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Effects of nitrogen deposition and litter layer management on soil CO₂, N₂O, and CH₄ emissions in a subtropical pine forestland

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Forestland soils play vital role in regulating global soil greenhouse gas (GHG) budgets, but the interactive effect of the litter layer management and simulated nitrogen (N) deposition on these GHG flux has not been elucidated clearly in subtropical forestland. A field trial was conducted to study these effects by using litter removal method under 0 and 40 kg N ha⁻¹ yr⁻¹ addition in a subtropical forestland in Yingtan, Jiangxi Province, China. Soil CO₂ emission was increased by N addition (18–24%) but decreased by litter removal (24–32%). Litter removal significantly ($P < 0.05$) decreased cumulative N₂O emission by 21% in treatments without N addition but only by 10% in treatments with 40 kg N ha⁻¹ yr⁻¹ addition. Moreover, litter-induced N₂O emission under elevated N deposition (0.094 kg N₂O-N ha⁻¹) was almost the same as without N addition (0.088 kg N₂O-N ha⁻¹). Diffusion of atmospheric CH₄ into soil was facilitated by litter removal, which increased CH₄ uptake by 55%. Given that the increasing trend of atmospheric N deposition in future, which would reduce litterfall in subtropical N-rich forest, the effect of surface litter layer change on soil GHG emissions should be considered in assessing forest GHG budgets and future climate scenario modeling.

Anthropogenic activities have greatly affected greenhouse gas (GHG) emissions from the terrestrial biosphere. During the last decade, atmospheric concentrations of CO₂, N₂O, and CH₄ have increased at rates of 1.9 ppm yr⁻¹, 0.8 ppb yr⁻¹, and 4.8 ppb yr⁻¹, respectively¹. Forestland, which covers 31% of land area and contains 365 Gt of carbon (C) in soils and litter layer², plays a vital role in regulating soil C and N dynamics and global GHG budgets since they mostly act as CO₂ and N₂O sources and CH₄ sinks¹.

Atmospheric nitrogen (N) deposition has increased dramatically since last century, mainly due to anthropogenic activities such as fossil fuel combustion and ammonia volatilization caused by N fertilizer application, and it is considered that this increasing trend will continue in the next few decades³. Increased N availability will significantly influence soil C and N dynamics, thus altering the exchange of GHGs between the biosphere and the atmosphere^{4–6}. Simulated N deposition mostly resulted in decreased CO₂ emission by inhibiting soil autotrophic and/or heterotrophic respiration and the decomposition of soil organic C (SOC)^{5,7}. Nitrate (NO₃⁻) could increase soil redox potential and thus decrease CH₄ production, while NH₄⁺ may inhibit CH₄ oxidation by methanotrophic bacteria to CO₂⁸. Increased soil N availability from N deposition greatly increased soil N₂O emissions. Liu and Greaver⁴ found that N addition (10–562 kg N ha⁻¹ yr⁻¹) significantly increased N₂O emission by 216% on average across different ecosystems by conducting a meta-analysis. A positive linear relationship between N rates and N₂O emission from the subtropical forest soils was primarily due to the promotion of soil denitrification rates caused by increased N availability^{9,10}. However, contrary effects or lacks of response of CO₂, CH₄, or N₂O emission to elevated N deposition have also been reported^{8,11}.

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In forest ecosystems, the litter layer, which contributes the largest C and nutrients input to soils, plays a vital role in regulating soil C and N dynamics and GHG emission. In temperate forests, litter layer decomposition contributes about 5% to 45% of total soil CO₂ emissions^{12,13}. Litter layer removal may decrease soil fungi: bacteria ratio and then affect soil CO₂ emissions, since litter layer decomposition is governed by fungi that can decompose cellulose and lignin¹⁴. Furthermore, well-aerated forest soils are considered as important CH₄ sinks because of the CH₄ consumption by methanotrophic bacteria¹⁵. Litter layer does not emit or uptake CH₄ by itself¹², but may affect soil CH₄ flux by controlling its diffusion between soil and atmosphere¹⁶.

The largest natural source of N₂O is from soils under natural vegetation, which accounted for 6.6 Tg N₂O-N yr⁻¹ of global terrestrial N₂O emissions¹. However, the effect of litter layer management on soil N₂O flux is not clear yet. It has been reported that litter layer removal either significantly reduced soil N₂O emission¹⁷⁻¹⁹ or had no impacts on soil N₂O emission in subtropical and tropical forests²⁰. Firstly, litter could provide organic C and N as substrate for nitrifiers and denitrifiers, but may also simulate microbial growth and activity, thus promoting N₂O production²¹. It has been reported that removals of litter layer reduced soil N₂O emissions by between 6% and 34% in forest ecosystems^{17,18}. Secondly, litter layer may act as a barrier, which could enhance the soil anaerobic environment and then promote soil N₂O production. Eickenscheidt and Brumme²² found that low soil gas diffusivity induced by litter layer, along with high N turnover rate, promoted high N₂O emission from acid beech forest soils, which explained 77% of the variation in N₂O fluxes. Therefore, the effect of litter layer on soil N₂O emission is controlled by the counterbalance between the promotion and inhibition effects mentioned above. However, to our knowledge, the distinct effect of the litter layer on soil GHG fluxes in forestland under elevated N deposition remains unclear. Therefore, precise quantification of the effect of litter layer on GHG flux with different N additions will help to understand how litter layer and N deposition influence soil processes and help to improve the biogeochemical models for GHG budget assessment.

Therefore, the objectives of this study were: (i) to quantify the effect of simulated N deposition, litter removal, and their interaction on soil CO₂, N₂O, and CH₄ emission; and (ii) to understand the key factors regulating soil GHG emissions in subtropical forestland.

Results

Climate and soil environmental variables. Mean air temperatures in the June 2011–May 2012 (18.2 °C) and June 2012–May 2013 (18.2 °C) periods were higher than the long-term MAT (17.8 °C), while daily mean air temperatures ranged from -0.88 °C on 4 January 2013 to 34.10 °C on 6 July 2012 during the 2-yr study period (Fig. 1a). Total precipitation during the 2011–2012 and 2012–2013 periods were 2116 and 2409 mm, respectively (Fig. 1a), which mainly fell during March to September, accounting for between 69% and 81% of the annual amount.

Annual and seasonal dynamics of soil temperature at 5 cm, 10 cm, and 15 cm depths followed daily air temperature (linear relationship, $r = 0.894-0.898$, $P < 0.001$), which was not affected by different treatments ($P > 0.05$, Fig. 1b). Soil WFPS varied from 21.7% to 93.4% with a mean of 59.3% in the March to September period (rainy season), which was significantly higher ($P < 0.001$) than that from October to February (from 26.6% to 81.4% with a mean of 51.5%, Fig. 1c). Soil WFPS dynamic was primarily governed by accumulated precipitation between the two gas measurements intervals for all treatments ($r = 0.53-0.55$, $P < 0.001$). Soil WFPS was significantly influenced by N addition and litter removal ($P < 0.001$) that N addition treatments (NL and NR) showed 7% higher WFPS than without N treatments (CL and CR) and litter retention treatments (CL and NL) showed 5% higher WFPS than litter removal treatments (CR and NR).

Soil NH₄⁺-N concentrations were 16.2 and 17.3 mg N kg⁻¹ on average in the NL and NR treatments, respectively and were notably higher than those in the CL and CR treatments (14.5 and 14.4 mg N kg⁻¹ on average, respectively) (Fig. 2a). Soil NO₃⁻ concentrations in the NL and NR treatments (5.0 and 5.9 mg N kg⁻¹ on average, respectively) were significantly higher than those in the CL and CR treatments (1.8 and 1.9 mg N kg⁻¹ on average, respectively) (Fig. 2b). However, no significant effect of litter removal on soil NH₄⁺ and NO₃⁻ concentration was observed in this study ($P > 0.05$).

Soil GHG fluxes. Similar seasonal and annual soil CO₂ flux dynamics were observed among different treatments (Fig. 3a), which followed the soil temperature dynamic that decreasing from July to February (Fig. 1b; Table 1). The averaged soil CO₂ fluxes were significantly influenced by both N addition and litter removal ($P < 0.001$), where N addition treatments (NL and NR) showed 22% higher mean soil CO₂ fluxes than without N treatments (CL and CR) and litter retention treatments (CL and NL) showed 38% higher mean soil CO₂ fluxes than litter removal treatments (CR and NR). Litter-induced CO₂ flux ranged from 0.70 mg CO₂-C m⁻² h⁻¹ in February to 59.84 mg CO₂-C m⁻² h⁻¹ in July, while no remarkable effect of N level on litter-induced CO₂ fluxes was observed during the study period ($P > 0.05$).

Over the 2-yr measurement period, a sharp increase in soil N₂O fluxes were observed after N addition, while N₂O fluxes were mostly lower than 10.0 μg N₂O-N m⁻² h⁻¹ for the rest of the study period (Fig. 3b). Averaged N₂O fluxes were 5.10 ± 2.78 and 4.00 ± 1.57 μg N₂O-N m⁻² h⁻¹ in CL and CR treatments, respectively, while N addition (40 kg N ha⁻¹ yr⁻¹) significantly increased N₂O fluxes by 2.3–2.7 times. Compared with CL and NL, litter removal (CR and NR) decreased the averaged N₂O flux by 21% and 8%, respectively, although this amount was not statistically significant. While no significant effect of N level on litter-induced N₂O fluxes was obtained in the present study (1.11 and 1.02 μg N₂O-N m⁻² h⁻¹ on average for N0 and N40, respectively), N addition showed much higher variation of litter-induced N₂O fluxes than without N addition (CV of 415% vs 223%).

Soil CH₄ fluxes ranged from -155 to 80 μg CH₄-C m⁻² h⁻¹ over the study period, with 70% of observations showing negative values (Fig. 3c), indicating that the study forestland soil mainly acted as an atmospheric CH₄ sink during the study period. Litter removal significantly increased soil CH₄ uptake by two-fold (i.e., more negative) with average CH₄ uptakes of 25.2–29.5 μg CH₄-C m⁻² h⁻¹ in litter removal treatments (CR and NR) and

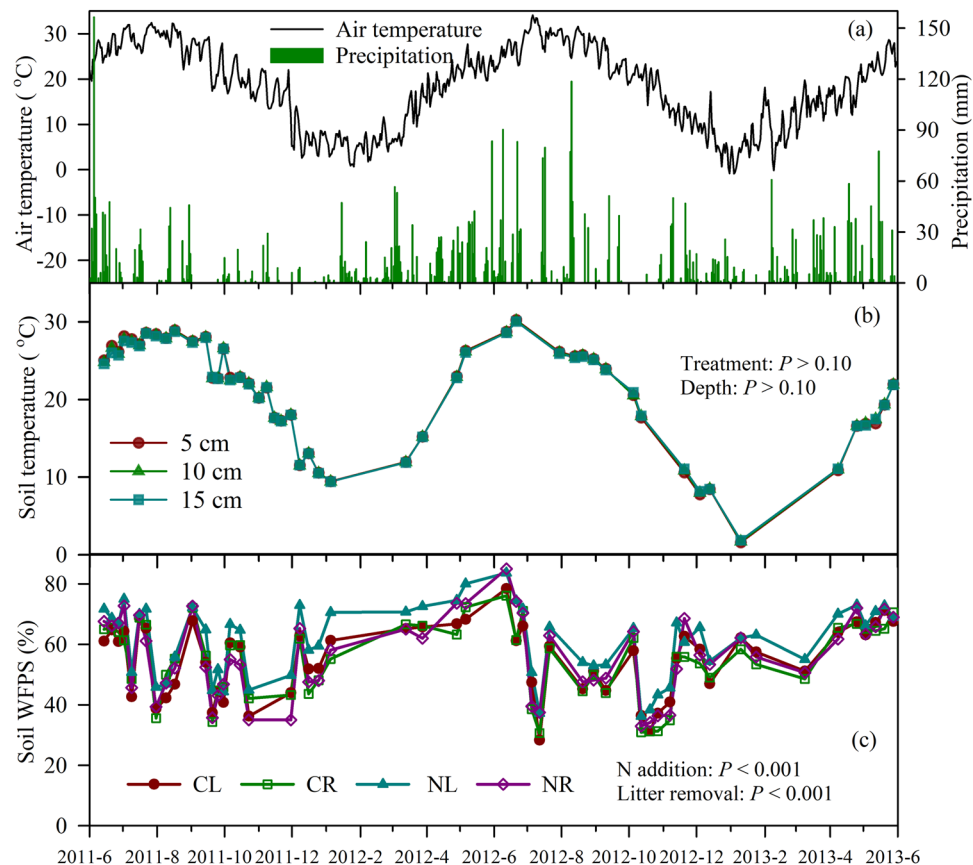


Figure 1. Temporal variations in daily air temperature and precipitation (a), soil temperature at different soil depth (b), and soil water-filled pore space (WFPS) at 5 cm depth (c) for different treatments over a 2-year period from 2011 to 2013. CL, no N addition with litter retention; CR, no N addition with removed litter layer; NL, 40 kg N ha⁻¹ yr⁻¹ addition with litter retention; NR, 40 kg N ha⁻¹ yr⁻¹ addition and removed litter layer.

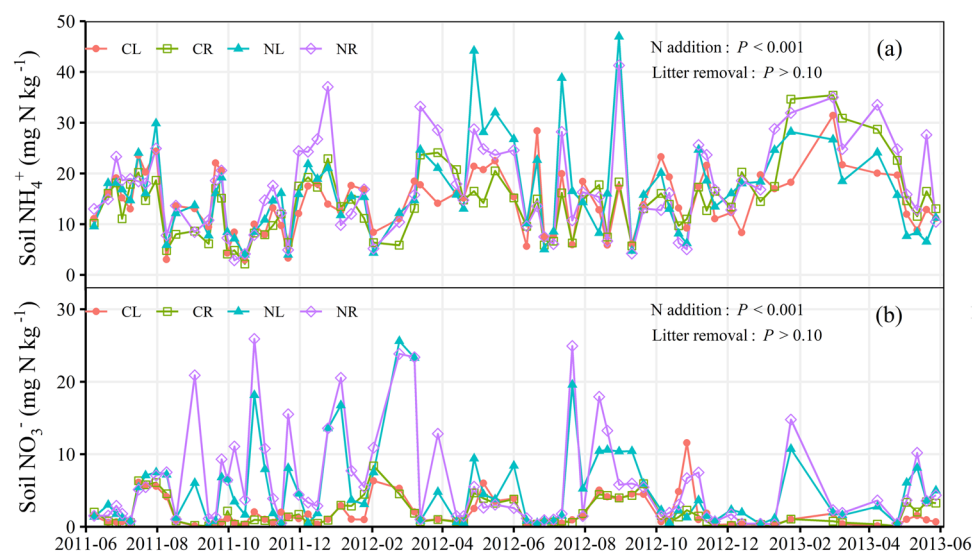


Figure 2. Temporal pattern of soil NH₄⁺ (a) and NO₃⁻ (b) concentrations (0–20 cm) over a two-year period from 2011 to 2013. CL, no N addition with litter retention; CR, no N addition with removed litter layer; NL, 40 kg N ha⁻¹ yr⁻¹ addition with litter retention; NR, 40 kg N ha⁻¹ yr⁻¹ addition and removed litter layer.

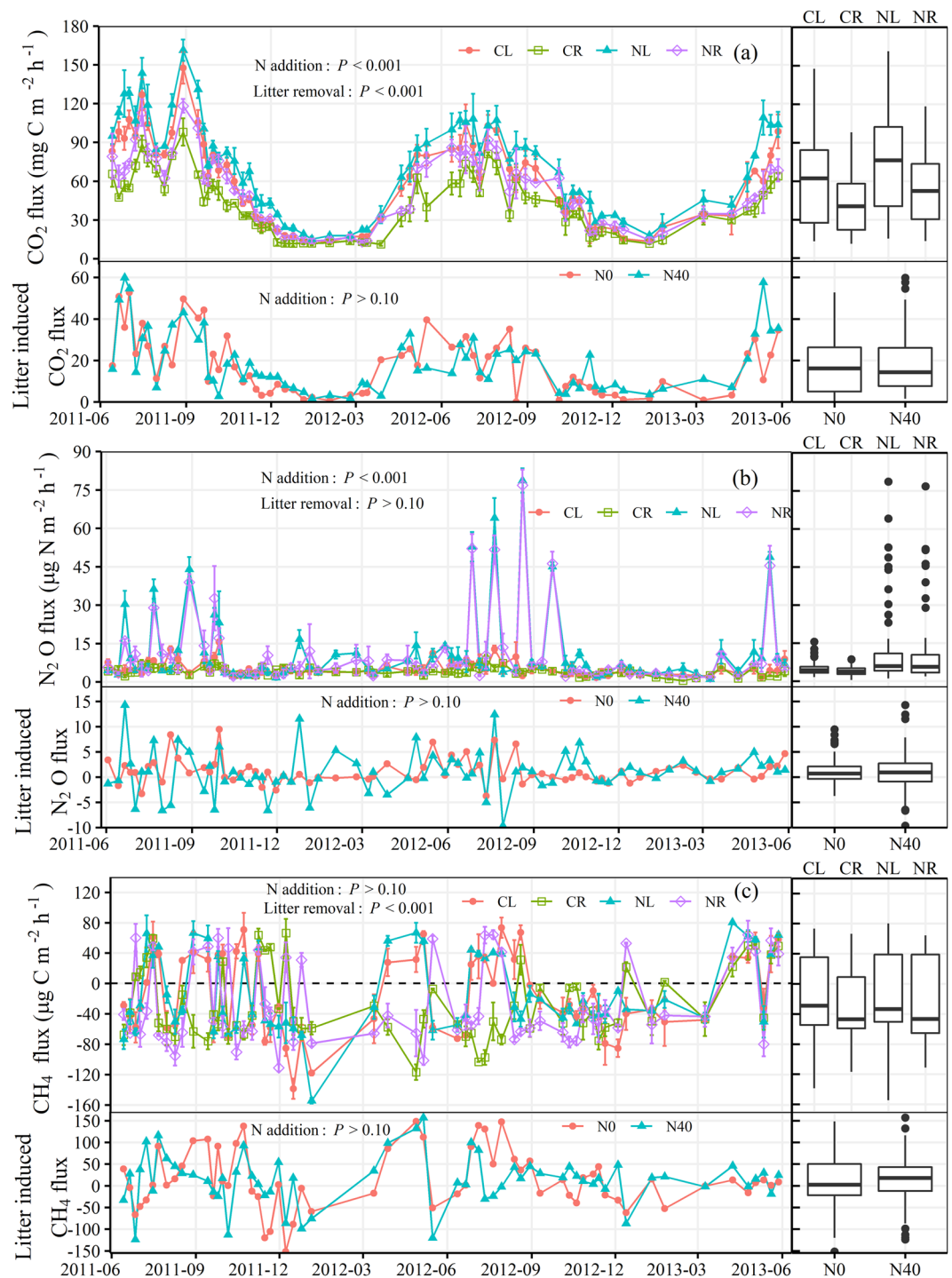


Figure 3. Temporal pattern of soil CO_2 , N_2O , and CH_4 fluxes from four treatments and litter induced CO_2 , N_2O , and CH_4 fluxes ($F_{\text{CL}} - F_{\text{CR}}$ and $F_{\text{NL}} - F_{\text{NR}}$) over a 2-year period from 2011 to 2013. Vertical bars denote the standard error ($n = 3$).

12.7–15.2 $\mu\text{g CH}_4\text{-C m}^{-2} \text{h}^{-1}$ in litter retention treatments (CL and NL). Furthermore, mean litter-induced CH_4 flux was not significantly affected by N addition ($P > 0.10$).

The natural logarithms of CO_2 and N_2O fluxes were significantly ($P < 0.05$) correlated with soil temperature at 5, 10, and 15 cm in all treatments (Table 1). Significantly positive correlations between CH_4 flux and soil temperature were observed in litter retention treatments (CL and NL), but negative correlations (although not significant) were observed in litter removal treatments (CR and NR). In contrast, there was no significant correlation between CO_2 , N_2O , or CH_4 fluxes and soil WFPS in all treatments.

	Treatment	$T_{5\text{cm}}$	$T_{10\text{cm}}$	$T_{15\text{cm}}$	WFPS	$\text{NO}_3^- \text{-N}$	$\text{NH}_4^+ \text{-N}$
ln CO_2	CL	0.884***	0.885***	0.879***	0.072	0.034	-0.006
	CR	0.839***	0.840***	0.839***	0.120	0.084	-0.134
	NL	0.832***	0.846***	0.846***	0.079	-0.198**	-0.069
	NR	0.843***	0.845***	0.844***	0.122	-0.228**	-0.122
ln N_2O	CL	0.288***	0.293***	0.296***	0.064	0.018	-0.013
	CR	0.218**	0.218**	0.219**	-0.108	0.086	-0.150
	NL	0.151*	0.165*	0.164*	0.075	0.186*	-0.148*
	NR	0.266**	0.274***	0.268**	0.115	0.162*	-0.178*
ln CH_4	CL	0.378***	0.384***	0.392***	0.050	0.092	-0.028
	CR	-0.092	-0.094	-0.099	0.105	0.056	0.057
	NL	0.198*	0.202*	0.200*	0.131	-0.074	0.113
	NR	-0.128	-0.127	-0.135	0.151*	0.030	0.114

Table 1. Correlations between the natural logarithm of soil GHG flux (CO_2 , N_2O , and CH_4) and soil parameters for different treatments over the experimental period. CL, no N addition with litter retention; CR, no N addition with removed litter layer; NL, 40 kg N $\text{ha}^{-1} \text{yr}^{-1}$ addition with litter retention; NR, 40 kg N $\text{ha}^{-1} \text{yr}^{-1}$ addition and removed litter layer. $T_{5\text{cm}}$, $T_{10\text{cm}}$, $T_{15\text{cm}}$ are soil temperature at 5, 10, 15 cm depth, respectively; WFPS, soil water-filled pore space WFPS. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

Cumulative GHG fluxes. Annual CO_2 fluxes were 4858 and 4652 kg $\text{CO}_2\text{-C ha}^{-1}$ for CL over 2011–2012 and 2012–2013, respectively, which were 33–48% higher ($P < 0.001$) than that in CR (Fig. 4a; Table 2). The N addition treatments remarkably ($P < 0.001$) increased the cumulative CO_2 flux to 5725–5732 kg $\text{CO}_2\text{-C ha}^{-1}$ for NL and to 4235–4355 kg $\text{CO}_2\text{-C ha}^{-1}$ for NR (18–23% and 24–29%, respectively). However, no yearly effect on annual CO_2 flux was observed in the present study (Table 2). Litter-induced CO_2 emissions were 1356 and 1434 kg $\text{CO}_2\text{-C ha}^{-1}$ in treatments with 0 and 40 kg N $\text{ha}^{-1} \text{yr}^{-1}$ addition, respectively.

Annual cumulative N_2O emission from June 2011 to May 2013 was significantly affected by N application ($P < 0.01$) and litter removal ($P < 0.05$) (Table 2; Fig. 4b). The lowest annual cumulative N_2O emission was observed in the CR treatment, ranging from 0.30 to 0.37 kg $\text{N}_2\text{O-N ha}^{-1}$ (0.34 kg $\text{N}_2\text{O-N ha}^{-1}$ on average), while the highest value was obtained in the NL treatment with an average of 0.94 kg $\text{N}_2\text{O-N ha}^{-1}$ (0.85–1.03 kg $\text{N}_2\text{O-N ha}^{-1}$). Annual cumulative N_2O emissions with N addition were 1.90–2.59 and 2.12–3.03 times higher than those without N addition for treatments with litter retention and treatments with litter layer removal, respectively. Furthermore, litter removal significantly ($P < 0.05$) decreased annual cumulative N_2O emission by 8–25% during the study period (Fig. 4b). Litter-induced N_2O emissions were 0.088 and 0.094 kg $\text{N}_2\text{O-N ha}^{-1}$ in treatments with 0 and 40 kg N $\text{ha}^{-1} \text{yr}^{-1}$ addition, respectively.

The forest soil acted as an atmospheric CH_4 sink from an annual perspective (Fig. 4c). Annual CH_4 uptake in litter retention treatments (CL and NL) ranged from 0.93 to 1.60 kg $\text{CH}_4\text{-C ha}^{-1}$, which was 44–64% lower ($P < 0.001$) than in litter removal treatments (CR and NR) (Table 2; Fig. 4c). However, no significant ($P > 0.05$) influence of N addition on annual CH_4 uptake was observed in the present study. Litter-induced CH_4 emissions were 1.38 and 1.75 kg $\text{CH}_4\text{-C ha}^{-1}$ in treatments with 0 and 40 kg N $\text{ha}^{-1} \text{yr}^{-1}$ addition, respectively.

Total annual GHG flux was significantly affected by both N application and litter removal ($P < 0.001$) (Table 2; Fig. 4d). Total annual GHG flux was significantly higher ($P < 0.001$) for treatments with litter retained (CL and NL, 17.2–21.4 Mg $\text{CO}_2\text{-eq. ha}^{-1} \text{yr}^{-1}$) than litter removal treatments (CR and NR, 12.1–16.3 Mg $\text{CO}_2\text{-eq. ha}^{-1} \text{yr}^{-1}$). N application (NL and NR) significantly ($P < 0.001$) increased annual GHG flux by 19–30%, compared with treatments without N addition (CL and CR). Litter-induced total GHG emissions were 5.06 and 5.36 Mg $\text{CO}_2\text{-eq. ha}^{-1} \text{yr}^{-1}$ in treatments with 0 and 40 kg N $\text{ha}^{-1} \text{yr}^{-1}$ addition, respectively. Furthermore, no interaction effect of N addition and litter removal on annual CO_2 , N_2O , CH_4 , or total GHG flux was obtained in this study (Table 2).

Discussion

Soil respiration rates were significantly increased by N addition (between 18% and 24%) in the studied subtropical forest soil, which is in line with previously results reported based on short-term studies^{23,24}. However, most of other studies reported a notable decrease of soil CO_2 emission after long-term N addition^{5,12,25,26}, mainly due to the decrease of soil microbial diversity and activity⁷, the depletion of labile C²⁷, and/or reduced root biomass²⁸. A 420-day incubation experiment that we conducted using the same forest soil showed that N addition significantly promoted fungal growth²⁹, which may explain the increase in CO_2 emission by N addition in the present study. However, the observed promotion of soil respiration by N addition likely represents the initial phase of the response, which may turn to decline in a long-term study. Conversely, litter removal significantly decreased CO_2 emission by between 24% and 32%, which agrees with the reported decrease of 34% by litter removal in a meta-analysis³⁰. Litter removal may reduce concentrations of dissolved organic carbon (DOC), easily mineralizable substrate for soil microbes, in both the litter layer and the mineral soil by between 22% and 31%³⁰, resulting in a decline in soil CO_2 emission.

Soil temperature was the dominant controlling factor of the seasonal CO_2 dynamics in the present study (Table 1) and many other studies^{12,31,32}. The temperature sensitivity of respiration, Q_{10} , was in the range of 1.85–2.02 (Table 3), which fell in the lower median of the global Q_{10} values of 1.3 to 3.3³³. This result indicated that the potential of temperature increase in the future may not exert a major influence on soil respiration in the subtropical *Pinus massoniana* plantation, which may be due to relative higher annual temperature in this subtropical

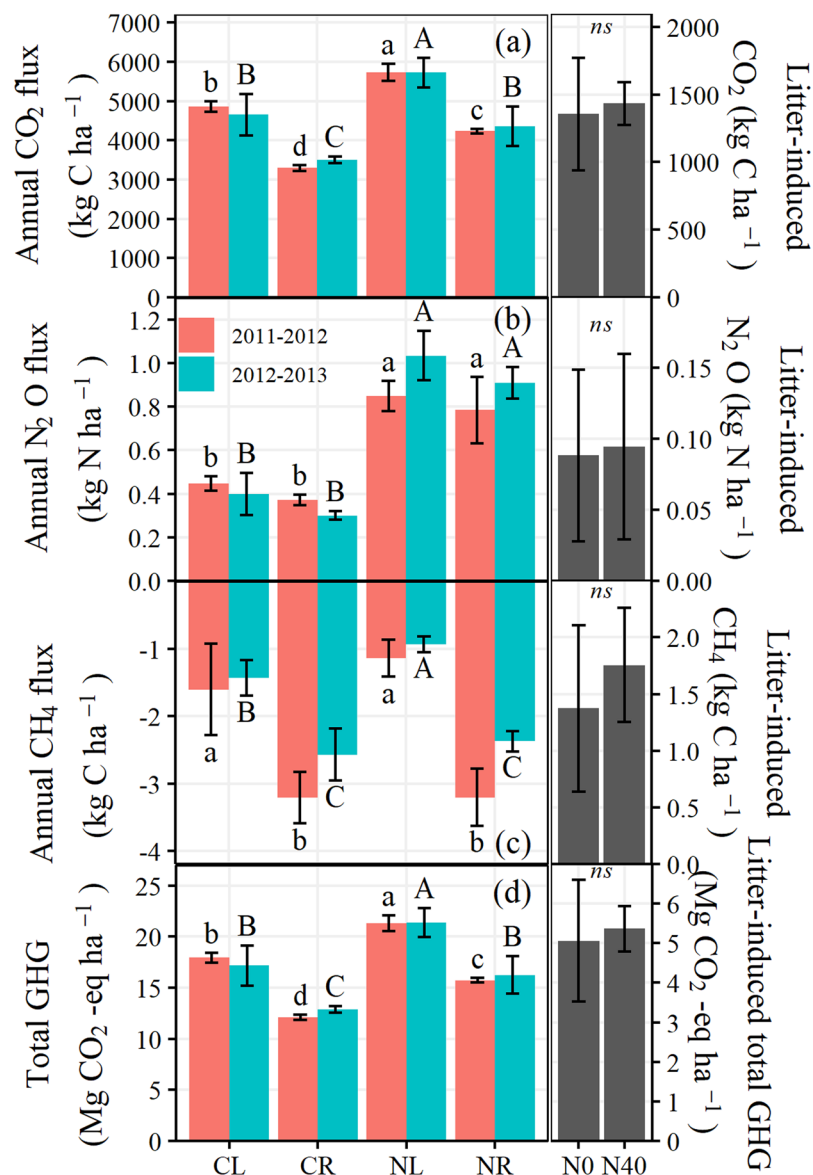


Figure 4. Cumulative CO₂, N₂O, and CH₄ emission under different treatments over a 2-year period from 2011 to 2013. Different lower case letters and capital letters indicate significant differences among treatments at $P < 0.05$ for the 2011–2012 and 2012–2013, respectively.

	CO ₂		N ₂ O		CH ₄		Total GHG	
	F	P	F	P	F	P	F	P
Intercept	1532.97	<0.001	732.21	<0.001	718.95	<0.001	1538.08	<0.001
N	78.52	<0.001	264.95	<0.001	3.67	0.074	87.39	<0.001
L	174.78	<0.001	8.32	0.011	104.06	<0.001	177.76	<0.001
Y	0.08	0.780	2.29	0.151	9.21	0.008	0.14	0.711
N × L	0.14	0.717	0.01	0.925	1.51	0.238	0.15	0.704
N × L × Y	0.12	0.886	5.99	0.012	1.10	0.359	0.16	0.857

Table 2. The effect of N addition (N), litter removal (L), study year (Y), and their interaction on the cumulative CO₂, N₂O, CH₄, and total GHG fluxes.

region (annual mean air temperature of 17.8°C) and the fact that soil respiration is more sensitive to warming in cold regions than in warm regions³⁴.

It has been suggested that subtropical forest soils, with an average N₂O emission rate of 0.9–3.6 Tg yr⁻¹, are an important source for the global N₂O budget⁹, with denitrification being regarded as the main process of

Treatment	Equation	Adj R ²	Q ₁₀
CL	$y = 14.58 \exp(0.067 T_{\text{soil}})$	0.672***	1.98
CR	$y = 9.38 \exp(0.070 T_{\text{soil}})$	0.661***	2.02
NL	$y = 19.57 \exp(0.061 T_{\text{soil}})$	0.616***	1.85
NR	$y = 13.39 \exp(0.064 T_{\text{soil}})$	0.655***	1.90
All	$y = 14.18 \exp(0.065 T_{\text{soil}})$	0.546***	1.91

Table 3. Relationship between CO₂ flux and soil temperature at 5 cm depth (T_{soil}) determined by van't Hoff equations, and temperature sensitivity (Q_{10}) of CO₂ in different treatments over the experimental period. CL, no N addition with litter retention; CR, no N addition with removed litter layer; NL, 40 kg N ha⁻¹ yr⁻¹ addition with litter retention; NR, 40 kg N ha⁻¹ yr⁻¹ addition and removed litter layer. *** $P < 0.001$.

N₂O production³⁵, accounting for between 54% and 76% of total soil N₂O production^{36,37}. Mean annual background N₂O emission in the CL treatment was 0.42 kg N₂O-N ha⁻¹ yr⁻¹ over 2 year in the present study (Fig. 4b), which was close to the values of 0.51 kg N₂O-N ha⁻¹ yr⁻¹ measured in the Notophyll vine forest of southeastern Queensland, Australia¹⁵ and 0.71 kg N₂O-N ha⁻¹ yr⁻¹ measured in the pine plantation of Heshengqiao station in Hubei province, China³⁸, but much lower than the range of 0.93–4.8 kg N₂O-N ha⁻¹ yr⁻¹ reported for other subtropical forest ecosystems^{9,20,37}. This low N₂O emission may be mainly attributed to the low soil N content (0.6 g kg⁻¹) of the test soil compared with other studies mentioned above (0.9–1.9 g kg⁻¹), since it has been demonstrated that annual background N₂O emission was significantly correlated with soil N and mineralized N³⁹. Furthermore, much lower soil C content in the present study (0.52%) would also be responsible for the low N₂O emission, since greater C content can enhance denitrification by stimulating the growth of denitrifying bacteria or by increasing the supply of the electron donor required by this process⁴⁰.

The increased N deposition (40 kg N ha⁻¹ yr⁻¹) significantly increased N₂O emission by 131% and 167% in treatments with and without litter, respectively (Fig. 4b). The present increase rate is lower than the results reported by Wang, *et al.*⁹, who found that 40 kg N ha⁻¹ yr⁻¹ addition (as NaNO₃) increased soil N₂O emission by 269% in a subtropical slash pine plantation in southern China. These results together suggested that soil N₂O emissions from subtropical forestland are highly sensitive to increased nitrate deposition, which may be due to its optimal hydrothermal conditions for denitrification. Averaged soil temperature (24.2 °C) and soil moisture (60.3% WFPS) at 5 cm depth during the rainy season (Fig. 1) were within the range of the optimum denitrification condition⁴¹. NO₃⁻-N input may not only supply more substrates for denitrification but also may decrease the reduction of N₂O to N₂ by suppressing the activity of nitrous oxide reductase⁴², which in turn increased soil N₂O emission.

In the present study, litter removal significantly ($P < 0.05$) decreased soil N₂O emission by 21% in treatments without N addition over 2 years (CR vs CL; Fig. 4b), which was in the range of 15% to 34% measured in subtropical forest^{17,18} but was lower than the range of 37% to 118% measured in temperate forest^{12,43}. The contribution of litter layer to soil N₂O emission could be mainly attributed to the mineralization of litter providing C and N substrates for nitrifiers and/or denitrifiers, thus promoting N₂O production²¹. The lower effect of litter removal on N₂O emission in the present study and in other subtropical forests^{17,18} compared with the temperate forest⁴³ might mainly be due to the difference in litter characteristics between subtropical and temperate forests. The needle litter of subtropical forest, characterized by high polyphenol contents⁴⁴ that would retard decomposition processes, was often less decomposable as that of temperate broad-leaved forests, especially in its early decomposition stage⁴⁵. This finding is in line with Papen and Butterbach-Bahl⁴⁶, who found that beech forest exhibited N₂O emissions 4–5 times higher than that in spruce forest, indicating that forest type was an important modulator of N₂O release from soil⁴⁷. In contrast, litter layer removal only decreased N₂O emission by 10% in treatments with 40 kg N ha⁻¹ yr⁻¹ addition, and litter-induced N₂O emission (0.09 kg N₂O-N ha⁻¹) under elevated N deposition (NR vs NL) was almost the same as that without N addition (CR vs CL; Fig. 4b). Our results suggested that the promotion effect of N addition on N₂O emission might be primarily derived from the enhancement of mineral soil N₂O emission rather than from litter decomposition and corresponding N₂O emission. The mineral soil was considered as the predominant contributor to N₂O emission in forest ecosystems^{17,20}. NO₃⁻-N input in the present study may supply more substrates for soil denitrifiers and promote corresponding N₂O emission. However, the insignificant effect of N addition on litter-induced N₂O emission may be due to the fact that the test acid soil (pH = 4.64) may not be favorable for litter decomposition. Litter layer had been characterized by its low turnover rate expressed by a high mean residence time of 19 years and only 8% of forest litter layer decayed in two years during a ¹⁵N experiment⁴⁸. Therefore, the effect of increased N deposition on litter layer decomposition and corresponding N₂O emission may not be observed in a relatively short study period, such as our 2-yr study. Therefore, with the increase of atmospheric N deposition in subtropical forests, elevated N deposition may promote soil N₂O emission by increasing its emission from mineral soils but not by stimulating litter-induced N₂O emission.

It has been reported that N deposition may increase²⁶, decrease³¹ or have no effect²⁵ on soil CH₄ flux. In our study, CH₄ flux was not significantly affected by N addition but was remarkably influenced by litter removal, which resulted in 55% higher CH₄ uptake in litter removal treatments. However, there are two potential explanations for the significant effect of litter removal on CH₄ uptake. Firstly, the monoterpenes released from decomposition of pine needles⁴⁹ may constrain the methanotrophs, then reducing the CH₄ consumption (40–100%) by soils⁵⁰. Secondly, litter layer may affect soil CH₄ emission or uptake by controlling CH₄ diffusion between soil and atmosphere¹⁶. Therefore, litter layer may act as a physical barrier against CH₄ diffusion into the soil, thus reducing CH₄ uptake in litter retention treatments. However, only net CH₄ fluxes, rather than CH₄ diffusion, were determined by static chamber in the present study, where further study is needed to verify this assumption.

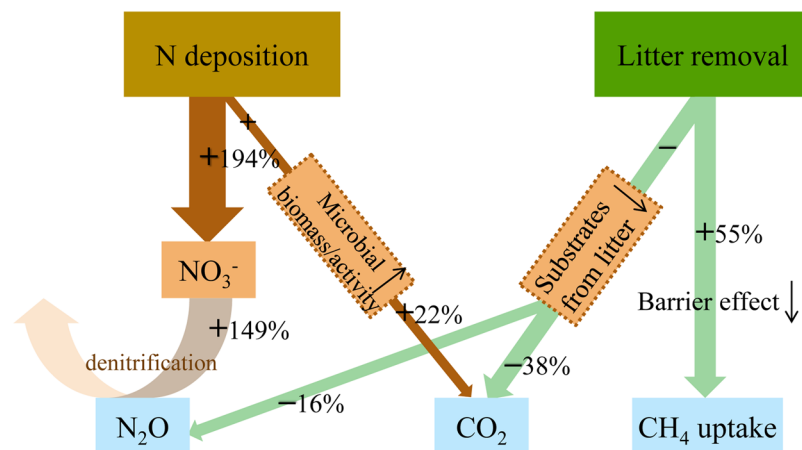


Figure 5. A stylized framework illustrating the main effect of N deposition and litter removal on soil CO_2 , N_2O , and CH_4 fluxes based on the mean values across 2-year period.

Emission of 1 kg of N_2O to the atmosphere is 298 times more effective than 1 kg of CO_2 , while 1 kg of CH_4 is 34 times more effective than 1 kg of CO_2 ¹. Therefore, the GWP of the three GHGs was calculated to identify the effect of N deposition and litter removal on global warming. Our results suggested that CO_2 was the predominant GHG in terms of GWP. In addition, significant effects of N deposition and litter removal on total GHGs were observed (Table 2; Fig. 4d), which was in line with the effect on CO_2 emission.

Increased N deposition has been expected to stimulate C sequestration in forests, where N deposition induced forest C sinks were estimated as 0.24 to 2.0 Pg C yr^{-1} by global biogeochemical models⁵¹. However, Quinn Thomas, *et al.*⁵² found that tree C storage in response to N deposition was dependent on tree species, where N deposition could decrease *Pinus resinosa* growth by 9% per kg N ha^{-1} yr^{-1} but enhanced the growth of 11 tree species as high as 16–18% per kg N ha^{-1} yr^{-1} . By conducting a meta-analysis, Chen, *et al.*⁵³ found that N addition (50 to 150 kg N ha^{-1} yr^{-1}) could decrease soil pH by 6.4%, which could directly damage root growth and inhibit tree growth, thus resulting in a 12.4% reduction of litter fall in N-rich subtropical forest. Our study site is located in south China, where, along with southwest China, has become the third-largest acid rain region in the world since 1990s⁵⁴ and has received quite high level of N deposition since last decade^{55,56}. Hence, a reduction of litter input in response to increased N deposition in subtropical pine forestland could be expected in future. Furthermore, extreme events may occur more and more frequently in future, which could also lead to either dramatic increase in litter fall input after hurricanes or severe storms⁵⁷, or rapid loss of litter layer after wildfires⁵⁸. Therefore, expected decrease of litter input in subtropical conifers forestland would decrease soil CO_2 and N_2O emission but promote CH_4 uptake as showed in the present study. It will be essential to consider the effect of surface litter layer change on soil GHG emissions in assessing forest GHG budgets and future climate scenario modeling.

An illustration summarizing the different effects of N deposition and litter removal on soil CO_2 , N_2O , and CH_4 emissions is presented in Fig. 5. Simulated N deposition promoted soil N_2O emission possibly by increasing denitrification substrates (NO_3^-) and promoted soil respiration by boosting microbial biomass and/or activity. Litter removal decreased the supply of C and N substrates that decomposed from litter layer, thus suppressing soil CO_2 and N_2O emissions. Furthermore, CH_4 uptake was only affected by litter removal since litter layer acts as a barrier against CH_4 diffusion. However, no interaction effect of N addition and litter removal on annual CO_2 , N_2O , CH_4 , or total GHG flux was observed in this study. Our results indicated that N deposition and litter layer influenced soil GHG emissions via different physical or chemical processes, which should be taken into account when quantifying GHG budgets for terrestrial ecosystems.

Methods

Site description and experimental design. A field experiment was conducted at Yingtan Ecological Experimental Station of Red Soil, Chinese Academy of Sciences, Yingtan, Jiangxi Province, Southeastern China (116°55'E, 28°15'N). The area is a hilly red soil region with a typical subtropical monsoon climate, where mean annual precipitation (MAP) is 1785 mm, mean annual air temperature (MAT) is 17.8 °C. The annual accumulative temperature (>10 °C) is 5528 °C with 262 days free of frost. The study site is a 30-year-old pine (*Pinus massoniana*) plantation with an average canopy height of 5 m and a stand density of 2600 stems ha^{-1} . Annual atmospheric wet N deposition is 20 kg N ha^{-1} yr^{-1} according to our field observation⁵⁶. The soil is characterized by an acid loamy clay texture with 36% clay, 43% silt, and 21% sand, and classified as Ferric Acrisols based on the USDA soil taxonomy. Before the experiment, the soil (0–20 cm) had a pH of 4.64 and a CEC of 84.22 mmol kg^{-1} and contained 5.23 g kg^{-1} organic C, 0.63 g kg^{-1} total N, 1.68 mg kg^{-1} NO_3^- -N, and 1.63 mg kg^{-1} NH_4^+ -N.

Two N levels of 0 and 40 kg N ha^{-1} yr^{-1} were established in the forest stand in 2011 to stimulate a future increase in atmospheric N deposition. To investigate the influence of litter layer on soil GHG emission, litter layer was removed using a method involving placing nylon nets (2 mm mesh) 50 cm above the soil surface after removing all detritus from the soil surface. In order to reduce soil disturbance, litter layer was removed carefully more than 1 month before the initiation of flux measurement. Fresh litter collected by nylon nets was removed once or twice per week during the study period. Therefore, four treatments were included in the present study: no N addition with

litter retention (CL); no N addition with removed litter layer (CR); 40 kg N ha⁻¹ yr⁻¹ addition with litter retention (NL); and 40 kg N ha⁻¹ yr⁻¹ addition and removed litter layer (NR). Each treatment was replicated three times. A total of 12 individual plots (3 m × 3 m) were selected on the flat area with a randomized block design with a 3-m-wide buffer strip surrounded each block. N (as NaNO₃) was weighed, mixed with 5 L of distilled water (equivalent to 0.56 mm precipitation), and applied to the NL and NR plots below the canopy using a sprayer. The solution was sprayed equally from March to September (rainy season), beginning in June 2011 and continuing throughout the study period. The same amount of distilled water was sprayed to CL and CR plots simultaneously.

Measurement protocols. Soil CO₂, N₂O, and CH₄ fluxes were determined using the closed-chamber method over a 2-yr period from 3 June 2011 to 28 May 2013 as reported by Fan, *et al.*¹⁰. Samples were taken in the morning between 09:00 and 12:00 once a week during the rainy season (March–September) and biweekly at other times. Concentrations of CO₂, N₂O, and CH₄ in samples were measured with a gas chromatograph (Agilent 7890, Santa Clara, CA, USA) equipped with a thermal conductivity detector (TCD) for CO₂, a ⁶³Ni electron capture detector (ECD) for N₂O, and a flame ionization detector (FID) for CH₄. The standards were purchased from the National Research Center for Certified Reference Materials, Beijing, China. GHG fluxes were calculated using a linear least squares fit to the four sampling points for each plot, where they were omitted if the fitting had $R^2 < 0.90$. Litter-induced CO₂, N₂O, and CH₄ fluxes were calculated as the difference between treatments with litter layer and treatments with removed litter layer (F_{CL} vs. F_{CR} and F_{NL} vs. F_{NR}). Cumulative fluxes were calculated by linear interpolation between measurement days.

Meteorological parameters, including daily air temperature and precipitation, were obtained from a nearby weather station (Milos 520, Vaisala, Finland). On every gas-sampling occasion, soil temperature (at 5, 10, and 15 cm) was determined using a digital thermometer, while soil water content was measured using a time domain reflectometry (TDR) probe at 5 cm depth (except when soil was frozen). Volumetric soil water content was converted to water-filled pore space (WFPS) according to the following equation:

$$WFPS = \text{volumetric water content (\%)} / \text{total soil porosity (cm}^{-3} \text{ cm}^{-3}\text{)}$$

where total soil porosity = 1 – soil bulk density (g cm⁻³)/2.65, with 2.65 g cm⁻³ being the assumed particle density of the soil.

Soil samples (0–20 cm) were collected weekly for the measurement of NH₄⁺ and NO₃⁻ concentrations.

Data calculation and statistical analysis. Fluxes of CO₂, N₂O, or CH₄ were calculated using a linear regression of GHG concentrations to four sampling time for each plot, by considering the chamber air temperature and atmospheric pressure. Cumulative CO₂ (E_{CO_2} , kg CO₂-C ha⁻¹), N₂O (E_{N_2O} , kg N₂O-N ha⁻¹), or CH₄ (E_{CH_4} , kg CH₄-C ha⁻¹) fluxes were calculated according to the following equation:

$$E_{CO_2}(\text{or } E_{N_2O} \text{ or } E_{CH_4}) = \sum_{i=1}^n (F_i + F_{i+1})/2 \times (t_{i+1} - t_i) \times 24$$

where F is the CO₂ flux (mg CO₂-C m⁻² h⁻¹), N₂O flux (μg N₂O-N m⁻² h⁻¹) or CH₄ flux (μg CH₄-C m⁻² h⁻¹), i is the i th measurement, the term $(t_{i+1} - t_i)$ is the days between two adjacent sampling, and n is the total times of sampling.

To evaluate the net global warming impact of CO₂, N₂O and CH₄ together induced by N deposition and litter management, the total GHG were calculated according to Jiang, *et al.*³¹, where they were summed up after converting N₂O and CH₄ fluxes to CO₂ equivalents (kg CO₂-eq. ha⁻¹ yr⁻¹) using global warming potential (GWP) values of 298 and 34 for N₂O and CH₄, respectively, at the 100-yr time horizon¹.

Soil temperature, soil WFPS, and GHG fluxes (CO₂, N₂O, CH₄, and total GHG) data were evaluated using mixed effects model with the *lme* function in the 'nlme' package, where N addition, litter removal, study year, and their interaction were treated as fixed effects, while blocks and/or sampling date were considered as random effect. GHG flux data were natural logarithm transformed as needed, to normalize the distributions prior to statistical analysis. Pearson correlation analysis was used to identify significant correlations between the natural logarithms of the GHG fluxes and the measured environmental variables with the *corr.test* function in the 'psych' package. The van't Hoff equation was established to calculate the temperature sensitivity ($Q_{10} = \exp(10b)$) of CO₂ fluxes to changes in soil temperature with the *nls* function in the 'stats' package. All statistical effects were considered significant at $P < 0.05$. Figures were prepared by 'ggplot2' package. All these analyses were performed with R software⁵⁹.

Data availability

All data generated or analysed during this study are included in this published article.

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References

1. IPCC. Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. (Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA, 1535 pp., 2013).
2. FAO. Global Forests Resources Assessment 2010–Main report. *FAO for Pap* **163**, 44–48 (2010).
3. Liu, X. J. *et al.* Nitrogen deposition and its ecological impact in China: An overview. *Env. Pollut.* **159**, 2251–2264 (2011).
4. Liu, L. & Greaver, T. L. A review of nitrogen enrichment effects on three biogenic GHGs: the CO₂ sink may be largely offset by stimulated N₂O and CH₄ emission. *Ecol. Lett.* **12**, 1103–1117 (2009).

5. Janssens, I. *et al.* Reduction of forest soil respiration in response to nitrogen deposition. *Nat. Geosci.* **3**, 315–322 (2010).
6. Song, X. *et al.* Nitrogen addition increased CO₂ uptake more than non-CO₂ greenhouse gases emissions in a Moso bamboo forest. *Sci. Adv.* **6**, eaaw5790 (2020).
7. Maris, S. C., Teira-Esmatges, M. R., Arbones, A. & Rufat, J. Effect of irrigation, nitrogen application, and a nitrification inhibitor on nitrous oxide, carbon dioxide and methane emissions from an olive (*Olea europaea* L.) orchard. *Sci. Total. Env.* **538**, 966–978 (2015).
8. Bodelier, P. L. E. & Laanbroek, H. J. Nitrogen as a regulatory factor of methane oxidation in soils and sediments. *FEMS Microbiol. Ecol.* **47**, 265–277 (2004).
9. Wang, Y. S. *et al.* Simulated nitrogen deposition reduces CH₄ uptake and increases N₂O emission from a subtropical plantation forest soil in southern China. *PLoS One* **9**, e93571 (2014).
10. Fan, J. *et al.* Sulfur deposition suppressed nitrogen-induced soil N₂O emission from a subtropical forestland in southeastern China. *Agr. For. Meteorol.* **233**, 163–170 (2017).
11. Zhu, X. Effects of warming, grazing/cutting and nitrogen fertilization on greenhouse gas fluxes during growing seasons in an alpine meadow on the Tibetan Plateau. *Agr. For. Meteorol.* **214–215**, 506–514 (2015).
12. Leitner, S., Sae-Tun, O., Kranzinger, L., Zechmeister-Boltenstern, S. & Zimmermann, M. Contribution of litter layer to soil greenhouse gas emissions in a temperate beech forest. *Plant. Soil.* **403**, 455–469 (2016).
13. Vivanco, L. & Austin, A. T. The importance of macro- and micro-nutrients over climate for leaf litter decomposition and nutrient release in Patagonian temperate forests. *For. Ecol. Manag.* **441**, 144–154 (2019).
14. Bonanomi, G. *et al.* Linking bacterial and eukaryotic microbiota to litter chemistry: Combining next generation sequencing with ¹³C CPMAS NMR spectroscopy. *Soil. Biol. Biochem.* **129**, 110–121 (2019).
15. Rowlings, D. W., Grace, P. R., Kiese, R. & Weier, K. L. Environmental factors controlling temporal and spatial variability in the soil-atmosphere exchange of CO₂, CH₄ and N₂O from an Australian subtropical rainforest. *Glob. Change Biol.* **18**, 726–738 (2012).
16. Wang, Y. *et al.* The litter layer acts as a moisture-induced bidirectional buffer for atmospheric methane uptake by soil of a subtropical pine plantation. *Soil. Biol. Biochem.* **66**, 45–50 (2013).
17. Wang, Y. D. *et al.* Effect of litter layer on soil-atmosphere N₂O flux of a subtropical pine plantation in China. *Atmos. Env.* **82**, 106–112 (2014).
18. Liu, H. *et al.* Greenhouse gas fluxes from soils of different land-use types in a hilly area of South China. *Agr. Ecosyst. Env.* **124**, 125–135 (2008).
19. Gao, J. *et al.* Effects of litter inputs on N₂O emissions from a tropical rainforest in Southwest China. *Ecosystems* **21**, 1013–1026 (2018).
20. Tang, X., Liu, S., Zhou, G., Zhang, D. & Zhou, C. Soil-atmospheric exchange of CO₂, CH₄, and N₂O in three subtropical forest ecosystems in southern China. *Glob. Change Biol.* **12**, 546–560 (2006).
21. Ley, M., Lehmann, M. F., Niklaus, P. A. & Luster, J. Alteration of nitrous oxide emissions from floodplain soils by aggregate size, litter accumulation and plant–soil interactions. *Biogeosciences* **15**, 7043–7057 (2018).
22. Eickenscheidt, N. & Brumme, R. Regulation of N₂O and NO_x emission patterns in six acid temperate beech forest soils by soil gas diffusivity, N turnover, and atmospheric NO_x concentrations. *Plant. Soil.* **369**, 515–529 (2013).
23. Contosta, A. R., Frey, S. D. & Cooper, A. B. Seasonal dynamics of soil respiration and N mineralization in chronically warmed and fertilized soils. *Ecosphere* **2**, art36 (2011).
24. Waldrop, M. P., Zak, D. R. & Sinsabaugh, R. L. Microbial community response to nitrogen deposition in northern forest ecosystems. *Soil. Biol. Biochem.* **36**, 1443–1451 (2004).
25. Zhao, Z. *et al.* Effects of warming and nitrogen deposition on CH₄, CO₂ and N₂O emissions in alpine grassland ecosystems of the Qinghai-Tibetan Plateau. *Sci. Total. Env.* **592**, 565–572 (2017).
26. Li, K. *et al.* Responses of CH₄, CO₂ and N₂O fluxes to increasing nitrogen deposition in alpine grassland of the Tianshan Mountains. *Chemosphere* **88**, 140–143 (2012).
27. Frey, S. D., Knorr, M. & Parrent, J. L. & al, e. Chronic nitrogen enrichment affects the structure and function of the soil microbial community in temperate hardwood and pine forests. *For. Ecol. Manag.* **196**, 159–171 (2004).
28. Litton, C. M., Raich, J. W. & Ryan, M. G. Carbon allocation in forest ecosystems. *Glob. Change Biol.* **13**, 2089–2109 (2007).
29. Xu, Y. *et al.* Stage-specific response of litter decomposition to N and S amendments in a subtropical forest soil. *Biol. Fert. Soils* **52**, 711–724 (2016).
30. Xu, S., Liu, L. L. & Sayer, E. J. Variability of above-ground litter inputs alters soil physicochemical and biological processes: a meta-analysis of litterfall-manipulation experiments. *Biogeosciences* **10**, 7423–7433 (2013).
31. Jiang, C., Yu, G., Fang, H., Cao, G. & Li, Y. Short-term effect of increasing nitrogen deposition on CO₂, CH₄ and N₂O fluxes in an alpine meadow on the Qinghai-Tibetan Plateau, China. *Atmos. Env.* **44**, 2920–2926 (2010).
32. Gagnon, B. *et al.* Soil-surface carbon dioxide emission following nitrogen fertilization in corn. *Can. J. Soil. Sci.* **96**, 219–232 (2016).
33. Raich, J. W. & Schlesinger, W. H. The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate. *Tellus B* **44**, 81–99 (1992).
34. Luo, Y., Wan, S., Hui, D. & Wallace, L. L. Acclimatization of soil respiration to warming in a tall grass prairie. *Nature* **413**, 622–625 (2001).
35. Braker, G. & Conrad, R. Diversity, structure, and size of N₂O-producing microbial communities in soils—what matters for their functioning? *Adv. Appl. Microbiol.* **75**, 33–70 (2011).
36. Russow, R., Spott, O. & Stange, C. F. Evaluation of nitrate and ammonium as sources of NO and N₂O emissions from black earth soils (Haplic Chernozem) based on ¹⁵N field experiments. *Soil. Biol. Biochem.* **40**, 380–391 (2008).
37. Zhu, J. *et al.* Spatial and temporal variability of N₂O emissions in a subtropical forest catchment in China. *Biogeosciences* **10**, 1309–1321 (2013).
38. Lin, S. *et al.* Differences in nitrous oxide fluxes from red soil under different land uses in mid-subtropical China. *Agr. Ecosyst. Env.* **146**, 168–178 (2012).
39. Ding, W. *et al.* Effect of long-term compost and inorganic fertilizer application on background N₂O and fertilizer-induced N₂O emissions from an intensively cultivated soil. *Sci. Total. Env.* **465**, 115–124 (2013).
40. Zhang, W. *et al.* Emissions of nitrous oxide from three tropical forests in Southern China in response to simulated nitrogen deposition. *Plant. Soil.* **306**, 221–236 (2008).
41. Xu, Y. B., Xu, Z. H., Cai, Z. C. & Reverchon, F. Review of denitrification in tropical and subtropical soils of terrestrial ecosystems. *J. Soils Sediment.* **13**, 699–710 (2013).
42. Dalal, R. C., Wang, W. J., Robertson, G. P. & Parton, W. J. Nitrous oxide emission from Australian agricultural lands and mitigation options: a review. *Aust. J. Soil. Res.* **41**, 165–195 (2003).
43. Xiao, D., Wang, M., Ji, L., Han, S. & Wang, Y. Variation characteristics of soil N₂O emission flux in broad-leaved Korean pine forest of Changbai Mountain. *Chinese Journal of Ecology* **23**, 46–52 (in Chinese) (2004).
44. Gallet, C. & Lebreton, P. Evolution of phenolic patterns in plants and associated litters and humus of a mountain forest ecosystem. *Soil. Biol. Biochem.* **27**, 157–165 (1995).
45. Gartner, T. B. & Cardon, Z. G. Decomposition dynamics in mixed-species leaf litter. *Oikos* **104**, 230–246 (2004).
46. Papen, H. & Butterbach-Bahl, K. A 3-year continuous record of nitrogen trace gas fluxes from untreated and limed soil of a N-saturated spruce and beech forest ecosystem in Germany: 1. N₂O emissions. *J. Geophys. Res.* **104**, 18487–18503 (1999).

47. Barrena, I. *et al.* Greenhouse gas fluxes (CO₂, N₂O and CH₄) from forest soils in the Basque Country: Comparison of different tree species and growth stages. *For. Ecol. Manag.* **310**, 600–611 (2013).
48. Eickenscheidt, N. & Brumme, R. Contribution of 15N-labelled leaf litter to N turnover, nitrous oxide emissions and N sequestration in a beech forest during eleven years. *Plant. Soil.* **362**, 67–77 (2013).
49. Lerdau, M., Litvak, M., Palmer, P. & Monson, R. Controls over monoterpene emissions from boreal forest conifers. *Tree Physiol.* **17**, 563–569 (1997).
50. Amaral, J. A. & Knowles, R. Inhibition of methane consumption in forest soils by monoterpenes. *J. Chem. Ecol.* **24**, 723–734 (1998).
51. Thornton, P. E., Lamarque, J.-F., Rosenbloom, N. A. & Mahowald, N. M. Influence of carbon-nitrogen cycle coupling on land model response to CO₂ fertilization and climate variability. *Glob. Biogeochem. Cy* **21**, GB4018 (2007).
52. Quinn Thomas, R., Canham, C. D., Weathers, K. C. & Goodale, C. L. Increased tree carbon storage in response to nitrogen deposition in the US. *Nat. Geosci.* **3**, 13–17 (2010).
53. Chen, H. *et al.* Effects of nitrogen deposition on carbon cycle in terrestrial ecosystems of China: A meta-analysis. *Env. Pollut.* **206**, 352–360 (2015).
54. Larssen, T. *et al.* Acid deposition and its effects in China: An overview. *Env. Sci. Policy* **2**, 9–24 (1999).
55. Yu, G. *et al.* Stabilization of atmospheric nitrogen deposition in China over the past decade. *Nat. Geosci.* **12**, 424–429 (2019).
56. Fan, J. L. *et al.* Atmospheric inorganic nitrogen deposition to a typical red soil forestland in southeastern China. *Env. Monit. Assess.* **159**, 241–253 (2009).
57. Ostertag, R., Scatena, F. N. & Silver, W. L. Forest floor decomposition following hurricane litter inputs in several Puerto Rican forests. *Ecosystems* **6**, 261–273 (2003).
58. Wardle, D. A., Hörnberg, G., Zackrisson, O., Kalela-Brundin, M. & Coomes, D. A. Long-term effects of wildfire on ecosystem properties across an island area gradient. *Science* **300**, 972–975 (2003).
59. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL, <https://www.R-project.org/> (2019).

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Author contributions

J.F. designed the experiment and wrote the drafts of the manuscript. R.L. conducted the experiment and analyzed the samples. J.F., R.L., B.G.M. and N.Z. reviewed and edited the drafts. All authors have read and agreed to the published version of the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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