and caledonine (65), have also been isolated from this genus; however, the structural elucidation of these compounds becomes more difficult as the number of units increases. Probably owing to this, some studies did not provide detailed attributions of their carbon signals. In such cases, mass spectrometry plays an important role in establishing the number of units present in their structures as well as the pattern of the junctions.

Table 3. Pyrrolidinoindoline alkaloids from *Psychotria* species.

Compounds	Species	References	¹³ C-NMR Data
Meso-chimonanthine (39)	<i>P. forsteriana</i> <i>P. muscosa</i> <i>P. colorata</i>	[41,49,50]	[50]
(+)-Chimonanthine (40)	<i>P. muscosa</i> <i>P. rostrata</i> <i>P. hoffmannseggiana</i>	[40–42]	[40]
Iso-calycanthine (41)	<i>P. forsteriana</i>	[50]	[50]
Calycanthine (42)	<i>P. forsteriana</i>	[50]	[50]
(8-8a),(8'-8'a)-tetrahydroisocalycanthine 3a(R), 3'a(R) (43)	<i>P. colorata</i>	[42]	[42]
N _b -desmethyl-meso-chimonanthine (44)	<i>P. lyciiflora</i>	[49]	[49]
Psychotriasine (45)	<i>P. calocarpa</i>	[43]	[43]
Psychohenin (46)	<i>P. henryi</i>	[47]	[47]
Compound (47)	<i>P. henryi</i>	[48]	[48]
Compound (48)	<i>P. henryi</i>	[48]	[48]
Glomerulatine A (49)	<i>P. glumerulata</i>	[51]	[51]
Glomerulatine B (50)	<i>P. glumerulata</i>	[51]	[51]
Glomerulatine C (51)	<i>P. glumerulata</i> <i>P. colorata</i> <i>P. oleoides</i> <i>P. lyciiflora</i>	[51]	[51]
Hodgkinsine (52)	<i>P. muscosa</i> <i>P. beccarioides</i> <i>P. rostrata</i>	[41–46]	[42]
Psychotrimine (53)	<i>P. rostrata</i>	[2]	[2]
Psychotripine (54)	<i>P. pilifera</i>	[52]	[52]
Quadrigemine A (55)	<i>P. forsteriana</i> <i>P. forsteriana</i>	[53]	[53]
Quadrigemine B (56)	<i>P. colorata</i> <i>P. rostrata</i>	[41,53]	[53]
Quadrigemine C (57)	<i>P. colorata</i> <i>P. oleoides</i>	[41–43,45,46, 50,54]	[45]
Quadrigemine I (58)	<i>P. oleoides</i>	[49]	[49]
Psycholeine (59)	<i>P. oleoides</i>	[46,54]	[46]
Psychopentamine (60)	<i>P. rostrata</i> <i>P. forsteriana</i> <i>P. oleoides</i>	[2]	[2]
Psychotridine (61)	<i>P. colorata</i> <i>P. beccarioides</i>	[41,44,45,53]	[45]
Isopsychotridine C (62)	<i>P. forsteriana</i>	[53,55]	[55]
Isopsychotridine B (63)	<i>P. oleoides</i>	[49,50]	[45]
Oleoidine (64)	<i>P. oleoides</i>	[49]	[49]
Caledonine (65)	<i>P. oleoides</i>	[49]	[49]

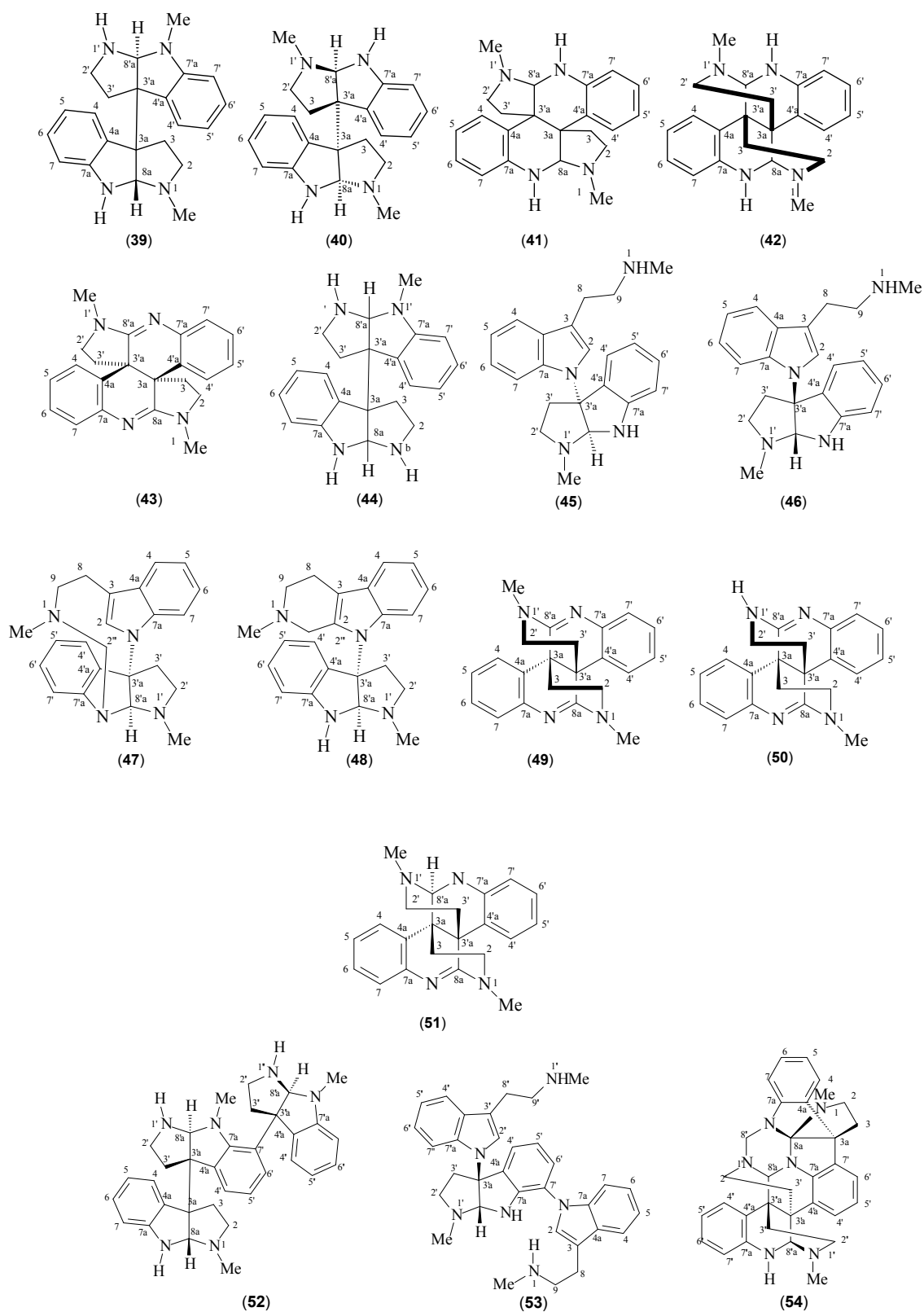


Figure 3. Cont.

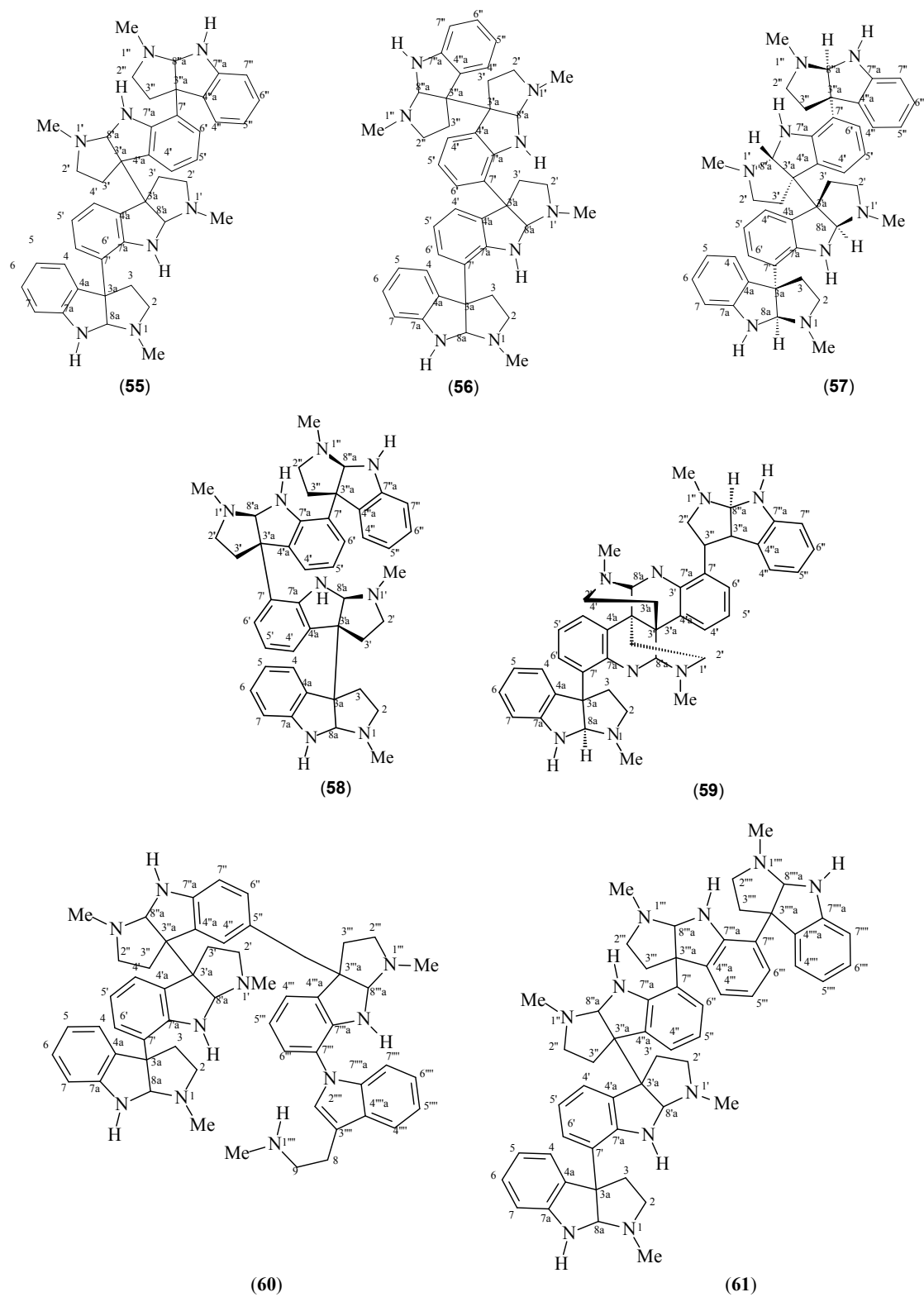


Figure 3. Cont.

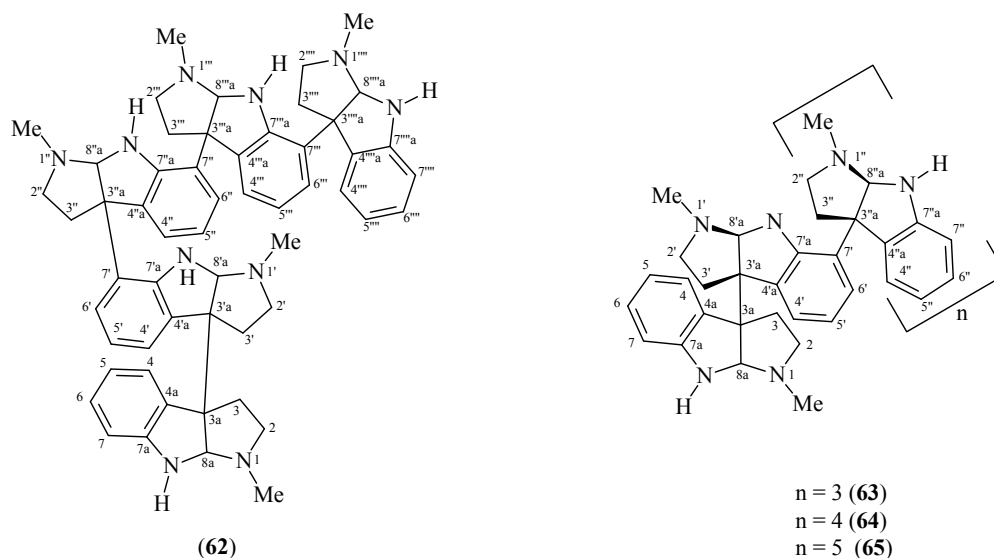


Figure 3. Structures of pyrrolidinoindoline alkaloids from *Psychotria* species.

Table 4. ^{13}C -NMR data of pyrrolidinoindoline alkaloids from *Psychotria* species.

Carbons	Compounds/ δ_{C} (ppm)								
	39 II	40 II	41 ns	42 II	43 II	44 II	45 I	46 I	47 II
C									
3	-	-	-	-	-	-	112.7	110.0	112.3
3a	64.7	63.6	37.8	36.8	48.9	62.8	-	-	-
4a	133.7	128.3	127.0	125.9	125.6	132.2	130.4	130.5	130.0
7a	152.5	150.5	145.3	146.2	145.8	151.7	137.7	138.0	135.0
8a	-	-	-	-	165.0	-	-	-	-
3'a	64.7	63.6	37.8	36.8	48.9	63.9	79.4	77.8	75.3
4'a	133.7	128.3	127.0	125.9	125.6	130.0	131.3	131.3	128.9
7'a	152.5	150.5	145.3	146.2	145.8	150.3	152.5	152.7	152.4
8'a	-	-	-	-	165.0	-	-	-	-
CH									
2	-	-	-	-	-	-	125.0	126.1	123.4
4	125.2	124.9	118.3	117.1	123.0	123.9	124.7	119.5	119.0
5	119.2	122.3	122.2 ^b	122.1	118.5	119.9	119.3	118.8 ^e	119.1 ^f
6	128.9	129.9	127.7	127.3	128.2	128.2	130.7	123.0	121.4
7	109.5	110.5	112.9	112.8	123.9	109.1	120.1	117.4	112.8
8a	83.9	84.6	71.7	71.82	-	79.3	-	-	-
4'	125.2	124.9	118.3	117.1	123.0	124.4	112.2	124.9	126.4
5'	119.2	119.8	125.2	125.2	121.9	117.9	122.4	119.8	118.7
6'	128.9	129.9	127.7	127.3	128.2	128.4	119.6	130.9	130.2
7'	109.5	110.5	112.9	112.8	123.9	108.2	110.0	110.4	108.8
8'a	83.9	84.6	71.7	71.82	-	82.4	87.0	87.3	86.6
CH₂									
2	53.1	52.4	46.9	47.3	48.5	44.9	-	-	-
3	36.4	33.2	34.9	32.5	29.9	35.3	-	-	-
2'	53.1	52.4	46.9	47.4	48.5	51.8	52.0	52.3	53.6
3'	36.4	33.2	34.9	32.5	29.9	38.1	39.9	40.0	37.7
2''	-	-	-	-	-	-	-	-	69.1
CH₃									
Me-N ¹ -	nd	33.8	46.9	43.4	31.1	-	36.3	33.9	40.6
MeN ^{1'} -	nd	33.8	46.9	43.4	31.1	35.12	35.7	36.4	37.1

Table 4. Cont.

Carbons	Compounds/ δ_C (ppm)								
	48 II	49 III	50 III	51 III	52 II	53 II	54 I + II	55 II,*	56 II,*
C									
2	129.6	-	-	-	-	-	-	-	-
3	109.4	-	-	-	-	114.9	-	-	-
3a	-	49.1	48.6	49.2	62.8	-	69.1	60.9 ^c	60.1 ^c
4a	128.0	126.4	126.1 ^a	129.5	131.7	128.3	133.8	132.3 ^d	133.2 ^e
7a	137.4	177.3	147.1	148.6	150.8	136.1	152.2	150.9 ^h	150.6 ^h
8a	-	165.1	164.7	166.5	-	-	106.9	-	-
3'a	76.7	49.1	48.6	45.3	63.0	76.7	37.0	63.2 ^j	63.9 ⁱ
4'a	130.5	126.4	125.4 ^a	122.3	132.3	132.0	122.0	132.4 ^d	132.9 ^e
7'	-	-	-	-	-	121.5	130.9	108.9 ^g	-
8'a	-	165.1	164.7	-	-	-	-	-	-
3''	-	-	-	-	-	112.5	-	-	-
3''a	-	-	-	-	60.0	-	38.4	62.9 ^j	63.3 ⁱ
8''	-	-	-	-	-	25.7 ^b	68.0	-	-
9''	-	-	-	-	-	52.0	-	-	-
4''a	-	-	-	-	131.7	129.8	122.3	132.6 ^d	-
7''a	-	-	-	-	151.1	136.1	144.4	-	-
3'''a	-	-	-	-	-	-	-	60.8 ^c	60.9 ^c
CH									
2	-	-	-	-	-	126.0	-	-	-
4	117.9	123.7	120.9	123.0	126.4	119.4 ^a	122.9	-	125.9 ^d
5	119.2	122.3	122.2 ^b	122.1	118.5	119.9	119.3	118.8 ^e	119.1 ^f
6	121.3	128.9	129.0 ^c	128.8	127.9	122.4	128.2	127.9 ^f	128.0 ^g
7	112.1	125.0	125.2 ^d	125.2	109.0	111.2	107.7	109.0 ^g	108.9
8a	-	-	-	-	86.4	-	-	86.9 ⁱ	85.9 ⁱ
4'	124.5	123.7	124.0 ^d	117.5	121.9	123.7	123.7	122.5	125.1 ^d
5'	119.0	122.3	122.6 ^b	124.9	116.8	119.3 ^a	122.0	116.3 ^k	118.3 ^f
6'	129.6	128.9	128.8 ^c	127.1	126.0	127.3	121.1	125.4	127.8 ^g
7'	108.9	125.0	124.4 ^d	114.4	-	-	-	-	-
8'a	86.5	-	-	76.5	81.7	86.1	69.7	86.1 ⁱ	83.3 ^j
2''	-	-	-	-	-	124.3	-	-	-
4''	-	-	-	-	124.2	119.3 ^a	125.4	-	-
5''	-	-	-	-	117.5	119.3 ^a	117.8	118.7 ^e	117.2 ^f
6''	-	-	-	-	127.4	121.7	127.6	-	-
7''	-	-	-	-	108.1	112.2	112.5	-	-
8''a	-	-	-	-	82.3	-	69.4	-	82.3 ^j
8'''a	-	-	-	-	-	-	-	-	87.1 ⁱ
5'''	-	-	-	-	-	-	-	116.2 ^k	116.8 ^f
6'''	-	-	-	-	-	-	-	126.4 ^f	-
CH₂									
2	-	48.2	48.1	48.5	51.7	-	54.9	52.6 ^a	52.3 ^a
3	-	30.3	30.3	31.7	37.6	-	36.3	38.8 ^b	38.5 ^b
2'	51.2	48.2	48.1	50.5	51.9	51.7	42.3	52.5 ^a	52.2 ^a
3'	40.6	30.3	30.3	34.0	36.7	39.1	33.1	38.7 ^b	36.6 ^b
2''	-	-	-	-	51.9	-	45.9	52.2 ^a	-
3''	-	-	-	-	38.0	-	33.7	38.5 ^b	-
3'''	-	-	-	-	-	-	-	36.6 ^b	-
CH₃									
Me-N ¹ -	44.8	30.9	30.8	30.7	35.2	36.3	36.4	35.7 ^l	35.8 ^k
MeN ^{1'} -	36.1	30.9	-	36.6	35.0	36.4	-	35.5 ^l	35.7 ^k
Me-N ^{1''} -	-	-	-	-	35.1	36.4	41.8	35.0 ^l	35.6 ^k
Me-N ^{1'''} -	-	-	-	-	-	-	-	-	35.2 ^k

Table 4. Cont.

Carbons	Compounds/ δ_C (ppm)								
	57 II,*	58 II,*	59 II,*	60 II	61 II,*	62 II,*	63 II,*	64 II,*	65 II,*
C									
3a	60.6	60.0	59.6 ^b	61.1	60.1 ^a	60.9 ^c	63.0 ^a	60.4 ^c	60.0 ^c
4a	-	132.0	132.4 ^c	132.9 ^b	-	132.7 ^d	-	132.8	132.1 ^e
7a	-	-	-	152.8	-	150.6 ^f	-	150.7 ^e	150.5 ^f
3'a	62.6	63.0	37.5 ^f	63.1	62.9	63.7 ^h	63.3 ^a	63.3 ^c	63.0
4'a	-	-	-	132.8 ^b	-	132.0 ^d	-	-	132.4 ^e
7'	-	110.0 ^c	-	123.8	-	-	-	-	108.8
7'a	-	-	-	151.0	-	148.9 ^f	-	150.3 ^e	148.9 ^f
5''	-	-	-	136.2	-	117.1	-	-	-
3''a	62.6	-	38.0 ^f	64.2	62.9	63.2 ^h	59.8 ^c	60.9 ^c	-
4''a	-	-	-	132.6	-	-	-	-	-
7''a	-	-	-	149.8	-	-	-	-	148.6
3'''a	60.6	-	60.6 ^b	62.3	60.6 ^a	60.1 ^c	59.8 ^c	-	60.5 ^c
4'''a	-	-	133.8 ^c	138.6	-	-	-	-	-
7'''	-	-	-	120.4	-	-	-	-	-
7'''a	-	-	-	144.7	-	-	-	-	-
3''''	-	-	-	114.5	-	-	-	-	-
3''''a	-	-	-	128.3	60.8 ^a	-	60.7 ^c	-	-
4''''a	-	-	-	-	-	-	-	-	-
CH									
4	-	126.0	-	126.9	-	123.6	-	126.1 ^d	125.3 ^d
4'	-	124.0	-	122.1	-	122.2	-	124.1	125.2 ^d
5	-	117.0 ^b	-	118.8	-	119.1 ^e	-	116.3	118.9
6	-	129.5	-	128.0	-	128.2	-	128.7	128.1
7	-	109.0 ^c	-	110.5	-	109.0	-	109.3	107.7
8a	85.8 ^a	88.0 ^d	88.5 ^d	87.3	86.0 ^b	87.2 ^g	81.8 ^b	86.6	86.9
5'	-	118.5 ^b	-	116.2	-	118.4 ^e	-	119.4	117.3
6'	-	127.5	-	126.5	-	126.1	-	126.2	125.3
8'a	82.3	83.0 ^d	74.0 ^g	82.4	82.6 ^c	85.8 ^g	86.8 ^b	82.8	86.0
4''	-	-	-	121.4	-	-	-	125.7 ^d	124.1
5''	-	119.0 ^b	-	-	-	117.1	-	-	-
6''	-	-	-	126.1	-	125.4	-	128.4	-
7''	-	-	-	108.5	-	-	-	-	-
8''a	82.3	-	72.0 ^g	83.4	82.3 ^c	83.1	86.8 ^d	83.0	-
4'''	-	-	-	123.3	-	-	-	122.7	123.6
5'''	-	119.5 ^b	-	118.8	-	-	-	-	-
6'''	-	128.5	-	125.1	-	-	-	-	-
7'''	-	-	-	120.4	-	-	-	-	-
8'''a	86.7 ^a	-	87.5 ^d	88.4	86.9 ^b	82.1	85.5 ^d	-	-
2''''	-	-	-	126.1	-	-	-	-	-
4''''	-	-	-	119.3	-	-	-	125.7	123.2
5''''	-	-	-	111.2	-	-	-	-	-
6''''	-	-	-	122.3	-	-	-	-	-
7''''	-	-	-	119.7	-	-	-	-	-
8''''a	-	-	-	-	85.1 ^b	-	84.8 ^d	-	-
CH ₂									
2	-	53.0	47.17 ^a	52.7 ^a	-	52.5 ^a	-	52 ^a	52.1 ^a
3	-	38.0 ^a	-	37.8	-	38.8 ^b	-	38.7 ^b	38.3 ^d
8	-	-	-	-	-	-	-	-	-
9	-	-	-	-	-	-	-	-	-
2'	-	-	-	52.7 ^a	-	52.0 ^a	-	52.8 ^a	52.4 ^a
3'	-	39.0 ^a	32.8 ^e	35.8	-	38.5 ^b	-	38.7 ^b	38.6 ^b
2''	-	-	-	52.6 ^a	-	-	-	52.9 ^a	-
3''	-	-	32.4 ^e	37.2	-	-	-	-	-
2'''	-	-	48.0 ^a	52.5 ^a	-	-	-	-	-
3'''	-	-	-	39.2	-	-	-	-	-
3''''	-	-	-	114.5	-	-	-	-	-

Table 4. Cont.

CH ₃	-	-	-	-	-	-	-	-
Me-N ^I -	36.0	36.1 ^h	34.8	-	35.6 ⁱ	-	35.7	35.4 ^g
Me-N ^{I'} -	-	42.6 ⁱ	35.3	-	35.1 ⁱ	-	-	35.6 ^g
Me-N ^{I''} -	-	42.6 ⁱ	35.8	-	-	-	-	-
Me-N ^{I'''} -	-	36.1 ^h	35.7	-	-	-	-	-
Me-N ^{I''''} -	-	-	36.5	-	-	-	-	-

^I CD₃OD, ^{II} CDCl₃ e ^{III} benzene-*d*₆, ^{ns} not specified; letters indicate signals that may be interchanged, nd = not detected; * indicates cases for which there was no complete detailed attribution of carbon signals.

2.3. ¹³C-NMR Chemical Shifts of Benzoquinolizidine Alkaloids Isolated from *Psychotria* Species

Muhammad et al. reported the isolation of five benzoquinolizidine alkaloids from *Psychotria klugii* [7] (Table 5). Among them, klugine (**66**) and 7'-*O*-demethylisocephaline (**67**) were reported for the first time, whereas cephaeline (**68**), isocephaline (**69**), and 7-*O*-methylpecoside (**70**) were previously isolated from *Cephaelis* species [56,57].

Table 5. Benzoquinolizidine Alkaloids from *P. klugii*.

Compound	Reference	¹³ C-NMR Data
Klugine (66)	[7]	[7]
7'- <i>O</i> -Demethylisocephaline (67)	[7]	[7]
Cephaeline (68)	[7]	[56]
Isocephaline (69)	[7]	[56]
7- <i>O</i> -Methylpecoside (70)	[7]	[57]

Compound **68** (ipecac alkaloid) as along with compounds **66**, **67**, and **69** possesses an unusual skeleton with two tetrahydroisoquinoline ring systems [10] characterized by the presence of four quaternary carbon signs at δ_C 147.2, 147.5 (C-9 and C-10, oxygenated *ortho*-substituted carbons), 126.8 (C-7a), and 130.1 (C-11a), two methine carbons at δ_C 108.6 (CH-11) and 111.5 (C-8), and signals at δ_C 62.4 (CH-11b), 52.3 (CH₂-6), and 29.2 (CH₂-7). A similar system is observed for the lower unit, with the exception of the absence of a methoxyl group attaching C-6' (a hydroxyl group in this position). The remarkable difference between compounds **68** and **69** (stereoisomers) is associated with the chemical shift of carbon C-1' at δ_C 51.9 and 55.3 respectively, whereas compounds **66** and **67** differ from **68** and **69** in the number and positions of the methoxyl groups. Interestingly, compound **70** exhibits carbon assignments consistent with a tetrahydroisoquinoline ring attached to a secologanin moiety at C-1. The chemical structures of compounds **68**–**70** are shown in Figure 4, and their ¹³C-NMR data are listed in Table 6.

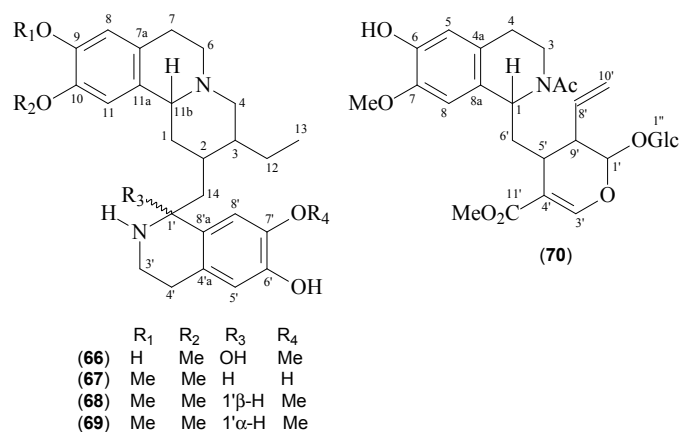
Figure 4. Structures of Benzoquinolizidine Alkaloids from *P. klugii*.

Table 6. ^{13}C -NMR Data of Benzoquinolizidine Alkaloids from *P. klugii*.

Carbons	Compound/ δ_{C} (ppm)				
	66 ^{ns}	67 ^{ns}	68 ^{II}	69 ^{II}	70 ^I
C					
6	-	-	-	-	146.5 ^a
7	-	-	-	-	147.8 ^a
9	146.5 ^a	146.8	147.2 ^a	147.2 ^a	-
10	147.8 ^b	148.0	147.5 ^a	147.4 ^a	-
4a	-	-	-	-	126.9
7a	127.8	126.9	126.8	126.5	-
8a	-	-	-	-	130.2
11a	129.7	127.9	130.1	129.9	-
1'	79.5	-	-	-	-
4'	-	-	-	-	111.7
4'a	127.7	123.2	127.6	127.9	-
6'	146.4 ^a	145.6	143.9 ^b	144.0	-
7'	-	-	-	-	-
11'	-	-	-	-	169.2
8'a	129.7	126.0	131.1	131.0	-
CH					
1	-	-	-	-	50.6
2	-	-	-	-	-
3	42.5	41.3	41.7	61.5	-
5	-	-	-	-	116.2
8	116.2	112.1	111.5	111.4	111.1
11	109.7	109.0	108.6	108.2	-
11b	63.8	62.7	62.4	62.8	-
1'	-	53.6	51.9	55.3	98.7
3'	-	-	-	-	153.1
4'	28.5	27.6	29.0	29.3	-
5'	116.4	115.2	114.7	114.8	27.5
8'	110.0	113.2	108.4	108.6	136.3
9'	-	-	-	-	45.1
CH₂					
1	40.6	36.9	36.9	39.3	-
3	-	-	-	-	36.1
4	62.2	61.6	61.3	52.6	29.1
6	53.3	51.9	52.3	52.6	-
7	29.3	25.3	29.2	29.1	-
12	24.4	23.3	23.6	24.0	-
14	37.0	38.0	40.9	40.4	-
3'	41.0	39.5	40.1	41.4	-
4'	28.5	27.6	29.0	29.3	-
6'	-	-	-	-	41.1
10'	-	-	-	-	120.1
CH₃					
13	11.5	10.1	11.2	11.3	-
Me ⁷ -O-	-	-	-	-	56.5
Me ⁹ -O-	-	55.4 ^d	55.8 ^e	55.8 ^f	-
Me ¹⁰ -O-	56.8 ^c	55.8 ^d	56.0 ^e	56.0 ^f	-
Me ^{7'} -O-	56.6 ^c	-	56.3 ^e	56.0 ^f	-
Glucose					
1''	-	-	-	-	100.5
2''	-	-	-	-	74.8
3''	-	-	-	-	78.2 ^b
4''	-	-	-	-	71.5
5''	-	-	-	-	78.3 ^b
6''	-	-	-	-	62.7
CO₂Me					51.7

^I CD₃OD, ^{II} CDCl₃ e^{ns} not specified; letters indicate signals that may be interchanged.

3. Conclusions

In this work, we attempted to compile ^{13}C -NMR data of alkaloids isolated from the *Psychotria* genus and provide information that may be useful in order to distinguish different types of skeletons. For monoterpene indole alkaloids (MIAs), mainly found in tropical species, a good strategy for their structural elucidation is to compare their spectral data with those observed for strictosidine (1). The monitoring of differences in specific parts of the spectrum, such as the signals of C-22, CH-17, CH-12, CH₂-5, and CH-1', may suggest alternative structural possibilities. Note that all comparisons performed in this work are restricted where possible to compounds whose ^{13}C -NMR experiments were run in the same solvent.

The main pyrrolidinoindoline alkaloids found in this genus are chimonanthine derivatives, with units linked mostly by C3a-C3'a or C-3a-C7a bonds. Some examples have shown different patterns of linkages between N (from tryptamine terminal units) and C-3a. For compounds with more than three units, such as quadrigemines A–C and psychotridine and its isomer, obtaining detailed assignments of these carbons is not possible owing to structural complexity. The occurrence of benzoquinolizidine alkaloids in *Psychotria* species is less common, comprising some compounds isolated from *Psychotria klugii*.

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References

1. Marques de Oliveira, A.; Lyra Lemos, R.P.; Conserva, L.M. β -Carboline alkaloids from *Psychotria barbiflora* DC. (Rubiaceae). *Biochem. Syst. Ecol.* **2013**, *50*, 339–341. [[CrossRef](#)]
2. Takayama, H.; Mori, I.; Kitajima, M.; Aimi, N.; Lajis, N.H. New Type of Trimeric and Pentameric Indole Alkaloids from *Psychotria rostrata*. *Org. Lett.* **2004**, *6*, 2945–2948. [[CrossRef](#)] [[PubMed](#)]
3. Benevides, P.J.C.; Young, M.C.M.; Bolzani, V.D.S. Biological Activities of Constituents from *Psychotria spectabilis*. *Pharm. Biol.* **2004**, *42*, 565–569. [[CrossRef](#)]
4. Amador, T.A.; Verotta, L.; Nunes, D.S.; Elisabetsky, E. Antinociceptive profile of hodgkinsine. *Planta Med.* **2000**, *66*, 770–772. [[CrossRef](#)] [[PubMed](#)]
5. Both, F.L.; Kerber, V.A.; Henriques, A.T.; Elisabetsky, E. Analgesic Properties of Umbellatine from *Psychotria umbellata*. *Pharm. Biol.* **2002**, *40*, 336–341. [[CrossRef](#)]
6. Matsuura, H.N.; Fett-Neto, A.G. The major indole alkaloid *N*, β -D-glucopyranosyl vincosamide from leaves of *Psychotria leiocarpa* Cham. & Schltdl. is not an antifeedant but shows broad antioxidant activity. *Nat. Prod. Res.* **2013**, *27*, 402–411. [[PubMed](#)]
7. Muhammad, I.; Dunbar, D.C.; Khan, S.I.; Tekwani, B.L.; Bedir, E.; Takamatsu, S.; Ferreira, D.; Walker, L.A. Antiparasitic Alkaloids from *Psychotria klugii*. *J. Nat. Prod.* **2003**, *66*, 962–967. [[CrossRef](#)] [[PubMed](#)]
8. Mahmud, Z.; Musa, M.; Ismail, N.; Lajis, N.H. Cytotoxic and Bacteriocidal Activities of *Psychotria rostrata*. *Int. J. Pharmacogn.* **1993**, *31*, 142–146. [[CrossRef](#)]
9. Adjibade, Y.; Kuballa, B.; Cabalion, P.; Jung, M.L.; Beck, J.P.; Anton, R. Cytotoxicity on human leukemic and rat hepatoma cell lines of alkaloid extracts of *Psychotria forsteriana*. *Planta Med.* **1989**, *55*, 567–568. [[CrossRef](#)] [[PubMed](#)]
10. Dewick, P.D. *Medicinal Natural Products: A Biosynthetic Approach*; Wiley: West Sussex, UK, 2002; p. 350.
11. O'Connor, S.E.; Maresh, J.J. Chemistry and biology of monoterpene indole alkaloid biosynthesis. *Nat. Prod. Rep.* **2006**, *23*, 532–547. [[CrossRef](#)] [[PubMed](#)]
12. Fasshuber, H. Strukturbestimmung sowie Untersuchung der Enzymkatalysierten Hydrolyse von Naturstoffen mit Glycosidstruktur aus der Pflanzengattung *Psychotria* L. Master's Thesis, Universität Wien, Vienna, Austria, 2011; p. 96.

13. Patthy-luka, A.; Károlyházy, L.; Szabó, L.F.; Podányi, B. First Direct and Detailed Stereochemical Analysis of Strictosidine. *J. Nat. Prod.* **1997**, *60*, 69–75. [[CrossRef](#)]
14. Achenbach, H.; Lottes, M.; Waibel, R.; Karikas, G.A.; Correa, M.D.; Gupta, M.P. Alkaloids and other compounds from *Psychotria correae*. *Phytochemistry* **1995**, *38*, 1537–1545. [[CrossRef](#)]
15. Pimenta, A.T.A.; Braz-Filho, R.; Delprete, P.G.; de Souza, E.B.; Silveira, E.R.; Lima, M.A.S. Structure elucidation and NMR assignments of two unusual monoterpene indole alkaloids from *Psychotria stachyoides*. *Magn. Reson. Chem.* **2010**, *48*, 734–737. [[CrossRef](#)] [[PubMed](#)]
16. Farias, F.M.; Konrath, E.L.; Zuanazzi, J.A.S.; Henriques, A.T. Strictosamide from *Psychotria nuda* (Cham. et Schltdl) Wawra (Rubiaceae). *Biochem. Syst. Ecol.* **2008**, *36*, 919–920. [[CrossRef](#)]
17. Paul, J.H.A.; Maxwell, A.R.; Reynolds, W.F. Novel Bis (monoterpene) Indole Alkaloids from *Psychotria bahiensis*. *J. Nat. Prod.* **2003**, *66*, 752–754. [[CrossRef](#)] [[PubMed](#)]
18. Faria, E.O.; Kato, L.; de Oliveira, C.M.A.; Carvalho, B.G.; Silva, C.C.; Sales, L.S.; Schuquel, I.T.A.; Silveira-Lacerda, E.P.; Delprete, P.G. Quaternary β -carboline alkaloids from *Psychotria prunifolia* (Kunth) Steyerem. *Phytochem. Lett.* **2010**, *3*, 113–116. [[CrossRef](#)]
19. Van De Santos, L.; Fett-Neto, A.G.; Kerber, V.A.; Elisabetsky, E.; Quirion, J.-C.; Henriques, A.T. Indole monoterpene alkaloids from leaves of *Psychotria suterella* Müll. Arg. (Rubiaceae). *Biochem. Syst. Ecol.* **2001**, *29*, 1185–1187. [[CrossRef](#)]
20. Kato, L.; De Oliveira, C.M.A.; Faria, E.O.; Ribeiro, L.C.; Carvalho, B.G.; Da Silva, C.C.; Schuquel, I.T.A.; Santin, S.M.O.; Nakamura, C.V.; Britta, E.A.; et al. Antiprotozoal Alkaloids from *Psychotria prunifolia* (Kunth) Steyerem. *J. Braz. Chem. Soc.* **2012**, *23*, 355–360.
21. Kerber, V.A.; Gregianini, T.S.; Paranhos, T.; Farias, F.; Fett, J.P.; Fett-neto, A.G.; Zuanazzi, S.; Quirion, J.; Elizabetsky, E.; Henriques, T. Brachycerine, a Novel Monoterpene Indole Alkaloid from *Psychotria brachyceras*. *J. Nat. Prod.* **2001**, *64*, 677–679. [[CrossRef](#)] [[PubMed](#)]
22. Kerber, V.A.; Passos, C.S.; Verli, H.; Quirion, J.P.; Henriques, A.T. Psychollatine, a Glucosidic Monoterpene Indole Alkaloid from *Psychotria umbellata*. *J. Nat. Prod.* **2008**, *71*, 697–700. [[CrossRef](#)] [[PubMed](#)]
23. Berger, A.; Fasshuber, H.; Schinnerl, J.; Brecker, L.; Greger, H. Various types of tryptamine-iridoid alkaloids from *Palicourea acuminata* (= *Psychotria acuminata*, Rubiaceae). *Phytochem. Lett.* **2012**, *5*, 558–562. [[CrossRef](#)]
24. Farias, F.M.; Passos, C.S.; Arbo, M.D.; Barros, D.M.; Gottfried, C.; Steffen, V.M.; Henriques, A.T. Strictosidinic acid, isolated from *Psychotria myriantha* Mull. Arg. (Rubiaceae), decreases serotonin levels in rat hippocampus. *Fitoterapia* **2012**, *83*, 1138–1143. [[CrossRef](#)] [[PubMed](#)]
25. Simoes-Pires, C.A.; Farias, F.M.; Martston, A.; Queiroz, E.; Chaves, C.G.; Henriques, A.T.; Hostettmann, K. Indole monoterpenes with antichemotactic activity from *Psychotria myriantha*: Chemotaxonomic significance. *Nat. Prod. Commun.* **2006**, *1*, 1101–1106.
26. Morita, H.; Ichihara, Y.; Takeya, K.; Watanabe, K.; Itokawa, H.; Motidome, M. A New Indole Alkaloid Glycoside from the Leaves of *Palicourea marcgravii*. *Planta Med.* **1989**, *55*, 288–289. [[CrossRef](#)] [[PubMed](#)]
27. Ferrari, F.; Messana, I.; Botta, B.; de Mello, J.F. Constituents of *Guettarda Platypoda*. *J. Nat. Prod.* **1986**, *49*, 1150–1151. [[CrossRef](#)]
28. Henriques, A.T.; Lopes, S.O.; Paranhos, J.T.; Gregianini, T.S.; Von Poser, G.L.; Fett-Neto, A.G.; Schripsema, J. *N*, β -D-Glucopyranosyl vincosamide, a light regulated indole alkaloid from the shoots of *Psychotria leiocarpa*. *Phytochemistry* **2004**, *65*, 449–454. [[CrossRef](#)] [[PubMed](#)]
29. Pimenta, A.T.A.; Braz-filho, R.; Silveira, E.R.; Lima, M.A.S. Alkaloid and other Chemical Constituents from *Psychotria stachyoides* Benth. *J. Braz. Chem. Soc.* **2011**, *22*, 2216–2219. [[CrossRef](#)]
30. Liew, S.Y.; Mukhtar, M.R.; Hadi, A.H.A.; Awang, K.; Mustafa, M.R.; Zaima, K.; Morita, H.; Litaudon, M. Nauline, a new indole alkaloid from the bark of *Nauclea officinalis*. *Molecules* **2012**, *17*, 4028–4036. [[CrossRef](#)] [[PubMed](#)]
31. Pimenta, A.A.T.; Braz-Filho, R.; Delprete, P.G.; De Souza, E.B.; Silveira, E.R.; Lima, M.A.S. Unusual monoterpene indole alkaloids from *Psychotria stachyoides* Benth. *Biochem. Syst. Ecol.* **2010**, *38*, 846–849. [[CrossRef](#)]
32. Kerber, V.A.; Passos, C.S.; Klein-Júnior, L.C.; Quirion, J.-C.; Pannecoucke, X.; Salliot-Maire, I.; Henriques, A.T. Three new monoterpene indole alkaloids from *Psychotria umbellata* Thonn. *Tetrahedron Lett.* **2014**, *55*, 4798–4800. [[CrossRef](#)]

33. Passos, C.S.; Simões-Pires, C.A.; Nurisso, A.; Soldi, T.C.; Kato, L.; de Oliveira, C.M.A.; de Faria, E.O.; Marcourt, L.; Gottfried, C.; Carrupt, P.-A.; et al. Indole alkaloids of *Psychotria* as multifunctional cholinesterases and monoamine oxidases inhibitors. *Phytochemistry* **2013**, *86*, 8–20. [[CrossRef](#)] [[PubMed](#)]
34. Sacconnay, L.; Ryckewaert, L.; Passos, C.S.; Guerra, M.C.; Kato, L.; De Oliveira, C.M.A.; Henriques, A.; Carrupt, P.-A.; Simões-Pires, C.; Nurisso, A. Alkaloids from *Psychotria* Target Sirtuins: In Silico and In Vitro Interaction Studies. *Planta Med.* **2015**, *81*, 517–524. [[CrossRef](#)] [[PubMed](#)]
35. Abreu, P.; Pereira, A. A new indole alkaloid from *Sarcocephalus latifolius*. *Heterocycles* **1998**, *48*, 885–891. [[CrossRef](#)]
36. Passos, C.S.; Soldi, T.C.; Abib, R.T.; Apel, M.A.; Simões-Pires, C.; Marcourt, L.; Gottfried, C.; Henriques, A.T. Monoamine oxidase inhibition by monoterpene indole alkaloids and fractions obtained from *Psychotria suterella* and *Psychotria laciniata*. *J. Enzyme Inhib. Med. Chem.* **2013**, *28*, 611–618. [[CrossRef](#)] [[PubMed](#)]
37. Levesque, J.; Jacquesy, R. Alcoyl—Glucosides: Nouveaux composés isolés de *Pauridiantha lyalii* Brem (Rubiaceae). *Tetrahedron* **1982**, *38*, 1417–1434. [[CrossRef](#)]
38. Paranhos, J.T.; Fragoso, V.; da Silveira, V.C.; Henriques, A.T.; Fett-Neto, A.G. Organ-specific and environmental control of accumulation of psychollatine, a major indole alkaloid glucoside from *Psychotria umbellata*. *Biochem. Syst. Ecol.* **2009**, *37*, 707–715. [[CrossRef](#)]
39. Brandt, V.; Tits, M.; Geerlings, A.; Frédéricich, M.; Penelle, J.; Delaude, C.; Verpoorte, R.; Angenot, L. β -Carboline glucoalkaloids from *Strychnos mellodora*. *Phytochemistry* **1999**, *51*, 1171–1176. [[CrossRef](#)]
40. Naves, R.F. Estudo Fitoquímico das Folhas de *Psychotria hoffmannseggiana* Roem & Schult (Rubiaceae). Master's Thesis, Universidade Federal de Goiás, Goiás, Brazil, 2014; pp. 1–212.
41. Verotta, L.; Peterlongo, F.; Elisabetsky, E.; Amador, T.A.; Nunes, D. High-performance liquid chromatography-diode array detection-tandem mass spectrometry analyses of the alkaloid extracts of Amazon *Psychotria* species. *J. Chromatogr. A* **1999**, *841*, 165–176. [[CrossRef](#)]
42. Verotta, L.; Pilati, T.; Tato, M.; Elisabetsky, E.; Amador, T.A. Pyrrolidinoindoline Alkaloids from *Psychotria colorata*. *J. Nat. Prod.* **1998**, *61*, 392–396. [[CrossRef](#)] [[PubMed](#)]
43. Zhou, H.; He, H.; Wang, Y.; Hao, X. A New Dimeric Alkaloid from the Leaf of *Psychotria calocarpa*. *Helv. Chim. Acta* **2010**, *93*, 1650–1652. [[CrossRef](#)]
44. Hart, N.; Johns, S.; Lambertson, J.; Summons, R. Psychotridine, a C₅₅H₆₂N₁₀, Alkaloid from *Psychotria beccarioides* (Rubiaceae). *Aust. J. Chem.* **1974**, *27*, 639–646. [[CrossRef](#)]
45. Libot, F.; Miet, C.; Kunesch, N.; Poisson, J.E. Rubiacées d'Océanie: Alcaloïdes de *Psychotria oleoides* de Nouvelle-Calédonie et de *Calycodendron milnei* du Vanuatu (Nouvelles-Hébrides). *J. Nat. Prod.* **1987**, *50*, 468–473. [[CrossRef](#)]
46. Gueritz-voegelein, F.; Sévenet, T.; Puset, J.; Adeline, M.-T.; Gillet, B.; Beloeil, J.-C.; Guénard, D.; Portier, P. Alkaloids from *Psychotria Oleoides* with Activity on Growth Hormone Release. *J. Nat. Prod.* **1992**, *55*, 923–930. [[CrossRef](#)]
47. Liu, Y.; Wang, J.-S.; Wang, X.-B.; Kong, L.-Y. Absolute configuration study of a new dimeric indole alkaloid from leaves and twigs of *Psychotria henryi*. *J. Asian Nat. Prod. Res.* **2014**, *16*, 29–33. [[CrossRef](#)] [[PubMed](#)]
48. Liu, Y.; Wang, J.-S.; Wang, X.-B.; Kong, L.-Y. Two novel dimeric indole alkaloids from the leaves and twigs of *Psychotria henryi*. *Fitoterapia* **2013**, *86*, 178–182. [[CrossRef](#)] [[PubMed](#)]
49. Jannic, V.; Guéritte, F.; Lapre, O.; Serani, L.; Martin, M.-T.; Sévenet, T.; Potier, P. Pyrrolidinoindoline Alkaloids from *Psychotria oleoides* and *Psychotria lyciiflora*. *J. Nat. Prod.* **1999**, *33*, 838–843. [[CrossRef](#)] [[PubMed](#)]
50. Adjibade, Y.; Weniger, B.; Quirion, J.C.; Kuballa, B.; Cabalion, P.; Anton, R. Dimeric Alkaloids from *Psychotria forsteriana*. *Phytochemistry* **1992**, *31*, 317–319. [[CrossRef](#)]
51. Solís, P.N.; Ravelo, A.G.; Palenzuela, J.A.; Gupta, M.P.; González, A.; Phillipson, J.D. Quinoline Alkaloids from *Psychotria glomerulata*. *Phytochemistry* **1997**, *44*, 963–969. [[CrossRef](#)]
52. Li, X.; Zhang, Y.; Cai, X.; Feng, T.; Liu, Y.; Li, Y.; Ren, J.; Zhu, H.-J.; Luo, X.-D. Psychotripine: A New Trimeric Pyrrolidinoindoline Derivative from *Psychotria pilifera*. *Org. Lett.* **2011**, *13*, 5896–5899. [[CrossRef](#)] [[PubMed](#)]
53. Roth, A.; Kuballa, B.; Bounthan, C.; Cabalion, P.; Sévenet, T.; Beck, J.P.; Anton, R. Cytotoxic Activity of Polyindoline Alkaloids of *Psychotria forsteriana* (Rubiaceae). *Planta Med.* **1986**, *6*, 450–453. [[CrossRef](#)]
54. Rasolonjanahary, R.; Sévenet, T.; Voegelein, F.G.; Kordon, C. Psycholeine, a natural alkaloid extracted from *Psychotria oleoides*, acts as a weak antagonist of somatostatin. *Eur. J. Pharmacol.* **1995**, *285*, 19–23. [[CrossRef](#)]
55. Roth, A.; Kuballa, B.; Cabalion, P.; Anton, R. Preliminary Study of the Alkaloids of *Psychotria forsteriana*. *Planta Med.* **1985**, *51*, 289. [[CrossRef](#)]

56. Itoh, A.; Ikuta, Y.; Baba, Y.; Tanahashi, T.; Nagakura, N. Ipecac alkaloids from *Cephaelis acuminata*. *Phytochemistry* **1999**, *52*, 1169–1176. [[CrossRef](#)]
57. Itoh, A.; Tanahashi, T.; Nagakura, N.; Nayeshiro, H. Tetrahydroisoquinoline-monoterpene glucosides from *Alangium lamarckii* and *Cephaelis ipecacuanha*. *Phytochemistry* **1994**, *36*, 383–387. [[CrossRef](#)]



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