



3 | Antimicrobial Chemotherapy | Full-Length Text

Clinical outcomes and pharmacokinetics/pharmacodynamics of intravenous polymyxin B treatment for various site carbapenem-resistant gram-negative bacterial infections: a prospective observational multicenter study

Zhenwei Yu,¹ Huangdu Hu,² Xiaofen Liu,^{3,4} Jieqiong Liu,¹ Lingyan Yu,⁵ Anqi Wei,⁶ Chuanwei Xin,⁷ Yongxiong Gan,⁸ Shu Lei,⁹ Li Zhuang,¹⁰ Yanfei Shen,¹¹ Xiaoxing Du,¹ Jianping Zhu,¹ Yi Yang,¹ Gang Liang,¹ Feng Guo,¹ Jing Zhang,^{3,4} Yunsong Yu^{1,2}

AUTHOR AFFILIATIONS See affiliation list on p. 16.

ABSTRACT Polymyxin B, a last resort for carbapenem-resistant gram-negative bacteria (CRGNB) infections, has infection site-specific pharmacokinetic/pharmacodynamic (PK/PD) properties. However, there is little clinical evidence to support optimal exposures of polymyxin B for different site infections. We performed a prospective, observational, multicenter study to evaluate the clinical outcomes and PK/PD of intravenous polymyxin B treatment for various site CRGNB infections. The main clinical outcomes were 14-day all-cause mortality and nephrotoxicity, and the secondary outcomes were 28-day mortality and clinical response. The area under curves (AUCs) of polymyxin B were determined, and their associations with clinical outcomes were analyzed by stratification based on the infection site. A total of 312 patients were ultimately enrolled from 10 research centers. The overall 14-day mortality was 29.5%, and those of patients with lower respiratory tract infection (LRTI), intra-abdominal infection (IAI), and bloodstream infection (BSI) were 32.3%, 19.7%, and 30.3%, respectively. The 28-day mortality rate was 38.1%, while LRTI patients had the highest mortality (41.4%) and IAI patients lowest (34.8%). The clinical response rate was 46.2%, which was similar among the subgroups. The overall AKI rate was 60.9%. An AUC greater than 50 mg·h/L was related to lower mortality in IAI patients but not in LRTI patients, which led to a lower but not significant difference in the overall analysis. The AUC of polymyxin B was an independent risk factor for 14-day mortality in IAI patients, and the cutoff value was 76 mg·h/L. The results would be helpful for personalized dosing and monitoring of polymyxin B.

CLINICAL TRIALS This study is registered with the Chinese Clinical Trial Registry as ChiCTR2200056667.

KEYWORDS polymyxin B, mortality, acute kidney injury, pharmacokinetic/pharmacodynamic, AUC

The emergence of multidrug-resistant pathogens has become a major challenge for healthcare providers (1). Infection caused by carbapenem-resistant gram-negative bacteria (CRGNB), mainly carbapenem-resistant *Klebsiella pneumoniae* (CRKP), carbapenem-resistant *Pseudomonas aeruginosa* (CRPA), and carbapenem-resistant *Acinetobacter baumannii* (CRAB), leads to increased mortality, longer hospital stays, and additional costs (2–5). Unfortunately, drugs that are susceptible to CRGNB are few in number (6).

Polymyxin, a fast bacterial-killing agent once abandoned due to its severe neurotoxicity and nephrotoxicity, had been brought back into the field of clinical application (7). It has good *in vitro* activity against CRGNBs and is considered one of the "last-resort"

Editor James E. Leggett, Providence Portland Medical Center, Portland, Oregon, USA

Address correspondence to Jing Zhang, zhangj61@fudan.edu.cn, or Yunsong Yu, yvys119@zju.edu.cn.

Zhenwei Yu, Huangdu Hu, and Xiaofen Liu contributed equally to this article. Author order was determined in order of increasing seniority.

The authors declare no conflict of interest.

See the funding table on p. 16.

Received 12 December 2024 **Accepted** 5 February 2025 **Published** 6 March 2025

Copyright © 2025 Yu et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

of CRGNB infection (8). Although novel beta-lactam/beta-lactamase inhibitors against CRGNB are available in recent years, polymyxins still play an important role, especially in developing countries (9). There are two main forms of polymyxins available on the market: colistin methanesulfonate (CMS) and polymyxin B sulfate. Although their molecular structures are similar, they involve distinct *in vivo* processes (10). Polymyxin B is administered in its active form and is considered to have superior pharmacological properties to CMS (11).

Polymyxin B has a narrow therapeutic window as low exposure leads to treatment failure and heterogeneous resistance, but high exposure can cause significant neurotoxicity and nephrotoxicity (12-14). It should be dosed under the guidance of the pharmacokinetic/pharmacodynamic (PK/PD) principle, but there are limited data to suggest an optimal exposure of polymyxin B to balance the efficacy and safety. A preclinical PK/PD study revealed that the free-drug area under the curve over the minimum inhibitory concentration (fAUC/MIC) was the PK/PD index associated with bacterial killing, and its value should be above 3.72-28 for 2 log kill of KP in thigh infection models (15). Thus, the 2019 international consensus guideline recommended an AUC target of 50-100 mg·h/L without sufficient clinical efficacy data (16). However, polymyxin B has site-specific PD properties. Studies in animal models have suggested that a much higher PK/PD target for lung infection is needed and that bacterial stasis cannot be achieved for some strains (15). Polymyxin B treatment also resulted in poorer clinical outcomes in patients with lower respiratory tract infection (LRTI) than in those with other site infections. Recent PK/PD studies including patients with various infection sites have suggested optimal AUC thresholds, but the results are inconclusive (17). Considering the viability of PKs and the uncertainty in PK/PD, it remains a challenge to dose polymyxin B in clinical practice (18). The Prato polymyxin consensus claimed that larger pharmacokinetic/pharmacodynamic and clinical studies of polymyxin B are urgently needed to develop improved dosing strategies with this drug (19).

Thus, we designed a prospective observational multicenter study to assess the clinical outcomes and PK/PD of intravenous polymyxin B treatment for various site CRGNB infection and to determine the optimal exposure range of polymyxin B, which would be helpful in the future clinical application of this drug.

RESULTS

Patient inclusion and characteristics and distribution of the AUC

As shown in Fig. 1, a total of 312 patients were ultimately enrolled from 10 research centers. All patients were eligible for clinical outcome evaluation, and 184 of those patients were eligible for nephrotoxicity analysis. The detailed patient characteristics are shown in Table 1. Most of the patients were old and admitted to the ICU. The main infection type was lower respiratory tract infection (LRTI), and some patients had multiple infection sites.

Clinical outcomes

The overall 14-day mortality was 29.5%, while patients with LRTI and bloodstream infections (BSI) had higher mortality rates (Fig. 2; Table 2). The 28-day mortality rate was 38.1%, while LRTI patients had the highest mortality (41.4%), and intra-abdominal infection (IAI) patients had the lowest (34.8%). However, the clinical response rate was 46.2%, which was similar among the subgroups. Among the patients eligible for nephrotoxicity analysis, the overall acute kidney injury (AKI) rate was 60.9%.

PK/PD analysis

The polymyxin B AUC distributions in whole patients and subgroups are shown in Fig. 3. Kaplan–Meier analysis was performed to test the differences in mortality between patients with different polymyxin B exposures. When we stratified the patients according to the AUC 50 mg·h/L threshold, the overall survival curve revealed that patients with

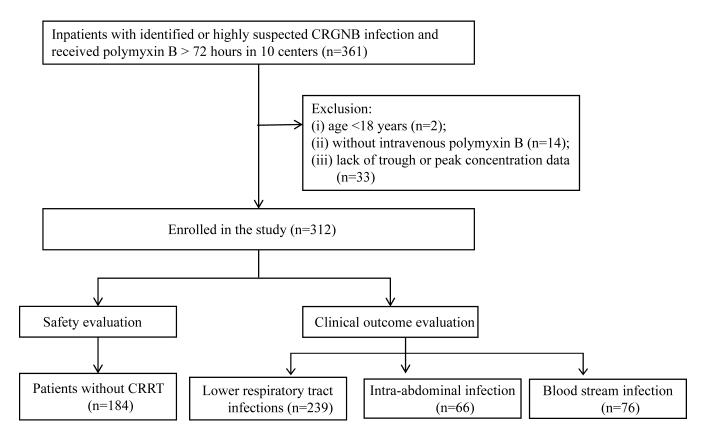


FIG 1 Flowchart of patient enrollment. CRGNB, carbapenem-resistant gram-negative bacteria; CRRT, continuous renal replacement treatment.

AUCs over 50 mg·h/L tended to have lower mortality, but the differences were not significant (Fig. 4A). However, the curves were similar in patients with LRTI (Fig. 4B). IAI patients with an AUC over 50 mg·h/L had significantly lower 14-day mortality (Fig. 4C). However, BSI patients had lower 28-day mortality if the AUC was greater than 50 mg·h/L (Fig. 4D).

The multivariate logistic regression identified several independent risk factors for overall 14-day mortality, but the AUC did not (Table 3). However, the AUC and use of CRRT were independent risk factors for 14-day mortality in IAI patients (Table 4). In patients with LRTI and BSI, the AUC was not associated with 14-day mortality (Tables S1 and S2). The results of multivariate logistic regression for secondary outcomes and other infection types are shown in Tables S3 to S11. Only in patients with IAI was the AUC associated with 28-day mortality (Table S8).

We performed ROC analysis for the area under the curve and 14-day mortality of IAI patients, and the sensitivity of the polymyxin B AUC for mortality was 0.768 (P < 0.001, Fig. 5A). The optimal cutoff value for the AUC obtained by ROC analysis was 76 mg·h/L. Subsequent Kaplan–Meier analysis revealed that an AUC over 76 mg·h/L was associated with much lower 14-day and 28-day mortality (Fig. 5B).

No significant association between the AUC and nephrotoxicity was identified in this study (Table 5).

DISCUSSION

To the best of our knowledge, this is the largest prospective observational multicenter study to assess the clinical outcomes and PK/PD of intravenous polymyxin B treatment for CRGNB infections with different infection sites. In terms of clinical outcomes, patients with IAI had lower mortality than those with other site infections. The AUC of polymyxin B was significantly associated with mortality for the first time, although only in IAI patients. A new polymyxin B AUC cut-off for IAI patients was provided by this study.

 TABLE 1
 Demographic characteristics of the included patients^a

Variable	Total (n = 312)	Subgroups				
		Non-CRRT (n = 184)	LRTIs (n = 239)	IAIs (n = 66)	BSIs (n = 76)	
Sex, n (%)						
Male	221 (70.8%)	128 (69.6%)	178 (74.5%)	41 (62.1%)	53 (69.7%)	
Female	91 (29.2%)	56 (30.4%)	61 (25.5%)	25 (37.9%)	23 (30.3%)	
Age (years)	69.0 (58.0, 80.0)	69.0 (58.0, 79.0)	72.0 (61.0, 82.0)	59.5 (51.0, 69.0)	68.0 (58.0, 77.0)	
Veight (kg)	60.5 (54.6, 70.0)	60.5 (54.1, 70.0)	60.0 (54.0, 70.0)	60.0 (55.0, 70.0)	65.0 (55.5, 70.0)	
BMI (kg/m²)	22.1 (19.8, 24.8)	22.0 (19.6, 24.8)	22.0 (19.6, 24.5)	22.9 (20.6, 25.1)	23.1 (20.8, 26.0)	
Days of hospital stay	45.0 (27.0, 76.0)	53.5 (30.0, 85.0)	43.0 (27.0, 70.0)	61.5 (38.0, 99.0)	43.0 (27.0, 80.0)	
Comorbidities, n (%)						
Dementia	14 (4.50%)	7 (3.80%)	13 (5.40%)	0 (0.00%)	3 (3.90%)	
Diabetes	85 (27.2%)	50 (27.2%)	69 (28.9%)	8 (12.1%)	24 (31.6%)	
Chronic kidney disease	35 (11.2%)	10 (5.40%)	30 (12.6%)	5 (7.60%)	9 (11.8%)	
Malignancy	75 (24.0%)	52 (28.3%)	53 (22.2%)	13 (19.7%)	16 (21.1%)	
Heart failure	40 (12.8%)	20 (10.9%)	37 (15.5%)	1 (1.50%)	7 (9.20%)	
COPD	30 (9.60%)	20 (10.9%)	28 (11.7%)	0 (0.00%)	4 (5.30%)	
Immunodeficiency	20 (6.40%)	6 (3.30%)	18 (7.50%)	3 (4.50%)	7 (9.20%)	
Hypertension	145 (46.5%)	76 (41.3%)	114 (47.7%)	21 (31.8%)	36 (47.4%)	
Hyperlipidemia	9 (2.90%)	6 (3.30%)	5 (2.10%)	5 (7.60%)	1 (1.30%)	
Acute pancreatitis	37 (11.9%)	21 (11.4%)	15 (6.30%)	31 (47.0%)	8 (10.5%)	
COVID-19	38 (12.2%)	21 (11.4%)	34 (14.2%)	5 (7.60%)	9 (11.8%)	
CU admission	290 (92.9%)	162 (88.0%)	230 (96.2%)	65 (98.5%)	68 (89.5%)	
Days of ICU stay	30.0 (19.0, 53.0)	35.5 (18.0, 60.0)	35.0 (19.0, 56.0)	37.0 (22.0, 51.0)	33.0 (20.0, 54.0)	
OFA score	8.00 (5.00, 12.0)	6.00 (4.00, 9.00)	9.00 (6.00, 12.0)	8.00 (4.00, 12.0)	8.00 (5.00, 12.0)	
APACHE II score	24.0 (17.0, 29.0)	21.0 (15.0, 27.0)	24.5 (18.0, 30.0)	24.5 (15.0, 28.0)	22.0 (17.0, 29.0)	
severely ill	160 (51.3%)	70 (38.0%)	132 (55.2%)	32 (48.5%)	38 (50.0%)	
Mechanical ventilation	229 (73.4%)	128 (69.6%)	190 (79.5%)	42 (63.6%)	51 (67.1%)	
Surgical removal of infection	43 (13.8%)	25 (13.6%)	23 (9.60%)	27 (40.9%)	12 (15.8%)	
nfected site, n (%)						
LRTIs	239 (76.6%)	137 (74.5%)	239 (100%)	28 (42.4%)	46 (60.5%)	
IAIs	66 (21.2%)	32 (17.4%)	28 (11.7%)	66 (100%)	11 (14.5%)	
BSIs	76 (24.4%)	42 (22.8%)	46 (19.2%)	11 (16.7%)	76 (100%)	
Other or undefined	36 (11.5%)	21 (11.4%)	23 (9.60%)	7 (10.6%)	10 (13.2%)	
athogen, n (%)						
CRKP	86 (27.6%)	51 (27.7%)	55 (23.0%)	34 (51.5%)	27 (35.5%)	
CRPA	65 (20.8%)	37 (20.1%)	52 (21.8%)	15 (22.7%)	10 (13.2%)	
CRAB	141 (45.2%)	73 (39.7%)	128 (53.6%)	30 (45.5%)	31 (40.8%)	
Others	137 (43.9%)	84 (45.7%)	107 (44.8%)	28 (42.4%)	47 (61.8%)	
CRRT	128 (41.0%)	0 (0.00%)	102 (42.7%)	34 (51.5%)	34 (44.7%)	
:CMO	23 (7.40%)	7 (3.80%)	23 (9.60%)	1 (1.50%)	6 (7.90%)	
Daily dose (mg/d)	150 (100, 150)	150 (100, 150)	150 (100, 150)	150 (150, 150)	150 (100, 150)	
Dose/weight (mg/kg/12 hours)	1.05 (0.833, 1.25)	1.00 (0.833, 1.25)	1.00 (0.833, 1.25)	1.14 (0.995, 1.36)	1.05 (0.833, 1.15	
Days of therapy	10.0 (7.00, 16.0)	10.0 (7.00, 16.0)	10.0 (7.00, 15.0)	14.0 (7.00, 22.0)	9.50 (7.00, 15.0)	
Nerosol inhalation	45 (14.4%)	23 (12.5%)	41 (17.2%)	2 (3.00%)	14 (18.4%)	
Concomitant drugs, n (%)	, ,	, ,	, ,	,	, ,	
Vasoactive drugs	151 (48.4%)	77 (41.8%)	123 (51.5%)	26 (39.4%)	38 (50.0%)	
Nephrotoxic drugs	101 (32.4%)	58 (31.5%)	74 (31.0%)	32 (48.5%)	19 (25.0%)	
Tigecycline	103 (33.0%)	59 (32.1%)	73 (30.5%)	25 (37.9%)	32 (42.1%)	
CAZ-AVI	43 (13.8%)	18 (9.80%)	32 (13.4%)	11 (16.7%)	13 (17.1%)	
Carbapenem	60 (19.2%)	30 (16.3%)	42 (17.6%)	17 (25.8%)	17 (22.4%)	
BLBLI	105 (33.7%)	64 (34.8%)	88 (36.8%)	19 (28.8%)	20 (26.3%)	
Aminoglycosides	10 (3.20%)	4 (2.20%)	8 (3.30%)	2 (3.00%)	1 (1.30%)	
Others	49 (15.7%)	29 (15.8%)	37 (15.5%)	14 (21.2%)	12 (15.8%)	

TABLE 1 Demographic characteristics of the included patients^a (Continued)

Variable	Total (n = 312)		Subgi	roups	
		Non-CRRT (n = 184)	LRTIs (n = 239)	IAIs (n = 66)	BSIs (n = 76)
None	69 (22.1%)	44 (23.9%)	56 (23.4%)	11 (16.7%)	18 (23.7%)
AUC _{ss,24 h} (mg·h/L)	76.4 (55.0, 107)	79.9 (59.6, 114)	76.5 (56.1, 106)	72.9 (52.8, 98.4)	79.7 (58.6, 109)
AUC ≥50 mg·h/L	251 (80.4%)	154 (83.7%)	194 (81.2%)	52 (78.8%)	67 (88.2%)
Laboratory data					
CRP (μg/L)	124 (74.6, 187)	111 (67.0, 158)	121 (69.5, 185)	134 (91.4, 187)	125 (83.7, 179)
PCT (ng/mL)	1.38 (0.476, 5.61)	0.830 (0.281, 2.62)	1.22 (0.422, 4.66)	1.92 (0.800, 13.1)	2.74 (0.905, 14.1)
RBC (10 ¹² /L)	2.50 (2.18, 2.92)	2.65 (2.28, 3.01)	2.55 (2.21, 2.95)	2.41 (2.00, 2.63)	2.57 (2.08, 2.96)
WBC (10 ⁹ /L)	10.1 (6.05, 15.0)	9.65 (6.48, 14.2)	10.3 (6.30, 14.6)	11.7 (6.19, 18.6)	10.6 (5.40, 16.4)
Neutrophil (10 ⁹ /L)	87.1 (78.7, 92.1)	86.5 (76.0, 91.4)	87.4 (79.0, 92.5)	89.0 (81.4, 91.8)	88.0 (74.9, 93.1)
SCr (µmol/L)	76.8 (54.0, 120)	63.5 (46.0, 95.6)	78.0 (54.0, 114)	75.0 (58.0, 122)	89.0 (60.8, 146)
BUN (mmol/L)	11.0 (6.50, 16.8)	8.97 (5.50, 13.6)	11.7 (7.20, 17.2)	9.68 (5.70, 15.3)	13.0 (6.99, 20.3)
ALB (g/L)	29.3 (26.3, 31.6)	29.3 (26.2, 31.6)	29.3 (26.6, 31.7)	29.5 (26.6, 32.5)	29.3 (25.9, 31.9)
TP (g/L)	54.6 (49.9, 60.1)	55.2 (50.1, 60.0)	54.6 (50.6, 60.1)	54.3 (49.3, 58.2)	54.9 (49.3, 60.7)
ALT (U/L)	30.0 (15.0, 60.0)	29.0 (15.0, 55.0)	30.0 (15.0, 57.3)	25.5 (14.8, 46.5)	32.0 (14.3, 73.8)
AST (U/L)	37.0 (23.5, 65.5)	34.0 (22.0, 56.0)	36.0 (24.0, 65.0)	38.5 (23.0, 56.8)	42.0 (24.0, 69.5)
ALP (U/L)	116 (80.0, 171)	102 (76.0, 162)	107 (77.5, 160)	143 (102, 203)	104 (84.0, 175)
TBIL (μmol/L)	16.1 (9.48, 39.6)	12.7 (8.70, 22.5)	15.3 (9.10, 36.4)	50.8 (16.8, 139)	18.0 (8.75, 47.9)

°OR, odds ratio; CRRT, continuous renal replacement therapy; LRTIs, lower respiratory tract infections; IAIs, intra-abdominal infections; BSIs, bloodstream infections; BMI, body mass index; COPD, chronic obstructive pulmonary disease; ICU, intensive care unit; SOFA, sequential organ failure assessment; APACHE, acute physiology and chronic health evaluation; CRKP, carbapenem-resistant *Klebsiella pneumoniae*; CRPA, carbapenem-resistant *Pseudomonas aeruginosa*; CRAB, carbapenem-resistant *Acinetobacter baumannii*; ECMO, extracorporeal membrane oxygenation; CAZ-AVI, ceftazidime-avibactam; BLBLI, beta-lactam/beta-lactamase inhibitor combinations; AUC_{55,24} h, area under the concentration-to-time curve across 24 hours at steady state; CRP; C-reactive protein; PCT, procalcitonin; RBC, red blood cell; WBC, white blood cell; SCr, serum creatinine; BUN, blood urea nitrogen; ALB, serum albumin; TP; total protein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase; TBIL, total bilirubin.

The results suggested that the infection site should be considered when dosing and monitoring polymyxin B.

The main strength of this study was the evaluation of PK/PD in different subgroups rather than the combination of patients with different characteristics. Polymyxins have significant infection site-specific PK/PD properties. A preclinical study evaluating the PK/PD relationship in *Klebsiella pneumoniae*-infected mouse models revealed that the fAUC/MIC was related to the antibacterial effect on thigh infection, and the target values of the fAUC/MIC for stasis and 1 log10 kill were 1.22–13.5 and 3.72–28.0, respectively. However, there was no relationship between the antibacterial effects of polymyxin B and the fAUC/MIC, and it was not possible to achieve stasis in lung infection, even at the highest dose tolerated by mice (15). Another study evaluating the PK/PD properties of colistin against PA and AB also reported similar results: for only some strains, it could achieve 2 log-kill in lung infection models at extremely high fAUC/MIC values (36.8–105) (20). They attributed the lack of responsiveness of lung infections to relatively low concentrations of polymyxins in the epithelial lining fluid (ELF) of mice that were systemically administered the drug.

However, recent clinical studies have reported confounding results (21, 22). Yang et al. performed a retrospective observational study and reported that an AUC of 50–100 mg·h/L was associated with decreased nephrotoxicity while ensuring clinical efficacy in critically ill patients. Tang et al. reported that the AUC/MIC was associated with polymyxin B in nosocomial pneumonia patients with CRO, and the cutoff value was 66.9 mg·h/L when polymyxin B was combined with other antibiotics (23). As we mentioned previously, these two studies included many pneumonia patients who received additional inhaled polymyxin B, which increased the concentration at the infection site and made the plasma concentration less relative (24). Thus, these results need further validation. A subsequent RCT revealed that this target AUC compliance was not correlated with clinical outcomes but with AKI only (25). It should be noted that these studies all used total plasma polymyxin B concentration to calculate the AUC due to the difficulty in determination of the concentration of unbound drugs in routine TDM

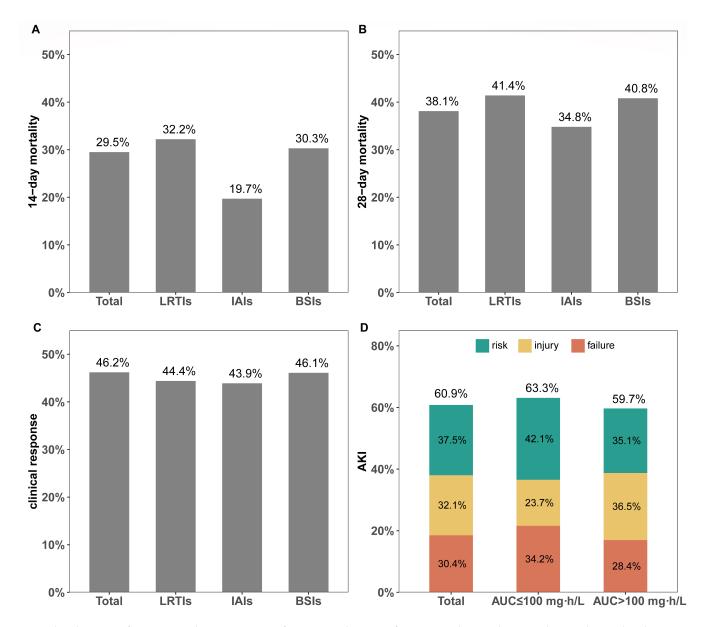


FIG 2 Clinical outcome of intravenous polymyxin B treatment for patients with CRGNB infection. (A) 14-day mortality; (B) 28-day mortality; (C) clinical response rate; (D) AKI rate. LRTI, lower respiratory tract infection; IAI, intra-abdominal infection; BSI, bloodstream infection; AKI, acute kidney injury; AUC, area under the curve of polymyxin B.

settings. Our study adequately explained the controversy of previous studies. Clinical PK/PD studies are biased by the inclusion of patients with different infection sites. Our results are in accordance with those of preclinical studies, which revealed that the plasma polymyxin B AUC was related to the clinical outcomes of some site infections other than LRTI. The results indicated that the use of reported AUC targets carefully as the infection site matters.

IAI is the only infection type whose mortality is significantly associated with the AUC of polymyxin B. It may be due to the relatively high distribution in intra-abdominal organs of this drug (26). This is a stronger evidence that supports the TDM of polymyxin B than observed in previous studies. The IAI types in the study were mainly infections after pancreatitis and post-surgery infections. Tube drainage was applied for nearly all patients; thus, we only recorded surgical removal during polymyxin B treatment in this

TABLE 2 Clinical outcomes of the included patients^a

Variable	Total (n = 312)	Subgroups				
		Non-CRRT (n = 184)	LRTIs (n = 239)	IAIs (n = 66)	BSIs (n = 76)	
Outcomes						
Clinical response	144 (46.2%)	99 (53.8%)	106 (44.4%)	29 (43.9%)	35 (46.1%)	
14-day all-cause mortality	92 (29.5%)	40 (21.7%)	77 (32.2%)	13 (19.7%)	23 (30.3%)	
28-day all-cause mortality	119 (38.1%)	51 (27.7%)	99 (41.4%)	23 (34.8%)	31 (40.8%)	
Occurrence of AKI						
No	72 (39.1%)	72 (39.1%)	51 (37.2%)	10 (31.2%)	15 (34.9%)	
Yes	112 (60.9%)	112 (60.9%)	86 (62.8%)	22 (68.8%)	28 (65.1%)	
AKI stage						
Risk	42 (37.5%)	42 (37.5%)	35 (40.7%)	5 (22.7%)	11 (39.3%)	
Injury	36 (32.1%)	36 (32.1%)	26 (30.2%)	11 (50.0%)	9 (32.1%)	
Failure	34 (30.4%)	34 (30.4%)	25 (29.1%)	6 (27.3%)	8 (28.6%)	

eCRRT, continuous renal replacement therapy; LRTIs, lower respiratory tract infections; IAIs, intra-abdominal infections; BSIs, bloodstream infections; AKI, acute kidney injury.

study. Finally, surgical removal was an independent risk factor associated with 14-day mortality, which suggested the importance of proactive source control in IAI patients.

BSI is considered an ideal disease model for PK/PD studies as the plasma concentration directly reflects the drug concentration at the infection site. Our previous study of CRKP BSI patients revealed that an AUC/MIC greater than 54.4 would benefit patients receiving polymyxin B combined with high-dose meropenem therapy (27). Survival analysis revealed that patients with an AUC less than 50 mg·h/L had increased mortality. However, the association between the AUC and 14-day mortality in patients with BSIs was not significant in this study. The reasons were that the sample size of BSI patients was limited, and most of them had comorbid pneumonia. On the basis of the available evidence, an AUC over 50 mg·h/L is a reasonable target for BSI patients.

Among all the infection sites, patients with LRTI had poorer clinical outcomes. These results were in accordance with those of previously mentioned preclinical studies and many clinical observations. A study of nosocomial pneumonia caused by MDRPs reported that the favorable clinical outcome was 47.3% in patients receiving intravenous polymyxin B, suggesting that polymyxin B is a reliable drug but can be used only as salvage therapy for nosocomial pneumonia caused by MDRPs (28). A prospective cohort study also revealed that polymyxin B treatment at the currently recommended dosage may result in inferior outcomes compared to other drugs in the treatment of VAP and VAT caused by organisms that are susceptible to this agent *in vitro* (29). Nebulized polymyxin B may be an option for LRTI patients because it can increase

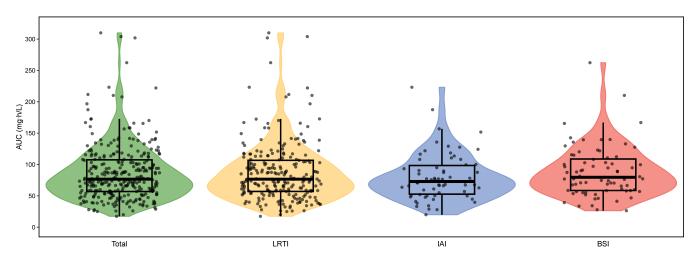


FIG 3 Distribution of polymyxin B steady-state 24-hour area under the curve in patients. LRTI, lower respiratory tract infection; IAI, intra-abdominal infection; BSI, bloodstream infection.

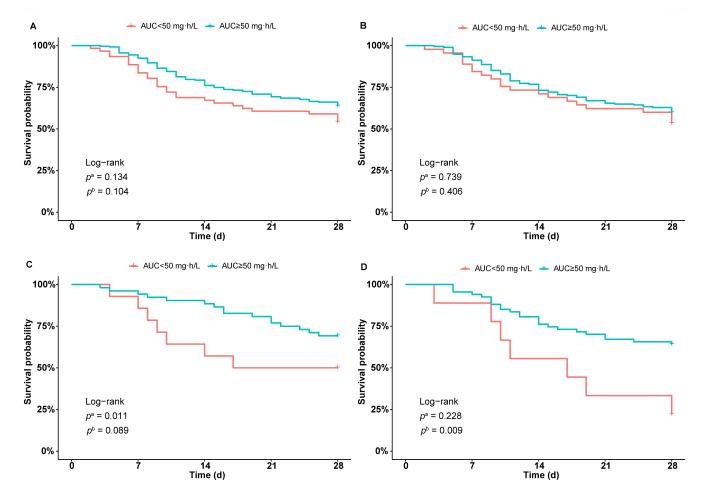


FIG 4 Survival analysis of patients with different levels of systemic polymyxin B exposure. (A) Overall patients; (B) patients with lower respiratory tract infection; (C) intra-abdominal infection; (D) bloodstream infection. AUC, area under the curve of polymyxin B. p^a and p^b indicate significant differences in 14-day and 28-day survival between the two groups, respectively.

the drug concentration at the infection site. The clinical benefit of inhaled polymyxin B is controversial in the current literature (30, 31). There are additional factors that could influence the effect of inhaled polymyxin B (32). In this multicenter study, inhaled polymyxin B did not have beneficial effects. The possible reasons may be that the protocols of inhaled polymyxin B differ among centers and that few patients have received inhaled treatment. Additional evidence is needed.

Combination with other antibiotics is a common strategy for enhancing the bacterial-killing effect and minimizing resistance emergence (33). For example, *in vitro* pharmacodynamics experiments revealed that polymyxin B and tigecycline have synergistic or additive effects on CRAB (34). The combination of antibiotics, namely, ceftazidime-avibactam and amikacin, was found to be an independent risk factor for overall 14-day mortality. Ceftazidime-avibactam is an effective and safe option for KPC-producing CRGNB, and its combined use could reduce the mortality in this study. It would be a confounding factor that makes interpretation of the results challenging. But AUC was not associated with overall mortality by excluding patients receiving ceftazidime-avibactam (Table S11). Polymyxin B combined with amikacin was reported to decrease 30-day mortality in patients with CRKP bloodstream infection (35). However, amikacin was associated with additional death incidences in our study, and the reason was unknown. The effect of combined antibiotics on the PK/PD of polymyxin B should be evaluated in future studies.

 TABLE 3
 Univariate and multivariate logistic regression analyses for 14-day mortality in all patients^a

	Survival	No survival	Univariate logistic regression	Multivariate logis	tic regression
	(n = 220)	(n = 92)	<i>P</i> -value	OR (95% CI)	<i>P</i> -value
Sex, n (%)					
Male	156 (70.9%)	65 (70.7%)	0.964		
Female	64 (29.1%)	27 (29.3%)			
Age (years)	68.0 (57.0, 78.0)	73.5 (61.0, 82.0)	0.063		0.660
Weight (kg)	60.5 (55.0, 70.0)	61.5 (51.4, 70.0)	0.407		
BMI (kg/m²)	22.0 (20.0, 24.8)	22.5 (19.5, 25.8)	0.277		
Comorbidities, n (%)					
Dementia	10 (4.50%)	4 (4.30%)	0.939		
Diabetes	53 (24.1%)	32 (34.8%)	0.054		0.584
CKD	23 (10.5%)	12 (13.0%)	0.510		
Malignancy	55 (25.0%)	20 (21.7%)	0.539		
Heart failure	26 (11.8%)	14 (15.2%)	0.414		
COPD	23 (10.5%)	7 (7.60%)	0.439		
Immunodeficiency	17 (7.70%)	3 (3.30%)	0.154		0.201
Hypertension	101 (45.9%)	44 (47.8%)	0.757		
Hyperlipidemia	6 (2.70%)	3 (3.30%)	0.798		
Acute pancreatitis	30 (13.6%)	7 (7.60%)	0.139		
COVID-19	17 (7.70%)	21 (22.8%)	<i>P</i> < 0.001	3.90 (1.70-8.97)	0.001
ICU admission	199 (90.5%)	91 (98.9%)	0.028		0.480
Severely ill	105 (47.7%)	55 (59.8%)	0.053		
Infected site, n (%)					
LRTIs	162 (73.6%)	77 (83.7%)	0.058		
IAIs	53 (24.1%)	13 (14.1%)	0.052		0.185
BSIs	53 (24.1%)	23 (25.0%)	0.865		
Other or undefined	24 (10.9%)	12 (13.0%)	0.591		
Pathogen, n (%)					
CRKP	68 (30.9%)	18 (19.6%)	0.043		0.368
CRPA	47 (21.4%)	18 (19.6%)	0.721		
CRAB	90 (40.9%)	51 (55.4%)	0.019		0.302
others	104 (47.3%)	33 (35.9%)	0.065		
CRRT	76 (34.5%)	52 (56.5%)	<i>P</i> < 0.001	2.65 (1.41-4.99)	0.002
ECMO	16 (7.30%)	7 (7.60%)	0.917		
Daily dose (mg/d)	150 (100, 150)	150 (100, 150)	0.630		
Dose/weight (mg/kg/12 hours)	1.05 (0.833, 1.25)	1.06 (0.833, 1.28)	0.909		
Aerosol inhalation	34 (15.5%)	11 (12.0%)	0.424		
Concomitant drugs, n (%)					
Tigecycline	70 (31.8%)	33 (35.9%)	0.488		
CAZ-AVI	36 (16.4%)	7 (7.60%)	0.046	0.179 (0.057-0.562)	0.003
Carbapenem	42 (19.1%)	18 (19.6%)	0.923		
BLBLI	73 (33.2%)	32 (34.8%)	0.785		
Aminoglycosides	5 (2.30%)	5 (5.40%)	0.161	6.27 (1.42-27.7)	0.016
Other	40 (18.2%)	9 (9.80%)	0.067		
None	51 (23.2%)	18 (19.6%)	0.483		
AUC _{ss,24 h} (mg·h/L)	76.6 (57.3, 108)	76.3 (51.8, 103)	0.976		
AUC ≥50 mg·h/L	181 (82.3%)	70 (76.1%)	0.211		
Laboratory data					
CRP (µg/L)	121 (73.1, 184)	131 (77.5, 193)	0.516		
PCT (ng/mL)	1.19 (0.402, 5.91)	1.88 (0.613, 5.44)	0.503		
RBC (10 ¹² /L)	2.51 (2.19, 2.95)	2.48 (2.16, 2.89)	0.366		
WBC (10 ⁹ /L)	9.60 (5.90, 14.2)	11.4 (6.60, 17.2)	0.024		0.063
Neutrophil (10 ⁹ /L)	85.8 (76.6, 91.5)	90.0 (83.5, 93.4)	0.046		

TABLE 3 Univariate and multivariate logistic regression analyses for 14-day mortality in all patients^a (Continued)

	Survival	No survival	Univariate logistic regression	Multivariate log	istic regression
	(n = 220)	(n = 92)	<i>P</i> -value	OR (95% CI)	<i>P</i> -value
SCr (µmol/L)	72.0 (53.0, 110)	96.3 (61.5, 152)	0.018		
BUN (mmol/L)	9.54 (6.18, 15.1)	13.6 (8.76, 22.8)	P < 0.001	1.04 (1.02-1.08)	0.003
ALB (g/L)	29.5 (26.9, 32.0)	28.0 (25.7, 30.7)	0.006		0.076
TP (g/L)	55.6 (51.2, 60.8)	53.2 (47.7, 57.9)	0.014		0.874
ALT (U/L)	29.0 (15.0, 60.0)	31.0 (14.0, 58.8)	0.412		
AST (U/L)	34.0 (21.0, 64.0)	40.0 (28.0, 69.5)	0.585		
ALP (U/L)	120 (82.3, 177)	104 (69.0, 151)	0.124		0.120
TBIL (µmol/L)	15.4 (9.83, 37.9)	17.9 (9.15, 40.2)	0.390		
AKI	88 (61.1%)	24 (60.0%)	0.899		

°OR, odds ratio; BMI, body mass index; CKD, chronic kidney disease; COPD, chronic obstructive pulmonary disease; ICU, intensive care unit; LRTIs, lower respiratory tract infections; IAIs, intra-abdominal infections; BSIs, bloodstream infections; CRKP, carbapenem-resistant *Klebsiella pneumoniae*; CRPA, carbapenem-resistant *Pseudomonas aeruginosa*; CRAB, carbapenem-resistant *Acinetobacter baumannii*; CRRT, continuous renal replacement therapy; ECMO, extracorporeal membrane oxygenation; CAZ-AVI, ceftazidime/avibactam; BLBLI, beta-lactam-beta-lactamase inhibitor combinations; AUC_{55,24} h, area under the concentration-to-time curve across 24 hours at steady state; CRP; C-reactive protein; PCT, procalcitonin; RBC, red blood cell; WBC, white blood cell; SCr, serum creatinine; BUN, blood urea nitrogen; ALB, serum albumin; TP; total protein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase; TBIL, total bilirubin; AKI, acute kidney injury.

CRRT was a risk factor associated with 14-day mortality in both overall patients and IAI patients. Previously, polymyxin B was thought to eliminate mainly the non-kidney pathway, and the dose does not need to be adjusted in patients with CRRT and renal insufficiency (16). However, recent studies reported that CRRT was a covariate in the PPK model and suggested that the dose used was slightly higher (36–38). But the AUCs of the CRRT patients were not lower than those of the other patients in this study. CRRT itself indicates kidney injury, and organ failure is a risk factor for mortality (39). Special attention should be given to these patients to improve their clinical outcomes.

Nephrotoxicity is a well-known dose-dependent toxicity of polymyxin B (40). Previous studies suggested that an AUC of 100 mg·h/L was the upper limit for polymyxin B exposure (17, 41). In contrast, we did not find a significant difference between patients with AUCs above or below 100 mg·h/L or the association between AKI and polymyxin B exposure in our study. As in previous studies, we have not analyzed the causality of polymyxin B use and AKI. Many factors in CRGNB-infected patients, such as septic shock, contribute to AKI (42). These factors interfere with the PK/PD analysis of polymyxin B, and future analysis on polymyxin B-induced AKI patients would be helpful. Moreover, the dose range of polymyxin B was narrow, which resulted in a narrow AUC distribution (averages: 77.2 vs 83.0 mg·h/L; range: 57.9 to 114 mg·h/L). The percentages of patients having an AUC over 100 mg·h/L were similar in both groups (Table 4). These issues may contribute to the negative finding in the exposure–nephrotoxicity relationship in this study.

There are several limitations to this study. The sample size for specific infection site patients was insufficient, and that may be the reason that the difference in the mortality among BSI patients with different AUCs was insignificant. The unbound polymyxin B in plasma samples was not determined. The influence of various confounding factors, such as the efficacy of combined antibiotics, needs to be verified in future studies. Polymyxin B MICs of isolated strains were not tested. The upper limit of polymyxin B exposure should be analyzed in those with drug-associated AKI, not all AKI patients.

Conclusion

We performed a large prospective observational multicenter study of polymyxin B for treating various site CRGNB infections. The clinical outcomes of polymyxin B, as well as nephrotoxicity, were provided. PK/PD analysis indicated that the AUC of polymyxin B was not associated with mortality in LRTI patients but was associated with that in IAI and BSI patients. A new AUC cutoff of polymyxin B was provided for IAI patients. These results would benefit personalized dosing and monitoring of this drug.

 $\textbf{TABLE 4} \quad \textbf{Univariate and multivariate logistic regression analyses of 14-day mortality in intra-abdominal infection patients}^a$

	Survival	No survival	Univariate logistic regression	Multivariate logis	tic regression
	(n = 53)	(n = 13)	P-value	OR (95% CI)	<i>P</i> -value
Sex, n (%)					
Male	33 (62.3%)	8 (61.5%)	0.961		
Female	20 (37.7%)	5 (38.5%)			
Age (years)	59.0 (53.0, 69.0)	63.0 (49.0, 71.0)	0.923		
Weight (kg)	63.0 (55.0, 70.0)	60.0 (55.0, 70.0)	0.137		0.826
BMI (kg/m²)	22.8 (20.6, 25.2)	23.3 (21.1, 24.8)	0.123		
Comorbidities, n (%)					
Diabetes	6 (11.3%)	2 (15.4%)	0.689		
CKD	4 (7.50%)	1 (7.70%)	0.986		
Malignancy	11 (20.8%)	2 (15.4%)	0.664		
Heart failure	1 (1.90%)	0 (0.00%)	0.992		
Immunodeficiency	3 (5.70%)	0 (0.00%)	0.994		
Hypertension	17 (32.1%)	4 (30.8%)	0.928		
Hyperlipidemia	3 (5.70%)	2 (15.4%)	0.254		
Acute pancreatitis	26 (49.1%)	5 (38.5%)	0.495		
COVID-19	5 (9.40%)	0 (0.00%)	0.993		
CU admission	52 (98.1%)	13 (100%)	0.992		
Severely ill	22 (41.5%)	10 (76.9%)	0.030		
Surgical removal of infection	25 (47.2%)	2 (15.4%)	0.027	0.004 (0.001–0.381)	0.017
nfected site, n (%)	23 (17.270)	2 (13.170)	0.027	0.001 (0.001 0.501)	0.017
LRTIs	21 (39.6%)	7 (53.8%)	0.356		
BSIs	9 (17.0%)	2 (15.4%)	0.890		
Other or undefined	5 (9.40%)	2 (15.4%)	0.536		
Pathogen, n (%)	3 (3.4070)	2 (13.470)	0.550		
CRKP	26 (49.1%)	8 (61.5%)	0.422		
CRPA	10 (18.9%)	5 (38.5%)	0.140		0.513
CRAB	22 (41.5%)	8 (61.5%)	0.200		0.515
Others	25 (47.2%)	3 (23.1%)	0.126		
CRRT	22 (41.5%)	12 (92.3%)	0.009		0.232
ECMO	0 (0.00%)	1 (7.70%)	0.991		0.232
		,			
Daily dose (mg/d)	150 (100, 150)	150 (150, 150)	0.197		
Dose/weight (mg/kg/12 hours) Aerosol inhalation		1.07 (0.833, 1.32)	0.338 0.993		
	2 (3.80%)	0 (0.00%)	0.993		
Concomitant drugs, n (%)	10 (25 00/)	C (4C 20/)	0.404		
Tigecycline	19 (35.8%)	6 (46.2%)	0.494		
CAZ-AVI	10 (18.9%)	1 (7.70%)	0.350		
Carbapenem	12 (22.6%)	5 (38.5%)	0.249		
BLBLI	17 (32.1%)	2 (15.4%)	0.246		
Aminoglycosides	2 (3.80%)	0 (0.00%)	0.993		
Others	10 (18.9%)	4 (30.8%)	0.352		
None	10 (18.9%)	1 (7.70%)	0.350		
AUC _{ss,24 h} (mg·h/L)	77.2 (62.8, 108)	50.5 (41.1, 66.6)	0.014	0.936 (0.886–0.989)	0.018
AUC ≥50 mg·h/L	45 (84.9%)	7 (53.8%)	0.020		
Laboratory data					
CRP (µg/L)	130 (87.2, 186)	147 (91.6, 217)	0.409		
PCT (ng/mL)	1.89 (0.800, 10.5)	2.62 (1.34, 14.0)	0.651		
RBC (10 ¹² /L)	2.29 (1.98, 2.67)	2.44 (2.21, 2.48)	0.698		
WBC (10 ⁹ /L)	11.2 (6.12, 16.4)	17.3 (8.60, 23.7)	0.092	1.144 (1.017–1.287)	0.026
Neutrophil (10 ⁹ /L)	89.0 (81.3, 92.6)	89.0 (82.6, 90.3)	0.934		
SCr (µmol/L)	73.0 (58.0, 116)	89.0 (62.0, 136)	0.816		
BUN (mmol/L)	9.00 (5.68, 15.5)	11.7 (6.20, 14.4)	0.976		

TABLE 4 Univariate and multivariate logistic regression analyses of 14-day mortality in intra-abdominal infection patients^a (Continued)

	Survival	No survival	Univariate logistic regression	Multivariate lo	gistic regression
	(n = 53)	(<i>n</i> = 13)	<i>P</i> -value	OR (95% CI)	<i>P</i> -value
ALB (g/L)	29.4 (26.5, 32.5)	29.7 (26.7, 32.5)	0.738		
TP (g/L)	54.6 (49.3, 58.0)	53.3 (50.5, 62.0)	0.818		
ALT (U/L)	24.0 (14.5, 44.5)	45.0 (22.0, 57.0)	0.629		
AST (U/L)	35.0 (22.0, 55.0)	48.0 (37.0, 69.0)	0.538		
ALP (U/L)	141 (104, 202)	143 (92.0, 204)	0.729		
TBIL (µmol/L)	38.3 (15.4, 126)	71.3 (31.0, 162)	0.549		
AKI	21 (67.7%)	1 (100.0%)	0.998		

°OR, odds ratio; BMI, body mass index; CKD, chronic kidney disease; ICU, intensive care unit; LRTIs, lower respiratory tract infections; BSIs, bloodstream infections; CRKP, carbapenem-resistant *Klebsiella pneumoniae*; CRPA, carbapenem-resistant *Pseudomonas aeruginosa*; CRAB, carbapenem-resistant *Acinetobacter baumannii*; CRRT, continuous renal replacement therapy; ECMO, extracorporeal membrane oxygenation; CAZ-AVI, ceftazidime/avibactam; BLBLI, beta-lactam-beta-lactamase inhibitor combinations; AUC_{sS,24} h, area under the concentration-to-time curve across 24 h at steady state; CRP; C-reactive protein; PCT, procalcitonin; RBC, red blood cell; WBC, white blood cell; SCr, serum creatinine; BUN, blood urea nitrogen; ALB, serum albumin; TP; total protein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase; TBIL, total bilirubin; AKI, acute kidney injury. Only surgical removal of infection during polymyxin B treatment was recorded.

MATERIALS AND METHODS

Study design and patient enrollment

This study was designed as a prospective, observational, multicenter study and was conducted in accordance with the guidelines of the Declaration of Helsinki. Ethics approval was obtained from the Ethics Committee of Sir Run Run Shaw Hospital, Zhejiang University School of Medicine (reference number 20220124-30). The study was registered in the Chinese Clinical Trial Registry (https://www.chictr.org.cn/) with the number ChiCTR2200056667 (registered 10 February 2022). Patients who met the following criteria were included: (i) identified or highly suspected infection caused by carbapenem-resistant gram-negative bacteria; (ii) received polymyxin B treatment for >72 hours; and (iii) were willing and able to collect plasma samples for concentration determination. The exclusion criteria were as follows: (i) patients aged <18 years; (ii) patients receiving nonintravenous polymyxin B only; and (iii) patients whose peak and trough concentrations were not obtained. Patients who received inhaled CMS or colistin sulfate were excluded because polymyxin B concentration determination was influenced as polymyxin E2 was used as internal standard. However, inhaled polymyxin B does not affect the determination of the plasma concentration, and these patients were included. Written informed consent was obtained from the patients before enrollment according to the institution's requirements.

Drug administration, sampling, concentration determination, and AUC calculation

Owing to the observational nature of the study, the dosing of polymyxin B and the regimen of combined antibiotics were not implemented and were decided by doctors. Two blood samples (peak and trough concentrations) were collected after four or more doses of intravenous polymyxin B. Plasma was isolated by centrifugation and stored at -80° C before determination. The polymyxin B concentrations, which were obtained by adding polymyxin B1 (including B1-I) and B2 concentrations, were determined via a previously established LC-MS/MS method (43). The 24-hour steady-state AUC was calculated via the first-order elimination-based equation method, which is recommended by the Chinese consensus guideline for polymyxin B therapeutic drug monitoring (44).

Follow-up and clinical outcomes

The patients were followed for up to 28 days, and the clinical evaluation was performed by two researchers. The primary outcomes were 14-day mortality and nephrotoxicity. The secondary outcomes were the clinical response rate and 28-day mortality. Clinical

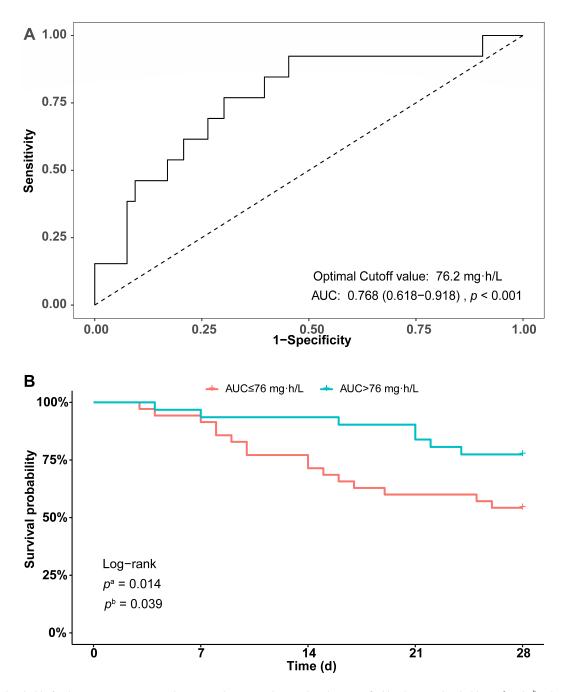


FIG 5 AUC threshold of polymyxin B in IAI patients by ROC analysis (A) and survival analysis stratified by the AUC threshold (B). p^a and p^b indicate significant differences in 14-day and 28-day survival between the two groups, respectively.

response was defined as the disappearance or improvement in signs and biochemical indicators of infection, which was evaluated at the end of treatment or on the 14th day of treatment if the duration of polymyxin B treatment was longer than 14 days. Nephrotoxicity was evaluated by the occurrence of AKI, which was evaluated based on changes in serum creatinine and the eGFR estimated via the Cockcroft–Gault equation during polymyxin B treatment (45). The severity of AKI was graded according to the RIFLE criteria (46). Patients who started continuous renal replacement therapy (CRRT) during polymyxin B treatment were defined as having F-AKI. Given that the treatment durations of the enrolled patients were longer than 3 days, all patients were evaluated for mortality and clinical response. Patients receiving CCRT before polymyxin B treatment

 ${\bf TABLE~5} \quad {\bf Univariate~and~multivariate~logistic~regression~analyses~for~AKIa}$

	Without AKI $(n = 72)$	AKI (n = 112)	Univariate logistic regression	Multivariate logistic regression	
			P-value	OR (95% CI)	<i>P</i> -value
Sex, n (%)					
Male	45 (62.5%)	83 (74.1%)	0.096	0.468 (0.228-0.960)	0.038
Female	27 (37.5%)	29 (25.9%)			
Age (years)	68.0 (56.0, 78.0)	70.0 (60.0, 80.0)	0.310		
Weight (kg)	60.0 (55.0, 68.0)	62.3 (53.3, 70.0)	0.615		
BMI (kg/m²)	21.6 (19.6, 24.4)	22.4 (19.6, 25.4)	0.573		
Comorbidities, n (%)					
Dementia	2 (2.80%)	5 (4.50%)	0.563		
Diabetes	21 (29.2%)	29 (25.9%)	0.626		
CKD	3 (4.20%)	7 (6.30%)	0.546		
Malignancy	18 (25.0%)	34 (30.4%)	0.431		
Heart failure	9 (12.5%)	11 (9.80%)	0.570		
COPD	10 (13.9%)	10 (8.90%)	0.295		
Immunodeficiency	2 (2.80%)	4 (3.60%)	0.768		
Hypertension	35 (48.6%)	41 (36.6%)	0.108	0.446 (0.225–0.884)	0.021
Hyperlipidemia	1 (1.40%)	5 (4.50%)	0.278	0.440 (0.223 0.004)	0.021
Acute pancreatitis	6 (8.30%)	15 (13.4%)	0.296		
COVID-19	7 (9.70%)	14 (12.5%)	0.564		
CU admission	62 (86.1%)	100 (89.3%)	0.518		
			0.903		
Severely ill	27 (37.5%)	43 (38.4%)	0.903		
nfected site, n (%)	F1 (70 00/)	06 (76 00/)	0.267		
LRTIs	51 (70.8%)	86 (76.8%)	0.367		
IAIs	10 (13.9%)	22 (19.6%)	0.317		
BSIs	15 (20.8%)	27 (24.1%)	0.606		
Other or undefined	7 (9.70%)	14 (12.5%)	0.564		
Pathogen, n (%)	47 (00 40)	2.4 (2.2.40()			
CRKP	17 (23.6%)	34 (30.4%)	0.320		
CRPA	10 (13.9%)	27 (24.1%)	0.095	3.50 (1.38–8.92)	0.009
CRAB	22 (30.6%)	51 (45.5%)	0.044	3.09 (1.47–6.47)	0.003
Others	38 (52.8%)	46 (41.1%)	0.121		
CRRT					
ECMO	1 (1.40%)	6 (5.40%)	0.202		
Daily dose (mg/d)	100 (100, 150)	150 (100, 150)	0.808		
Oose/weight	1.00 (0.833, 1.36)	1.00 (0.833, 1.24)	0.606		
(mg/kg/12 hours)					
Aerosol inhalation	10 (13.9%)	13 (11.6%)	0.648		
Concomitant drugs, n (%)					
Tigecycline	13 (18.1%)	46 (41.1%)	0.001	4.14 (1.89–9.08)	< 0.001
CAZ-AVI	6 (8.30%)	12 (10.7%)	0.597		
Carbapenem	11 (15.3%)	19 (17.0%)	0.763		
BLBLI	28 (38.9%)	36 (32.1%)	0.349		
Aminoglycosides	1 (1.40%)	3 (2.70%)	0.565		
Others	12 (16.7%)	17 (15.2%)	0.787		
None	21 (29.2%)	23 (20.5%)	0.182		
AUC _{ss,24 h} (mg·h/L)	77.2 (57.9, 114)	82.0 (63.9, 113)	0.348		
AUC >100 mg·h/L	50 (69.4%)	74 (66.1%)	0.634		
aboratory data		•			
CRP (μg/L)	108 (54.9, 156)	111 (73.8, 161)	0.541		
PCT (ng/mL)	0.870 (0.246, 1.90)	0.810 (0.305, 2.69)	0.955		
RBC (10 ¹² /L)	2.63 (2.31, 3.02)	2.65 (2.27, 2.97)	0.826		
WBC (10° /L)	9.41 (6.28, 12.8)	10.1 (6.58, 14.6)	0.261		

TABLE 5 Univariate and multivariate logistic regression analyses for AKI^a (Continued)

	Without AKI (n = 72)	AKI (n = 112)	Univariate logistic regression	Multivariate lo	gistic regression
			<i>P</i> -value	OR (95% CI)	<i>P</i> -value
Neutrophil (10 ⁹ /L)	86.5 (77.1, 91.5)	86.5 (75.4, 91.4)	0.966		
SCr (µmol/L)	64.0 (53.7, 98.1)	62.5 (44.0, 92.8)	0.293		
BUN (mmol/L)	8.38 (5.20, 14.2)	9.89 (5.74, 13.6)	0.615		
ALB (g/L)	30.5 (26.6, 31.6)	28.4 (26.0, 31.5)	0.334		
TP (g/L)	56.3 (49.4, 63.2)	54.0 (50.6, 58.8)	0.090		0.322
ALT (U/L)	30.0 (17.3, 56.0)	28.0 (15.0, 53.5)	0.416		
AST (U/L)	35.0 (24.0, 64.5)	34.0 (20.5, 53.5)	0.499		
ALP (U/L)	96.0 (71.5, 195)	110 (78.0, 154)	0.275		
TBIL (µmol/L)	11.1 (7.83, 19.1)	15.0 (9.25, 22.9)	0.652		

°OR, odds ratio; BMI, body mass index; CKD, chronic kidney disease; COPD, chronic obstructive pulmonary disease; ICU, intensive care unit; LRTIs, lower respiratory tract infections; IAIs, intra-abdominal infections; BSIs, bloodstream infections; CRKP, carbapenem-resistant *Klebsiella pneumoniae*; CRPA, carbapenem-resistant *Pseudomonas aeruginosa*; CRAB, carbapenem-resistant *Acinetobacter baumannii*; CRRT, continuous renal replacement therapy; ECMO, extracorporeal membrane oxygenation; CAZ-AVI, ceftazidime/avibactam; BLBLI, beta-lactam-beta-lactamase inhibitor combinations; AUC_{55,24} h, area under the concentration-to-time curve across 24 hours at steady state; CRP; C-reactive protein; PCT, procalcitonin; RBC, red blood cell; WBC, white blood cell; SCr, serum creatinine; BUN, blood urea nitrogen; ALB, serum albumin; TP; total protein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase; TBIL, total bilirubin.

were excluded from nephrotoxicity evaluation because of the difficulty in evaluating AKI with RIFLE.

PK/PD and statistical analysis

To explore the association between polymyxin B exposure (AUC) and clinical outcomes, Kaplan–Meier analysis was performed for each outcome by stratifying patients according to the preset AUC threshold. For mortality and clinical response, the AUC threshold was 50 mg·h/L, whereas for nephrotoxicity, the threshold was 100 mg·h/L. Subgroup analysis was carried out based on the infection site, including LRTI, intra-abdominal infection (IAI), and bloodstream infection (BSI). Data are presented as the means and standard errors or medians and interquartile ranges for continuous variables and as frequencies (percentages) for categorical variables. Student's t test and the Mann–Whitney test were used for comparison of normally and nonnormally distributed continuous variables, respectively. The χ test or Fisher's exact test was used for comparison of categorical variables. Multivariate logistic regression was used to identify any independent variables associated with primary and secondary outcomes, which included variables with P < 0.2 in the univariate analysis. If the AUC was an independent variable, ROC analysis was performed. P < 0.05 was considered statistically significant.

ACKNOWLEDGMENTS

We thank all collaborators for their help in patient enrollment and sample and data collection.

This work was supported by the Development Center for Medical Science and Technology National Health Commission of China (WKZX2023CX200004), the Zhejiang Natural and Science Foundation of China (LYY21H310006), the Zhejiang Pharmaceutical Association (2021ZYY02) and the Zhejiang Medical Doctors Association (YS2022-2-002), and Hangzhou Biomedicine and Health Industry Development Project (2023WJC091).

Conceptualization, J.Z. and Y.Y.; data curation, X.L., S.L., C.X., L.Z., Y.G., J.Z., Y.Y., G.L., A.W., Y.S., X.D., and F.G.; formal analysis, Z.Y., X.L., and J.L.; funding acquisition, Z.Y. and A.W.; investigation, Z.Y., X.L., J.L., and L.Y.; methodology, Z.Y., X.L., and J.L.; project administration, J.Z. and Y.Y.; resources, J.Z. and Y.Y.; software, Z.Y.; supervision, J.Z. and Y.Y.; validation, Z.Y., X.L., and J.L.; visualization, Z.Y. and J.L.; roles/writing-original draft, Z.Y., H.H., and J.L.; writing - review & editing, J.Z. and Y.Y.

AUTHOR AFFILIATIONS

¹Sir Run Run Shaw Hospital, School of Medicine, Zhejiang University, Hangzhou, China ²Center for General Practice Medicine, Department of Infectious Diseases, Zhejiang Provincial People's Hospital (Affiliated People's Hospital, Hangzhou Medical College), Hangzhou, China

³Institute of Antibiotics, National Clinical Research Center for Aging and Medicine, Huashan Hospital, Fudan University, Shanghai, China

⁴Key Laboratory of Clinical Pharmacology of Antibiotics, Shanghai, China

⁵Second Affiliated Hospital, School of Medicine, Zhejiang University, Hangzhou, China

⁶Department of Intensive Care Unit, Hangzhou Red-Cross Hospital, Hangzhou, China

⁷Tongde Hospital of Zhejiang Province, Hangzhou, China

⁸The First Affiliated Hospital of Ningbo University, Ningbo, China

⁹The First Affiliated Hospital of Zhejiang Chinese Medical University, Hangzhou, China

¹⁰Shulan (Hangzhou) Hospital, Hangzhou, China

¹¹Zhejiang Hospital, Hangzhou, China

AUTHOR ORCIDs

Zhenwei Yu http://orcid.org/0000-0002-3776-2290

Xiaofen Liu http://orcid.org/0000-0002-5526-1502

Xiaoxing Du https://orcid.org/0000-0003-3608-195X

Jing Zhang http://orcid.org/0000-0003-2966-9149

Yunsong Yu http://orcid.org/0000-0003-2903-918X

FUNDING

Funder	Grant(s)	Author(s)
National Health Commission of the People's Republic of China (NHC)	WKZX2023CX200004	Zhenwei Yu
MOST NSFC NSFC-Zhejiang Joint Fund 浙江省科学技术厅 Natural Science Foundation of Zhejiang Province (ZJNSF)	LYY21H310006	Zhenwei Yu
Zhejiang Pharmaceutical Association (ZIPA)	2021ZYY02	Zhenwei Yu
Zhejiang Medical Doctor Association	YS2022-2-002	Zhenwei Yu
Hangzhou Science and Technology Bureau	2023WJC091	Anqi Wei

AUTHOR CONTRIBUTIONS

Zhenwei Yu, Data curation, Funding acquisition, Investigation, Methodology, Software, Validation, Visualization, Writing – original draft | Huangdu Hu, Writing – original draft | Xiaofen Liu, Formal analysis, Investigation, Methodology, Validation | Jieqiong Liu, Formal analysis, Investigation, Methodology, Validation, Visualization, Writing – original draft | Lingyan Yu, Investigation | Anqi Wei, Data curation, Funding acquisition | Chuanwei Xin, Data curation | Yongxiong Gan, Data curation | Shu Lei, Data curation | Li Zhuang, Data curation | Yanfei Shen, Data curation | Xiaoxing Du, Data curation | Jianping Zhu, Data curation | Yi Yang, Data curation | Gang Liang, Data curation | Feng Guo, Data curation | Jing Zhang, Conceptualization, Project administration, Supervision, Writing – review and editing | Yunsong Yu, Conceptualization, Project administration, Supervision, Writing – review and editing

DATA AVAILABILITY

The data sets used and analyzed during the current study are available from the corresponding author on reasonable request.

ETHICS APPROVAL

Ethics approval was obtained from the Ethics Committee of Sir Run Run Shaw Hospital, Zhejiang University School of Medicine (reference number 20220124-30). Written informed consent was obtained from the patients before enrollment according to the institution's requirements.

ADDITIONAL FILES

The following material is available online.

Supplemental Material

Tables S1 to S11 (AAC01859-24-s0001.docx). Multivariate logistic analysis for secondary outcomes and other infection types.

REFERENCES

- Mills JP, Marchaim D. 2021. Multidrug-resistant Gram-negative bacteria: infection prevention and control update. Infect Dis Clin North Am 35:969–994. https://doi.org/10.1016/j.idc.2021.08.001
- Reyes J, Komarow L, Chen L, Ge L, Hanson BM, Cober E, Herc E, Alenazi T, Kaye KS, Garcia-Diaz J, et al. 2023. Global epidemiology and clinical outcomes of carbapenem-resistant *Pseudomonas aeruginosa* and associated carbapenemases (POP): a prospective cohort study. Lancet Microbe 4:e159–e170. https://doi.org/10.1016/S2666-5247(22)00329-9
- Babiker A, Clarke LG, Saul M, Gealey JA, Clancy CJ, Nguyen MH, Shields RK. 2021. Changing epidemiology and decreased mortality associated with carbapenem-resistant Gram-negative bacteria, 2000–2017. Clin Infect Dis 73:e4521–e4530. https://doi.org/10.1093/cid/ciaa1464
- Li R, Tang H, Xu H, Cui K, Li S, Shen J. 2021. Effect on 30-day mortality and duration of hospitalization of empirical antibiotic therapy in CRGNBinfected pneumonia. Ann Clin Microbiol Antimicrob 20:15. https://doi.or g/10.1186/s12941-021-00421-2
- Murray CJL, Ikuta KS, Sharara F, Swetschinski L, Robles Aguilar G, Gray A, Han C, Bisignano C, Rao P, Wool E, et al. 2022. Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. Lancet 399:629– 655. https://doi.org/10.1016/S0140-6736(21)02724-0
- Jean SS, Harnod D, Hsueh PR. 2022. Global threat of carbapenemresistant Gram-negative bacteria. Front Cell Infect Microbiol 12:823684. https://doi.org/10.3389/fcimb.2022.823684
- Kassamali Z, Jain R, Danziger LH. 2015. An update on the arsenal for multidrug-resistant *Acinetobacter* infections: polymyxin antibiotics. Int J Infect Dis 30:125–132. https://doi.org/10.1016/j.ijid.2014.10.014
- Yang Q, Pogue JM, Li Z, Nation RL, Kaye KS, Li J. 2020. Agents of last resort: an update on polymyxin resistance. Infect Dis Clin North Am 34:723–750. https://doi.org/10.1016/j.idc.2020.08.003
- Rodríguez-Baño J, Gutiérrez-Gutiérrez B, Machuca I, Pascual A. 2018. Treatment of infections caused by extended-spectrum-beta-lactamase-, AmpC-, and carbapenemase-producing *Enterobacteriaceae*. Clin Microbiol Rev 31:e00079-17. https://doi.org/10.1128/CMR.00079-17
- Nation RL, Velkov T, Li J. 2014. Colistin and polymyxin B: peas in a pod, or chalk and cheese? Clin Infect Dis 59:88–94. https://doi.org/10.1093/cid/c in 213
- Tran TB, Velkov T, Nation RL, Forrest A, Tsuji BT, Bergen PJ, Li J. 2016. Pharmacokinetics/pharmacodynamics of colistin and polymyxin B: are we there yet? Int J Antimicrob Agents 48:592–597. https://doi.org/10.101 6/i.iiantimicag.2016.09.010
- Roch M, Sierra R, Andrey DO. 2023. Antibiotic heteroresistance in ESKAPE pathogens, from bench to bedside. Clin Microbiol Infect 29:320–325. htt ps://doi.org/10.1016/j.cmi.2022.10.018
- Liu X, Chen Y, Yang H, Li J, Yu J, Yu Z, Cao G, Wu X, Wang Y, Wu H, Fan Y, Wang J, Wu J, Jin Y, Guo B, Hu J, Bian X, Li X, Zhang J. 2021. Acute toxicity is a dose-limiting factor for intravenous polymyxin B: a safety and pharmacokinetic study in healthy Chinese subjects. J Infect 82:207–215. https://doi.org/10.1016/j.jinf.2021.01.006
- Azad MAK, Nation RL, Velkov T, Li J. 2019. Mechanisms of polymyxininduced nephrotoxicity. Adv Exp Med Biol 1145:305–319. https://doi.org/10.1007/978-3-030-16373-0_18
- Landersdorfer CB, Wang J, Wirth V, Chen K, Kaye KS, Tsuji BT, Li J, Nation RL. 2018. Pharmacokinetics/pharmacodynamics of systemically

- administered polymyxin B against *Klebsiella pneumoniae* in mouse thigh and lung infection models. J Antimicrob Chemother 73:462–468. https://doi.org/10.1093/jac/dkx409
- 16. Tsuji BT, Pogue JM, Zavascki AP, Paul M, Daikos GL, Forrest A, Giacobbe DR, Viscoli C, Giamarellou H, Karaiskos I, Kaye D, Mouton JW, Tam VH, Thamlikitkul V, Wunderink RG, Li J, Nation RL, Kaye KS. 2019. International consensus guidelines for the optimal use of the polymyxins: endorsed by the American College of Clinical Pharmacy (ACCP), European Society of Clinical Microbiology and Infectious Diseases (ESCMID), Infectious Diseases Society of America (IDSA), International Society for Anti-infective Pharmacology (ISAP), Society of Critical Care Medicine (SCCM), and Society of Infectious Diseases Pharmacists (SIDP). Pharmacotherapy 39:10–39. https://doi.org/10.1002/phar.2209
- Yang J, Liu S, Lu J, Sun T, Wang P, Zhang X. 2022. An area under the concentration–time curve threshold as a predictor of efficacy and nephrotoxicity for individualizing polymyxin B dosing in patients with carbapenem-resistant gram-negative bacteria. Crit Care 26:320. https://doi.org/10.1186/s13054-022-04195-7
- Chen H, Zhang Z, Yu Z. 2024. How can polymyxin B be dosed based on current pharmacokinetic knowledge? Eur J Clin Pharmacol 80:1421– 1423. https://doi.org/10.1007/s00228-024-03708-3
- Nation RL, Li J, Cars O, Couet W, Dudley MN, Kaye KS, Mouton JW, Paterson DL, Tam VH, Theuretzbacher U, Tsuji BT, Turnidge JD. 2015. Framework for optimisation of the clinical use of colistin and polymyxin B: the Prato polymyxin consensus. Lancet Infect Dis 15:225–234. https://doi.org/10.1016/S1473-3099(14)70850-3
- Cheah SE, Wang J, Nguyen VTT, Turnidge JD, Li J, Nation RL. 2015. New pharmacokinetic/pharmacodynamic studies of systemically administered colistin against *Pseudomonas aeruginosa* and *Acinetobacter baumannii* in mouse thigh and lung infection models: smaller response in lung infection. J Antimicrob Chemother 70:3291–3297. https://doi.org /10.1093/jac/dkv267
- Zhang B, Li X, Chen Y, Chen B, Cheng Y, Lin H, Que W, Liu M, Zhou L, Zhang H, Qiu H, Wu C. 2023. Determination of polymyxin B in human plasma and epithelial lining fluid using LC-MS/MS and its clinical application in therapeutic drug monitoring. J Pharm Biomed Anal 227:115291. https://doi.org/10.1016/j.jpba.2023.115291
- Yapa SWS, Li J, Porter CJH, Nation RL, Patel K, McIntosh MP. 2013. Population pharmacokinetics of colistin methanesulfonate in rats: achieving sustained lung concentrations of colistin for targeting respiratory infections. Antimicrob Agents Chemother 57:5087–5095. htt ps://doi.org/10.1128/AAC.01127-13
- Tang T, Li Y, Xu P, Zhong Y, Yang M, Ma W, Xiang D, Zhang B, Zhou Y. 2023. Optimization of polymyxin B regimens for the treatment of carbapenem-resistant organism nosocomial pneumonia: a real-world prospective study. Crit Care 27:164. https://doi.org/10.1186/s13054-023-04448-z
- 24. Zhao Y, Chen H, Yu Z. 2023. Trough concentration may not be a good target for polymyxin B therapeutic drug monitoring. Crit Care 27:41. https://doi.org/10.1186/s13054-023-04326-8
- Liu S, Wu Y, Qi S, Shao H, Feng M, Xing L, Liu H, Gao Y, Zhu Z, Zhang S, Du Y, Lu Y, Yang J, Chen P, Sun T. 2023. Polymyxin B therapy based on therapeutic drug monitoring in carbapenem-resistant organisms sepsis:

- the PMB-CROS randomized clinical trial. Crit Care 27:232. https://doi.org/10.1186/s13054-023-04522-6
- Manchandani P, Zhou J, Ledesma KR, Truong LD, Chow DS-L, Eriksen JL, Tam VH. 2016. Characterization of polymyxin B biodistribution and disposition in an animal model. Antimicrob Agents Chemother 60:1029– 1034. https://doi.org/10.1128/AAC.02445-15
- Yu Z, Liu X, Du X, Chen H, Zhao F, Zhou Z, Wang Y, Zheng Y, Bergen PJ, Li X, Sun R, Fang L, Li W, Fan Y, Wu H, Guo B, Li J, Yu Y, Zhang J. 2022. Pharmacokinetics/pharmacodynamics of polymyxin B in patients with bloodstream infection caused by carbapenem-resistant *Klebsiella pneumoniae*. Front Pharmacol 13:975066. https://doi.org/10.3389/fphar. 2022.975066
- Furtado GHC, d'Azevedo PA, Santos AF, Gales AC, Pignatari ACC, Medeiros EAS. 2007. Intravenous polymyxin B for the treatment of nosocomial pneumonia caused by multidrug-resistant *Pseudomonas* aeruginosa. Int J Antimicrob Agents 30:315–319. https://doi.org/10.1016 /i.ijantimicag.2007.05.017
- Rigatto MH, Ribeiro VB, Konzen D, Zavascki AP. 2013. Comparison of polymyxin B with other antimicrobials in the treatment of ventilatorassociated pneumonia and tracheobronchitis caused by *Pseudomonas* aeruginosa or Acinetobacter baumannii. Infection 41:321–328. https://doi. org/10.1007/s15010-012-0349-z
- Ding P, Li H, Nan Y, Liu C, Wang G, Cai H, Yu W. 2024. Outcome of intravenous and inhaled polymyxin B treatment in patients with multidrug-resistant gram-negative bacterial pneumonia. Int J Antimicrob Agents 64:107293. https://doi.org/10.1016/j.ijantimicag.2024 107293
- Liu J, Shao M, Xu Q, Liu F, Pan X, Wu J, Xiong L, Wu Y, Tian M, Yao J, Huang S, Zhang L, Chen Y, Zhang S, Wen Z, Du H, Liu Y, Li W, Xu Y, Teboul J-L, Chen D, TaoWang. 2022. Low-dose intravenous plus inhaled versus intravenous polymyxin B for the treatment of extensive drugresistant Gram-negative ventilator-associated pneumonia in the critical illnesses: a multi-center matched case-control study. Ann Intensive Care 12:72. https://doi.org/10.1186/s13613-022-01033-5
- 32. Rouby J-J, Zhu Y, Torres A, Rello J, Monsel A. 2022. Aerosolized polymyxins for ventilator-associated pneumonia caused by extensive drug resistant Gram-negative bacteria: class, dose and manner should remain the trifecta. Ann Intensive Care 12:97. https://doi.org/10.1186/s13613-022-01068-8
- Cheah SE, Li J, Tsuji BT, Forrest A, Bulitta JB, Nation RL. 2016. Colistin and polymyxin B dosage regimens against Acinetobacter baumannii: differences in activity and the emergence of resistance. Antimicrob Agents Chemother 60:3921–3933. https://doi.org/10.1128/AAC.02927-1
- Hagihara M, Housman ST, Nicolau DP, Kuti JL. 2014. In vitro pharmacodynamics of polymyxin B and tigecycline alone and in combination against carbapenem-resistant Acinetobacter baumannii. Antimicrob Agents Chemother 58:874–879. https://doi.org/10.1128/AAC.01624-13

- Medeiros GS, Rigatto MH, Falci DR, Zavascki AP. 2019. Combination therapy with polymyxin B for carbapenemase-producing *Klebsiella* pneumoniae bloodstream infection. Int J Antimicrob Agents 53:152–157. https://doi.org/10.1016/j.ijantimicag.2018.10.010
- Pi M-Y, Cai C-J, Zuo L-Y, Zheng J-T, Zhang M-L, Lin X-B, Chen X, Zhong G-P, Xia Y-Z. 2023. Population pharmacokinetics and limited sampling strategies of polymyxin B in critically ill patients. J Antimicrob Chemother 78:792–801. https://doi.org/10.1093/jac/dkad012
- Surovoy YA, Burkin MA, Galvidis IA, Sobolev MA, Rende OC, Tsarenko SV. 2023. Comparative polymyxin B pharmacokinetics in critically ill patients with renal insufficiency and in continuous veno-venous hemodialysis. Eur J Clin Pharmacol 79:79–87. https://doi.org/10.1007/s00228-022-0341
- Hanafin PO, Kwa A, Zavascki AP, Sandri AM, Scheetz MH, Kubin CJ, Shah J, Cherng BPZ, Yin MT, Wang J, Wang L, Calfee DP, Bolon M, Pogue JM, Purcell AW, Nation RL, Li J, Kaye KS, Rao GG. 2023. A population pharmacokinetic model of polymyxin B based on prospective clinical data to inform dosing in hospitalized patients. Clin Microbiol Infect 29:1174–1181. https://doi.org/10.1016/j.cmi.2023.05.018
- Belcher JM, Garcia-Tsao G, Sanyal AJ, Bhogal H, Lim JK, Ansari N, Coca SG, Parikh CR, TRIBE-AKI Consortium. 2013. Association of AKI with mortality and complications in hospitalized patients with cirrhosis. Hepatology 57:753–762. https://doi.org/10.1002/hep.25735
- Nation RL, Rigatto MHP, Falci DR, Zavascki AP. 2019. Polymyxin acute kidney injury: dosing and other strategies to reduce toxicity. Antibiotics (Basel) 8:24. https://doi.org/10.3390/antibiotics8010024
- Lakota EA, Landersdorfer CB, Nation RL, Li J, Kaye KS, Rao GG, Forrest A. 2018. Personalizing Polymyxin B dosing using an adaptive feedback control algorithm. Antimicrob Agents Chemother 62:e00483-18. https:// doi.org/10.1128/AAC.00483-18
- Poston JT, Koyner JL. 2019. Sepsis associated acute kidney injury. BMJ 364:k4891. https://doi.org/10.1136/bmj.k4891
- Liu X, Yu Z, Wang Y, Wu H, Bian X, Li X, Fan Y, Guo B, Zhang J. 2020. Therapeutic drug monitoring of polymyxin B by LC-MS/MS in plasma and urine. Bioanalysis 12:845–855. https://doi.org/10.4155/bio-2020-005
- 44. Liu X, Huang C, Bergen PJ, Li J, Zhang J, Chen Y, Chen Y, Guo B, Hu F, Hu J, et al. 2023. Chinese consensus guidelines for therapeutic drug monitoring of polymyxin B, endorsed by the Infection and Chemotherapy Committee of the Shanghai Medical Association and the Therapeutic Drug Monitoring Committee of the Chinese Pharmacological Society. J Zhejiang Univ Sci B 24:130–142. https://doi.org/10.1631/jzus.B2200466
- Cockcroft DW, Gault MH. 1976. Prediction of creatinine clearance from serum creatinine. Nephron 16:31–41. https://doi.org/10.1159/00018058 0
- Venkataraman R, Kellum JA. 2007. Defining acute renal failure: the RIFLE criteria. J Intensive Care Med 22:187–193. https://doi.org/10.1177/08850 66607299510