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Impact of type 2 diabetes variants identified through genome-wide association studies in early-onset type 2 diabetes from South Indian population

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The prevalence of early-onset type 2 diabetes (EOT2D) is increasing in Asian countries. Genome-wide association studies performed in European and various other populations have identified associations of numerous variants with type 2 diabetes in adults. However, the genetic component of EOT2D which is still unexplored could have similarities with late-onset type 2 diabetes. Here in the present study we aim to identify the association of variants with EOT2D in South Indian population. Twenty-five variants from 18 gene loci were genotyped in 1,188 EOT2D and 1,183 normal glucose tolerant subjects using the MassARRAY technology. We confirm the association of the *HHEX* variant rs1111875 with EOT2D in this South Indian population and also the association of *CDKN2A/2B* (rs7020996) and *TCF7L2* (rs4506565) with EOT2D. Logistic regression analyses of the *TCF7L2* variant rs4506565(A/T), showed that the heterozygous and homozygous carriers for allele 'T' have odds ratios of 1.47 (95% confidence interval [CI], 1.17 to 1.83; $p = 0.001$) and 1.65 (95% CI, 1.18 to 2.28; $p = 0.006$) respectively, relative to AA homozygote. For the *HHEX* variant rs1111875 (T/C), heterozygous and homozygous carriers for allele 'C' have odds ratios of 1.13 (95% CI, 0.91 to 1.42; $p = 0.27$) and 1.58 (95% CI, 1.17 to 2.12; $p = 0.003$) respectively, relative to the TT homozygote. For *CDKN2A/2B* variant rs7020996, the heterozygous and homozygous carriers of allele 'C' were protective with odds ratios of 0.65 (95% CI, 0.51 to 0.83; $p = 0.0004$) and 0.62 (95% CI, 0.27 to 1.39; $p = 0.24$) respectively, relative to TT homozygote. This is the first study to report on the association of *HHEX* variant rs1111875 with EOT2D in this population.

Keywords: early-onset type 2 diabetes, genome-wide association studies, *HHEX* gene, MassARRAY genotyping, South Asians, *TCF7L2* gene

Introduction

Early-onset type 2 diabetes (EOT2D) a relatively new phenomenon recognized in the past few decades and is caused by the complex interplay between genetic and environmental factors [1,2]. Currently 425 million people are living with diabetes worldwide and the number is expected to reach 629 million by 2045 with nearly 60% of the affected peo-

ple living in Asian countries [3,4]. Asians have an earlier age of diagnosis and a higher prevalence of diabetes for the same body mass index (BMI) than Europeans. India alone is presently home to 72 million people with diabetes [4,5]. Epidemiological studies performed in Indians showed a 25.3% increase in individuals developing type 2 diabetes (T2D) at < 40 years [6]. A recent nationwide population-based study estimating the national prevalence of diabetes and prediabetes in India also indicates 25–34 years as the take-off point for diabetes both in urban and rural areas [7].

Genome-wide association studies (GWAS) and subsequent meta-analyses of these studies have increased the list of T2D associated genetic variants to more than a hundred [8]. However, these variants identified by the large scale GWAS were mostly with the late-onset type 2 diabetes (LOT2D) subtype that develops after 40 years of age. Though the T2D subtype that develops at earlier ages (EOT2D) has a considerably larger heritable component, very few studies have looked at the genetic component of EOT2D. T2D develops at an earlier age a decade or two earlier in Asian Indians and often coincides with the monogenic form of diabetes namely maturity-onset diabetes of the young (MODY) [9,10]. Indeed previous studies have demonstrated association of some MODY variants also with EOT2D [11–13]. Genetic variants in *TCF7L2* [14–16], *HNF1A* [17], *ABCA1* [18], *DIO2* [19], *PCLC* [20], *TRIB3* [21], *ADIPOQ*, and *LEPR* [22] identified with LOT2D in various populations also showed association with EOT2D. Our group also replicated the association of the variants in *TCF7L2*, *CDKN2A/2B* and an intergenic single nucleotide polymorphism (SNP) on chromosome 1p31 identified with LOT2D in various population also with EOT2D in Asian Indians [23]. With this background, the present study was designed to study the association of 25 variants within 18 distinct gene loci, previously identified with the LOT2D subtype in various GWAS, on South Indians with EOT2D.

Methods

Study subjects

The study group comprised of 1,188 unrelated EOT2D subjects and 1,183 normal glucose tolerant (NGT) subjects recruited from Chennai Urban Rural Epidemiology Study (CURES) and from Dr. Mohan's Diabetes Specialties Centre (DMDSC) tertiary diabetes center in Chennai in South India. Subjects for the study were selected based on the World Health Organization (WHO) criteria. NGT was defined as fasting plasma glucose < 100 mg/dL and 2-h post glucose value \leq 140 mg/dL. Diabetes was diagnosed if the fasting plasma glucose was \geq 126 mg/dL or 2-h post glucose value \geq 200 mg/dL or if the participant was on drug therapy for diabe-

tes after diagnosis by a physician. The following criteria were used for selection of EOT2D subjects: patients having early-onset diabetes if they were diagnosed before the age of 35 years, responding to oral hypoglycemic agents, fasting C-peptide > 1.0, stimulated C-peptide > 2.0 pmol/mL, and glutamic acid decarboxylase antibodies negative. Only unrelated individuals were included in this study. Subjects with ketoacidosis at diagnosis, exocrine pancreatic disease (fibrocalculous pancreatic diabetes), pregnant women and subjects known to have confirmed maturity-onset diabetes of the young, were excluded from the study. Written consent was obtained from all the individuals participating in the study and the study was approved by the Institutional Ethics Committee of the Madras Diabetes Research Foundation (RHN/Adhoc/19/2011-2012).

Anthropometric and biochemical measurements

Anthropometric measurements including weight, height, and waist measurements were obtained using standardized techniques. The BMI was calculated using the formula, weight (kg)/(height \times height)(m²). Blood pressure (BP) was measured with a mercury sphygmomanometer (Diamond Deluxe BP apparatus, Pune, India) from the left arm in a sitting position. Fasting plasma glucose (glucose oxidase-peroxidase method), serum cholesterol (cholesterol oxidase-peroxidase-amidopyrine method), serum triglycerides (glycerol phosphate oxidase-peroxidase-amidopyrine method), and high-density lipoprotein cholesterol (direct method polyethylene glycol-pretreated enzymes) was measured using Hitachi-912 Auto analyzer (Hitachi, Mannheim, Germany). Low-density lipoprotein cholesterol was calculated using the Friedewald formula. Glycated hemoglobin was estimated by high-pressure liquid chromatography using the variant machine (Bio-Rad, Hercules, CA, USA) and the intra- and inter-assay coefficient of variation of glycated hemoglobin was less than 10%.

SNP selection

Twenty-five variants representing eighteen different gene loci identified in various GWAS studies including *ADAMTS9* [24], *CDC123* [25], *CDKAL1* [26–28], *CDKN2A/2B* [24,29,30], *COBLL1* [31], *GRB14* [32], *HNF1A* [33], *HNF4A* [34], *IGF2BP2* [26,28], *JAZF1* [24,33], *HHEX* [35], *PPARG* [35], *RBMS1* [36], *SLC30A8* [27], *TCF7L2* [37], *THADA* [24], *TP53INP1* [34], and *TSPAN8* [34] were selected for the study.

Genotyping

Genomic DNA was extracted from peripheral blood leucocytes by proteinase K digestion followed by phenol-chloroform method. Genotyping was done using MassARRAY system (Sequenom, San Diego, CA, USA) following the manufacturer's instructions as

published elsewhere [38]. SpectroTYPER software (Sequenom) automatically called the genotypes and only conservative and moderate calls were accepted for the study. Ten percent of the samples genotyped were replicated and discordance rate observed was less than 0.4% for the replicated samples. All the variants genotyped had call rate ranging between 90%–99%.

Statistical analysis

Hardy-Weinberg equilibrium (HWE) was performed by using Pearson χ^2 statistics in controls for each variant separately. Logistic regression analysis was performed assuming additive model to determine the association between variants and the risk for EOT2D, with and without adjusting for parametric confounders such as age, sex, and BMI using SPSS version 20.0 (IBM Corp., Armonk, NY, USA). The power of the study was estimated using PS Power and Sample Size program (Vanderbilt University, Nashville, TN, USA) calculations (with type I error probability $\alpha = 0.05$). Linkage disequilibrium (LD) and haplotype frequencies were estimated using Haploview software (<http://www.broad.mit.edu/mpg/haploview/>) [39].

Ethical approval

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional ethics committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Results

Clinical and biochemical parameters of the study subjects

Table 1 summarizes the clinical and biochemical parameters of the subjects studied. Mean age of the EOT2D and NGT subjects were 32 ± 6 and 31 ± 8 (mean \pm SD), respectively. The fasting plasma glucose, 2-h post plasma glucose, and glycated hemoglobin were

significantly ($p < 0.001$) higher among the EOT2D subjects when compared with the NGT subjects.

Comparison of minor allele frequencies of the studied polymorphisms in South Indian population with frequencies from the 1000 Genomes Project populations

Minor allele frequencies (MAF) of the SNPs studied in the present study were compared with the reported frequencies of 1000 Genomes Project (Global, European, and South Asian population), representative of the genetic diversity that exists within various population in the world and is shown in Supplementary Table 1. According to the 1000 Genome Project database, MAF of most of the studied SNPs in the present study was similar to the South Asian population, thus supporting the fact that this study population presented a high South Asian component. Similarly, all the SNPs included for the present study were common in South Asian population with allele frequency (MAF) ≥ 0.05 .

LD estimation

LD analysis was performed for SNPs in *IGF2BP2* (rs4402960, rs1470579, and rs6769511), *CDKAL1* (rs4712523, rs4712524, and rs7754840), *JAZF1* (rs868745 and rs849134), and *CDKN2A/2B* (rs564398, rs7020996, and rs2383208). Fig. 1 shows the r^2 values for the studied SNPs. The r^2 values were found to be at least 0.83 between the SNPs in *IGF2BP2*, *CDKAL1*, and *JAZF1*.

Since the r^2 values between SNPs in *CDKN2A/2B* was less than 0.35, haplotypes were constructed and the difference in the haplotype frequencies between cases and controls were analyzed. For the SNPs within *CDKN2A/2B*, although the frequency of the TCA haplotype was higher in EOT2D subjects when compared with the NGT subjects ($p = 0.027$), the significance was lost after Bonferroni correction ($p < 0.05/7 = 0.007$). Table 2 shows the haplotype frequencies of the SNPs within *CDKN2A/2B* gene.

Table 1. Clinical and biochemical characteristics of the study subjects

| Parameter | NGT (n = 1,183) | EOT2D (n = 1,188) | p-value |
|---|-----------------|-------------------|---------|
| Age (yr) | 31 \pm 8 | 32 \pm 6 | 0.022* |
| Body mass index (kg/m ²) | 23.5 \pm 4.8 | 26.0 \pm 5.4 | 0.98 |
| Fasting blood sugar (mg/dL) | 86 \pm 9 | 177 \pm 62 | <0.001 |
| 2-h post plasma glucose (mg/dL) | 99 \pm 19 | 260 \pm 85 | <0.001 |
| Glycosylated hemoglobin (%) | 5.5 \pm 0.5 | 8.9 \pm 1.9 | <0.001 |
| Total cholesterol (mg/dL) | 167 \pm 42 | 173 \pm 46 | 0.002 |
| Log transformed serum triglycerides (mg/dL) | 106 \pm 65 | 168 \pm 145 | <0.001 |
| HDL cholesterol (mg/dL) | 44 \pm 16 | 39 \pm 11 | <0.001 |

Values are presented as mean \pm SD.

NGT, normal glucose tolerant; EOT2D, early onset type 2 diabetes; HDL, high-density lipoprotein; SD, standard deviation.

Association of studied SNPs with EOT2D

Genotypic distributions of all the variants studied were in HWE and none of the variants studied showed monoallelic condition. As shown in Table 3, six SNPs within five distinct loci rs7020996 (*CDKN2A/2B*), rs7607980 (*COBLL1*), rs6769511, rs1470579, rs4402960 (*IGF2BP2*), rs4812829 (*HNF4A*), rs1111875 (*HHEX*) and rs4506565 (*TCF7L2*) were found to be significantly associated ($p < 0.05$) with EOT2D in our South Indian population. However, after Bonferroni correction ($p < 0.05/25 = 0.002$) the association with EOT2D remained significant only for three SNPs within

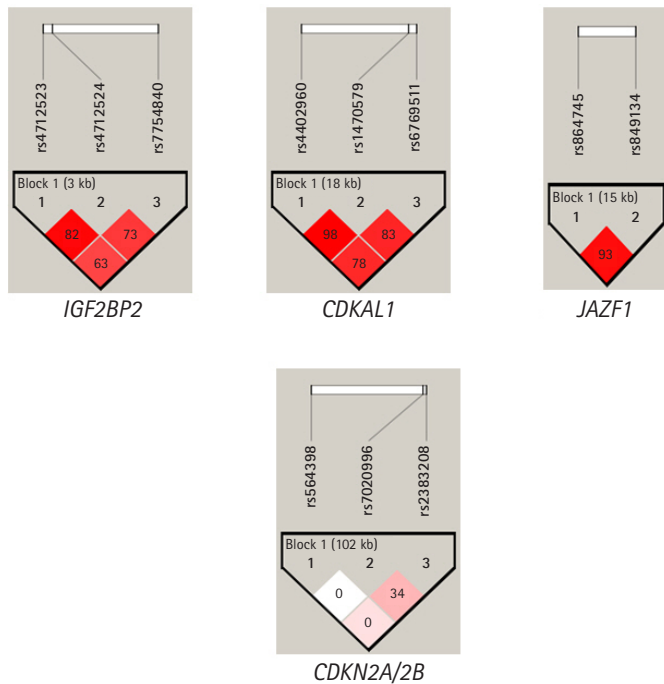


Fig. 1. Linkage disequilibrium plot for the single nucleotide polymorphisms (SNPs) of *IGF2BP2*, *CDKAL1*, *JAF1*, and *CDKN2A/2B*, genes. R^2 values mentioned in the linkage disequilibrium (LD) plot. LD is seen only between SNPs of the same gene, not across the genes.

Table 2. Haplotype frequencies of SNPs in *CDKN2A/2B*

| Haplotype | Cases Freq | Controls Freq | χ^2 | p-value |
|---|------------|---------------|----------|---------|
| <i>CDKN2A/2B</i> (rs564398, rs7020996, rs2383208) | | | | |
| TCA | 0.621 | 0.590 | 4.901 | 0.027* |
| CCA | 0.216 | 0.221 | 0.188 | 0.664 |
| TTG | 0.058 | 0.072 | 3.989 | 0.046 |
| TCG | 0.039 | 0.036 | 0.318 | 0.573 |
| TTA | 0.024 | 0.033 | 3.130 | 0.769 |
| CTG | 0.018 | 0.020 | 0.216 | 0.642 |
| CTA | 0.014 | 0.020 | 1.952 | 0.162 |

SNP, single nucleotide polymorphism.

* $p < 0.05$, statistically significant.

three distinct gene loci rs1111875 (*HHEX*: $p = 2.0 \times 10^{-4}$), rs4506565 (*TCF7L2*: $p = 1.0 \times 10^{-5}$) and rs7020996 (*CDKN2A/2B*: $p = 6.0 \times 10^{-4}$). A tendency to association ($p < 0.05$) with EOT2D was also observed with variants in *COBLL1* (rs7607980), *IGF2BP2* (rs6769511, rs1470579, and rs4402960), and *HNF4A* (rs4812829) in the present study. The risk allele frequencies for all the variants in the EOT2D and NGT subjects are shown in Table 3.

Logistic regression analyses were performed under the additive model for the variants with significant association to EOT2D identified in the present study: rs1111875 (*HHEX*), rs4506565 (*TCF7L2*), and rs7020996 (*CDKN2A/2B*) after adjusting for potential confounders like age, sex, and BMI (Table 4). The heterozygous and homozygous carriers of allele 'T' of the *TCF7L2* variant rs4506565 (A/T) had an odds ratio of 1.47 (95% confidence interval [CI], 1.17 to 1.83; $p = 0.001$) and 1.65 (95% CI, 1.18 to 2.28; $p = 0.006$) respectively relative to AA homozygote. In the case of the *HHEX* variant rs1111875 (T/C), heterozygous and homozygous carriers for allele 'C' had an odds ratio of 1.13 (95% CI, 0.91 to 1.42; $p = 0.27$) and 1.58 (95% CI, 1.17 to 2.12; $p = 0.003$) respectively relative to TT homozygote. However, for the *CDKN2A/2B* variant rs7020996 heterozygous carrier for the 'T' allele showed an association that was protective in nature with odds ratios of 0.65 (95% CI, 0.51 to 0.83; $p = 0.0004$) while the homozygous carrier showed no significant association (OR, 0.62; 95% CI, 0.27 to 1.39; $p = 0.24$) relative to the CC homozygote with EOT2D.

Table 5 shows the comparison of the clinical and biochemical characteristics of NGT subjects and the SNPs associated with EOT2D based on their genotype. For the rs4506565 of the *TCF7L2* gene, NGT subjects homozygous for the 'TT' genotype had increased glycated hemoglobin levels (mean \pm SD, 5.5 ± 0.4) when compared with the carriers of the 'AA' genotype (5.4 ± 0.4 , $p = 0.01$). In case of the *CDKN2A/2B* variant rs7020996, carriers of

Table 3. The allelic distribution of the variants genotyped in cases and controls

| No. | Variant | Gene/nearest gene | Chromosome No. | Risk allele | RAF (%) | | p-value |
|-----|------------------|-------------------|----------------|-------------|---------|-------|--------------------------|
| | | | | | NGT | EOT2D | |
| 1 | rs7607980 (T/C) | <i>COBLL1</i> | 2 | T | 91.8 | 93.7 | 0.010 |
| 2 | rs3923113 (A/C) | <i>GRB14</i> | 2 | A | 82.6 | 84.6 | 0.06 |
| 3 | rs7593730 (C/T) | <i>RBMS1</i> | | C | 77.2 | 79 | 0.14 |
| 4 | rs7578597 (T/C) | <i>THADA</i> | | C | 89.7 | 89.3 | 0.07 |
| 5 | rs1801282 (C/G) | <i>PPARG</i> | 3 | G | 90.1 | 90.3 | 0.82 |
| 6 | rs4607103 (C/T) | <i>ADAMTS9</i> | | C | 46.3 | 47.5 | 0.43 |
| 7 | rs6769511 (T/C) | <i>IGF2BP2</i> | | C | 48.9 | 52.3 | 0.03 ^b |
| 8 | rs1470579 (A/C) | | | A | 47.7 | 50.9 | 0.04 ^b |
| 9 | rs4402960 (G/T) | | | G | 46.7 | 50.7 | 0.01 ^b |
| 10 | rs7754840 (G/C) | <i>CDKAL1</i> | 6 | C | 28.2 | 28.9 | 0.62 |
| 11 | rs4712523 (A/G) | | | G | 29.1 | 30.7 | 0.24 |
| 12 | rs4712524 (A/G) | | | A | 26.3 | 27.8 | 1.08 |
| 13 | rs864745 (T/C) | <i>JAZF1</i> | 7 | T | 73.9 | 75.9 | 0.14 |
| 14 | rs849134 (A/G) | | | A | 74.2 | 76.3 | 0.10 |
| 15 | rs13266634 (C/T) | <i>SLC30A8</i> | 8 | C | 77.5 | 79.1 | 0.20 |
| 16 | rs896854 (C/T) | <i>TP53INP1</i> | | T | 56.4 | 59.1 | 0.07 |
| 17 | rs7020996 (C/T) | <i>CDKN2A/2B</i> | 9 | C | 85.5 | 88.9 | 6.00 × 10 ⁻⁴ |
| 18 | rs2383208 (A/G) | | | A | 86.8 | 87.4 | 0.52 |
| 19 | rs564398 (T/C) | | | C | 78.7 | 79.5 | 0.51 |
| 20 | rs1111875 (C/T) | <i>HHEX</i> | 10 | T | 37 | 42.5 | 2.00 × 10 ^{-4a} |
| 21 | rs4506565 (A/T) | <i>TCF7L2</i> | | T | 30.1 | 36.4 | 1.00 × 10 ^{-5a} |
| 22 | rs10906115 (A/G) | <i>CDC123</i> | | G | 48.4 | 50 | 0.32 |
| 23 | rs1800574 (C/T) | <i>HNF1A</i> | 12 | C | 7.6 | 8.7 | 0.17 |
| 24 | rs4760790 (G/A) | <i>TSPAN8</i> | | G | 36.1 | 38.5 | 0.08 |
| 25 | rs4812829 (A/G) | <i>HNF4A</i> | 20 | A | 32.1 | 35.5 | 0.008 ^b |

RAF, risk allele frequency; NGT, normal glucose tolerant; EOT2D, early onset type 2 diabetes.

^ap-values with significance threshold after Bonferroni correction $p = 0.05/25 = 0.002$.

^bVariants showing only borderline significance ($p < 0.05$).

Table 4. Association of variants with early onset type 2 diabetes with OR and CI (adjusted for age, sex, and BMI)

| Variant | Genotype | Genotype frequency, n (%) | | OR (95% CI) (adjusted for age, sex and BMI) | p-value ^a |
|--------------------------------|----------|---------------------------|------------|---|----------------------|
| | | NGT | EOT2D | | |
| rs4506565 (<i>TCF7L2</i>) | AA | 506 (49.2) | 457 (41.3) | Reference | |
| | TA | 412 (40.0) | 490 (44.3) | 1.47 (1.17–1.83) | 0.001 |
| | TT | 110 (10.7) | 158 (14.3) | 1.65 (1.18–2.28) | 0.006 |
| rs1111875 (<i>HHEX</i>) | TT | 438 (40.3) | 408 (35.7) | Reference | |
| | TC | 493 (45.4) | 505 (44.1) | 1.13 (0.91–1.42) | 0.27 |
| | CC | 156 (14.4) | 231 (20.2) | 1.58 (1.17–2.12) | 0.003 |
| rs7020996 (<i>CDKN2A/2B</i>) | CC | 805 (73.4) | 879 (79.3) | Reference | |
| | TC | 266 (24.2) | 214 (19.3) | 0.65 (0.51–0.83) | 0.0004 |
| | TT | 26 (2.4) | 15 (1.4) | 0.62 (0.27–1.39) | 0.24 |

OR, odds ratio; CI, confidence interval; BMI, body mass index; NGT, normal glucose tolerant; EOT2D, early onset type 2 diabetes.

^ap-values were adjusted for age, sex, and BMI.

Table 5. Clinical and biochemical characteristics of the NGT subjects stratified based on rs4506565 (A/T) and rs7020996 (C/T) genotypes

| Parameter | TCF7L2-rs4506565 (A/T) | | | CDKN2A/2B-rs7020996 (C/T) | | |
|--------------------------------|------------------------|------------|------------------------|---------------------------|-----------------------|-----------------------|
| | AA | AT | TT | CC | TC | TT |
| BMI (kg/m ²) | 23.6 ± 4.8 | 23.7 ± 4.6 | 23.0 ± 4.7 | 23.4 ± 4.6 | 22.9 ± 4.7 | 23.7 ± 6.2 |
| Fasting plasma glucose (mg/dL) | 86 ± 8 | 88 ± 9 | 86 ± 9 | 87 ± 9 ^a | 85 ± 9 | 86 ± 8 |
| 2-h plasma glucose (mg/dL) | 98 ± 20 | 99 ± 19 | 101 ± 18 | 99.11 ± 19 | 98 ± 20 | 97 ± 18 |
| Total cholesterol (mg/dL) | 170 ± 51 | 168 ± 33 | 161 ± 34 | 168 ± 45 | 165 ± 33 | 164 ± 35 |
| Triglycerides (mg/dL) | 106 ± 61 | 111 ± 79 | 97 ± 45 | 104 ± 56 ^b | 107 ± 83 ^b | 117 ± 80 ^b |
| HDL cholesterol (mg/dL) | 45 ± 16 | 45 ± 15 | 44 ± 17 | 44 ± 15 | 44 ± 15 | 47 ± 28 |
| LDL cholesterol (mg/dL) | 101 ± 30 | 101 ± 30 | 96 ± 31 | 101 ± 30 | 99 ± 30 | 93 ± 26 |
| Glycated hemoglobin (%) | 5.4 ± 0.4 | 5.4 ± 0.4 | 5.5 ± 0.4 ^c | 5.4 ± 0.6 | 5.5 ± 0.5 | 5.3 ± 0.8 |

NGT, normal glucose tolerant; BMI, body mass index; HDL, high-density lipoprotein; LDL, low density lipoprotein.

^ap = 0.03 compared to TC (adjusted for age and sex).

^bLog transformed values.

^cp = 0.01 compared to AA (adjusted for age and sex).

the ‘CC’ genotype had significantly higher fasting plasma glucose levels (mean ± SD, 87 ± 9 mg/dL), compared to the carriers of the ‘TC’ genotype (85 ± 9 mg/dL, p = 0.03). None of the other biochemical parameters showed any significant differences among the genotypes in either the NGT or the diabetic subjects.

Discussion

There is a rapid increase in the number of subjects diagnosed with T2D below the age of 40 years. However, only few studies have investigated the association of genetic determinants of LOT2D with EOT2D. The present study aimed at investigating the association of 25 variants from 18 distinct gene loci with EOT2D in this South Indian population, has shown association of variants in *TCF7L2*, *CDKN2A/2B*, and *HHEX* with EOT2D with p-values of 1.00×10^{-5} , 6.00×10^{-4} , and 2.00×10^{-4} respectively with power ranging from 67%–83%.

Transcription factor-7-like 2 (*TCF7L2*) spans 217kb region on chromosome 10q25.3. *TCF7L2* is a transcription factor involved in the Wnt signaling pathway and is expressed not only in the β-cells but also in other cell lineages and glucose- metabolizing tissues, including the liver [40]. *TCF7L2* identified by Grant et al. [41] is the gene with greater susceptibility to LOT2D in various populations [42–48]. Association of the *TCF7L2* variant rs4506565 (A/T) with LOT2D was initially reported by the Wellcome Trust Case Control Consortium with odds ratio of 1.88 (1.56–2.27, p = 5.1×10^{-12}) [37]. The association of rs4506565 (*TCF7L2*) with T2D was later replicated in Middle east [49,50], Tunsanian Arabs [51], Lebanese [52], and Indian population [48,53,54]. While in Europeans, the *TCF7L2* variant rs4506565 showed evidence for association with EOT2D exceeding genome-wide significance, thus clearly establishing *TCF7L2* as a

T2D susceptibility gene of substantial importance [42]. Table 6 shows the comparison of the p-value and odds ratio of the SNPs with association to EOT2D identified in the present study with p-value and odds ratio in other population with T2D [24,35,37,49,52,55,56]. The risk allele frequency of the *TCF7L2* (rs4506565) in South Indian EOT2D subjects was observed to be 36.4%, compared with 37% in North Indian subjects [48], 49% in Saudi Arabian subjects [49], 44% in Tunsanian Arab subjects [51], 46% in Lebanese subjects [52] and 39% in European subjects with LOT2D [37]. In the present study, we have shown a strong association of the *TCF7L2* variant with EOT2D in the South Indian population. A previous study by Chidambaram et al. [23] has shown only marginal association of rs4506565 (*TCF7L2*) with EOT2D in Asian Indians. While, the limitation of the previous study was the small sample size, in the present study, we used a much larger sample size thus increasing the power of the study. Rs4506565 (*TCF7L2*) also showed a significant association with fasting glucose in non-diabetic subjects in European population [57]. A comprehensive pathway analysis with 529 of the 548 genes within 5 kb of a *TCF7L2* binding site by Zhao et al. [58] has shown enriched metabolism-related pathway categories in genes bound by *TCF7L2*. Lyssenko et al. [59] using an adenovirus system showed 2-fold increased expression of *TCF7L2* in human islets, associated with increased insulin gene expression and reduced glucose-stimulated insulin secretion compared with control islets. These studies thus provide evidence for increased expression of *TCF7L2* in human islets with altered insulin but not glucagon secretion. *TCF7L2* also plays a crucial role in coordinating the expression of proinsulin and its subsequent processing to form mature insulin [60]. In mouse models, removal of *TCF4* from B cells in newborn *Tcf7l2*^{-/-} mice and in adult B cell-specific *Tcf7l2* mutants, challenged by fasting or by high-fat diet did not show any affect in their function [61].

Table 6. Comparison of association results for T2D loci in various population with the present study

| Gene | Present study | | European | | Lebanese | | Saudi Arabian | | Japanese | | Korean | |
|-----------|-----------------------|---------------------|-----------------------|--------------------------|----------------------|--------------------------|---------------|--------------------------|----------|-------------------------|-----------------------|--------------------------|
| | p-value | OR (95% CI) | European | OR (95% CI) | p-value | OR (95% CI) | p-value | OR (95% CI) | p-value | OR (95% CI) | p-value | OR (95% CI) |
| rs4506565 | 1.13×10^{-5} | 1.33 (1.17–1.51) | 5.7×10^{-12} | 1.88 (1.56–2.27) [37] | 5.0×10^{-6} | 1.43 (1.20–1.82) [52] | 0.007 | 1.39 (1.09–1.77) [49] | - | - | - | - |
| rs7020996 | 0.0006 | 1.36 (1.14–.63) | 1.8×10^{-7} | - | - | - | - | - | - | - | - | - |
| rs1111875 | 0.0002 | 1.26 (1.11–1.42) | 5.7×10^{-10} | 1.13 (1.09–1.17) [35] | - | - | - | - | 0.0013 | 1.3 (1.11–1.52) [55] | 1.89×10^{-4} | 1.43 (1.18–1.72) [56] |

T2D, type 2 diabetes; OR, odds ratio; CI, confidence interval.

The present study has also confirmed the association of the *CDKN2A/2B* variant rs7020996 with EOT2D in the South Indian population, which was also earlier suggested by Chidambaram et al. [23] in Asian Indian population. The *CDKN2A/2B* locus at chromosome 9p21 was tagged as hot spot for association with LOT2D in a series of GWAS [24,26,30,35]. Zeggini et al. [24] in a meta-analysis study initially reported on the association of rs7020996 of the *CDKN2A/2B* gene with T2D in European population with OR 1.26 (1.15–1.38) ($p = 1.8 \times 10^{-7}$). Though *CDKN2A/2B* was reported to influence diabetes risk across varied ethnicities, not many studies have replicated the association of the *CDKN2A/2B* variant rs7020996 with T2D. Replication of the *CDKN2A/2B* variant rs7020996 both with EOT2D and LOT2D by our own group has shown significant association [23,54].

The risk allele frequency of *CDKN2A/2B* (rs7020996) in South Indian EOT2D subjects was observed to be 88.9% in EOT2D subjects, compared with 88% in Asian Indian EOT2D subjects [23] and 91% in South Indian LOT2D subjects [53]. The *CDKN2A/2B* genes are expressed in adipocytes and pancreatic islets. *CDKN2A* and *CDKN2B* encodes p16INK4a and p15INK4b and inhibit the activity of CDK4 and CDK6, respectively. The p16INK4a encoded by *CDKN2A* is a tumor suppressor and inhibits CDK4 (cyclin-dependent kinase) influencing pancreatic β cell proliferation, through decreased cell mass and subsequent decreased insulin release. The increased insulin demand possibly increases the susceptibility to T2D [43]. In murine models, overexpression of *Cdkn2a* leads to decreased islet proliferation in aging mice and that of *Cdkn2b* leads to islet hypoplasia and diabetes [62]. Study by Kong et al. [63] investigating the mechanism through which the GWAS identified *CDKN2A/2B* variants increase the T2D risk showed the impact of *CDKN2A/2B* SNPs mediated through β -cell mass but not β -cell function.

HHEX, located on chromosome 10q23.33 encodes a 270 amino-acid protein and was identified to be strongly associated with LOT2D in European populations by Scott et al. [35] with odds ratio (OR) 1.13 (1.09–1.17), $p = 5.7 \times 10^{-10}$. The association was later replicated in Danish [43], Japanese [55,64,65], Korean [56], Han Chinese [66], and in Tunisian population [67]. However, studies performed in Indians [53,54,68–70] and African American population [71] failed to replicate the association observed in various populations. The lack of association of the *HHEX* variant among various Indian populations Khatri Sikhs [68], Hyderabad population [69], an endogamous North Indian population [53] and South Indian population [54] could possibly be due to the insufficient sample size, population stratification/admixture or due to confounders. Meta-analysis of 26 studies with 45,792 cases and 65,083 controls, also revealed a stronger association between

rs1111875 and risk for T2D in East Asian population (OR, 1.19) than in white populations (OR, 1.15) and Indian population (OR, 1.13) [72]. Intriguingly, a meta-analysis by Chauhan et al. [73] performed in Indian population successfully replicated the association of the *HHEX* variant with LOT2D in North Indian population. The risk allele frequency of *HHEX* (rs1111875) in South Indian was observed to be 42.3% in EOT2D subjects, compared with 32% in Japanese [55], 36% in Korean [56], 32% in Han Chinese [66], and 52% in European [35] subjects with T2D. Giannini et al. [74] showed association of the *HHEX* variant rs1111875 with prediabetes among obese youth. In European and Finnish population, the rs1111875 (*HHEX*) showed association with lower birth weight providing evidence for the ‘fetal programming hypothesis’ suggestive of decreased insulin secretion or action with reduced intrauterine growth and thereby lower birth weight as well as susceptibility to LOT2D [75,76]. *HHEX* genes encodes a transcription factor involved in the Wnt signaling pathway and also plays important role in many biological processes including cell cycle regulation, organ development, and cell differentiation via both transcriptional activation and repression [77]. Recent functional studies have identified *HHEX* as the first transcription factor required for δ -cell maintenance mediated through paracrine regulation of β -cell activity. The same study also showed misregulated *HHEX* expression with paracrine control of insulin secretion, leading to accelerated β -cell exhaustion and failure [78]. In *HHEX*-null mice pancreatic β cells was defined for its involvement in β -cell differentiation and function and failure of ventral pancreas development [79]. However, the exact mechanism through which the *TCF7L2*, *CDKN2A/2B*, and *HHEX* exerts its effect on T2D is still unclear. Additionally, a recent study by Mohan et al. [80] has also suggested the predominant role of beta-cell dysfunction than insulin resistance in the pathogenesis of T2D among Asian Indian youth.

With regard to the other variants genotyped in the present study rs6769511, rs1470579, rs4402960 of *IGF2BP2*, rs7607980 of *COBLL1*, and rs4812829 of *HNF4A* showed only a nominal association ($p < 0.05$) in terms of the association with EOT2D in this South Indian population with power ranging from 34% to 46%. The nominal association of these variants rs6769511, rs1470579, rs4402960 (*IGF2BP2*), rs7607980 (*COBLL1*), and rs4812829 (*HNF4A*) observed with EOT2D in the present study may however be due to the relatively small sample size, which is one of the major limitations of this study. Moreover, we have replicated only 25 gene variants with EOT2D in the present study out of the several hundred gene variants identified with LOT2D.

The significance of EOT2D is that, due to the earlier onset of diabetes these individuals are at increased susceptibility to complica-

tions of diabetes includes neuropathy, retinopathy, and cardiovascular disease compared to LOT2D. Our results highlight the need for larger prospective studies to identify the effect of genetic variants implicated in the development of EOT2D. To conclude, the present study is the first study to confirm the association of gene variants associated with EOT2D in South Indian population, and shows the importance of the *HHEX* variants with EOT2D.

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Authors' Contribution

Conceptualization: VR. Data curation: VR. Formal analysis: SL. Funding acquisition: VM, VR. Methodology: MC, SL. Writing – original draft: VR, SL. Writing – review & editing: VR, VM.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

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Supplementary Materials

Supplementary data can be found with this article online at <http://www.genominfo.org>.

References

1. Constantino MI, Molyneaux L, Limacher-Gisler F, Al-Saeed A, Luo C, Wu T, et al. Long-term complications and mortality in young-onset diabetes: type 2 diabetes is more hazardous and lethal than type 1 diabetes. *Diabetes Care* 2013;36:3863-3869.
2. Wilmot E, Idris I. Early onset type 2 diabetes: risk factors, clinical

- impact and management. *Ther Adv Chronic Dis* 2014;5:234-244.
3. International Diabetes Federation. *IDF Diabetes Atlas*. 8th ed. Brussels: International Diabetes Federation, 2017.
 4. Anjana RM, Pradeepa R, Deepa M, Datta M, Sudha V, Unnikrishnan R, et al. Prevalence of diabetes and prediabetes (impaired fasting glucose and/or impaired glucose tolerance) in urban and rural India: phase I results of the Indian Council of Medical Research-India DIABetes (ICMR-INDIAB) study. *Diabetologia* 2011;54:3022-3027.
 5. Joshi SR. Diabetes care in India. *Ann Glob Health* 2015;81:830-838.
 6. Praveen PA, Madhu SV, Mohan V, Das S, Kakati S, Shah N, et al. Registry of youth onset diabetes in India (YDR): rationale, recruitment, and current status. *J Diabetes Sci Technol* 2016;10:1034-1041.
 7. Anjana RM, Deepa M, Pradeepa R, Mahanta J, Narain K, Das HK, et al. Prevalence of diabetes and prediabetes in 15 states of India: results from the ICMR-INDIAB population-based cross-sectional study. *Lancet Diabetes Endocrinol* 2017;5:585-596.
 8. Mahajan A, Taliun D, Thurner M, Robertson NR, Torres JM, Rayner NW, et al. Fine-mapping type 2 diabetes loci to single-variant resolution using high-density imputation and islet-specific epigenome maps. *Nat Genet* 2018;50:1505-1513.
 9. Mohan V. Why are Indians more prone to diabetes? *J Assoc Physicians India* 2004;52:468-474.
 10. Gujral UP, Pradeepa R, Weber MB, Narayan KM, Mohan V. Type 2 diabetes in South Asians: similarities and differences with white Caucasian and other populations. *Ann N Y Acad Sci* 2013;1281:51-63.
 11. Anuradha S, Radha V, Deepa R, Hansen T, Carstensen B, Pedersen O, et al. A prevalent amino acid polymorphism at codon 98 (Ala98Val) of the hepatocyte nuclear factor-1alpha is associated with maturity-onset diabetes of the young and younger age at onset of type 2 diabetes in Asian Indians. *Diabetes Care* 2005;28:2430-2435.
 12. Anuradha S, Radha V, Mohan V. Association of novel variants in the hepatocyte nuclear factor 4A gene with maturity onset diabetes of the young and early onset type 2 diabetes. *Clin Genet* 2011;80:541-549.
 13. Ang SF, Tan CSH, Wang L, Dorajoo R, Fong JCW, Kon WY, et al. PAX4 R192H is associated with younger onset of type 2 diabetes in East Asians in Singapore. *J Diabetes Complications* 2019;33:53-58.
 14. Wegner L, Hussain MS, Pilgaard K, Hansen T, Pedersen O, Vaag A, et al. Impact of *TCF7L2* rs7903146 on insulin secretion and action in young and elderly Danish twins. *J Clin Endocrinol Metab* 2008;93:4013-4019.
 15. Dabelea D, Dolan LM, D'Agostino R Jr, Hernandez AM, McAteer JB, Hamman RF, et al. Association testing of *TCF7L2* polymorphisms with type 2 diabetes in multi-ethnic youth. *Diabetologia* 2011;54:535-539.
 16. Silbernagel G, Renner W, Grammer TB, Hugel SR, Bertram J, Kleber ME, et al. Association of *TCF7L2* SNPs with age at onset of type 2 diabetes and proinsulin/insulin ratio but not with glucagon-like peptide 1. *Diabetes Metab Res Rev* 2011;27:499-505.
 17. Hegele RA, Cao H, Harris SB, Hanley AJ, Zinman B. The hepatic nuclear factor-1alpha G319S variant is associated with early-onset type 2 diabetes in Canadian Oji-Cree. *J Clin Endocrinol Metab* 1999;84:1077-1082.
 18. Villarreal-Molina MT, Flores-Dorantes MT, Arellano-Campos O, Villalobos-Comparan M, Rodriguez-Cruz M, Miliar-Garcia A, et al. Association of the ATP-binding cassette transporter A1 R230C variant with early-onset type 2 diabetes in a Mexican population. *Diabetes* 2008;57:509-513.
 19. Nair S, Muller YL, Ortega E, Kobes S, Bogardus C, Baier LJ. Association analyses of variants in the *DIO2* gene with early-onset type 2 diabetes mellitus in Pima Indians. *Thyroid* 2012;22:80-87.
 20. Ma L, Hanson RL, Que LN, Guo Y, Kobes S, Bogardus C, et al. *PCLO* variants are nominally associated with early-onset type 2 diabetes and insulin resistance in Pima Indians. *Diabetes* 2008;57:3156-3160.
 21. Prudente S, Scarpelli D, Chandalia M, Zhang YY, Morini E, Del Guerra S, et al. The *TRIB3* Q84R polymorphism and risk of early-onset type 2 diabetes. *J Clin Endocrinol Metab* 2009;94:190-196.
 22. Liao WL, Chen CC, Chang CT, Wu JY, Chen CH, Huang YC, et al. Gene polymorphisms of adiponectin and leptin receptor are associated with early onset of type 2 diabetes mellitus in the Taiwanese population. *Int J Obes (Lond)* 2012;36:790-796.
 23. Chidambaram M, Liju S, Saboo B, Sathyavani K, Viswanathan V, Pankratz N, et al. Replication of genome-wide association signals in Asian Indians with early-onset type 2 diabetes. *Acta Diabetol* 2016;53:915-923.
 24. Zeggini E, Scott LJ, Saxena R, Voight BF, Marchini JL, Hu T, et al. Meta-analysis of genome-wide association data and large-scale replication identifies additional susceptibility loci for type 2 diabetes. *Nat Genet* 2008;40:638-645.
 25. Shu XO, Long J, Cai Q, Qi L, Xiang YB, Cho YS, et al. Identification of new genetic risk variants for type 2 diabetes. *PLoS Genet* 2010;6:e1001127.
 26. Diabetes Genetics Initiative of Broad Institute of Harvard and MIT, Lund University, and Novartis Institutes of BioMedical Re-

- search, Saxena R, Voight BF, Lyssenko V, Burt NP, de Bakker PI, et al. Genome-wide association analysis identifies loci for type 2 diabetes and triglyceride levels. *Science* 2007;316:1331-1336.
27. Rung J, Cauchi S, Albrechtsen A, Shen L, Rocheleau G, Cavalcanti-Proenca C, et al. Genetic variant near *IRS1* is associated with type 2 diabetes, insulin resistance and hyperinsulinemia. *Nat Genet* 2009;41:1110-1115.
28. Unoki H, Takahashi A, Kawaguchi T, Hara K, Horikoshi M, Andersen G, et al. SNPs in *KCNQ1* are associated with susceptibility to type 2 diabetes in East Asian and European populations. *Nat Genet* 2008;40:1098-1102.
29. Takeuchi F, Serizawa M, Yamamoto K, Fujisawa T, Nakashima E, Ohnaka K, et al. Confirmation of multiple risk loci and genetic impacts by a genome-wide association study of type 2 diabetes in the Japanese population. *Diabetes* 2009;58:1690-1699.
30. Zeggini E, Weedon MN, Lindgren CM, Frayling TM, Elliott KS, Lango H, et al. Replication of genome-wide association signals in UK samples reveals risk loci for type 2 diabetes. *Science* 2007;316:1336-1341.
31. Albrechtsen A, Grarup N, Li Y, Sparso T, Tian G, Cao H, et al. Exome sequencing-driven discovery of coding polymorphisms associated with common metabolic phenotypes. *Diabetologia* 2013;56:298-310.
32. Kooner JS, Saleheen D, Sim X, Sehmi J, Zhang W, Frossard P, et al. Genome-wide association study in individuals of South Asian ancestry identifies six new type 2 diabetes susceptibility loci. *Nat Genet* 2011;43:984-989.
33. Saxena R, Elbers CC, Guo Y, Peter I, Gaunt TR, Mega JL, et al. Large-scale gene-centric meta-analysis across 39 studies identifies type 2 diabetes loci. *Am J Hum Genet* 2012;90:410-425.
34. Voight BF, Scott LJ, Steinthorsdottir V, Morris AP, Dina C, Welch RP, et al. Twelve type 2 diabetes susceptibility loci identified through large-scale association analysis. *Nat Genet* 2010;42:579-589.
35. Scott LJ, Mohlke KL, Bonnycastle LL, Willer CJ, Li Y, Duren WL, et al. A genome-wide association study of type 2 diabetes in Finns detects multiple susceptibility variants. *Science* 2007;316:1341-1345.
36. Qi L, Cornelis MC, Kraft P, Stanya KJ, Linda Kao WH, Pankow JS, et al. Genetic variants at 2q24 are associated with susceptibility to type 2 diabetes. *Hum Mol Genet* 2010;19:2706-2715.
37. Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature* 2007;447:661-678.
38. Gabriel S, Ziaugra L, Tabbaa D. SNP genotyping using the Sequenom MassARRAY iPLEX platform. *Curr Protoc Hum Genet* 2009;Chapter 2:Unit 2.12.
39. Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 2005;21:263-265.
40. Cropano C, Santoro N, Groop L, Dalla Man C, Cobelli C, Galderisi A, et al. The rs7903146 variant in the *TCF7L2* gene increases the risk of prediabetes/type 2 diabetes in obese adolescents by impairing beta-cell function and hepatic insulin sensitivity. *Diabetes Care* 2017;40:1082-1089.
41. Grant SF, Thorleifsson G, Reynisdottir I, Benediktsson R, Manolescu A, Sainz J, et al. Variant of transcription factor 7-like 2 (*TCF7L2*) gene confers risk of type 2 diabetes. *Nat Genet* 2006;38:320-323.
42. Groves CJ, Zeggini E, Minton J, Frayling TM, Weedon MN, Rayner NW, et al. Association analysis of 6,736 U.K. subjects provides replication and confirms *TCF7L2* as a type 2 diabetes susceptibility gene with a substantial effect on individual risk. *Diabetes* 2006;55:2640-2644.
43. Grarup N, Rose CS, Andersson EA, Andersen G, Nielsen AL, Albrechtsen A, et al. Studies of association of variants near the *HHEX*, *CDKN2A/B*, and *IGF2BP2* genes with type 2 diabetes and impaired insulin release in 10,705 Danish subjects: validation and extension of genome-wide association studies. *Diabetes* 2007;56:3105-3111.
44. Cauchi S, El Achhab Y, Choquet H, Dina C, Kremler F, Weitgasser R, et al. *TCF7L2* is reproducibly associated with type 2 diabetes in various ethnic groups: a global meta-analysis. *J Mol Med (Berl)* 2007;85:777-782.
45. Ng MC, Tam CH, Lam VK, So WY, Ma RC, Chan JC. Replication and identification of novel variants at *TCF7L2* associated with type 2 diabetes in Hong Kong Chinese. *J Clin Endocrinol Metab* 2007;92:3733-3737.
46. Tabara Y, Osawa H, Kawamoto R, Onuma H, Shimizu I, Miki T, et al. Replication study of candidate genes associated with type 2 diabetes based on genome-wide screening. *Diabetes* 2009;58:493-498.
47. Bodhini D, Radha V, Dhar M, Narayani N, Mohan V. The rs12255372(G/T) and rs7903146(C/T) polymorphisms of the *TCF7L2* gene are associated with type 2 diabetes mellitus in Asian Indians. *Metabolism* 2007;56:1174-1178.
48. Chandak GR, Janipalli CS, Bhaskar S, Kulkarni SR, Mohankrishna P, Hattersley AT, et al. Common variants in the *TCF7L2* gene are strongly associated with type 2 diabetes mellitus in the Indian population. *Diabetologia* 2007;50:63-67.
49. Acharya S, Al-Elq A, Al-Nafaie A, Muzaaheed M, Al-Ali A. Type 2 diabetes mellitus susceptibility gene *TCF7L2* is strongly associated with hyperglycemia in the Saudi Arabia Population of the eastern province of Saudi Arabia. *Eur Rev Med Pharmacol Sci* 2015;

- 19:3100-3106.
50. O'Beirne SL, Salit J, Rodriguez-Flores JL, Staudt MR, Abi Khalil C, Fakhro KA, et al. Type 2 diabetes risk allele loci in the Qatari population. *PLoS One* 2016;11:e0156834.
 51. Turki A, Al-Zaben GS, Mtraoui N, Marmmuoch H, Mahjoub T, Almawi WY. Transcription factor-7-like 2 gene variants are strongly associated with type 2 diabetes in Tunisian Arab subjects. *Gene* 2013;513:244-248.
 52. Nemr R, Turki A, Echtay A, Al-Zaben GS, Daher HS, Irani-Hakime NA, et al. Transcription factor-7-like 2 gene variants are strongly associated with type 2 diabetes in Lebanese subjects. *Diabetes Res Clin Pract* 2012;98:e23-e27.
 53. Gupta V, Khadawat R, Ng HK, Kumar S, Aggarwal A, Rao VR, et al. A validation study of type 2 diabetes-related variants of the *TCF7L2*, *HHEX*, *KCNJ11*, and *ADIPOQ* genes in one endogamous ethnic group of north India. *Ann Hum Genet* 2010;74:361-368.
 54. Chidambaram M, Radha V, Mohan V. Replication of recently described type 2 diabetes gene variants in a South Indian population. *Metabolism* 2010;59:1760-1766.
 55. Horikoshi M, Hara K, Ito C, Shojima N, Nagai R, Ueki K, et al. Variations in the *HHEX* gene are associated with increased risk of type 2 diabetes in the Japanese population. *Diabetologia* 2007;50:2461-2466.
 56. Lee YH, Kang ES, Kim SH, Han SJ, Kim CH, Kim HJ, et al. Association between polymorphisms in *SLC30A8*, *HHEX*, *CDKN2A/B*, *IGF2BP2*, *FTO*, *WFS1*, *CDKAL1*, *KCNQ1* and type 2 diabetes in the Korean population. *J Hum Genet* 2008;53:991-998.
 57. Dupuis J, Langenberg C, Prokopenko I, Saxena R, Soranzo N, Jackson AU, et al. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. *Nat Genet* 2010;42:105-116.
 58. Zhao J, Schug J, Li M, Kaestner KH, Grant SF. Disease-associated loci are significantly over-represented among genes bound by transcription factor 7-like 2 (*TCF7L2*) in vivo. *Diabetologia* 2010;53:2340-2346.
 59. Lyssenko V, Lupi R, Marchetti P, Del Guerra S, Orho-Melander M, Almgren P, et al. Mechanisms by which common variants in the *TCF7L2* gene increase risk of type 2 diabetes. *J Clin Invest* 2007;117:2155-2163.
 60. Zhou Y, Park SY, Su J, Bailey K, Ottosson-Laakso E, Shcherbina L, et al. *TCF7L2* is a master regulator of insulin production and processing. *Hum Mol Genet* 2014;23:6419-6431.
 61. Boj SF, van Es JH, Huch M, Li VS, Jose A, Hatzis P, et al. Diabetes risk gene and Wnt effector Tcf7l2/TCF4 controls hepatic response to perinatal and adult metabolic demand. *Cell* 2012;151:1595-1607.
 62. Hannou SA, Wouters K, Paumelle R, Staels B. Functional genomics of the *CDKN2A/B* locus in cardiovascular and metabolic disease: what have we learned from GWASs? *Trends Endocrinol Metab* 2015;26:176-184.
 63. Kong Y, Sharma RB, Ly S, Stamateris RE, Jesdale WM, Alonso LC. *CDKN2A/B* T2D genome-wide association study risk SNPs impact locus gene expression and proliferation in human islets. *Diabetes* 2018;67:872-884.
 64. Omori S, Tanaka Y, Takahashi A, Hirose H, Kashiwagi A, Kaku K, et al. Association of *CDKAL1*, *IGF2BP2*, *CDKN2A/B*, *HHEX*, *SLC30A8*, and *KCNJ11* with susceptibility to type 2 diabetes in a Japanese population. *Diabetes* 2008;57:791-795.
 65. Furukawa Y, Shimada T, Furuta H, Matsuno S, Kusuyama A, Doi A, et al. Polymorphisms in the *IDE-KIF11-HHEX* gene locus are reproducibly associated with type 2 diabetes in a Japanese population. *J Clin Endocrinol Metab* 2008;93:310-314.
 66. Lin Y, Li P, Cai L, Zhang B, Tang X, Zhang X, et al. Association study of genetic variants in eight genes/loci with type 2 diabetes in a Han Chinese population. *BMC Med Genet* 2010;11:97. 20550665
 67. Kifagi C, Makni K, Boudawara M, Mnif F, Hamza N, Abid M, et al. Association of genetic variations in *TCF7L2*, *SLC30A8*, *HHEX*, *LOC387761*, and *EXT2* with type 2 diabetes mellitus in Tunisia. *Genet Test Mol Biomarkers* 2011;15:399-405.
 68. Sanghera DK, Ortega L, Han S, Singh J, Ralhan SK, Wander GS, et al. Impact of nine common type 2 diabetes risk polymorphisms in Asian Indian Sikhs: *PPARG2* (Pro12Ala), *IGF2BP2*, *TCF7L2* and *FTO* variants confer a significant risk. *BMC Med Genet* 2008;9:59.
 69. Kommoju UJ, Samy SK, Maruda J, Irgam K, Kotla JP, Velaga L, et al. Association of *CDKAL1*, *CDKN2A/B* & *HHEX* gene polymorphisms with type 2 diabetes mellitus in the population of Hyderabad, India. *Indian J Med Res* 2016;143:455-463.
 70. Phani NM, Adhikari P, Nagri SK, D'Souza SC, Satyamoorthy K, Rai PS. Replication and relevance of multiple susceptibility loci discovered from genome wide association studies for type 2 diabetes in an Indian Population. *PLoS One* 2016;11:e0157364.
 71. Lewis JP, Palmer ND, Hicks PJ, Sale MM, Langefeld CD, Freedman BI, et al. Association analysis in african americans of European-derived type 2 diabetes single nucleotide polymorphisms from whole-genome association studies. *Diabetes* 2008;57:2220-2225.
 72. Wang Y, Qiao W, Zhao X, Tao M. Quantitative assessment of the influence of hematopoietically expressed homeobox variant (rs1111875) on type 2 diabetes risk. *Mol Genet Metab* 2011;102:194-199.

73. Chauhan G, Spurgeon CJ, Tabassum R, Bhaskar S, Kulkarni SR, Mahajan A, et al. Impact of common variants of *PPARG*, *KCNJ11*, *TCF7L2*, *SLC30A8*, *HHEX*, *CDKN2A*, *IGF2BP2*, and *CDKAL1* on the risk of type 2 diabetes in 5,164 Indians. *Diabetes* 2010;59:2068-2074.
74. Giannini C, Dalla Man C, Groop L, Cobelli C, Zhao H, Shaw MM, et al. Co-occurrence of risk alleles in or near genes modulating insulin secretion predisposes obese youth to prediabetes. *Diabetes Care* 2014;37:475-482.
75. Freathy RM, Bennett AJ, Ring SM, Shields B, Groves CJ, Timpson NJ, et al. Type 2 diabetes risk alleles are associated with reduced size at birth. *Diabetes* 2009;58:1428-1433.
76. Andersson EA, Pilgaard K, Pisinger C, Harder MN, Grarup N, Faerch K, et al. Type 2 diabetes risk alleles near *ADCYS*, *CDKAL1* and *HHEX-IDE* are associated with reduced birthweight. *Diabetologia* 2010;53:1908-1916.
77. Arterbery AS, Bogue CW. Hhex is necessary for the hepatic differentiation of mouse ES cells and acts via Vegf signaling. *PLoS One* 2016;11:e0146806.
78. Zhang J, McKenna LB, Bogue CW, Kaestner KH. The diabetes gene Hhex maintains delta-cell differentiation and islet function. *Genes Dev* 2014;28:829-834.
79. Bort R, Martinez-Barbera JP, Beddington RS, Zaret KS. Hex homeobox gene-dependent tissue positioning is required for organogenesis of the ventral pancreas. *Development* 2004;131:797-806.
80. Mohan V, Amutha A, Ranjani H, Unnikrishnan R, Datta M, Anjana RM, et al. Associations of beta-cell function and insulin resistance with youth-onset type 2 diabetes and prediabetes among Asian Indians. *Diabetes Technol Ther* 2013;15:315-322.