

Review

# Role of Secreted Frizzled-Related Protein 1 in Early Mammary Gland Tumorigenesis and Its Regulation in Breast Microenvironment

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**Abstract:** In mice, the lack of *secreted frizzled-related protein* 1 (*SFRP1*) is responsible for mammogenesis and hyperplasia, while, in bovines, its overexpression is associated with post-lactational mammary gland involution. Interestingly, there are no reports dealing with the role of *SFRP1* in female involution. However, *SFRP1* dysregulation is largely associated with human tumorigenesis in the literature. Indeed, the lack of *SFRP1* is associated with both tumor development and patient prognosis. Considering the increased risk of breast tumor development associated with incomplete mammary gland involution, it is crucial to demystify the "grey zone" between physiological age-related involution and tumorigenesis. In this review, we explore the functions of *SFRP1* involved in the breast involution processes to understand the perturbations driven by the disappearance of *SFRP1* in mammary tissue. Moreover, we question the presence of recurrent microcalcifications identified by mammography. In bone metastases from prostate primary tumor, overexpression of *SFRP1* results in an osteolytic response of the tumor cells. Hence, we explore the hypothesis of an osteoblastic differentiation of mammary cells induced by the lack of *SFRP1* during lobular involution, resulting in a new accumulation of hydroxyapatite crystals in the breast tissue.

**Keywords:** breast cancer; *secreted frizzled-related protein 1; SFRP1*; lobular involution; breast involution; microcalcifications; inflammation; apoptosis; osteomimic microenvironment; osteoblast-like cells

# 1. Introduction

*Secreted frizzled-related protein* 1 (*SFRP1*) encodes one of the eight known SFRP family members. This 3873 base pair coding gene is localized on chromosome 8p11, from nucleotide 41,261,957 to 41,309,473 (Figure 1A). Composed of 3 exons, it can be spliced into two different isoforms described in Figure 1B. Translation of *SFRP1* results in a 314 amino acid (aa) protein composed of three major domains, schematized in Figure 1C. The first domain (1 to 31 aa) is a peptide signal (PS) that allows the protein to be transported to the extracellular matrix after synthesis. The second domain (57 to 171aa), the frizzled (FRI), is an extracellular cysteine rich domain (CRD) largely conserved among species [1]. It is composed of 10 cysteines forming disulfide bonds visible on the alpha-helical crystal structure of the domain [2]. Due to the fact of its homology, the *SFRP1* CRD binds the Wnt-binding site of frizzled proteins (Fz) [3–5]. Bafico et al. [6] reported that *SFRP1* is also able to directly interact with Fz resulting in a non-functional complex.





**Figure 1.** *Secreted frizzled-related protein 1* (*SFRP1*) identity. (**A**) Schematic representation of *SFRP1* position on chromosome 8. (**B**) Schematic representation of *SFRP1* isoforms. (**C**) Schematic representation of *SFRP1* protein domains. (**D**) Top 10 of protein–protein interactions between SFRP1 and Wnt signaling pathways. Figure drawn using Simple modular architecture research tool (SMART) [7]. Wnt canonical (**E**) and non-canonical (**F**) pathways in physiological context.

## 2. SFRP1 in Wnt Signaling Pathway

In 1997, Finch et al. [8] described *SFRP1* as a secreted Wnt antagonist. In fact, *SFRP1* PS allows the protein to be transported to the extracellular matrix. In this compartment, *SFRP1* fixes Wnt molecules, inducing a downregulation of the Wnt signaling pathway. The Wnt molecules caption induces a decrease of the intracellular  $\beta$ -catenin level which is responsible for the downregulation of Wnt target genes' transcription. As a primordial regulator of cell growth, cell polarity, cell fate determination, and malignant transformation, *SFRP1* is widely expressed in human cells [8]. Later, in 2000, Uren et al. [9] reported that low concentration of SFRP1 could potentiate Wnt signaling

pathway activity rather than inhibiting it. This result suggests the existence of a high-affinity site of liaison involved in low SFRP1 concentration and conversely a low affinity site of liaison in high SFRP1 concentration. *Secreted frizzled-related protein 1* (*SFRP1*) is known to downregulate canonical Wnt pathway. Moreover, the important homology between Wnt molecules suggests that *SFRP1* could be able to regulate the Wnt non-canonical pathway as well (Figure 1D–F). In fact, in prostate epithelial cells, the overexpression of *SFRP1* had no impact on intracellular  $\beta$ -catenin level. However, it activated the WNT/JNK pathway [10]. To date, the processes used by *SFRP1* to regulate cell activity remain poorly understood. Nevertheless, the Wnt signaling pathway, highly conserved during evolution, has a crucial role in embryonic development and in adult tissues proliferation, differentiation, and apoptosis. Because of these crucial roles, dysregulation of the Wnt signaling pathway antagonists is strongly associated with many diseases and most importantly cancer [11–13].

### 3. SFRP1 Expression Is Deregulated in Breast Cancer

Many evidence that SFRP1 dysregulation is involved in tumorigenesis is available in the literature [10,14–37]. Hence, understanding the mechanism of SFRP1 remains essential in order to develop cancer treatments. Several studies have demonstrated that SFRP1 in breast cancer is downregulated at both the mRNA and protein levels (Table 1). Moreover, this downregulation is also associated with poor survival outcome [25]. It has also been reported that SFRP1 is wildly downregulated in breast malignant lesions. Under this scenario, the activation of canonical and non-canonical Wnt pathways depends on breast cancer subtypes or vice versa [29]. In fact, Huth et al. [29] reported that the canonical Wnt pathway is affected in basal-like breast cancer while the non-canonical Wnt pathway is affected in luminal-like breast cancer cells. This diversity in the operating mode makes the underlying molecular mechanisms of SFRP1 particularly difficult to understand. However, the studies described in Table 1 reported that SFRP1 is under-expressed in breast cancer tissue compared to normal tissue. In fact, as a down-regulator of the Wnt signaling pathway, SFRP1 is involved in the negative control of cell proliferation, differentiation, and survival. In other words, SFRP1 is a tumor-suppressor in physiological conditions. Furthermore, in triple-negative breast cancer cases, the over-expression of SFRP1 is responsible for an increasing sensitivity to chemotherapy [32]. By its pro-apoptotic effect, SFRP1 is a potential therapeutic tool that should be further investigated. Interestingly, if we focus specifically on breast tissue, the downregulation of SFRP1 in both human and mice SFRP1 -/- is responsible for the increasing estrogen-induced response and hyperplasia [30]. In normal mammary epithelial cells, estrogen positively regulates SFRP1 expression [25]. A genome-wide association study reported that, in the SFRP1-modulated gene interaction network, 27 genes were modulated in a cluster involved in estrogen stimulus response [31]. To exacerbate the evidence of estrogen pathway involvement in SFRP1-related breast tumorigenesis, Bernemann et al. [32] showed that SFRP1 is overexpressed in triple-negative breast tumors compared with other breast tumors. The relationship between SFRP1 and estrogen is an important facet to investigate. Indeed, breast evolution during a woman's life is hormone-dependent. Furthermore, estrogen decreases dramatically during menopause which is responsible for a negative regulation of the bone resorption and formation, resulting in loss of bone mass [33,34]. That decrease of bone mass was associated with the increasing expression of miR-542-3p, involved in the downregulation of osteoblast differentiation by targeting SFRP1 in rats after ovariectomy-related osteoporosis [35]. Understanding the cross-regulation between SFRP1 and estrogen sensitivity could allow us to better understand both breast tumorigenesis and menopause-related osteoporosis.

Alterations in SFRP1	Role of SFRP1	Population/Breast Cancer Subtypes	Signaling Pathway Involved in Tumorigenesis	Reference
Higher methylation and lower expression of SFRP1 mRNA in tumors compared to non-tumoral tissues	Tumor-suppressor	NA */all subtypes	NA	Veeck et al. (2006) [28]
Under-expression of <i>SFRP1</i> mRNA and protein in tumors compared to non-tumoral tissues	Tumor-suppressor	NA/ductal and lobular carcinomas	NA	Dahl et al. (2007) [15]
Higher methylation, deletion, and under-expression of <i>SFRP1</i> mRNA in tumors compared to non-tumoral tissues	Tumor-suppressor	Indians/67% ER **/PR ***	Canonical Wnt signaling pathway	Mukherjee et al. (2012) [27]
Higher expression in TNBC ****compared to other BC subtypes	Higher sensitivity to chemotherapy in TNBC overexpressing <i>SFRP1</i>	NA/all subtypes	Wnt and TGF-β ***** signaling pathways	Bernemann et al. (2014) [32]
Under-expression of <i>SFRP1</i> mRNA in tumors compared to non-tumoral tissues	Tumor-suppressor	NA/basal-like	Canonical Wnt signaling pathway	Huth et al. (2014) [29]
Under-expression of <i>SFRP1</i> mRNA in tumors compared to non-tumoral tissues	Tumor-suppressor	NA/luminal-like HER2 positive	Non-canonical Wnt signaling pathway	-
Estrogen-mediated signaling is differentially affected by the expression levels of Sfrp1 in mammary epithelial cells: Estrogen signaling and <i>SFRP1</i> expression	Tumor-suppressor	NA/luminal-like	Estrogen-mediated and Wnt signaling pathways	Gregory and Schneider (2015) [25]
Higher methylation and lower expression of <i>SFRP1</i> mRNA in tumors compared to non-tumoral tissues	Tumor-suppressor	Chineses /all subtypes	NA	Li et al. (2016) [26]
Copy number alterations induce under-expression of <i>SFRP1</i> mRNA in tumors compared to non-tumoral tissues	Tumor-suppressor	NA/all subtypes	NA	Zhu et al. (2018) [36]
Under-expression of <i>SFRP1</i> is associated with aggressiveness	Tumor-suppressor	NA/breast cancer risk continuum	NA	Kothari et al. (2018) [37]
Higher expression in begnin tissues compared to hyperplasias	Tumor-suppressor	NA/ductal and lobular hyperplasias	Estrogen-mediated and Wnt signaling pathways	Gregory et al. (2019) [30]

# **Table 1.** Dysregulation of *SFRP1* in human breast lesions or cell lines reported in the literature.

\* NA = non-applicable, \*\* ER = estrogen receptor, \*\*\* PR = progesterone receptor, \*\*\*\* TNBC = triple-negative breast cancer, \*\*\*\*\* TGF-  $\beta$  = transforming growth factor beta

To better understand which process induces SFRP1 expression dysregulation, some studies focused on the epigenetic profile of the gene. As described in Table 1, SFRP1 downregulation is associated with the hypermethylation of the SFRP1 promoter region's CpG islands, suggesting that epigenetics could be as crucial as gene aberrations in breast tumorigenesis [15,26–28]. In 2006, Veeck et al. [28] reported that the hypermethylation of SFRP1 promoter was the principal cause of gene silencing in breast cancer. Furthermore, they associated this hypermethylation to an unfavorable prognosis for patients. This was also demonstrated in 2007 in renal cell carcinomas by Dahl et al. [15] suggesting that the hypermethylation of SFRP1 promoter contributes to the development of multiple human tumors. Later in 2012, Mukherjee et al. [27] reported that 67% of breast cancer samples were altered in SFRP1. In the same cohort, 56% of samples were hypermethylated in the SFRP1 promoter region, suggesting once more that the hypermethylation of SFRP1 promoter is responsible for SFRP1 silencing in breast cancer [27]. More recently, the integration of expression data and next-generation sequencing allowed Li et al. [26] to characterize the correlation between SFRP1 promoter methylation and gene expression regulation. They demonstrated a strong negative correlation between both, suggesting that hypermethylation of SFRP1 promoter is responsible for SFRP1 silencing in breast cancer. Furthermore, they reported that this methylation quantification was a better parameter to improve the diagnosis of the disease [26]. The SFRP1 epigenetic seems to be a crucial player in early breast tumorigenesis and needs to be explored to better understand the mechanisms of tumor development.

# 4. SFRP1 Has a Major Role in the Lobular Involution Process

# 4.1. Involution

Mammary glands are continuously evolving during a woman's life and hormone cycles (Figure 2A). In the course of lobular evolution steps, lobular involution in both post-lactation and peri-menopause contexts is strongly associated with breast cancer risk. Indeed, many studies have demonstrated an inverse relationship between the degree of lobular involution and breast cancer risk in both normal and benign breast lesions [38-42]. The word "involution" is commonly used to describe an inward folding. In the breast tissue, the term "involution" can be used to describe the regression of the mammary gland. Such process occurs at the end of the breastfeeding period, to return to the pre-pregnancy mammary gland size. During this reversible involution, useless acinis are degraded by the immune system and replaced by collagen and adipose tissue. During the peri-menopausal period, an irreversible involution of the mammary gland occurs. In fact, with the impossibility to procreate comes the futility of milk production by the breast. To limit the risk of malignant transformation, useless epithelial cells are eliminated from the breast by the immune system and replaced by adipose tissue. This process remains poorly understood, notably because of the important intrapersonal variability. In fact, some women benefit from a total disappearance of types 2–3 lobules, resulting in a total absence of acinis at the end of the lobular involution, while others have an incomplete mammary gland regression increasing their number of epithelial cells in the breast tissue hence increasing, at the same time, their risk in developing a breast tumor. There is as yet no study reporting evidence of a relationship between SFRP1 and lobular involution in women. However, SFRP1 appears to have a major impact on breast hyperplasia in mice, as Gauger et al. [43] reported that, in virgin knock-down SFRP1  $^{-/-}$  mice, the ductal branching and the lobulo-alveolar activity of the breast was comparable to mid-pregnant wild-type mice. On the other hand, Zheng et al. [44] described that, in bovines, SFRP1 is overexpressed in the late lactation process compared to the early peak of lactation, suggesting the implication of SFRP1 in reversible lobular involution following lactation. Taken together, these results suggest a causative role of SFRP1 in both early regulation of breast development and reversible post-lactation involution. Regarding a potential role of SFRP1 in breast involution and considering the protective role of mammary involution against breast tumorigenesis [39–42,45–51], SFRP1 could be a crucial player, to optimize lobular involution and decrease breast cancer risk. To better understand

the role of *SFRP1* in breast involution, we describe the implication of this protein in each molecular step involved in the lobular involution hereafter.



Figure 2. SFRP1 downregulation in breast tissue induces a chain reaction responsible for the malignant transformation of the cells. (A) Mammary glands evolve throughout a woman's life until reaching their growth peak during breastfeeding. Afterwards, acinis remain useful, so the breast starts a reversible involution to destroy the excessive glandular and epithelial tissue. During the peri-menopausal step of a woman's life, another involution appears to eliminate the rest of the useful tissue, decreasing at the same time the risk of breast cancer development. As a down-regulator of cell growth, SFRP1 under-expression could be responsible for the incomplete age-related breast involution resulting in a malignant transformation of the breast in the presence of microcalcifications. (B) The over-activation of the Wnt signaling pathway due to the SFRP1 down-regulation in breast tissue is responsible for the increasing proliferation and migration of mammary cells. The local inflammation needed to start the physiological age-related breast involution is responsible for the recruitment of the immune system. In the presence of microcalcifications, monocytes are able to differentiate in osteoclasts, while mesenchymal stem cells can differentiate in osteoblasts responsible for de novo microcalcifications formation. Furthermore, the replacement of epithelial tissue by adipose tissue needs pre-adipocytes maturation which is downregulated by the under-expression of SFRP1. The production of adipokines by immature adipocytes generates a chronic inflammation exacerbated by the hypersensitivity of epithelial cells to TGF-  $\beta$  due to the SFRP1 under-expression.

#### 4.2. Adipogenesis

The mammary gland involution is described by the gradual replacement of breast epithelium and stroma by collagen and adipose tissue and by the regression of type 2–3 lobules to type 1 lobules [52]. Many studies focus on obesity-related unbalance resulting in the disturbed secretion of inflammatory molecules named adipokines by the adipose tissue [53–58]. Interestingly, adipogenesis also occurs during breast involution, so that adipokines overexpression could appear during this specific step. In fact, the interaction between cancer cells and adipocytes could result in their reprogramming in cancer-associated adipocytes (CAAs), responsible for an abundant secretion of adipokines which upregulate the adhesion, migration, and invasion of the cells [54,55]. Furthermore, in mice, evidence of adipocytes delipidation to produce de novo lipids by lactating epithelial cells was reported [54,59]. Co-cultures of adipocyte stem cells with MCF7 cells showed an enhancing proliferation of the tumoral cells due to the estrogen-activated response mediated through leptin [53]. Interestingly, in mice adipocytes, SFRP1 expression increases during adipogenesis [60]. In humans, SFRP1 is also upregulated in early adipogenesis [61], both at the mRNA and protein levels. Legathu et al. [61] reported that SFRP1 expression is increased dramatically following induction of adipocytes differentiation. This SFRP1 overexpression induces the decrease of intracellular  $\beta$ -catenin levels, suggesting that adipogenesis is activated by the negative regulation of the Wnt canonical signaling pathway by SFRP1 [61]. In SFRP1 <sup>-/-</sup> mice, when obesity is induced, there is an increase in the inflammatory response associated with adipogenesis, and an upregulation of macrophage activity [62]. This inflammatory exacerbation is also associated with mammary branching [63]. Furthermore, body weight gain in post-menopausal women is associated with higher breast cancer risk [64,65]. Age-related weight gain in women, combined with the replacement of breast epithelium by adipose tissue during involution suggest that adipose tissue could play an important role in early tumorigenesis. In the case of an abnormal under-expression of SFRP1, we can hypothesize that pre-adipocytes and adipocytes stem cells are not able to differentiate in mature adipocytes, resulting in an overexpression of cytokines and adipokines responsible for a chronic inflammation of the breast tissue. The incomplete involution potentially associated with the lack of SFRP1 results in the presence of numerous epithelial cells able to delipidate adipocytes to make more energy. That lack of SFRP1 results also in the hyper-activation of the Wnt signaling pathway, which is responsible for enhancing cell proliferation, adhesion, and survival. The SFRP1 gene seems crucial at many steps for maintaining the balance between normal adipose tissue remodeling and tumorigenesis.

#### 4.3. Inflammation

The SFRP1 gene is upregulated in physiological inflammatory conditions, such as during post-lactation involution, suggesting a role in the degradation of acini by the immune system [66]. Interestingly, in periodontal tissue, the inhibition of SFRP1 at the protein level with a specific antibody is associated with less inflammation and a significant reduction of apoptosis in periodontitis context [67]. The existence of a crosslink between Wnt signaling pathway and TGF- $\beta$  signaling pathway seems to be the explanation of such cross-regulations between inflammation and the Wnt signaling pathway. In fact, the under-expression of SFRP1 in non-malignant cell lines results in the increasing sensitivity of the cell to the anti-inflammatory molecule TGF- $\beta$  [68]. Furthermore, Dzialo et al. [69] proposed a model in which TGF- $\beta$  activates the Wnt canonical signaling pathway by producing Wnt molecules. Interestingly, chronic inflammation of the breast tissue is inversely associated with lobular involution [52]. This suggests that anti-inflammatory processes could also promote cell proliferation and differentiation. In a breast cancer context, the accumulation of Wnt molecules due to the TGF- $\beta$ pathway, combined with the lack of SFRP1, result in a dramatic over-activation of the Wnt signaling pathway and, consequently, in an increasing cell proliferation (Figure 2B). This suggests a sensitive balance between the inflammation needed to initiate the destruction of the epithelial cells by the immune system and the chronic inflammation responsible for decreasing breast involution and increasing hyperplasia. We hypothesize that the lack of SFRP1 is responsible for an upregulation of TGF- $\beta$ sensitivity which is responsible for the increase of Wnt molecule production and so—an abnormal

hyperplasia. The combination of increasing TGF- $\beta$  sensitivity and adipocyte-related inflammation could result in chronic inflammation resulting in an incomplete breast involution and a higher risk of breast tumorigenesis. Furthermore, high estrogen levels decrease immune reaction in the breast tissue [70]. This evidence suggests that a post-menopausal lobular involution could result in a higher risk of chronic inflammation, promoting breast tumorigenesis.

# 4.4. Apoptosis

Lobular involution is also characterized by both apoptosis of epithelial cells and lobulo-alveolar remodeling [71,72]. Regarding apoptosis, few studies have reported an association between SFRP1 expression and apoptosis in breast tissue. In fact, the lack of SFRP1 affects apoptotic gene expression and activity, suggesting a major role for this gene in the regulation of cell survival and proliferation in the breast [73]. In human colorectal cancer cell lines, Wang et al. [23] demonstrated that SFRP1 overexpression promotes apoptosis. Similar results have been reported in a cholangiocarcinoma context using miR-191 to knockdown SFRP1 [74]. In bone marrow-derived mesenchymal stem cells, the use of miR-144 targeting SFRP1 inhibited apoptosis as well [75]. Genome-wide identification of key modulators implicated in breast tumorigenesis demonstrated a significant effect of immunity processes in breast tumor development. In fact, among the top clusters of gene ontology (GO) [76,77] terms enriched in modulator genes, the majority were directly involved in immune cell activation, apoptosis, and inflammatory response to immunity [29,31,78]. Furthermore, another cluster highlighted by these studies is involved in tissue remodeling, specifically in cell adhesion. Moreover, Chiu et al. [31] focused their analyses on specific SFRP1-modulated genes in breast cancer, and among the top six clusters of GO [76,77] terms, four were involved in tissue remodeling like extracellular structure or cytoskeleton organization. Regarding the top cluster enriched for microRNAs in human primary breast nodules, Yang et al. [78] concluded, once more, that apoptosis and inflammation were dramatically dysregulated. These results taken together suggest that the lack of SFRP1 in breast could be responsible for a downregulation of epithelial cells apoptosis and tissue remodeling which is responsible for an incomplete involution associated with an increasing breast cancer risk and, in worst cases, in an hyperplasia responsible for the early development of breast tumor.

# 5. SFRP1 Expression Dysregulation Is Responsible for an Osteoblastic Differentiation of Breast Cells and an Accumulation of Microcalcifications

## 5.1. SFRP1 and Osteoblast-Like Cells in the Breast

The SFRP1 gene is a well-known important regulator of bone remodeling. Indeed, SFRP1 is expressed by osteoblasts and inhibits osteoclast formation through its binding with receptor activator of nuclear factor kappa-B ligand (RANKL) [79,80]. In human fetal osteoblastic cell lines, the inhibition of SFRP1 resulted in the promotion of osteoblastic differentiation due to the activation of the Wnt/ $\beta$ -catenin signaling pathway [81]. Tang et al. [75] demonstrated in bone marrow-derived mesenchymal stem cells that a decrease in SFRP1 expression using miR-144 induces osteoblastic differentiation of the cells. Interestingly, the lack of SFRP1 also seems to be involved in cells activity. Once differentiated, the lack of *SFRP1* is responsible for the upregulation of trabecular bone formation by the osteoblasts in mice [82]. Furthermore, many microRNAs-related dysregulations in breast tumor concern the bone morphogenic protein response, vitamin D response, and osteoblasts proliferation [78]. These pathways are directly related to the phospho-calcium homeostasis [83-85]. Furthermore, vitamin D is also known as a down-regulator of the Wnt pathway and an up-regulator of the TGF- $\beta$  pathway, suggesting a crosstalk between bone homeostasis, inflammation, and apoptosis [86] that could be involved in breast cancer tumorigenesis. Kothari et al. [37] described another evidence of this potential osteoblast differentiation in the breast. The authors performed transcriptome analyses of breast lesions at different stages of tumor aggressiveness. Expression of SFRP1 decreases with tumor progression. Interestingly, this decrease is accompanied with the increase of Secreted Phosphoprotein 1

(*SPP1*) and *Periostin* (*POSTN*) expression [37]. *Secreted Phosphoprotein* 1, also named *Osteopontin*, is expressed by osteoblasts to promote bone resorption by increasing osteoclasts bone adhesion [87,88]. *Periostin* is also produced by osteoblasts and plays a preponderant role in tissue development and regeneration including wound healing [89,90]. In the context of inflammatory bone disease, resolving the resorption-induced inflammation by decreasing *SFRP1* expression induces the Wnt canonical signaling pathway activation which is essential for osteoblasts differentiation and activity [91–94]. With this evidence, we hypothesize that osteoblast differentiation in the breast tissue could result in a de novo bone construction.

## 5.2. SFRP1 and Microcalcifications

Breast microcalcifications are present in 30% to 50% of all malignant breast lesions. Most breast calcifications visible at mammography are made of a combination of calcium and phosphate, the exact composition of the mineral part of bones. Moreover, evidences of osteoblast-like cells in breast tissue suggest a potential for breast cells differentiation in a microcalcification environment [37,95–103]. In 2005, Morgan et al. [99] reported that microcalcifications were responsible for increasing the mitogenesis properties of breast cells, resulting in an amplification of the malignant process. This evidence was validated in ductal carcinoma in situ in 2019 by He et al. [100]. In 2012, Cox et al. [101] described in vitro that breast cancer cells were able to adapt to the bone microenvironment by developing osteomimetic characteristics. To further evidence the capability of breast cells to osteomimicry, Scimeca et al. [102] demonstrated, in 2019, that the presence of microcalcifications combined with the presence of activated monocytes induce an epithelial to mesenchymal transition. Furthermore, they proved an osteoblast phenotype acquisition of the cells producing hydroxyapatite by performing immunohistochemical analysis. This capacity of breast cells to develop osteomimetic properties were also reported in 2019 by O'Grady et al. [103]. In fact, they demonstrated the ability of breast cancer cell lines to mineralize, and doing so, proved the capacity of breast cancer cells to produce de novo microcalcifications in osteogenic conditions [103]. Furthermore, microcalcifications are described as an indirect sign of pathological process, such as inflammation, and, as described previously, inflammation is also associated with breast involution in both post-lactation and peri-menopausal contexts, but also in tumorigenesis and tumoral growth. These results taken together suggest that a lack of SFRP1 in breast tissue could be responsible for osteoblastic differentiation, resulting in accumulation of new microcalcifications in the breast tissue.

#### 6. Conclusions

We propose that the post-menopausal lobular involution initiation is a risk factor of breast cancer development due to the concomitant falling of estrogens and SFRP1. Lack of SFRP1 induces a decrease of the adipogenesis needed for the replacement of epithelial cells in breast during lobular involution. Associated with that, the decrease of pre-adipocytes and adipocytes stem cells maturation, also due to the lack of SFRP1, increases dramatically the expression of adipokines resulting in an important inflammatory reaction. Furthermore, the low expression of SFRP1 is responsible for an exacerbation of TGF- $\beta$  sensitivity which is responsible for the chronic inflammation installation and the upregulation of Wnt molecule production. Increasing Wnt molecule quantity, added to the absence of SFRP1 as a down-regulator of the Wnt signaling pathway, results in an in-chain reaction in which proliferation, adhesion, and migration are dramatically exacerbated. Furthermore, microcalcifications added to the Wnt signaling pathway over-stimulation result in osteoblastic differentiation of the cells and the production of new hydroxyapatite crystals in breast tissue (Figure 2B). Finally, it is essential to understand the exact role of SFRP1 in both physiological lobular involution and tumor development. The *SFRP1* gene could become an important key player to optimize the protective effect of breast involution. Moreover, understanding the underlying mechanisms used by SFRP1 to regulate mammary gland evolution could be crucial to prevent early breast tumorigenesis.

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# References

- Rehn, M.; Pihlajaniemi, T. Identification of three N-terminal ends of type XVIII collagen chains and tissue-specific differences in the expression of the corresponding transcripts. *J. Biol. Chem.* 1995, 270, 4705–4711. [CrossRef]
- 2. Dann, C.E.; Hsieh, J.-C.; Rattner, A.; Sharma, D.; Nathans, J.; Leahy, D.J. Insights into Wnt binding and signaling from the structures of two Frizzled cysteine-rich domains. *Nature* **2001**, *412*, 86–90. [CrossRef]
- 3. Bhanot, P.; Brink, M.; Samos, C.H.; Hsieh, J.-C.; Wang, Y.; Macke, J.P.; Andrew, D.; Nathans, J.; Nusse, R. A new member of the frizzled family from Drosophila functions as a Wingless receptor. *Nature* **1996**, *382*, 225–230. [CrossRef]
- Lin, K.; Wang, S.; Julius, M.A.; Kitajewski, J.; Moos, M.; Luyten, F.P. The cysteine-rich frizzled domain of Frzb-1 is required and sufficient for modulation of Wnt signaling. *Proc. Natl. Acad. Sci. USA* 1997, 94, 11196–11200. [CrossRef]
- 5. Rehn, M.; Pihlajaniemi, T.; Hofmann, K.; Bucher, P. The frizzled motif: In how many different protein families does it occur? *Trends Biochem. Sci.* **1998**, *23*, 415–417. [CrossRef]
- Bafico, A.; Gazit, A.; Pramila, T.; Finch, P.W.; Yaniv, A.; Aaronson, S.A. Interaction of Frizzled Related Protein (FRP) with Wnt Ligands and the Frizzled Receptor Suggests Alternative Mechanisms for FRP Inhibition of Wnt Signaling. *J. Biol. Chem.* 1999, 274, 16180–16187. [CrossRef]
- Letunic, I.; Bork, P. 20 years of the SMART protein domain annotation resource. *Nucleic Acids Res.* 2018, 46, D493–D496. [CrossRef]
- 8. Finch, P.W.; He, X.; Kelley, M.J.; Uren, A.; Schaudies, R.P.; Popescu, N.C.; Rudikoff, S.; Aaronson, S.A.; Varmus, H.E.; Rubin, J.S. Purification and molecular cloning of a secreted, Frizzled-related antagonist of Wnt action. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 6770–6775. [CrossRef]
- Üren, A.; Reichsman, F.; Anest, V.; Taylor, W.G.; Muraiso, K.; Bottaro, D.P.; Cumberledge, S.; Rubin, J.S. Secreted Frizzled-related Protein-1 Binds Directly to Wingless and Is a Biphasic Modulator of Wnt Signaling. *J. Biol. Chem.* 2000, 275, 4374–4382. [CrossRef]
- 10. Joesting, M.S.; Cheever, T.R.; Volzing, K.G.; Yamaguchi, T.P.; Wolf, V.; Naf, D.; Rubin, J.S.; Marker, P.C. Secreted frizzled related protein 1 is a paracrine modulator of epithelial branching morphogenesis, proliferation, and secretory gene expression in the prostate. *Dev. Biol.* **2008**, *317*, 161–173. [CrossRef]
- 11. Kawano, Y. Secreted antagonists of the Wnt signaling pathway. J. Cell Sci. 2003, 116, 2627–2634. [CrossRef]
- 12. Klaus, A.; Birchmeier, W. Wnt signaling and its impact on development and cancer. *Nat. Rev. Cancer* **2008**, *8*, 387–398. [CrossRef]
- 13. Reya, T.; Clevers, H. Wnt signaling in stem cells and cancer. Nature 2005, 434, 843–850. [CrossRef]
- Caldwell, G.M.; Jones, C.; Gensberg, K.; Jan, S.; Hardy, R.G.; Byrd, P.; Chughtai, S.; Wallis, Y.; Matthews, G.M.; Morton, D.G. The Wnt Antagonist sFRP1 in Colorectal Tumorigenesis. *Cancer Res.* 2004, 64, 883–888. [CrossRef]
- 15. Dahl, E.; Wiesmann, F.; Woenckhaus, M.; Stoehr, R.; Wild, P.J.; Veeck, J.; Knüchel, R.; Klopocki, E.; Sauter, G.; Simon, R.; et al. Frequent loss of SFRP1 expression in multiple human solid tumours: Association with aberrant promoter methylation in renal cell carcinoma. *Oncogene* **2007**, *26*, 5680–5691. [CrossRef]
- Delic, S.; Lottmann, N.; Stelzl, A.; Liesenberg, F.; Wolter, M.; Götze, S.; Zapatka, M.; Shiio, Y.; Sabel, M.C.; Felsberg, J.; et al. MiR-328 promotes glioma cell invasion via SFRP1-dependent Wnt-signaling activation. *Neuro Oncol.* 2014, *16*, 179–190. [CrossRef]
- 17. Kawano, Y.; Diez, S.; Uysal-Onganer, P.; Darrington, R.S.; Waxman, J.; Kypta, R.M. Secreted Frizzled-related protein-1 is a negative regulator of androgen receptor activity in prostate cancer. *Br. J. Cancer* **2009**, *100*, 1165–1174. [CrossRef]

- Kierulf-Vieira, K.S.; Sandberg, C.J.; Grieg, Z.; Günther, C.-C.; Langmoen, I.A.; Vik-Mo, E.O. Wnt inhibition is dysregulated in gliomas and its re-establishment inhibits proliferation and tumor sphere formation. *Exp. Cell Res.* 2016, 340, 53–61. [CrossRef]
- 19. Liang, J.; Kang, X.; Halifu, Y.; Zeng, X.; Jin, T.; Zhang, M.; Luo, D.; Ding, Y.; Zhou, Y.; Yakeya, B.; et al. Secreted frizzled-related protein promotors are hypermethylated in cutaneous squamous carcinoma compared with normal epidermis. *BMC Cancer* **2015**, *15*, 641. [CrossRef]
- 20. Qu, Y.; Ray, P.S.; Li, J.; Cai, Q.; Bagaria, S.P.; Moran, C.; Sim, M.-S.; Zhang, J.; Turner, R.R.; Zhu, Z.; et al. High levels of secreted frizzled-related protein 1 correlate with poor prognosis and promote tumourigenesis in gastric cancer. *Eur. J. Cancer* **2013**, *49*, 3718–3728. [CrossRef]
- 21. Ren, J.; Wang, R.; Huang, G.; Song, H.; Chen, Y.; Chen, L. sFRP1 Inhibits Epithelial–Mesenchymal Transition in A549 Human Lung Adenocarcinoma Cell Line. *Cancer Biother. Radiopharm.* **2013**, *28*, 565–571. [CrossRef]
- 22. Tian, J.; He, H.; Lei, G. Wnt/β-catenin pathway in bone cancers. *Tumor Biol.* **2014**, *35*, 9439–9445. [CrossRef]
- 23. Wang, Z.; Li, R.; He, Y.; Huang, S. Effects of secreted frizzled-related protein 1 on proliferation, migration, invasion, and apoptosis of colorectal cancer cells. *Cancer Cell Int.* **2018**, *18*, 48. [CrossRef] [PubMed]
- Yang, H.; Liu, G.; Liu, Y.; Zhao, H.; Yang, Z.; Zhao, C.; Zhang, X.; Ye, H. Over-expression of microRNA-940 promotes cell proliferation by targeting GSK3β and sFRP1 in human pancreatic carcinoma. *Biomed. Pharmacother.* 2016, *83*, 593–601. [CrossRef] [PubMed]
- Gregory, K.J.; Schneider, S.S. Estrogen-mediated signaling is differentially affected by the expression levels of Sfrp1 in mammary epithelial cells: Estrogen signaling and *Sfrp1* expression. *Cell Biol. Int.* 2015, *39*, 873–879. [CrossRef] [PubMed]
- Li, Z.; Heng, J.; Yan, J.; Guo, X.; Tang, L.; Chen, M.; Peng, L.; Wu, Y.; Wang, S.; Xiao, Z.; et al. Integrated analysis of gene expression and methylation profiles of 48 candidate genes in breast cancer patients. *Breast Cancer Res. Treat.* 2016, 160, 371–383. [CrossRef]
- Mukherjee, N.; Bhattacharya, N.; Alam, N.; Roy, A.; Roychoudhury, S.; Panda, C.K. Subtype-specific alterations of the Wnt signaling pathway in breast cancer: Clinical and prognostic significance. *Cancer Sci.* 2012, *103*, 210–220. [CrossRef]
- Veeck, J.; Niederacher, D.; An, H.; Klopocki, E.; Wiesmann, F.; Betz, B.; Galm, O.; Camara, O.; Dürst, M.; Kristiansen, G.; et al. Aberrant methylation of the Wnt antagonist SFRP1 in breast cancer is associated with unfavourable prognosis. *Oncogene* 2006, 25, 3479–3488. [CrossRef]
- 29. Huth, L.; Rose, M.; Kloubert, V.; Winkens, W.; Schlensog, M.; Hartmann, A.; Knüchel, R.; Dahl, E. BDNF Is Associated with SFRP1 Expression in Luminal and Basal-Like Breast Cancer Cell Lines and Primary Breast Cancer Tissues: A Novel Role in Tumor Suppression? *PLoS ONE* **2014**, *9*, e102558. [CrossRef]
- Gregory, K.J.; Roberts, A.L.; Conlon, E.M.; Mayfield, J.A.; Hagen, M.J.; Crisi, G.M.; Bentley, B.A.; Kane, J.J.; Makari-Judson, G.; Mason, H.S.; et al. Gene expression signature of atypical breast hyperplasia and regulation by SFRP1. *Breast Cancer Res.* 2019, 21, 76. [CrossRef]
- 31. Chiu, Y.-C.; Wang, L.-J.; Hsiao, T.-H.; Chuang, E.Y.; Chen, Y. Genome-wide identification of key modulators of gene-gene interaction networks in breast cancer. *BMC Genom.* **2017**, *18*, 679. [CrossRef] [PubMed]
- 32. Bernemann, C.; Hülsewig, C.; Ruckert, C.; Schäfer, S.; Blümel, L.; Hempel, G.; Götte, M.; Greve, B.; Barth, P.J.; Kiesel, L.; et al. Influence of secreted frizzled receptor protein 1 (SFRP1) on neoadjuvant chemotherapy in triple negative breast cancer does not rely on WNT signaling. *Mol. Cancer* **2014**, *13*, 174. [CrossRef] [PubMed]
- 33. Kalervo Väänänen, H.; Härkönen, P.L. Estrogen and bone metabolism. Maturitas 1996, 23, S65–S69. [CrossRef]
- 34. Epstein, S. Update of current therapeutic options for the treatment of postmenopausal osteoporosis. *Clin. Ther.* **2006**, *28*, 151–173. [CrossRef] [PubMed]
- 35. Zhang, X.; Zhu, Y.; Zhang, C.; Liu, J.; Sun, T.; Li, D.; Na, Q.; Xian, C.J.; Wang, L.; Teng, Z. miR-542-3p prevents ovariectomy-induced osteoporosis in rats via targeting SFRP1. *J. Cell. Physiol.* **2018**, 233, 6798–6806. [CrossRef] [PubMed]
- 36. Zhu, Q.-N.; Renaud, H.; Guo, Y. Bioinformatics-based identification of miR-542-5p as a predictive biomarker in breast cancer therapy. *Hereditas* **2018**, *155*, 17. [CrossRef]
- Kothari, C.; Ouellette, G.; Labrie, Y.; Jacob, S.; Diorio, C.; Durocher, F. Identification of a gene signature for different stages of breast cancer development that could be used for early diagnosis and specific therapy. *Oncotarget* 2018, *9*, 37407–37420. [CrossRef]

- Ghosh, K.; Vachon, C.M.; Pankratz, V.S.; Vierkant, R.A.; Anderson, S.S.; Brandt, K.R.; Visscher, D.W.; Reynolds, C.; Frost, M.H.; Hartmann, L.C. Independent Association of Lobular Involution and Mammographic Breast Density With Breast Cancer Risk. *JNCI J. Natl. Cancer Inst.* 2010, *102*, 1716–1723. [CrossRef]
- 39. Henson, D.E.; Tarone, R.E.; Nsouli, H. Lobular Involution: The Physiological Prevention of Breast Cancer. *JNCI J. Natl. Cancer Inst.* **2006**, *98*, 1589–1590. [CrossRef]
- 40. Milanese, T.R.; Hartmann, L.C.; Sellers, T.A.; Frost, M.H.; Vierkant, R.A.; Maloney, S.D.; Pankratz, V.S.; Degnim, A.C.; Vachon, C.M.; Reynolds, C.A.; et al. Age-Related Lobular Involution and Risk of Breast Cancer. *JNCI J. Natl. Cancer Inst.* **2006**, *98*, 1600–1607. [CrossRef]
- Radisky, D.C.; Visscher, D.W.; Frank, R.D.; Vierkant, R.A.; Winham, S.; Stallings-Mann, M.; Hoskin, T.L.; Nassar, A.; Vachon, C.M.; Denison, L.A.; et al. Natural history of age-related lobular involution and impact on breast cancer risk. *Breast Cancer Res. Treat.* 2016, 155, 423–430. [CrossRef] [PubMed]
- 42. Radisky, D.C.; Hartmann, L.C. Mammary Involution and Breast Cancer Risk: Transgenic Models and Clinical Studies. *J. Mammary Gland Biol. Neoplasia* **2009**, *14*, 181–191. [CrossRef] [PubMed]
- 43. Gauger, K.J.; Shimono, A.; Crisi, G.M.; Schneider, S. Loss of sfrp1 promotes ductal branching in the murine mammary gland. *BMC Dev. Biol.* **2012**, *12*, 25. [CrossRef] [PubMed]
- 44. Zheng, X.; Ning, C.; Dong, Y.; Zhao, P.; Li, J.; Fan, Z.; Li, J.; Yu, Y.; Mrode, R.; Liu, J. Quantitative proteome analysis of bovine mammary gland reveals protein dynamic changes involved in peak and late lactation stages. *Biochem. Biophys. Res. Commun.* **2017**, *494*, 292–297. [CrossRef] [PubMed]
- 45. Chollet-Hinton, L.; Puvanesarajah, S.; Sandhu, R.; Kirk, E.L.; Midkiff, B.R.; Ghosh, K.; Brandt, K.R.; Scott, C.G.; Gierach, G.L.; Sherman, M.E.; et al. Stroma modifies relationships between risk factor exposure and age-related epithelial involution in benign breast. *Mod. Pathol.* **2018**, *31*, 1085–1096. [CrossRef]
- Figueroa, J.D.; Pfeiffer, R.M.; Patel, D.A.; Linville, L.; Brinton, L.A.; Gierach, G.L.; Yang, X.R.; Papathomas, D.; Visscher, D.; Mies, C.; et al. Terminal Duct Lobular Unit Involution of the Normal Breast: Implications for Breast Cancer Etiology. JNCI J. Natl. Cancer Inst. 2014, 106. [CrossRef]
- Gierach, G.L.; Patel, D.A.; Pfeiffer, R.M.; Figueroa, J.D.; Linville, L.; Papathomas, D.; Johnson, J.M.; Chicoine, R.E.; Herschorn, S.D.; Shepherd, J.A.; et al. Relationship of Terminal Duct Lobular Unit Involution of the Breast with Area and Volume Mammographic Densities. *Cancer Prev. Res. Phila. Pa.* 2016, *9*, 149–158. [CrossRef]
- 48. Ginsburg, O.M.; Martin, L.J.; Boyd, N.F. Mammographic density, lobular involution, and risk of breast cancer. *Br. J. Cancer* **2008**, *99*, 1369–1374. [CrossRef]
- 49. Henson, D.E. On the possible role of involution in the natural history of breast cancer. *Cancer* **1993**, *71*, 2154–2156. [CrossRef]
- 50. Henson, D.E.; Tarone, R.E. Involution and the etiology of breast cancer. Cancer 1994, 74, 424–429. [CrossRef]
- 51. Walker, R.; Martin, C. The aged breast. J. Pathol. 2007, 211, 232–240. [CrossRef] [PubMed]
- 52. Hanna, M.; Dumas, I.; Orain, M.; Jacob, S.; Têtu, B.; Sanschagrin, F.; Bureau, A.; Poirier, B.; Diorio, C. Association between local inflammation and breast tissue age-related lobular involution among premenopausal and postmenopausal breast cancer patients. *PLoS ONE* 2017, *12*, e0183579. [CrossRef] [PubMed]
- 53. Strong, A.L.; Strong, T.A.; Rhodes, L.V.; Semon, J.A.; Zhang, X.; Shi, Z.; Zhang, S.; Gimble, J.M.; Burow, M.E.; Bunnell, B.A. Obesity associated alterations in the biology of adipose stem cells mediate enhanced tumorigenesis by estrogen dependent pathways. *Breast Cancer Res.* **2013**, *15*, R102. [CrossRef]
- 54. Tan, J.; Buache, E.; Chenard, M.-P.; Dali-Youcef, N.; Rio, M.-C. Adipocyte is a non-trivial, dynamic partner of breast cancer cells. *Int. J. Dev. Biol.* **2011**, *55*, 851–859. [CrossRef]
- 55. Nieman, K.M.; Romero, I.L.; Van Houten, B.; Lengyel, E. Adipose tissue and adipocytes support tumorigenesis and metastasis. *Biochim. Biophys. Acta BBA Mol. Cell Biol. Lipids* **2013**, *1831*, 1533–1541. [CrossRef] [PubMed]
- 56. Delort, L.; Rossary, A.; Farges, M.-C.; Vasson, M.-P.; Caldefie-Chézet, F. Leptin, adipocytes and breast cancer: Focus on inflammation and anti-tumor immunity. *Life Sci.* **2015**, *140*, 37–48. [CrossRef]
- 57. Chamras, H.; Bagga, D.; Elstner, E.; Setoodeh, K.; Koeffler, H.P.; Heber, D. Preadipocytes stimulate breast cancer cell growth. *Nutr. Cancer* **1998**, *32*, 59–63. [CrossRef]
- 58. Christodoulatos, G.S.; Spyrou, N.; Kadillari, J.; Psallida, S.; Dalamaga, M. The Role of Adipokines in Breast Cancer: Current Evidence and Perspectives. *Curr. Obes. Rep.* **2019**, *8*, 413–433. [CrossRef]

- 59. Hovey, R.C.; Aimo, L. Diverse and Active Roles for Adipocytes during Mammary Gland Growth and Function. *J. Mammary Gland Biol. Neoplasia* **2010**, *15*, 279–290. [CrossRef]
- 60. Wang, L.; Wang, Y.; Meng, Y.; Zhang, C.; Di, L. GSK3-activated STAT5 regulates expression of SFRPs to modulate adipogenesis. *FASEB J.* 2018, *32*, 4714–4726. [CrossRef]
- 61. Lagathu, C.; Christodoulides, C.; Tan, C.Y.; Virtue, S.; Laudes, M.; Campbell, M.; Ishikawa, K.; Ortega, F.; Tinahones, F.J.; Fernández-Real, J.-M.; et al. Secreted frizzled-related protein 1 regulates adipose tissue expansion and is dysregulated in severe obesity. *Int. J. Obes.* **2010**, *34*, 1695–1705. [CrossRef] [PubMed]
- Gauger, K.J.; Bassa, L.M.; Henchey, E.M.; Wyman, J.; Bentley, B.; Brown, M.; Shimono, A.; Schneider, S.S. Mice Deficient in Sfrp1 Exhibit Increased Adiposity, Dysregulated Glucose Metabolism, and Enhanced Macrophage Infiltration. *PLoS ONE* 2013, *8*, e78320. [CrossRef] [PubMed]
- 63. Gauger, K.J.; Bassa, L.M.; Henchey, E.M.; Wyman, J.; Ser-Dolansky, J.; Shimono, A.; Schneider, S.S. The effects of diet induced obesity on breast cancer associated pathways in mice deficient in SFRP1. *Mol. Cancer* **2014**, *13*, 117. [CrossRef] [PubMed]
- 64. Stoll, B. Adiposity as a risk determinant for postmenopausal breast cancer. *Int. J. Obes.* **2000**, *24*, 527–533. [CrossRef]
- Welti, L.M.; Beavers, D.P.; Caan, B.J.; Sangi-Haghpeykar, H.; Vitolins, M.Z.; Beavers, K.M. Weight Fluctuation and Cancer Risk in Postmenopausal Women: The Women's Health Initiative. *Cancer Epidemiol. Biomark. Prev.* 2017, 26, 779–786. [CrossRef]
- Seol, M.B.; Bong, J.J.; Baik, M. Expression Profiles of Apoptosis Genes in Mammary Epithelial Cells. *Mol. Cells.* 2005, 20, 97–104.
- 67. Li, C.H.; Amar, S. Inhibition of SFRP1 Reduces Severity of Periodontitis. J. Dent. Res. 2007, 86, 873–877. [CrossRef]
- 68. Gauger, K.J.; Chenausky, K.L.; Murray, M.E.; Schneider, S.S. SFRP1 reduction results in an increased sensitivity to TGF-b signaling. *BMC Cancer* **2011**, *11*, 59. [CrossRef]
- 69. Działo, E.; Tkacz, K.; Błyszczuk, P. Crosstalk between TGF-β and WNT signaling pathways during cardiac fibrogenesis. *Acta Biochim. Pol.* **2018**, *65*, 341–349. [CrossRef]
- Quigley, D.A.; Tahiri, A.; Lüders, T.; Riis, M.H.; Balmain, A.; Børresen-Dale, A.-L.; Bukholm, I.; Kristensen, V. Age, estrogen, and immune response in breast adenocarcinoma and adjacent normal tissue. *Oncolmmunology* 2017, 6, e1356142. [CrossRef]
- 71. Furth, P.A.; Bar-Peled, U.; Li, M. Apoptosis and mammary gland involution: Reviewing the process. *Apoptosis* **1997**, 2, 19–24. [CrossRef] [PubMed]
- 72. Jindal, S.; Gao, D.; Bell, P.; Albrektsen, G.; Edgerton, S.M.; Ambrosone, C.B.; Thor, A.D.; Borges, V.F.; Schedin, P. Postpartum breast involution reveals regression of secretory lobules mediated by tissue-remodeling. *Breast Cancer Res.* **2014**, *16*, R31. [CrossRef] [PubMed]
- 73. Gauger, K.J.; Schneider, S.S. Tumour supressor secreted frizzled related protein 1 regulates p53-mediated apoptosis: SFRP1 loss reduces p53-mediated apoptosis. *Cell Biol. Int.* **2014**, *38*, 124–130. [CrossRef] [PubMed]
- 74. Kang, P.-C.; Leng, K.-M.; Liu, Y.-P.; Liu, Y.; Xu, Y.; Qin, W.; Gao, J.-J.; Wang, Z.-D.; Tai, S.; Zhong, X.-Y.; et al. miR-191 Inhibition Induces Apoptosis Through Reactivating Secreted Frizzled-Related Protein-1 in Cholangiocarcinoma. *Cell. Physiol. Biochem.* **2018**, *49*, 1933–1942. [CrossRef] [PubMed]
- 75. Tang, L.; Lu, W.; Huang, J.; Tang, X.; Zhang, H.; Liu, S. miR-144 promotes the proliferation and differentiation of bone mesenchymal stem cells by downregulating the expression of SFRP1. *Mol. Med. Rep.* **2019**, *20*, 270–280. [CrossRef]
- 76. Ashburner, M.; Ball, C.A.; Blake, J.A.; Botstein, D.; Butler, H.; Cherry, J.M.; Davis, A.P.; Dolinski, K.; Dwight, S.S.; Eppig, J.T.; et al. Gene Ontology: Tool for the unification of biology. *Nat. Genet.* 2000, 25, 25–29. [CrossRef]
- 77. Gene Ontology Consortium. The Gene Ontology Consortium the Gene Ontology Resource: 20 years and still GOing strong. *Nucleic Acids Res.* **2019**, *47*, D330–D338. [CrossRef]
- 78. Yang, Y.; Xing, Y.; Liang, C.; Hu, L.; Xu, F.; Chen, Y. Crucial microRNAs and genes of human primary breast cancer explored by microRNA-mRNA integrated analysis. *Tumor Biol.* **2015**, *36*, 5571–5579. [CrossRef]
- 79. Boudin, E.; Fijalkowski, I.; Piters, E.; Van Hul, W. The role of extracellular modulators of canonical Wnt signaling in bone metabolism and diseases. *Semin. Arthritis Rheum.* **2013**, *43*, 220–240. [CrossRef]

- Häusler, K.D.; Horwood, N.J.; Chuman, Y.; Fisher, J.L.; Ellis, J.; Martin, T.J.; Rubin, J.S.; Gillespie, M.T. Secreted Frizzled-Related Protein-1 Inhibits RANKL-Dependent Osteoclast Formation. *J. Bone Miner. Res.* 2004, 19, 1873–1881. [CrossRef]
- 81. Guo, D.; Li, Q.; Lv, Q.; Wei, Q.; Cao, S.; Gu, J. MiR-27a Targets sFRP1 in hFOB Cells to Regulate Proliferation, Apoptosis and Differentiation. *PLoS ONE* **2014**, *9*, e91354. [CrossRef] [PubMed]
- 82. Bodine, P.V.N.; Zhao, W.; Kharode, Y.P.; Bex, F.J.; Lambert, A.-J.; Goad, M.B.; Gaur, T.; Stein, G.S.; Lian, J.B.; Komm, B.S. The Wnt Antagonist Secreted Frizzled-Related Protein-1 Is a Negative Regulator of Trabecular Bone Formation in Adult Mice. *Mol. Endocrinol.* **2004**, *18*, 1222–1237. [CrossRef] [PubMed]
- 83. DeLuca, H.F. Overview of general physiologic features and functions of vitamin D. *Am. J. Clin. Nutr.* **2004**, *80*, 1689S–1696S. [CrossRef] [PubMed]
- 84. Goltzman, D. Functions of vitamin D in bone. Histochem. Cell Biol. 2018, 149, 305–312. [CrossRef]
- 85. Lips, P. Vitamin D physiology. Prog. Biophys. Mol. Biol. 2006, 92, 4-8. [CrossRef]
- 86. Jeon, S.-M.; Shin, E.-A. Exploring vitamin D metabolism and function in cancer. *Exp. Mol. Med.* **2018**, *50*, 20. [CrossRef]
- 87. Kiefer, M.C.; Bauer, D.M.; Barr, P.J. The cDNA and derived amino acid sequence for human osteopontin. *Nucleic Acids Res.* **1989**, 17, 3306. [CrossRef]
- 88. Reinholt, F.P.; Hultenby, K.; Oldberg, A.; Heinegard, D. Osteopontin—A possible anchor of osteoclasts to bone. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 4473–4475. [CrossRef]
- Gillan, L.; Matei, D.; Fishman, D.A.; Gerbin, C.S.; Karlan, B.Y.; Chang, D.D. Periostin Secreted by Epithelial Ovarian Carcinoma Is a Ligand for αVβ3 and αVβ5 Integrins and Promotes Cell Motility. *Cancer Res.* 2002, 62, 5358–5364.
- 90. Takeshita, S.; Kikuno, R.; Tezuka, K.; Amann, E. Osteoblast-specific factor 2: Cloning of a putative bone adhesion protein with homology with the insect protein fasciclin I. *Biochem. J.* **1993**, 294, 271–278. [CrossRef]
- 91. Wang, F.-S.; Lin, C.-L.; Chen, Y.-J.; Wang, C.-J.; Yang, K.D.; Huang, Y.-T.; Sun, Y.-C.; Huang, H.-C. Secreted Frizzled-Related Protein 1 Modulates Glucocorticoid Attenuation of Osteogenic Activities and Bone Mass. *Endocrinology* **2005**, *146*, 2415–2423. [CrossRef] [PubMed]
- 92. Larson, S.R.; Zhang, X.; Dumpit, R.; Coleman, I.; Lakely, B.; Roudier, M.; Higano, C.S.; True, L.D.; Lange, P.H.; Montgomery, B.; et al. Characterization of osteoblastic and osteolytic proteins in prostate cancer bone metastases. *The Prostate* 2013, 73, 932–940. [CrossRef] [PubMed]
- Johnson, R.W.; Merkel, A.R.; Page, J.M.; Ruppender, N.S.; Guelcher, S.A.; Sterling, J.A. Wnt signaling induces gene expression of factors associated with bone destruction in lung and breast cancer. *Clin. Exp. Metastasis* 2014, 31, 945–959. [CrossRef] [PubMed]
- 94. Matzelle, M.M.; Gallant, M.A.; Condon, K.W.; Walsh, N.C.; Manning, C.A.; Stein, G.S.; Lian, J.B.; Burr, D.B.; Gravallese, E.M. Resolution of inflammation induces osteoblast function and regulates the Wnt signaling pathway. *Arthritis Rheum.* **2012**, *64*, 1540–1550. [CrossRef] [PubMed]
- 95. Bonfiglio, R.; Scimeca, M.; Toschi, N.; Pistolese, C.A.; Giannini, E.; Antonacci, C.; Ciuffa, S.; Tancredi, V.; Tarantino, U.; Albonici, L.; et al. Radiological, Histological and Chemical Analysis of Breast Microcalcifications: Diagnostic Value and Biological Significance. *J. Mammary Gland Biol. Neoplasia* 2018, 23, 89–99. [CrossRef] [PubMed]
- 96. Cox, R.F.; Morgan, M.P. Microcalcifications in breast cancer: Lessons from physiological mineralization. *Bone* **2013**, *53*, 437–450. [CrossRef] [PubMed]
- 97. Scimeca, M.; Giannini, E.; Antonacci, C.; Pistolese, C.A.; Spagnoli, L.G.; Bonanno, E. Microcalcifications in breast cancer: An active phenomenon mediated by epithelial cells with mesenchymal characteristics. *BMC Cancer* **2014**, *14*, 286. [CrossRef]
- 98. Sharma, T.; Radosevich, J.A.; Pachori, G.; Mandal, C.C. A Molecular View of Pathological Microcalcification in Breast Cancer. *J. Mammary Gland Biol. Neoplasia* **2016**, *21*, 25–40. [CrossRef]
- 99. Morgan, M.P.; Cooke, M.M.; McCarthy, G. Microcalcifications associated with breast cancer: An epiphenomenon or biologically significant feature of selected tumors? *J. Mammary Gland Biol. Neoplasia* 2005, 10, 181–187. [CrossRef]
- 100. He, F.; Springer, N.L.; Whitman, M.A.; Pathi, S.P.; Lee, Y.; Mohanan, S.; Marcott, S.; Chiou, A.E.; Blank, B.S.; Iyengar, N.; et al. Hydroxyapatite mineral enhances malignant potential in a tissue-engineered model of ductal carcinoma in situ (DCIS). *Biomaterials* 2019, 224, 119489. [CrossRef]

- 101. Cox, R.F.; Jenkinson, A.; Pohl, K.; O'Brien, F.J.; Morgan, M.P. Osteomimicry of mammary adenocarcinoma cells in vitro; increased expression of bone matrix proteins and proliferation within a 3D collagen environment. *PLoS ONE* 2012, 7, e41679. [CrossRef] [PubMed]
- 102. Scimeca, M.; Bonfiglio, R.; Menichini, E.; Albonici, L.; Urbano, N.; De Caro, M.T.; Mauriello, A.; Schillaci, O.; Gambacurta, A.; Bonanno, E. Microcalcifications drive breast cancer occurrence and development by macrophage-mediated epithelial to mesenchymal transition. *Int. J. Mol. Sci.* 2019, 20, 5633. [CrossRef] [PubMed]
- 103. O'Grady, S.; Morgan, M.P. Deposition of calcium in an in vitro model of human breast tumour calcification reveals functional role for ALP activity, altered expression of osteogenic genes and dysregulation of the TRPM7 ion channel. *Sci. Rep.* **2019**, *9*, 542.



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