# Endoplasmic reticulum stress and pulmonary hypertension

# Yanan Hu<sup>1,\*</sup>, Wenhao Yang<sup>2,3,4,\*</sup>, Liang Xie<sup>3,4</sup>, Tao Liu<sup>1</sup>, Hanmin Liu<sup>2,3,4</sup> and Bin Liu<sup>1</sup>

<sup>1</sup>Department of Pediatrics, The Affiliated Hospital of Southwest Medical University, Luzhou, China; <sup>2</sup>Department of Pediatrics, West China Second University Hospital, Sichuan University, Chengdu, China; <sup>3</sup>Key Laboratory of Birth Defects and Related Diseases of Women and Children (Sichuan University), Ministry of Education, Chengdu, China; <sup>4</sup>The Vascular Remodeling and Developmental Defects Research Unit, West China Institute of Women and Children's Health, West China Second University Hospital, Sichuan University, Chengdu, China

#### Abstract

Pulmonary hypertension is a fatal disease of which pulmonary vasculopathy is the main pathological feature resulting in the mean pulmonary arterial pressure higher than 25 mmHg. Moreover, pulmonary hypertension remains a tough problem with unclear molecular mechanisms. There have been dozens of studies about endoplasmic reticulum stress during the onset of pulmonary hypertension in patients, suggesting that endoplasmic reticulum stress may have a critical effect on the pathogenesis of pulmonary hypertension. The review aims to summarize the rationale to elucidate the role of endoplasmic reticulum stress in pulmonary hypertension. Started by reviewing the mechanisms responsible for the unfolded protein response following endoplasmic reticulum stress, the potential link between endoplasmic reticulum stress and pulmonary hypertension were introduced, and the contributions of endoplasmic reticulum stress to different vascular cells, mitochondria, and inflammation were described, and finally the potential therapies of attenuating endoplasmic reticulum stress for pulmonary hypertension were discussed.

### **Keywords**

pulmonary hypertension, endoplasmic reticulum stress, unfolded protein response, vascular remodeling

Date received: 24 September 2019; accepted: 19 December 2019

Pulmonary Circulation 2020; 10(1) 1–11 DOI: 10.1177/2045894019900121

# Introduction

Pulmonary hypertension (PH) is a highly hazardous disease, and gradually increasing pulmonary artery (PA) pressure is one of the main characteristics. The symptoms accompanied with high PA pressure include the remodeling of pulmonary vessels, enhanced vasoconstriction, and the compensatory hypertrophy of right ventricle (RV). In the final phase of this disease, heart failure and even death will occur. According to the current clinical classification system from World Health Organization, PH is classified into five categories based on presumed molecular etiologies, histopathology, and clinical associations.<sup>1</sup> Group 1 includes a severe form of PH called pulmonary artery hypertension (PAH). Other groups affect a much larger global population and reflect a wide variety of conditions, such as congenital or acquired left heart disease, lung diseases and/or hypoxic, chronic thromboembolism, and unclear multifactorial mechanisms. In the past, landmark studies have shown similar changes of PASMCs (pulmonary artery smooth muscle

cells)/ECs (endothelial cells) in the pathology of cancer and PH, which, to a certain extent, explains the pivotal mechanism of PH.<sup>2–4</sup> That is to say, some cancer-related studies could provide some references for PH research.<sup>5</sup>

At present, endoplasmic reticulum (ER) stress is a hot topic in the researches of cancer pathogenesis.<sup>6</sup> In recent years, several studies have shown that the glucose-regulated protein 78 kDa (GRP78), a molecular chaperone in ER stress, is involved in proliferation and survival of cancer cells and angiogenesis in tumor tissues.<sup>6,7</sup> Angiogenic transformation with tumor angiogenesis was discovered as a downstream target of the unfolded protein response

\*These authors contributed equally to this article.

Corresponding authors:

Email: Iblyfy@126.com

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (http://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).

© The Author(s) 2020. Article reuse guidelines: sagepub.com/journals-permissions journals.sagepub.com/home/pul



Hanmin Liu and Bin Liu, Department of Pediatrics, The Affiliated Hospital of Southwest Medical University, No. 25 Taiping Street, Luzhou 646000, Sichuan, China.

(UPR) pathway, emphasizing the importance of ER stress in tumor angiogenesis.<sup>8,9</sup> Therefore, some studies focused on the role of ER stress in PH and the results suggested that UPR functioned in the development of PH.<sup>10–12</sup> Nowadays, the inhibition of ER stress was considered as a new potential intervention in clinical treatment of PH. Some studies have already demonstrated that the treatment of using the chemical chaperone, such as 4-phenylbutyrate (PBA), to decrease ER stress could reverse or treat animal models of PH.<sup>11,12</sup> However, this therapy has not been used in clinical or preclinical studies yet. In a word, the relevant research results of ER stress presented a glimmer of hope for exploring the new targets for the future treatment of PH.

## The endoplasmic reticulum stress

#### Endoplasmic reticulum

ER is the central organelle for intracellular secretion. It is responsible for post-translational modification, folding and maturation, and secretion of transmembrane and secreted proteins. The proteins are then further transported to the Golgi and eventually secreted as the vesicles or displayed on the surface of the plasma.<sup>13</sup> Moreover, ER is also crucial for other cellular functions like biosynthesis of lipids (including triglycerides, phospholipids, and cholesterol), Ca<sup>2+</sup> buffering, and carbohydrate synthesis. However, the speed of proteins transportation and folding is affected by intracellular and extracellular factors, and changes among different cell types. Thus, by enlarging the entire size of ER preferentially and increasing the production of chaperone proteins, cells will adjust to the need for the entering of numerous nascent proteins into the lumen of ER for folding.<sup>14</sup>

# ER stress

Stress conditions of ER refer to situations such as the status of high glucose or lack of energy, hypoxia, Ca<sup>2+</sup> overload, oxidative stress, and exposure to chemicals that will cause imbalances in the homeostasis.<sup>15</sup> These stimulations activate the related signals to promote new proteins synthesis for dealing with stress, while these signals will reduce the general protein synthesis.<sup>16,17</sup> When the amount of translating proteins exceeds that which ER folding can handle, it will cause misfolded proteins to accumulate in the ER. Due to heaping up of these misfolded proteins in ER, an evolutionarily conserved response of stress which is called unfolded protein response (UPR) will be activated. The adaptive response that takes place in the initial phase of UPR aims to rehabilitate protein folding homeostasis.<sup>18–23</sup> When cells cannot recover from ER stress, UPR will terminate this adaptive response and trigger cell apoptosis.19,24-28 The function of UPR prevented the damaged and non-functional proteins heaping up in the ER by re-establishing body homeostasis or triggering cell death.<sup>29</sup>

There are three types of UPR signal transducers in mammalian cells, all of which are ER-resident transmembrane proteins. The three proteins are inositol-requiring enzyme 1  $\alpha/\beta$  (IRE1 $\alpha/\beta$ ), protein kinase RNA-like ER kinase (PERK), and activating transcription factor 6 (ATF6) (Fig. 1). IRE1 is essential for UPR in plants and animals.<sup>30-33</sup> As a multidomain protein of the ER transmembrane, IRE1 has the properties of both a kinase and an endoribonuclease. Under ER stress, IRE1 RNase is acticonformational vated through autophosphorylation. oligomerization.34-36 changes, and higher order The second sensor of UPR is PERK which primarily attenuates the translation of protein and modulates oxidative stress. This protein can phosphorylate and activate a transcription factor- nuclear factor erythroid-2-related factor 2 (NRF2) and eukaryotic initiation factor 2a (eIF2a) to alleviate unfolded proteins or misfolded proteins.<sup>29,37-41</sup> ATF6, the last sensor of the three branches of UPR, is a transmembrane protein and it also functions as a transcription factor when it is cleaved.<sup>42–44</sup> The direct target of the cleaved ATF6 is the UPR proteins, such as chaperones.<sup>45</sup> At present, there is a more comprehensive understanding of the signaling pathways in ER stress, but the connections between ER stress and many diseases are still waiting for more researchers to explore.

#### The relationship between ER stress and PH

It is widely accepted that the vascular remodeling is a major feature in the pathogenesis of PH and may be present in a wide range of diseased tissues. The corresponding pathological process causing vascular remodeling in PH is very complicated and involves a variety of environmental factors and genes.<sup>46–48</sup> This feature was associated with hyperplasia of PASMCs, excessive proliferation of PAECs, microthrombus formation, and persistent pulmonary vasoconstriction. And the uncontrolled over-proliferation of PASMCs and PAECs in complicated vascular lesions will eventually lead to occlusion of the PA, which further cause the adverse rise in pulmonary blood pressure.<sup>49–51</sup> In addition, these individual and collective changes were related to excessive cell proliferation, apoptosis resistance, circulating inflammatory cells recruitment, and phenotypic switching.<sup>52,53</sup>

Studies have shown that vascular remodeling is closely related to ER stress, especially the proliferation of PASMCs.<sup>12</sup> Other aspects of PH, such as proliferation of PAECs and the subcellular connections, are also closely related to ER stress.

#### Pulmonary artery smooth muscle cells

As indicated by the above description, vascular remodeling is one of the significant features in PH, but the mechanisms are still under investigating. In several studies, it has been pointed out that the imbalance of proliferation and apoptosis in PASMCs contributes to medial thickening.<sup>50,51</sup>



**Fig. 1.** The unfolded protein response (UPR) pathway. The three types of UPR signal transducers are PERK, IRE1, and ATF6. IRE1 has both kinase and endoribonuclease identity. When IRE1 is activated, a small intron of XBP1 is removed to form sXBP1, which involves the transcription of genes that restores ER folding ability. PERK phosphorylates eIF2a attenuating mRNA translation, but it specifically induces the transcription factors ATF4 and CHOP, which will induce the occurrence of related reactions such as protein synthesis and apoptosis. ATF6 is translocated into the Golgi where it is cleaved to expose the transcriptionally active cytoplasmic domain of ATF6. The direct target of the cleaved ATF6 is the UPR proteins, such as chaperones. Also, ATF6 can also induce CHOP and XBP1 genes.

PERK: protein kinase RNA-like ER kinase; IRE1: inositolrequiring enzyme 1; ATF6: activating transcription factor 6; BIP: Immunoglobulin binding protein; eIF2a: eukaryotic initiation factor 2a; XBP1: X-box binding protein; S1P: protease site 1 protease; S2P: protease site 2 protease; CHOP: transcription factor C/EBP homologous protein; mTOR: mammalian target of rapamycin; JNK: c-Jun N-terminal kinase.

Pathological studies have also shown that some cells in the intraluminal occlusion express SMC markers.<sup>54</sup> Therefore, some features such as over-proliferation and anti-apoptosis of PASMCs made the concept of carcinoid appearing. Meanwhile, there was increasing evidence that the hypertrophy, proliferation, and apoptosis resistance of PASMCs were critical components of abnormal vascular remodeling in PH<sup>5</sup> (Fig. 2).

Recent studies on the role of ER stress in PH have focused on PASMC. It has been suggested that PASMCs proliferation and resistance to apoptosis are essential for vascular remodeling in PH.<sup>55–57</sup> ER stress is a basic cellular response that promotes the proliferation and enhances inflammatory response of PASMCs.<sup>58</sup> The abnormal proliferation of PASMCs is the most important cause of pulmonary vascular remodeling in PAH.<sup>59,60</sup> Recently, some studies have shown that autophagy is involved in the development of PAH induced by MCT.<sup>61,62</sup> And eIF2 $\alpha$  can activate ER autophagy after ER stress.<sup>37–39</sup> At the same time, eIF2 $\alpha$  plays a key role in regulating cell proliferation and hypertrophy, and participated in the regulation of SMC proliferation and migration.<sup>63,64</sup> According to Wang et al., they observed that after transfected with eIF2 $\alpha$  siRNA in hypoxia-promoted PASMCs proliferation, it can significantly inhibit the proliferation of PASMCs.<sup>65,66</sup> Moreover, Cao et al. found that an inhibitor of the IRE1 $\alpha$ /XBP1 pathway, 4u8c, promoted apoptosis and repressed cell proliferation and migration of PASMCs.<sup>67</sup> All of the above indicate that ER stress is involved in the abnormal proliferation of PASMC in PH.

There was evidence indicating that recruiting the inflammatory cells and continuous development of inflammation in PH are two crucial components of pathological vascular remodeling.<sup>68</sup> Recently, it has been reported that ER stress may trigger smooth muscle cells to produce HA.<sup>69,70</sup> In the rat PH model induced by MCT, the activity of hyaluronidase-1 (HYAL1) increased significantly at the beginning of the disease, while its degradation and synthesis increased in the late stage of PH.<sup>71</sup> However, apart from the increase in the production of HA, there is no systematic description of the mechanism of HA clearance in the (diseased) lungs, and no complete description of the mechanism of the accumulation of HA in a balanced state. It is well known that the clearance of HA is critical to organ homeostasis.<sup>72</sup> Previous studies have shown that PH was related to the reducing degradation and increasing synthesis of HA, especially isomers of high molecular weight.<sup>71,73</sup> Yeager et al.<sup>74</sup> recently pointed out that the activation of UPR and the generation of pro-inflammatory biomolecule were attributed to ET-1 signaling in rat PASMCs. The occurrence of ER stress promotes the proliferation and inflammatory state of PASMCs, which will promote the onset of PH. These experimental



**Fig. 2.** The main pathological changes of pulmonary artery (PA) during endoplasmic reticulum (ER) stress reaction. ER stress can be induced by multiple adverse physiologic conditions. When ER stress occurs, ECs and SMCs in the PA play an extremely important role. In addition to hypertrophy, hyperplasia and apoptosis resistance, PASMCs also release inflammatory mediators. PAECs are in a dysfunctional state: in the early stage of PH, apoptosis of ECs increased, and in the later period, excessive proliferation occurred. These pathological processes will lead to the generation of microthrombi, persistent pulmonary vasoconstriction, and pulmonary vascular remodeling, eventually leading to PH and even death. PASMCs: pulmonary artery smooth muscle cells; PAECs: pulmonary artery endothelial cells.

results further demonstrate the importance of PASMC in the inflammatory process of PH development, especially in recruiting and preserving the immune cells. The persistent inflammation produced by PASMCs is related to vascular remodeling and inhibition of this process is a potential method of treating PH.

Dromparis et al. found that the ATF6 pathway could be activated in the vasculature of both hypoxic models in vivo and cultured vascular SMCs.<sup>11</sup> Surprisingly, they did not mention if the CHOP pathway was also activated. Meanwhile, the ATF6 signaling pathway has been shown to up-regulate Nogo to disrupt mitochondrial-ER units in PASMC, leading to the occurrence of PH.<sup>44,75</sup> Although the function of ER stress in PASMCs have been extensively studied, the mechanism is still unknown. In summary, inhibiting ER stress that stimulates the above PASMCs responses is also an extremely appealing treatment option.

#### Pulmonary arterial endothelial cells

The endothelium secretes many mediators which are necessary for normal vascular functioning, including the mediators of regulating vascular tone and coagulation, modulating immune responses, and controlling vascular cell growth. Moreover, vascular ECs compose a barrier between the vessel lumen and the wall, through which liquid, gas, and macromolecular substances could be selectively transported. At the same time, it also maintains the homeostasis of blood vessels and mediates the passage of nutrients and white blood cells through the vessel wall. Vascular intima can maintain the balance between vasodilation and vasoconstriction in physiological conditions. ECs play an important role in the early stage of PH due to their apoptosis.<sup>76</sup> Furthermore, in the studies of late-stage PH, it was confirmed that the over-proliferation endothelial cells, vasoconstriction, the formation of microthrombus, and eventual vascular remodeling are crucial for pathological changes of PH<sup>76</sup> (Fig. 2).

Changes of endothelial function in the pulmonary vasculature contribute to the neointimal formation, thickening of the intima, and occlusion of the distal pulmonary artery, which is of great significance for PH.<sup>51</sup> One feature in some patients with severe PH is a complex vascular lesion called a plexiform lesion.<sup>77</sup> These lesions can be seen in the myofibroblastic stroma in which the monoclonal EC of the lesion is proliferating.<sup>78–80</sup> The plexiform or complex vascular lesions are characterized by anti-apoptotic and phenotypic altered EC.<sup>81–85</sup>

ECs produce the vasodilator/anti-mitotic agents and vasoconstrictors/mitosis, such as prostacyclin, NO, thromboxane A2 and ET-1, to regulate SMCs activity.<sup>86</sup> Endothelial dysfunction is thought to be an imbalance that favors the production of vasoconstrictors and proliferative factors, including pro-inflammatory and thrombogenic effects, which has been clearly shown in PH.<sup>87</sup> Under the normal conditions, the main players are prostacyclin, nitric oxide (NO), and thrombomodulin. These participants inhibit SMC proliferation, platelet aggregation, and the expression of leukocyte adhesion molecule.<sup>88</sup>

Some dysfunctions are related to the reducing activity of endothelial NO synthase (eNOS), increasing endotheliumproduced ROS, decreasing anticoagulant properties, adhesion molecules up-expression, and release of chemokines and cytokines. It has been confirmed in previous studies that the release of the vasoconstrictor thromboxane A2 increased in these patients, in contrast, the release of prostacyclin is depressed.<sup>89</sup>

At the same time, more and more evidence showed that the imbalance of signal transduction of ROS and NO was related with PH.<sup>90</sup> Moreover, the decreasing expression of prostacyclin synthase in the pulmonary arteries could be detected in some patients with primary PH,<sup>91</sup> and the expression of NO synthase decreased in lung ECs in some PH patients.<sup>92</sup> Recently, the studies with independent PH model and muscles of CHOP<sup>-/-</sup> mice have suggested that in the ECs the expression of eNOS induced by ER stress has negative impacts on ECs function.<sup>93,94</sup>

There are also some patients with elevating plasma ET-1 levels.<sup>95</sup> In addition, elevating plasma ET-1 levels have also been reported in experimental models of PH.<sup>96</sup> Interestingly, ER stress also induces changes in ECs secretion.<sup>97</sup> In a study to confirm whether the UPR pathway was activated in ECs, Lenna et al.<sup>98</sup> examined the expression of pulmonary vascular UPR markers in the systemic sclerosis-associated PH (SSC-PH) patients. In this study, they observed that the expressions of BIP and CHOP as markers of ER stress in SSC-PH pulmonary vasculature and macrophages were significantly higher than those in healthy subjects. Interestingly, the elevating CHOP levels existed mainly in the ECs of SSC-PH lung.<sup>88</sup> In SSC-PH model of the mouse, the elevating ER stress markers could also be found in Gata6-KO mice.<sup>99</sup> The experimental result of Lenna et al.<sup>98</sup> suggested that macrophages and ECs might be the reason for increasing ER stress/UPR in hypoxic mice. According to the study of Lenna et al., thapsigargin, the ER stress inducer, could up-regulate the expression of ET-1 by increasing the formation of the ATF4/c-Jun transcription complex in ECs.<sup>100</sup> These findings have important implications for the study of the mechanism of ER stress in the endothelium of PH. Perhaps it is a good idea to focus on the ER stress in the treatment of PH.

# Subcellular networks

Increasing evidence indicated that the ER stress/UPR pathway had a relationship with other subcellular networks.<sup>88</sup> These networks included the ER-mitochondrial units, inflammatory response network and other subcellular factors.

Among all other organelles associated with ER, mitochondria are the most prominent organelle that regulates metabolism and cell survival. Previous studies have shown that mitochondrial dysfunction may lead to vascular remodeling, thus affecting the occurrence of PH.<sup>101-103</sup> Mitochondria-associated membrane (MAM) is a special contact formed by the interaction of ER and mitochondrion in physical structure.<sup>104</sup> Membrane and luminal components can mix and exchange in the MAM, and the composition of MAM is adapted in response to a variety of internal and external stimuli.<sup>104–106</sup> The structure of MAM is complicated and involves a large number of proteins with widely various functions.<sup>107</sup> IP3R (inositol 1.4.5-triphate receptor) and VDAC (voltage-dependent anion channel) are the major ER-mitochondrial calcium (Ca<sup>2+</sup>) transfer channels (Fig. 3). They are located on both the ER side and mitochondria side of the MAM and physically connect the two organelles by forming a complex with the chaperone GRP75, respectively.<sup>108</sup> Ca<sup>2+</sup> has been identified as an important regulator of cell proliferation and apoptosis, and it also plays an important role in PASMC proliferation and vasoconstriction of PH.<sup>109-113</sup> And the ER is the main intracellular reservoir of  $Ca^{2+}$ , while many ER chaperone proteins are dependent on  $Ca^{2+}$ , thus the regulation of  $Ca^{2+}$ homeostasis in ER is very important.<sup>114,115</sup> Alterations of calcium manipulate by the ER/mitochondrial couple providing a pathway to activate apoptosis.<sup>116</sup> A key process linking apoptosis to ER-mitochondrial interactions is the change of the Ca<sup>2+</sup> homeostasis, which results in massive and/or prolonged mitochondrial Ca<sup>2+</sup> overload.<sup>107</sup> Moreover, Mitochondria are accepted as indispensable oxygen sensor for hypoxic pulmonary vasoconstriction in cells.<sup>117</sup> And mitochondrial signals will promote apoptosis-resistance and proliferative diathesis of vascular remodeling in PH.<sup>118</sup>

Recently, several studies have shown that ER stress have involved in the development of inflammation during the PH, but all these are preliminary studies.<sup>12,74,98</sup> However, the roles of inflammation both in ER stress and PH are highly correlated. Proinflammatory cytokines and chemokines have been shown to be involved in the animal model of monocrotaline-induced PH.119-122 Most patients with primary PH have evidence of autoimmune and/or active inflammation. In these patients, circulating antinuclear antibodies and increasing serum levels of interleukin (IL)-1 and IL-6 can be detected.<sup>123,124</sup> And several studies have demonstrated that in ER stress IRE1 and PERK signaling pathways also participate in the inflammatory response. 125,126 The UPR pathway also can trigger the activation of NF-kB, another major inflammatory mediator.<sup>127</sup> The specific relationship among ER stress, inflammation and PH could contribute to reveals the pathogenesis of PH, which requires further research.

Among the other various factors affecting PH, the PDGF- $\beta$  receptor is one of the most concerned focuses in PH researches and it is responsible for proliferation of PASMCs and neointimal hyperplasia of the PA.<sup>128–130</sup> Other experiments of cultured PASMCs have indicated that using platelet-derived growth factor (PDGF)-BB to



**Fig. 3.** Correlation between endoplasmic reticulum (ER) and mitochondria. Under physiological conditions, Ca2 + can be transferred from the ER into the mitochondria. Early stage of ER stress triggers an increase in mitochondrial metabolism which depends critically upon mitochondria-associated membrane (MAMS) and Ca2 + transfer. If stress persists, this response leads to mitochondrial collapse and triggers apoptotic cell death.

IP3R: inositol 1,4,5-triphate receptor; VDAC: voltage-dependent anion channel; ATP: adenosine triphosphate.

treat cells will activate the UPR pathway.<sup>12</sup> These results suggest that PDGF-BB may be one of the contributors to activate the UPR pathway in PH. There has been increasing evidence showing that the ER stress and UPR pathways have a critical effect on regulating the expression of proangiogenic factors. They also serve as novel mediators of angiogenesis. Up-regulation of angiogenic factors such as VEGF-A, FGF, IL-8, and ET-1 has been reported to occur during the UPR.<sup>131</sup> Moreover, according to reports, as a regulatory ER structural protein, <sup>132,133</sup> Nogo-B is involved in vascular remodeling and plays a role in PAH.<sup>75,134</sup> At the same time, it has also been demonstrated that PH-related loss of function mutations of BMPRII and the resultant protein transporting dysfunction can also induce ER stress.<sup>135</sup> Furthermore, recent researches on metabolic abnormalities of the pulmonary vascular system and the right ventricle have also attracted more and more attention.<sup>136,137</sup>

# **Potential therapies**

Recently, several studies have suggested that the chemical chaperones of ER stress, like 4-PBA or tauroursodeoxycholic acid (TUDCA), could reduce the mPAP significantly in PH animal model.<sup>10–12</sup> Chemical chaperones are often defined as low molecular weight compounds.<sup>138,139</sup> So far, the exact mechanism of chemical chaperones in ER stress has not been elucidated. Most likely, these two molecules exert their effects by stabilizing the structure of incorrectly folded

proteins, stimulating molecular chaperones to obtain more efficient protein trafficking, reducing protein aggregation, and preventing the interactions of non-specific proteins and other proteins.<sup>140–142</sup> 4-PBA, One of the chemical chaperones which most commonly mentioned, is a low molecular weight fatty acid and a non-toxic pharmacological compound.143-146 It has three main pharmacological effects as an ammonia scavenger, <sup>143,147</sup> a weak histone deacetylase inhibitor, <sup>148,149</sup> and an ER stress inhibitor.<sup>150,151</sup> Due to its ammonia scavenger properties, it has been approved by the FDA for clinical use in pathological diseases of the urea cycle<sup>147,152,153</sup> At present, it has been widely studied as a small chemical chaperone to modulate restoration of ER homeostasis and multiple concerning pathological conditions.<sup>12,154</sup> Furthermore, using 4-PBA can down-regulate several key proteins in ER stress, thereby improving PA and RV remodeling.<sup>3,10,143,155</sup> Another chemical chaperone, TUDCA, a safe hydrophilic bile acid. Although the exact mechanism of TUDCA as a chemical chaperone is still unclear, it has been shown to prevent UPR dysfunction and reduce ER stress in various cell types.<sup>156-160</sup> Recently, there has been shown that TUDCA binded with the hydrophobic regions of proteins to prevent subsequent protein aggregation and unfolded protein accumulation, thus attenuating ER stress.<sup>161,162</sup> And it has been reported that TUDCA also exerts these effects partially by assisting in the transfer of mutant proteins and partially by improving protein folding capacity through the activation of ATF6.163

These above results showed that attenuating ER stress could be an effective treatment strategy to protect the PA from damage. If the association between ER stress and PH was figured out, the use of these drugs which have non-toxic and does not burden other organs would be a nice choice for clinical treatment of PH.

# Summary

A review of all the literature mentioned in the text makes it evident that ER stress was involved in the PH, and may play an important role. These above content in this article show that attenuating ER stress may be an effective treatment to protect the PA. It is also clear that the exact molecular mechanisms of ER stress in the pathology of PH remains unclear. More studies are needed in future.

#### Authors' contribution

YH and WY contributed in original draft preparation, LX, BL and HL in review and editing, YH and TL in literature correction, and BL and HL in supervision.

#### Guarantor

Yanan Hu, Bin Liu, Hanmin Liu.

#### **Ethical approval**

No ethical statement will be required for this study because there is no direct involvement of human.

# **Conflict of interest**

The author(s) declare that there is no conflict of interest.

#### Funding

This research was supported by Sichuan Science and Technology Program "Key Research and Development Projects of Sichuan Science and Technology Department" (19ZDYF1169). This research was also supported by Science and technology project of Chengdu Science and Technology Bureau (2016-HM01-00102-SF).

#### References

- Simonneau G, Gatzoulis MA, Adatia I, et al. Updated clinical classification of pulmonary hypertension. J Am Coll Cardiol 2013; 62: D34–41.
- Rai PR, Cool CD, King JA, et al. The cancer paradigm of severe pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2008; 178: 558–564.
- 3. Sutendra G and Michelakis ED. The metabolic basis of pulmonary arterial hypertension. *Cell Metab* 2014; 19: 558–573.
- Ruffenach G, Bonnet S, Rousseaux S, et al. Identity crisis in pulmonary arterial hypertension. *Pulm Circ* 2018; 8: 2045893217746054.
- 5. Boucherat O, Vitry G, Trinh I, et al. The cancer theory of pulmonary arterial hypertension. *Pulm Circ* 2017; 7: 285–299.
- Dong D, Ni M, Li J, et al. Critical role of the stress chaperone GRP78/BiP in tumor proliferation, survival, and tumor angiogenesis in transgene-induced mammary tumor development. *Cancer Res* 2008; 68: 498–505.

- Cullen SJ, Fatemie S and Ladiges W. Breast tumor cells primed by endoplasmic reticulum stress remodel macrophage phenotype. *Am J Cancer Res* 2013; 3: 196–210.
- Ghosh R, Lipson KL, Sargent KE, et al. Transcriptional regulation of VEGF-A by the unfolded protein response pathway. *PLoS One* 2010; 5: e9575.
- Wang YG, Alam GN, Ning Y, et al. The unfolded protein response induces the angiogenic switch in human tumor cells through the PERK/ATF4 pathway. *Cancer Research* 2012; 72: 5396–5406.
- Wu Y, Adi D, Long M, et al. 4-Phenylbutyric acid induces protection against pulmonary arterial hypertension in rats. *PLoS One* 2016; 11: e0157538.
- 11. Dromparis P, Paulin R, Stenson TH, et al. Attenuating endoplasmic reticulum stress as a novel therapeutic strategy in pulmonary hypertension. *Circulation* 2013; 127: 115.
- Koyama M, Furuhashi M, Ishimura S, et al. Reduction of endoplasmic reticulum stress by 4-phenylbutyric acid prevents the development of hypoxia-induced pulmonary arterial hypertension. *Am J Physiol Heart Circ Physiol* 2014; 306: H1314–1323.
- 13. Sano R and Reed JC. ER stress-induced cell death mechanisms. *Biochim Biophys Acta* 2013; 1833: 3460–3470.
- Schuck S, Prinz WA, Thorn KS, et al. Membrane expansion alleviates endoplasmic reticulum stress independently of the unfolded protein response. *J Cell Biol* 2009; 187: 525–536.
- 15. Mao SZ, Fan XF, Xue F, et al. Intermedin modulates hypoxic pulmonary vascular remodeling by inhibiting pulmonary artery smooth muscle cell proliferation. *Pulm Pharmacol Ther* 2014; 27: 1–9.
- Ron D and Walter P. Signal integration in the endoplasmic reticulum unfolded protein response. *Nat Rev Mol Cell Biol* 2007; 8: 519–529.
- 17. Hetz C, Chevet E and Oakes SA. Proteostasis control by the unfolded protein response. *Nat Cell Biol* 2015; 17: 829–838.
- Feldman M and van der Goot FG. Novel ubiquitin-dependent quality control in the endoplasmic reticulum. *Trends Cell Biol* 2009; 19: 357–363.
- 19. Merksamer PI and Papa FR. The UPR and cell fate at a glance. *J Cell Sci* 2010; 123: 1003–1006.
- Hoseki J, Ushioda R and Nagata K. Mechanism and components of endoplasmic reticulum-associated degradation. *J Biochem* 2010; 147: 19–25.
- Fonseca SG, Gromada J and Urano F. Endoplasmic reticulum stress and pancreatic beta-cell death. *Trends Endocrinol Metab* 2011; 22: 266–274.
- 22. Walter P and Ron D. The unfolded protein response: from stress pathway to homeostatic regulation. *Science* 2011; 334: 1081–1086.
- Smith MH, Ploegh HL and Weissman JS. Road to ruin: targeting proteins for degradation in the endoplasmic reticulum. *Science* 2011; 334: 1086–1090.
- 24. Woehlbier U and Hetz C. Modulating stress responses by the UPRosome: a matter of life and death. *Trends Biochem Sci* 2011; 36: 329–337.
- Shore GC, Papa FR and Oakes SA. Signaling cell death from the endoplasmic reticulum stress response. *Curr Opin Cell Biol* 2011; 23: 143–149.
- Hetz C. The unfolded protein response: controlling cell fate decisions under ER stress and beyond. *Nat Rev Mol Cell Bio* 2012; 13: 89–102.

- 27. Appenzeller-Herzog C and Hall MN. Bidirectional crosstalk between endoplasmic reticulum stress and mTOR signaling. *Trends Cell Biol* 2012; 22: 274–282.
- Jager R, Bertrand MJM, Gorman AM, et al. The unfolded protein response at the crossroads of cellular life and death during endoplasmic reticulum stress. *Biol Cell* 2012; 104: 259–270.
- 29. McQuiston A and Diehl JA. Recent insights into PERKdependent signaling from the stressed endoplasmic reticulum. *F1000Res* 2017; 6: 1897.
- Hetz C and Glimcher LH. Fine-tuning of the unfolded protein response: assembling the IRE1 alpha interactome. *Molecular Cell* 2009; 35: 551–561.
- Hetz C, Martinon F, Rodriguez D, et al. The unfolded protein response: integrating stress signals through the stress sensor IRE1 alpha. *Physiol Rev* 2011; 91: 1219–1243.
- 32. Nagashima Y, Mishiba K, Suzuki E, et al. Arabidopsis IRE1 catalyses unconventional splicing of bZIP60 mRNA to produce the active transcription factor. *Sci Rep* 2011; 1: 29.
- Chen YN and Brandizzi F. AtIRE1A/AtIRE1B and AGB1 independently control two essential unfolded protein response pathways in Arabidopsis. *Plant J* 2012; 69: 266–277.
- Ali MMU, Bagratuni T, Davenport EL, et al. Structure of the Ire1 autophosphorylation complex and implications for the unfolded protein response. *Embo J* 2011; 30: 894–905.
- Aragon T, van Anken E, Pincus D, et al. Messenger RNA targeting to endoplasmic reticulum stress signalling sites. *Nature* 2009; 457: 736–740.
- Korennykh AV, Egea PF, Korostelev AA, et al. The unfolded protein response signals through high-order assembly of Ire1. *Nature* 2009; 457: 687–693.
- Harding HP, Zhang Y, Zeng H, et al. An integrated stress response regulates amino acid metabolism and resistance to oxidative stress. *Mol Cell* 2003; 11: 619–633.
- Tsai YC and Weissman AM. The unfolded protein response, degradation from endoplasmic reticulum and cancer. *Genes Cancer* 2010; 1: 764–778.
- Han J, Back SH, Hur J, et al. ER-stress-induced transcriptional regulation increases protein synthesis leading to cell death. *Nat Cell Biol* 2013; 15: 481–490.
- B'chir W, Maurin AC, Carraro V, et al. The eIF2 alpha/ATF4 pathway is essential for stress-induced autophagy gene expression. *Nucleic Acids Res* 2013; 41: 7683–7699.
- Harding HP, Novoa I, Zhang YH, et al. Regulated translation initiation controls stress-induced gene expression in mammalian cells. *Mol Cell* 2000; 6: 1099–1108.
- Yoshida H, Okada T, Haze K, et al. ATF6 activated by proteolysis binds in the presence of NF-Y (CBF) directly to the cis-acting element responsible for the mammalian unfolded protein response. *Mol Cell Biol* 2000; 20: 6755–6767.
- Gupta A, Hossain MM, Read DE, et al. PERK regulated miR-424(322)-503 cluster fine-tunes activation of IRE1 and ATF6 during unfolded protein response. *Sci Rep* 2015; 5: 18304.
- 44. Yoshida H, Matsui T, Yamamoto A, et al. XBP1 mRNA is induced by ATF6 and spliced by IRE1 in response to ER stress to produce a highly active transcription factor. *Cell* 2001; 107: 881–891.
- Shen J, Snapp EL, Lippincott-Schwartz J, et al. Stable binding of ATF6 to BiP in the endoplasmic reticulum stress response. *Mol Cell Biol* 2005; 25: 921–932.

- Parpaleix A, Amsellem V, Houssaini A, et al. Role of interleukin-1 receptor 1/MyD88 signalling in the development and progression of pulmonary hypertension. *Eur Respir J* 2016; 48: 470–483.
- Pugh ME and Hemnes AR. Metabolic and hormonal derangements in pulmonary hypertension: from mouse to man. *Int J Clin Pract* 2010; 64: 5–13.
- 48. Assad TR and Hemnes AR. Metabolic dysfunction in pulmonary arterial hypertension. *Curr Hypertens Rep* 2015; 17.
- Humbert M, Morrell NW, Archer SL, et al. Cellular and molecular pathobiology of pulmonary arterial hypertension. *J Am Coll Cardiol* 2004; 43: 13S–24S.
- Ranchoux B, Harvey LD, Ayon RJ, et al. Endothelial dysfunction in pulmonary arterial hypertension: an evolving landscape (2017 Grover Conference Series). *Pulm Circ* 2018; 8: 2045893217752912.
- Humbert M, Montani D, Perros F, et al. Endothelial cell dysfunction and cross talk between endothelium and smooth muscle cells in pulmonary arterial hypertension. *Vasc Pharmacol* 2008; 49: 113–118.
- Schermuly RT, Ghofrani HA, Wilkins MR, et al. Mechanisms of disease: pulmonary arterial hypertension. *Nat Rev Cardiol* 2011; 8: 443–455.
- 53. Rabinovitch M. Molecular pathogenesis of pulmonary arterial hypertension. *J Clin Invest* 2012; 122: 4306–4313.
- 54. Yi ES, Kim H, Ahn H, et al. Distribution of obstructive intimal lesions and their cellular phenotypes in chronic pulmonary hypertension – a morphometric and immunohistochemical study. *Am J Resp Crit Care* 2000; 162: 1577–1586.
- 55. Burg ED, Remillard CV and Yuan JX. Potassium channels in the regulation of pulmonary artery smooth muscle cell proliferation and apoptosis: pharmacotherapeutic implications. *Br J Pharmacol* 2008; 153(Suppl 1): S99–S111.
- Perros F, Montani D, Dorfmuller P, et al. Platelet-derived growth factor expression and function in idiopathic pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2008; 178: 81–88.
- 57. Wang G, Liu X, Meng L, et al. Up-regulated lipocalin-2 in pulmonary hypertension involving in pulmonary artery SMC resistance to apoptosis. *Int J Biol Sci* 2014; 10: 798–806.
- Chen R, Zhong W, Shao C, et al. Docosahexaenoic acid inhibits monocrotaline-induced pulmonary hypertension via attenuating endoplasmic reticulum stress and inflammation. *Am J Physiol Lung Cell Mol Physiol* 2018; 314: L243–L255.
- Santos-Ribeiro D, Mendes-Ferreira P, Maia-Rocha C, et al. Pulmonary arterial hypertension: Basic knowledge for clinicians. Arch Cardiovasc Dis 2016; 109: 550–561.
- Yu L, Tu Y, Jia X, et al. Resveratrol protects against pulmonary arterial hypertension in rats via activation of silent information regulator 1. *Cell Physiol Biochem* 2017; 42: 55–67.
- Long L, Yang X, Southwood M, et al. Chloroquine prevents progression of experimental pulmonary hypertension via inhibition of autophagy and lysosomal bone morphogenetic protein type II receptor degradation. *Circ Res* 2013; 112: 1159–1170.
- 62. Kim D and George MP. Pulmonary hypertension. *Med Clin North Am* 2019; 103: 413–423.
- Jiang L, Zang D, Yi S, et al. A microRNA-mediated decrease in eukaryotic initiation factor 2α promotes cell survival during PS-341 treatment. *Sci Rep* 2016; 6: 21565.

- Liu X, Bennett RL, Cheng X, et al. PKR regulates proliferation, differentiation, and survival of murine hematopoietic stem/progenitor cells. *Blood* 2013; 121: 3364–3374.
- Wang AP, Li XH, Yang YM, et al. A critical role of the mTOR/eIF2α pathway in hypoxia-induced pulmonary hypertension. *PLoS One* 2015; 10: e0130806.
- Guo L, Li Y, Tian Y, et al. eIF2alpha promotes vascular remodeling via autophagy in monocrotaline-induced pulmonary arterial hypertension rats. *Drug Des Devel Ther* 2019; 13: 2799–2809.
- 67. X C, Y H, X L, et al. The IRE1α-XBP1 pathway function in hypoxia-induced pulmonary vascular remodeling, is upregulated by quercetin, inhibits apoptosis and partially reverses the effect of quercetin in PASMCs. *Am J Transl Res* 2019; 11: 641–654.
- Hassoun PM, Mouthon L, Barbera JA, et al. Inflammation, growth factors, and pulmonary vascular remodeling. J Am Coll Cardiol 2009; 54: S10–19.
- Majors AK, Austin RC, de la Motte CA, et al. Endoplasmic reticulum stress induces hyaluronan deposition and leukocyte adhesion. J Biol Chem 2003; 278: 47223–47231.
- ME L, D M, C F, et al. Primary murine airway smooth muscle cells exposed to poly(I,C) or tunicamycin synthesize a leukocyte-adhesive hyaluronan matrix. *J Biol Chem* 2009; 284: 5299–5312.
- Ormiston ML, Slaughter GR, Deng Y, et al. The enzymatic degradation of hyaluronan is associated with disease progression in experimental pulmonary hypertension. *Am J Physiol Lung Cell Mol Physiol* 2010; 298: L148–157.
- Fischer JW and Schror K. Regulation of hyaluronan synthesis by vasodilatory prostaglandins. Implications for atherosclerosis. *Thromb Haemost* 2007; 98: 287–295.
- Aytekin M, Comhair SA, de la Motte C, et al. High levels of hyaluronan in idiopathic pulmonary arterial hypertension. *Am J Physiol Lung Cell Mol Physiol* 2008; 295: L789–799.
- 74. Yeager ME, Belchenko DD, Nguyen CM, et al. Endothelin-1, the unfolded protein response, and persistent inflammation: role of pulmonary artery smooth muscle cells. *Am J Respir Cell Mol Biol* 2012; 46: 14–22.
- 75. Sutendra G, Dromparis P, Wright P, et al. The role of Nogo and the mitochondria-endoplasmic reticulum unit in pulmonary hypertension. *Sci Transl Med* 2011; 3: 88ra55.
- Voelkel NF and Cool C. Pathology of pulmonary hypertension. *Cardiol Clin* 2004; 22: 343–351, v.
- Voelkel NF Tuder RM and Weir EK. Pathophysiology of primary pulmonary hypertension: from physiology to molecular mechanisms. In: Rubin LJ and Rich S (ed) *Primary pulmonary hypertension*. New York: Marcel Dekker, Inc, 1997, pp. 83–129.
- Voelkel NF and Tuder RM. Severe pulmonary hypertensive diseases: a perspective. *Eur Respir J* 1999; 14: 1246–1250.
- Cool CD, Stewart JS, Werahera P, et al. Three-dimensional reconstruction of pulmonary arteries in plexiform pulmonary hypertension using cell-specific markers. *Am J Pathol* 1999; 155: 411–419.
- Sakao S, Tatsumi K and Voelkel NF. Reversible or irreversible remodeling in pulmonary arterial hypertension. *Am J Respir Cell Mol Biol* 2010; 43: 629–634.
- 81. Levy M, Maurey C, Celermajer DS, et al. Impaired apoptosis of pulmonary endothelial cells is associated with intimal

proliferation and irreversibility of pulmonary hypertension in congenital heart disease. J Am Coll Cardiol 2007; 49: 803-810.

- Yuan JXJ and Rubin LJ. Pathogenesis of pulmonary arterial hypertension – the need for multiple hits. *Circulation* 2005; 111: 534–538.
- Sakao ST-SL, Cool CD, Tada Y, et al. VEGF-R blockade causes endothelial cell apoptosis, expansion of surviving CD341 precursor cells and transdifferentiation to smooth muscle-like and neuronal-like cells. *FASEB J* 2007; 21: 3640–3652.
- 84. Taraseviciene-Stewart L, Kasahara Y, Alger L, et al. Inhibition of the VEGF receptor 2 combined with chronic hypoxia causes cell death-dependent pulmonary endothelial cell proliferation and severe pulmonary hypertension. *FASEB J* 2001; 15: 427–438.
- 85. Sakao S, Taraseviciene-Stewart L, Lee JD, et al. Initial apoptosis is followed by increased proliferation of apoptosis-resistant endothelial cells. *Faseb J* 2005; 19: 1178.
- Galie N. The endothelin system in pulmonary arterial hypertension. *Cardiovasc Res* 2004; 61: 227–237.
- Archer S and Rich S. Primary pulmonary hypertension: a vascular biology and translational research "Work in progress". *Circulation* 2000; 102: 2781–2791.
- Lenna S, Han R and Trojanowska M. Endoplasmic reticulum stress and endothelial dysfunction. *IUBMB Life* 2014; 66: 530–537.
- Christman BW, McPherson CD, Newman JH, et al. An imbalance between the excretion of thromboxane and prostacyclin metabolites in pulmonary hypertension. *N Engl J Med* 1992; 327: 70–75.
- Tabima DM, Frizzell S and Gladwin MT. Reactive oxygen and nitrogen species in pulmonary hypertension. *Free Radic Biol Med* 2012; 52: 1970–1986.
- Giaid A and Saleh D. Reduced expression of endothelial nitric oxide synthase in the lungs of patients with pulmonary hypertension. N Engl J Med 1995; 333: 214–221.
- Tuder RM, Cool CD, Geraci MW, et al. Prostacyclin synthase expression is decreased in lungs from patients with severe pulmonary hypertension. *Am J Resp Crit Care* 1999; 159: 1925–1932.
- Lenna S, Townsend DM, Tan FK, et al. HLA-B35 upregulates endothelin-1 and downregulates endothelial nitric oxide synthase via endoplasmic reticulum stress response in endothelial cells. *J Immunol* 2010; 184: 4654–4661.
- Loinard C, Zouggari Y, Rueda P, et al. C/EBP homologous protein-10 (CHOP-10) limits postnatal neovascularization through control of endothelial nitric oxide synthase gene expression. *Circulation* 2012; 125: 1014–1026.
- Stewart DJ, Levy RD, Cernacek P, et al. Increased plasma endothelin-1 in pulmonary-hypertension – marker or mediator of disease. *Ann Intern Med* 1991; 114: 464–469.
- 96. Miyauchi T, Yorikane R, Sakai S, et al. Contribution of endogenous endothelin-1 to the progression of cardiopulmonary alterations in rats with monocrotaline-induced pulmonaryhypertension. *Circ Res* 1993; 73: 887–897.
- Galan M, Kassan M, Kadowitz PJ, et al. Mechanism of endoplasmic reticulum stress-induced vascular endothelial dysfunction. *Biochim Biophys Acta* 2014; 1843: 1063–1075.
- Lenna S, Farina AG, Martyanov V, et al. Increased expression of endoplasmic reticulum stress and unfolded protein response

genes in peripheral blood mononuclear cells from patients with limited cutaneous systemic sclerosis and pulmonary arterial hypertension. *Arthritis Rheum* 2013; 65: 1357–1366.

- Ghatnekar A, Chrobak I, Reese C, et al. Endothelial GATA-6 deficiency promotes pulmonary arterial hypertension. *Am J Pathol* 2013; 182: 2391–2406.
- 100. Lenna S, Chrobak I, Farina GA, et al. HLA-B35 and dsRNA induce endothelin-1 via activation of ATF4 in human microvascular endothelial cells. *PLoS One* 2013; 8: e56123.
- Ten VS and Ratner V. Mitochondrial bioenergetics and pulmonary dysfunction: current progress and future directions. *Paediatr Respir Rev* 2019; pii: S1526-0542(19)30036–3.
- Culley MK and Chan SY. Mitochondrial metabolism in pulmonary hypertension: beyond mountains there are mountains. *J Clin Invest* 2018; 128: 3704–3715.
- Marshall JD, Bazan I, Zhang Y, et al. Mitochondrial dysfunction and pulmonary hypertension: cause, effect, or both. *Am J Physiol Lung Cell Mol Physiol* 2018; 314: L782–L796.
- Vance JE. Phospholipid synthesis in a membrane fraction associated with mitochondria. J Biol Chem 1990; 265: 7248–7256.
- Myhill N, Lynes EM, Nanji JA, et al. The subcellular distribution of calnexin is mediated by PACS-2. *Mol Biol Cell* 2008; 19: 2777–2788.
- 106. Bui M, Gilady SY, Fitzsimmons REB, et al. Rab32 modulates apoptosis onset and mitochondria-associated membrane (MAM) properties. J Biol Chem 2010; 285: 31590–31602.
- 107. Giorgi C, De Stefani D, Bononi A, et al. Structural and functional link between the mitochondrial network and the endoplasmic reticulum. *Int J Biochem Cell B* 2009; 41: 1817–1827.
- 108. Szabadkai G, Bianchi K, Varnai P, et al. Chaperonemediated coupling of endoplasmic reticulum and mitochondrial Ca2+ channels. J Cell Biol 2006; 175: 901–911.
- Hadri L, Kratlian RG, Benard L, et al. Therapeutic efficacy of AAV1.SERCA2a in monocrotaline-induced pulmonary arterial hypertension. *Circulation* 2013; 128: 512–523.
- 110. Aguero J, Ishikawa K, Hadri L, et al. Characterization of right ventricular remodeling and failure in a chronic pulmonary hypertension model. *Am J Physiol Heart Circ Physiol* 2014; 307: H1204–1215.
- 111. Aguero J, Ishikawa K, Hadri L, et al. Intratracheal gene delivery of SERCA2a ameliorates chronic post-capillary pulmonary hypertension: a large animal model. J Am Coll Cardiol 2016; 67: 2032–2046.
- 112. Bonnet S, Rochefort G, Sutendra G, et al. The nuclear factor of activated T cells in pulmonary arterial hypertension can be therapeutically targeted. *Proc Natl Acad Sci USA* 2007; 104: 11418–11423.
- Zhao L, Oliver E, Maratou K, et al. The zinc transporter ZIP12 regulates the pulmonary vascular response to chronic hypoxia. *Nature* 2015; 524: 356–360.
- Bravo R, Gutierrez T, Paredes F, et al. Endoplasmic reticulum: ER stress regulates mitochondrial bioenergetics. *Int J Biochem Cell Biol* 2012; 44: 16–20.
- Schroder M. Endoplasmic reticulum stress responses. Cell Mol Life Sci 2008; 65: 862–894.
- 116. Pinton P, Giorgi C, Siviero R, et al. Calcium and apoptosis: ER-mitochondria Ca2+ transfer in the control of apoptosis. Oncogene 2008; 27: 6407–6418.
- 117. Weir EK, Lopez-Barneo J, Buckler KJ, et al. Acute oxygensensing mechanisms. *N Engl J Med* 2005; 353: 2042–2055.

- 118. Dromparis P, Paulin R, Sutendra G, et al. Uncoupling protein 2 deficiency mimics the effects of hypoxia and endoplasmic reticulum stress on mitochondria and triggers pseudohypoxic pulmonary vascular remodeling and pulmonary hypertension. *Circ Res* 2013; 113: 126–136.
- Voelkel NF, Tuder RM, Bridges J, et al. Interleukin-1 receptor antagonist treatment reduces pulmonary-hypertension generated in rats by monocrotaline. *Am J Resp Cell Mol* 1994; 11: 664–675.
- Bhargava A, Kumar A, Yuan N, et al. Monocrotaline induces interleukin-6 mRNA expression in rat lungs. *Heart Dis* 1999; 1: 126–132.
- 121. Humbert M, Monti G, Brenot F, et al. Increased interleukin-1 and interleukin-6 serum concentrations in severe primary pulmonary hypertension. *Am J Respir Crit Care Med* 1995; 151: 1628–1631.
- 122. Kimura H, Kasahara Y, Kurosu K, et al. Alleviation of monocrotaline-induced pulmonary hypertension by antibodies to monocyte chemotactic and activating factor monocyte chemoattractant protein-1. *Lab Invest* 1998; 78: 571–581.
- 123. Isern RA, Yaneva M, Weiner E, et al. Autoantibodies in patients with primary pulmonary-hypertension association with anti-Ku. *Am J Med* 1992; 93: 307–312.
- 124. Humbert M, Monti G, Brenot F, et al. Increased interleukin-1 and interleukin-6 serum concentrations in severe primary pulmonary-hypertension. *Am J Resp Crit Care* 1995; 151: 1628–1631.
- 125. Urano F, Wang XZ, Bertolotti A, et al. Coupling of stress in the ER to activation of JNK protein kinases by transmembrane protein kinase IRE1. *Science* 2000; 287: 664–666.
- 126. Zhang KZ and Kaufman RJ. From endoplasmic-reticulum stress to the inflammatory response. *Nature* 2008; 454: 455–462.
- 127. Garg AD, Kaczmarek A, Krysko O, et al. ER stress-induced inflammation: does it aid or impede disease progression? *Trends Mol Med* 2012; 18: 589–598.
- 128. Rabinovitch M. Molecular pathogenesis of pulmonary arterial hypertension. *J Clin Invest* 2008; 118: 2372–2379.
- Archer SL, Weir EK and Wilkins MR. Basic science of pulmonary arterial hypertension for clinicians new concepts and experimental therapies. *Circulation* 2010; 121: 2045–2066.
- Zhang L, Ma J, Shen TT, et al. Platelet-derived growth factor (PDGF) induces pulmonary vascular remodeling through 15-LO/15-HETE pathway under hypoxic condition. *Cell Signal* 2012; 24: 1931–1939.
- 131. Paridaens A, Laukens D, Vandewynckel YP, et al. Endoplasmic reticulum stress and angiogenesis: is there an interaction between them? *Liver Int* 2014; 34: e10–18.
- Hu J, Shibata Y, Voss C, et al. Membrane proteins of the endoplasmic reticulum induce high-curvature tubules. *Science* 2008; 319: 1247–1250.
- 133. Zurek N, Sparks L and Voeltz G. Reticulon short hairpin transmembrane domains are used to shape ER tubules. *Traffic* 2011; 12: 28–41.
- 134. Muñoz JP and Zorzano A. Endoplasmic reticulum stress enters a Nogo zone. *Sci Transl Med* 2011; 3: 88ps26.
- 135. Sobolewski A, Rudarakanchana N, Upton PD, et al. Failure of bone morphogenetic protein receptor trafficking in pulmonary arterial hypertension: potential for rescue. *Hum Mol Genet* 2008; 17: 3180–3190.

- 136. Whyte C, Thies F, Peyrol L, et al. N-3 long-chain polyunsaturated fatty acids inhibit smooth muscle cell migration by modulating urokinase plasminogen activator receptor through MEK/ERK-dependent and -independent mechanisms. J Nutr Biochem 2012; 23: 1378–1383.
- Sutendra G and Michelakis ED. The metabolic basis of pulmonary arterial hypertension. *Cell Metabolism* 2014; 19: 558–573.
- Cortez L and Sim. The therapeutic potential of chemical chaperones in protein folding diseases. *Prion* 2014; 8: pii: 28938.
- 139. Uppala JK, Gani AR and Ramaiah KVA. Chemical chaperone, TUDCA unlike PBA, mitigates protein aggregation efficiently and resists ER and non-ER stress induced HepG2 cell death. *Sci Rep* 2017; 7: 3831.
- Kusaczuk M. Tauroursodeoxycholate bile acid with chaperoning activity: molecular and cellular effects and therapeutic perspectives. *Cells* 2019; 8: 1471.
- 141. Perlmutter DH. Chemical chaperones: a pharmacological strategy for disorders of protein folding and trafficking. *Pediatr Res* 2002; 52: 832–836.
- 142. Dandage R, Bandyopadhyay A, Jayaraj GG, et al. Classification of chemical chaperones based on their effect on protein folding landscapes. ACS Chem Biol 2015; 10: 813–820.
- 143. Kolb PS, Ayaub EA, Zhou W, et al. The therapeutic effects of 4-phenylbutyric acid in maintaining proteostasis. *Int J Biochem Cell Biol* 2015; 61: 45–52.
- 144. Wang WJ, Mulugeta S, Russo SJ, et al. Deletion of exon 4 from human surfactant protein C results in aggresome formation and generation of a dominant negative. J Cell Sci 2003; 116: 683–692.
- 145. de Almeida SF, Picarote G, Fleming JV, et al. Chemical chaperones reduce endoplasmic reticulum stress and prevent mutant HFE aggregate formation. J Biol Chem 2007; 282: 27905–27912.
- 146. Chamcheu JC, Navsaria H, Pihl-Lundin I, et al. Chemical chaperones protect epidermolysis bullosa simplex keratinocytes from heat stress-induced keratin aggregation: involvement of heat shock proteins and MAP kinases. *J Invest Dermatol* 2011; 131: 1684–1691.
- 147. Lichter-Konecki U, Diaz G, Merritt JL, et al. Ammonia control in children with urea cycle disorders (UCDs); phase 2 comparison of sodium phenylbutyrate and glycerol phenylbutyrate. *Mol Genet Metab* 2011; 103: 323–329.
- Miller AC, Cohen S, Stewart M, et al. Radioprotection by the histone deacetylase inhibitor phenylbutyrate. *Radiat Environ Biophys* 2011; 50: 585–596.
- 149. Shi X, Zheng C, Li C, et al. 4-Phenybutyric acid promotes gastric cancer cell migration via histone deacetylase inhibition-mediated HER3/HER4 up-regulation. *Cell Biol Int* 2018; 42: 53–62.
- 150. Xiao C, Giacca A and Lewis GF. Sodium phenylbutyrate, a drug with known capacity to reduce endoplasmic reticulum stress, partially alleviates lipid-induced insulin resistance and beta-cell dysfunction in humans. *Diabetes* 2011; 60: 918–924.

- Hong YP, Guo WY, Wang WX, et al. 4-Phenylbutyric acid attenuates pancreatic beta-cell injury in rats with experimental severe acute pancreatitis. *Int J Endocrinol* 2016; 2016: 4592346.
- 152. Monteleone JP, Mokhtarani M, Diaz GA, et al. Population pharmacokinetic modeling and dosing simulations of nitrogen-scavenging compounds: disposition of glycerol phenylbutyrate and sodium phenylbutyrate in adult and pediatric patients with urea cycle disorders. *J Clin Pharmacol* 2013; 53: 699–710.
- 153. Lee B, Rhead W, Diaz GA, et al. Phase 2 comparison of a novel ammonia scavenging agent with sodium phenylbutyrate in patients with urea cycle disorders: safety, pharmacokinetics and ammonia control. *Mol Genet Metab* 2010; 100: 221–228.
- 154. Mimori S, Ohtaka H, Koshikawa Y, et al. 4-Phenylbutyric acid protects against neuronal cell death by primarily acting as a chemical chaperone rather than histone deacetylase inhibitor. *Bioorg Med Chem Lett* 2013; 23: 6015–6018.
- 155. Park CS, Cha H, Kwon EJ, et al. The chemical chaperone 4-phenylbutyric acid attenuates pressure-overload cardiac hypertrophy by alleviating endoplasmic reticulum stress. *Biochem Biophys Res Commun* 2012; 421: 578–584.
- 156. Falasca L, Tisone G, Palmieri G, et al. Protective role of tauroursodeoxycholate during harvesting and cold storage of human liver: a pilot study in transplant recipients. *Transplantation* 2001; 71: 1268–1276.
- 157. Xie Q, Khaoustov VI, Chung CC, et al. Effect of tauroursodeoxycholic acid on endoplasmic reticulum stress-induced caspase-12 activation. *Hepatology* 2002; 36: 592–601.
- 158. Schoemaker MH, Conde de la Rosa L, Buist-Homan M, et al. Tauroursodeoxycholic acid protects rat hepatocytes from bile acid-induced apoptosis via activation of survival pathways. *Hepatology* 2004; 39: 1563–1573.
- 159. Lee YY, Hong SH, Lee YJ, et al. Tauroursodeoxycholate (TUDCA), chemical chaperone, enhances function of islets by reducing ER stress. *Biochem Bioph Res Co* 2010; 397: 735–739.
- 160. da-Silva WS, Ribich S, Arrojo e Drigo R, et al. The chemical chaperones tauroursodeoxycholic and 4-phenylbutyric acid accelerate thyroid hormone activation and energy expenditure. *FEBS Lett* 2011; 585: 539–544.
- 161. Gani AR, Uppala JK and Ramaiah KV. Tauroursodeoxycholic acid prevents stress induced aggregation of proteins in vitro and promotes PERK activation in HepG2 cells. Arch Biochem Biophys 2015; 568: 8–15.
- Paridaens A, Raevens S, Colle I, et al. Combination of tauroursodeoxycholic acid and N-acetylcysteine exceeds standard treatment for acetaminophen intoxication. Liver Int 2017; 37: 748–756.
- 163. Omura T, Asari M, Yamamoto J, et al. Sodium tauroursodeoxycholate prevents paraquat-induced cell death by suppressing endoplasmic reticulum stress responses in human lung epithelial A549 cells. *Biochem Bioph Res Co* 2013; 432: 689–694.