

Epidemiology

Ian M. Mackay^{1,2}, Katherine E. Arden^{1,2} and Stephen B. Lambert^{1,2}

¹*Queensland Paediatric Infectious Diseases Laboratory, Sir Albert Sakzewski Virus Research Centre, Queensland Children's Medical Research Institute, Royal Children's Hospital, Brisbane, Australia*

²*Clinical Medical Virology Centre, University of Queensland, Brisbane, Australia*

Abstract

The common cold is the result of an upper respiratory tract infection causing an acute syndrome characterised by a combination of non-specific symptoms, including sore throat, cough, fever, rhinorrhoea, malaise, headache, and myalgia. Respiratory viruses, alone or in combination, are the most common cause. The course of illness can be complicated by bacterial agents, causing pharyngitis or sinusitis, but they are a rare cause of cold and flu-like illnesses (CFLIs). Our understanding of CFLI epidemiology has been enhanced by molecular detection methods, particularly polymerase chain reaction (PCR) testing. PCR has not only improved detection of previously known viruses, but within the last decade has resulted in the detection of many divergent novel respiratory virus species. Human rhinovirus (HRV) infections cause nearly all CFLIs and they can be responsible for asthma and chronic obstructive pulmonary disease exacerbations. HRVs are co-detected with other respiratory viruses in statistically significant patterns, with HRVs occurring in the lowest proportion of co-detections, compared to most other respiratory viruses. Some recently identified rhinoviruses may populate an entirely new putative HRV species; HRV C. Further work is required to confirm a causal role for these newly identified viruses in CFLIs. The burden of illness associated with CFLIs is poorly documented, but where data are available, the impact of CFLIs is considerable. Individual infections, although they do not commonly result in more severe respiratory tract illness, are associated with substantial direct and indirect resource use. The product of frequency and burden for CFLIs is likely to be greater in magnitude than for any other respiratory syndrome, but further work is required to document this. Our understanding of the viral causes of CFLIs, although incomplete, has improved in recent years. Documenting burden is also an important step in progress towards improved control and management of these illnesses.

Introduction

The common cold is the result of an upper respiratory tract infection (URTI) resulting in an acute syndrome best described as cold and flu-like illness (CFLI). It is characterised by a combination of non-specific symptoms, including sore throat, cough, fever, rhinorrhoea, malaise, headache

and myalgia. It is usually due to infection by one or more of many viruses detected in the respiratory tract [1]. Bacterial commensals or those causing pharyngitis or sinusitis may complicate clinical CFLI diagnoses due to an overlap in detection or symptoms but overall, bacteria are rare causes of CFLI [2, 3] and are not reviewed here.

In 2001, the first of many divergent novel respiratory virus species were described for the first time with the aid of polymerase chain reaction (PCR)-based molecular techniques. Discovery of human metapneumovirus (HMPV) [4] was followed by other newly identified viruses (NIVs) including the human coronaviruses (HCoVs) NL63 [5] and HKU1 [6], human bocavirus (HBoV) [7] and many new human rhinovirus (HRV) strains [8, 9] populating an entirely new putative HRV species; HRV C [10, 11]. Some NIVs are yet to be clearly associated with specific clinical syndromes, but all have been detected in patients with CFLIs [12]. Because there have been no case-controlled studies of the common cold for some time, it is unclear what the combined impact of molecular diagnostic testing for respiratory viruses and the increasing number of NIVs will be for our understanding of the syndrome, but it is likely to be significant.

In this chapter we review the epidemiology of the common cold. This includes several aspects of the incidence and disease distribution of the common cold by discussing which, when and how the viral causative agents are detected, focussing on the HRVs. We briefly examine causal association of some complications following CFLIs which include asthma [13] and chronic obstructive pulmonary disease (COPD) [14] and describe the impact and cost of CFLIs.

Epidemiology of viral causes of CFLIs

The viruses consistently causing most CFLIs are HRVs [15] comprising 50–80% [3, 16] of relevant symptomatic respiratory illnesses. However, the human influenza viruses (IFVs; IFAV, IFBV and IFCV), the human parainfluenza viruses (HPIVs, 1–4), the HCoVs, 229E and OC43, human respiratory syncytial virus (HRSV), human adenoviruses (HAdV) and human enteroviruses (HEV) [17] have also been associated with 8–15% of CFLIs [18–20], despite some being traditionally considered more ‘serious’ causes of respiratory syndromes, including acute lower respiratory tract illnesses (LRTIs).

Factors affecting the circulation patterns and clinical impact of respiratory viruses

The reported peak activity and rate of different viral infections associated with CFLIs varies with the manner in which illnesses are defined, recorded,

documented aetiologically, and tracked longitudinally [21]. However, historical detection rate data do not comprehensively represent HRV circulation patterns because sequential infections by different strains occur and may appear as unbroken symptomatic episodes during a single observation period [22, 23]; an occurrence which is rarely examined. In other instances, multiple HRV strains can be isolated [24, 25] or detected [26–28] from a single specimen, indicating a capacity for HRV co-infection which is similarly overlooked.

Studies seeking to explain epidemics of the colloquially termed ‘respiratory viruses’ often attribute them to the season [29]. Commonly, the circulation pattern for each virus that can recur annually is dominated by a different strain or species, which changes each year depending on the nature of pre-existing immunity within that location’s population. When we documented the seasonal characteristics of HMPV detections over 4 years, we found that among over 700 HMPV-positive specimens, the four genetically defined HMPV subtypes exchanged dominance each year and detection frequencies cycled up to, or down from, their peak at other times [30, 31]. Herd immunity contributes to controlling epidemics of viruses, especially those which elicit strong and long-lasting immune responses in their hosts. The age of a population is therefore a significant factor in the epidemiology of respiratory viruses. In general terms, adults suffer fewest severe outcomes from respiratory virus infection and children most. In the very young a portion of the pathology of severe illness can be attributed to the small and developing airways. Individual immunity is accrued over time for the adults but is relatively weak among neonates and young children since it is during the early years of life that this response is developed by repeated exposure to infection. A relative increase in the prevalence of symptomatic illnesses is also seen among the elderly, often attributed to the waning of immunity with age.

For some respiratory viruses, including IFV and HRSV, a strict pattern of peak activity occurring with colder or wetter weather is common [33–35]. Exceptions can occur during which higher temperatures and greater daily temperature fluctuations parallel the respiratory virus epidemic period [34]. Some of these epidemic peaks can be seen in Figure 1 exemplified by HRSV, HMPV and IFAV detected in a paediatric hospital-based population during 2003. The defined peaks usually recur at a similar time each year, whereas HEVs and HAdVs are more consistently present and the HRVs, apart from dominating the overall number of detections in this population, often peak in spring and autumn. HRSV activity also correlates with complex interactions of latitude, temperature, humidity and UVB radiance [29]. Apart from weather conditions, cohorting of populations can also occur during return from long school or university holidays, which is a particularly common trigger for HRV epidemics in the young [36–39]. The accompanying increased risk of transmission due to aerosols in close quarters and shared contact with contaminated surfaces are implicated as the cause of

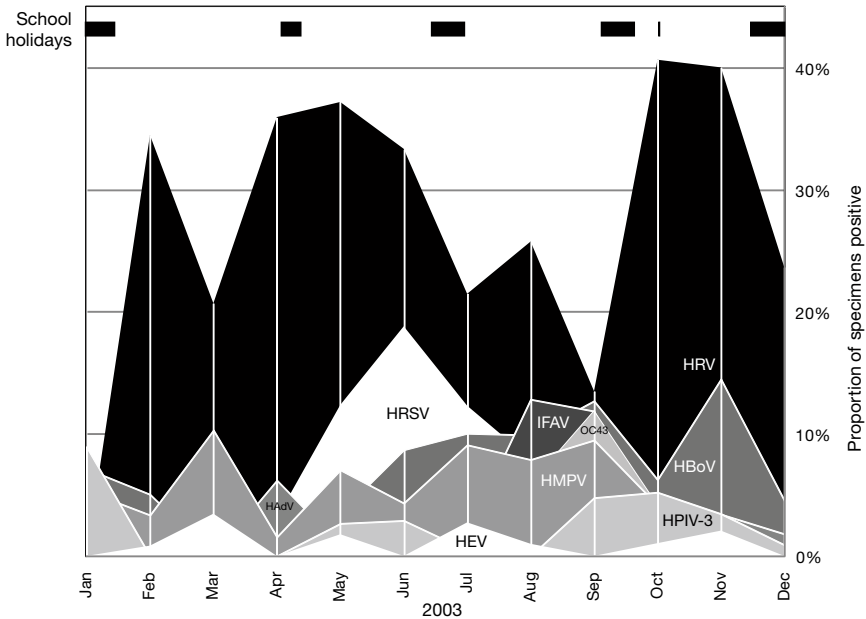


Figure 1. Virus detections plotted by month for 2003. The virus detected is indicated, as are school holidays in Queensland, Australia, during the year of the study at this location. Data were derived from a paediatric hospital-based, in- and outpatient population [8, 32].

rapid increases in the numbers of symptomatic illnesses. Susceptibility to CFLIs may also be directly influenced by weather conditions affecting the respiratory epithelium [40].

HRV detections occur throughout the year but are usually seen to peak in spring and autumn [41–48] depending on the method of detection, the length of the study period and the type of population investigated [22, 49]. One study indicated that HRVs of any given strain might be sporadically detected ahead of an epidemic, providing warning of the impending wide-spread activity by that strain [50]. Few studies examine whether every strain recurs each year at a single location or whether herd immunity protects against reinfection by a previous epidemic strain, and if so, how long such an effect might last. In Brisbane (Queensland, Australia) during 2003, HRV B strains circulated during winter and HRV C strains predominantly circulated during spring [8]. In contrast, the HRV As occurred in all seasons.

The role of common cold viruses in potentiating bacterial infections

It has long been known that bacterial adherence is enhanced by preceding infection of a respiratory virus [51], particularly HRSV and IFAV [51]. Such

studies began after the influenza pandemic of 1918 [52] during which death due to secondary bacterial infection was a significant contributor to total morbidity. Since then, prior infection by IFAV and HRV has been shown to increase the number of staphylococci, streptococci and pneumococci adhering to a pharyngeal cell line, while measles virus decreased adherence and HAdV had no effect [53]. Possible mechanisms, identified using animal studies, include IFV neuraminidase-mediated removal of sialic acid moieties permitting bacteria access to otherwise hidden receptor molecules [52, 54] on the cell surface, and expression of haemagglutinin which enhances group A streptococcal binding [55]. HRV-14 infection can also increase subsequent bacterial adherence by up-regulating streptococcal receptor expression on human tracheal cells, partly mediated by transcription factor activation after HRV infection [56]. Both infectious HRV-16 and HRV-2 reduced the capacity of human alveolar macrophages to respond to lipopolysaccharide and lipoteichoic acids *in vitro* [57], which may, *in vivo*, permit worsening of an HRV infection *via* concomitant bacterial superinfection. In children, but not adults, peak HRSV activity was significantly and positively correlated with peak *Streptococcus pneumoniae*-mediated pneumococcal disease activity in Australia and New Zealand [58, 59]. In the Netherlands, both children and adults were found to have higher rates of pneumococcal and meningococcal disease during peak IFV and HRSV seasons [35]. Vaccination to prevent pneumococcal disease successfully prevented nearly one third of cases of pneumonia associated with the major respiratory viruses [60], presumably by preventing bacterial super infections.

Methods of detecting viruses in CFLIs

Robust detection methods that are kept up-to-date have been of paramount importance to our evolving understanding of the epidemiology of the CFLIs. Co-culture of patient secretions with “permissive” cells lines has been the longest serving method of detection but it is now well known for being insensitive [48, 61, 62]. Examples of previously unknown respiratory viruses believed to be endemic rather than recently emerged are being reported with increasing frequency. For the HRVs this was exemplified by the molecular identification of a large number of highly divergent, and at writing, unculturable HRV C strains [8, 10, 11, 32]. Insensitive and inefficient testing of the HRV super-group is to be blamed for delaying characterisation of the rhinoviruses, thus leading to a significant underestimation of the total number and nature of strains, which has been undeniably detrimental for all previous epidemiology studies of CFLI. With the introduction of nucleic acid testing and improvements to PCR product detection methods, a quantum leap in respiratory virus detection frequencies has been achieved. Even so, no single assay has been shown to robustly detect all HRVs and no panel of assays has achieved 100% laboratory diagnoses. Time to specimen

delivery was once thought to be a cause for reduced aetiologies but even the use of PCR, not requiring infectious pathogens, still misses a large proportion of suspected infections [63].

Cell culture methods

Cell culture techniques are limited by poor sensitivity due to slow growth or poorly cytopathic viruses, reduced viability due to poor specimen handling, narrow detection windows, complex result interpretation requiring high levels of operator expertise, host immunosuppression, antimicrobial therapies, high levels of background signal and non-specific cross-reactions [64, 65]. Nonetheless, both microbial culture and rapid immunofluorescence assays can be used to produce valuable epidemiological data, reveal new, uncharacterised or atypical microbes and yield intact or infectious organisms for further study [66].

Viruses have been isolated in cell cultures since the 1950s but diagnostic services were limited for a further two decades [67]. Microscopic examination of degenerative changes brought about by virus replication (cytopathic effects), a sometimes slow and always technically demanding skill, was later augmented by haemadsorption tests to identify the extent of haemagglutinating protein expression, which indicates the replication of certain respiratory viruses. Subsequently, shell vial methods were employed, which, when used with specific antibodies, identified viral antigens in 1–2 days compared to 2–10 days for haemagglutination methods [68]. In 1953 Andrewes and colleagues at the Common Cold Unit (Salisbury, UK) described the first isolation of an HRV strain [69, 70]. Later, improved culture systems permitted viral replication to be more easily identified and maintained [71, 72]. Nonetheless, even using cell-culture conditions normally favouring the appearance of cytopathicity, instances of non-cytopathic HRV strains have been found by other methods [73], which may have included HRV C-like viruses.

Because the respiratory tract is a cellularly diverse environment and because a wide variety of viruses with diverse tropism cause CFLIs, cell culture methods require the use of a broad range of cell types. For methods to be useful, they must encompass virus concentrations ranging from 10^1 to 10^5 TCID₅₀/ml [74–77]. Additionally, successful isolation and higher viral yields require monitoring of cell age after plating (< 72 h), inoculum volume, culture medium pH (6.8–7.3) and cell density [78–81]. Therefore, culture can be expensive, not just for the labour required to inoculate, maintain under sometimes fastidious conditions [70, 80, 82–84] and monitor the cultures, but also to ensure that the diverse range of cell stocks and culture media are available and fresh. Even with these criteria met, HRVs and most of the respiratory NIVs have proven to be very poor targets for isolation methods based on cell culture [67]. Despite the challenges [85], virus isolation is

reportedly a more sensitive indicator of infection than an antibody rise in paired sera [86].

Antibody-based methods

To date, antibody-based methods have proven the most diversely commercialised and robust diagnostic format either for the indirect detection of a host response to a respiratory virus, or the direct detection of viral antigen in culture or from infected cells present in specimens, such as nasopharyngeal aspirates or bronchoalveolar lavage. Antibody-based results augment both general diagnostic molecular data and those data provided by research studies aiming at better characterising respiratory viruses. Apart from speed, cost-benefit and familiarity, an obvious advantage derived from use of a protein-based system is the existence of conserved antigenic regions among related viruses; regions that do not vary significantly among strains of the same species or other relevant taxonomic grouping. Such conservation is infrequently reflected at the nucleotide level making these regions troublesome targets for nucleic acid-based systems but ideal for antibodies. Unfortunately, antigenic conservation can also be manifested as cross-reaction; difficulty discriminating between infections caused by closely related viruses. Such discrimination is important when searching for the role of each individual respiratory virus in illness [87].

Antigen detection methods may be performed with or without a biological amplification step such as *in vitro* cell culture. If culture is not being employed, then it is necessary to collect cellular specimens since the cells confine virions to a small, easily identified space that aids/allows immunofluorescent detection; but such cellular specimens are not always available. Rapid respiratory virus antigen detection is relatively insensitive and, depending on the clinical priorities for the particular virus, negative results may need to be confirmed using another assay, which largely abrogates the benefits of speed [88, 89]. Furthermore, antibody-based methods have not kept up with the recent flurry of NIVs and so reliable diagnostic reagents are not available for the latest viral discoveries [67].

Seroclassification or 'serotyping' of an HRV infection was once the gold standard for strain identification of the 'common cold viruses' but serotyping became impractical as the number of distinct strains grew beyond convenience [79, 90]. Antibodies are essential for strain-specific neutralisation of infection [91], techniques around which the HRV nomenclature system evolved in 1967 [92]. These determine whether co-incubation of a characterised antibody with a preparation of an unknown virus can preclude its cellular entry and replication. If successful, the antibody chosen confers some degree of identification upon the unknown virus. Such techniques have found that a large number of distinct strains circulate each year and that a selection of them predominate in a given season, replaced by others in

subsequent years [70, 93]. Today, PCR-based sequencing methods can do the same job at the genetic level with increased objectivity and speed compared to the complex and lengthy neutralisation methods [94, 95].

Polymerase chain reaction

The improved sensitivity of PCR-based assays dramatically increased the frequency of viral detection compared to cultivation methods [96], which has meant that many previous studies are incomparable to today's findings. This improvement is especially noticeable for the HRVs [3, 26, 42, 97–99] but also for other viruses that are fastidious or, to date, impossible, to culture. Because of PCR it is becoming commonplace to find reports of HRVs predominating in CFLIs [100–102], despite the incomplete validation of published assays against all picornavirus (HRV and HEV) strains using clinical material. Nonetheless, many assays successfully detect the currently circulating HRV strains at levels as low as 10^2 TCID₅₀/sample. This amount is commonly shed during experimental infections [103, 104]. Because HRV strains are now being detected beyond their traditionally understood symptomatic context of the CFLI syndrome [16, 105], it is becoming more important to define a qualitative and quantitative correlation between HRV nucleic acid detection and the presence of infectious virus at the sampling site. Unfortunately, the latter is problematic when using PCR to study respiratory viruses because of the inability to normalise the amount of starting RNA template [106].

Improved detection by PCR compared with traditional methods means that less invasive specimen types can be used for research, and in most circumstances, diagnostic testing. For example, prior to PCR, there were problems with the sensitivity of RSV detection using less-invasive specimen types. Using antigen detection, a reduction in positives by approximately one-third was seen when nasal swabs were used, compared with nasopharyngeal aspirates [107, 108]. Use of PCR has largely overcome this issue [109], to the point where less-invasive specimen types can be easily collected by lay people in community settings for research purposes [47, 110], or used instead of invasive techniques in clinic or outpatient settings [109].

When they are included in the PCR testing menu, HRVs raise the frequency of pathogen detection above one per sample [111]. Studies find that HRV strains are very frequent contributors to co-infections [112] and co-detections [113], sometimes presenting this in terms of their minor contributing role in serious respiratory disease [112, 113]. More likely this reflects the insensitivity of old cell culture-based methods that simply failed to propagate many HRV strains and in the process created paradigms for the HRVs that reduced their profile for further study. In one study, half of all HRV detections were found concurrently with another virus, on the surface, a significant fraction, and yet 80% or more of HRSV, HMPV, HEV

and IFV detections and 71% of HCoV-NL63 detections were found in the company of another virus [114].

The use of a multiplex real-time PCR (m-rtPCR) or a suite of individual rtPCR assays [113] that encompass the majority of regularly detected viral targets is being steadily embraced by diagnostic laboratories that receive respiratory secretions and a number of these panels include a capacity to detect HRVs [115]. Multiplexing PCRs increases result throughput and reduces costs associated with labour and time but also requires significant research and developmental time and may still perform at a reduced clinical sensitivity compared to individual assays.

Innovative, but less well evaluated, multitarget molecular laboratory tools now exist including the MultiCode-PLx system, which employs a synthetic nucleobase pair, multiplex PCR and microsphere flow cytometry [116]. It permits the discrete detection of 17 respiratory viral targets and two assay controls, although it returns an unusually low HRV detection rate. Similar technology also provides a sensitive, 20-target, 2-step RT-PCR-based assay [117]. The Seeplex[®] respiratory virus detection kit targets 12 respiratory viruses [118] using dual priming oligonucleotides [119] and detecting the amplicon by capillary electrophoresis (Seegene Inc.). It has compared favourably to culture-based testing [120]. The ResPlex II assay (Qiagen) employs a proprietary multiplex RT-nPCR [121] approach followed by amplicon detection using a Luminex suspension array to identify 12 targets [122, 123]. The xTAG[™] respiratory viral panel combines PCR and the Luminex array system and detects more than 20 different targets including controls (Luminex Corporation). PCR amplicon detection by MassTag technology can discriminate 20–30 viral and bacterial agents of illness [124] using oligonucleotides tagged with a unique compound that is released *via* a photolabile link (Qiagen). The MassTag approach has been able to detect HRV C strains [9, 26, 125]. Microarrays can detect thousands of viral targets (US\$ 30–300 per sample) but still require a pre-hybridisation PCR amplification because of their insufficient sensitivity to directly detect viral nucleic acids from clinical specimens. Arrays are still low-throughput, high-turnaround time diagnostic options. At their most robust, microarrays, like PCR, rely on the existence of conserved regions of sequence to detect unknown viruses and they too can detect previously unknown HRV strains [126], although nothing vastly different from what is already known. Rapid protein- or virion-based assays are not (yet) adequately sensitive [127, 128].

PCR does have some downsides, some of which have been mentioned already. Detection of microbial genomic nucleic acids cannot yield the same information about infectivity as cell culture, but there have been good correlations reported between infectivity and viral genome detection for yellow fever virus [129] and in a comprehensive birth cohort study characterising frequent respiratory infections in which PCR data were found to correlate very well with symptomatic respiratory illness [130]. Despite the ‘closed’

nature of the modern generation of rtPCR techniques, they are PCR-based and as such still subject to contamination by amplicon from previous runs and template from extraction areas or infected technologists. Efficient PCR relies entirely on conserved sequence targets and thus an extensive foreknowledge of each virus is being sought. If the region targeted by oligonucleotides is subject to genetic variation, PCR will continue the diagnostic trend towards underestimating viruses in CFLIs. Even for conserved targets, PCR primer pair designs that yield a single specific amplicon from clinical specimen extracts can be extremely difficult to achieve when faced with the highly variable cellular and microbial content of respiratory tract specimens and the sequence similarities between viruses and humans for some targets. Non-specificity can render quantification methods useless [131] as can the absence of suitable housekeeping gene targets to permit normalisation of viral nucleic acid input. Because the success of PCR has led to an increase in the number of virus detections and a reduction in the number of virions required for a positive result, positive PCR methods are sometimes greeted with scepticism due to them being perceived as too sensitive. Such concerns must be addressed by careful epidemiology.

Questions raised by the co-detection of viruses among CFLIs

When thorough screening is conducted for all relevant viruses in each specimen, multiple virus detections are a frequent result. In particular, this has been the case since the more widespread adoption of PCR as the diagnostic method of choice because it is significantly more sensitive than the traditional methods of culture and direct or indirect fluorescent antibody assays. PCR is also better than other diagnostic methods at rapidly and specifically discriminating multiple targets representing different viral genes or strains, fuelling an increasing number of reports of microbial co-detections in 20% or more of specimens [114, 132–137]. For viral co-detections that include HRSV, interferon gamma (IFN- γ) levels are reduced [100]. This suggests a mechanism of immune intervention that creates a beachhead in the host's innate immune response, which subsequently permits additional viruses to gain a foothold, thereby increasing the frequency of co-detections. Although a description of all of a patient's viruses is necessary before the significance of co-detections can be determined, it does complicate the interpretation of results and the traditional assignation of a "causal" virus. Historically, to save labour and costs, causality has been associated with a "first-past-the-post" approach in which the initial virus to be detected is assigned the causal role [122]. Many laboratories have yet to completely adopt PCR, and so the occurrence of co-detections is not globally acknowledged, further complicating their impact compared to single detections.

What the detection of more than one virus, as well as the particular mix of viruses involved, means to the clinical outcome is controversial, with

studies describing illness severity that is worsened [138–140] or unchanged [137, 141] by multiple detections. Among infants hospitalised with bronchiolitis, there was a 2.7-fold increased likelihood of infants with viral co-detections being admitted to a paediatric intensive care unit than those with single detections [114]. Considering their ubiquity, it is interesting that relatively low numbers of concurrent detections of other respiratory viruses occur with HRV strains [47, 142]. In fact, HRV strains are co-detected with other pathogens in reproducible, but clinically undefined, patterns [111]. Nonetheless, there is an increasing number of single HRV detections being made from patients with significant LRTIs and with acute otitis media [143]: it is becoming clearer that the HRV infection process can directly cause illness and that HRVs are not merely passengers in the clinical outcome of the infection [44].

The increasing proportion of viruses found in the company of other viruses, and also with bacteria, raises some interesting questions. Is it possible that a certain number, or certain mix, of viruses, or both, is necessary to tip the host into a state of symptomatic illness? This question may be especially relevant for viruses traditionally thought of as causing more mild respiratory illness, such as the HAdVs and perhaps HBoV. It is noteworthy that the proportion of asymptomatic episodes decreases with the number of micro-organisms detected and increases with age in children [138]. If not the nature then perhaps it is the order of infection that is important as has been suggested for some viral and bacterial pairings. This is poorly addressed by examining data from the clinical microbiology laboratory since such testing is only a cross-sectional snapshot of the host's condition. To address this question accurately, carefully planned, longitudinal cohort studies are required. In a study of 27 children during the first year of life who contracted five or more moderate-to-severe respiratory illnesses, it was apparent that the same viral species or strain did not usually recur during a 12-month period [130]. Another question is whether infection by one virus or bacterium predisposes the host to infection by one or more others.

The proportion of asymptomatic PCR positives is virus specific and occurs in more than a third of children during a CFLI season [144]. A particularly confounding and relevant issue for viral epidemiology is that raised by the criteria used to define an illness in some studies. Some criteria are so stringent that they may miss mild, but nonetheless common and virally induced CFLI symptoms such as headaches [144]. Such omissions are likely to contribute to the number of 'asymptomatic' cases reported by some investigations [114] and to the severity scores used for studies linking single and multiple detections to clinical outcome. It might be simple coincidence that two or more viruses can be detected in the same specimen, reflecting an overlap of their seasonal peaks [114] when it is more likely that hosts will come into contact with more than one virus in the community. However, we believe this is not the case for two reasons. We have not seen a

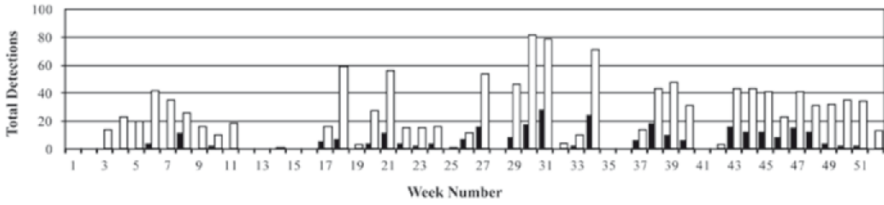


Figure 2. The total number of virus detections (open bars) and co-detections (filled bars) during each week of 2003. Viruses tested included, HRSV, HMPV, IFAV, IFBV, HAAdV, HBoV, HPIVs, non-SARS HCoVs and respiratory picornaviruses. Data are derived from [8] and [32].

seasonal trend towards more co-detections in certain seasons (Fig. 2) but we have seen patterns that indicate virus-specific factors drive the association between co-detected viruses.

When we statistically analysed co-detections from earlier studies of HRV-QPM [32], which also included screening for traditional respiratory viruses and NIVs, we identified that patterns existed that particularly involved the association of certain viruses. Specifically, HRVs were the virus or virus group with the lowest statistically significant proportion of co-detections. We believe that this may be an example of a strong HRV interference effect. Others have shown that epidemics of HRSV may be interrupted or apparently staved off by an epidemic of IFV [33, 145]. A possible mechanism for the separation often seen between an epidemic peak due to one virus and that from another could be competition between different viruses for replication in the same host cells or tissues or for use of the same, or very similar, receptor molecules required for infection. Interference may also be due to the nature of the immune response elicited by the infected host in response to infection by the first virus [73, 146]. Seasonal variation in the prevalence of any virus may be influenced by interference, whereby the peak prevalence of one respiratory virus impedes or prevents the processes that let other viruses establish themselves at the same time, in the same host population [145].

Despite extensive investigation of respiratory specimens taken from patients requiring hospitalisation, oxygen therapy and/or drug treatment, we noted the retention of a large proportion (34%) of specimens from which no virus could be detected [8]. Other studies have found similar frequencies of negative specimens and such findings indicate the likely existence of yet-to-be characterised viral causes of respiratory illness. Extrapolating from all the known respiratory viruses and recent research findings, it is reasonable to assume that any new agents of respiratory disease will be associated with CFLIs as well as possibly more severe disease in some populations and also both as sole agents and in the company of other viruses and bacteria.

Associations between acute virus infections and chronic respiratory disease

CFLI is linked with a number of more serious clinical conditions that may require hospitalisation, invasive testing procedures and the use of drugs and other supportive measures. An URTI may develop into a LRTI or it may acutely exacerbate pre-existing chronic conditions including asthma and COPD. Such exacerbations mask CFLI epidemiology by favouring the clinical diagnosis of the LRTI. Acute LRTIs contribute to more morbidity and mortality than HIV infection, malaria, cancer or heart attack [147] worldwide. Because of equivalent isolation frequencies from well and ill children, the presence of potential bacterial pathogens cannot reliably be correlated with LRT symptoms [145].

As we stated earlier in the chapter, many respiratory viruses that are associated with serious disease are also associated with milder common cold-like illnesses; the converse is also true. Viruses, especially the HRVs, that were previously deemed to be capable of causing only mild illness [138] are now being frequently associated with costly and distressing illnesses and CFLI complications. In particular, respiratory viruses often cause more severe LRT symptoms in neonates and infants, because of airway swelling, excessive secretions and smooth muscle contraction in their narrow immature airways resulting from infection [148].

The importance of HRV infection associated with LRT morbidity during the first year of life is both significant [13] and underappreciated [149]. HRVs replicate in non-nasal tissues including smooth muscle [150] and bronchial epithelial cells [151, 152]. In addition, the immunopathological effect of viral replication in the upper airways may be transmitted systemically [148]. If HRVs naturally replicate in the LRT, as has been reported [83], then a local host inflammatory effect is a likely pathogenic mechanism.

In one example, a German birth cohort study found a positive association between repeated LRTI (pneumonia, bronchitis, pertussis, tracheo-bronchitis, 'flu', croup and bronchitis) before the age of 3 and wheeze at the age of 7 [13]. Nonetheless, the impact of LRTIs on immune development and the contribution of genetic predisposition to LRTIs remain unclear [14, 153, 154]. This study also found a significant inverse relationship between recurrent "runny nose" episodes and subsequent atopic sensitisation, and these repeated infections imparted most of their protective effect during the first year of life [13]. A study of infants found that a sixth of HRV isolate-positive patients exhibited symptoms of LRTIs (mostly wheezing) [155]. In adults ≥ 40 years of age, the duration of symptoms and frequency of LRTIs associated with HRV isolation starts to increase with age [156].

Although HRVs have been associated with threefold more LRT and wheezy LRT illnesses than HRSV [149], the risk of obstructive airway disease is similar whether an HRV or HRV and HRSV are detected [100].

Studies of children in hospital-based populations usually report more significant clinical outcomes, especially those relating to the LRT [157]. These data can be considered a condensed sampling of illness among community-based populations but conclusions should be interpreted cautiously. LRT illness has also been identified in other age and patient groups [74, 91, 99, 100, 155, 158–161]; nonetheless, hospital-based populations retain importance for probing the potential of a virus to cause severe clinical outcomes, especially due to a first infection. This environment provides cases with the strongest influence on future prioritisation of therapeutic developments [145].

HRVs and expiratory wheezing exacerbations

Acute wheezing episodes (including bronchiolitis and acute asthma, which share similar pathologies) are a common epidemic and seasonal LRT manifestation of respiratory virus infection of the URT and LRT of children from all ages, but especially among males and during the first year of life [145, 158, 162, 163]. The mechanisms underlying the induction or exacerbation of asthma are not yet fully understood [148, 164] but wheezing is blamed for excessive use of antibiotics, for being the primary cause of hospitalisation among children and, rarely, for death [48, 165, 166]. Exacerbations of asthma and COPD are often preceded by a symptomatic rather than asymptomatic HRV episode [166–171], although, in some instances, an exacerbation is the only evidence of symptoms [172]. Reduced peak expiratory volume in children is especially associated with detection of respiratory picornaviruses [170].

Traditionally, it is HRSV infection that is causally associated with expiratory wheezing because of the virus's well-known ability to infect the LRT, but periods of epidemic wheezing unaccompanied by high rates of HRSV detection are common [163, 173]. The Childhood Origins of Asthma Study (COAST) used sampling criteria that were designed to intentionally investigate the role of HRSV in illness, but instead of HRSV, the data indicated that HRVs were the most important predictor of subsequent wheezing in early childhood [174, 175]. Although the total number of symptomatic respiratory illnesses did not differ significantly, asthmatics had more HRV infections, while their siblings had more bacterial infections. Since asthmatics are more often treated with antibiotics, bacterial detection rates may be falsely lowered in some reports [176]. Significantly higher rates of HRV detection with more obvious LRT symptoms are more common in asthmatic children than in non-asthmatic populations [166, 176–178]. History of asthma in children also appears to be a risk factor for more frequent symptomatic viral infections. However, the presence of atopy or allergy does not appear to be a common feature [162, 166] since only a small proportion of allergic children have asthma [179].

Impact and cost of the common cold

For any illness or syndrome, mapping the epidemiology and burden of disease is needed for a number of reasons, but key amongst them is prioritising the need for prevention, treatment, and further research efforts. There are three pieces of evidence required by those developing health policies in assessing whether to recommend or implement a publicly funded prevention or treatment program: epidemiology of the targeted illness, the efficacy of the intervention, and the cost effectiveness of the intervention [180]. Evaluations of cost effectiveness consist of a number of key components, including how common the illness is, the cost associated with illness, and the cost of any intervention, either prevention or treatment [181]. Given the ubiquitous nature of the common cold syndrome, there has been little attention paid to documenting impact. This is a feature colds have in common with the less frequent, but more severe end of the respiratory infection spectrum. Based on estimates from the Global Burden of Disease study, acute respiratory diseases, despite being one of the largest contributors to disability-adjusted life-years (DALYs), receive a discouragingly low proportion of health-related research funds [182].

The value used in cost-effectiveness evaluations is a product of counts of illness and impact of individual illness, often presented as DALYs [181]. Even though CFLIs have lower severity compared with complicated URTIs and LRTIs, due to the frequency their burden cannot be ignored. Acute respiratory infection incidence is highest in the first 2 years of life, with up to 13 episodes per year, and it is not uncommon to average close to one infection per child-month [130, 183]. Whereas illnesses can often be managed in the community with supportive care from parents, complications requiring a medical visit in which antibiotic therapy is prescribed, such as otitis media (30%) and sinusitis (8%), are common [184]. In pre-school aged children, nearly 50% of general practitioner visits are for acute respiratory infections [185], many of which will only involve self-limiting URT symptoms.

The availability of preventive vaccines and therapeutic antivirals means that inter-pandemic influenza is the most studied of respiratory viruses associated with the cold. Estimates around the cost impact of other respiratory viruses are rare – particularly compared to their relative frequency. Some estimates about the cost impact of non-influenza viruses are available from the US. Using a telephone survey of over 4000 households, researchers collected self-reported incidence and resource use during non-influenza, viral respiratory infections [186]. These figures were extrapolated to the US population and costs attached to resource use. The direct costs associated with viral respiratory infections were US\$17 billion annually, with these being outweighed by the indirect cost burden of US\$22.5 billion. The indirect cost component was made up of missed workdays due to illness, totalling 70 million days, and missed workdays while caring for a household member, totalling 189 million days [186]. The annual cost burden of antibiotic use for

acute respiratory tract illness in the US is over US\$1.3 billion alone [187]. This compares with a recent modelling assessment of seasonal influenza suggesting annual costs US\$87.1 billion, with 83% of this cost due to annual deaths [188]. Information about HRSV impact is more common than other non-influenza viruses, but pertains mainly to those groups of children who are currently eligible for preventive interventions: those born prematurely with associated lung disease, or with specific congenital cardiopulmonary malformations [189–191]. A US study using three national databases and an assumption that 15% of all acute otitis media was due to HRSV calculated direct medical costs from HRSV to be over US\$1.3 billion (2002 dollars) per annum, with 98% of these costs associated with illness in the less than 5-year age group [192].

Although national data are rare, community-level impact is even less commonly measured. Two recent community-level studies have included an assessment of acute respiratory illness in children using a sensitive definition for influenza-like illness [47, 110, 193–195]. The threshold for burden data collection for study children could be met with a combination of two non-specific symptoms, such as nasal stuffiness and decreased activity [47, 194]. Standard costs were applied to burden data to derive a syndrome cost [193] and a virus-specific cost of illness [195]. A mean cost for community-managed illness from each study was AUD\$241 from the 2001 pilot study [193], and AUD\$309 from the 2003/2004 main study [195] (average exchange rates during study period: United Kingdom pound £1 = AUD\$2.49, Euro €1 = AUD\$1.73, and US\$1 = AUD\$1.50) [196]. The main study included an influenza season of higher than normal activity with H3N2 influenza A (drifted strain subtype A/Fujian/411/2002-like) being the predominant circulating type [197]. Virus-specific cost of illness for all viruses other than influenza fell within a relatively narrow band, and picornaviruses (not further differentiated) had an mean cost of AUD\$267 per illness [195]. A recent UK study looking at the cost impact of individual cough illnesses in children aged 3–59 months, without detailed recording of indirect costs, reported a mean cost per episode to the National Health Service (NHS) of £27, a mean cost for the family of £15, and an annual cost to the NHS £31.5 million [198].

These findings show that, although there are some data on illnesses associated with more serious outcomes and specific viruses, there continues to be little in the way of targeted research at the national or community level documenting the simple burden associated with the common cold. Future community-based studies into the common cold and associated respiratory tract illness, integrating epidemiology and economic methods, are required [199].

Conclusions

The common cold is the syndromic child of many parents. The nature of the child, its epidemiology, severity, and impact, is determined by interaction of host, pathogen, and environmental effects. HRVs are the agents most commonly associated with CFLIs, but other respiratory viruses, including influenza viruses and HRSV, can be associated with the syndrome. The recent expansion in the use of PCR has brought improved detection of known viruses, but also detection of NIVs. Through these means, the diagnostic gap in all respiratory illnesses is reduced. The contribution of HRV Cs in respiratory illness appears to overshadow that of other known RVs; however, it is difficult to judge given the paucity of data from the other species, and further documentation of HRV epidemiology and impact are research priorities for the coming years. Although our knowledge of the causes of CFLIs has improved in past few years, the collation of impact data is some way behind. Documenting burden is an important step in the progress towards improved control and management of these illnesses.

Acknowledgements.

This work was possible because of funding from NH&RMC project grant 455905 and RCHF Project Seeding grant 10281.

References

- 1 Wat D (2004) The common cold: A review of the literature. *Eur J Intern Med* 15: 79–88
- 2 Mäkelä MJ, Puhakka T, Ruuskanen O, Leinonen M, Saikku P, Kimpimäki M, Blomqvist S, Hyypiä T, Arstila P (1998) Viruses and bacteria in the etiology of the common cold. *J Clin Microbiol* 36: 539–542
- 3 Arruda E, Pitkäranta A, Witek TJ, Doyle CA, Hayden FG (1997) Frequency and natural history of rhinovirus infections in adults during autumn. *J Clin Microbiol* 35: 2864–2868
- 4 van den Hoogen BG, de Jong JC, Groen J, Kuiken T, de Groot R, Fouchier RAM, Osterhaus ADME (2001) A newly discovered human pneumovirus isolated from young children with respiratory tract disease. *Nat Med* 7: 19–24
- 5 van der Hoek L, Pyrc K, Jebbink MF, Vermeulen-Oost W, Berkhout RJM, Wolthers KC, Wertheim-van Dillen PME, Kaandorp J, Spaargaren J, Berkhout B (2004) Identification of a new human coronavirus. *Nat Med* 10: 368–373
- 6 Woo PCY, Lau SKP, Chu C-M, Chan K-H, Tsoi H-W, Huang Y, Wong BHL, Poon RWS, Cai JJ, Luk W-K et al. (2005) Characterization and complete genome sequence of a novel coronavirus, coronavirus HKU1, from patients with pneumonia. *J Virol* 79: 884–895
- 7 Allander T, Tammi MT, Eriksson M, Bjerkner A, Tiveljung-Lindell A, Andersson

- B (2005) Cloning of a human parvovirus by molecular screening of respiratory tract samples. *Proc Nat Acad Sci USA* 102: 12891–12896
- 8 Arden KE, McErlean P, Nissen MD, Sloots TP, Mackay IM (2006) Frequent detection of human rhinoviruses, paramyxoviruses, coronaviruses, and bocavirus during acute respiratory tract infections. *J Med Virol* 78: 1232–1240
- 9 Lamson D, Renwick N, Kapoor V, Liu Z, Palacios G, Ju J, Dean A, St GK, Briese T, Lipkin WI (2006) MassTag polymerase-chain-reaction detection of respiratory pathogens, including a new rhinovirus genotype, that caused influenza-like illness in New York State during 2004–2005. *J Infect Dis* 194: 1398–1402
- 10 Lau SKP, Yip CCY, Tsoi H-W, Lee RA, So L-Y, Lau Y-L, Chan K-H, Woo PCY, Yuen K-Y (2007) Clinical features and complete genome characterization of a distinct human rhinovirus genetic cluster, probably representing a previously undetected HRV species, HRV-C, associated with acute respiratory illness in children. *J Clin Microbiol* 45: 3655–3664
- 11 McErlean P, Shackleton LA, Andrewes E, Webster DR, Lambert SB, Nissen MD, Sloots TP, Mackay IM (2008) Distinguishing molecular features and clinical characteristics of a putative new rhinovirus species, human rhinovirus C (HRV C). *PLoS One* 3: e1847
- 12 Esposito S, Bosis S, Niesters HG, Tremolati E, Sabatini C, Porta A, Fossali E, Osterhaus AD, Principi N (2008) Impact of human bocavirus on children and their families. *J Clin Microbiol* 46: 1337–1342
- 13 Illi S, von ME, Lau S, Bergmann R, Niggemann B, Sommerfeld C, Wahn U (2001) Early childhood infectious diseases and the development of asthma up to school age: A birth cohort study. *BMJ* 322: 390–395
- 14 Hershenson MB, Johnston SL (2006) Rhinovirus infections: More than a common cold. *Am J Respir Crit Care Med* 174: 1284–1285
- 15 Heikkinen T, Järvinen A (2003) The common cold. *Lancet* 361: 51–59
- 16 Johnston SL, Sanderson G, Pattemore PK, Smith S, Bardin PG, Bruce CB, Lambden PR, Tyrrell DAJ, Holgate ST (1993) Use of polymerase chain reaction for diagnosis of picornavirus infection in subjects with and without respiratory symptoms. *J Clin Microbiol* 31: 111–117
- 17 Turner RB (1998) The common cold. *Pediatr Ann* 27: 790–795
- 18 Pappas DE, Hendley JO, Hayden FG, Winther B (2008) Symptom profile of common colds in school-aged children. *Pediatr Infect Dis J* 27: 8–11
- 19 Eccles R (2007) Mechanisms of symptoms of the common cold and influenza. *Br J Hosp Med* 68: 578–582
- 20 Pizzichini MMM, Pizzichini E, Efthimiadis A, Chauhan AJ, Johnston SL, Hussack P, Mahony J, Dolovich J, Hargreave FE (1998) Asthma and natural colds. *Am J Respir Crit Care Med* 158: 1178–1184
- 21 Lemanske RF, Gern JE, Gangnon RE (2006) Viral specimen collection by parents increases response rate in population-based virus studies. *J Allergy Clin Immunol* 117: 956–957
- 22 Phillips CA, Melnick JL, Grim CA (1968) Rhinovirus infections in a student population: Isolation of the five new serotypes. *Am J Epidemiol* 87: 447–456
- 23 Minor TE, Dick EC, Peterson JA, Docherty DE (1974) Failure of naturally acquired rhinovirus infections to produce temporal immunity to heterologous serotypes. *Infect Immun* 10: 1192–1193

- 24 Cooney MK, Kenny GE (1977) Demonstration of dual rhinovirus infection in humans by isolation of different serotypes in human heteroploid (HeLa) and human diploid fibroblast cell cultures. *J Clin Microbiol* 5: 202–207
- 25 Cooney MK, Hall CB, Fox JP (1972) The Seattle virus Watch. III. Evaluation of isolation methods and summary of infections detected by virus isolations. *Am J Epidemiol* 96: 286–305
- 26 Renwick N, Schweiger B, Kapoor V, Liu Z, Villari J, Bullmann R, Miething R, Briese T, Lipkin WI (2007) A recently identified rhinovirus genotype is associated with severe respiratory-tract infection in children in Germany. *J Infect Dis* 196: 1754–1760
- 27 Lee W-M, Kiesner C, Pappas T, Lee I, Grindle K, Jartti T, Jakiela B, Lemanske RF, Shult PA, Gern JE (2007) A diverse group of previously unrecognized human rhinoviruses are common causes of respiratory illness in infants. *PLoS One* 2: e966
- 28 Peltola V, Waris M, Österback R, Susi P, Ruuskanen O, Hyypiä T (2008) Rhinovirus transmission within families with children: Incidence of symptomatic and asymptomatic infections. *J Infect Dis* 197: 382–389
- 29 Yusuf S, Piedimonte G, Auais A, Demmler G, Krishnan S, van Caseele P, Singleton R, Broor S, Parveen S, Avendano L et al. (2007) The relationship of meteorological conditions to the epidemic activity of respiratory syncytial virus. *Epidemiol Infect* 135: 1077–1090
- 30 Mackay IM, Waliuzzaman Z, Chidlow GR, Fegredo DC, Laingam S, Adamson P, Harnett GB, Rawlinson W, Nissen MD, Sloots TP (2004) Use of the P gene to genotype human metapneumovirus identifies 4 viral subtypes. *J Infect Dis* 190: 1913–1918
- 31 Mackay IM, Bialasiewicz S, Jacob KC, McQueen E, Arden KE, Nissen MD, Sloots TP (2006) Genetic diversity of human metapneumovirus over 4 consecutive years in Australia. *J Infect Dis* 193: 1630–1633
- 32 McErlean P, Shackleton LA, Lambert SB, Nissen MD, Sloots TP, Mackay IM (2007) Characterisation of a newly identified human rhinovirus, HRV-QPM, discovered in infants with bronchiolitis. *J Clin Virol* 39: 67–75
- 33 Glezen WP, Paredes A, Taber LH (1980) Influenza in children relationship to other respiratory agents. *JAMA* 243: 1345–1349
- 34 Chew FT, Doraisingam S, Ling AE, Kumarasinghe G, Lee BW (1998) Seasonal trends of viral respiratory tract infections in the tropics. *Epidemiol Infect* 121: 121–128
- 35 Jansen AG, Sanders EA, Van der Ende A, van Loon AM, Hoes AW, Hak E (2008) Invasive pneumococcal and meningococcal disease: Association with influenza virus and respiratory syncytial virus activity? *Epidemiol Infect* 136: 1448–1454
- 36 Johnson HE, Altman R, Hamre D, Ward T (1964) Viral infections and the common cold. *Chest* 45: 46–53
- 37 Al-Sunaidi M, Williams CH, Hughes PJ, Schnurr DP, Stanway G (2007) Analysis of a new human parechovirus allows the definition of parechovirus types and the identification of RNA structural domains. *J Virol* 81: 1013–1021
- 38 Hamre D, Connelly AP, Procknow JJ (1966) Virologic studies of acute respira-

- tory disease in young adults. IV. Virus isolations during four years of surveillance. *Am J Epidemiol* 83: 238–249
- 39 Johnston NW, Johnston SL, Norman GR, Dai J, Sears MR (2006) The September epidemic of asthma hospitalization: School children as disease vectors. *J Allergy Clin Immunol* 117: 557–562
- 40 Deal EC Jr, McFadden ER Jr, Ingram RH Jr, Breslin FJ, Jaeger JJ (1980) Airway responsiveness to cold air and hyperpnea in normal subjects and in those with hay fever and asthma. *Am Rev Respir Dis* 121: 621–628
- 41 Winther B, Hayden FG, Hendley JO (2006) Picornavirus infections in children diagnosed by RT-PCR during longitudinal surveillance with weekly sampling: Association with symptomatic illness and effect of season. *J Med Virol* 78: 644–650
- 42 Vesa S, Kleemola M, Blomqvist S, Takala A, Kilpi T, Hovi T (2001) Epidemiology of documented viral respiratory infections and acute otitis media in a cohort of children followed from two to twenty-four months of age. *Pediatr Infect Dis J* 20: 574–581
- 43 Silva MJ, Ferraz C, Pissarra S, Cardoso MJ, Simões J, Vitór AB (2007) Role of viruses and atypical bacteria in asthma exacerbations among children in Oporto (Portugal). *Allergol Immunopathol* 35: 4–9
- 44 Miller EK, Lu X, Erdman DD, Poehling KA, Zhu Y, Griffin MR, Hartvert TV, Anderson LJ, Weinberg GA, Hall CB et al. (2007) Rhinovirus-associated hospitalizations in young children. *J Infect Dis* 195: 773–781
- 45 Gwaltney JM Jr, Hendley JO, Simon G, Jordan WS Jr (1966) Rhinovirus infections in an industrial population. I. The occurrence of illness. *N Engl J Med* 275: 1261–1268
- 46 Fox JP, Cooney MK, Hall CE (1975) The Seattle virus watch. V. Epidemiologic observation of rhinovirus infections, 1965–1969 in families with young children. *Am J Epidemiol* 101: 122–143
- 47 Lambert SB, Allen KM, Druce JD, Birch CJ, Mackay IM, Carlin JB, Carapetis JP, Sloots TP, Nissen MD, Nolan TM (2007) Community epidemiology of human metapneumovirus, human coronavirus NL63, and other respiratory viruses in healthy preschool-aged children using parent-collected specimens. *Pediatrics* 120: e929–e937
- 48 Jartti T, Lehtinen P, Vuorinen T, Österback R, van den Hoogen B, Osterhaus ADME, Ruuskanen O (2004) Respiratory picornaviruses and respiratory syncytial virus as causative agents of acute expiratory wheezing in children. *Emerg Infect Dis* 10: 1095–1101
- 49 Wald TG, Shult P, Krause P, Miller BA, Drinka P, Gravenstein S (1995) A rhinovirus outbreak among residents of a long-term care facility. *Ann Intern Med* 123: 588–593
- 50 Dick EC, Blumer CR, Evans AS (1967) Epidemiology of infections with rhinovirus types 43 and 55 in a group of university of Wisconsin student families. *Am J Epidemiol* 86: 386–400
- 51 Hament JM, Kimpen JL, Fleer A, Wolfs TF (1999) Respiratory viral infection predisposing for bacterial disease: A concise review. *FEMS Immunol Med Microbiol* 26: 189–195

- 52 Peltola VT, McCullers JA (2004) Respiratory viruses predisposing to bacterial infections: Role of neuraminidase. *Pediatr Infect Dis J* 23: S87–S97
- 53 Selinger DS, Reed WP, McLaren LC (1981) Model for studying bacterial adherence to epithelial cells infected with viruses. *Infect Immun* 32: 941–944
- 54 McCullers JA, Bartmess KC (2003) Role of neuraminidase in lethal synergism between influenza virus and *Streptococcus pneumoniae*. *J Infect Dis* 187: 1000–1009
- 55 Okamoto S, Kawabata S, Nakagawa I, Okuno Y, Goto T, Sano K, Hamada S (2003) Influenza A virus-infected hosts boost an invasive type of *Streptococcus pyogenes* infection in mice. *J Virol* 77: 4104–4112
- 56 Ishizuka S, Yamaya M, Suzuki T, Takahashi H, Ida S, Sasaki T, Inoue D, Sekizawa K, Nishimura H, Sasaki H (2003) Effects of rhinovirus infection on the adherence of *Streptococcus pneumoniae* to cultured human airway epithelial cells. *J Infect Dis* 188: 1928–1939
- 57 Oliver BG, Lim S, Wark P, Laza-Stanca V, King N, Black JL, Burgess JK, Roth M, Johnston SL (2008) Rhinovirus exposure impairs immune responses to bacterial products in human alveolar macrophages. *Thorax* 63: 519–525
- 58 Watson M, Gilmour R, Menzies R, Ferson M, McIntyre P (2006) The association of respiratory viruses, temperature, and other climatic parameters with the incidence of invasive pneumococcal disease in Sydney, Australia. *Clin Infect Dis* 42: 211–215
- 59 Murdoch DR, Jennings LC (2009) Association of respiratory virus activity and environmental factors with the incidence of invasive pneumococcal disease. *J Infect* 58: 37–46
- 60 Madhi SA, Klugman KP (2004) A role for *Streptococcus pneumoniae* in virus-associated pneumonia. *Nat Med* 10: 811–813
- 61 Kaiser L, Aubert J-D, Pache J-C, Deffernez C, Rochat T, Garbino J, Wunderli W, Meylan P, Yerly S, Perrin L et al. (2006) Chronic rhinoviral infection in lung transplant recipients. *Am J Respir Crit Care Med* 174: 1392–1399
- 62 Larson HE, Reed SE, Tyrrell DAJ (1980) Isolation of rhinoviruses and coronaviruses from 38 colds in adults. *J Med Virol* 5: 221–229
- 63 Denny FW, Clyde WA Jr (1986) Acute lower respiratory tract infections in nonhospitalized children. *J Pediatr* 108: 635–646
- 64 Whelen AC, Persing DH (1996) The role of nucleic acid amplification and detection in the clinical microbiology laboratory. *Annu Rev Microbiol* 50: 349–373
- 65 Carman WF, Wallace LA, Walker J, McIntyre S, Noone A, Christie P, Millar J, Douglas JD (2000) Rapid virological surveillance of community influenza infection in general practice. *Br Med J* 321: 736–737
- 66 Ogilvie M (2001) Molecular techniques should not now replace cell culture in diagnostic virology laboratories. *Rev Med Virol* 11: 351–354
- 67 Leland DS, Ginocchio CC (2007) Role of cell culture for virus detection in the age of technology. *Clin Microbiol Rev* 20: 49–78
- 68 Mahony JB (2008) Detection of respiratory viruses by molecular methods. *Clin Microbiol Rev* 21: 716–747
- 69 Andrewes CH, Chaponiere DM, Gompels AEH, Pereira HG, Roden AT

- (1953) Propagation of common-cold virus in tissue cultures. *Lancet* 265: 546–547
- 70 Andrewes CH (1966) Rhinoviruses and common colds. *Annu Rev Med* 17: 361–370
- 71 Pelon W, Mogabgab WJ, Phillips LA, Pierce WE (1957) A cytopathogenic agent isolated from naval recruits with mild respiratory illnesses. *Proc Soc Exp Biol Med* 94: 262–267
- 72 Price WH (1956) The isolation of a new virus associated with respiratory clinical disease in humans. *Proc Natl Acad Sci USA* 42: 892–896
- 73 Olson LC, Willhight M, Buescher EL (1972) Recovery and characterization of non-cytopathogenic rhinoviruses. *J Gen Virol* 17: 237–240
- 74 Douglas RG Jr, Cate TR, Gerone PJ, Couch RB (1966) Quantitative rhinovirus shedding patterns in volunteers. *Am Rev Respir Dis* 94: 159–167
- 75 D’Alessio DJ, Meschievitz CK, Peterson JA, Dick CR, Dick EC (1984) Short-duration exposure and the transmission of rhinoviral colds. *J Infect Dis* 150: 189–194
- 76 Cate TR, Couch RB, Fleet WF, Griffith WR, Gerone PJ, Knight V (1965) Production of tracheobronchitis in volunteers with rhinovirus in a small-particle aerosol. *Am J Epidemiol* 81: 95–105
- 77 Hendley JO, Wenzel RP, Gwaltney JM Jr (1973) Transmission of rhinovirus colds by self-inoculation. *N Engl J Med* 288: 1361–1364
- 78 Sethi SK (1978) Reproducible plaquing system for rhinovirus serotypes in HeLa cells – Agarose suspension. *Acta Virol* 22: 60–65
- 79 Gwaltney JM Jr (1966) Micro-neutralization test for identification of rhinovirus serotypes. *Proc Soc Exp Biol Med* 122: 1137–1141
- 80 Behbehani AM, Lee LH (1964) Growth, plaque production and cationic stabilization of rhinovirus type 1 (Echovirus 28). *J Bacteriol* 88: 1608–1611
- 81 Fiala M, Kenny GE (1966) Enhancement of rhinovirus plaque formation in human heteroploid cell cultures by magnesium and calcium. *J Bacteriol* 92: 1717–1715
- 82 Parsons R, Tyrrell DAJ (1961) A plaque method for assaying some viruses isolated from common colds. *Nature* 189: 640–642
- 83 Papadopoulos NG, Sanderson G, Hunter J, Johnston SL (1999) Rhinoviruses replicate effectively at lower airway temperatures. *J Med Virol* 58: 100–104
- 84 Rosenbaum MJ, De Berry P, Sullivan EJ, Pierce WE, Mueller RE, Peckinpaugh RO (1971) Epidemiology of the common cold in military recruits with emphasis on infections by rhinovirus types 1A, 2, and two unclassified rhinoviruses. *Am J Epidemiol* 93: 183–193
- 85 Mogabgab WJ, Pelon W (1957) Problems in characterizing and identifying an apparently new virus found in association with mild respiratory disease in recruits. *Ann N Y Acad Sci* 67: 403–412
- 86 Hendley JO, Edmondson WP Jr, Gwaltney JM Jr (1972) Relation between naturally acquired immunity and infectivity of two rhinoviruses in volunteers. *J Infect Dis* 125: 243–248
- 87 Relman DA (2003) Shedding light on microbial detection. *N Engl J Med* 349: 2162–2163
- 88 Kuypers J, Wright N, Ferrenberg J, Huang M-L, Cent A, Corey L, Morrow R

- (2006) Comparison of real-time PCR assays with fluorescent-antibody assays for diagnosis of respiratory virus infections in children. *J Clin Microbiol* 44: 2382–2388
- 89 Falsey AR, Walsh EE (2006) Viral pneumonia in older adults. *Clin Infect Dis* 42
- 90 Johnston SL, Bardin PG, Pattemore PK (1993) Viruses as precipitants of asthma symptoms. III. Rhinoviruses: Molecular biology and prospects for future intervention. *Clin Exp Allergy* 23: 237–246
- 91 Ketler A, Hamparian VV, Hilleman MR (1962) Characterization and classification of ECHO 28-rhinovirus-coryzavirus agents. *Proc Soc Exp Biol Med* 110: 821–831
- 92 Conant RM, Hamparian VV (1968) Rhinoviruses: Basis for a numbering system. II. Serologic characterization of prototype strains. *J Immunol* 100: 107–113
- 93 Gwaltney JM Jr, Hendley JO (1978) Rhinovirus transmission: One if by air, two if by hand. *Am J Epidemiol* 107: 357–361
- 94 Ledford RM, Patel NR, Demenczuk TM, Watanyar A, Herbertz T, Collett MS, Pevear DC (2004) VP1 sequencing of all human rhinovirus serotypes: Insights into genus phylogeny and susceptibility to antiviral capsid-binding compounds. *J Virol* 78: 3663–3674
- 95 Oberste MS, Maher K, Kilpatrick DR, Pallansch LA (1999) Molecular evolution of the human enteroviruses: Correlation of serotype with VP1 sequence and application to picornavirus classification. *J Virol* 73: 1941–1948
- 96 van de Pol AC, van Loon AM, Wolfs TF, Jansen NJ, Nijhuis M, Breteler EK, Schuurman R, Rossen JW (2007) Increased detection of respiratory syncytial virus, influenza viruses, parainfluenza viruses, and adenoviruses with real-time PCR in samples from patients with respiratory symptoms. *J Clin Microbiol* 45: 2260–2262
- 97 Pitkäranta A, Arruda E, Malmberg H, Hayden FG (1997) Detection of rhinovirus in sinus brushings of patients with acute community-acquired sinusitis by reverse transcription-PCR. *J Clin Microbiol* 35: 1791–1793
- 98 Kämmerer U, Kunkel B, Korn K (1994) Nested PCR for specific detection and rapid identification of human picornaviruses. *J Clin Microbiol* 32: 285–291
- 99 Andeweg AC, Bestebroer TM, Huybreghs M, Kimman TG, de Jong JC (1999) Improved detection of rhinoviruses in clinical samples by using a newly developed nested reverse transcription-PCR assay. *J Clin Microbiol* 37: 524–530
- 100 Aberle JH, Aberle SW, Pracher E, Hutter H-P, Kundi M, Popw-Kraupp T (2005) Impact on clinical course of disease and interferon- γ response. *Pediatr Infect Dis J* 24: 605–610
- 101 Versteegh FGA, Weverling GJ, Peeters MF, Wilbrink B, Veenstra-van Schie MTM, van Leewen-Gerritsen JM, Mooi-Kokenberg EANM, Schellekens JFP, Roord JJ (2005) Community-acquired pathogens associated with prolonged coughing in children: A prospective cohort study. *Clin Microbiol Infect* 11: 801–807
- 102 Hutchinson AF, Ghimire AK, Thompson MA, Black JF, Brand CA, Lowe AJ, Smallwood DM, Vlahos R, Bozinovski S, Brown GV et al. (2007) A communi-

- ty-based, time-matched, case-control study of respiratory viruses and exacerbations of COPD. *Respir Med* 101: 2472–2481
- 103 Arruda E, Hayden FG (1993) Detection of human rhinovirus RNA in nasal washings by PCR. *Mol Cell Probe* 7: 373–379
- 104 Lu X, Holloway B, Dare RK, Kuypers J, Yagi S, Williams JV, Hall CB, Erdman DD (2008) Real-time reverse transcription-PCR assay for comprehensive detection of human rhinoviruses. *J Clin Microbiol* 46: 533–539
- 105 Suvilehto J, Roivainen M, Seppänen M, Meri S, Hovi T, Carpén O, Pitkäranta A (2006) Rhinovirus/enterovirus RNA in tonsillar tissue of children with tonsillar disease. *J Clin Virol* 35: 292–297
- 106 Mackay IM, Bustin S, Andrade JM, Nissen MD, Sloots TP (2007) Quantification of microorganisms: Not human, not dimple, not quick. In: IM Mackay (ed): *Real-time PCR in microbiology*. Caister Academic Press, Norfolk, 133–182
- 107 Stensballe LG, Trautner S, Kofoed PE, Nante E, Hedegaard K, Jensen IP, Aaby P (2002) Comparison of nasopharyngeal aspirate and nasal swab specimens for detection of respiratory syncytial virus in different settings in a developing country. *Trop Med Int Health* 7: 317–321
- 108 Macfarlane P, Denham J, Assous J, Hughes C (2005) RSV testing in bronchiolitis: Which nasal sampling method is best? *Arch Dis Child* 90: 634–635
- 109 Lambert SB, Whiley DM, O'Neill NT, Andrews EC, Canavan FM, Bletchly C, Siebert DJ, Sloots TP, Nissen MD (2008) Comparing nose-throat swabs and nasopharyngeal aspirates collected from children with symptoms for respiratory virus identification using real-time polymerase chain reaction. *Pediatrics* 122: e615–e620
- 110 Lambert SB, Allen KM, Nolan TM (2008) Parent-collected respiratory specimens – A novel method for respiratory virus and vaccine efficacy research. *Vaccine* 26: 1826–1831
- 111 Brunstein JD, Cline CL, McKinney S, Thomas E (2008) Evidence from multiplex molecular assays for complex multipathogen interactions in acute respiratory infections. *J Clin Microbiol* 46: 97–102
- 112 Stott EJ, Eadie MB, Grist NR (1969) Rhinovirus infections of children in hospital: Isolation of three possibly new rhinovirus serotypes. *Am J Epidemiol* 90: 45–52
- 113 Tiveljung-Lindell A, Rotzen-Ostlund M, Gupta S, Ullstrand R, Grillner L, Zwegyberg-Wirgart B, Allander T (2009) Development and implementation of a molecular diagnostic platform for daily rapid detection of 15 respiratory viruses. *J Med Virol* 81: 167–175
- 114 Richard N, Komurian-Pradel F, Javouhey E, Perret M, Rajoharison A, Bagnaud A, Billaud G, Vernet G, Lina B, Floret D et al. (2008) The impact of dual viral infection in infants admitted to a pediatric intensive care unit associated with severe bronchiolitis. *Pediatr Infect Dis J* 27: 1–5
- 115 Gunson RN, Collins TC, Carman WF (2005) Real-time RT-PCR detection of 12 respiratory viral infections in four triplex reactions. *J Clin Virol* 33: 341–344
- 116 Nolte FS, Marshall DJ, Rasberry C, Schievelbein S, Banks GG, Storch GA, Arens MQ, Buller RS, Prudent JR (2007) MultiCode-PLx system for multiplexed detection of seventeen respiratory viruses. *J Clin Microbiol* 45: 2779–2786

- 117 Mahony J, Chong S, Merante F, Yaghoubian S, Sinha T, Lisle C, Janeczko R (2007) Development of a respiratory virus panel test for detection of twenty human respiratory viruses by use of multiplex PCR and a fluid microbead-based assay. *J Clin Microbiol* 45: 2965–2970
- 118 Drews SJ, Blair J, Lombos E, DeLima C, Burton L, Mazzulli T, Low DE (2008) Use of the Seeplex RV Detection kit for surveillance of respiratory viral outbreaks in Toronto, Ontario, Canada. *Ann Clin Lab Sci* 38: 376–379
- 119 Yoo SJ, Kuak EY, Shin BM (2007) Detection of 12 respiratory viruses with two-set multiplex reverse transcriptase-PCR assay using a dual priming oligonucleotide system. *Korean J Lab Med* 27: 420–427
- 120 Roh KH, Kim J, Nam MH, Yoon S, Lee CK, Lee K, Yoo Y, Kim MJ, Cho Y (2008) Comparison of the Seeplex reverse transcription PCR assay with the R-mix viral culture and immunofluorescence techniques for detection of eight respiratory viruses. *Ann Clin Lab Sci* 38: 41–46
- 121 Han J, Swan DC, Smith SJ, Lum SH, Sefers SE, Unger ER, Tang YW (2006) Simultaneous amplification and identification of 25 human papillomavirus types with Tempex technology. *J Clin Microbiol* 44: 4157–4162
- 122 Brunstein J, Thomas E (2006) Direct screening of clinical specimens for multiple respiratory pathogens using the Genaco Respiratory Panels 1 and 2. *Diagn Mol Pathol* 15: 169–173
- 123 Li H, McCormac MA, Estes RW, Sefers SE, Dare RK, Chappell JD, Erdman DD, Wright PF, Tang Y-W (2007) Simultaneous detection and high-throughput identification of a panel of RNA viruses causing respiratory tract infections. *J Clin Microbiol* 45: 2105–2109
- 124 Briese T, Palacios G, Kokoris M, Jabado O, Liu Z, Renwick N, Kapoor V, Casas I, Pozo F, Limberger R et al. (2005) Diagnostic system for rapid and sensitive differential detection of pathogens. *Emerg Infect Dis* 11: 310–313
- 125 Dominguez SR, Briese T, Palacios G, Hui J, Villari J, Kapoor V, Tokarz R, Glode MP, Anderson MS, Robinson CC et al. (2008) Multiplex MassTag-PCR for respiratory pathogens in pediatric nasopharyngeal washes negative by conventional diagnostic testing shows a high prevalence of viruses belonging to a newly recognized rhinovirus clade. *J Clin Virol* 43: 219–222
- 126 Wang D, Coscoy L, Zylberberg M, Avila PC, Boushey HA, Ganem D, DeRisi JL (2002) Microarray-based detection and genotyping of viral pathogens. *Proc Natl Acad Sci USA* 99: 15687–15692
- 127 Ostroff R, Ettinger A, La H, Rihanek M, Zalman L, Meador III J, Patick AK, Worland S, Polisky B (2001) Rapid multiserotype detection of human rhinoviruses on optically coated silicon surfaces. *J Clin Virol* 21: 105–117
- 128 Shanmukh S, Jones L, Driskell J, Zhao Y, Dluhy R, Tripp RA (2006) Rapid and sensitive detection of respiratory virus molecular signatures using a silver nanorod array SERS substrate. *Nano Lett* 6: 2630–2636
- 129 Bae HG, Nitsche A, Teichmann A, Biel SS, Niedrig M (2003) Detection of yellow fever virus: A comparison of quantitative real-time PCR and plaque assay. *J Virol Methods* 110: 185–191
- 130 Jartti T, Lee W-M, Pappas T, Evans M, Lemanske RF, Gern JE (2008) Serial viral infections in infants with recurrent respiratory illnesses. *Eur Respir J* 32: 314–320

- 131 Mackay IM, Arden KE, Nissen MD, Sloots TP (2007) Challenges facing real-time PCR characterization of acute respiratory tract infections. In: IM Mackay (ed): *Real-Time PCR in Microbiology: From Diagnosis to Characterization*. Caister Academic Press, Norfolk, 269–318
- 132 Brouard J, Freymuth F, Vabret A, Jokic M, Guillois B, Duhamel JF (2000) Viral co-infections in immunocompetent infants with bronchiolitis: Prospective epidemiologic study (in French). *Arch Pediatr* 7 (Suppl 3): 531s–535s
- 133 Papadopoulos NG, Moustaki M, Tsolia M, Bossios A, Astra E, Prezerakou A, Gourgiotis D, Kafetzis D (2002) Association of rhinovirus infection with increased disease severity in acute bronchiolitis. *Am J Respir Crit Care Med* 165: 1285–1289
- 134 Maggi F, Pifferi M, Vatteroni M, Fornai C, Tempestini E, Anzilotti S, Lanini L, Andreoli E, Ragazzo V, Pistello M et al. (2003) Human metapneumovirus associated with respiratory tract infections in a 3-year study of nasal swabs from infants in Italy. *J Clin Microbiol* 41: 2987–2991
- 135 Peltola J, Waris M, Hyypiä T, Ruuskanen O (2006) Respiratory viruses in children with invasive pneumococcal disease. *Clin Infect Dis* 43: 266–268
- 136 Juvén T, Mertsola J, Waris M, Leinonen M, Meurman O, Roivainen M, Eskola J, Saikku P, Ruuskanen O (2000) Etiology of community-acquired pneumonia in 254 hospitalized children. *Pediatr Infect Dis J* 19: 293–298
- 137 Garcia-Garcia ML, Calvo C, Perez-Brena P, De Cea JM, Acosta B, Casas I (2006) Prevalence and clinical characteristics of human metapneumovirus infections in hospitalized infants in Spain. *Pediatr Pulmonol* 41: 863–871
- 138 van der Zalm MM, van Ewijk BE, Wilbrink B, Uiterwaal CSPM, Wolfs TFW, van der Ent CK (2009) Respiratory pathogens in children with and without respiratory symptoms. *J Pediatr* 154: 396–400
- 139 Aberle JH, Aberle SW, Pracher E, Hutter H-P, Kundi M, Popw-Kraupp T (2005) Single versus dual respiratory virus infections in hospitalized infants: Impact on clinical course of disease and interferon-gamma response. *Pediatr Infect Dis J* 24: 605–610
- 140 Greensill J, McNamara PS, Dove W, Flanagan B, Smyth RL, Hart CA (2003) Human metapneumovirus in severe respiratory syncytial virus bronchiolitis. *Emerg Infect Dis* 9: 372–375
- 141 Simon A, Wilkesmann A, Muller A, Schildgen O (2007) HMPV infections are frequently accompanied by co-infections. *Pediatr Pulmonol* 42: 98
- 142 Mackay IM (2007) Human bocavirus: Multisystem detection raises questions about infection. *J Infect Dis* 196: 968–970
- 143 Winther B, Alper CM, Mandel EM, Doyle WJ, Hendley JO (2007) Temporal relationships between colds, upper respiratory viruses detected by polymerase chain reaction, and otitis media in young children followed through a typical cold season. *Pediatrics* 119: 1069–1075
- 144 Alper CM, Doyle WJ, Winther B, Hendley JO (2008) Upper respiratory virus detection without parent-reported illness in children is virus-specific. *J Clin Virol* 43: 120–122
- 145 Glezen WP, Denny FW (1973) Epidemiology of acute lower respiratory disease in children. *N Engl J Med* 288: 498–505

- 146 Hitchcock G, Tyrrell DA (1960) Some virus isolations from common colds. II. Virus interference in tissue cultures. *Lancet* 1: 237–239
- 147 Mizgerd JP (2006) Lung infection—a public health priority. *PloS Medicine* 3: e76
- 148 Bardin PG, Johnston SL, Pattemore PK (1992) Viruses as precipitants of asthma symptoms. II. Physiology and mechanisms. *Clin Exp Allergy* 22: 809–822
- 149 Kusel MMH, de Klerk NH, Holt PG, Keadze T, Johnston SL, Sly PD (2006) Role of respiratory viruses in acute upper and lower respiratory tract illness in the first year of life. *Pediatr Infect Dis J* 25: 680–686
- 150 Hakonarson H, Maskeri N, Carter C, Hodinka RL, Campbell D, Grunstein MM (1998) Mechanism of rhinovirus-induced changes in airway smooth muscle responsiveness. *J Clin Invest* 102: 1732–1741
- 151 Gern JE, Galagan DM, Jarjour NN, Dick EC, Busse WW (2008) Detection of rhinovirus RNA in lower airway cells during experimentally induced infection. *Am J Respir Crit Care Med* 155: 1159–1161
- 152 Jakiela B, Brockman-Schneider R, Amineva S, Lee W-M, Gern JE (2008) Basal cells of differentiated bronchial epithelium are more susceptible to rhinovirus infection. *Am J Respir Cell Mol Biol* 38: 517–523
- 153 Gern JE, Busse WW (2002) Relationship of viral infections to wheezing illnesses and asthma. *Nat Rev Immunol* 2: 132–138
- 154 Martin JG, Siddiqui S, Hassan M (2006) Immune responses to viral infections: Relevance for asthma. *Paediatr Respir Rev* 7S: S125–S127
- 155 Krilov L, Pierik L, Keller E, Mahan K, Watson D, Hirsch M, Hamparian V, McIntosh K (1986) The association of rhinoviruses with lower respiratory tract disease in hospitalized patients. *J Med Virol* 19: 345–352
- 156 Monto AS, Bryan ER, Ohmit S (1987) Rhinovirus infections in Tecumseh, Michigan: Frequency of illness and number of serotypes. *J Infect Dis* 156: 43–49
- 157 El-Sahly HM, Atmar RL, Glezen WP, Greenberg SB (2000) Spectrum of clinical illness in hospitalized patients with “Common cold” virus infections. *Clin Infect Dis* 31: 96–100
- 158 Glezen WP, Loda FA, Clyde WA, Senior RJ, Sheaffer CI, Conley WG, Denny FW (1971) Epidemiologic patterns of acute lower respiratory disease of children in a pediatric group practice. *J Pediatr* 78: 397–406
- 159 Bloom HH, Forsyth BR, Johnson KM, Chanock RM (1963) Relationship of rhinovirus infection to mild upper respiratory disease. *JAMA* 186: 38–45
- 160 Collinson J, Nicholson KG, Cancio E, Ashman J, Ireland DC, Hammersley V, Kent J, O’Callaghan C (1996) Effects of upper respiratory tract infections in patients with cystic fibrosis. *Thorax* 51: 1115–1122
- 161 Andréoletti L, Lesay M, Deschildre A, Lambert V, Dewilde A, Wattré P (2000) Differential detection of rhinoviruses and enteroviruses RNA sequences associated with classical immunofluorescence assay detection of respiratory virus antigens in nasopharyngeal swabs from infants with bronchiolitis. *J Med Virol* 61: 341–346
- 162 Rakes GP, Arruda E, Ingram JM, Hoover GE, Zambrano JC, Hayden FG, Platts-Mills TAE, Heymann PW (1999) Rhinovirus and respiratory syncytial

- virus in wheezing children requiring emergency care. *Am J Respir Crit Care Med* 159: 785–790
- 163 Henderson FW, Clyde WA, Collier AM, Denny FW, Senior RJ, Sheaffer CI, Conley WG, Christian RM (1979) The etiologic and epidemiologic spectrum of bronchiolitis in pediatric practice. *J Pediatr* 95: 183–190
- 164 Martinez FD (2007) Gene-environment interactions in asthma. *Proc Am Thorac Soc* 4: 26–31
- 165 Mallia P, Johnston SL (2006) How viral infections cause exacerbation of airway diseases. *Chest* 130: 1203–1210
- 166 Pattemore PK, Johnston SL, Bardin PG (1992) Viruses as precipitants of asthma symptoms. I. Epidemiology. *Clin Exp Allergy* 22: 325–336
- 167 Heymann PW, Platts-Mills TAE, Johnston SL (2005) Role of viral infections, atopy and antiviral immunity in the etiology of wheezing exacerbations among children and young adults. *Pediatr Infect Dis J* 24: S217–S222
- 168 Lidwell OM, Sommerville T (1951) Observations on the incidence and distribution of the common cold in a rural community during 1948 and 1949. *J Hyg (Lond)* 49: 365–381
- 169 Green RM, Custovic A, Sanderson G, Hunter J, Johnston SL, Woodcock A (2007) Synergism between allergens and viruses and risk of hospital admission with asthma: Case-control study. *Br Med J* 324: 1–5
- 170 Johnston SL, Pattemore PK, Sanderson G, Smith S, Lampe F, Josephs L, Symington P, O’Toole S, Myint SH, Tyrrell DAJ et al. (1995) Community study of role of viral infections in exacerbations of asthma in 9–11 year old children. *Br Med J* 310: 1225–1229
- 171 Minor TE, Dick EC, DeMeo AN, Ouellette JJ, Cohen M, Reed CE (1974) Viruses as precipitants of asthmatic attacks in children. *JAMA* 227: 292–298
- 172 Roldaan AC, Masural N (1982) Viral respiratory infections in asthmatic children staying in a mountain resort. *Eur J Respir Dis* 63: 140–150
- 173 Lemanske RF (2002) The childhood origins of asthma (COAST) study. *Pediatr Allergy Immunol* 15: 1–6
- 174 van der Zalm MM, Uiterwaal CSPM, de Jong BM, Wilbrink B, van der Ent CK (2006) Viral specimen collection by parents increases response rate in population-based virus studies. *J Allergy Clin Immunol* 117: 955–957
- 175 Lemanske RF, Jackson DJ, Gangnon RE, Evans MD, Li Z, Shult PA, Kirk CJ, Reisdorf E, Roberg KA, Anderson EL et al. (2005) Rhinovirus illnesses during infancy predict subsequent childhood wheezing. *J Allergy Clin Immunol* 116: 571–577
- 176 Minor TE, Baker JW, Dick EC, DeMeo AN, Ouellette JJ, Cohen M, Reed CE (1974) Greater frequency of viral; respiratory infections in asthmatic children as compared with their nonasthmatic siblings. *J Pediatr* 85: 472–477
- 177 Nicholson KG, Kent J, Ireland DC (1993) Respiratory viruses and exacerbations of asthma in adults. *Br Med J* 307: 982–986
- 178 Rawlinson WD, Waliuzzaman Z, Carter IW, Belessis YC, Gilbert KM, Morton JR (2003) Asthma exacerbations in children are associated with rhinovirus but not human metapneumovirus infection. *J Infect Dis* 187: 1314–1318
- 179 Yoo J, Tcheurekdjian H, Lynch SV, Cabana M, Boushey HA (2007) Microbial

- manipulation of immune function for asthma prevention. Inferences from clinical trials. *Proc Am Thorac Soc* 4: 277–282
- 180 Fedson DS, Nichol KL (2006) Influenza vaccination: Policy *versus* evidence: No gap between policy and evidence. *BMJ* 333: 1020
- 181 Drummond M, Sculpher M, Torrance G (2005) *Methods for the economic evaluation of health care programmes*. Oxford University Press: Oxford
- 182 Michaud CM, Murray CJ, Bloom BR (2001) Burden of disease – implications for future research. *JAMA* 285: 535–539
- 183 Monto AS (2002) Epidemiology of viral respiratory infections. *Am J Med* 112: 4S–12S
- 184 Revai K, Dobbs LA, Nair S (2007) Incidence of acute otitis media and sinusitis complicating upper respiratory tract infection: The effect of age. *Pediatrics* 119: e1408–1412
- 185 Bridges-Webb C, Britt H, Miles DA, Neary S, Charles J, Traynor V (1993) Morbidity and treatment in general practice in Australia. *Aust Fam Physician* 22: 336–339–342–346
- 186 Fendrick AM, Monto AS, Nightengale B (2003) The economic burden of non-influenza-related viral respiratory tract infection in the United States. *Arch Intern Med* 163: 487–494
- 187 Bertino JS (2002) Cost burden of viral respiratory infections: Issues for formulary decision makers. *Am J Med* 112: 42S–49S
- 188 Molinari NA, Ortega-Sanchez IR, Messonnier ML (2007) The annual impact of seasonal influenza in the US: Measuring disease burden and costs. *Vaccine* 25: 5086–5096
- 189 O’Shea TM, Sevick MA, Givner LB (1998) Costs and benefits of respiratory syncytial virus immunoglobulin to prevent hospitalization for lower respiratory tract illness in very low birth weight infants. *Pediatr Infect Dis J* 17: 587–593
- 190 Numa A (2000) Outcome of respiratory syncytial virus infection and a cost-benefit analysis of prophylaxis. *J Paediatr Child Health* 36: 422–427
- 191 Rietveld E, DeJonge HC, Polder JJ (2004) Anticipated costs of hospitalization for respiratory syncytial virus infection in young children at risk. *Pediatr Infect Dis J* 23: 523–529
- 192 Paramore LC, Ciuryla V, Ciesla G (2004) Economic impact of respiratory syncytial virus-related illness in the US: An analysis of national databases. *Pharmacoeconomics* 22: 275–284
- 193 Lambert S, O’Grady K-A, Gabriel S, Carter R, Nolan T (2004) The cost of seasonal respiratory illnesses in Australian children: The dominance of patient and family costs and implications for vaccine use. *Commun Dis Intell* 28: 510–516
- 194 Lambert SB, Allen KM, Druce JD (2005) Respiratory illness during winter: A cohort study of urban children from temperate Australia. *J Paediatr Child Health* 41: 125–129
- 195 Lambert SB, Allen KM, Carter RC, Nolan TM (2008) The cost of community-managed viral respiratory illnesses in a cohort of healthy preschool-aged children. *Respir Res* 9: 1–11
- 196 Australian Bureau of Statistics (2007) Australian Bureau of Statistics: Australian Economic Indicators: May 2004

- 197 Turner J, Tran T, Birch C (2004) Higher than normal seasonal influenza activity in Victoria, 2003. *Commun Dis Intell* 28: 175–180
- 198 Hollinghurst S, Gorst C, Fahey T (2008) Measuring the financial burden of acute cough in pre-school children: A cost of illness study. *BMC Fam Pract* 9: 10
- 199 Coleman MS, Washington ML, Orenstein WA (2006) Interdisciplinary epidemiologic and economic research needed to support a universal childhood influenza vaccination policy. *Epidemiol Rev* 28: 41–46