

Research Article

Two-Photon Microscopy Allows Imaging and Characterization of Cochlear Microvasculature In Vivo

Friedrich Ihler,^{1,2} Mattis Bertlich,^{1,2} Bernhard Weiss,¹ Steffen Dietzel,² and Martin Canis^{1,2}

¹Department of Otorhinolaryngology, University of Göttingen Medical Center, Georg-August-Universität Göttingen, Robert-Koch-Strasse 40, 37099 Göttingen, Germany

²Walter-Brendel-Zentrum für Experimentelle Medizin (WBex), Ludwig-Maximilians-Universität München, Marchioninistrasse 15, 81377 München, Germany

Correspondence should be addressed to Martin Canis; martin.canis@med.uni-goettingen.de

Received 1 December 2014; Revised 26 February 2015; Accepted 2 March 2015

Academic Editor: Peter S. Roland

Copyright © 2015 Friedrich Ihler et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Impairment of cochlear blood flow has been discussed as factor in the pathophysiology of various inner ear disorders. However, the microscopic study of cochlear microcirculation is limited due to small scale and anatomical constraints. Here, two-photon fluorescence microscopy is applied to visualize cochlear microvessels. Guinea pigs were injected with Fluorescein isothiocyanate- or Texas red-dextrane as plasma marker. Intravital microscopy was performed in four animals and explanted cochleae from four animals were studied. The vascular architecture of the cochlea was visualized up to a depth of $90.0 \pm 22.7 \mu\text{m}$. Imaging yielded a mean contrast-to-noise ratio (CNR) of 3.3 ± 1.7 . Mean diameter in vivo was $16.5 \pm 6.0 \mu\text{m}$ for arterioles and $8.0 \pm 2.4 \mu\text{m}$ for capillaries. In explanted cochleae, the diameter of radiating arterioles and capillaries was measured with $12.2 \pm 1.6 \mu\text{m}$ and $6.6 \pm 1.0 \mu\text{m}$, respectively. The difference between capillaries and arterioles was statistically significant in both experimental setups ($P < 0.001$ and $P = 0.022$, two-way ANOVA). Measured vessel diameters in vivo and ex vivo were in agreement with published data. We conclude that two-photon fluorescence microscopy allows the investigation of cochlear microvessels and is potentially a valuable tool for inner ear research.

1. Introduction

The pathogenesis of most inner ear disorders is still largely unknown and subject to controversial discussion [1]. Hearing depends on a homeostasis of fluid and ions in the inner ear which is maintained by cochlear microcirculation [2]. Inner ear blood supply is provided by the labyrinthine artery as a branch of the anterior inferior cerebellar artery. Its terminal vessel, the spiral modiolar artery, radiates arterioles to the lateral cochlear wall forming a capillary system in the stria vascularis which lies within the spiral ligament [3]. Impairment of cochlear blood flow has been considered as a factor in the pathophysiology of various kinds of inner ear disorders [4–6] like noise-induced hearing loss and presbycusis [7]. Owing to the typical clinical features, this is also discussed as the cause for sudden sensorineural hearing loss [7, 8].

However, in humans direct evidence for this hypothesis is lacking due to limits for clinical investigation. Difficulties

result from the complexity and the hidden localization of the cochlea within the temporal bone, thus precluding visualization of cochlear microcirculation. Well-established clinical imaging modalities such as magnetic resonance imaging (MRI) [9], single photon emission computed tomography (SPECT) [10], and ultrasound imaging [6] have been adopted for vascular imaging but lack either sufficient spatial resolution, satisfactory contrast, or both to be effective for cochlear microvascular imaging.

Data about the configuration of cochlear blood supply had been gathered at first by ex vivo examination of cast or dissected preparations of animal and human cochleae [3, 11–13]. Intravital microscopy was developed to study cochlear microvessels in guinea pigs in vivo [14, 15], and various animal models were established to assess cochlear blood flow. Methods applied include laser-Doppler flowmetry [16, 17], microelectrode oxygen tension determination [18], labeled [19] or unlabeled [20] microsphere techniques, laser speckle contrast

imaging in combination with Doppler optical microangiography [21], and intravital fluorescence microscopy [14, 22–24]. Conventional fluorescence microscopy provides the best spatial resolution of these techniques and thereby allows selective quantification of blood flow in stria vascularis capillaries. However, it requires ablation of the overlying bone to access cochlear microvessels. Moreover, a specific investigation is limited to capillaries, which lie in the outermost part of the cochlear turn in the spiral ligament. Cochlear radiating arterioles cannot be investigated directly so far. This is because of their anatomical location deep within the ceiling of the scala vestibuli [3, 25]. Moreover, conventional fluorescence microscopy requires preparatory thinning of the cochlear lateral wall [14], with so far undefined but potentially severe consequences on local physiology [26].

Two-photon laser scanning fluorescence microscopy was first described by Denk and coworkers [27]. It is a light microscopy technique that allows *in vivo* imaging up to a depth of one millimeter from the surface of a specimen in some tissues [28–30], providing subcellular resolution and good light penetration as well as low phototoxicity [31, 32]. An important constraint of conventional light microscopy is scattering and absorption of photons. Multiphoton fluorescence microscopy overcomes this by exciting with near-infrared light in frequencies that have superior scattering characteristics. Beyond this, absorption of biological molecules is minimal at the frequencies applied, allowing deeper tissue penetration [33]. The application of two simultaneous photons with high frequency and low energy additionally allows a more focused approach with imaging confined to small volumes [34]. Thus, two-photon microscopy may circumvent invasive preparation while enabling volumetric visualization of a specimen.

In the present study we applied two-photon microscopy to the cochlea of guinea pigs to test whether it allows visualization of the cochlear microvasculature without prior removal of the overlaying cochlear bone.

2. Material and Methods

2.1. Experimental Setup. Six female adult albino Dunkin-Hartley guinea pigs (weight range, 250–300 g) were obtained from Charles River Laboratories (Sulzfeld, Germany). The experiments were performed according to Bavarian state regulations for animal experimentation and were approved on July 27, 2006, by the District Government of Upper Bavaria (animal license number: 55.2-1-54-2531-57-06).

Guinea pigs were anesthetized using intraperitoneal injections of ketamine 85 mg/kg b.w. (Ketavet Parke-Davis, Berlin, Germany) and xylazine 8.5 mg/kg b.w. (Rompun; Bayer, Leverkusen, Germany). A surgical level of anaesthesia was maintained by supplementary half doses of ketamine and xylazine injected every 45 min. This protocol was shown earlier to reliably maintain systemic blood pressure [22].

During the experiments, replacement fluid was infused (NaCl, 8 μ L/100 g b.w./min) to maintain renal blood flow and to compensate for fluid loss encountered during anaesthesia. The animals were placed on a thermostatically controlled

heating pad which maintained temperature at 38°C. Heart rate and oxygen saturation were continuously monitored by pulseoxymetry. During surgical procedures, middle arterial pressure was 67 ± 12 mmHg and heart rate was 230 ± 26 /min on average ($n = 6$). A polyethylene catheter (PE 50) was placed in the right jugular vein for intravenous injections. Head fixation of the animals was realized by a custom-made head holder as described previously [22].

2.2. Surgical Preparations. Through a postauricular incision, the right auditory bulla was opened by a dorsolateral approach. The auricle, overlying soft tissue, the lateral bony part of the bulla including the tympanic membrane, posterior annulus, and tensor tympani tendon as well as parts of the bony bulla were removed. To provide access of the objective to the cochlea, the perpendicular portion of the ramus of the mandibular was sacrificed as well as the middle ear ossicles in order to allow a direct line of visualization. Mucosa and mucosal vessels overlying the second and the third turns of the cochlea were gently wiped off by a piece of gel foam. Anatomic landmarks such as the facial nerve and the semicircular canals were identified (Figure 1).

Following *in vivo* measurements, animals were euthanized by intraperitoneal injection of 800 mg/kg b.w. of sodium pentobarbital. For the examination of explanted cochleae, after systemic injection of fluorescent dye and euthanasia, the petrous part of the temporal bone was removed as a whole. Then, the cochlea was exposed by careful dissection of the bulla. The specimen was studied unfixed in a solution of 0.9% NaCl.

2.3. Two-Photon Microscopy. Fluorescein isothiocyanate-(FITC-) labeled dextran (order number 46947; molecular weight 500 kDa; 0.05 to 0.1 mL of a 5% solution in 0.9% NaCl; Sigma, Deisenhofen, Germany) or Texas red-labeled dextran (order number D1830; molecular weight 70 kDa; 1.0 mL of a 5% solution in 0.9% NaCl; Life Technologies, Carlsbad, CA, USA) was injected intravenously as a plasma marker to visualize cochlear microcirculation. Multiphoton microscopy was performed on a TriMScope (LaVision BioTec, Bielefeld, Germany) described elsewhere [35, 36].

Two water immersion objectives were used for image acquisition, either 20x (numerical aperture 0.95, working distance 2 mm, field number 22 mm, and field of view in current study 0.5 mm \times 0.5 mm) or 10x magnification (numerical aperture 0.3, working distance 3.5 mm, field number 26.5 mm, and field of view in current study 1 mm \times 1 mm). 0.9% NaCl or ultrasound gel was applied as immersion liquid. Excitation was achieved with 800, 860, or 1180 nm.

2.4. Image Processing and Analysis. Two-dimensional images of fluorescence intensity (XY) in successive depths were processed as three-dimensional stacks (XYZ) in Tagged Image File Format (TIFF) by ImageJ 1.47v [37], the distribution Fiji [38], or Imaris 7.6 (Bitplane, Zürich, Switzerland). Stitching of neighboring stacks to one was performed with a plug-in for Fiji [39].

Vessels were classified by their branching characteristics as radiating arteriole before the first offshoot or as capillary

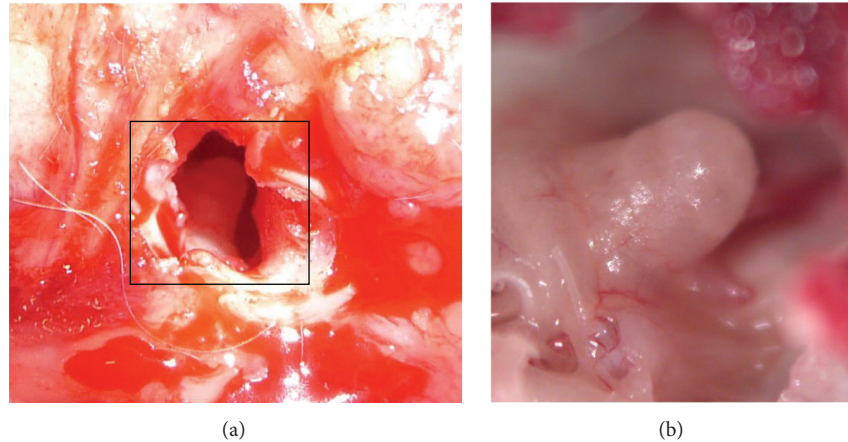


FIGURE 1: Exposure of a guinea pig cochlea. (a) The right bulla was opened through a lateral and ventral approach to allow access to the cochlea (box) for two-photon microscopy. (b) Explanted cochlea in a larger magnification.

thereafter. Radial arterioles have been described to run within the spiral osseous lamina perpendicular to the midmodiolar axis of the cochlea while capillaries are branching from them in the lateral cochlear wall with an angle of about 60 degrees [3]. These characteristics were readily identifiable in our experimental setup.

Depth of visualization was determined by identifying the deepest visible vessel in each stack and averaging the depth value across stacks. Vessel borders could be easily identified and blurred edges constituted no major obstacle due to viable resolution. However, to rule out any remaining variability, values for vessel diameters were obtained by averaging 5 independent measurements along the visible sector on each identified vessel, thereby correcting for variations along the vessel axis. The mean coefficient of variation of the individual measurements of diameter for the respective vessels was 0.05 (standard deviation 0.04, range 0.01–0.19), signifying a low variation.

Contrast-to-noise ratio (CNR) was calculated by measuring mean grey value (M) and standard deviation of grey value (SD) in representative regions of interest within (M_W , SD_W) and outside (M_O , SD_O) the studied vessels by application of the formula

$$CNR = \frac{(M_W - M_O)}{\text{sqrt}(SD_W^2 + SD_O^2)} \quad (1)$$

as described before [40, 41].

2.5. Statistical Analysis. Statistical analysis was performed using SigmaPlot 12 (Systat Software, Chicago, IL, USA). To identify differences greater than expected by chance, result values were compared using two-way analysis of variance (ANOVA). Thereby, vessel diameter and contrast-to-noise ratio (CNR) were analysed for differences dependent on vessel type (capillary or arteriole), specimen (in vivo measurement or explant), and dye (FITC or Texas red-dextran). To correct for multiple comparisons, the Holm-Sidak method was applied. A P value < 0.05 was considered to be statistically significant.

3. Results

After surgical exposure of the cochlea (Figure 1), intravital observation of cochlear microcirculation with two-photon laser scanning microscopy was feasible through the intact cochlear bony wall without ablation of the bone. Visualization was possible up to a depth of $90.0 \pm 22.7 \mu\text{m}$ from the outer surface of the bone. Optical sections of the stria vascularis vasculature were recorded along the Z -direction from the third turn (Figure 2).

Four cochleae from four animals were examined by intravital two-photon microscopy. From the respective images, six arterioles and six capillaries were selected for quantification. As an alternative to intravital microscopy, four explanted cochleae from four animals were subjected to two-photon microscopy. The elimination of animal movement allowed recording of higher resolution images with slower scan rates (Figure 3).

Regarding image quality, two-photon microscopy yielded a mean contrast-to-noise ratio (CNR) of 3.3 ± 1.7 in all images. CNR for images obtained from explanted cochleae was 3.7 ± 1.7 while CNR was 3.1 ± 1.8 in in vivo images. Fluorescein isothiocyanate (FITC) as a fluorescent dye resulted in a CNR of 3.6 ± 1.6 while CNR of images obtained with Texas red was 3.1 ± 1.8 . Image quality with Texas red was higher in images from explanted cochleae compared to in vivo images ($P = 0.015$). Contrastingly, in measurements on living animals, FITC was superior to Texas red ($P = 0.022$). An overview of contrast-to-noise values is given in Table 1.

Mean diameter in vivo was $16.5 \pm 6.0 \mu\text{m}$ for six arterioles and $8.0 \pm 2.4 \mu\text{m}$ for six capillaries. The difference was statistically significant ($P < 0.001$, two-way ANOVA). An example of a time series of a XY -image is presented in Video 1 (see Video 1 in Supplementary Material available online at <http://dx.doi.org/10.1155/2015/154272>). In freshly explanted, unfixed cochleae, the diameter of five radiating arterioles and five capillaries was measured with $12.2 \pm 1.6 \mu\text{m}$ and $6.6 \pm 1.0 \mu\text{m}$, respectively. This difference was statistically significant ($P = 0.022$, two-way ANOVA). Although a change

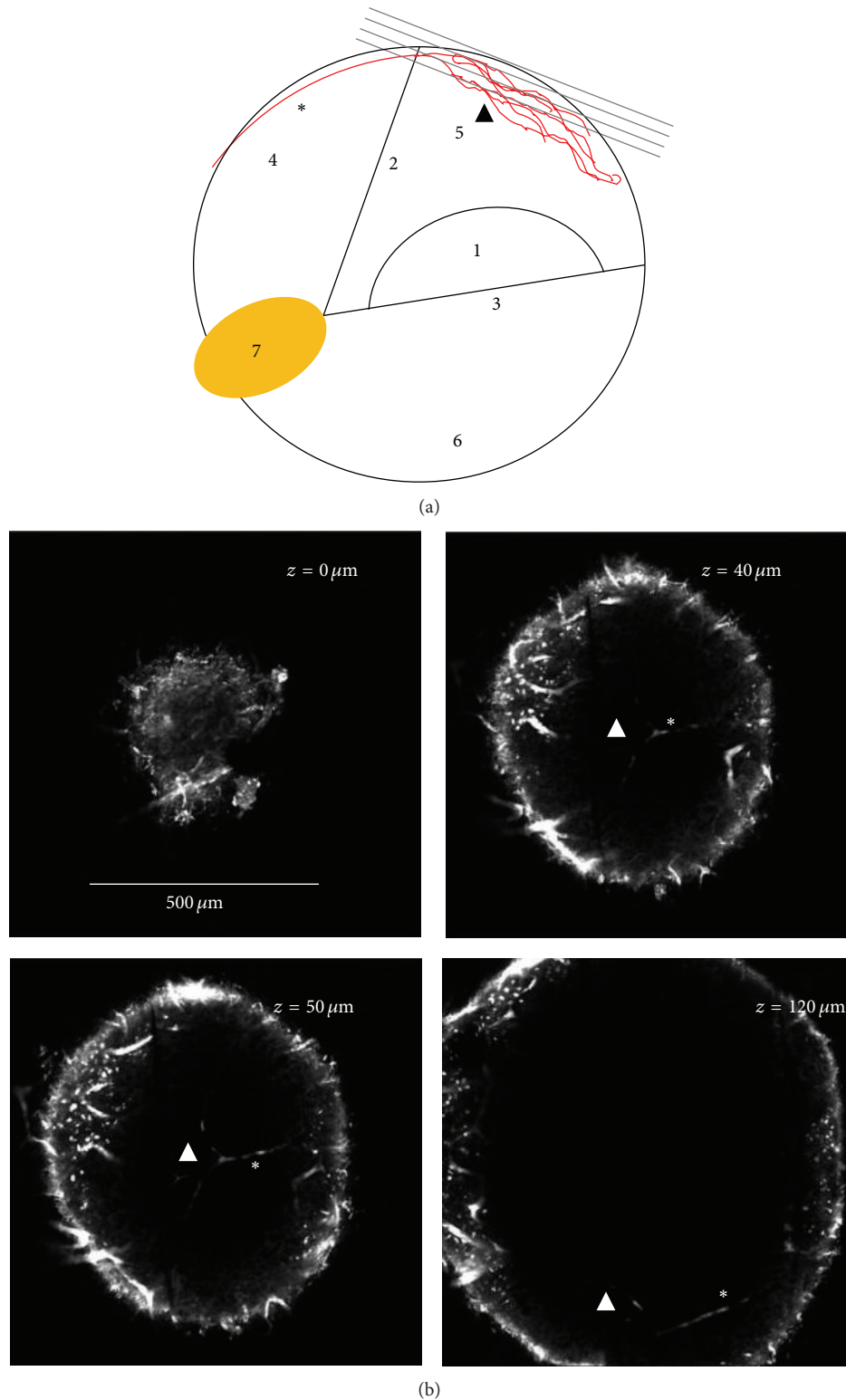


FIGURE 2: Three-dimensional two-photon imaging of an explanted cochlea. (a) The schematic drawing shows the location of successive *XY*-images of the guinea pig cochlear lateral wall (dark grey) in relation to neighboring structures in a cross section of a cochlear turn. The images are recorded along the *Z*-direction using two-photon microscopy after application of Texas red-labeled dextran and excitation with 1180 nm. (b) Four of the equidistantly acquired optical sections were selected for representation in this figure to show specific features. A strong autofluorescence signal visualizes the bony capsule of the cochlea. *, radiating arteriole; ▲, stria vascularis capillaries; 1, organ of Corti; 2, membrane of Reissner; 3, basilar membrane; 4, scala vestibuli; 5, scala media; 6, scala tympani; 7, spiral ganglion.

TABLE 1: Image quality in two-photon microscopy of cochlear microvessels ex vivo and in vivo.

Specimen/dye		FITC	Texas red	<i>P</i> (two-way ANOVA)
Ex vivo examination (explanted cochlea)	Number of vessels	5	5	=0.161
	CNR	3.0 ± 1.0	4.4 ± 2.1	
In vivo examination	Number of vessels	6	6	=0.022*
	CNR	4.2 ± 2.0	2.0 ± 0.4	
P (two-way ANOVA)		=0.227	=0.015*	

ANOVA: analysis of variance; CNR: contrast-to-noise ratio; FITC: fluorescein isothiocyanate; * difference statistically significant with $P < 0.05$.

TABLE 2: Diameter of cochlear microvessels ex vivo and in vivo by two-photon microscopy.

Specimen/vessel		Arterioles	Capillaries	<i>P</i> (two-way ANOVA)
Ex vivo examination (explanted cochlea)	Number of vessels	5	5	=0.022*
	Diameter [μm]	12.2 ± 1.6	6.6 ± 1.0	
In vivo examination	Number of vessels	6	6	<0.001*
	Diameter [μm]	16.5 ± 6.0	8.0 ± 2.4	
P (two-way ANOVA)		=0.060	=0.512	

ANOVA: analysis of variance; * difference statistically significant with $P < 0.05$.

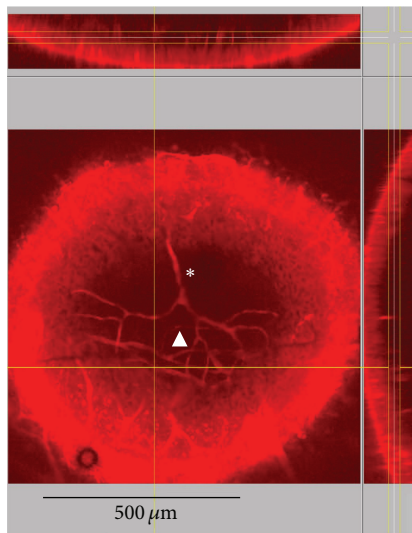


FIGURE 3: Vascular architecture of the guinea pig cochlea. Visualization of microvessels in an explanted cochlea by two-photon microscopy. The main image shows an *XY*-projection of those sections which are between the vertical yellow lines in the *YZ*-section on the right. This *YZ*-section is placed at the vertical yellow line in the main image. The horizontal yellow line indicates the position of the *XZ*-section shown at the top, mostly covered by the inset. *, radiating arteriole; ▲, stria vascularis capillaries.

in diameter might be expected for vessels in explants, for example, due to postmortal shrinking, the difference to in vivo diameters was not significant in the present data. An overview of the vessel diameters obtained can be found in Table 2.

4. Discussion

Conventional intravital fluorescent microscopy after surgical reduction of the bony cochlear lateral wall was to date the only published approach for quantification of vessel count, diameter, length, density, permeability, and blood flow velocity in cochlear microcirculation [14, 22–24]. However, surgical preparation of the cochlear lateral wall as required for conventional microscopy bears the risk of disturbing local physiology [26], thus potentially invalidating the measurements performed. Here we demonstrate that investigations are possible through the intact cochlear lateral wall with two-photon fluorescence microscopy. We were able to visualize the three-dimensional anatomy of the cochlear vasculature, assess vessel diameters in vivo and ex vivo, and show that in principle blood flow measurements are possible with this approach.

In microcirculatory experiments under anesthesia, an independent effect of anesthetics on microvessels is to be discussed. Xylazine and ketamine are known to provide reliably stable cardiorespiratory parameters during surgical experiments [42]. There are currently no studies focusing explicitly on the effect of this combination on cochlear microcirculation. In an experimental setup employing a dorsal skinfold chamber, ketamine was supposed to cause a constrictive state during induction and maintenance of anesthesia compared to the awake state [43]. Contrasting reports have shown a negligible effect of xylazine on vessel diameters [44]. The anesthesia protocol applying ketamine and xylazine in the present report has been shown to provide stable narcosis without confounding experiments in several studies to date [22, 45–50].

The definition of cochlear capillaries is an issue not yet finally resolved. A considerable fraction of the studies on

cochlear microvasculature do not elaborate on the classification system applied, most likely since *in vivo* only capillaries were visible [11–14, 24, 51]. Preparations of the cochlea by cast in earlier studies allowed following the complete vessel tree and thereby identifying different segments. The radiating arterioles have been described as singular vessels branching from the spiral modiolar artery and running perpendicularly from the midmodiolar axis within the spiral lamina towards the cochlear lateral wall [3, 52], where they feed into the stria vascularis in regular intervals [11, 52]. Capillaries have been described to branch off these vessels with an angle of about 60 degrees [3]. Other investigators acknowledged the classification of cochlear microvessels as an issue in *in vivo* experiments but concluded that visual classification by known branching characteristics is a method with acceptable accuracy [15].

A mean contrast-to-noise ratio (CNR) of 3.3 ± 1.7 in all images of the present study represents an acceptable level of quality. Other optical or combined approaches that attempted to achieve subsurface imaging *in vivo* reported a CNR of 1.95 for a penetration depth of up to 7 mm [41] or 3.5 to 10 for up to 8 mm [53]. However, no previous study had to deal with the challenging situation of bone as an optically dense substance overlying a liquid as in the guinea pig cochlea in the present study.

Previously published data stated the diameter of stria vascularis capillaries in guinea pigs from 7.5 to 16.0 μm *in vivo* [15, 24, 51] and 9.2 to 21.8 μm *ex vivo* [11–13], while radiating arterioles were reported to measure 12.0 to 19.3 μm *ex vivo* [3, 12, 52]. Our results of $8.0 \pm 2.4 \mu\text{m}$ for capillaries *in vivo* and $12.2 \pm 1.6 \mu\text{m}$ for arterioles *ex vivo* are well in line with those earlier reports. The diameter of stria vascularis capillaries in unfixed, freshly explanted cochleae of $6.6 \pm 1.0 \mu\text{m}$ *ex vivo* in the present study was considerably smaller than values published on fixed material. An explanation for that could be an enlargement of the vasculature by increased pressure that was discussed to occur during perfusion with fixation agents [12]. No prior *in vivo* data on radiating arterioles are available, most likely due to their location within the roof of the scala vestibuli inaccessible for traditional methods. Therefore, this was assessed with $16.5 \pm 6.0 \mu\text{m}$ *in vivo* for the first time in the present report.

There is an ongoing debate where blood flow regulation in the cochlea takes place. Arteriolar resistance vessels surrounded by smooth muscle cells have been suggested to play an important part [2]. In contrast, capillaries show no smooth muscle cells but contain a high density of pericytes [2, 54] which exhibit vasocontractility under both *in vivo* and *in vitro* conditions [2, 54, 55] and thereby may be an important factor in the regulation of cochlear microcirculation [2, 54]. Conventional intravital microscopy lacks the depth resolution that is crucial for characterizing three-dimensional microvascular morphology of the vascular bed on a cellular level, thereby making two-photon microscopy a valuable novel approach to address those questions. A possible restriction, however, might be imaging speed, as visualization by a laser-scanning application like two-photon microscopy might limit the image section accessible for real-time imaging [33]. A further limitation is the required amount of surgical

removal of tissue that does not allow repeated assessments over several days or longer. However, during the experiments the surgical procedures and consecutive measurements were tolerated quite well and cardiovascular parameters were stable.

The feasibility of observation of cochlear vasculature through the intact cochlear bony wall as demonstrated here bears the potential to develop into a valuable animal model for research on inner ear microcirculation. In addition to superior penetration depth compared to conventional confocal microscopy [29], two-photon excitation minimizes photobleaching, the destruction of fluorophores, and tissue damage by phototoxicity, ameliorating further problems of earlier approaches. Two-photon-excitation line-scanning approaches [56–59] may allow measurement of red blood cell velocity, shear stress, and related hemodynamic parameters of cochlear blood flow.

5. Conclusion

Two-photon microscopy allows the differentiation of radiating arterioles and stria vascularis capillaries of the cochlea *in vivo* and *ex vivo* and has potentially broad applications in imaging and characterization of cochlear microcirculation in health and disease models.

Disclosure

The authors declare no financial interests in companies or other entities that have an interest in the information in the contribution (including grants, advisory boards, employment, consultancies, contracts, honoraria, royalties, expert testimony, partnerships, or stock ownership in medically related fields). Parts of this work were presented at the 82nd Annual Meeting of the German Society of Oto-Rhino-Laryngology, Head and Neck Surgery, June 1–5, 2011, in Freiburg im Breisgau, Germany. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the paper.

Conflict of Interests

The authors declare that there is no conflict of interests.

Authors' Contribution

Conception and design of the study were done by Friedrich Ihler, Steffen Dietzel, and Martin Canis; acquisition of data was done by Friedrich Ihler and Steffen Dietzel; analysis and interpretation of data were done by Friedrich Ihler, Mattis Bertlich, Bernhard Weiss, and Martin Canis. Drafting the manuscript was done by Friedrich Ihler and Martin Canis; revising the paper critically for important intellectual content was done by Friedrich Ihler, Mattis Bertlich, Bernhard Weiss, Steffen Dietzel, and Martin Canis. All authors approved the final version of the paper. Steffen Dietzel and Martin Canis contributed equally to this work.

Acknowledgments

FI and MC were supported by the German Federal Ministry of Education and Research under the Grant code 01 EO 0901. This study was also supported by the BioImaging Network of the Ludwig-Maximilians-Universität München (<http://www.bin.lmu.de>), the Excellence Initiative of the Deutsche Forschungsgemeinschaft (<http://www.dfg.de>), and the Bundesministerium für Bildung und Forschung (<http://www.bmbf.de>). FI, MB, BW, and MC were supported by the Deutsche Forschungsgemeinschaft under the grant code CA 629/2-1. The publication of this study was supported by the Deutsche Forschungsgemeinschaft (<http://www.dfg.de>) and the Open Access Publication Funds of the Georg-August-Universität Göttingen. Parts of this work were presented at the 82nd Annual Meeting of the German Society of Oto-Rhino-Laryngology, Head and Neck Surgery, June 1st–5th 2011 in Freiburg im Breisgau, Germany.

References

- [1] F. M. Byl Jr., "Sudden hearing loss: eight years' experience and suggested prognostic table," *Laryngoscope*, vol. 94, no. 5, part 1, pp. 647–661, 1984.
- [2] X. Shi, "Physiopathology of the cochlear microcirculation," *Hearing Research*, vol. 282, no. 1-2, pp. 10–24, 2011.
- [3] A. Axelsson, "The vascular anatomy of the cochlea in the guinea pig and in man," *Acta Oto-Laryngologica*, vol. 66, supplement 243, pp. 3–134, 1968.
- [4] A. Johnson, M. Hawke, and G. Berger, "Sudden deafness and vertigo due to inner ear hemorrhage—a temporal bone case report," *Journal of Otolaryngology*, vol. 13, no. 4, pp. 201–207, 1984.
- [5] J. M. Miller and H. Dengerink, "Control of inner ear blood flow," *American Journal of Otolaryngology: Head and Neck Medicine and Surgery*, vol. 9, no. 6, pp. 302–316, 1988.
- [6] J. M. Miller, T.-Y. Ren, and A. L. Nuttall, "Studies of inner ear blood flow in animals and human beings," *Otolaryngology—Head and Neck Surgery*, vol. 112, no. 1, pp. 101–113, 1995.
- [7] T. Nakashima, S. Naganawa, M. Sone et al., "Disorders of cochlear blood flow," *Brain Research Reviews*, vol. 43, no. 1, pp. 17–28, 2003.
- [8] A. Glacet-Bernard, W. Roquet, A. Coste, R. Peynègre, G. Coscas, and G. Soubrane, "Central retinal vein occlusion and sudden deafness: a possible common pathogenesis," *European Journal of Ophthalmology*, vol. 11, no. 2, pp. 197–199, 2001.
- [9] J. Thornton, Q. Bashir, V. A. Aletich, G. M. Debrun, F. Charbel, and M. F. Mafee, "Role of magnetic resonance imaging and diagnostic and interventional angiography in vascular and neoplastic diseases of the skull base associated with vestibulo-cochlear symptoms," *Topics in Magnetic Resonance Imaging*, vol. 11, no. 2, pp. 123–137, 2000.
- [10] M. Koyuncu, A. R. Elhami, H. Akan, M. Sahin, T. Basoglu, and M. Simsek, "Investigation of the vertebrobasilar arterial system in vertigo by vestibulocochlear test, SPECT and angiography," *Auris Nasus Larynx*, vol. 28, no. 1, pp. 23–28, 2001.
- [11] L. Carlisle and A. Forge, "The vessels of the stria vascularis: quantitative comparison of three rodent species," *Hearing Research*, vol. 38, no. 1-2, pp. 111–117, 1989.
- [12] M. Moriguchi, H. Masutani, M. Sugita, K. Matsunaga, J. Okamoto, and Y. Nakai, "Measurement of inner ear vessel diameters with casting method," *Acta Oto-Laryngologica, Supplement*, vol. 486, pp. 39–44, 1991.
- [13] L. Fechter and L. Carlisle, "Auditory dysfunction and cochlear vascular injury following trimethyltin exposure in the guinea pig," *Toxicology and Applied Pharmacology*, vol. 105, no. 1, pp. 133–143, 1990.
- [14] A. L. Nuttall, "Techniques for the observation and measurement of red blood cell velocity in vessels of the guinea pig cochlea," *Hearing Research*, vol. 27, no. 2, pp. 111–119, 1987.
- [15] F. P. Miles and A. L. Nuttall, "In vivo capillary diameters in the stria vascularis and spiral ligament of the guinea pig cochlea," *Hearing Research*, vol. 33, no. 2, pp. 191–200, 1988.
- [16] T. Ren, P. B. Brechtelsbauer, J. M. Miller, and A. L. Nuttall, "Cochlear blood flow measured by averaged laser Doppler flowmetry (ALDF)," *Hearing Research*, vol. 77, no. 1-2, pp. 200–206, 1994.
- [17] P. C. Goodwin, J. M. Miller, H. A. Dengerink, J. W. Wright, and A. Axelsson, "The laser Doppler: a non-invasive measure of cochlear blood flow," *Acta Oto-Laryngologica*, vol. 98, no. 5-6, pp. 403–412, 1984.
- [18] A. L. Nuttall, E. Hultcrantz, and M. Lawrence, "Does loud sound influence the intracochlear oxygen tension?" *Hearing Research*, vol. 5, no. 2-3, pp. 285–293, 1981.
- [19] E. Hultcrantz and C. Angelborg, "Cochlear blood circulation studied with microspheres," *ORL*, vol. 40, no. 2, pp. 65–76, 1978.
- [20] J. Prazma, S. G. Vance, and G. Rodgers, "Measurement of cochlear blood flow—new technique," *Hearing Research*, vol. 14, no. 1, pp. 21–28, 1984.
- [21] R. Reif, J. Qin, L. Shi et al., "Monitoring hypoxia induced changes in cochlear blood flow and hemoglobin concentration using a combined dual-wavelength laser speckle contrast imaging and Doppler optical microangiography system," *PLoS ONE*, vol. 7, no. 12, Article ID e52041, 2012.
- [22] M. Canis, W. Arpornchayanon, C. Messmer, M. Suckfuell, B. Olzowy, and S. Strieth, "An animal model for the analysis of cochlear blood flow disturbance and hearing threshold in vivo," *European Archives of Oto-Rhino-Laryngology*, vol. 267, no. 2, pp. 197–203, 2010.
- [23] A. Axelsson, A. L. Nuttall, and J. M. Miller, "Observations of cochlear microcirculation using intravital microscopy," *Acta Oto-Laryngologica*, vol. 109, no. 3-4, pp. 263–270, 1990.
- [24] A. L. Nuttall, "Velocity of red blood cell flow in capillaries of the guinea pig cochlea," *Hearing Research*, vol. 27, no. 2, pp. 121–128, 1987.
- [25] J. E. Hawkins Jr., "Microcirculation in the labyrinth," *Archives of Oto-Rhino-Laryngology*, vol. 212, no. 4, pp. 241–251, 1976.
- [26] S. Hu and L. V. Wang, "Photoacoustic imaging and characterization of the microvasculature," *Journal of Biomedical Optics*, vol. 15, no. 1, Article ID 011101, 2010.
- [27] W. Denk, J. H. Strickler, and W. W. Webb, "Two-photon laser scanning fluorescence microscopy," *Science*, vol. 248, no. 4951, pp. 73–76, 1990.
- [28] K. König, "Multiphoton microscopy in life sciences," *Journal of Microscopy*, vol. 200, no. 2, pp. 83–104, 2000.
- [29] T. Ishii and M. Ishii, "Intravital two-photon imaging: a versatile tool for dissecting the immune system," *Annals of the Rheumatic Diseases*, vol. 70, supplement 1, pp. i113–i115, 2011.
- [30] F. Helmchen and W. Denk, "Deep tissue two-photon microscopy," *Nature Methods*, vol. 2, no. 12, pp. 932–940, 2005.

- [31] K. Svoboda and R. Yasuda, "Principles of two-photon excitation microscopy and its applications to neuroscience," *Neuron*, vol. 50, no. 6, pp. 823–839, 2006.
- [32] B.-G. Wang, K. Konig, and K. J. Halbhuer, "Two-photon microscopy of deep intravital tissues and its merits in clinical research," *Journal of Microscopy*, vol. 238, no. 1, pp. 1–20, 2010.
- [33] P. Pantazis and W. Supatto, "Advances in whole-embryo imaging: a quantitative transition is underway," *Nature Reviews Molecular Cell Biology*, vol. 15, no. 5, pp. 327–339, 2014.
- [34] D. M. Small, W. Y. Sanchez, S. Roy, M. J. Hickey, and G. C. Gobe, "Multiphoton fluorescence microscopy of the live kidney in health and disease," *Journal of Biomedical Optics*, vol. 19, no. 2, Article ID 020901, 2014.
- [35] M. Rehberg, F. Krombach, U. Pohl, and S. Dietzel, "Signal improvement in multiphoton microscopy by reflection with simple mirrors near the sample," *Journal of Biomedical Optics*, vol. 15, no. 2, Article ID 026017, 2010.
- [36] M. Rehberg, F. Krombach, U. Pohl, and S. Dietzel, "Label-free 3D visualization of cellular and tissue structures in intact muscle with second and third harmonic generation microscopy," *PLoS ONE*, vol. 6, no. 11, Article ID e28237, 2011.
- [37] C. A. Schneider, W. S. Rasband, and K. W. Eliceiri, "NIH Image to ImageJ: 25 years of image analysis," *Nature Methods*, vol. 9, no. 7, pp. 671–675, 2012.
- [38] J. Schindelin, I. Arganda-Carreras, E. Frise et al., "Fiji: an open-source platform for biological-image analysis," *Nature Methods*, vol. 9, no. 7, pp. 676–682, 2012.
- [39] S. Preibisch, S. Saalfeld, and P. Tomancak, "Globally optimal stitching of tiled 3D microscopic image acquisitions," *Bioinformatics*, vol. 25, no. 11, pp. 1463–1465, 2009.
- [40] J. J. Vaquero, S. Redondo, E. Lage et al., "Assessment of a new high-performance small-animal X-ray tomograph," *IEEE Transactions on Nuclear Science*, vol. 55, no. 3, pp. 898–905, 2008.
- [41] X. Yang, H. Gong, G. Quan, Y. Deng, and Q. Luo, "Combined system of fluorescence diffuse optical tomography and micro-computed tomography for small animal imaging," *Review of Scientific Instruments*, vol. 81, no. 5, Article ID 054304, 2010.
- [42] D. O. Schwenke and P. A. Cragg, "Comparison of the depressive effects of four anesthetic regimens on ventilatory and cardiovascular variables in the Guinea pig," *Comparative Medicine*, vol. 54, no. 1, pp. 77–85, 2004.
- [43] Z. L. S. Brookes, N. J. Brown, and C. S. Reilly, "Intravenous anaesthesia and the rat microcirculation: the Dorsal Microcirculatory Chamber," *British Journal of Anaesthesia*, vol. 85, no. 6, pp. 901–903, 2000.
- [44] R. C. Sloan, M. Rosenbaum, D. O'Rourke et al., "High doses of ketamine-xylazine anesthesia reduce cardiac ischemia-reperfusion injury in guinea pigs," *Journal of the American Association for Laboratory Animal Science*, vol. 50, no. 3, pp. 349–354, 2011.
- [45] W. Arpornchayanon, M. Canis, F. Ihler, C. Settevendemie, and S. Strieth, "TNF-alpha inhibition using etanercept prevents noise-induced hearing loss by improvement of cochlear blood flow in vivo," *International Journal of Audiology*, vol. 52, no. 8, pp. 545–552, 2013.
- [46] W. Arpornchayanon, M. Canis, M. Suckfuell, F. Ihler, B. Olzowy, and S. Strieth, "Modeling the measurements of cochlear microcirculation and hearing function after loud noise," *Otolaryngology—Head and Neck Surgery*, vol. 145, no. 3, pp. 463–469, 2011.
- [47] M. Bertlich, F. Ihler, K. Sharaf, B. G. Weiss, M. Strupp, and M. Canis, "Betahistine metabolites, aminoethylpyridine, and hydroxyethylpyridine increase cochlear blood flow in guinea pigs in vivo," *International Journal of Audiology*, vol. 53, no. 10, pp. 753–759, 2014.
- [48] F. Ihler, M. Bertlich, K. Sharaf, S. Strieth, M. Strupp, and M. Canis, "Betahistine exerts a dose-dependent effect on cochlear stria vascularis blood flow in guinea pigs in vivo," *PLoS ONE*, vol. 7, no. 6, Article ID e39086, 2012.
- [49] F. Ihler, K. Sharaf, M. Bertlich et al., "Etanercept prevents decrease of cochlear blood flow dose-dependently caused by tumor necrosis factor alpha," *Annals of Otolaryngology & Laryngology*, vol. 122, no. 7, pp. 468–473, 2013.
- [50] F. Ihler, S. Strieth, N. Pieri, P. Göhring, and M. Canis, "Acute hyperfibrinogenemia impairs cochlear blood flow and hearing function in guinea pigs in vivo," *International Journal of Audiology*, vol. 51, no. 3, pp. 210–215, 2012.
- [51] W. S. Quirk, G. Avinash, A. L. Nuttall, and J. M. Miller, "The influence of loud sound on red blood cell velocity and blood vessel diameter in the cochlea," *Hearing Research*, vol. 63, no. 1-2, pp. 102–107, 1992.
- [52] M. Lawrence, "Control mechanisms of inner ear microcirculation," *The American Journal of Otolaryngology*, vol. 1, no. 4, pp. 324–333, 1980.
- [53] D. Kepshire, S. C. Davis, H. Dehghani, K. D. Paulsen, and B. W. Pogue, "Fluorescence tomography characterization for sub-surface imaging with protoporphyrin IX," *Optics Express*, vol. 16, no. 12, pp. 8581–8593, 2008.
- [54] M. Dai, A. Nuttall, Y. Yang, and X. Shi, "Visualization and contractile activity of cochlear pericytes in the capillaries of the spiral ligament," *Hearing Research*, vol. 254, no. 1-2, pp. 100–107, 2009.
- [55] M. Dai and X. Shi, "Fibro-vascular coupling in the control of cochlear blood flow," *PLoS ONE*, vol. 6, no. 6, Article ID e20652, 2011.
- [56] D. Kleinfeld, P. P. Mitra, F. Helmchen, and W. Denk, "Fluctuations and stimulus-induced changes in blood flow observed in individual capillaries in layers 2 through 4 of rat neocortex," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 95, no. 26, pp. 15741–15746, 1998.
- [57] E. Chaigneau, M. Oheim, E. Audinat, and S. Charpak, "Two-photon imaging of capillary blood flow in olfactory bulb glomeruli," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 22, pp. 13081–13086, 2003.
- [58] F. Helmchen and D. Kleinfeld, "In vivo measurements of blood flow and glial cell function with two-photon laser-scanning microscopy," in *Angiogenesis: In Vivo Systems, Part A*, D. A. Cheresh, Ed., pp. 231–254, Elsevier, 2008.
- [59] W. S. Kamoun, S.-S. Chae, D. A. Lacorre et al., "Simultaneous measurement of RBC velocity, flux, hematocrit and shear rate in vascular networks," *Nature Methods*, vol. 7, no. 8, pp. 655–660, 2010.