

Role of Embryo Glue as a transfer medium in the outcome of fresh non-donor *in-vitro* fertilization cycles

**Neeta Singh,
Monica Gupta,
Alka Kriplani,
Perumal Vanamail**
Department of Obstetrics
and Gynaecology, AIIMS,
New Delhi, India

Address for correspondence:

Dr. Neeta Singh,
Department of Obstetrics and
Gynecology, Room No-3090A,
Third Floor, Teaching Block,
All India Institute of Medical
Sciences, Ansari Nagar,
New Delhi, India.
E-mail: drneetasingh
@yahoo.com

Received: 16.03.2015
Review completed: 19.05.2015
Accepted: 25.09.2015

ABSTRACT

BACKGROUND: EmbryoGlue is a hyaluronan-enriched embryo transfer (ET) medium which aids in implantation of embryos, hence, improves pregnancy rates in *in-vitro* fertilization-ET cycles (IVF-ET). **AIM:** To evaluate the role of EmbryoGlue in improving implantation and pregnancy rates. **DESIGN AND SETTING:** A prospective case-control study conducted at assisted reproductive center of a tertiary care hospital. **METHOD:** In 42 women undergoing IVF, embryos were transferred into 50 µL of EmbryoGlue for 10 min prior to transfer inside uterine cavity. In the control group ($n = 42$), embryos were transferred to conventional blastocyst culture medium. Statistical analysis was performed using SPSS IBM version 19.0. **RESULTS:** Clinical pregnancy rate in the study group was 7% higher than the control group. The difference, however, was not statistically significant. In addition, no improvement in implantation rates was observed in the study group. However, significant difference ($P = 0.04$) in clinical pregnancy rate was observed with the EmbryoGlue in patients with previous IVF failure. In the study group, 50% patients (6/12) with previous IVF failure had successful implantation, but in the control group none of the patients (0/11) with previous implantation failure could achieve pregnancy. **CONCLUSION:** It is difficult to conclude a favourable role of EmbryoGlue in IVF-ET cycles with a good prognosis. However, in patients with recurrent implantation failure, it may be considered as a useful transfer medium.

KEY WORDS: Embryo transfer, hyaluronan, implantation, *in-vitro* fertilization, pregnancy, recurrent implantation failure

INTRODUCTION

Assisted reproductive technologies have become an integral part of infertility treatment in the present decade. To increase the efficacy of *in-vitro* fertilization/embryo transfer (IVF-ET) cycles, improvements have been made in techniques of ovarian stimulation, fertilization, culture and transfer of embryos. Despite these advances, implantation of embryos still presents a challenge to the clinician. Implantation is a multistage process which includes zona hatching, apposition and adhesion of blastocyst followed by an invasion of trophoblast into the uterine endometrium. There are several reasons why embryos fail to implant despite a favourable transfer. One of the reasons for unsuccessful implantation being failure to develop a sticky matrix between blastocyst and endometrium. Various modifications have been made in ET medium to improve implantation and

pregnancy rates. Protein supplementation has been widely used in ET medium, most common of which is albumin.^[1] Albumin is present abundantly in the female reproductive tract and serves as a source of energy, hormones, vitamins, and metals. In addition, albumin not only provides viscosity to culture medium, but also acts as a lubricant promoting easy handling and preventing adherence of embryos to the culture dish.^[2] Hyaluronan (HA) supplementation of culture medium was

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

How to cite this article: Singh N, Gupta M, Kriplani A, Vanamail P. Role of Embryo Glue as a transfer medium in the outcome of fresh non-donor *in-vitro* fertilization cycles. *J Hum Reprod Sci* 2015;8:214-7.

Access this article online

Quick Response Code:



Website:

www.jhrsonline.org

DOI:

10.4103/0974-1208.170398

introduced to improve implantation and pregnancy rate in IVF-ET cycles.^[3] HA, a glycosaminoglycan, is present in the oviduct and uterine fluid and increases at the time of implantation. EmbryoGlue is a human ET medium, which contains high concentration of HA (0.5 mg/mL) and low concentration of recombinant human albumin (rHA = 2.5 mg/mL). Standard blastocyst culture medium (G-2™, Vitrolife, Sweden) is supplemented with 10 mg/mL of rHA and a lower concentration of HA (0.125 mg/mL) compared to EmbryoGlue. G-2™ medium also contains ethylenediaminetetraacetic acid (EDTA), which is absent in EmbryoGlue. EmbryoGlue is supplemented with both nonessential and essential amino acids, whereas G-2™ contains only nonessential amino acids. EDTA is required for cleavage stage embryos, but inhibits blastocyst development.^[4] Similarly, only nonessential amino acids promote the development of cleavage stage embryos, whereas both essential and nonessential amino acids are required for blastocyst development.^[5,6] Hence, EmbryoGlue is thought to be the ideal medium for blastocyst development. Accordingly, the present case-control clinical trial was undertaken to evaluate the efficacy of EmbryoGlue as a human ET medium and to compare the pregnancy outcomes versus those of G-2™ medium.

MATERIALS AND METHODS

Ethical approval was taken from the Institute's Ethics Committee prior to the initiation of the study.

Type of study

Prospective, case-control clinical trial.

Study population and site

Women in the reproductive age group undergoing IVF at Assisted Reproductive Unit in the Department of Obstetrics and Gynaecology at a tertiary care center were enrolled in this study.

Study duration

The study was conducted over a period of 3 months between March and May 2014.

Sample size

Eighty-four women undergoing IVF were included in this study. In 42 women undergoing IVF or intracytoplasmic sperm injection (ICSI), embryos were transferred to HA-enriched transfer medium. In the control group ($n = 42$), embryos were transferred to conventional blastocyst culture medium. Women aged >35 years, those with poor ovarian reserve, and possible causes for failure of implantation such as diabetes mellitus, hypertension, and autoimmune diseases were excluded from the study.

Ovarian stimulation

For ovarian stimulation, we either used the gonadotropin-releasing hormone-agonist (GnRH) "long protocol" or flexible GnRH-antagonist regimen. In the long protocol, pituitary down-regulation was achieved with injectable GnRH-agonist, leuprolide acetate (Luprofact; Bayer Zydus, Mumbai, India) 1 mg/day subcutaneous (s.c) from the midluteal phase prior to the treatment cycle. Gonadotropins, most commonly Recombinant Follicle Stimulating Hormone (Gonal F; Merck Serono, Italy) injected s.c. or rarely intramuscular (i.m.) human menopausal gonadotropin (Humog; Bharat Serums and Vaccines Limited, Ambernath) was used for ovarian stimulation after adequate down-regulation was achieved, that is, serum estradiol <30 pg/ml and luteinizing hormone <2 IU/L. During stimulation, the dose of leuprolide acetate was reduced to 0.5 mg/day. In the flexible GnRH-antagonist protocol, Cetorelix acetate (Cetrotide; Merck Serono, Italy) 0.25 mg/day was administered s.c in the stimulated cycle when the diameter of leading follicle reached 14 mm. Recombinant human chorionic gonadotropin (hCG, Ovitrelle; Merck Serono, Italy) 250 µg was administered s.c for inducing ovulation when 2–3 follicles were more than 18 mm in size. Oocyte retrieval was performed 34–36 h after hCG administration under ultrasound guidance. Retrieved oocytes were fertilized either by conventional IVF or ICSI technique. Embryos were cultured in sequential media using the "G-series" produced by Vitrolife (Goteborg, Sweden).

Embryo transfer

Embryos were transferred on day 2–5 depending on the number and quality of embryos. The best or good quality embryos were selected for intrauterine transfer (maximum 3–4 embryos in each ET cycle). In the control group, the selected embryos were transferred into 50 µl of G-2™ culture medium, whereas in the test group, they were transferred into 50 µl of EmbryoGlue (Vitrolife, Sweden). The embryos were incubated for at least 10 min in the transfer medium in an environment of 6% CO₂ at 37°C prior to transfer into uterine cavity. ET was performed using a Labotect catheter (Labotect, Germany). Luteal phase support in the form of progesterone 100 mg i.m. or vaginal pessary 300 mg twice daily was administered to both the groups.

Outcome measures

To calculate the fertilization rate, we divided the number of fertilized zygotes by the total number of oocytes in conventional IVF or injected metaphase II oocytes in ICSI. We also checked the serum beta-hCG in all women on day 14 of transfer. Those with positive beta-hCG underwent sonography for confirmation of clinical pregnancy. The number of gestational sacs divided by the number of embryos transferred was used to calculate the implantation

rate. Follow-up of pregnancy was done until 20 weeks period of gestation.

Data analysis

Statistical analysis was performed using SPSS IBM version 19.0 (Armonk, NY: IBM Corp). Descriptive statistics such as mean, median, standard deviation, and range values were calculated. Mean values of normally distributed data were compared using Student's *t*-independent test. For nonnormal distributed data, median values were compared using Mann-Whitney U-test. Frequency data of categorical variables were compared using Chi-square/Fisher's exact test as appropriate. For all statistical tests, $P < 0.05$ was considered for statistical significance. Main outcome measures were clinical pregnancy rate and implantation rate.

RESULTS

A total of 84 patients undergoing IVF-ET cycle participated in this study. The overall clinical pregnancy rate was 27.4%. In 42 patients, HA-enriched EmbryoGlue was used as a transfer medium and embryos of similar number of age-matched controls were transferred to conventional G-2™ medium. Patient characteristics were as depicted in Table 1. There was no significant difference in between the two groups in relation to mean age, indication of infertility, number of days of stimulation, total dose of gonadotropins, serum estradiol levels on day of trigger, and number of oocytes collected. The frequency of IVF and ICSI cycles was similar in both the groups. There was also no difference among the number of oocytes fertilized by IVF/ICSI, the fertilization rate, and the mean number of embryos transferred. The clinical pregnancy rate was slightly higher in the study group compared to the control group (31% vs. 23.8%). The difference, however, was not statistically significant [Table 2]. There was no difference in the rate of multiple pregnancy and implantation rate between the two groups. In the study group, nine patients who conceived had day 5 embryos transferred, whereas four had day 3. In the control group, six pregnancies were with day 5 embryos and four with day 3. The difference in pregnancy rate between day 5 and day 3 embryos was not significant. The pregnant patients were followed until 20 weeks period of gestation. One patient in the study group had a missed abortion at 8 weeks period of gestation. Rest of the pregnancies were uneventful until the follow-up period. In the study group, there were 12 patients with recurrent implantation failure, out of which 6 (50%) patients had successful implantation. Whereas in the control group, there were 11 patients with recurrent implantation failure and none could achieve pregnancy. The difference in implantation rate was significant ($P = 0.04$) between the two groups in patients with prior implantation failure.

Table 1: Patient characteristics in both the groups

	EmbryoGlue group	Control group	P
Number of IVF/ICSI cycles	42	42	
Mean age±SD	31.26±3.0	32.2±4	0.2
Indication (%)			
Tubal factor	21 (50)	6 (14.2)	0.2
Male factor	7 (16.6)	7 (16.6)	1.0
Endometriosis	2 (4)	2 (4)	1.0
PCOS	3 (7.1)	2 (4)	0.5
Multiple factors	3 (7.1)	3 (7.1)	1.0
Idiopathic	6 (14.2)	13 (30.9)	0.06
Days of stimulation (mean±SD)	10.9±1.4	11.5±1.4	0.07
Total dose of gonadotropins (mean±SD)	3454.9±1086.1	3525.4±1040.6	0.7
Serum E2 level on day of trigger (mean±SD)	3336.1±2194.1	3316.1±1864.7	0.9
Number of oocytes collected (mean±SD)	9.8±4.6	8.5±4.1	0.2
Type of fertilization (%)			
IVF	29 (69)	22 (52.3)	0.1
ICSI	13 (30.9)	20 (47.6)	0.1
Number of oocytes fertilized (mean±SD)	6.9±3.6	6.8±6.5	0.9
Fertilization rate (%)	291/400 (72.75)	252/353 (71.38)	0.84
Cleavage rate (%)	260/291 (89.3)	231/252 (91.7)	0.360
Number of embryos transferred (mean±SD)	3.4±0.7	3.7±3.3	0.6

IVF= *In-vitro* fertilization, SD= Standard deviation, ICSI= Intracytoplasmic sperm injection, PCOS= Polycystic ovary syndrome

Table 2: Clinical outcome in two groups

	EmbryoGlue	Control group	P
Number of IVF/ICSI cycles	42	42	
Clinical pregnancy (%)	13 (31)	10 (23.8)	0.463
Multiple pregnancy (%)	4 (9.5)	5 (11.9)	0.50
Implantation rate (%)	18/44 (40.9)	13/34 (38.2)	0.81

IVF= *In-vitro* fertilization, ICSI= Intracytoplasmic sperm injection

DISCUSSION

HA-enriched culture medium is hypothesized to be best suited for blastocyst development. EmbryoGlue contains higher concentration of HA and lower concentration of albumin. HA is a high molecular weight linear polysaccharide, comprised of repetitive units of n-acetyl-D-glucosamine and d-glucuronic acid.^[7] HA interacts in autocrine and paracrine manner and also with certain cell surface receptors such as CD 44 expressed on preimplantation embryo and in the endometrium.^[8] HA is known to be involved in physiological processes such as embryological development, migration, adhesion, proliferation, and differentiation of cells. It also indirectly promotes angiogenesis.^[1] By virtue of these properties, it improves apposition and attachment of embryos which are key steps in the process of implantation. It also increases viscosity, which promotes

easy handling, facilitating ET process, and by virtue of apposition, it prevents expulsion of embryos from uterine cavity posttransfer. Simon *et al.*^[1] found better pregnancy and implantation rate with HA-enriched media compared to albumin supplementation, though the difference was not significant and the sample size was small. Several other studies have also demonstrated higher pregnancy and implantation rate with HA-enriched media.^[9-12]

In our study, there was no significant difference in pregnancy and implantation rate with EmbryoGlue. However, the clinical pregnancy rate was 7% higher in the glue group. Similar finding of higher clinical pregnancy, but without statistical significance has been demonstrated in several studies which could be due to small sample sizes in these studies.^[13-16] Fancsovičs *et al.*^[16] used HA-enriched media in 290 patients and found 3% higher clinical pregnancy rate in the study group, though the difference was not significant. Addition of HA is supposed to improve implantation of embryos; hence, if more number of embryos are transferred, chances of multiple pregnancy increases. Majority of the studies did not report significant difference in multiple pregnancy rate in between the two groups.^[1,9,17] However, Urman *et al.*^[10] reported 15% higher multiple pregnancy rate with HA-enriched media. We did not find any significant difference in multiple pregnancy rate between the two groups in our study.

When patients with previous IVF failure were evaluated in the study group, it was found that 50% had successful implantation with HA-enriched media compared to the control group. Our finding of improved implantation in recurrent IVF failure is in agreement with few other studies which included patients with recurrent IVF failure.^[14,18] Beneficial effect of HA-enriched media in patients of advanced age and recurrent implantation failure has been demonstrated by Urman *et al.*^[10] We excluded patients >35 years; hence, we cannot comment on implantation rate in elderly patients.

We can conclude that the use of EmbryoGlue is advantageous in patients with recurrent implantation failure. However, a large randomized control trial is required to evaluate its role in all patients.

Acknowledgment

We acknowledge the colleagues who supported us while conducting this study. We also acknowledge the patients who participated in the study.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Simon A, Safran A, Revel A, Aizenman E, Reubinoff B, Porat-Katz A, *et al.* Hyaluronic acid can successfully replace albumin as the sole macromolecule in a human embryo transfer medium. *Fertil Steril* 2003;79:1434-8.
2. Leese HJ. The formation and function of oviduct fluid. *J Reprod Fertil* 1988;82:843-56.
3. Stojkovic M, Kölle S, Peinl S, Stojkovic P, Zakhartchenko V, Thompson JG, *et al.* Effects of high concentrations of hyaluronan in culture medium on development and survival rates of fresh and frozen-thawed bovine embryos produced *in vitro*. *Reproduction* 2002;124:141-53.
4. Gardner DK, Lane MW, Lane M. EDTA stimulates cleavage stage bovine embryo development in culture but inhibits blastocyst development and differentiation. *Mol Reprod Dev* 2000;57:256-61.
5. Lane M, Gardner DK. Nonessential amino acids and glutamine decrease the time of the first three cleavage divisions and increase compaction of mouse zygotes *in vitro*. *J Assist Reprod Genet* 1997;14:398-403.
6. Lane M, Hooper K, Gardner DK. Effect of essential amino acids on mouse embryo viability and ammonium production. *J Assist Reprod Genet* 2001;18:519-25.
7. Scott JE, Heatley F. Biological properties of hyaluronan in aqueous solution are controlled and sequestered by reversible tertiary structures, defined by NMR spectroscopy. *Biomacromolecules* 2002;3:547-53.
8. Campbell S, Swann HR, Aplin JD, Seif MW, Kimber SJ, Elstein M. CD44 is expressed throughout pre-implantation human embryo development. *Hum Reprod* 1995;10:425-30.
9. Friedler S, Schachter M, Strassburger D, Esther K, Ron El R, Raziel A. A randomized clinical trial comparing recombinant hyaluronan/recombinant albumin versus human tubal fluid for cleavage stage embryo transfer in patients with multiple IVF-embryo transfer failure. *Hum Reprod* 2007;22:2444-8.
10. Urman B, Yakin K, Ata B, Isiklar A, Balaban B. Effect of hyaluronan-enriched transfer medium on implantation and pregnancy rates after day 3 and day 5 embryo transfers: A prospective randomized study. *Fertil Steril* 2008;90:604-12.
11. Hambiliki F, Ljunger E, Karlström PO, Stavreus-Evers A. Hyaluronan-enriched transfer medium in cleavage-stage frozen-thawed embryo transfers increases implantation rate without improvement of delivery rate. *Fertil Steril* 2010;94:1669-73.
12. Nakagawa K, Takahashi C, Nishi Y, Jyuen H, Sugiyama R, Kuribayashi Y, *et al.* Hyaluronan-enriched transfer medium improves outcome in patients with multiple embryo transfer failures. *J Assist Reprod Genet* 2012;29:679-85.
13. Loutradi KE, Prassas I, Bili E, Sanopoulou T, Bontis I, Tarlatzis BC. Evaluation of a transfer medium containing high concentration of hyaluronan in human *in vitro* fertilization. *Fertil Steril* 2007;87:48-52.
14. Korosec S, Virant-Klun I, Tomazevic T, Zech NH, Meden-Vrtovec H. Single fresh and frozen-thawed blastocyst transfer using hyaluronan-rich transfer medium. *Reprod Biomed Online* 2007;15:701-7.
15. Hazlett WD, Meyer LR, Nasta TE, Mangan PA, Karande VC. Impact of EmbryoGlue as the embryo transfer medium. *Fertil Steril* 2008;90:214-6.
16. Fancsovičs P, Lehner A, Murber A, Kaszas Z, Rigo J, Urbancsek J. Effect of hyaluronan-enriched embryo transfer medium on IVF outcome: A prospective randomized clinical trial. *Arch Gynecol Obstet* 2015;291:1173-9.
17. Sifer C, Mour P, Tranchant S, Visentin E, Hafhouf E, Sermondade N, *et al.* Is there an interest in the addition of hyaluronan to human embryo culture in IVF/ICSI attempts? *Gynecol Obstet Fertil* 2009;37:884-9.
18. Valojerdi MR, Karimian L, Yazdi PE, Gilani MA, Madani T, Baghestani AR. Efficacy of a human embryo transfer medium: A prospective, randomized clinical trial study. *J Assist Reprod Genet* 2006;23:207-12.