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Diel CO₂ cycles reduce severity of behavioural abnormalities in coral reef fish under ocean acidification

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Elevated CO₂ levels associated with ocean acidification (OA) have been shown to alter behavioural responses in coral reef fishes. However, all studies to date have used stable pCO₂ treatments, not considering the substantial diel pCO₂ variation that occurs in shallow reef habitats. Here, we reared juvenile damselfish, *Acanthochromis polyacanthus*, and clownfish, *Amphiprion percula*, at stable and diel cycling pCO₂ treatments in two experiments. As expected, absolute lateralization of *A. polyacanthus* and response to predator cue of *Am. percula* were negatively affected in fish reared at stable, elevated pCO₂ in both experiments. However, diel pCO₂ fluctuations reduced the negative effects of OA on behaviour. Importantly, in experiment two, behavioural abnormalities that were present in fish reared at stable 750 μatm CO₂ were largely absent in fish reared at 750 ± 300 μatm CO₂. Overall, we show that diel pCO₂ cycles can substantially reduce the severity of behavioural abnormalities caused by elevated CO₂. Thus, past studies may have over-estimated the impacts of OA on the behavioural performance of coral reef fishes. Furthermore, our results suggest that diel pCO₂ cycles will delay the onset of behavioural abnormalities in natural populations.

Increasing atmospheric CO₂ levels are expected to cause a reduction of ocean surface water pH by 0.3–0.4 of a unit by the year 2100, a process commonly referred to as ocean acidification (OA)¹. Ocean acidification projections are based on open ocean environments that are relatively stable over time¹. In contrast, coastal and shallow water habitats can experience substantial natural fluctuations in pCO₂ on a variety of temporal scales^{2,3}. These fluctuations are driven by a range of biological and physical processes⁴ and in some instances their magnitude can exceed mean CO₂ levels projected to occur over the next century^{2,3}. Furthermore, natural pCO₂ fluctuations are expected to increase in size throughout the century, as increased CO₂ uptake by the oceans leads to reduced seawater buffering capacity^{5,6}. Consequently, as mean oceanic pCO₂ levels rise, shallow water marine organisms will be exposed to higher pCO₂ levels for longer periods of time in addition to experiencing a greater range of pCO₂ levels.

Our current understanding of how natural pCO₂ fluctuations will interact with rising mean oceanic pCO₂ levels to affect the performance of shallow water marine organisms under future OA is limited. This is because most OA experiments have used stable pCO₂ levels consistent with open ocean projections, instead of pCO₂ levels naturally relevant to the study organism^{7,8}. While such experiments have demonstrated a range of impacts on traits across various taxa^{9–11}, their ecological relevance is uncertain. Indeed, a handful of studies have shown that natural pCO₂ fluctuations can significantly modify the biological responses of shallow water marine organisms to OA^{12–19}. Consequently, there has been a call for experiments on shallow water marine organisms that include pCO₂ treatments representative of their natural habitats^{7,8,20,21}. Results from such experiments will be vital for improving predictions of when the negative effects caused by elevated pCO₂ will become evident in natural populations²².

Some of the most notable effects of stable, elevated pCO₂ levels have been observed in coral reef fishes. Specifically, exposure to pCO₂ levels between 700–1000 μatm have been shown to impair a range of sensory systems and alter ecologically important behaviours^{10,23,24}. Alterations include impaired anti-predator responses^{25–29}, loss of lateralization^{30,31}, loss of learning^{32,33} and increased activity/boldness²⁷. Such behavioural abnormalities are expected to have significant ecological consequences for fish populations. For example, as a consequence of

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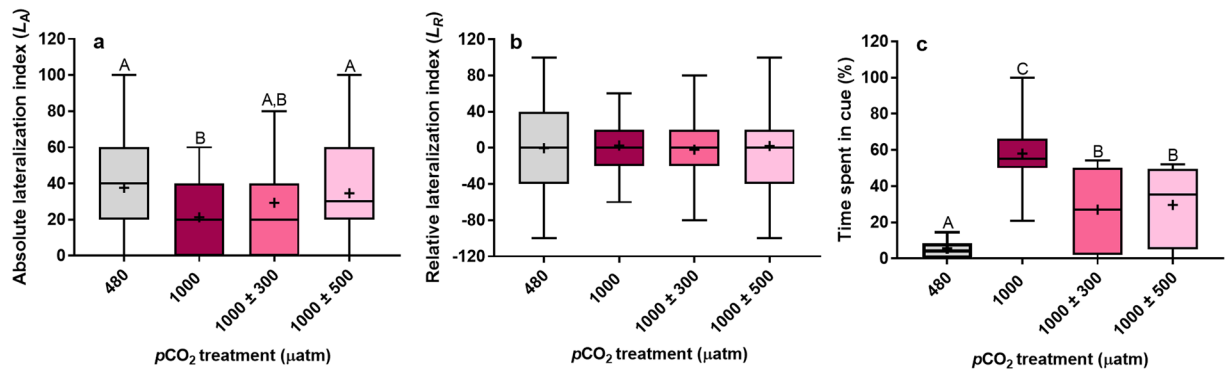


Figure 1. Effects of stable vs diel cycling elevated $p\text{CO}_2$ on behavioural responses in experiment one. (a) Absolute lateralization and (b) relative lateralization in juvenile *Acanthochromis polyacanthus* ($n = 60$ per treatment) were determined using a two-way T-maze. (c) Response to predator cue of juvenile *Amphiprion percula* ($n = 16$ per treatment) was determined using a two-choice flume. Different letters represent significant differences between treatments (Tukey, $P < 0.05$). Boxplots are sized according to the 25th and 75th quartiles, where the line identifies the median and the whiskers indicate the minimum and maximum values. + signs represent means.

exhibiting riskier behaviour, predation-related mortality was significantly higher when settlement stage damselfish were exposed to elevated $p\text{CO}_2$ in the laboratory and released into their native habitat, inferring that recruitment and population sustainability will be threatened by projected future CO_2 levels in the ocean²⁷. Furthermore, the impacts that behavioural abnormalities have on predator-prey dynamics^{26, 34} and competitive interactions³⁵ will likely cause shifts in community structure with unknown consequences for ecosystem functioning.

Coral reefs are highly dynamic shallow water habitats that experience diel cycles in $p\text{CO}_2$. These daily CO_2 cycles are driven by the processes of photosynthesis/respiration and calcification/dissolution over a day-night cycle, but are also influenced by physical controls such as water flow and residence time^{36–38}. In shallow reef areas, diel variation in $p\text{CO}_2$ can range anywhere from ± 50 to 600 μatm around the mean^{38–41}. Although the $p\text{CO}_2$ of coral reef waters is not in perfect equilibrium with the atmosphere over a daily timescale, the carbonate system is still heavily influenced by flushing with offshore waters and thus the mean $p\text{CO}_2$ of reef waters will rise in line with rising atmospheric CO_2 ³⁶. To our knowledge, only three studies (all on calcifying corals) have explicitly considered diel $p\text{CO}_2$ variation when investigating the potential impacts of OA on coral reef organisms. Importantly, they found that the negative impacts of OA on growth and calcification were buffered by the presence of a diel cycling $p\text{CO}_2$ regime^{12, 13, 42}. The behavioural alterations that have been observed in coral reef fishes are likely to be sensitive to the interactive effects of diel $p\text{CO}_2$ cycles and rising mean $p\text{CO}_2$ levels for two reasons. Firstly, previous work has shown that it takes between 24–96 h of exposure to stable elevated $p\text{CO}_2$ levels for behavioural abnormalities to manifest, with shorter onset times at higher $p\text{CO}_2$ levels²⁷. Secondly, the negative effects of elevated $p\text{CO}_2$ on behavioural responses are concentration-dependent^{27, 28, 31}. Consequently, diel $p\text{CO}_2$ cycles could reduce the severity of behavioural abnormalities, or prevent them from manifesting, by providing fish with a recovery period, especially if $p\text{CO}_2$ levels drop below the onset threshold (600–700 μatm). Alternatively, experiencing higher maximum $p\text{CO}_2$ levels daily may lead to more severe behavioural abnormalities.

To determine how diel $p\text{CO}_2$ cycles affect the behavioural responses of coral reef fishes under OA, we reared juvenile damselfish, *Acanthochromis polyacanthus* (Bleeker, 1855), and clownfish, *Amphiprion percula* (Lacepède, 1802), under a series of stable and diel cycling $p\text{CO}_2$ treatments in two different experiments. The aim of the first experiment was to determine if the magnitude of diel $p\text{CO}_2$ cycles affects the behavioural performance of coral reef fishes under OA. The aim of the second experiment was to determine if the presence of diel $p\text{CO}_2$ cycles affects the mean CO_2 level at which behavioural abnormalities occur (i.e. the onset of behavioural abnormalities). Specifically, in experiment one, the behaviour of fish reared at two stable CO_2 levels (480 and 1000 μatm) was compared with the behaviour of fish reared in two cycling CO_2 treatments of different magnitude (1000 \pm 300 and 1000 \pm 500 μatm). Therefore, this experiment enabled us to test if the magnitude of diel $p\text{CO}_2$ fluctuations affected the behaviour of fish under OA. In experiment two, the behaviour of fish reared at three stable CO_2 levels (460, 750 and 1000 μatm) was compared with the behaviour of fish reared in diel cycling CO_2 treatments at two different mean CO_2 levels (750 \pm 300 and 1000 \pm 300 μatm). Therefore, this experiment enabled us to test if the effect of diel $p\text{CO}_2$ cycles was dependent on the mean CO_2 level experienced by the fish. In both experiments, we measured behavioural lateralization in *A. polyacanthus* and the response to a predator cue by *Am. percula*. These traits were chosen for each species as previous studies have demonstrated clear negative impacts of exposure to stable, elevated $p\text{CO}_2$ conditions^{25, 27, 31, 43}. It was predicted that diel $p\text{CO}_2$ fluctuations could reduce the overall severity and delay the onset of behavioural abnormalities under OA conditions.

Results

Experiment one. Absolute lateralization (L_A) was significantly influenced by CO_2 treatment (Fig. 1a, $\chi^2 = 15.75$, $df = 3$, $P = 0.001$). As expected, juveniles reared under stable, elevated $p\text{CO}_2$ were less lateralized compared to those reared at control levels ($P = 0.001$). However, diel $p\text{CO}_2$ cycles significantly increased how lateralized juvenile *A. polyacanthus* were at 1000 μatm . L_A of juveniles reared under small fluctuations (± 300 μatm) was

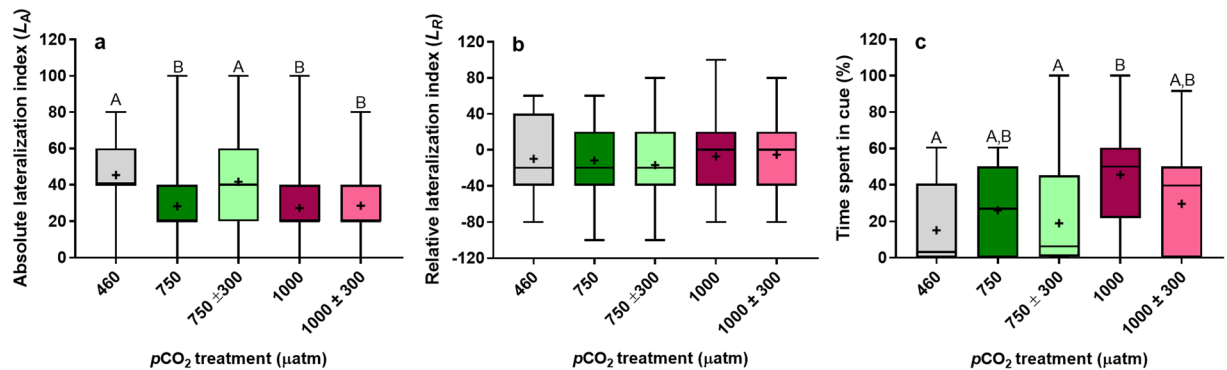


Figure 2. Effects of stable vs diel cycling elevated $p\text{CO}_2$ on behavioural responses in experiment two. (a) Absolute lateralization and (b) Relative lateralization in juvenile *Acanthochromis polyacanthus* ($n = 60$ per treatment) were determined using a two-way T-maze. (c) Response to predator cue of juvenile *Amphiprion percula* ($n = 24$ per treatment) was determined using a two-choice flume. Different letters represent significant differences between treatments (Tukey, $P < 0.05$). Boxplots are sized according to the 25th and 75th quartiles, where the line identifies the median and the whiskers indicate the minimum and maximum values. + signs represent means.

intermediate, but not significantly different, to those reared at control and stable, elevated $p\text{CO}_2$ (min. $P = 0.214$). L_A of juveniles reared under large fluctuations ($\pm 500 \mu\text{atm}$) was fully restored to control levels being significantly greater than those reared at stable, elevated $p\text{CO}_2$ ($P = 0.01$). Mean relative lateralization (L_R) in juvenile *A. polyacanthus* was unaffected by CO_2 treatment (Fig. 1b, $\chi^2 = 0.52$, $df = 3$, $P = 0.914$). Furthermore, no group exhibited a preference for left or right turning (Fig. S1, max. $\chi^2 = 0.84$, $P = 0.358$). Juveniles reared under stable, elevated $p\text{CO}_2$ tended to have a narrower L_R distribution compared to the other treatments (Fig. S3), although these differences were not significant (Max. KS = 0.15, $P = 0.510$).

Mean percentage time that juvenile *Am. percula* spent in predator cue water was significantly affected by CO_2 treatment (Fig. 1c, $\chi^2 = 51.45$, $df = 3$, $P < 0.001$). As expected, juveniles reared at stable, elevated $p\text{CO}_2$ spent a greater amount of time in predator cue water compared to those reared at control levels ($P < 0.001$). However, diel $p\text{CO}_2$ cycles significantly reduced the amount of time that juvenile *Am. percula* spent in predator cue water at 1000 μatm . Juveniles reared under both small ($\pm 300 \mu\text{atm}$) and large ($\pm 500 \mu\text{atm}$) fluctuations demonstrated partial restoration of antipredator behaviour spending an amount of time in predator cue water which was intermediate, and significantly different, to juveniles reared at control and stable, elevated $p\text{CO}_2$ (max. $P < 0.001$).

Experiment two. As was observed in experiment one, mean L_A was significantly affected by CO_2 treatment (Fig. 2a, $\chi^2 = 75.25$, $df = 4$, $P < 0.001$), with juveniles reared under stable, elevated $p\text{CO}_2$ (750 and 1000 μatm) being less lateralized compared to those reared at control levels (max. $P < 0.001$). Diel $p\text{CO}_2$ cycles did not affect how lateralized juvenile *A. polyacanthus* were at mean $p\text{CO}_2$ level of 1000 μatm ($P = 0.986$). In contrast, diel $p\text{CO}_2$ cycles fully restored lateralization in juveniles reared at a mean CO_2 of 750 μatm , being similar to those reared at control levels ($P = 0.710$) and significantly greater than both the stable, elevated CO_2 treatments (max. $P < 0.001$). Also, as observed in experiment one, mean L_R in juvenile *A. polyacanthus* was unaffected by CO_2 treatment (Fig. 2b, $\chi^2 = 4.86$, $df = 4$, $P = 0.302$), and no group exhibited a preference for left or right turning (Fig. S2, max. $\chi^2 = 3.43$, $P = 0.064$). However, there were more individuals which were less lateralized in the 750, 1000 and 1000 $\pm 300 \mu\text{atm}$ CO_2 treatments (Fig. S2).

Similar to experiment one, CO_2 treatment significantly affected the mean percentage time that juvenile *Am. percula* spent in predator cue water (Fig. 2c, $\chi^2 = 15.95$, $df = 4$, $P = 0.003$). As expected, juveniles reared at 1000 μatm CO_2 spent a greater amount of time in predator cue water compared to those reared at control levels ($P = 0.004$). The percentage time juveniles reared at 750 μatm CO_2 spent in predator cue water was intermediate, but not significantly different, to those reared at control and 1000 μatm $p\text{CO}_2$ (min. $P = 0.194$). Diel $p\text{CO}_2$ cycles influenced the predator cue response of juvenile *Am. percula* at mean $p\text{CO}_2$ levels of 750 and 1000 μatm . Juveniles reared at 750 $\pm 300 \mu\text{atm}$ CO_2 spent a percentage of time in predator cue water which was more similar to those reared at control levels compared to those reared at 750 μatm CO_2 . Finally, juveniles reared at 1000 $\pm 300 \mu\text{atm}$ CO_2 demonstrated partial restoration of antipredator behaviour, with juveniles spending a percentage of time in predator cue water which was intermediate to those reared at 460 and 1000 μatm CO_2 (min. $P = 0.309$).

Discussion

This study demonstrates for the first time that diel $p\text{CO}_2$ cycles can significantly modify the behavioural responses of fishes under OA. The negative impacts of elevated CO_2 on coral reef fish behaviour have been well documented and are expected to have significant ecological consequences for reef fish populations through effects on recruitment, predator-prey interactions, competition and habitat preference^{10,23,24}. However, all studies to date have exposed fish to stable levels of elevated CO_2 , not considering the natural diel $p\text{CO}_2$ cycles that occur on coral reefs. Here we show that the severity of two behavioural abnormalities commonly observed under elevated CO_2 are reduced when fish experience a diel cycling $p\text{CO}_2$ regime. The extent of reduction was influenced by both the magnitude of fluctuation and mean $p\text{CO}_2$ level experienced, as well as the behavioural trait. Overall, our results

indicate that previous studies have probably over-estimated the behavioural impacts of OA on coral reef fishes once they have settled to reef habitats where diel CO₂ cycles are prevalent.

Previous research using stable pCO₂ treatments has found that behavioural abnormalities start to manifest in coral reef fish between 600–700 μatm. Our results indicate that diel pCO₂ cycles will delay the onset of behavioural abnormalities. In experiment two, we show that behavioural abnormalities present in fish reared at a stable level of 750 μatm CO₂ were absent in fish reared at 750 ± 300 μatm CO₂. However, in both experiments, although less severe, behavioural abnormalities were still present in the fluctuating 1000 μatm CO₂ treatments. Thus, it appears that mean oceanic pCO₂ levels closer to 1000 μatm will need to be reached before behavioural abnormalities could manifest in natural populations of reef fishes. Furthermore, we observed full restoration of behavioural lateralization in juvenile *A. polyacanthus* reared under 1000 ± 500 μatm CO₂, inferring that some behavioural abnormalities may not manifest at all for populations living in habitats with large CO₂ fluctuations, such as shallow reef flats and closed lagoons, even when average oceanic conditions reach 1000 μatm CO₂. The observation that diel pCO₂ variation can reduce and/or delay the onset of behavioural abnormalities in juvenile coral reef fish under OA is particularly important given the ecological consequences of behavioural abnormalities and past research that has shown a limited capacity for acclimation of behavioural traits to stable, elevated pCO₂^{31,44}. However, it is important to mention that behavioural abnormalities are still likely to occur in the pelagic larval phase of coral reef fish as they occupy a more stable CO₂ environment in the open ocean. Consequently, population replenishment and sustainability of reef fish populations could still be threatened by near-future OA due to impaired behaviour in the larval phase^{27,45,46}, even if behavioural effects are less severe in juveniles that have already settled to reef habitats. Finally, in experiment one, and to a lesser extent in experiment two, we observed more individual variation in predator cue responses of *Am. percula* at 1000 μatm CO₂ if fish were reared under cycling conditions. This level of individual variation has previously been observed only at a mean stable CO₂ of 700 μatm²⁷. Thus, in addition to potentially providing more time for reef fish populations to adapt to future OA conditions, by delaying the onset of behavioural abnormalities, diel pCO₂ cycles may also increase the adaptive potential of fish populations at higher CO₂ levels by increasing the range of individual variation upon which selection can act.

The underlying mechanism of behavioural abnormalities in fish under OA conditions is linked to the effects of acid-base regulation on the function of type A γ-aminobutyric acid (GABA_A) neurotransmitter receptors^{23,43}. GABA_A receptors are gated ion channels with specific conductance for HCO₃⁻ and Cl⁻. Under elevated pCO₂ fish increase intracellular and extracellular HCO₃⁻ concentrations to prevent plasma and tissue acidosis^{47–49}. In a recent study on *A. polyacanthus* this compensatory mechanism was shown to be sufficient to reverse the transmembrane gradients of HCO₃⁻ in brain tissue, which could interfere with GABA_A receptor function and cause behavioural alterations⁵⁰. For coral reef fish it appears that complete acid-base regulation in the brain under stable, elevated pCO₂ levels may take between 24–96 h, as this is the exposure period required before behavioural abnormalities manifest²⁷. Our results suggest that for fish reared under diel pCO₂ cycles, exposure to lower CO₂ levels for several hours each day is sufficient to prevent the physiological changes that would normally occur at a stable, high CO₂. Extracellular and intracellular pH regulation take place at different rates, occurring more quickly in the former. For example, in gulf toadfish (*Opsanus beta*) exposed to 1900 μatm CO₂, complete blood pH compensation was achieved after 2 h, whereas muscle intracellular pH was not adjusted until after 24 h⁴⁹, see also⁵¹. Thus, based on the onset times under stable, elevated pCO₂ (24–96 h), it would appear that behavioural abnormalities do not manifest in coral reef fish until brain intracellular pH compensation is complete, although further testing is required. This could explain why diel pCO₂ cycles alleviated the negative impacts of OA. Although no data is available, we assume coral reef fish would achieve pH compensation as fast, or faster, than toadfish in the example above, due to their higher metabolic rates and more active lifestyle. Therefore, we hypothesise that fish reared under diel pCO₂ cycles were able to track disturbances in extracellular pH, but weren't exposed to higher pCO₂ levels long enough for full brain intracellular pH compensation to occur.

It has been suggested that behavioural abnormalities may also be influenced by alterations in gene expression related to ion regulation^{32,52}. Ion-regulation in blood and tissues is under circadian control in fishes^{53,54}. In a recent study on *A. polyacanthus*, variation in behavioural tolerance to stable, elevated pCO₂ (754 μatm) was linked to the differential expression of genes related to circadian rhythm control⁵⁵. For example, offspring of CO₂ sensitive parents (i.e., those that exhibited behavioural abnormalities) upregulated the enzyme that catalyses the final reaction in the synthesis of melatonin, a key regulator of the circadian rhythm, which plays an important role in controlling ion-regulation⁵⁶. This indicates that CO₂ sensitive individuals might display more pronounced acid-base compensation if exposed to a sustained elevation of CO₂ due to a stronger influence of circadian rhythm control, leading to larger changes of the neuronal ion gradients that determine GABA_A receptor function. Our observations that diel pCO₂ cycles can alleviate the negative behavioural effects of OA suggests that fish were displaying normal, or less, circadian control over acid-base regulation and thus did not respond so strongly to internal pH changes caused by elevated CO₂ therefore avoiding altered brain ion gradients. Consequently, it appears that internal circadian rhythm control of acid-base regulation in coral reef fish is disrupted under stable, elevated pCO₂, indicating that this process may be linked to the natural diel pCO₂ cycles occurring in shallow reef habitats.

In this study we repeated the control, stable 1000 μatm CO₂ and 1000 ± 300 μatm CO₂ treatments in two different experiments. Although we observed similar responses to predator cue in *Am. percula* in both experiments, there were some differences in the effects of CO₂ cycles on behavioural lateralization. In experiment one behavioural lateralization was partially restored in juvenile *A. polyacanthus* reared at 1000 ± 300 μatm CO₂, whereas no restoration was observed in experiment two. The reason for the different results between experiments is unclear, but one possible reason is differences in the duration that the high CO₂ peaks lasted. In experiment one the high peaks lasted approximately three hours, whereas in experiment two they lasted close to eight hours. Consequently, fish in experiment one had less time to adjust their acid-base status during the high peak, which may have resulted in them exhibiting less severe behavioural impairments. The reason we did not observe a

similar difference between experiments for response to predator cue in juvenile *Am. percula* may be because the effect of elevated $p\text{CO}_2$ on this trait was concentration dependent, as seen in experiment two. Due to logistical constraints, it was not possible to have duplicated experimental systems in experiment one. In contrast, experiment two had duplicate systems for each $p\text{CO}_2$ treatment. As similar results were observed in each experiment we are confident that the pseudo-replication in experiment one did not affect the results. In general, the effects of stable, elevated $p\text{CO}_2$ on lateralization and response to a predator cue observed in this study are consistent with previous work on the same species^{25, 27, 29, 31}, with one exception. Previous studies have reported a clear attraction of *Am. percula* to a predator cue (>80% of time in predator cue water) at 1000 μatm CO_2 , whereas *Am. percula* in the current experiments exhibited neither attraction of avoidance of the predator cue (45–58% of time in predator cue water) at this CO_2 level. The same observation was also reported in adult goldskinny wrasse, *Ctenolabrus rupestris*⁵⁷. Why fish in the current experiments exhibited a less dramatic change in antipredator behaviour at high CO_2 compared with previous experiments is unknown, but could be related to some differences in protocol. In contrast to past studies that reset each fish to the starting position when the direction of the water sources was switched, fish were not disturbed during trials in this study. Another potential factor is the life-stage that was tested. Previous studies tested settlement stage larvae, whereas settled juveniles were used in this study. Age-specific responses to predator cues, as well as expression of odourant receptor genes, have been observed in other species of fish^{58, 59}.

In this study, we show that a diel $p\text{CO}_2$ cycle can substantially reduce the severity of behavioural abnormalities caused by elevated CO_2 in coral reef fishes. In contrast, behavioural impairments were still present in a temperate shark species reared under elevated CO_2 in a mesocosm that experienced diel CO_2 variation, although there was no stable, elevated CO_2 treatment to compare against⁶⁰. A handful of other studies have also shown that daily $p\text{CO}_2$ fluctuations can significantly modify the biological responses of shallow water marine organisms to OA^{12–15, 17, 19}. This highlights the importance of considering natural $p\text{CO}_2$ variability when trying to determine the response of shallow water marine organisms to OA. While our understanding of the magnitude and frequency of $p\text{CO}_2$ fluctuations *in situ* is growing, many shallow water habitats remain under- or un-sampled⁸. Consequently, there is a need for more high resolution *in situ* studies that characterise natural CO_2 variability both spatially and temporally. Such data will establish ecologically relevant $p\text{CO}_2$ treatments to be used in laboratory experiments and allow us to better interpret results from past OA studies that have employed stable $p\text{CO}_2$ levels^{7, 61}. This will be critical for accurately assessing the likely effects of OA on shallow water marine organisms and which species and ecosystems may be at greatest risk.

Materials and Methods

Study species. *Acanthochromis polyacanthus* and *Amphiprion percula* are common throughout the Indo-Pacific region. Both species are demersal spawners, laying their eggs within small caves and crevices in the reef matrix. In *A. polyacanthus*, eggs hatch into small juveniles, with both parents providing care to the eggs and offspring for up to 45 d post-hatching⁶². In contrast, *Am. percula* has a relatively short larval phase of approximately 11 d before settling on the reef⁶³. Both species can be bred and reared in captivity with high success, which has led to their establishment as models for investigating the potential impacts of OA on coral reef fishes^{25, 27, 31, 50, 55}.

Brood-stock and general rearing protocol. Adult *A. polyacanthus* were collected using hand nets from the Bramble Reef area (site 1: 18°22'S, 146°40'E; site 2: 18°25'S, 146°40'E) of the Great Barrier Reef in July 2015. Fish were transported to an environmentally controlled aquarium research facility at James Cook University (JCU) (Townsville, Australia) where they were housed as breeding pairs in 60 L aquaria at temperature conditions matching the collection location. An existing brood-stock of *Am. percula* at JCU was used. These pairs had been collected from the Cairns Region of the Great Barrier Reef and housed at JCU for four years. Adult *A. polyacanthus* and *Am. percula* pairs were maintained under stable, ambient $p\text{CO}_2$ (~490 μatm). Temperatures were increased at a rate of 0.5 °C *per* week until the summer breeding temperature of 29 °C was reached in the first week of November 2015. Adult pairs were provided with half a terracotta pot to act as a shelter and spawning site. Aquaria were checked each morning for the presence of newly laid clutches. Pairs were fed *ad libitum* on commercial fish feed pellets (INVE Aquaculture Nutrition NRD 12/20) once daily outside the breeding season and twice daily during the breeding season (November–May).

Acanthochromis polyacanthus juveniles were fed a combination of freshly hatched *Artemia* naupli and weaning fish feed (INVE Aquaculture Nutrition Wean-S 200–400 μm) daily for the first four days post hatch (dph). 5–21 dph they were fed daily on the weaning feed and then switched to a small pellet fish feed (INVE Aquaculture Nutrition NRD 5/8) at 22 dph. Rearing of larval *Am. percula* was performed using methods described by Munday *et al.*²⁸. Settled juveniles were fed daily on the weaning fish feed.

Experimental design. Experiment one was carried out at the aquarium research facility at JCU. For details on the experimental system refer to Supporting Information. Fish were reared at two stable (480 and 1000 μatm) and two cycling (1000 \pm 300 and 1000 \pm 500 μatm) CO_2 treatments (Table 1 and Figure S3). The stable 1000 μatm $p\text{CO}_2$ treatment represented the open ocean projection for the end of this century, typically used in many OA experiments¹¹. The cycling $p\text{CO}_2$ treatments matched levels that have been observed in some tidal lagoons³⁸. Diel $p\text{CO}_2$ fluctuations of between \pm 50–150 μatm are more typical in other reef areas^{40, 41}, however, the magnitude of fluctuations seen in tidal lagoons today may become more common in other reef areas by the year 2100, as a amplification in diel $p\text{CO}_2$ fluctuations is predicted to occur over this time period⁵. Mean values for seawater parameters in experiment one are presented in Table 1.

Experiment two was carried at the National Sea Simulator (SeaSim) facility at the Australian Institute of Marine Science (AIMS) (Cape Cleveland, Australia). Fish were reared at three stable (460, 750 and 1000 μatm) and two cycling (750 \pm 300 and 1000 \pm 300 μatm) CO_2 treatments (Table 2 and Figure S4). For details on the

Parameter	$p\text{CO}_2$ treatment (μatm)			
	480	1000	1000 \pm 300	1000 \pm 500
Average pH_T	8.01 \pm 0.01	7.75 \pm 0.02	7.77 \pm 0.08	7.80 \pm 0.16
Min. pH_T	—	—	7.64 \pm 0.03	7.57 \pm 0.02
Max. pH_T	—	—	7.89 \pm 0.01	8.01 \pm 0.01
pH_T range	—	—	0.24 \pm 0.03	0.44 \pm 0.02
Average $p\text{CO}_2$ (μatm)	480 \pm 20	990 \pm 46	961 \pm 195	934 \pm 389
Min. $p\text{CO}_2$ (μatm)	—	—	681 \pm 21	482 \pm 19
Max $p\text{CO}_2$ (μatm)	—	—	1304 \pm 102	1591 \pm 98
$p\text{CO}_2$ range (μatm)	—	—	623 \pm 96	1109 \pm 95
TA ($\mu\text{mol kg}^{-1}$)	2570 \pm 54	2574 \pm 42	2582 \pm 43	2583 \pm 43
Temperature ($^\circ\text{C}$)	28.7 \pm 0.3	28.9 \pm 0.3	28.9 \pm 0.3	29.0 \pm 0.2
Salinity	36.7 \pm 0.4	36.7 \pm 0.4	36.7 \pm 0.4	36.7 \pm 0.4

Table 1. Seawater parameters for experiment one. Values are means \pm 1 SD for daily average, minimum, maximum and range of pH_T and $p\text{CO}_2$. Mean \pm 1 SD for total alkalinity (TA), temperature ($^\circ\text{C}$) and salinity over the experiment are also shown.

Parameter	$p\text{CO}_2$ treatment (μatm)				
	460	750	750 \pm 300	1000	1000 \pm 300
Average $p\text{CO}_2$ (μatm)	458 \pm 7	748 \pm 9	788 \pm 203	994 \pm 23	1042 \pm 256
Min. $p\text{CO}_2$ (μatm)	443 \pm 26	721 \pm 29	527 \pm 27	926 \pm 121	667 \pm 33
Max $p\text{CO}_2$ (μatm)	477 \pm 42	773 \pm 28	1025 \pm 86	1060 \pm 128	1328 \pm 78
$p\text{CO}_2$ range (μatm)	49 \pm 101	59 \pm 112	498 \pm 90	130 \pm 216	661 \pm 94
TA ($\mu\text{mol kg}^{-1}$)	2322 \pm 20	2325 \pm 23	2326 \pm 21	2330 \pm 22	2330 \pm 23
Temperature ($^\circ\text{C}$)	28.5 \pm 0.1	28.5 \pm 0.1	28.5 \pm 0.1	28.5 \pm 0.1	28.5 \pm 0.1
Salinity	35.4 \pm 0.4	35.4 \pm 0.4	35.4 \pm 0.4	35.4 \pm 0.4	35.4 \pm 0.4

Table 2. Seawater parameters for experiment two. Values are means \pm 1 SD for daily average, minimum, maximum and range of $p\text{CO}_2$. Mean \pm 1 SD for total alkalinity (TA), temperature ($^\circ\text{C}$) and salinity over the experiment are also shown.

experimental system refer to Supplementary Information. Previous experiments indicate that behavioural abnormalities are first evident at around 700 μatm CO_2 , although the magnitude of effect is often not as large as observed at higher CO_2 levels^{27, 28, 31}. Therefore, the inclusion of the 750 and 750 \pm 300 μatm CO_2 treatments enabled us to determine how diel $p\text{CO}_2$ cycles may affect the onset threshold of behavioural abnormalities. Mean values for seawater parameters in experiment two are presented in Table 2.

A similar protocol was employed in both experiments. Three offspring clutches were used per species, each from a different parental pair. In experiment one, *A. polyacanthus* and *Am. percula* clutches were transferred to the experimental system and split between $p\text{CO}_2$ treatments in duplicate tanks (12–15 *A. polyacanthus* per tank and 10 *Am. percula* per tank) at 1 and 12 dph respectively. In experiment two, offspring clutches were transferred to the experimental system and split between $p\text{CO}_2$ treatments in duplicate tanks (one tank per line; 15 *A. polyacanthus* per tank and 13–15 *Am. percula* per tank) at 14 and 12 dph respectively. *A. polyacanthus* clutches were transferred at 14 dph in experiment two, compared with 1 dph in experiment one, due to logistical reasons.

Behavioural lateralization trials on *A. polyacanthus* were performed 40–42 dph, which equated to approximately six and four weeks of exposure to $p\text{CO}_2$ treatments in experiments one and two respectively. Predator cue trials on *Am. percula* were performed 18–20 dph, which equated to approximately 1 week of exposure to $p\text{CO}_2$ treatments in both experiments. All behavioural trials were performed between 09:00 and 17:00. Fish were gently transferred to the behavioural arenas using a glass beaker to minimise handling stress. Fish from each $p\text{CO}_2$ treatment were tested at random times throughout the day to account for any possible time of day effects in the fluctuating treatments. Each fish was tested once, being placed in an isolation chamber within their experimental tank after a trial for the rest of the day. Research was carried out under approval of the James Cook University animal ethics committee (permit: A2210) and according to the University's animal ethics guidelines.

Behavioural assays. *Behavioural lateralization trials.* Behavioural lateralization (i.e., favoring the left or right side during behavioural activities) is an expression of brain functional asymmetry and a strong determinant of fish behaviour. Lateralized individuals show higher performance in cognitive tasks⁶⁴, schooling behaviour⁶⁵ and escape reactivity⁶⁶. Lateralization in juvenile *A. polyacanthus* was determined using a detour test in a two-way T-maze using methods similar to those described by³¹. The two-way T-maze consisted of an experimental arena (60 cm \times 30 cm \times 20 cm), with a runway in the middle (25 cm \times 2 cm, length \times width), and at both ends of the runway (2 cm ahead of the runway) an opaque barrier (12 cm \times 12 cm \times 1 cm) was positioned perpendicular to the runway. The maze was filled to a depth of 4 cm with the respective treatment water of the fish being tested, being changed after each trial. A single fish was placed at one end of the T-maze and given a 3 min habituation

period, during which time it could explore the apparatus. At the end of the habituation period the fish was gently guided into the runway using a plastic rod with the observer standing directly behind the fish (the plastic rod was never placed closer than approximately twice the body length of the fish). At this point to minimise human interference affecting direction turned the observer slowly stepped back from the maze and the fish was allowed to swim to the end of the runway. In instances when a fish did not swim to the end, encouragement was provided by gently moving the plastic rod around at the beginning of the runway. Direction choice was recorded as the first direction turned when the fish exited the runway. Ten consecutive runs were recorded per fish. Twenty fish from each clutch (ten per tank) were tested per CO₂ treatment. To account for any possible asymmetry in the maze, turns were recorded alternately on the two ends of the runway. Turning preference (i.e. bias in left or right turns) at the population level was assessed using the relative lateralization index (L_R , from -100 to $+100$, indicating complete preference for left and right turning, respectively) according to the following formula: $L_R = [(Turn\ to\ the\ right - Turn\ to\ the\ left) / (Turn\ to\ the\ right + Turn\ to\ the\ left)] * 100$. The strength of lateralization (irrespective of its direction) was also assessed at the individual-level using the absolute lateralization index L_A (ranging from 0 (an individual that turned in equal proportion to the right and to the left) to 100 (an individual that turned right or left on all 10 trials)). Lateralization trials in experiment two were performed with the observer blinded to the experimental treatments.

Predator cue trials. The ability to detect and elicit appropriate antipredator behaviour is critical for survival, especially in early life-stages that experience a greater predation threat⁶⁷. The response of juvenile *Am. percula* to a predator cue was tested in a two-channel choice flume using methods similar to those described by²⁹. The flume combination was predator cue water *versus* untreated water. Water at the same pCO_2 level from two different sources (9 L buckets) was gravity fed into the choice flume, which was divided down half of its length. A constant flow rate of $100\ ml\ min^{-1}$ was maintained and monitored using a flow meter and dye test after every water change. Water was changed after each trial. Fish were tested under the mean pCO_2 level of their respective treatments (i.e. fish reared under both 1000 and $1000 \pm 300\ \mu atm$ were tested at $1000\ \mu atm$), due to the logistical difficulties involved in manipulating predator cue water pH across a daily cycle. While this resulted in fish from cycling treatments experiencing a change in pCO_2 between experimental and test water, recent work has shown this has no effect on the response of *Am. percula* to a predator cue at far greater changes than experienced in this study²⁹. For each trial, a single test fish was placed in the centre of the downstream end of the choice flume and given a 2 min acclimation period. The position of the fish was then recorded every five seconds for a total of 2 min. A rest period of 4 min followed, during which time the water sources were switched to eliminate potential side preferences. The position of the fish was then once again recorded every five seconds for a total of 2 min. Fish were not disturbed during the trial. Temperatures during the trials were kept within $1\ ^\circ C$ of the temperature in the rearing tanks. Eight fish from each clutch were tested per pCO_2 treatment (4 per tank). Predator cues were obtained from three common coral-cod, *Cephalopholis miniatus*, as described by ref. 29. Response to predator cue was assessed as the percentage of time spent in the cue water. In experiment one, the control fish from one clutch exhibited no response to the predator cue (i.e. did not avoid the predator cue) and so this clutch was excluded from data analysis.

Statistical analyses. The effects of pCO_2 treatment on absolute lateralization (L_A), relative lateralization (L_R) and percentage time spent in cue water were tested using mixed-effects logistic regressions⁶⁸. Models for L_A data from experiment one and predator cue data from experiments one and two were over dispersed and so were re-run using a penalised quasi-likelihood. In all models, parental pair and tank were included as random factors, with tank nested within parental pair. Pairwise comparisons were performed using Tukey's post hoc tests. To determine if a treatment group demonstrated a turning direction preference Pearson's Chi-square tests were used, where we expected a 50:50 ratio for left/right turning preference. Finally, differences in the relative frequency distribution of L_R between treatments were tested using Kolmogorov-smirnov tests. Mixed-effects logistic regressions with and without penalised quasi-likelihood were conducted in R version 3.3.2⁶⁹ using the lme4⁷⁰ and MASS⁷¹ packages respectively. Pairwise comparisons were conducted using the multcomp⁷² package. Pearson's Chi-square tests were performed using Minitab 17.

Data availability. The datasets generated during and analysed during the current study are available from the corresponding author on request or via the Tropical Research Data Hub (doi:10.4225/28/5923bfed71f8d).

References

1. Doney, S. C., Fabry, V. J., Feely, R. A. & Kleypas, J. A. Ocean acidification: the other CO₂ problem. *Ann. Rev. Mar. Sci.* **1**, 169–192 (2009).
2. Hofmann, G. E. *et al.* High-frequency dynamics of ocean pH: a multi-ecosystem comparison. *PLoS One* **6**, e28983 (2011).
3. Duarte, C. M. *et al.* Is ocean acidification an open-ocean syndrome? Understanding anthropogenic impacts on seawater pH. *Estuaries and Coasts* **36**, 221–236 (2013).
4. Waldbusser, G. G. & Salisbury, J. E. Ocean acidification in the coastal zone from an organism's perspective: multiple system parameters, frequency domains, and habitats. *Ann. Rev. Mar. Sci.* **6**, 221–247 (2014).
5. Shaw, E. C., McNeil, B. I., Tilbrook, B., Matear, R. & Bates, M. L. Anthropogenic changes to seawater buffer capacity combined with natural reef metabolism induce extreme future coral reef CO₂ conditions. *Glob. Chang. Biol.* **19**, 1632–1641 (2013).
6. McNeil, B. I. & Sasse, T. P. Future ocean hypercapnia driven by anthropogenic amplification of the natural CO₂ cycle. *Nature* **529**, 383–386 (2016).
7. McElhany, P. & Busch, S. D. Appropriate pCO_2 treatments in ocean acidification experiments. *Mar. Biol.* **160**, 1807–1812 (2013).
8. Wahl, M., Saderne, V. & Sawall, Y. How good are we at assessing the impact of ocean acidification in coastal systems? Limitations, omissions and strengths of commonly used experimental approaches with special emphasis on the neglected role of fluctuations. *Mar. Freshw. Res.* **67**, 25–36 (2016).
9. Wittmann, A. C. & Pörtner, H. Sensitivities of extant animal taxa to ocean acidification. *Nat. Clim. Chang.* **3**, 995–1001 (2013).

10. Nagelkerken, I. & Munday, P. L. Animal behaviour shapes the ecological effects of ocean acidification and warming: moving from individual to community-level responses. *Glob. Chang. Biol.* **22**, 974–989 (2016).
11. Kroeker, K. J. *et al.* Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. *Glob. Chang. Biol.* **19**, 1884–1896 (2013).
12. Dufault, A. M., Cumbo, V. R., Fan, T. & Edmunds, P. J. Effects of diurnally oscillating pCO₂ on the calcification and survival of coral recruits. *Proc. R. Soc. B Biol. Sci.* **279**, 2951–2958 (2012).
13. Comeau, S., Edmunds, P. J., Spindel, N. B. & Carpenter, R. C. Diel pCO₂ oscillations modulate the response of the coral *Acropora hyacinthus* to ocean acidification. *Mar. Ecol. Prog. Ser.* **501**, 99–111 (2014).
14. Frieder, C. A., Gonzalez, J. P., Bockmon, E. E., Navarro, M. O. & Levin, L. A. Can variable pH and low oxygen moderate ocean acidification outcomes for mussel larvae? *Glob. Chang. Biol.* **20**, 754–764 (2014).
15. Cornwall, C. E. *et al.* Diurnal fluctuations in seawater pH influence the response of a calcifying macroalga to ocean acidification. *Proc. R. Soc. B Biol. Sci.* **280**, 20132201 (2013).
16. Eriander, L., Wrangle, A. & Havenhand, J. N. Simulated diurnal pH fluctuations radically increase variance in - but not the mean of - growth in the barnacle *Balanus improvisus*. *ICES J. Mar. Sci.*, doi:10.1093/icesjms/fsv214 (2015).
17. Ou, M. *et al.* Responses of pink salmon to CO₂-induced aquatic acidification. *Nat. Clim. Chang.* **5**, 950–955 (2015).
18. Alenius, B. & Munguia, P. Effects of pH variability on the intertidal isopod. *Paradella diana*. *Mar. Freshw. Behav. Physiol.* **45**, 245–259 (2012).
19. Clark, H. R. & Gobler, C. J. Do diurnal fluctuations in CO₂ and dissolved oxygen concentrations provide a refuge from hypoxia and acidification for early life stage bivalves? *Mar. Ecol. Prog. Ser.* **558**, 1–14 (2016).
20. Boyd, P. W. *et al.* Biological responses to environmental heterogeneity under future ocean conditions. *Glob. Chang. Biol.* **22**, 2633–2650 (2016).
21. Vargas, C. A. *et al.* Species-specific responses to ocean acidification should account for local adaptation and adaptive plasticity. *Nat. Ecol. Evol.* **1**, doi:10.1038/s41559-017-0084 (2017).
22. Shaw, E. C., Munday, P. L. & McNeil, B. I. The role of CO₂ variability and exposure time for biological impacts of ocean acidification. *Geophys. Res. Lett.* **40**, 4685–4688 (2013).
23. Heuer, R. M. & Grosell, M. Physiological impacts of elevated carbon dioxide and ocean acidification on fish. *AJP Regul. Integr. Comp. Physiol.* **307**, R1061–R1084 (2014).
24. Clements, J. C. & Hunt, H. L. Marine animal behaviour in a high CO₂ ocean. *Mar. Ecol. Prog. Ser.* **536**, 259–279 (2015).
25. Dixson, D. L., Munday, P. L. & Jones, G. P. Ocean acidification disrupts the innate ability of fish to detect predator olfactory cues. *Ecol. Lett.* **13**, 68–75 (2010).
26. Allan, B. J. M., Domenici, P., McCormick, M. I., Watson, S.-A. & Munday, P. L. Elevated CO₂ affects predator-prey interactions through altered performance. *PLoS One* **8**, e58520 (2013).
27. Munday, P. L. *et al.* Replenishment of fish populations is threatened by ocean acidification. *Proc. Natl. Acad. Sci. USA* **107**, 12930–12934 (2010).
28. Ferrari, M. C. O. *et al.* Intrageneric variation in antipredator responses of coral reef fishes affected by ocean acidification: implications for climate change projections on marine communities. *Glob. Chang. Biol.* **17**, 2980–2986 (2011).
29. Munday, P. L. *et al.* Effects of elevated CO₂ on predator avoidance behaviour by reef fishes is not altered by experimental test water. *PeerJ*, doi:10.7717/peerj.2501 (2016).
30. Domenici, P., Allan, B., McCormick, M. I. & Munday, P. L. Elevated carbon dioxide affects behavioural lateralization in a coral reef fish. *Biol. Lett.* **8**, 78–81 (2012).
31. Welch, M. J., Watson, S.-A., Welsh, J. Q., McCormick, M. I. & Munday, P. L. Effects of elevated CO₂ on fish behaviour undiminished by transgenerational acclimation. *Nat. Clim. Chang.* **4**, 1086–1089 (2014).
32. Chivers, D. P. *et al.* Impaired learning of predators and lower prey survival under elevated CO₂: a consequence of neurotransmitter interference. *Glob. Chang. Biol.* **20**, 515–522 (2014).
33. Ferrari, M. C. O. *et al.* Effects of ocean acidification on learning in coral reef fishes. *PLoS One* **7**, e31478 (2012).
34. Ferrari, M. C. O. *et al.* Putting prey and predator into the CO₂ equation - qualitative and quantitative effects of ocean acidification on predator-prey interactions. *Ecol. Lett.* **14**, 1143–1148 (2011).
35. McCormick, M. I., Watson, S.-A. & Munday, P. L. Ocean acidification reverses competition for space as habitats degrade. *Sci. Rep.* **3**, 3280 (2013).
36. Falter, J. L., Lowe, R. J., Zhang, Z. & McCulloch, M. Physical and biological controls on the carbonate chemistry of coral reef waters: effects of metabolism, wave forcing, sea level, and geomorphology. *PLoS One* **8**, e53303 (2013).
37. Anthony, K. R. N., Kleypas, J. A. & Gattuso, J.-P. Coral reefs modify their seawater carbon chemistry – implications for impacts of ocean acidification. *Glob. Chang. Biol.* **17**, 3655–3666 (2011).
38. Shaw, E. C., McNeil, B. I. & Tilbrook, B. Impacts of ocean acidification in naturally variable coral reef flat ecosystems. *J. Geophys. Res.* **117**, C03038 (2012).
39. Kayanne, H., Suzuki, A. & Saito, H. Diurnal changes in the partial pressure of carbon dioxide in coral reef water. *Science*. **269**, 214–216 (1995).
40. Albright, R., Langdon, C. & Anthony, K. R. N. Dynamics of seawater carbonate chemistry, production, and calcification of a coral reef flat, Central Great Barrier Reef. *Biogeosciences* **10**, 6747–6758 (2013).
41. Kline, D. I. *et al.* Six month *in situ* high-resolution carbonate chemistry and temperature study on a coral reef flat reveals asynchronous pH and temperature anomalies. *PLoS One* **10**, e0127648 (2015).
42. Chan, W. Y. & Eggins, S. M. Calcification responses to diurnal variation in seawater carbonate chemistry by the coral *Acropora formosa*. *Coral Reefs*, doi:10.1007/s00338-017-1567-8 (2017).
43. Nilsson, G. E. *et al.* Near-future carbon dioxide levels alter fish behaviour by interfering with neurotransmitter function. *Nat. Clim. Chang.* **2**, 201–204 (2012).
44. Allan, B. J. M., Miller, G. M., McCormick, M. I., Domenici, P. & Munday, P. L. Parental effects improve escape performance of juvenile reef fish in a high-CO₂ world. *Proc. R. Soc. B Biol. Sci.* **281**, 20132179 (2014).
45. Munday, P. L. *et al.* Ocean acidification impairs olfactory discrimination and homing ability of a marine fish. *Proc. Natl. Acad. Sci. USA* **106**, 1848–1852 (2009).
46. Devine, B. M., Munday, P. L. & Jones, G. P. Rising CO₂ concentrations affect settlement behaviour of larval damselfishes. *Coral Reefs* **31**, 229–238 (2012).
47. Heuer, R. M. & Grosell, M. Physiological impacts of elevated carbon dioxide and ocean acidification on fish. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **307**, R1061–R1084 (2014).
48. Baker, D. W. *et al.* Complete intracellular pH protection during extracellular pH depression is associated with hypercapnia tolerance in white sturgeon, *Acipenser transmontanus*. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **296**, 1868–1880 (2009).
49. Esbaugh, A. J., Heuer, R. & Grosell, M. Impacts of ocean acidification on respiratory gas exchange and acid – base balance in a marine teleost. *Opsanus beta*. *J. Comp. Physiol. B* **182**, 921–934 (2012).
50. Heuer, R. M., Welch, M. J., Rummer, J. L., Munday, P. L. & Grosell, M. Altered brain ion gradients following compensation for elevated CO₂ are linked to behavioural alterations in a coral reef fish. *Sci. Rep.* **6**, 33216 (2016).
51. Wood, C. M., Turner, J. D., Munger, R. S. & Graham, M. S. Control of ventilation in the hypercapnic skate *Raja ocellata*: II. Cerebrospinal fluid and intracellular pH in the brain and other tissues. *Respir. Physiol.* **80**, 279–298 (1990).

52. Lai, F., Jutfelt, F. & Nilsson, G. E. Altered neurotransmitter function in CO₂-exposed stickleback (*Gasterosteus aculeatus*): a temperate model species for ocean acidification research. *Conserv. Physiol.* **3**, doi:10.1093/conphys/cov018 (2015).
53. Peterson, M. S. & Gilmore, R. G. Hematocrit, osmolality and ion concentration in fishes: consideration of circadian patterns in the experimental design. *J. Exp. Mar. Biol. Ecol.* **121**, 73–78 (1988).
54. Dmitriev, A. V. & Mangel, S. C. A circadian clock regulates the pH of the fish retina. *Journal Physiol.* **522**, 77–82 (2000).
55. Schunter, C. *et al.* Molecular signatures of transgenerational response to ocean acidification in a species of reef fish. *Nat. Clim. Chang.* **6**, 1014–1018 (2016).
56. López-patiño, M. A., Rodríguez-illamola, A., Gesto, M., Soengas, J. L. & Míguez, J. M. Changes in plasma melatonin levels and pineal organ melatonin synthesis following acclimation of rainbow trout (*Oncorhynchus mykiss*) to different water salinities. *J. Exp. Biol.* **214**, 928–936 (2011).
57. Sundin, J. & Jutfelt, F. 9–28 d of exposure to elevated pCO₂ reduces avoidance of predator odour but had no effect on behavioural lateralization or swimming activity in a temperate wrasse (*Ctenolabrus rupestris*). *ICES J. Mar. Sci.*, doi:10.1093/icesjms/fsv101 (2015).
58. Hawkins, L. A., Magurran, A. E. & Armstrong, J. D. Ontogenetic learning of predator recognition in hatchery-reared Atlantic salmon. *Salmo salar. Anim. Behav.* **75**, 1663–1671 (2008).
59. Johnstone, K. A., Lubieniecki, K. P., Koop, B. F. & Davidson, W. S. Expression of olfactory receptors in different life stages and life histories of wild Atlantic salmon (*Salmo salar*). *Mol. Ecol.* **20**, 4059–4069 (2011).
60. Pistevidos, J. C. A., Nagelkerken, I., Rossi, T., Olmos, M. & Connell, S. D. Ocean acidification and global warming impair shark hunting behaviour and growth. *Sci. Rep.*, doi:10.1038/srep16293 (2015).
61. Challener, R. C., Robbins, L. L. & McClintock, J. B. Variability of the carbonate chemistry in a shallow, seagrass-dominated ecosystem: implications for ocean acidification experiments. *Mar. Freshw. Res.* **67**, 163–172 (2016).
62. Kavanagh, K. D. Larval brooding in the marine damselfish *Acanthochromis polyacanthus* (Pomacentridae) is correlated with highly divergent morphology, ontogeny and life-history traits. *Bull. Mar. Sci.* **66**, 321–337 (2000).
63. Bay, L. K., Crozier, R. H. & Caley, M. J. The relationship between population genetic structure and pelagic larval duration in coral reef fishes on the Great Barrier Reef. *Mar. Biol.* **149**, 1247–1256 (2006).
64. Dadda, M. & Bisazza, A. Does brain asymmetry allow efficient performance of simultaneous tasks? *Anim. Behav.* **72**, 523–529 (2006).
65. Bisazza, A. & Dadda, M. Enhanced schooling performance in lateralized fishes. *Proc. R. Soc. B Biol. Sci.* **272**, 1677–1681 (2005).
66. Dadda, M., Koolhaas, W. H. & Domenici, P. Behavioural asymmetry affects escape performance in a teleost fish. *Biol. Lett.* **6**, 414–417 (2010).
67. Almany, G. R. & Webster, M. S. The predation gauntlet: early post-settlement mortality in reef fishes. *Coral Reefs* **25**, 19–22 (2006).
68. Warton, D. I. & Hui, F. K. C. The arcsine is asinine: the analysis of proportions in ecology. *Ecology* **92**, 3–10 (2011).
69. R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/> (2016).
70. Bates, D., Mächler, M., Bolker, B. M. & Walker, S. C. Fitting linear mixed-effects models using lme4. *J. Statistical Softw.* **67**, 1–48 (2015).
71. Venables, W. & Ripley, B. Modern applied statistics with S. (Springer, 2012).
72. Hothorn, T., Bretz, F. & Westfall, P. Simultaneous inference in general parametric models. *Biometric J.* **50**, 346–363 (2008).

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Author Contributions

M.D.J. and P.L.M. designed the experiment. M.D.J. performed all experiments and statistical analysis. C.H. assisted with experimental procedures at SeaSim. M.D.J. drafted the first version of the manuscript with input from P.L.M., M.I.M. and C.H.

Additional Information

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