

LETTER TO THE EDITOR

Detection of *Rickettsia amblyommii* in ticks collected from Missouri, USA

Meghan Hermance¹, Rodrigo I Marques dos Santos¹, Dar Heinze¹, Nicole Hausser^{1,2}, Donald H Bouyer¹ and Saravanan Thangamani^{1,2}

Emerging Microbes and Infections (2014) 3, e34; doi:10.1038/emi.2014.31; published online 21 May 2014

Dear Editor,

In September 2012, we collected ticks from Missouri with the goal of isolating the novel phlebovirus, Heartland virus (HRTV). HRTV was described in two farmers from northwestern Missouri who presented with thrombocytopenia and severe febrile illness.¹ These patients were both bitten by ticks 5–7 days before the onset of their clinical symptoms. Our hypothesis was that we would isolate HRTV from ticks collected in Missouri by inoculation of cell culture and/or by detection of viral RNA on polymerase chain reaction (PCR) assay. Furthermore, we used this field surveillance study as an opportunity to screen for other potential viral and bacterial pathogens in the tick samples we collected.

Information from the published literature¹ was used to identify three geographically relevant collection sites across the central and western region of the state (Supplementary Figure S1A). One thousand two hundred and sixty-nine total ticks were collected from the three locations. Of these, 1191 (93.9%) were *Dermacentor albipictus*, 74 (5.8%) were *Amblyomma americanum* and four (0.3%) were *Ixodes scapularis* (Supplementary Figure S1B). Two nymphs were collected at location 2, but all other ticks collected during this study were larvae.

The speciated ticks were pooled into groups of twenty and screened for tick-borne pathogens (Supplementary Methods and Supplementary Table S1). Using the primers specific for HRTV,¹ Powassan virus² and deer tick virus,³ we were unable to generate any positive PCR amplicons in the viral PCR screening. At location 2, one larval pool and one nymphal pool of ticks generated positive PCR amplicons when screened with the *Rickettsia*-specific primers for the outer membrane protein A (*ompA*) and citrate synthase (*gltA*) genes.⁴

Sequence analysis demonstrated that the two *Rickettsia*-positive samples aligned with the *ompA* and *gltA* genes of *Candidatus Rickettsia amblyommii* [GenBank: 378930552]. *ompA* gene sequences for *R. amblyommii*, *R. raoultii*, *R. slovaca* and *R. rickettsii* were obtained from GenBank. These sequences were trimmed and then underwent ClustalW alignment in the MegAlign program. Specifically, *ompA*-positive samples from both the larval and nymphal pools shared 100% sequence identity across a 431 bp segment of the *ompA* gene of *Candidatus Rickettsia amblyommii* [GenBank: 378930552] (Figure 1A). Additional sequence analysis demonstrated that there was 100% sequence identity between both the larval and nymphal *gltA*-positive samples and *Candidatus Rickettsia amblyommii*

[GenBank: 378930552] across a 628 bp segment of the *gltA* gene (Supplementary Figure S2). Furthermore, the presence of *Rickettsia* in a larval pool of ticks collected at location 2 indicates the occurrence of transovarial transmission of *R. amblyommii*.

To further confirm our molecular identification of *R. amblyommii*, we screened the tick samples with *Rickettsia*-specific primers for the outer membrane protein B (*ompB*) gene⁵ and for the 17 kDa gene.⁶ The tick samples aligned perfectly with the *Candidatus Rickettsia amblyommii* [GenBank: 378930552] *ompB* gene sequence (Supplementary Figure S3). Our *R. amblyommii*-positive samples also completely aligned with the *Candidatus Rickettsia amblyommii* [GenBank: 378930552] 17 kDa gene sequence (Supplementary Figure S4).

No cytopathic effect was detected in any of the cell lines inoculated with the tick homogenates. However, we used an immunofluorescence assay (Supplementary Methods) to confirm the detection of *R. amblyommii* antigens in the *A. americanum* tick homogenates (Figure 1B). Tick mitochondrial 16S rRNA sequence analysis confirmed that the *R. amblyommii*-positive samples were isolated from *A. americanum* ticks. The 16S rRNA sequence from our *Rickettsia*-positive tick pools shared 100% sequence identity with *A. americanum* 16S rRNA (Supplementary Figure S5).

This field surveillance study was unable to isolate HRTV in any of the ticks we collected from Missouri. The lack of HRTV found in this study may be the result of our small sample sizes; only 5.8% of the total collected ticks were *A. americanum*. After our field collection of ticks was completed, another group published their findings of detecting HRTV from *A. americanum* collected in Missouri during 2012.⁷ This group conducted tick collections ranging from April to early-August 2012. As our collection did not occur until mid-September, the majority of ticks collected in our study were larvae, as would be expected.⁸

We did confirm the presence of *R. amblyommii* in two pooled *A. americanum* tick homogenates. To date, no definitive role has been defined for *R. amblyommii* in human pathogenesis, but a recent study has shown that *A. americanum* ticks parasitizing humans are frequently infected with *R. amblyommii*.⁹ Two *A. americanum* ticks collected in Kansas were found to be concurrently infected with *R. rickettsii*, which causes Rocky Mountain spotted fever, and with *R. amblyommii*.¹⁰ The co-infection of these *A. americanum* ticks with *R. rickettsii* and *R. amblyommii* raises interesting questions about the epidemiology of spotted fever group rickettsiae and Rocky Mountain

¹Department of Pathology, University of Texas Medical Branch, Galveston, TX 77555, USA and ²Insectary Services Division, Galveston National Laboratory, University of Texas Medical Branch, Galveston, TX 77555, USA

Correspondence: S Thangamani

E-mail: sathanga@utmb.edu

Received 22 November 2013; revised 14 February 2014; accepted 14 March 2014

- 2 Anderson JF, Armstrong PM. Prevalence and genetic characterization of Powassan virus strains infecting *Ixodes scapularis* in Connecticut. *Am J Trop Med Hyg* 2012; **87**: 754–759.
- 3 Brackney DE, Nofchissey RA, Fitzpatrick KA, Brown IK, Ebel GD. Stable prevalence of Powassan virus in *Ixodes scapularis* in a northern Wisconsin focus. *Am J Trop Med Hyg* 2008; **79**: 971–973.
- 4 Regnery RL, Spruill CL, Plikaytis BD. Genotypic identification of rickettsiae and estimation of interspecies sequence divergence for portions of two rickettsial genes. *J Bacteriol* 1991; **173**: 1576–1589.
- 5 Roux V, Raoult D. Phylogenetic analysis of members of the genus *Rickettsia* using the gene encoding the outer-membrane protein rOmpB (ompB). *Int J Syst Evol Micr* 2000; **50**: 1449–1455.
- 6 Heise SR, Elshahed MS, Little SE. Bacterial diversity in *Amblyomma americanum* (Acari: Ixodidae) with a focus on members of the genus *Rickettsia*. *J Med Entomol* 2010; **47**: 258–268.
- 7 Savage HM, Godsey MS, Lambert A *et al*. First detection of heartland virus (Bunyaviridae: Phlebovirus) from field collected arthropods. *Am J Trop Med Hyg* 2013; **89**: 445–452.
- 8 Kollars TM, Oliver JH, Durden LA *et al*. Host association and seasonal activity of *Amblyomma americanum* (Acari: Ixodidae) in Missouri. *J Parasitol* 2000; **86**: 1156–1159.
- 9 Jiang J, Yarina T, Miller MK, Stromdahl EY, Richards AL. Molecular detection of *Rickettsia amblyommii* in *Amblyomma americanum* parasitizing humans. *Vector-Borne Zoonot* 2010; **10**: 329–340.
- 10 Berrada ZL, Goethert HK, Cunningham J, Telford SR 3rd. *Rickettsia rickettsii* (Rickettsiales: Rickettsiaceae) in *Amblyomma americanum* (Acari: Ixodidae) from Kansas. *J Med Entomol* 2011; **48**: 461–467.
- 11 Apperson CS, Engber B, Nicholson WL *et al*. Tick-borne diseases in North Carolina: is “*Rickettsia amblyommii*” a possible cause of rickettsiosis reported as Rocky Mountain spotted fever? *Vector-Borne Zoonot* 2008; **8**: 597–606.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivs 3.0 Unported License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-nd/3.0/>

Supplementary Information for this article can be found on *Emerging Microbes and Infections*' website (<http://www.nature.com/emi/>)